

In vivo antimicrobial assay of the PPO system using horseshoe crab as the model animal

Naxin Jiang

National University of Singapore

Nguan Soon Tan

National University of Singapore

Bow Ho

National University of Singapore

Jeak Ling Ding

National University of Singapore

Method Article

Keywords: horseshoe crab, prophenoloxidase (PPO), antimicrobial

Posted Date: October 31st, 2007

DOI: <https://doi.org/10.1038/nprot.2007.482>

License: © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Abstract

Introduction

To demonstrate the ability of microbial factor-activated PPO activity in clearing the invading pathogen *in vivo*, we infected horseshoe crabs in the presence or absence of PO-specific inhibitor, PTU¹ or kojic acid². A comparison of the remnant bacterial load under these conditions should help to clarify the specific contribution of PO, if any, to the antimicrobial activity. Previously, it was reported that HMC/PPO is activated by host intracellular factors released through LPS-dependent degranulation of hemocytes. To avoid provocation of PPO by such cellular components and to unequivocally demonstrate that the microbial factor-activated PPO contributes to the antimicrobial defense, Gram-positive bacteria lacking LPS were used to avoid LPS-induced hemocyte lysis. To this end, the *S. aureus* laboratory strains, PC1839 (V8 protease-producing) and AK3 (V8 protease inactive mutant), were injected into the animals.

Procedure

1. Culture the Gram-positive bacterial strains under pyrogen-free condition as described in protocols #4 and #5.
2. Adjust the bacterial population to 10^6 - 10^7 cfu/ml with pyrogen free 3% NaCl (isotonic to the horseshoe crab hemolymph).
3. Inject the horseshoe crab intracardially with 10^5 - 10^6 cfu bacteria /100 gram body, using #23 needle.
4. At 30 min post injection, collect hemolymph from the horseshoe crab by cardiac puncture using #18 needle.
5. Immediately after collection, remove the hemocytes by centrifuging the hemolymph at 150 x g for 10 min at room temperature.
6. Quantify the remnant bacterial load in the extracellular milieu by applying 100 μ l of the cell-free hemolymph (from step 5) to the nutrient agar plate and incubate at 37 °C overnight.
7. In order to confirm the contribution of PO activity in the bacterial clearance, include 5 mM PTU or 5 mM kojic acid in the bacterial injection to block *in vivo* PO activity, if any.

Anticipated Results

As shown in Figure 7, at 30 min post-injection, the remnant bacterial load in the cell free hemolymph (injected without PTU or kojic acid) is less than 10^4 cfu/ml. Co-injection of PTU or kojic acid with the bacteria results in significantly higher bacterial load of the extracellular-protease positive strains such as *S. aureus* PC1839. In contrast, the clearance of the extracellular-protease-negative strains such as AK3, is unaffected by PTU or kojic acid.

References

1. Nellaiappan, K. & Sugumaran, M. On the presence of prophenoloxidase in the hemolymph of the horseshoe crab, *Limulus*. *Comp Biochem Physiol B Biochem Mol Biol*. **113**, 163-168 (1996).
2. Dowd, P. F. Relative inhibition of insect phenoloxidase by cyclic fungal metabolites from insect and plant pathogens. *Natural Toxins*, **7** (6), 337-341 (1999)

Figures

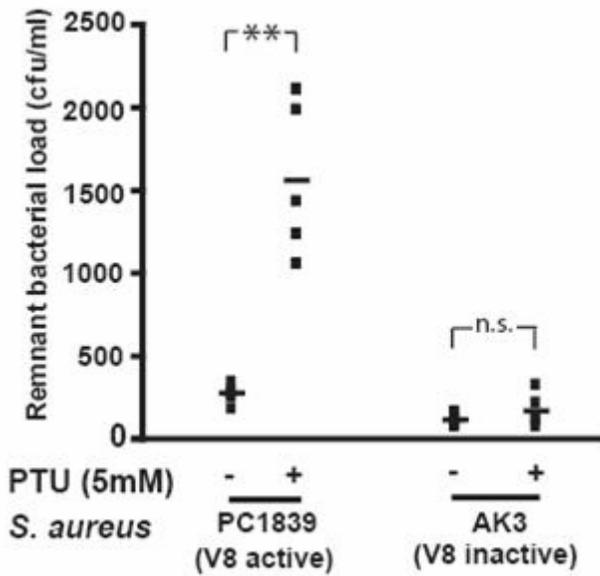


Figure 1

The PO triggered by the microbial protease contributes to in vivo antimicrobial activity. Injection of the *S. aureus* laboratory strains, PC1839 and AK3, which are active V8 protease-positive and -negative, respectively into the horseshoe crab at 105 cfu/100 gram body weight, in the presence or absence of 5 mM PTU. At 30 min post injection, the remaining bacterial load in the hemolymph was measured. The protease-positive strain which specifically evoked the ROS-production by HMC/PPO, is in turn killed effectively. However, co-injection with PTU inhibited the HMC/PPO activity, and allowed the bacteria to remain viable in the host. On the other hand, the clearance of the V8-inactive strain was unaffected by PTU. This is probably due to the antimicrobial effects of parallel PO-independent mechanisms (see main text for further explanations).