

# Immunostaining of spindle checkpoint proteins on *Drosophila* mitotic chromosomes from larval brains

Mariarosaria Musar (✉ [mariarosaria.musaro@uniroma1.it](mailto:mariarosaria.musaro@uniroma1.it))

DiSTeBA, Universit del Salento, Lecce, 73100 Italy; Istituto di Biologia e Patologia Molecolari del CNR and Dipartimento di Genetica e Biologia Molecolare, Universit di Roma La Sapienza, Roma, 00185 Italy;

---

## Method Article

**Keywords:** chromosome cytology, immunostaining

**Posted Date:** February 6th, 2008

**DOI:** <https://doi.org/10.1038/nprot.2008.25>

**License:**  This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

---

# Abstract

## Introduction

We describe a very simple method to immunostain *Drosophila* mitotic chromosomes for spindle checkpoint proteins and kinetochore components. By using appropriate primary antibodies, this procedure allowed us to detect Zw10, Zwilch, Cenp-C, Cenp-meta and BubR1 on kinetochores of larval brain chromosomes. However, the same method can be used for detecting also structural components of chromosomes (our unpublished results).

## Reagents

Saline (0.7% NaCl), Hypotonic solution (0.5% sodium citrate). Fixing solution (1.8% Formaldehyde, 45% acetic acid in dH<sub>2</sub>O) Liquid nitrogen Ethanol PBS-T (PBS containing 0.1% TritonX). Appropriate primary and secondary antibodies Vectashield medium H-1200 with DAPI (4,6 diamidino-2-phenylindole)

## Equipment

Slides and coverslips Humid box Tweezers Dissecting Scope Epifluorescence Microscope

## Procedure

1. Dissect *Drosophila* larval brains in saline (0.7% NaCl),
2. Treat dissected brains for 10 min with a hypotonic solution of 0.5% sodium citrate.
3. Fix for 5 min in a drop of fixing solution (1.8% Formaldehyde, 45% acetic acid) on a coverslip.
4. Gently lean a clean slide over the coverslip and squash the brain in the same fixing solution
5. Freeze the slide in liquid nitrogen
6. Remove the coverslip and immerse the slide in cold ethanol (-20°C) for 10 minutes.
7. Wash the slide in PBS-T (PBS containing 0.1% TritonX).
8. Incubate the slide overnight with appropriate primary antibody in a humid box at 4°C
9. The next day, wash the slide twice in PBS-T for 10 minutes
10. Incubate the slide with the secondary antibody for 2 h at room temperature, in a humid box.
11. Wash the slide twice in PBS-T for 10 minutes and let it air dry.
12. Mount the slide in Vectashield medium H-1200 with DAPI (4,6 diamidino-2-phenylindole; Vector Laboratories, Burlingame, CA) to stain DNA and reduce fluorescence fading.
13. Analyze the immunostaining with a epifluorescence microscope