

1 **The double rhythm development and characteristics of the**
2 ***Gasterophilus pecorum* (Diptera: Gasterophilidae) population in**
3 **desert steppe**

4 Ke Zhang^{1, #}, Heqing Huang^{2, #}, Ran Zhou¹, Boru Zhang³, Chen Wang⁴, Make Ente⁵,
5 Boling Li⁶, Dong Zhang¹, Kai Li^{1*}

6
7 # Ke Zhang and Heqing Huang contributed equally to this work.

8 ¹ Key Laboratory of Non-Invasive Research Technology for Endangered Species, College of
9 Nature Conservation, Beijing Forestry University, Beijing 100083, China

10 ² Chongqing Academy of Environmental Science, Chongqing 401147, China

11 ³ Qinhuangdao Forestry Bureau, Qinhuangdao 066004, Hebei, China.

12 ⁴ Mt. Kalamaili Ungulate Nature Reserve, Changji 381100, Xinjiang, China

13 ⁵ Xinjiang Research Centre for Breeding Przewalski's Horse, Urumqi 831700, Xinjiang, China.

14 ⁶ China National Environment Monitoring Centre, Beijing 100012, China.

15 *Corresponding author: likai_sino@sina.com, 86 13811778282

16
17 **Abstract**

18 **Background:** The departure of the mature larvae of the horse stomach bot fly from the host indicates
19 the beginning of a new infection period, and the *Gasterophilus pecorum* becomes the dominant
20 species in the desert steppe, showing its special biological characteristics. The population dynamics
21 of *G. pecorum* were studied to reveal the population development rule of *G. pecorum* in the arid
22 desert steppe.

23 **Method:** The larvae were collected and recorded in the newly excreted feces by tracking the
24 Przewalski's horses (*Equus przewalskii*), meanwhile, the larval pupation experiments were carried
25 out under natural conditions.

26 **Results:** (a) There was a positive correlation between the survival rate and the number of larvae (r
27 = 0.630, $p < 0.01$), indicating that the species development had the characteristics of centralized
28 occurrence; (b) The main periods of mature larvae discharge were from early April to early May

29 (peak I) and from mid-August to early September (peak II), and the larval population curve showed
30 a sudden spike in increase and gradual decrease at both peaks; under higher temperature, the number
31 of adults from peak II had higher survival rate, higher pupation rate, higher emergence rate and less
32 eclosion time than that of peak I; (c) Although it has one generation a year, the occurrence peak
33 twice annually displaying a unique “ double rhythm development ” phenomenon, which forms a
34 “ dual rhythm parasitism ” pressure on the local host. This phenomenon is very rare in the study of
35 insect life history, especially in the parasite epidemiology.

36 **Conclusion:** The natural discharge period of the *G. pecorum* larvae in Kalamaili Nature Reserve
37 (KNR) is longer than 7 months and have the potentially long term infection effect on the host. The
38 above phenomenon is one important reason for the local equine animals to be severely infected with
39 equine myiasis.

40 **Keywords:** Desert Steppe, *Gasterophilus pecorum*, Mature larvae, Population dynamics, Double
41 rhythm development, Survival rate

42

43 **Background**

44 The horse stomach bot flies (*Gasterophilus* spp.) are common obligate parasites in equids [1–2].
45 *Gasterophilus* larvae parasitize the digestive tract of equine animals and causes inflammation, ulcer,
46 hernia and other phenomena [3–5]. The larvae absorb nutrition from the host and secrete toxins, and
47 may leads to host death when infected severely [6–7]. The genus *Gasterophilus* includes nine
48 species in the world [8–9], and six of them are recorded in China [10]. The infection of bot flies in
49 local equine is 100% in Kalamaili Nature Reserve (KNR), Xinjiang located in the desert grassland
50 [11]. Among the 6 species of horse stomach bot flies, *G. pecorum* infections are extremely high,
51 accounting for 89-98% of all bot flies infections [11–12], which is different from the occurrence of
52 *G. intestinalis* and *G. nasalis* in other regions as the dominant species [13–14].

53 The horse stomach flies are insect of complete metamorphosis and have four developmental stages,
54 namely, egg, larva, pupa and adult, one generation a year [8]. Them develop in the digestive tract of
55 equines until the mature larva leaves the host, thus starting a new life cycle. The mature larva is the
56 first state of the host’s life in vitro, and its population dynamics determines the subsequent
57 population development process. The most significant feature in the life cycle of the *G. pecorum* is

58 its "unusual" reproductive strategy of laying eggs on grass [8]. Most of the studies on *G. pecorum*
59 originated from the early biological observation by Chereshev in Kazakhstan [15], the mature
60 larvae mainly appear in early August to early September. However, a study reported that a
61 significant number of *G. pecorum* larvae were collected in KNR to study the development period
62 of pupae, and it was considered that the high incidence period of mature larvae was in spring [16].
63 Based on the differences between some recent findings and the existing life history records of *G.*
64 *pecorum*, a systematic study of the population dynamics and developmental process of the local *G.*
65 *pecorum* in vitro of the host was carried out in order to understand the development characteristics
66 of this species in the desert steppe.

67 Insect are poikilothermal animals whose metabolism, life cycle and lifespan are subject to the
68 temperature of the environment [17–18].The horse stomach flies have a relatively stable
69 environment in the digestive tract of host, and once discharged from the host, it will be affected by
70 the external temperature [19]. Horse stomach flies are thermophilic insects that tend to live at high
71 temperatures (in warm regions or on a warm-blooded host) [20].The higher temperature is
72 conducive to accelerate the development rate of insects in the suitable temperature range [21–22].
73 In addition, the survival rate of metamorphosis insects between different states is also affected by
74 temperature [23–25]. The developmental response of insects to temperature is important in
75 understanding the ecology of insect life histories [26]. So, we conducted an experiment to
76 investigate the effect of different temperatures on the developmental rate, survival, pupation,
77 emergence of *G. pecorum* in order to provide parameters for forecasting and management systems
78 in KNR.

79

80 **Result**

81 **Population dynamics of mature larvae of *G. pecorum***

82 A total of 2,021 piles of equine feces were examined, where 443 piles (21.92%) contained *G.*
83 *pecorum* larvae (Table 1). The proportion of feces containing larvae (PF) in early April and mid-to-
84 late August were 56.03% and 53.23%, respectively. There were two obvious larval peaks with a
85 significant range of changes. In the following May and September, the PF gradually decreased and
86 remained low. Among them, the PF was less than 20% during the three periods of March, mid-to-

87 late May to the end of July, and mid-to-late September.

88 Table 1 The number of equine feces and *G. pecorum* larvae collected in KNR in 2018

Date	Feces investigated (N _i)	Feces containing larvae (n _i)	PF n _i /N _i (%)	Larvae (m _i)	NL (m _i /N _i)	Proportion of larvae (m _i /Σm _i) (%)
3.15	98	2	2.04	2	0.02	0.28
3.22	94	6	6.38	7	0.07	0.99
3.29	48	3	6.25	5	0.10	0.71
4.05	59	34	57.63	103	1.75	14.63
4.12	82	45	54.88	94	1.15	13.35
4.19	71	37	52.11	62	0.87	8.81
4.26	58	18	31.03	40	0.69	5.68
5.3	59	27	45.76	30	0.51	4.26
5.10	54	13	24.07	14	0.26	1.99
5.17	43	8	18.60	8	0.19	1.14
5.24	55	9	16.36	9	0.16	1.28
5.31	51	4	7.84	4	0.08	0.57
6.7	47	5	10.64	5	0.11	0.71
6.14	57	3	5.26	3	0.05	0.43
6.21	38	4	10.53	5	0.13	0.71
6.28	88	5	5.68	5	0.06	0.71
7.5	75	5	6.67	5	0.07	0.71
7.12	100	3	3.00	3	0.03	0.43
7.19	50	2	4.00	2	0.04	0.28
7.26	100	4	4.00	4	0.04	0.57
8.2	80	1	1.25	1	0.01	0.14
8.9	39	3	7.69	3	0.08	0.43
8.16	54	18	33.33	30	0.56	4.26
8.23	108	61	56.48	108	1.00	15.34
8.30	93	46	49.46	60	0.65	8.52
9.6	79	28	35.44	36	0.46	5.11
9.13	72	22	30.56	25	0.35	3.55
9.20	100	17	17.00	20	0.20	2.84
9.27	69	10	14.49	11	0.16	1.56
Total	2021	443	21.92	704	0.35	100

89 Note: The corresponding information of each date represents the larva data between the time point and the previous
 90 time point, for example, 9.27 represents the larva collection during 9.21-9.27.

91

92 A total of 704 larvae of *G. pecorum* were collected in this study. The average number of larvae per
 93 feces (NL) was 0.35 during the entire investigation period, among which the highest was in early
 94 April with an average of 1.40, followed by mid-to-late April and mid-to-late August, with 0.79 and

95 0.84 respectively. The NL showed the same trend as the PF (Figure 1).
96 The percentage of larvae in April and August were 42.47% and 28.55%, respectively. Among them,
97 27.98% and 14.79% were in early and mid-to-late April, and 23.86% in mid-to-late August,
98 respectively, which was significantly higher than the other periods ($p < 0.05$). The number of larvae
99 collected from early April to early May and from mid-August to early September accounted for
100 48.72% and 32.52% of the total number of larvae, respectively, and the cumulative proportion of
101 the two stages accounted for 81.24%. Therefore, the main periods of mature larvae discharge in
102 KNR were from early April to early May (peak I) and from mid-August to early September (peak
103 II) (Figure 2) while maintaining continuous larvae production throughout March to September.
104 The NL was derived from obtaining an average of 10 piles of feces per adult Przewalski's horse per
105 day, as observed from 10 wild horses in temporary captivity. These horses defecated an average of
106 10.1 piles of feces daily. It is estimated that 749 larvae of *G. pecorum* were discharged from each
107 horse, which was close to the annual average infection amount of *G. pecorum* released locally [11].
108 The results showed that with time, the cumulative number of larvae collected during the survey
109 period showed a "double S" growth curve (Figure 3). The two spikes were concentrated in April and
110 August, the slope of the curve showed that the first growth of the cumulative growth curve of total
111 larvae (Figure 3, A) and survival larvae (Figure 3, B) were increased fastest on April 5, and the
112 second growth curve (Figure 3, C) and (Figure 3, D) were both on August 23.

113

114 **Population development analysis**

115 The results showed that the survival rates of the two peaks were 69.57% (peak I) and 73.27% (peak
116 II) ($p = 0.183$, $t = -1.727$), with higher pupation rate from peak II (89.19%) than that of peak I
117 (66.83%) ($p = 0.002 < 0.05$, $t = -10.547$). The eclosion rate of peak II (87.88%) was also higher than
118 that of peak I (63.31%) ($p = 0.002$, $t = -9.525$) (Figure 4, B).

119 The average pupal period of the two peak stages lasted for 34.05 days and 20.2 days respectively (p
120 < 0.001 , $t = 15.513$) (Figure 4, A). The longest and shortest development period of peak I was 39
121 days and 26 days, and the longest and shortest development time of peak II was 24 days and 18 days.
122 Spearman correlation analysis showed that there was a positive correlation between the survival rate
123 and the number of larvae ($r = 0.630$, $p < 0.01$). That is, the survival rate of *G. pecorum* larvae was

124 higher during the two peak periods.

125 Population decreases in three stages throughout the development from larvae (N) to adults (N');
126 survival rate (α), pupation rate (β) and eclosion rate (γ), yielding:

$$127 \quad N' = \alpha\beta\gamma N \quad (6)$$

128 The number of adults developed from larvae in peak I and peak II were N₁' and N₂', respectively,
129 thus:

$$130 \quad N_1' = \alpha_1\beta_1\gamma_1 N_1 = 69.57\% \times 66.83\% \times 63.31\% \times (51.70\% \times N) = 15.22\%N$$

$$131 \quad N_2' = \alpha_2\beta_2\gamma_2 N_2 = 73.27\% \times 89.19\% \times 87.88\% \times (41.76\% \times N) = 23.98\%N$$

132 The results showed that the larvae that emerged into adults from peak I accounted for 15.22% of the
133 total larvae, and peak II was 23.98%. The number of larvae in peak I was 1.48 times as much as that
134 in peak II, but the higher survival rate, pupation rate and emergence rate in peak II resulted in 1.32
135 times more of adults than that of peak I.

136 Table 2 The sex ratio of *G. pecorum* adults in the two peak periods

Period	Proportion of females (%)	Proportion of males (%)	Total (%)	Male to female ratio
Peak I	57.95	42.05	100	0.73
Peak II	43.10	56.90	100	1.32

137

138 The number of female adults in peak I and peak II were N_{FI} and N_{FII}, respectively, therefore:

$$139 \quad N_{FI} = 15.22\%N \times 57.95\% = 0.09\%N$$

$$140 \quad N_{FII} = 23.98\%N \times 43.10\% = 0.10\%N$$

141 In addition, there were differences in the sex ratio between the two peak periods. The ratio of male
142 to female in peak I was 0.73, which was lower than that in peak II (0.76) (Table 2). When comparing
143 the proportion of female adults in the total larvae in the two peak periods, it was found that peak II
144 (0.10%) was slightly higher than peak I (0.09%), signifying that peak II had greater infective
145 potential to equine animals in KNR.

146

147 **Temperature characteristics during larvae discharge period**

148 The daily temperature of KNR depicted that the temperature rose rapidly from March to April, and
149 slowed down from April to July, and began to decline after reaching the highest temperature of 38 °C
150 (July 20) (Figure 5, A). The maximum daily temperature difference is 20 °C, the minimum and the
151 average daily temperature differences are 3 °C and 12.78 °C respectively. After fitting the trend of
152 temperature change, it was found that the curve showed a parabola characteristic of “rising first and
153 then decreasing” ($y = 0.8570 + 3.7449x + 0.1031x^2 - 0.0240x^3$, $R^2=91.18\%$) (Figure 5, B).
154 The maximum value of the fitting curve was found to be July 19.

155

156 **Discussion**

157 Insect population dynamics is an important part of insect ecology research [30–31]. In the reports
158 on the fauna of horse stomach bot fly, the prevalence of *G. pecorum* was low in most countries and
159 regions [2, 13, 32], but it is more common in the digestive tract of equines in Mongolia-Xinjiang
160 and Qinghai-Tibet regions of China [12, 33], central and northern Kazakhstan [14], the Republic of
161 Mongolia [34] and the Yakut Republic of Russia [35]. This high incidence of this species showed a
162 certain regionality. In terms of basic research on stomach bot fly, a study was carried out on the
163 pupal development period of *G. pecorum* and presented a prediction model for adult occurrence
164 period [16]. The study on the egg development period and the survival period of the first instar
165 larvae of *G. pecorum* has also been completed (in submission). The results will further improve the
166 understanding of this species and contribute to the understanding of its epidemiology.

167 The parasites are also adaptable to the environment and have different performance characteristics
168 in different environments [36–38]. The KNR is a desert grassland with high temperature and
169 drought in summer, severe cold and long winter, and little annual precipitation. The unique
170 environmental conditions have formed a specific occurrence of *G. pecorum*. This special situation
171 is different from that of the early discovery in Kazakhstan that the larvae of *G. pecorum* were
172 excreted only in August [15]. It is also different from the report that the larvae of *G. pecorum* were
173 found in January, March and July in southern Italy [32]. And different from that reported in South
174 Africa that the adult of *G. pecorum* only appeared in February to May and August [8].

175 Phenology affects the development dynamics of insects [39–40]. The development rhythm of some
176 insects of one generation a year is closely related to phenology [41–42]. With the change of

177 phenology, the population dynamics showed a peak period of correlation [43]. In this study, the
178 larval population of *G. pecorum* indicated two peaks annually, which were closely related to the
179 special arid climate of the desert steppe. In the desert steppe, water is the most crucial factor for life
180 [44–45], therefore, many plants derived characteristics of rapid development of ephemeral plants.
181 Due to the characteristics of local precipitation [46], some *Stipa* sp. showed a special secondary
182 growth phenomenon after adapting to the environment [47]. Through field investigation, we found
183 that the local dominant plant *Stipa* began to resume growth at the end of March and early April,
184 head sprouting in May, and began to regenerate tender leaves in late August, which confirmed the
185 phenomenon of secondary growth. Some studies have shown that once water conditions are suitable,
186 many desert plants formed the characteristics of rapid development and life cycle [48–50], and the
187 effects are also reflected in the ecosystem based on this: the phenomenon of simultaneous
188 occurrence of *G. pecorum* and vector plants. Different from the other five species which lay eggs
189 directly on the horse, the female of *G. pecorum* lay eggs on *Stipa* [51]. Under the influence of
190 phenology, the secondary growth of *Stipa* was observed, and parasite population dynamics matched
191 perfectly with the growth of vector plants and phenological changes, which also led to the local
192 double rhythm development phenomenon of *G. pecorum*. This phenomenon indicates that there are
193 two centralized infections per year for the host, which is different from the annual infection of
194 parasitic linear animals [52–53], also differs from the special infection types associated with
195 phenology, such as most arthropod infections [54–56].

196 In the appropriate temperature range, the higher the temperature, the more beneficial the
197 development of insects [57]. The average temperature of peak I (11.3 ± 5.3 °C) was significantly
198 lower than peak II (24.4 ± 2.7 °C) ($t = -11.083, p < 0.001$), which affected the survival rate of mature
199 larvae. In the subsequent pupal development stage, the environmental temperature in peak II is more
200 conducive to the life process of this stage. This resulted in mature larvae of peak II with higher
201 survival rate (SR, PR and ER) and shorter pupal stage, thus eventually exceeding the number of the
202 adults in peak I.

203

204 **Conclusion**

205 The natural discharge period of the *G. pecorum* larvae in Kalamaili Nature Reserve is longer

206 than 7 months, and it's the first time that the unique phenomenon of "double rhythm
207 development" has been found in stomach bot flies. The close relationship between the double
208 rhythm development of *G. pecorum* and the secondary growth of *Stipa* in the desert steppe is an
209 important reason for the high infection rate, high infection amount of *G. pecorum* and the absolute
210 dominance of *Gasterophilus* spp. in local equine animals. The population in peak II had higher
211 survival rate than peak I because of the suitable development conditions. This study demonstrated
212 the highly co-evolutionary phenomenon of the desert steppe ecosystem and revealed the tenacious
213 adaptability of life under adverse conditions.

214

215 **Methods**

216 **Study area**

217 Kalamaili Nature Reserve (KNR) is located in the desert subregion of Northwest China
218 (44°36'~46°00'N, 88°30'~90°03'E), with an area of 18,000 km², an altitude of 600-1,464 m, an
219 average annual temperature of 2.4 °C, an average annual precipitation of 159 mm and an annual
220 evaporation of 2,090 mm. It has a typical temperate continental climate [27–28]. The protected
221 animals are the reintroduced species *Equus Przewalskii* (EN), the endangered species *Equus*
222 *hemionus* (NT), as well as *Gazella subgutturosa* (VU) and *Ovis ammon* (NT), There are also
223 domestic livestock such as horses there seasonally [29].

224

225 **Material**

226 From March to September 2018, the feces of the Przewalski's horses were inspected, with 50-80
227 piles per week. The larvae of *G. pecorum* in the feces were collected, and the number of stomach
228 bot flies was recorded.

$$229 \text{ Proportion of feces containing larvae (PF, \%)} = \frac{\text{Number of feces with larvae}}{\text{Number of feces investigated}} \times 100\%(1)$$

$$230 \text{ Number of larvae per fece (NL)} = \frac{\text{Number of larvae in feces}}{\text{Number of feces investigated}} \quad (2)$$

231 Transparent plastic cups (8 cm in diameter, 6 cm in height) were used as the pupation container of
232 *G. pecorum* larvae, five larvae were placed in each cup, the cup mouth was sealed with gauze and
233 cultured in outdoor shade, the pupation and eclosion behavior of the larvae were observed

234 simultaneously. The number of each insect state was recorded, the survival rate, pupation rate and
235 eclosion rate were counted. The daily temperature was also recorded.

236 **Data analysis**

237 To evaluate the development status of stomach bot flies, the following formulas were utilized:

$$238 \quad \text{Survival rate (SR, \%)} = \frac{\text{Number of surviving larvae in feces}}{\text{Total number of larvae in feces}} \times 100\% \quad (3)$$

$$239 \quad \text{Pupation rate (PR, \%)} = \frac{\text{Number of pupae}}{\text{Number of surviving larvae}} \times 100\% \quad (4)$$

$$240 \quad \text{Eclosion rate (ER, \%)} = \frac{\text{Number of adults}}{\text{Number of pupae}} \times 100\% \quad (5)$$

241

242 Spearman correlation analysis was used to analyze the relationship between the number of mature
243 larvae and its survival rate, and the significant level was set as $\alpha = 0.05$. Data analysis was performed
244 in SPSS 20.0, and the graph was drawn by Sigmaplot 12.0 and Graphpad prism 7.

245

246

247 **List of Abbreviations**

248 KNR - Kalamaili Nature Reserve

249 PF - Proportion of feces containing larvae

250 NL - Number of larvae per feces

251 SR - Survival rate

252 PR - Pupation rate

253 ER - Eclosion rate

254

255 **Acknowledgements**

256 We are grateful to the staff of Kalamaili Nature Reserve for their support and valuable technical
257 assistance.

258

259 **Funding**

260 This work was supported by the National Science Foundation of China (No. 31670538), and the

261 Species Project (2018) of Department for Wildlife and Forest Plants Protection, NFGA of China.

262

263 **Availability of Data and Material**

264 All data generated or analysed during this study are included in this article.

265

266 **Author contributions**

267 K.L., K.Z. and H.Q.H. conceived the study, K.Z. drafted the manuscript, K.Z., H.Q.H. and R.Z.

268 conducted the experiment, K.Z., B.R.Z., C.W. and M.E. carried out the statistics, B.L.L., D.Z. and

269 K.L. revised the manuscript. All authors reviewed the manuscript and approved the final version.

270

271 **Ethics Statement**

272 The study was performed in accordance with the relevant guidelines and regulations regarding

273 animal welfare. All experimental protocols were approved by Wildlife Conservation Office of

274 Altay Prefecture and Beijing Forestry University.

275

276 **Consent for publication**

277 Not applicable.

278

279 **Competing interests**

280 The authors declare no competing interests.

281

282 **Reference**

- 283 1. Royce LA, Rossignol PA, Kubitz ML, Burton FR. Recovery of a second instar *Gasterophilus*
284 larva in a human infant: a case report. *Am J Trop Med Hyg.* 1999;60:403–4.
- 285 2. Mukbel R, Torgerson PR, Abo-Shehada M. Seasonal variations in the abundance of
286 *Gasterophilus* spp. larvae in donkeys in northern Jordan. *Trop Anim Health Pro.* 2001;33:501–9.

- 287 3. Cogley TP, Cogley MC. Inter-relationship between *Gasterophilus* larvae and the horse's
288 gastric and duodenal wall with special reference to penetration. *Vet Parasitol.* 1999;86:127–
289 42.
- 290 4. Gökçen A, Sevgili M, Altaş MG, Camkerten I. Presence of *Gasterophilus* species in Arabian
291 horses in Sanliurfa region. *Turk Soc Parasitol.* 2008;32:337–9.
- 292 5. Moshaverinia A, Baratpour A, Abedi V, Mohammadi-Yekta M. Gasterophilosis in Turkmen
293 horses caused by *Gasterophilus pecorum* (Diptera, Oestridae). *Scientia Parasitologica.*
294 2016;17:49–52.
- 295 6. Smith MA, Mcgarry JW, Kelly DF, Proudman CJ. *Gasterophilus pecorum* in the soft palate
296 of a British pony. *Vet Rec.* 2005;156:283–4.
- 297 7. Pawlas M, Sotysiak Z, Nicpoń J. Existence and pathomorphological picture of gasterophilosis
298 in horses from north-east Poland. *Med Weter.* 2007;63:1377–80.
- 299 8. Zumpt F. Myiasis in Man and Animals in the Old World. In: Morphology, biology and
300 pathogenesis of myiasis-producing flies in systematic order. London: Butterworths; 1965. P.
301 110–128.
- 302 9. Cogley TP. Key to the eggs of the equid stomach bot flies *Gasterophilus* Leach 1817 (Diptera:
303 Gasterophilidae) utilizing scanning electron microscopy. *Syst Entomol.* 1991;16:125–33.
- 304 10. Zhang BR, Huang HQ, Zhang D, Chu HJ, Ma XP, Li K, et al. Genetic diversity of common
305 *Gasterophilus* spp. from distinct habitats in China. *Parasite Vector.* 2018;11:474–85.
- 306 11. Liu SH, Li K, Hu DF. The incidence and species composition of *Gasterophilus* (Diptera,
307 Gasterophilidae) causing equine myiasis in northern Xinjiang, China. *Vet Parasitol*
308 2016;217:36–8.
- 309 12. Huang HQ, Zhang BR, Chu HJ, Zhang D, Li K. *Gasterophilus* (Diptera, Gasterophilidae)
310 infestation of equids in the Kalamaili Nature Reserve, China. *Parasite.* 2016;23:36.
- 311 13. Pandey VS, Ouhelli H, Verhulst A. Epidemiological observations on *Gasterophilus intestinalis*
312 and *Gasterophilus nasalis* in donkeys from Morocco. *Veterinary Parasitology* 1992;41:285–92.
- 313 14. Ibrayev B, Lider L, Bauer C. *Gasterophilus* spp. infections in horses from northern and central
314 Kazakhstan. *Vet Parasitol.* 2015;207:94–8.
- 315 15. Chereshev NA. Biological peculiarities of the botfly *Gasterophilus pecorum* Fabr (Diptera:
316 Gasterophilidae). *Dokl Akad Nauk SSSR.* 1951;77:765–8 (in Russian).
- 317 16. Wang KH, Zhang D, Hu DF, Chu HJ, Cao J, Li K, et al. Developmental threshold temperature
318 and effective accumulated temperature for pupae of *Gasterophilus pecorum*. *Chin J Vector*
319 *Biol & Control.* 2015;26:572–5(in Chinese).
- 320 17. Gordon P, Harder LD, Mutch RA. Development of aquatic insect eggs in relation to
321 temperature and strategies for dealing with different thermal environments. *Biol J Linn Soc.*

- 1996;58:221–44.
18. Potter K, Davidowitz G, Woods HA. Insect eggs protected from high temperatures by limited homeothermy of plant leaves. *J Exp Biol.* 2009;212:3448–54.
19. Knapp FW, Sukhapesna V, Lyons ET, Drudge JH. Development of third-instar *Gasterophilus intestinalis* artificially removed from the stomachs of horses. *Ann Entomol Soc Am.* 1979;72:331–3.
20. Vincent HR, Ring TC. Encyclopedia of insects (Second edition). In: Temperature, effects on development and growth. Oxford: Academic Press; 2009. p. 990–993.
21. Ikemoto T. Intrinsic optimum temperature for development of insects and mites. *Environ Entomol.* 2015;34:1377–87.
22. Wu TH, Shiao SF, Okuyama T. Development of insects under fluctuating temperature: a review and case study. *J Appl Entomol.* 2015;139:592–9.
23. Verloren MC. On the comparative influence of periodicity and temperature upon the development of insects. *Ecol Entomol.* 2010;11:63–9.
24. Roe A, Higley LG. Development modeling of *Lucilia sericata* (Diptera: Calliphoridae). *Peer J.* 2015;3:1–14.
25. Karol G, Monika F. Effect of temperature treatment during development of *Osmia rufa* L. on mortality, emergence and longevity of adults. *J Apic Sci.* 2016;60:221–32.
26. Régnière J, Powell J, Bentz B, Nealis V. Effects of temperature on development, survival and reproduction of insects: experimental design, data analysis and modeling. *J Insect Physiol.* 2012;58:634–47.
27. Zang S, Cao J, Alimujiang K, Liu SH, Zhang YJ, Hu DF. Food patch particularity and foraging strategy of reintroduced Przewalski's horse in North Xinjiang, China. *Turk J Zool.* 2017;41:924–30.
28. Zhou R, Zhang K, Zhang TG, Zhou T, Chu HJ, Li K, et al. Identification of volatile components from oviposition and non-oviposition plants of *Gasterophilus pecorum* (Diptera: Gasterophilidae). *Scientific Reports.* 2020;10:15731.
29. Liu G, Aaron BA, Zimmermann W, Hu DF, Wang WT, Chu HJ, et al. Evaluating the reintroduction project of Przewalski's horse in China using genetic and pedigree data. *Biological Conservation.* 2014;171:288–98.
30. Heino J, Peckarsky BL. Integrating behavioral, population and large-scale approaches for understanding stream insect communities. *Curr Opin Insect Sci.* 2014;2:7–13.

- 354 31. Modlmeier AP, Keiser CN, Wright CM, Lichtenstein JLL, Pruitt JN. Integrating animal
355 personality into insect population and community ecology. *Curr Opin Insect Sci.* 2015;9:77–
356 85.
- 357 32. Otranto D, Milillo P, Capelli G, Colwell DD. Species composition of *Gasterophilus* spp.
358 (Diptera, Oestridae) causing equine gastric myiasis in southern Italy: parasite biodiversity and
359 risks for extinction. *Vet Parasitol.* 2005;133:111–8.
- 360 33. Wang WT, Xiao S, Huang HQ, Li K, Zhang D, Chu HJ, et al. Diversity and infection of
361 *Gasterophilus* spp. in Mongol-Xinjiang region and Qinghai Tibet region. *Sci Silva Sin.*
362 2016;52:134–9 (in Chinese).
- 363 34. Dorzh C, Minár J. Warble flies of the families Oestridae and Gasterophilidae (Diptera) found
364 in the Mongolian People's Republic. *Folia Parasit.* 1971;18:161–4.
- 365 35. Reshetnikov AD, Barashkova AI, Prokopyev ZS. Infestation of horses by the causative agents
366 of gasterophilosis (Diptera: Gasterophilidae): the species composition and the north-eastern
367 border of the area in the Republic (Sakha) of Yakutia of the Russian Federation. *Life Sci J.*
368 2014;11:587–90.
- 369 36. Gandon S, Ebert D, Olivieri I, Michalakis Y. Differential adaptation in spatially heterogeneous
370 environments and host-parasite coevolution. In: Mopper, S. and Strauss, S.Y. (eds), *Genetic
371 structure and local adaptation in natural insect populations.* London: Chapman and Hall; 1998.
372 p. 325–42.
- 373 37. Thomas F, Renaud F, Guégan JF. Parasitism and ecosystems. In:
374 *Parasitism and hostile environments.* New York: Oxford University Press; 2015. p. 85–112.
- 375 38. Machado TO, Braganca MAL, Carvalho ML, Andrade FJD. Species diversity of sandflies
376 (Diptera: Psychodidae) during different seasons and in different environments in the district of
377 Taquaruçú, state of Tocantins, Brazil. *Mem I Oswaldo Cruz.* 2012;107:955–9.
- 378 39. Aliakbarpour H, Che SMR, Dieng H. Species composition and population dynamics of thrips
379 (Thysanoptera) in mango orchards of Northern Peninsular Malaysia. *Environ Entomol.*
380 2010;39:1409–19.
- 381 40. Palomo LAT, Martinez NB, Napoles JR, Leon OS, Arroyo HS, Graziano JV, et al. Population
382 fluctuations of thrips (Thysanoptera) and their relationship to the phenology of vegetable crops
383 in the central region of Mexico. *Fla Entomol.* 2015;98:430–8.
- 384 41. Shibata E. Seasonal fluctuation and spatial pattern of the adult population of the Japanese pine
385 sawyer, *Monochamus alternatus* Hope (Coleoptera : Cerambycidae), in Young Pine Forests.
386 *Appl Entomol Zool.* 2008;16:306–9.
- 387 42. Haack RA, Lawrence RK, Heaton GC. Seasonal shoot-feeding by *Tomicus piniperda*
388 (Coleoptera: Scolytidae) in Michigan. *Great Lakes Entomologist.* 2018;33:1–8.
- 389 43. Lamb RJ, Mackay PA. Seasonal dynamics of a population of the aphid *Uroleucon rudbeckiae*
390 (Hemiptera: Aphididae): implications for population regulation. *Can Entomol.* 2016;149:300–
391 14.

- 392 44. Huxman TE, Smith MD, Fay PA, Knapp AK, Shaw MR, Loik ME, et al. Convergence across
393 biomes to a common rain-use efficiency. *Nature*. 2004;429:651–4.
- 394 45. Cleland EE, Collins ST, Dickson TL, Farrer EC, Gross KL, Gherardi LA, et al. Sensitivity of
395 grassland plant community composition to spatial vs. temporal variation in precipitation.
396 *Ecology*. 2013;94:1687–96.
- 397 46. Yong SP, Zhu ZY. A certain fundamental characteristics of gobi desert vegetation in the centre
398 Asia. *J Inner Mongolia University*. 1992;23:235–44(in Chinese).
- 399 47. Cui NR. The Flora Records of Main Forage Grass Crops in Xinjiang. In: Book one. Urumqi:
400 Xinjiang People’s Publishing House; 1990. p. 140–157 (in Chinese).
- 401 48. Ogle K, Reynolds JF. Plant responses to precipitation in desert ecosystems: integrating
402 functional types, pulses, thresholds, and delays. *Oecologia*. 2004;141:282–94.
- 403 49. Mckenna MF, Houle G. Why are annual plants rarely spring ephemerals? *New Phytol*.
404 2010;148:295–302.
- 405 50. Tielbörger K, Valleriani A. Can seeds predict their future? Germination strategies of density-
406 regulated desert annuals. *Oikos*. 2010;111:235–44.
- 407 51. Liu SH, Hu DF, Li K. Oviposition site selection by *Gasterophilus pecorum* (Diptera:
408 Gasterophilidae) in its habitat in Kalamaili Nature Reserve, Xinjiang, China. *Parasite*.
409 2015;22:34.
- 410 52. Epe C, Kings M, Stoye M, Böer M. The Prevalence and transmission to exotic equids (*Equus*
411 *quagga antiquorum*, *Equus przewalskii*, *Equus africanus*) of intestinal nematodes in
412 contaminated pasture in two wild animal parks. *J Zoo Wildlife Med*. 2001;32:209–16.
- 413 53. Hu XL, Liu G, Zhang TX, Yang S, Hu DF, Liu SQ, et al. Regional and seasonal effects on the
414 gastrointestinal parasitism of captive forest musk deer. *Acta Trop*. 2018;177:1–8
- 415 54. Teel PD, Marin SL, Grant WE. Simulation of host-parasite-landscape interactions: influence
416 of season and habitat on cattle fever tick (*Boophilus* sp.) population dynamics. *J Am Soc*
417 *Nephrol*. 1996;14:855–62.
- 418 55. James PJ, Moon RD, Brown DR. Seasonal dynamics and variation among sheep in densities
419 of the sheep biting louse, *Bovicola ovis*. *Int J Parasitol*. 1998;28:283–92.
- 420 56. Taylor B, Rahman PM, Murphy ST, Sudheendrakumar VV. Within-season dynamics of red
421 palm mite (*Raoiella indica*) and phytoseiid predators on two host palm species in south-west
422 India. *Exp Appl Acarol*. 2012;57:331–45.
- 423 57. Hercus MJ, Loeschcke V, Rattan SIS. Lifespan extension of *Drosophila melanogaster* through
424 hormesis by repeated mild heat stress. *Biogerontology*. 2003;4:149–56.
- 425

426 Figure legends

427 Figure 1 The population dynamics of *G. pecorum* in 2018 (mean/week).

428 Figure 2 Proportion of larvae of *G. pecorum* collected weekly (mean/week).

429 Figure 3 The accumulative collection of *G. pecorum* larvae in 2018.

430 Figure 4 Comparison of development in vitro of *G. pecorum* between two peak periods. (A) The

431 pupal days of the two peak periods; (B) The survival rate (SR), pupation rate (PR) and eclosion

432 rate (ER) of the two peak periods.

433 Figure 5 The temperature changes in KNR in 2018. (A) The average daily temperature, the highest

434 temperature, the lowest temperature, and the temperature difference in KNR; (B) The temperature

435 fitting curve.