

A Fair and EMG-validated Comparison of Recruitment Criteria, Musculotendon Models and Muscle Coordination Strategies, for the Inverse-dynamics Based Optimization of Muscle Forces During Gait

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1 **A fair and EMG-validated comparison of recruitment criteria, musculotendon models and**
2 **muscle coordination strategies, for the inverse-dynamics based optimization of muscle**
3 **forces during gait**

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10

11 **Abstract**

12 Experimental studies and EMG collections suggest that a specific strategy of muscle coordination
13 is chosen by the central nervous system to perform a given motor task. A popular mathematical
14 approach for solving the muscle recruitment problem is optimization. Optimization-based
15 methods minimize or maximize some criterion (objective function or cost function) which reflects
16 the mechanism used by the central nervous system to recruit muscles for the movement
17 considered. The proper cost function is not known a priori, so the adequacy of the chosen function
18 must be validated according to the obtained results. In addition of the many criteria proposed,
19 several physiological representations of the musculotendon actuator dynamics along with
20 different musculoskeletal models can be found in the literature, which hinders the selection of the
21 best neuromusculotendon model for each application. Seeking to provide a fair base for
22 comparison, this study measures the efficiency and accuracy of: i) four different criteria; ii) one
23 static and three physiological representations of the musculotendon actuator dynamics; iii) a
24 synergy-based method; all of them within the framework of inverse-dynamics based optimization.
25 Motion/force/EMG gait analyses were performed on ten healthy subjects. A musculoskeletal
26 model of the right leg actuated by 43 Hill-type muscles was scaled to each subject and used to
27 calculate joint moments, musculotendon kinematics and moment arms. Muscle activations were
28 then estimated using the different approaches, and these estimates were compared with EMG
29 measurements. Although similar results were obtained with all the methods, it must be pointed
30 out that a higher complexity of the method does not guarantee better results, as the best
31 correlations with experimental values were obtained with two simplified approaches.

32 **1. Introduction**

33

34 Determination of muscle forces during gait is of great interest to extract the principles of the
35 central nervous system (CNS) control, to facilitate assessment of pathological gait, or to estimate
36 the loads on bones and joints (prevention of injuries in sports, surgical planning to reconstruct
37 diseased joints) [1]–[3]. The invasive character of in vivo experimental measurements, and the

38 uncertain relation between muscle force and EMG, makes computer modeling and simulation a
39 useful substitutive approach. Determination of muscle forces by computer modeling and
40 simulation was extensively treated and numerous approaches can be found in the literature to
41 solve the redundancy problem of the muscle recruitment, as well as to represent the
42 musculotendon actuator dynamics [4]–[8]. Each author highlights the advantages of his own
43 approach. However, results do not depend only on the approach used, but also, on the
44 experimental data collection and on the musculoskeletal model used, which makes more difficult
45 for the readers to select objectively which approach to use for a certain application.

46 To objectively compare different approaches, it is necessary to test them under the same
47 conditions. When proposing a new approach, authors generally make a comparison with
48 experimental measurements in order to validate it [9], [10], and, in some cases, they compare their
49 results with those provided by a previous approach which gives confidence to readers [11], [12].
50 In some applications, a benchmark problem can be found that establishes some defined
51 conditions, so that researchers can get a fair comparison [13]. In the case of the resolution of the
52 muscle force-sharing problem, the authors only found a benchmark where the computational
53 speed and biological accuracy of three musculotendon models was compared during simple
54 muscle-driven simulations [14]. The physiological effect of static and dynamic optimization
55 during gait was compared by Pandy and Anderson [9] and De Groote et al. [15], but none of the
56 studies offered an experimental validation that allowed to conclude which method provided the
57 most realistic results.

58 Few years ago, a grand challenge competition to predict in vivo knee loads was organized by
59 some researchers who shared their experimental data collections for the analysis and its evaluation
60 [16]. However, the musculoskeletal modeling could differ between participants so that, by using
61 a different multibody model (with different degrees of freedom) and different muscle geometry
62 (which implies different arm moments), the results could not dissociate the effect of
63 neuromusculotendon models.

64 In this work, a comparison of the efficiency and accuracy of: i) four different criteria; ii) one static
65 and three physiological representations of the musculotendon actuator dynamics; iii) a synergy-
66 based method; all of them within the framework of inverse-dynamics based optimization, was
67 conducted. Motion/force/EMG gait analyses were performed on ten healthy subjects. A
68 musculoskeletal model of the right leg actuated by 43 Hill-type muscles was scaled to each subject
69 and used to calculate joint moments, musculotendon kinematics and moment arms. Therefore, the
70 muscle force-sharing problem was solved under the same conditions and using the same inputs.
71 Muscle activations were then estimated using the different approaches, and these estimates were
72 compared with EMG measurements which served as experimental reference.

73 **2. Methods**

74 **2.1 Experimental data collection**

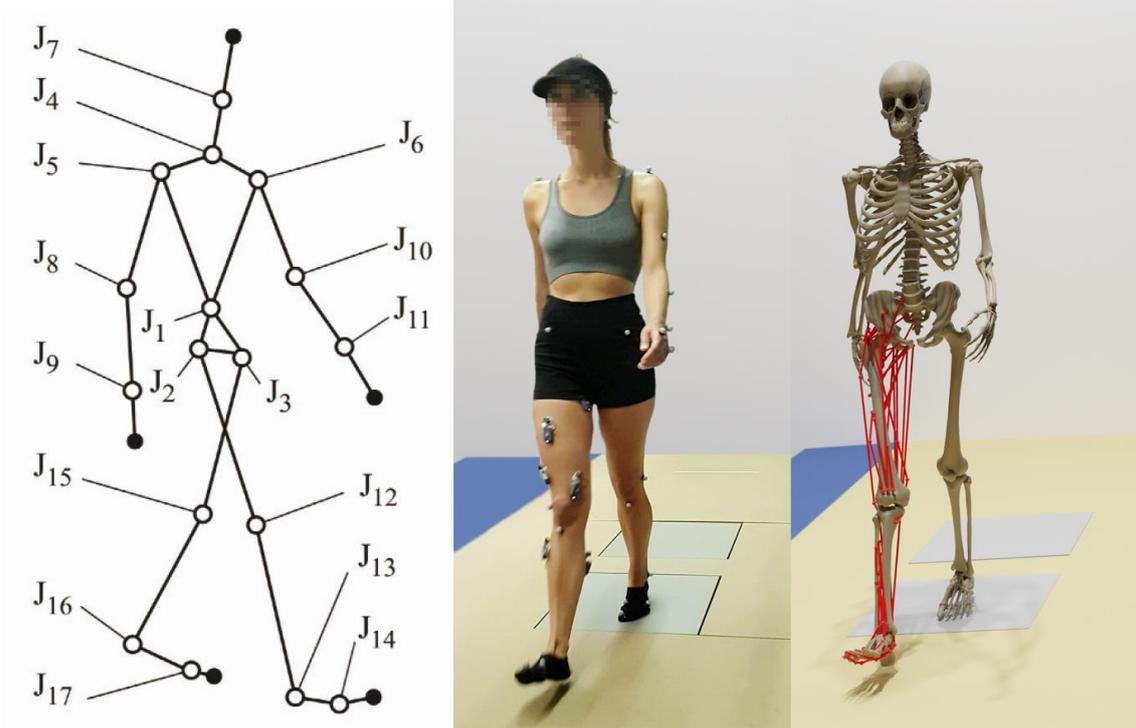
75 Ten subjects (seven males, three females, age 42 ± 16 years, height 173 ± 16 cm, body mass $73 \pm$
76 26 kg) were recruited for this study. All subjects gave written informed consent for their
77 participation. Subjects walked at their self-selected speed (1.1 ± 0.2 m/s) along a walkway with
78 two embedded force plates (AMTI, AccuGait sampling at 100 Hz). The motion was captured
79 using 12 optical infrared cameras (Natural Point, OptiTrack FLEX 3 also sampling at 100 Hz)
80 that computed the position of 37 optical markers (Figure 1). Additionally, 9 surface EMG signals
81 were recorded from the right leg at 1 kHz (BTS, FREEEMG). Each EMG signal was rectified,
82 filtered by singular spectrum analysis (SSA) with a window length of 250 [17] (equivalent to the
83 common forward and reverse low-pass 5th order Butterworth filter with a cut-off frequency of 6
84 Hz), and, then, normalized with respect to its maximal value, as recommended in [18]. This cut-
85 off frequency value is consistent with the ranges reported in previous studies using EMG data
86 [18], [19].

87 **2.2 Musculoskeletal model**

88 The human body was modeled as a three-dimensional multibody system formed by rigid bodies
89 (Figure 1, left and center). The model consisted of 18 anatomical segments [20]: two hindfeet,

90 two forefeet, two shanks, two thighs, a pelvis, a torso, a neck, a head, two arms, two forearms,
91 and two hands. The segments were linked by ideal spherical joints, thus defining a model with 57
92 degrees of freedom (DOFs). The axes of the global reference frame were defined as follows: x -
93 axis in the anterior–posterior direction, y -axis in the medial–lateral direction, and z -axis in the
94 vertical direction. The computational model was defined with 228 mixed (natural + angular)
95 coordinates. The subset of natural coordinates comprised the three Cartesian coordinates of 22
96 points and the three Cartesian components of 36 unit vectors, thus yielding a total of 174 variables.

97 Matrix-R formulation [21] was used to perform an inverse-dynamics analysis to obtain the joint
98 torques along the motion by means of the in-house developed MBSLIM library [22] programmed
99 in FORTRAN, as described in [23]. Once the joint torques were computed, it was assumed that
100 43 right leg muscles contributed to the following six right-leg inverse-dynamics moments: the
101 three rotational DOFs at the hip, the flexion/extension DOF at the knee, and the plantar/dorsi
102 flexion and inversion/eversion at the ankle. Muscles were modeled as one or more straight-line
103 segments with via points. These points corresponded to the attachments of muscle and tendon to
104 bone and were defined as the origin (i.e., proximal attachment) and insertion (i.e., distal
105 attachment). Muscle properties and local coordinates for these points were obtained from
106 OpenSim (model Gait2392) [24] and scaled to each subject from the generic reference OpenSim
107 model, as commented further in 2.4.



108

109

Figure 1: Gait of healthy subject: multibody model (left); acquired motion (middle);

110

computational model (right).

111

2.3 Optimization problem

112

As introduced before, the fundamental problem is that there are more muscles serving each degree

113

of freedom of the system than those strictly necessary from the mechanical point of view. In this

114

case, there are 43 muscles at the leg to actuate 6 degrees of freedom (other degrees of freedom of

115

the leg are controlled by joint structures as bones and ligaments, yielding a reaction moment

116

instead a drive torque). Consequently, there is an infinite number of solutions for this problem

117

and, in order to reproduce the specific strategy of muscle coordination adopted by the CNS,

118

optimization is used.

119

The inverse-dynamics based optimization problem that serves to determine the muscle forces at

120

each time-point can be formulated in general form as:

121

$$\begin{aligned}
 & \min C \\
 & \text{subject to } \mathbf{J}^T \mathbf{F}^{MT} = \mathbf{Q}^{ID} \\
 & F_i^{Min} \leq F_i^{MT} \leq F_i^{Max} \quad i = 1, 2, \dots, m
 \end{aligned} \tag{1}$$

122 where C is the cost function, \mathbf{Q}^{ID} is the vector of inverse-dynamics joint moments at the right leg
 123 (where the force-sharing problem is addressed), \mathbf{F}^{MT} is the vector of muscle forces, \mathbf{J} is the
 124 Jacobian whose transpose projects the muscle forces into the joint drive torques space, F_i^{Min} and
 125 F_i^{Max} are the instantaneous minimum and maximum allowed forces in muscle i , respectively, and
 126 m is the number of muscles. Expression of the objective function C depends on the muscle
 127 recruitment criterion used. In the literature, several muscle recruitment criteria have been
 128 suggested to represent the CNS behavior. In this work, four of them have been considered in the
 129 context of static optimization.

130 **2.3.1 Static optimization (SO)**

131 **Nonlinear polynomial criteria**

132 The polynomial criterion can be written as

$$133 \quad \min \sum_{i=1}^m \left(\frac{F_i^{MT}}{k_i} \right)^w \quad (2)$$

134 where k_i denotes a positive weighting factor and w is the power of the polynomial. According to
 135 [25], the muscle force prediction obtained by minimizing the sum of muscle stresses raised to a
 136 power w whose value ranges between 1.4 and 5.1 is physiologically analogous to minimizing
 137 muscle fatigue. As Anderson and Pandy did in their study [9], a power of 2 was chosen.

138 **Criterion I - minimization of the sum of the squares of muscle forces**

$$139 \quad \min \sum_{i=1}^m \left(F_i^{MT} \right)^2 ; \quad (3)$$

140 **Criterion II - minimization of the sum of the squares of relative muscle forces**

$$141 \quad \min \sum_{i=1}^m \left(\frac{F_i^{MT}}{F_{0,i}^M} \right)^2 ; \quad (4)$$

142 with $F_{0,i}^M$ the maximum isometric force from [24].

143 **Criterion III - minimization of the sum of the squares of muscle stresses**

144
$$\min \sum_{i=1}^m \left(\frac{F_i^{MT}}{PCSA_i} \right)^2 ; \quad (5)$$

145 with $PCSA$ the physiological cross sectional area from [26].

146 **Min/max criterion**

147 The min/max criterion distributes the collaborative muscle forces in such a way that the maximum
148 relative muscle force is as small as possible. Therefore, the largest endurance for a task is attained
149 when the maximum relative muscle force [27] or the maximum muscle stress [28] is as small as
150 possible. The min/max criterion takes the form:

151
$$\min \left(\max \left(\frac{F_i^{MT}}{k_i} \right) \right), i = 1, \dots, m ; \quad (6)$$

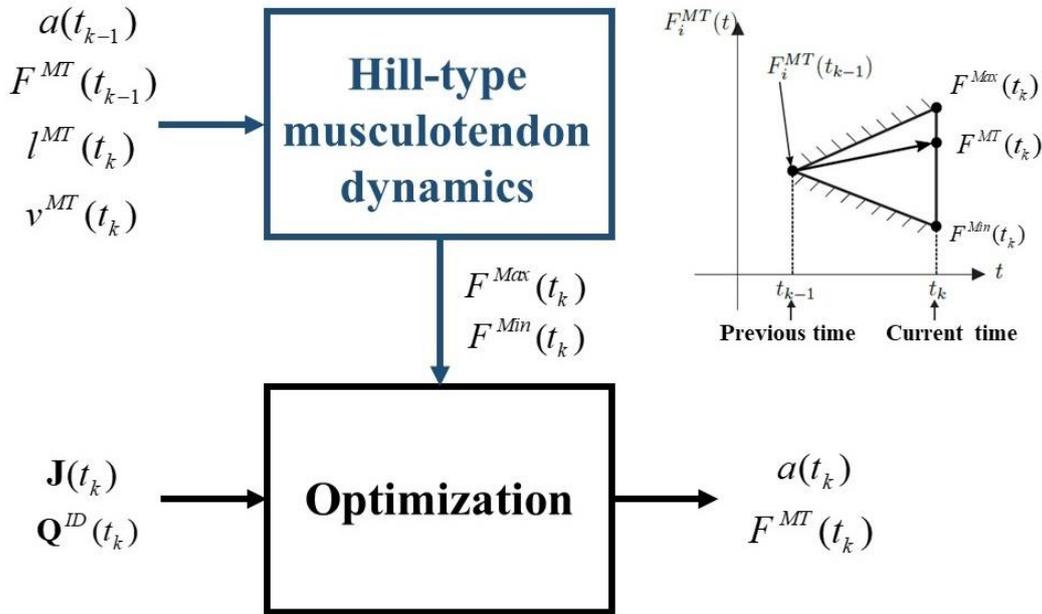
152 For this study, the following criterion is used:

153 **Criterion IV - minimization of the largest relative muscle force**

154
$$\min \left(\max \left(\frac{F_i^{MT}}{F_{0,i}^M} \right) \right), i = 1, \dots, m ; \quad (7)$$

155 For SO, the physiological behavior of the musculotendon actuator dynamics is not considered,
156 so, the limit values of the muscular forces are $F_i^{Min} = 0$ and $F_i^{Max} = F_{0,i}^M$.

157



159

160 Figure 2: Physiological inverse-dynamics approach.

161

162 At physiological level, musculotendon actuator dynamics introduces muscle force constraints.

163 Whereas the static optimization approach disregards these constraints in order to simplify the

164 problem, the so-called physiological approach [29] takes them into consideration. This approach

165 applies optimization techniques at each time-point, and prescribes minimal and maximal

166 constraints for the forces by extrapolating the force values from the previous time-point through

167 feasible muscle dynamics (Figure 2).

168

169 The dynamics of musculotendon actuators can be divided into two parts. First, the activation

170 dynamics which corresponds to the transformation of a neural excitation sent by the brain into an

171 activation of the contractile apparatus. Activation dynamics is described by a first-order ordinary

172 differential equation that contains the relationship among the muscle activation a , its derivative

173 \dot{a} , and the neural excitation u as:

174
$$\dot{a}(t) = \frac{u(t) - a(t)}{\tau}, \quad (8)$$

175 with $\tau = \tau_{act}$ when $a(t_{k-1}) \leq u(t_k)$ and $\tau = \tau_{deact}$ when $a(t_{k-1}) > u(t_k)$. The activation and
 176 deactivation time constants τ_{act} and τ_{deact} are set to 15 ms and 50 ms, respectively [30], [31].

177 Second, this activation is transformed into a muscle force by the second phase, the contraction
 178 dynamics. The force generated by a muscle is constrained by its force-length-velocity properties,
 179 related to the Hill-type musculotendon model used, which is defined by this second differential
 180 equation:

181
$$F^{MT}(t) = f(a(t), F^{MT}(t), l^{MT}(t), v^{MT}(t)). \quad (9)$$

182 The musculotendon length l^{MT} and velocity v^{MT} depend on the position and velocity of the body
 183 segments and, in turn, the generated tendon force F^{MT} affects the motion of the body segments.
 184 Thus, there exists interaction between muscles and body segments.

185 The complete musculotendon dynamics can be expressed as a system of two differential equations
 186 which can be written, in a simplified form, as

187
$$\dot{x}(t) = \begin{bmatrix} \dot{a}(t) \\ F^{MT}(t) \end{bmatrix} = h(x(t), u(t), l^{MT}(t), v^{MT}(t)). \quad (10)$$

188 This system is used to define the minimal and maximal constraints for the forces by extrapolating
 189 the force values from the previous time-point using feasible muscle dynamics, integrating (10)
 190 with $u(t) = 0$ to estimate $F^{Min}(t)$ and with $u(t) = 1$ to estimate $F^{Max}(t)$ (the muscular excitation
 191 is assumed to be constant between the two time frames). Using the physiological approach, the
 192 initial activations and muscles forces are needed. The determination of initial activations and
 193 muscular forces is based on the static condition which states that the initial fiber velocity of each
 194 muscle is set to zero and, F^{Min} and F^{Max} correspond to $a = 0$ and $a = 1$, respectively.

195 Integration is carried out with Matlab *Ode23t*. Two integrations per muscle are required at each
 196 time step, which make the optimization and integration process heavy and slow. By programming

197 the muscular functions of the Hill-type musculotendon model into a FORTRAN mex file, the
 198 computational time is reduced by a factor of 10. However, the computational time is still long
 199 and, in addition, the high tendon stiffness makes really difficult to use this approach (a suitable
 200 scaling of muscle parameters is needed) [32]. Therefore, in order to simplify the problem while
 201 keeping some physiological characteristics, most authors prefer to use a Hill-type musculotendon
 202 model with a rigid tendon [12], [15], [33].

203 **Physiological approach with rigid tendon (PHY1)**

204 In this way, the tendon length is constant and, consequently, the muscle fiber length and velocity
 205 depend only on the musculoskeletal geometry as well as on body segment configurations (which
 206 affect l^{MT} and v^{MT}) and not the musculotendon force. Consequently, the force-length-velocity
 207 allowed is expressed as:

$$208 \quad F^{MT}(t) = a(t)g(l^{MT}(t), v^{MT}(t)). \quad (11)$$

209 Use of the rigid tendon model avoids the two integrations needed to calculate the limits of the
 210 muscle force at each instant. Then, in order to further reduce the computational burden, the first-
 211 order ordinary differential equation (8) used to estimate the muscular activation, a , can be
 212 simplified as follows:

213 **Time response considered (PHY2)**

214 In order to keep the muscular time response relation given by (8), the first-order ordinary
 215 differential equation can be converted into:

$$216 \quad a(t_k) = u(t_k) + (a(t_{k-1}) - u(t_k))e^{(-\Delta t/\tau)}, \quad (13)$$

217 with $\tau = \tau_{act}$ when $a(t_{k-1}) \leq u(t_k)$, $\tau = \tau_{deact}$ when $a(t_{k-1}) > u(t_k)$ and Δt is the time step.

218 Therefore, the minimal and maximal muscular force constraints of the optimization problem can
 219 be obtained through integration with any of the described approaches by extrapolating the force
 220 values from the previous time-point using feasible muscle dynamics.

221 **Time response ignored (PHY3)**

222 However, authors who consider the tendon as a rigid element usually choose to ignore the
223 muscular time response and assume that:

$$224 \quad a(t) = u(t) . \quad (12)$$

225 In this work, the minimization of the sum of the squares of muscle forces (Criterion I) was used
226 as objective function for the three physiological models.

227 **2.3.3 Synergy optimization (SynO)**

228 The fact that synergies take a high dimensional control space and reduce it to a low dimensional
229 space is potentially useful for reducing the amount of indeterminacy when estimating muscle
230 forces via optimization. For this reason, some authors started to investigate how to include it to
231 solve the muscle force-sharing problem.

232 The synergy optimization (SynO) approach used in [12] estimates muscle forces during human
233 walking using synergy-constructed muscle activations, similar to the more complex approach
234 proposed in [34]. SynO finds muscle forces that match the inverse-dynamics joint moments as
235 closely as possible through the moment tracking error term in the cost function. In SynO,
236 synergies couple muscle activations across time frames, requiring the optimization to be
237 performed over all the time frames simultaneously as follows:

$$238 \quad a_{fxm}^* = T_{fxn_s}(T_p)V_{n_s \times m} \quad (14)$$

239 where $T_{fxn_s}(T_p)$ and $V_{n_s \times m}$ are the time-varying synergy activations defined by B-spline nodes,
240 and the corresponding time-invariant synergy vectors, respectively. Each muscle activation
241 synergy is composed of a single time-varying synergy activation defined by $p = (f-1)/5 + 1$ (nearest
242 integer, f = number of frames) B-spline nodal points along with its corresponding time-invariant
243 synergy vector defined by $m = 43$ weights specifying inter-muscle activation coupling. Thus, for
244 n_s synergies ($n_s = 3$ in this study), the number of design variables is $n_s^*(p+m)$. Muscle synergy
245 quantities are used as the design variables for synergy optimization. Each optimization problem

246 is theoretically over-determined. However, in practice, the problems remain under-determined
 247 since neighboring time frames are not completely independent from one another.

248 Using these design variables, the SynO cost function is formulated as follows:

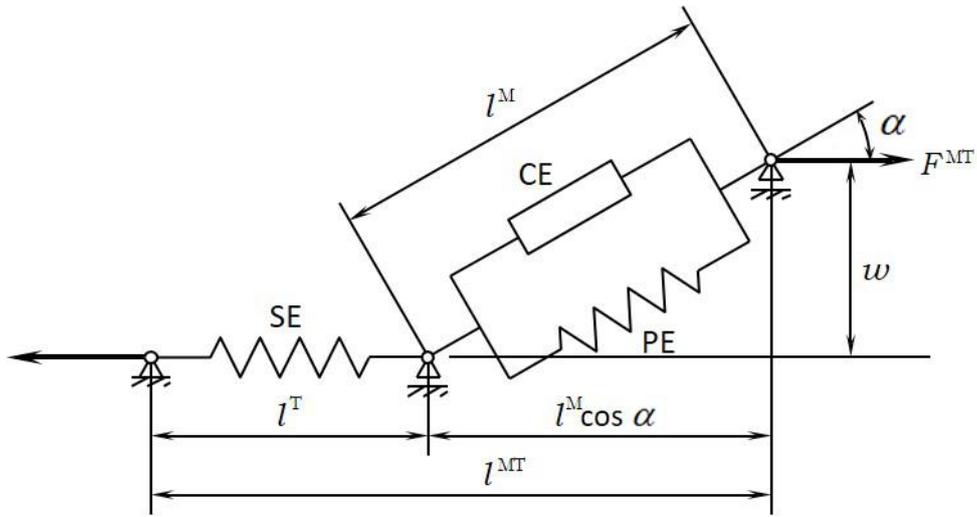
$$249 \quad C_{C_p, V}^{SynO} = \sum_{j=1}^n \left(\beta \sum_{k=1}^6 \left[\frac{Q_{jk}^{MT} - Q_{jk}^{ID}}{\max(|Q_k^{ID}|)} \right]^2 + \sum_{i=1}^m (a_{ij}^{*2} + \lambda_{ij, pen} (a_{ij}^* - 1)^2) \right) \quad (15)$$

250 Where a_{ij}^* are the synergy-based muscle activations, and $\lambda_{ij, pen} = \begin{cases} 0 & 0 \leq a_{ij}^* \leq 1 \\ 10^5 & \text{otherwise} \end{cases}$ are

251 penalization factors for muscle i at the time frame j to ensure that muscle activations stay between
 252 0 and 1. While previous approaches enforce the muscle forces to exactly reproduce the inverse-
 253 dynamics joint moments through its equality constraints, the SynO approach minimizes the error
 254 between Q^{ID} and Q^{MT} , being Q^{MT} the joint moments produced by the muscle forces estimated
 255 by SynO. A scale factor $\beta = 100$ is applied to achieve the best compromise between joint
 256 moment tracking and activation minimization [35]. Using three synergies, the mean joint
 257 intersegmental moment matching between Q^{ID} and Q^{MT} across subjects was higher than 96%.

258 The objective function is programmed as a FORTRAN mex file to reduce computation time (16
 259 times faster than the original Matlab function). Linear equality constraints enforce that the sum
 260 of weights within each synergy vector is equal to 1, which makes the synergy construction unique,
 261 while lower bound constraints enforce the synergy activation B-spline nodes and synergy vector
 262 weights to be non-negative. The same musculotendon model used for the PHY3 approach is used
 263 here.

264



266

267 Figure 3: Hill-type muscle model. The muscle fibers are modeled as an active contractile element
 268 (CE) in parallel with a passive elastic component (PE). These elements are in series with a
 269 nonlinear elastic tendon (SE). The pennation angle denotes the angle between the muscle fibers
 270 and the tendon. Superscripts MT, M, and T indicate musculotendon, muscle fiber, and tendon,
 271 respectively.

272 Due to the sensitivity of physiological approaches [32], a suitable scaling of musculotendon
 273 parameters is needed. In addition to the high tendon stiffness which makes implementation really
 274 difficult, some Hill-muscle equations become numerically stiff when numerical singularities are
 275 approached [14]. Since these conditions are often encountered during a simulation, to prevent that
 276 the solver gets stuck at points that were numerically feasible yet not physiologically sound [36]
 277 (which slows the process of numerical integration), a scaling correction was applied. Length
 278 parameters were scaled in two steps. First, for each muscle, the tendon slack length (l_s^T) and the
 279 optimal muscle fiber length (l_0^M) were scaled with a scale factor calculated as the relation between
 280 the subject's musculotendon length in standing position and that of the generic model in the same
 281 position. As the pennation angle of the reference, α_0 , is kept, the scaled distance between the
 282 aponeuroses of muscle origin and insertion, w (which remains constant during the muscle
 283 contraction), is given by:

284
$$w = l^M \sin(\alpha) = l_0^M \sin(\alpha_0). \quad (16)$$

285 Then, because tendons are so stiff that their lengths do not change significantly during movement,
 286 the approximated muscle fiber length, l^{M*} , can be calculated for each muscle during the complete
 287 gait cycle as follows:

288
$$l^{M*}(t) = (l^{MT}(t) - l_S^T) / \cos(\alpha(t)) \quad (17)$$

289 with $\alpha(t) = \arctan\left(\frac{w}{l^{MT}(t) - l_S^T}\right)$. Finally, in order to keep the normalized muscle lengths

290 ($\bar{l}^M = \frac{l^M}{l_0^M}$) within the physiological optimal conditions ($0.5 < \bar{l}^M \leq 1.2$) [37], the final scaled l_0^M

291 was set to the maximum approximated muscle fiber length along the motion.

292 **2.5 Optimization protocol and EMG comparison**

293 Optimization seeks to find the best solution from all the feasible ones by minimizing the objective
 294 function. Finding the global minima of a function is really difficult because of the many local
 295 minima. In order to get the best possible results, the following protocol is used in this work.
 296 Although each optimization problem is solved using the Matlab's *fmincon* nonlinear constrained
 297 optimization algorithm, five global optimizations are run using Matlab's *ga* genetic optimization
 298 algorithm with a population size of 50 to provide random initial guesses for *fmincon*. The solution
 299 with the lowest objective function value is chosen as initial guess for the initial time point.
 300 Thereafter, as muscle activation is normally smooth and continuous during gait, the optimal
 301 solution from the previous time frame is used as the initial guess for the current time frame [38].

302 Matching between estimated muscle activations and EMG was quantified via cross-correlation
 303 using the Pearson correlation coefficient r (Matlab's function *corrcoef*) with a maximum time
 304 delay of 150 ms [39]. The correlation coefficient r was chosen to compare muscle activations and
 305 EMG data so as to focus on shape rather than on magnitude discrepancies, as there is no direct
 306 relationship between EMG and muscle force amplitude [40], [41].

307 **3. Results**

308 The different approaches presented in this study were compared with EMG measurements for the
309 ten healthy subjects. Normalized muscle activations during a gait cycle of one healthy subject
310 estimated by SO using all the criteria are plotted in Figure 3 along with the corresponding
311 normalized EMG measurements. Comparison of muscle activations estimated with the different
312 criteria are significantly different but show some similarities too.

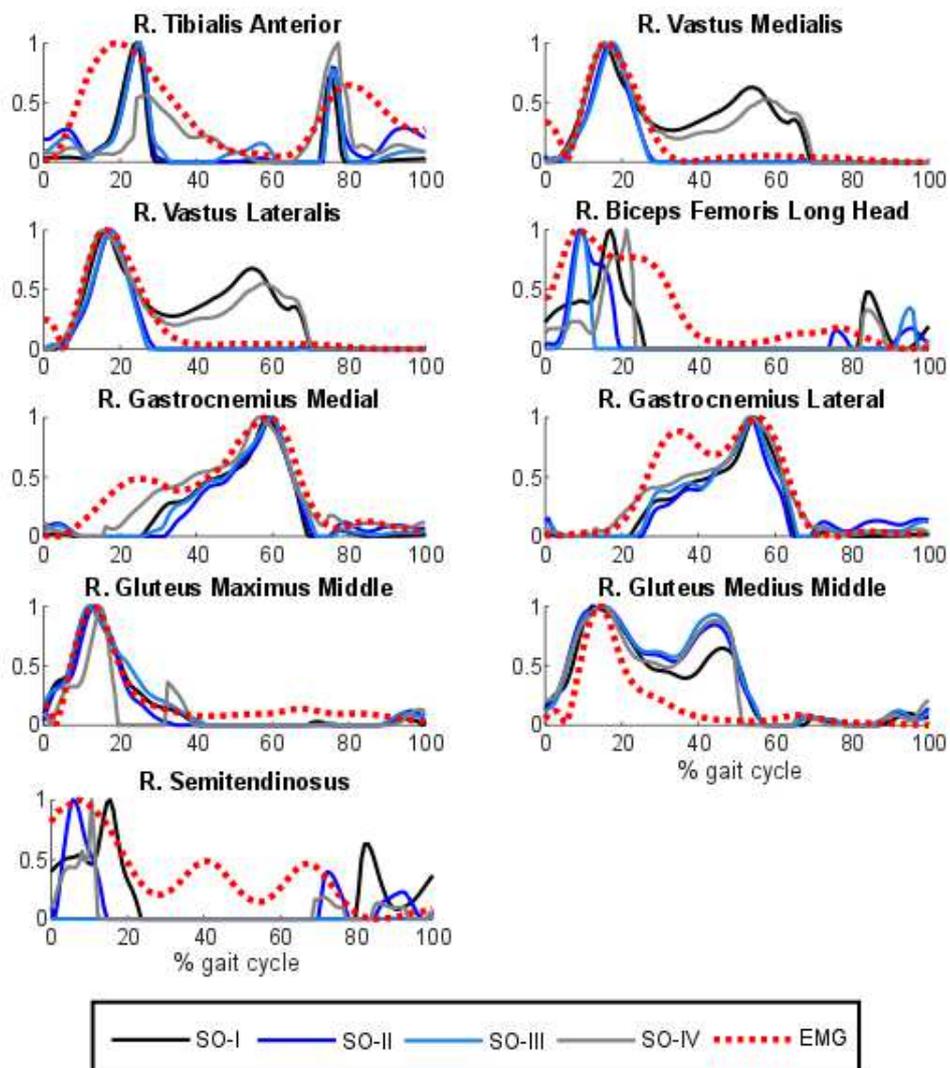
313 Then, to observe the physiological effect of the musculotendon model, four approaches using
314 criterion I were compared: normalized muscle activations during a gait cycle of one healthy
315 subject estimated by static optimization (SO-I) and the three physiological approaches (PHY1,
316 PHY2 and PHY3) are represented in Figure 4. While PHY3 presented distinct results, muscle
317 activations estimated by SO-I, PHY1 and PHY2 were very similar. PHY1 and PHY2 showed
318 almost the same results.

319 Furthermore, the normalized muscle activations during a gait cycle of one healthy subject
320 estimated by PHY3 and synergy optimization with 3 synergies (SynO3) are compared in Figure
321 5 to highlight the effect of the synergy structure. Both used the same musculotendon model
322 (physiological approach with rigid tendon and activation time response ignored). However, results
323 are significantly different.

324 Mean across subjects Pearson correlation coefficient r values between EMG vs. muscle
325 activations of all the approaches of this study are reported in Table 1. Correlations of the many
326 approaches did not present such differences. Mean values of the different approaches are close,
327 between 0.61 (SO-IV and SynO3) and 0.74 (SO-I and PHY2). Approaches SO-I, PHY1 and PHY2
328 show almost the same correlations (means of 0.73 and 0.74).

329 Besides, the computational efficiency of the different approaches studied in this work is compared
330 in Table 1. All calculations were performed on an Intel® Core™ i7-6700K processor running at
331 4.00 GHz with 16 GB of RAM. With a mean computational time of 2.5 s to simulate a gait cycle,
332 SO-I is the fastest approach, while PHY1 is the slowest one (225.2 s).

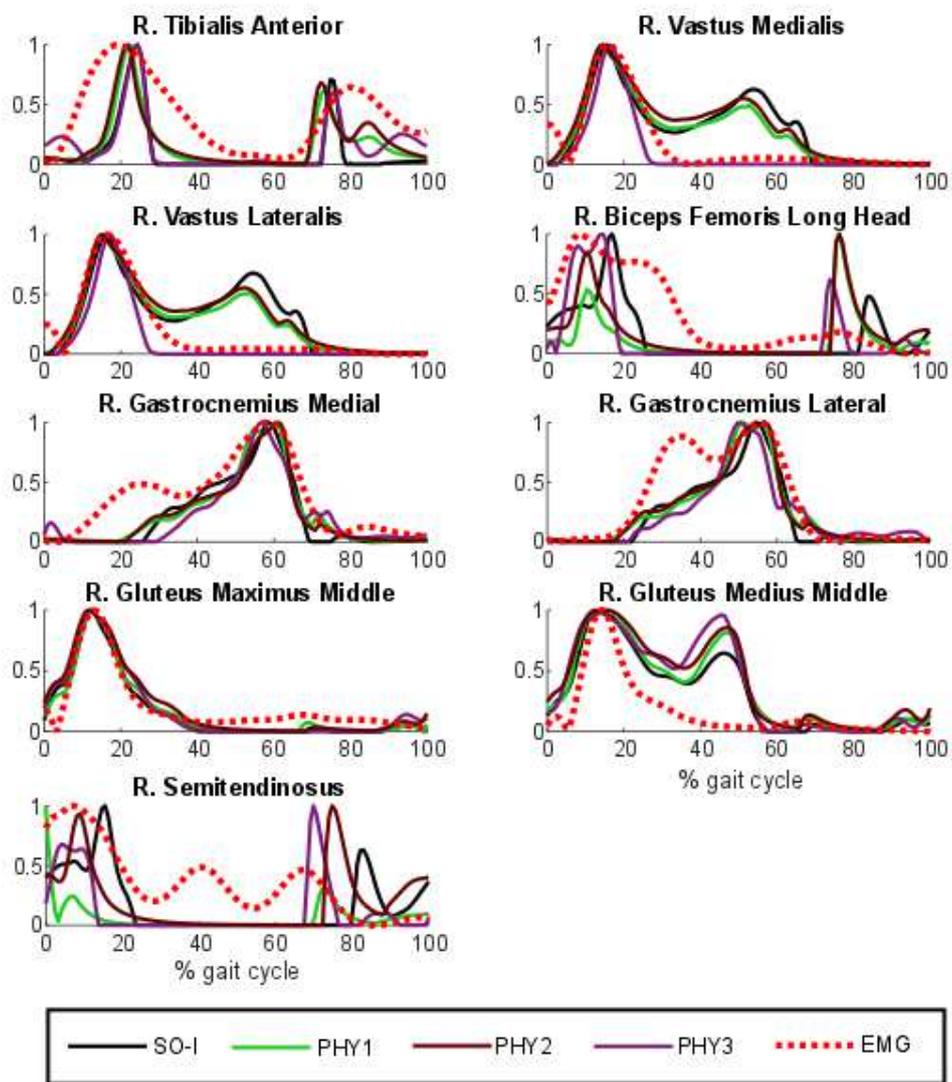
333 Finally, the resulting joint reaction forces at hip, knee and ankle for the different muscle
 334 recruitment approaches are compared (Figure 6 and Table 2). Similar joint reaction forces are
 335 obtained with the non-synergy-based approaches: SO-I, PHY1 and PHY2 offer almost the same
 336 results, while SO-II and PHY3 show joint reactions slightly higher than their counterparts at hip
 337 and knee levels. However, the joint reaction forces at hip, knee and ankle calculated from the
 338 muscle forces estimated with SynO3 are much higher than those obtained with the other
 339 approaches.



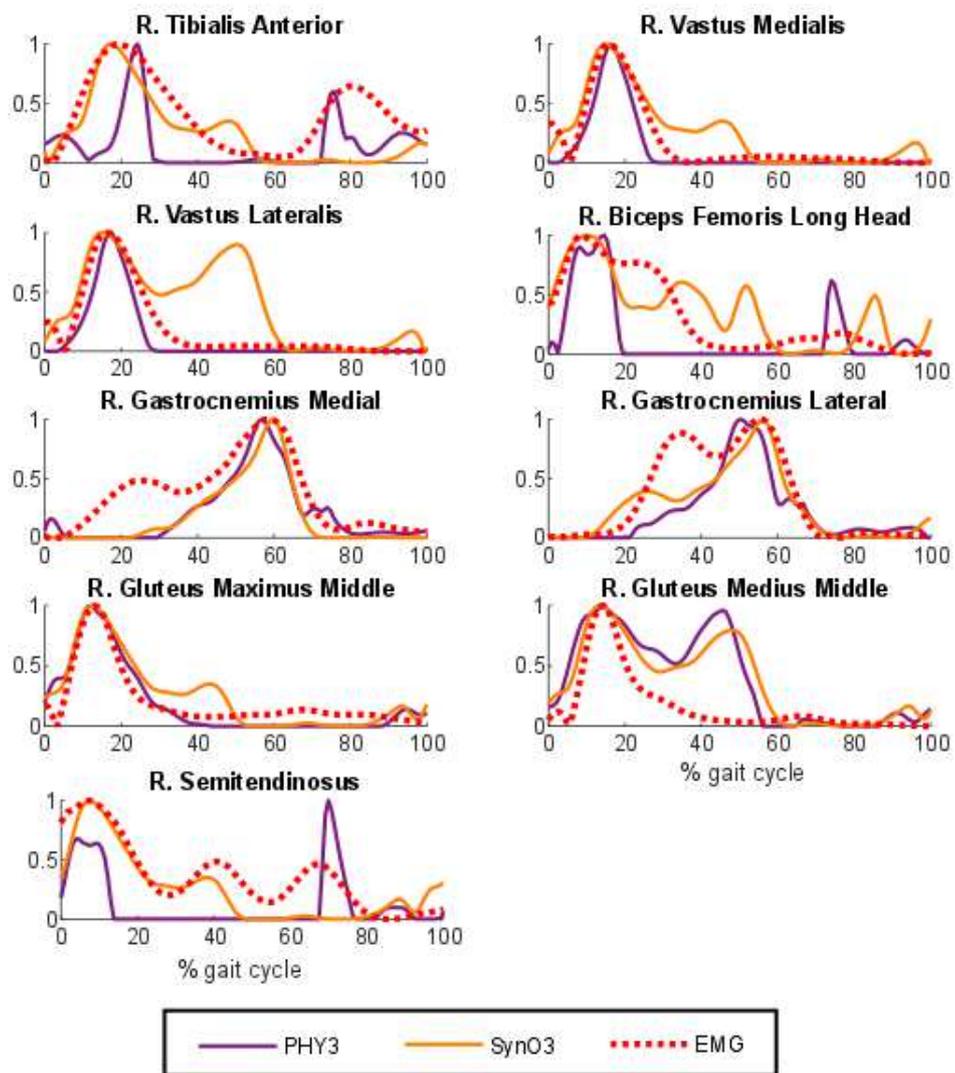
340

341 Figure 4: Normalized muscular activations obtained with static optimization (criteria I-IV) vs.
 342 normalized EMG for a healthy subject.

343



344
 345 Figure 5: Normalized muscular activations obtained through static optimization with criterion I
 346 (SO-I) and physiological optimization with criterion I (original approach and two simplified
 347 alternatives) vs. normalized EMG for a healthy subject. PHY1: physiological approach; PHY2:
 348 physiological approach with rigid tendon and activation time response considered; PHY3:
 349 physiological approach with rigid tendon and activation time response ignored.



350

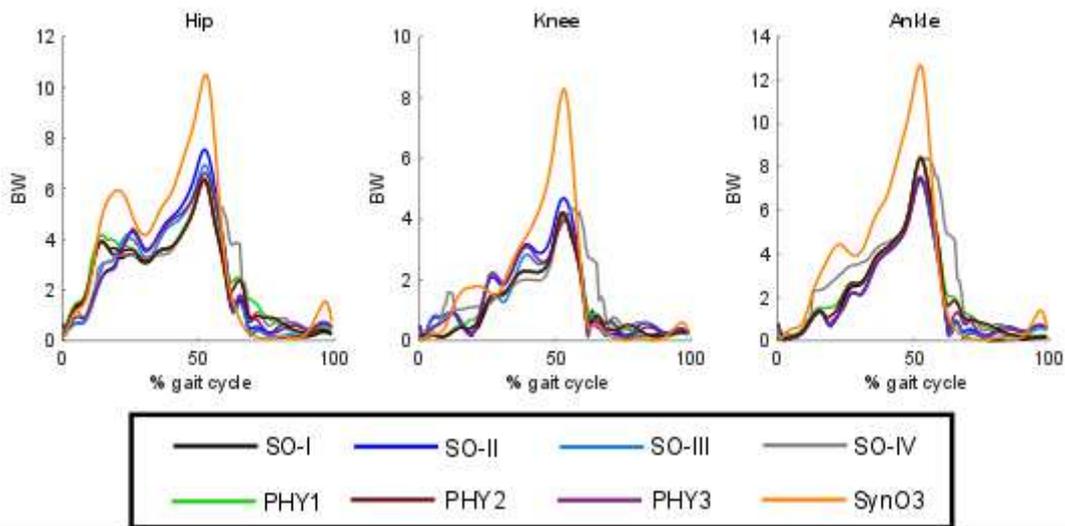
351 Figure 6: Normalized muscular activations obtained from physiological optimization with rigid
 352 tendon and activation time response ignored (PHY2) and synergy optimization with 3 synergies
 353 (SynO3) vs. normalized EMG for a healthy subject.

354

355 Table 1: Mean across subjects Pearson correlation coefficient r values between EMG vs. muscle
 356 activations ($r < 0.40$ in red and $r > 0.60$ in green) and computational time of the different
 357 approaches.

	MEAN VALUES							
	Pearson correlation coefficient r between across-subject mean EMG vs. muscle activations							
	SO-I	SO-II	SO-III	SO-IV	PHY1	PHY2	PHY3	Syn03
R. Tibialis Anterior	0.61	0.65	0.52	0.23	0.62	0.65	0.67	0.37
R. Vastus Medialis	0.68	0.67	0.67	0.55	0.79	0.79	0.75	0.73
R. Vastus Lateralis	0.68	0.70	0.70	0.63	0.79	0.79	0.82	0.66
R. Gastrocnemius Medial	0.86	0.76	0.84	0.80	0.79	0.80	0.76	0.85
R. Gastrocnemius Lateral	0.75	0.67	0.73	0.65	0.69	0.70	0.67	0.74
R. Semitendinosus	0.68	0.65	0.49	0.59	0.61	0.64	0.58	0.56
R. Biceps Femoris Long Head	0.78	0.77	0.59	0.56	0.70	0.74	0.65	0.37
R. Gluteus Maximus Middle	0.89	0.82	0.87	0.81	0.89	0.88	0.86	0.66
R. Gluteus Medius Middle	0.71	0.63	0.61	0.62	0.69	0.69	0.56	0.58
Mean	0.74	0.71	0.67	0.61	0.73	0.74	0.70	0.61
Mean computational time	2.5	27.6	16.1	19.2	225.2	3.6	3.2	48.7

358



359

360 Figure 7: Joint reaction forces at hip, knee and ankle obtained with different muscle recruitment
 361 approaches.

362

363 Table 2: Mean across subjects of the maximum joint reaction forces at hip, knee and ankle for
 364 different muscle recruitment approaches.

	SO-I	SO-II	SO-III	SO-IV	PHY1	PHY2	PHY3	Syn03
Hip	7.1	9.0	6.6	8.9	7.4	7.3	9.2	11.9
Knee	4.4	5.6	3.7	5.5	4.5	4.5	5.7	7.1
Ankle	8.4	8.1	8.0	8.8	8.5	8.5	8.1	11.5

365

366 4. Discussion

367 This work offers a comparison of the efficiency and accuracy of: i) four different criteria; ii) three
 368 different physiological representations of the musculotendon actuator dynamics; iii) a synergy-
 369 based method; all of them in the framework of inverse-dynamics based optimization. All the
 370 approaches were used under the same conditions by taking the same inputs from
 371 motion/force/EMG gait analyses performed on ten healthy subjects. Results obtained with the
 372 different methods do not present large discrepancies. Higher complexity of the method does not
 373 guarantee better results, as the best correlations with experimental values were obtained with two
 374 simplified approaches.

375 First, muscles activations obtained from SO and four different criteria exhibit visually different
 376 shapes along with some similarities. In addition, mean across subjects Pearson correlation
 377 coefficient r values between EMG vs. muscle activations of all the criteria do not present
 378 significant discrepancies. The best correlations were obtained with the simplest and fastest
 379 criterion (SO-I), which yielded a correlation of 74%, while the worst correlations were obtained
 380 with the most involved criterion (SO-IV).

381 Second, it was observed that the physiological representation of the musculotendon actuator
 382 dynamics does not affect the estimation of muscle forces during gait. Muscle activation shapes,
 383 experimental correlations and joint reaction forces are almost the same as those obtained through
 384 the non-physiological method (SO). The same conclusion was drawn by De Groote et al. [15],
 385 Anderson and Pandy [6] and Millard et al. [14]. However, for faster, higher-powered tasks, like
 386 running or jumping, a compliant tendon model could be preferable. Moreover, despite its

387 disadvantages (harder to implement and higher computational time), the physiological approach
388 served to implement some Hill-based energy expenditure methods [42], [43], since it provides the
389 muscular variables required as inputs.

390 Third, as previously observed in [35], the synergy structure imposed within the SynO approach
391 did not improve prediction of muscle activations during gait. The muscle synergy hypothesis has
392 been notoriously difficult to prove or falsify [44], and results of this study do not allow to draw a
393 conclusion in this regard. It can only be said that the SynO approach offers reasonable prediction
394 of muscle activations and that its reduced dimensional control space could be beneficial for
395 applications such as epidural electrical stimulation [45] or motion control and prediction [33].

396 Finally, all the estimated joint reaction forces at the hip were higher than the direct experimental
397 measurements reported in the literature [46]–[48]. Brand et al. reported that hip contact-force
398 predictions in the literature are higher than force measurements because of modeling assumptions
399 [48]. In this work, and in the literature [48], it has been shown that, paradoxically, physiological
400 representation of the musculotendon actuator dynamics increases rather than reduces the
401 discrepancies between force predictions and measurements, due to its constraints. Same
402 conclusion can be drawn for the SynO approach, which disproportionately increases the joint
403 forces due to its imposed synergy structure and reduced dimensional control space. Shourijeh and
404 Fregly observed that the joint stiffness results were visibly different between the SynO and SO
405 solutions, and that the stiffness decreased as the number of synergies was increased [12].

406 **Conclusion**

407 In conclusion, this study evaluated several approaches to predict muscle activations during gait
408 by comparing them with EMG measurements obtained experimentally, and found that higher
409 complexity of the method does not guarantee better results. No significant differences among
410 predicted EMG patterns were found. However, the simplified physiological approach with rigid
411 tendon and activation time considered, presented the best accuracy and a very competitive
412 computational time.

413 **Availability of supporting data**

414 The datasets generated for this study are available on request to the corresponding author.

415 **Ethical Approval and Consent to participate**

416 The studies involving human participants were reviewed and approved by the Committee of
417 Ethics of the University of A Coruña. The participants provided their written informed consent
418 to participate in this study.

419 **Consent for publication**

420 The participants provided their written informed consent for publication.

421 **Authors' contribution**

422 F.M. designed the experiments with the supervision of U.L. and performed the experiments with
423 the help of M.L. F.M. derived the models and analyzed the data. F.M. and J.C. wrote the
424 manuscript in consultation with U.L. and M.L.

425 **Conflict of interest**

426 No conflicts of interest lie with any of the authors.

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489

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Figures

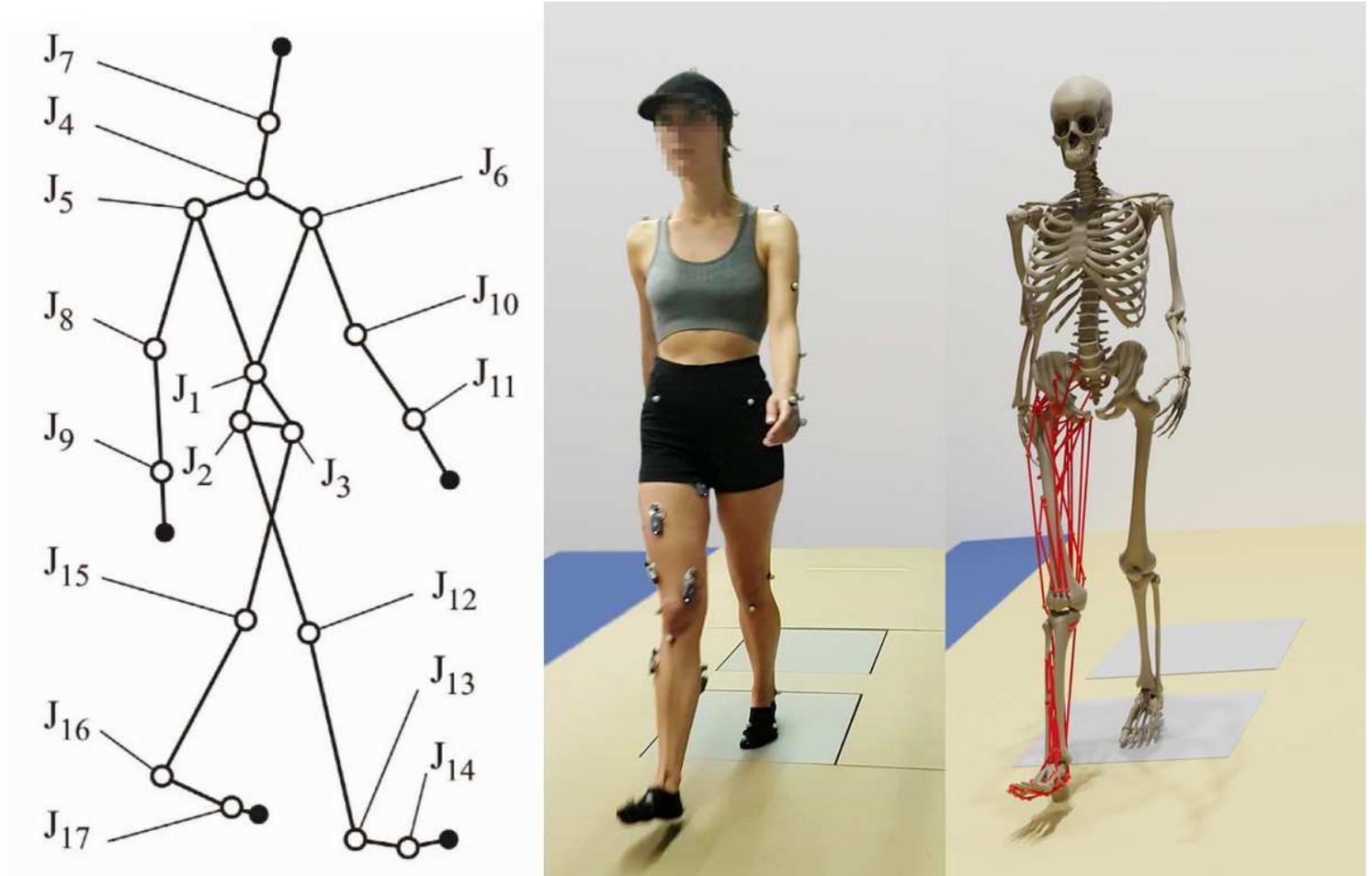


Figure 1

Gait of healthy subject: multibody model (left); acquired motion (middle); computational model (right).

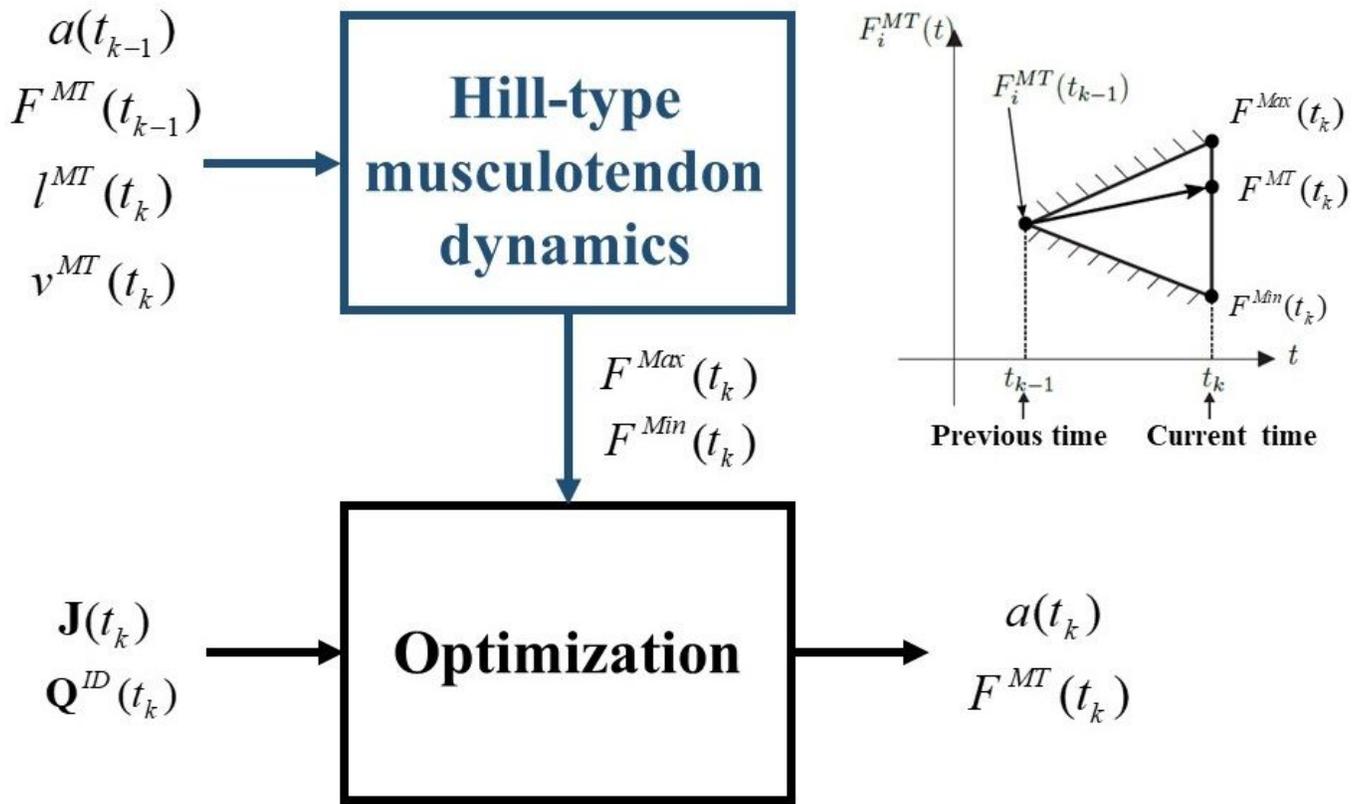


Figure 2

Physiological inverse-dynamics approach.

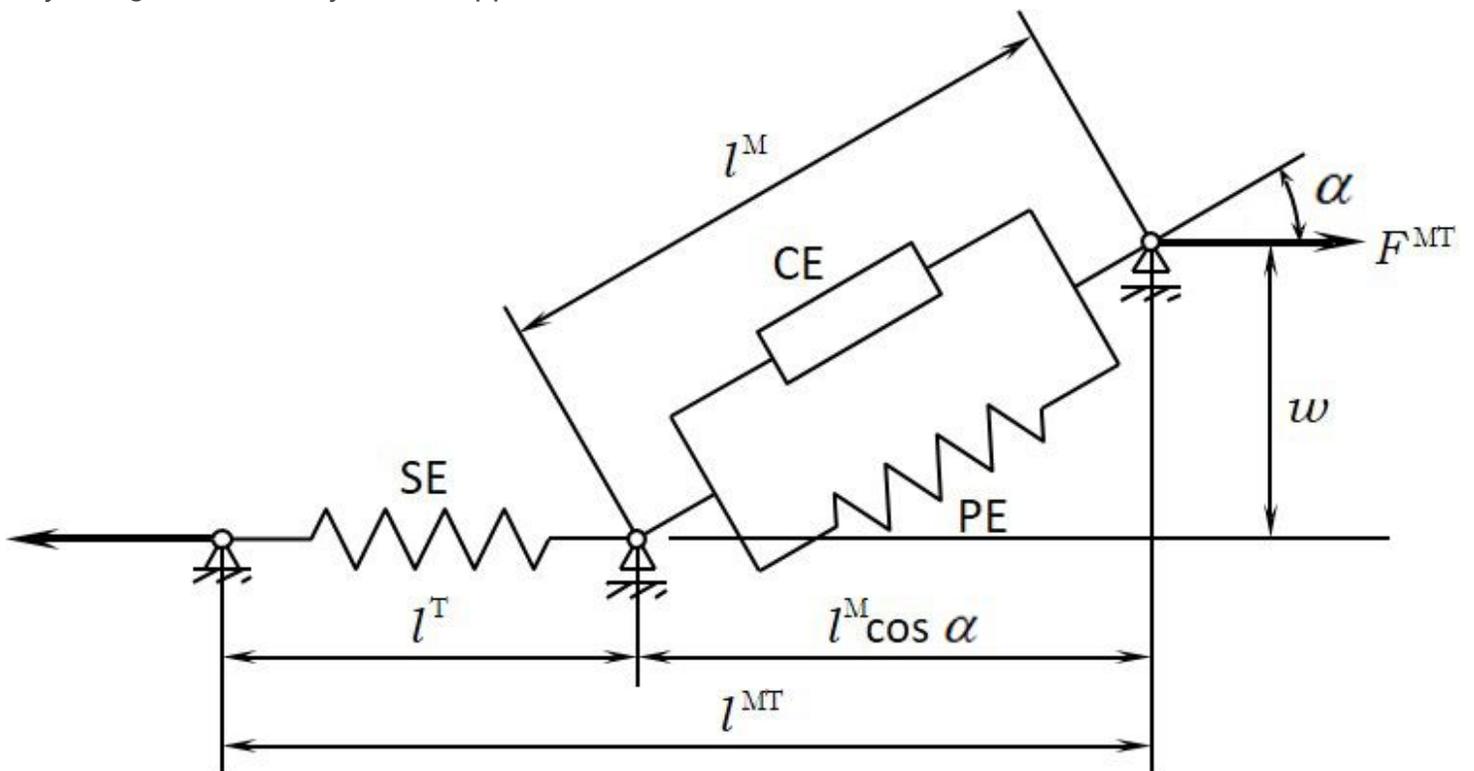


Figure 3

Hill-type muscle model. The muscle fibers are modeled as an active contractile element (CE) in parallel with a passive elastic component (PE). These elements are in series with a nonlinear elastic tendon (SE). The pennation angle denotes the angle between the muscle fibers and the tendon. Superscripts MT, M, and T indicate musculotendon, muscle fiber, and tendon, respectively.

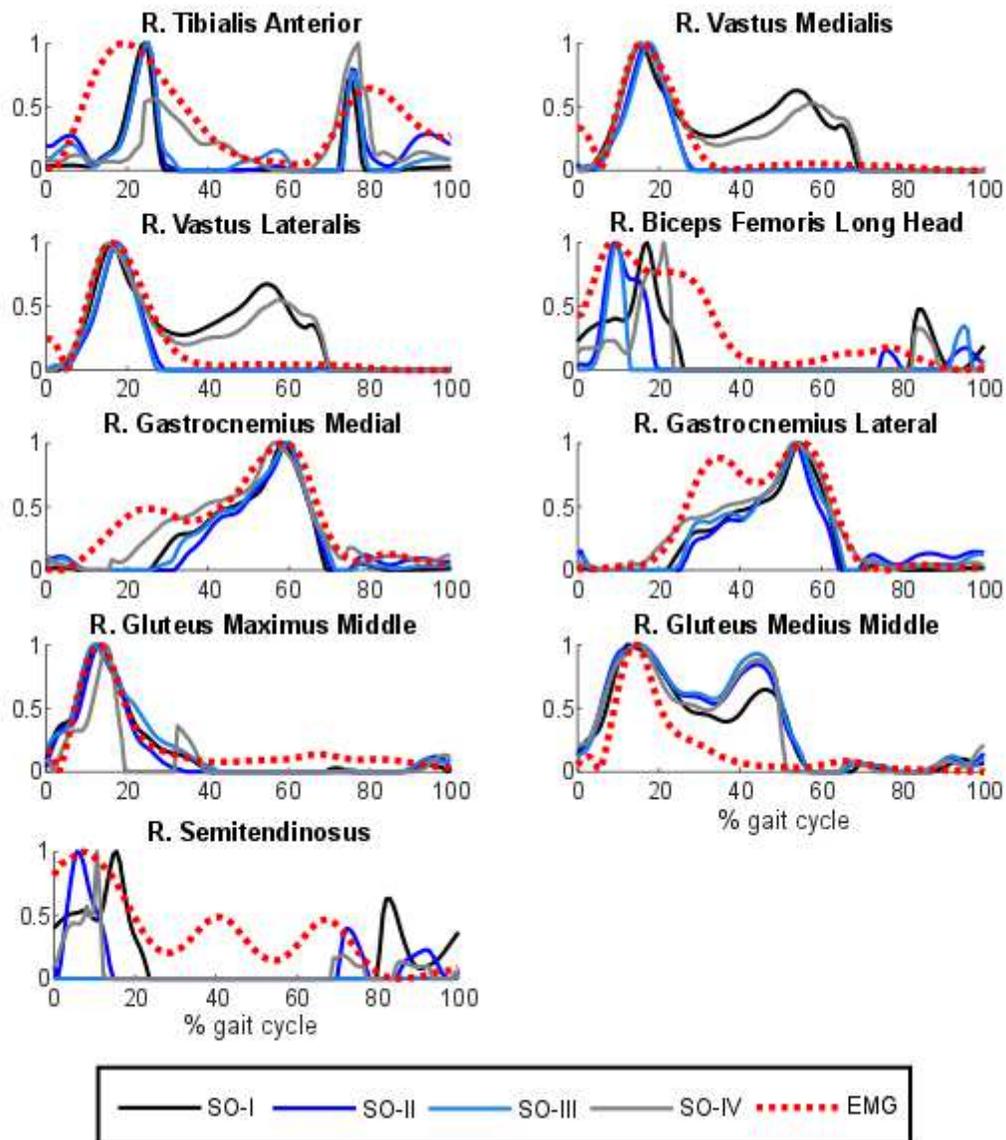


Figure 4

Normalized muscular activations obtained with static optimization (criteria I-IV) vs. normalized EMG for a healthy subject.

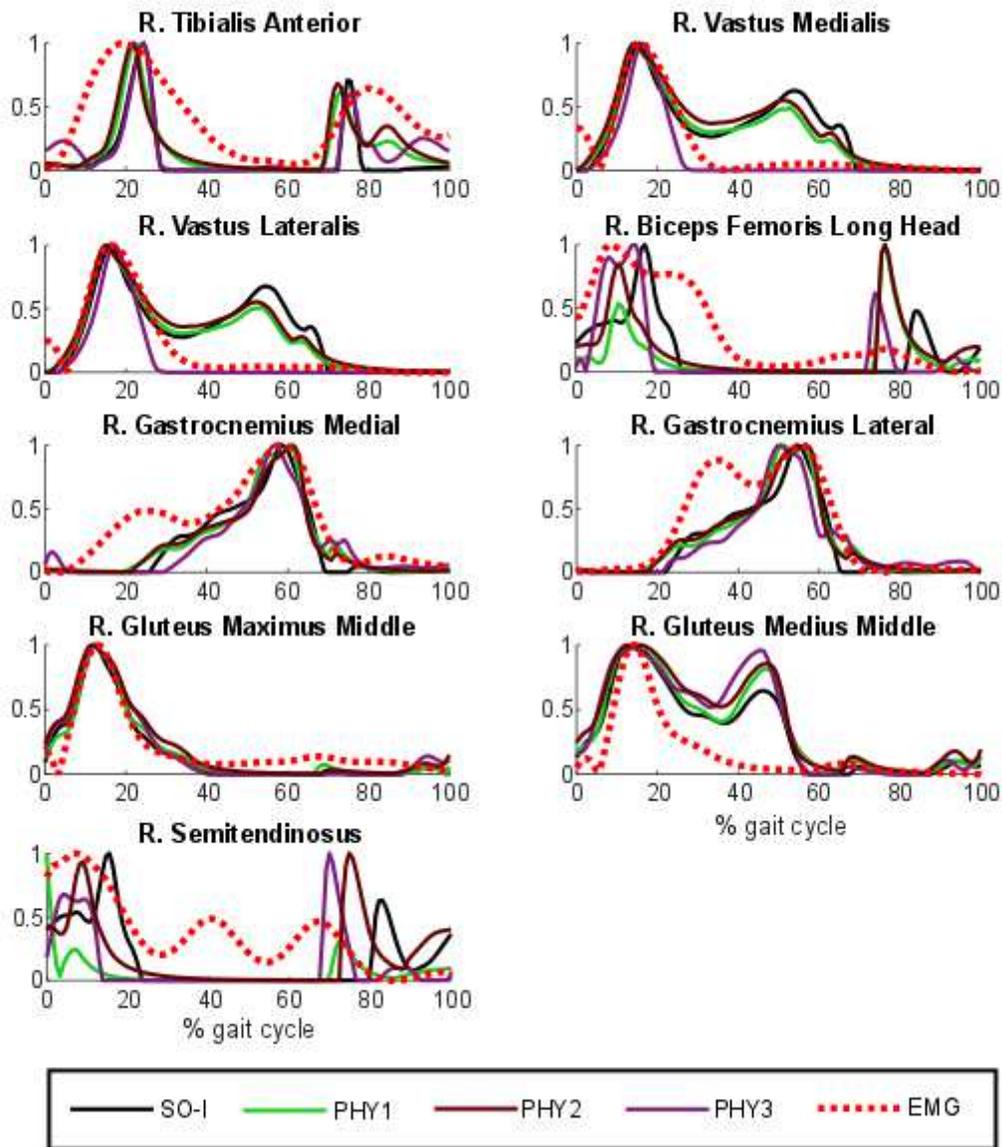


Figure 5

Normalized muscular activations obtained through static optimization with criterion I (SO-I) and physiological optimization with criterion I (original approach and two simplified alternatives) vs. normalized EMG for a healthy subject. PHY1: physiological approach; PHY2: physiological approach with rigid tendon and activation time response considered; PHY3: physiological approach with rigid tendon and activation time response ignored.

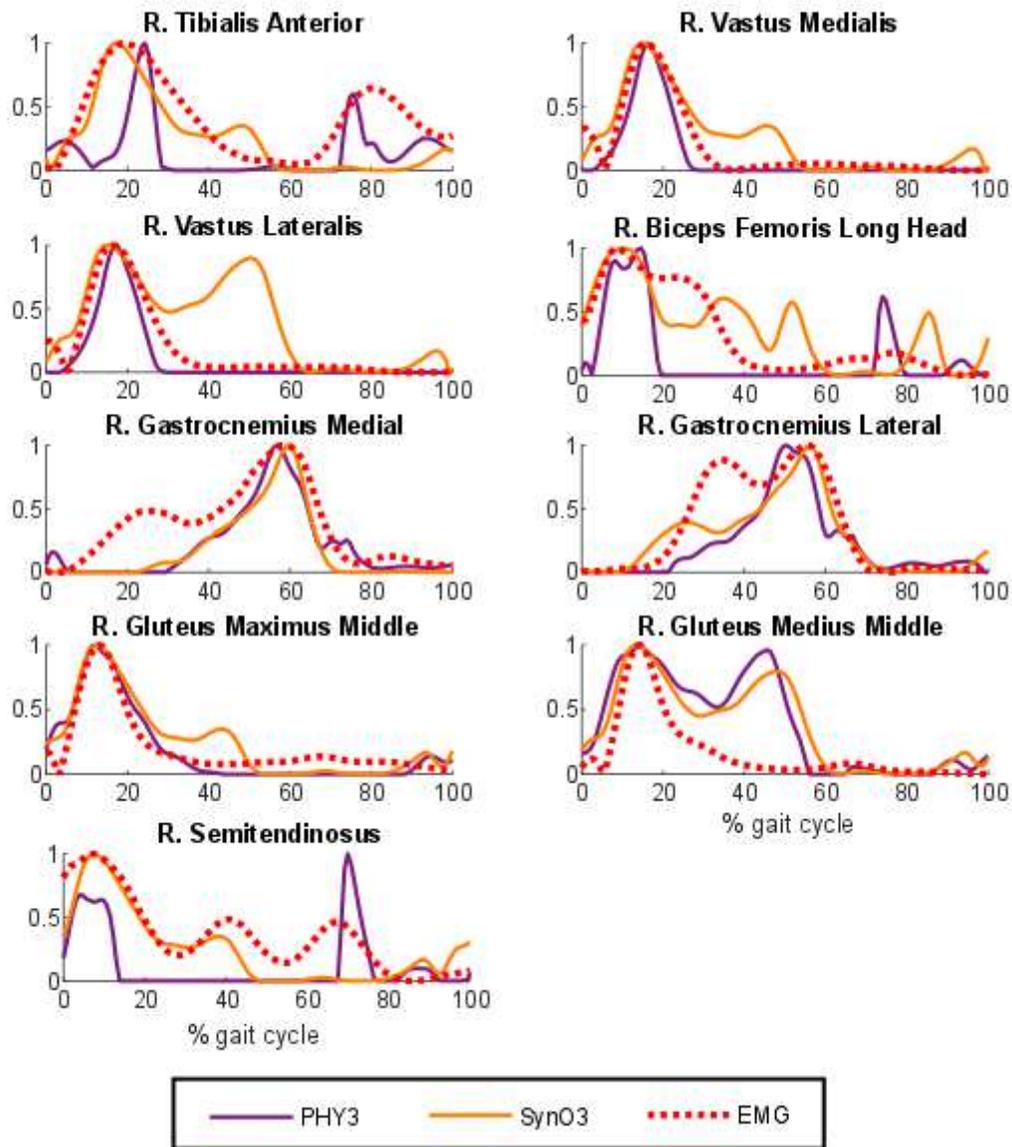


Figure 6

Normalized muscular activations obtained from physiological optimization with rigid tendon and activation time response ignored (PHY2) and synergy optimization with 3 synergies (Syn03) vs. normalized EMG for a healthy subject.

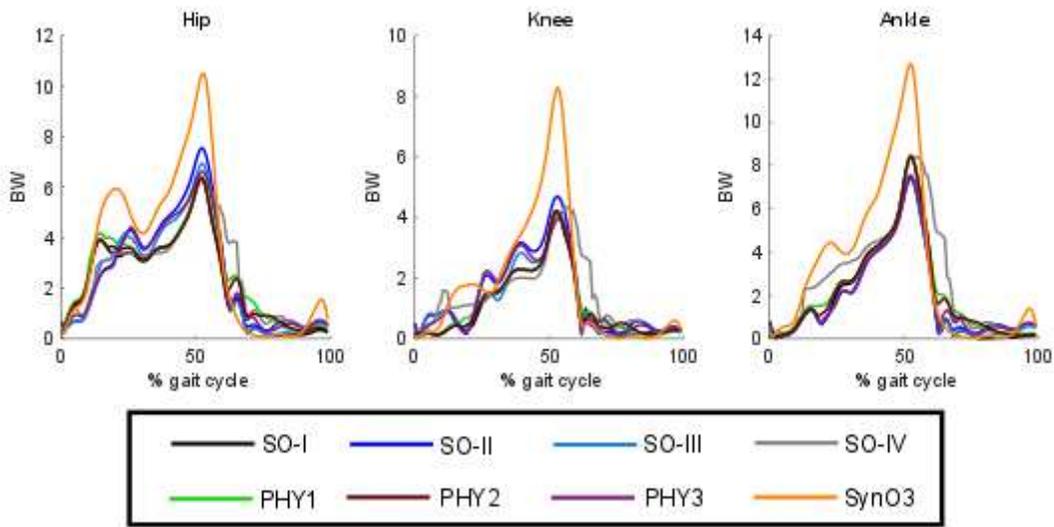


Figure 7

Joint reaction forces at hip, knee and ankle obtained with different muscle recruitment approaches.