

Description of a Rat Model of Polymicrobial Abdominal Sepsis Mimicking Human Colon Perforation

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Research note

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Abstract

Objective: Standard rodent sepsis models as cecal ligation and puncture models (CLP) or cecal ligation and incision models (CLI) are frequently not suited experiments, mainly because they lack surgical repair, and they are difficult to control for severity. The colon ascendens stent peritonitis model (CASP) overcomes some of these limitations.

Result: Here we present our modification of the rodent CASP model, where severity of sepsis can be controlled by timing of surgical repair and treatment, and by diameter of the stent. Further, basic hemodynamic monitoring (blood pressure and heart rate) and frequent blood sampling can be achieved, which might guide further treatment.

Introduction

Sepsis and septic shock frequently occur all over the world [1] and are associated with high mortality and unfavorable outcome [2, 3]. Although great efforts are made to improve sepsis recognition and treatment, neither sepsis burden nor mortality has changed in the past decade [3]. Given the higher risk of acquiring sepsis and of adverse outcome in the elderly [4], with the aging population in many countries sepsis will remain and even will rise as one of the most important health care threats.

With this perspective, new therapeutic options are urgently needed. Since sepsis is a complex syndrome defined as organ dysfunction associated with dysregulated host response to infection [4], preclinical research has to rely on animal models to explore the complex interaction of infectious agents and host factors. Commonly used research models are endotoxin/toxin administration, exogenously administered pathogenic bacteria, and host-barrier disruption models, where the animals' protective endogenous host barrier is altered. For practical and financial reasons, rodent models dominate over other species. The far most frequent models within this category are models where the intestinum is disrupted, and the intestinal content spills into the normally sterile peritoneal cavity and thus induces a polymicrobial sepsis. Cecal ligation and puncture (CLP models) and cecal ligation and incision (CLI models) are regarded as gold standard models because they resemble initiation and progression with characteristics of human sepsis, but this view has been challenged [5]. CLP and CLI do not completely mimic the representative course of human perforated intestine and are difficult to control in terms of severity and predictability. The colon ascendens stent peritonitis (CASP) model, first described in mice [6] and later in rats [7], overcomes this limitation.

In this manuscript, we describe the adaptations made to improve the CASP model in terms of monitoring hemodynamics and facilitating frequent blood sampling as well as non-surgical treatment.

Materials And Methods

These experiments were performed as a pilot of a project aiming to explore the effects of a new endotoxin scavenger as a randomized placebo-controlled trial, the results of the main experiments are not

published in this manuscript.

The experiment was reviewed and approved by the Animal Care and Experimentation Committee of the Canton of Bern, Switzerland (26169 BE6/15) and followed the Swiss national guidelines for the performance of animal experiments. Male Wistar rats aged 8–9 weeks, weighing 380 g in average were obtained from Janvier Labs (Le Genest-Saint-Isle, France) and kept in individually ventilated cages (IVC) with controlled 12 hours light/dark cycles at 22 ± 2 °C at the Central Animal Facility of the University of Bern. Food and water were provided ad libitum. A total of 56 animals were included (4 groups: 20 animals sham surgery ± scavenger/placebo, 36 animals CASP ± scavenger/placebo).

Anesthesia

Anesthesia was induced with 6% sevoflurane (Sevorane, AbbVie, Baar, Switzerland) in oxygen in an induction chamber, and 50 µg/kg fentanyl (Janssen-Cilag, Zug, Switzerland) was added intraperitoneally. The animals were intubated using a modified 2.0 mm Angiocath (BD, Allschwil; Switzerland) and mechanically ventilated with a small animal ventilator (KTR-4 Rodent Ventilator, Hugo Sachs, March, Germany). Anesthesia was maintained with sevoflurane 2.5–3.5%, and analgesia with additional 50 µg/kg fentanyl administered subcutaneously 30 minutes later. The animal was placed on an operation table with an in-built feedback controlled heater system (TCAT-2, Harvard Apparatus, Hugo Sachs, March, Germany) aiming for a constant body temperature of 37 °C.

Preparation

First, we incised the right groin, and then tunneled a Redon needle/wound drainage trocar from the neck to the groin. Two polyurethan catheters (PU 40, SAI Infusion Technologies, Lake Villa, IL, USA) were inserted in the subcutaneous tunnel. The catheters were then equipped with a 1.5 to 2 cm tip made of PE-50 for vessel cannulation, and filled with heparinized sterile normal saline solution (5000 I.U. heparin in 1000 ml NaCl 0.9%) via a blunt needle and luer lock syringe on the cranial part. The femoral artery and vein were microsurgically exposed, cannulated and secured with a suture. Before skin closure, the catheters were additionally bend as a loop and secured twice with sutures. The animals were turned in prone position and the catheters secured with a subcutaneous loop and suture at the neck. We channeled the catheters trough a tethered harness (SAI Infusion Technologies, Lake Villa, IL, USA) and connected them via a two channel swivel with 2 purge systems (syringe drivers) to guarantee free movement of the animal and integrity of the catheters. After drawing a first arterial and venous blood sample, both purge lines were continuously flushed with 1.5 ml/h Glucose 5% in H₂O 2:1 and heparin (5000 I.U. heparin in 1000 ml GS), to the arterial line purge we added buprenorphine (Temgesic, Reckitt Benckiser Switzerland, Wallisellen) to achieve a concentration of 1 µg/ml. The animals were weaned from the ventilator, extubated and placed for recovery from surgery in a cage equipped with a swivel mount. Duration of surgery was 50–60 minutes.

Colon Ascendens Stent Peritonitis

6 hours after finishing the preparation the animals were re-anesthetized with sevoflurane 6% in oxygen in the induction chamber, supplemented with 20 µg/kg buprenorphine subcutaneously. The animals were placed on the heated operation table, and anesthesia was maintained with 2.5 to 3.5% sevoflurane in oxygen via facemask. A median laparotomy was performed, and the coecum and the ascending colon identified and exposed. A small incision was made about 1 cm distal the iliocecal valve at the anti-mesenteric side, and a 12 French PU catheter of 1.5 cm length was inserted. This stent was additionally fixed to the colon by a suture. Patency was tested by instillation of saline into the stent, with spontaneous emptying of feces afterwards, or gentle squeezing the coecum until intestinal content was visible in the opening of the stent. The animals in the sham group received the same surgery, omitting colostomy, with the stent only sutured at the outer side of the colon ascendens. The intestine was packed back into the abdominal cavity and the abdominal wall was closed in two layers. Surgical time was below 20 min. The animal was placed back into its cage equipped with the swivel mount. Volume replacement was started, the buprenorphine dosing was resumed. Additionally the animals were provided with water and nourishment ad libitum.

Surgical repair and treatment

16 hours after peritonitis induction and randomization, animals were again re-anesthetized as described before using induction chamber and facemask. The abdominal sutures were re-opened, the stent identified, which could necessitate exploration with sterile cotton swabs to remove fibrin and detritus. Cultures from the abundant purulent ascites and detritus were taken at this time point for the main experiment. Once the stent was identified, the additional suture was cut, the stent removed and the intestinal wall closed with simple interrupted stitchings. The abdominal cavity was then rinsed with warm Ringer's solution, and the abdominal wall closed with sutures. Surgical time was below 20 minutes.

Then the rats were placed in their cages with the swivel mount. All animals received an intravenous infusion of meropenem (Meronem, Pfizer Switzerland, Zurich, 75 mg per kg BW, diluted to 5 mg/ml, in 15 min) followed by a continuous infusion via the central venous line (225 mg/kg BW/24 hours). Analgesia was provided continuously with buprenorphine administered via the intraarterial purge line.

Blood pressure was measured continuously with a standard transducer system linked to a BIOPAC MP100 analog-digital-converter and was acquired and analyzed with the Biopac AcqKnowledge Data Acquisition Software (BIOPAC, Goleta, CA 93117, USA). Heart rate was calculated with the blood pressure tracing.

Blood samples were drawn 16 and 40 hours post peritonitis induction, and after the last blood sampling. The animals were euthanized with an intravenous infusion of pentobarbital (150 mg/kg BW, Esconarkon, Streuli Pharma AG, Uznach, Switzerland).

Results

All but 2 animals survived the initial surgery. In the CASP group, 4 animals (out of 36) died or were euthanized according to the score sheet after sepsis induction, before randomization. No further premature death occurred after randomization and start of the treatment. Blood sampling was achieved before peritonitis induction and before start of meropenem (6 hours) in all animals. Line patency decreased over time, after 24 hours, blood samples could not be obtained in 9 animals, but did not decrease further.

In the 6 animals where microbiological samples from the abdominal cavity were drawn at repair, polymicrobial growth of gram negative and gram positive bacteria in great quantity could be documented as a proof of the infection with intestinal pathogens.

Because of the relatively long tubing system and connection via the swivel system, blood pressure tracings were damped. Tracing of reliable systolic and diastolic blood pressure reading were not possible, we relied on mean arterial pressure. Although clear signs of peritonitis were seen, and cultures grew abundant bacteria, systemic hypotension did not occur (mixed effects model for repeated measures data, $p = \text{ns}$, Fig. 1). Heart rate was significantly higher in the septic animals (mixed effects model, $p = 0.005$ for group, $p = 0.02$ for time, group x time factor: $p = 0.04$).

In conclusion, the addition of the arterial and venous catheters in the CASP model expands the research possibilities of the model. With these catheters, fluid management and anti-infective as well as analgesic treatment can be individualized, and blood sampling is facilitated. With potential adaptions of stent diameter and timing of surgery, this model can be used in a large variety of clinically relevant experiments.

Limitation

The model simulates the frequent clinical scenario of intestinal perforation, with subsequent surgical repair, adequate use of fluid resuscitation and antibiotic treatment. Addition of continuous arterial blood pressure and heart rate monitoring, continuous intravenous fluid administration over several hours and frequent blood sampling possibility this model approaches a frequent complex clinical scenario.

Traditional models have disadvantages depending on the research questions. Lipopolysaccharide injection models do not produce sepsis, but endotoxin shock; injection of bacteria or a defined amount of bacteria or fecal slurry lack the pathophysiology of gut discontinuation. Surgical models of abdominal sepsis (cecal ligation CL, cecal ligation and puncture CLP, cecal ligation and incision CLI and cecal ligation and dissection CLD) are moderately well controllable in terms of mortality and sepsis severity, and in surviving animals, abscess formation frequently occurs. This does not reflect the current medical scenario, where surgical treatment with closure of the gut discontinuation and lavage is attempted. The CASP model overcomes this drawback with surgical repair of the damage. Continuous bacterial contamination renders this model moderately controllable, but adaptation of the model can be

accomplished by different sizes of the stent [8], and by different timing of surgical repair. This model requires meticulous anesthesiology techniques because the repair surgery, albeit quickly done, is performed in a septic animal. The addition of arterial and venous catheters expand the possibilities of the model. Continuous blood pressure and heart rate monitoring can guide fluid management with frequent adjustments. Together with the venous catheter, both lines enable blood sampling without posing additional stress to the animal, and safe and reliable dosing of medication.

In this pilot experiment we found that combining the vessel cannulation and the CASP surgery in one single operation leads to an unacceptable high mortality. After modification and introducing a recovery period between both procedures the mortality during surgery declined. Another problem was failure to sample blood because of blocked catheters. Although we added heparin to the purge fluids, in some animals the catheter was partially clotted or developed a valve-like phenomena, where patency for injecting liquids was possible, but aspiration was not. With two lines in place, we could use the second line in some instances, but in 12%, no blood sampling was possible. This occurred exclusively in the 40 h post peritonitis and in the final blood sampling. If the experiment depends on frequent blood sampling, we recommend a different tubing system with biocompatible heparin bonded surfaces.

Because of the long tubing and the connection with a swivel, the arterial blood pressure curve was damped, and could not reliably be read as systolic/diastolic blood pressure anymore. Since tissue perfusion depends on mean arterial blood pressure this is not a major drawback[9].

List Of Abbreviations

CLP: cecal ligation and puncture

CLI: cecal ligation and incision models

CASP: colon ascendens stent peritonitis

CLD: cecal ligation and dissection

Declarations

Ethics approval and consent to participate:

The experiment was reviewed and approved by the Animal Care and Experimentation Committee of the Canton of Bern, Switzerland (26169 BE6/15) and followed the Swiss national guidelines for the performance of animal experiments.

Consent to publish:

N/A

Availability of data and materials:

the datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request

Competing interests:

The Department of Intensive Care Medicine, University Hospital Bern, has, or has had in the past, research contracts with Orion Corporation, Abbott Nutrition International, B. Braun Medical AG, CSEM SA, Edwards Lifesciences Services GmbH, Kenta Biotech Ltd, Maquet Critical Care AB, Omnicare Clinical Research AG and research and development/consulting contracts with Edwards Lifesciences SA, Maquet Critical 3Care AB and Nestlé. The money was paid into a departmental fund; no author received personal financial gain.

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Authors' contributions:

JU developed the protocol, performed the experiments, analyzed the data and drafted the manuscript; MG conceived the study, performed the experiments, analyzed the data and drafted the manuscript , AL performed the experiments and drafted the manuscript, MH conceived the experiment, oversaw development of the protocol and was responsible for the overall conduct of the trial, helped analyzing the data and edited the manuscript. All authors read and approved the final manuscript

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References

1. Fleischmann-Struzek C, Mikolajetz A, Schwarzkopf D, Cohen J, Hartog CS, Pletz M, Gastmeier P, Reinhart K: **Challenges in assessing the burden of sepsis and understanding the inequalities of sepsis outcomes between National Health Systems: secular trends in sepsis and infection incidence and mortality in Germany.** *Intensive Care Med* 2018, **44**(11):1826-1835.
2. Liu V, Escobar GJ, Greene JD, Soule J, Whippy A, Angus DC, Iwashyna TJ: **Hospital deaths in patients with sepsis from 2 independent cohorts.** *JAMA* 2014, **312**(1):90-92.
3. Rhee C, Dantes R, Epstein L, Murphy DJ, Seymour CW, Iwashyna TJ, Kadri SS, Angus DC, Danner RL, Fiore AE *et al*: **Incidence and Trends of Sepsis in US Hospitals Using Clinical vs Claims Data, 2009-2014.** *JAMA* 2017, **318**(13):1241-1249.
4. Gotts JE, Matthay MA: **Sepsis: pathophysiology and clinical management.** *BMJ* 2016, **353**:i1585.
5. Dejager L, Pinheiro I, Dejonckheere E, Libert C: **Cecal ligation and puncture: the gold standard model for polymicrobial sepsis?** *Trends in microbiology* 2011, **19**(4):198-208.
6. Zantl N, Uebe A, Neumann B, Wagner H, Siewert JR, Holzmann B, Heidecke CD, Pfeffer K: **Essential role of gamma interferon in survival of colon ascendens stent peritonitis, a novel murine model of abdominal sepsis.** *Infection and immunity* 1998, **66**(5):2300-2309.
7. Lustig MK, Bac VH, Pavlovic D, Maier S, Grundling M, Grisk O, Wendt M, Heidecke CD, Lehmann C: **Colon ascendens stent peritonitis—a model of sepsis adopted to the rat: physiological, microcirculatory and laboratory changes.** *Shock* 2007, **28**(1):59-64.
8. Maier S, Traeger T, Entlein M, Westerholt A, Kleist B, Hüser N, Holzmann B, Stier A, Pfeffer K, Heidecke CD: **Cecal ligation and puncture versus colon ascendens stent peritonitis: two distinct animal models for polymicrobial sepsis.** *Shock* 2004, **21**(6):505-511.
9. Leone M, Asfar P, Radermacher P, Vincent JL, Martin C: **Optimizing mean arterial pressure in septic shock: a critical reappraisal of the literature.** *Crit Care* 2015, **19**(1):101.

Figures

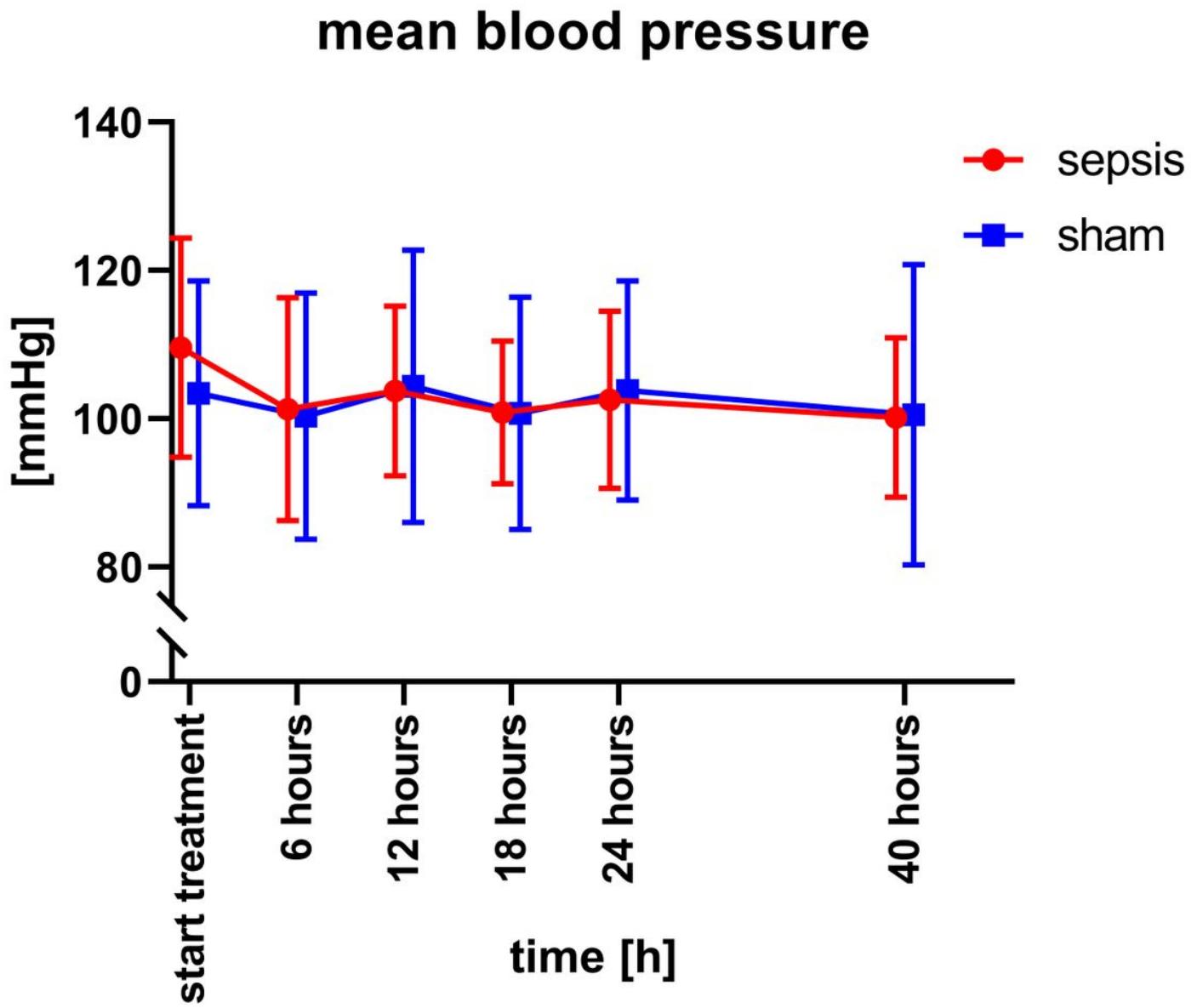


Figure 1

Blood pressure Time course of blood pressure und septic (red) and sham (blue) animals. Note that measurement of blood pressure began 16 hours after sepsis induction /sham surgery, when treatment with surgery and meropenem started. There are no significant differences between the groups

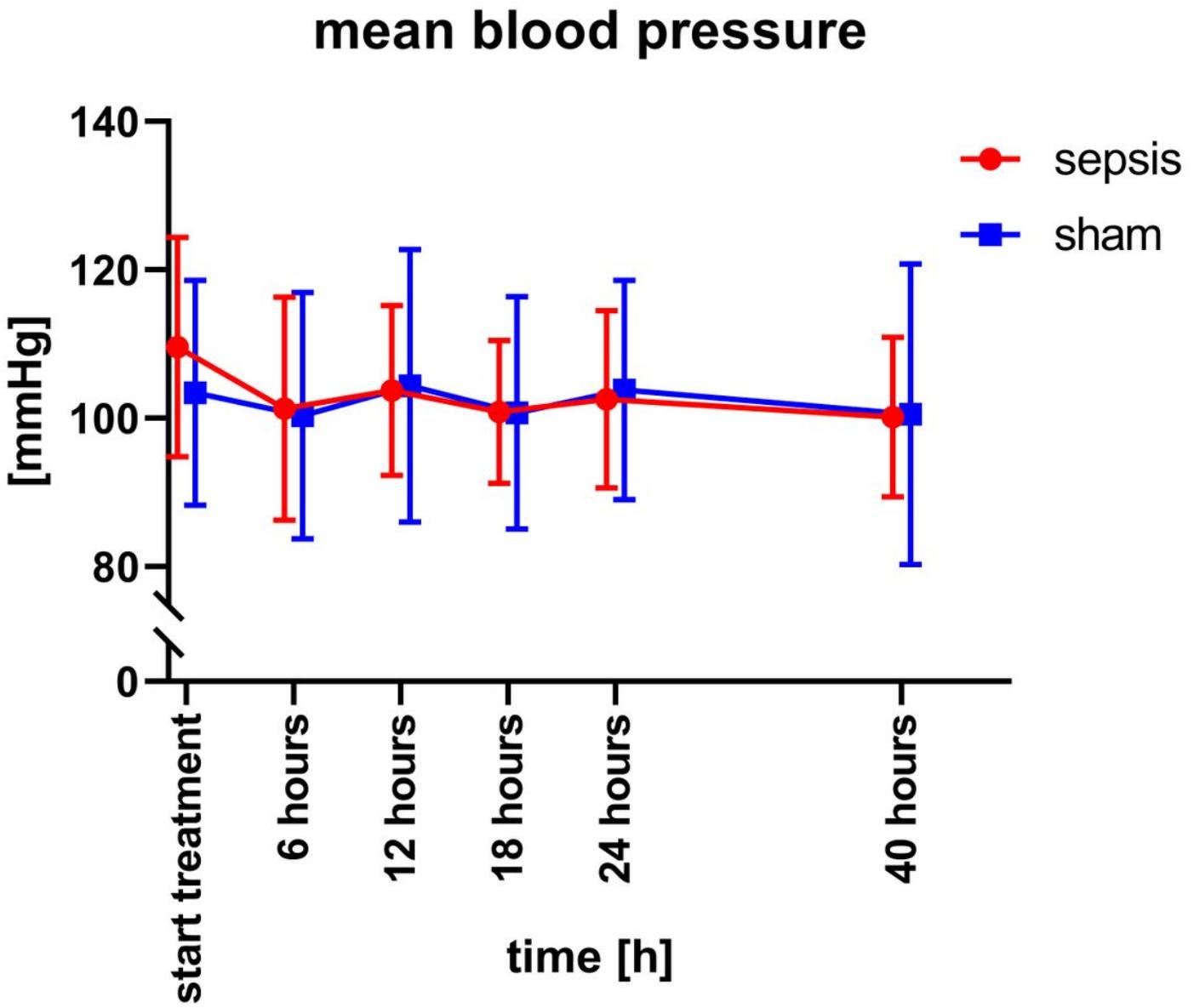


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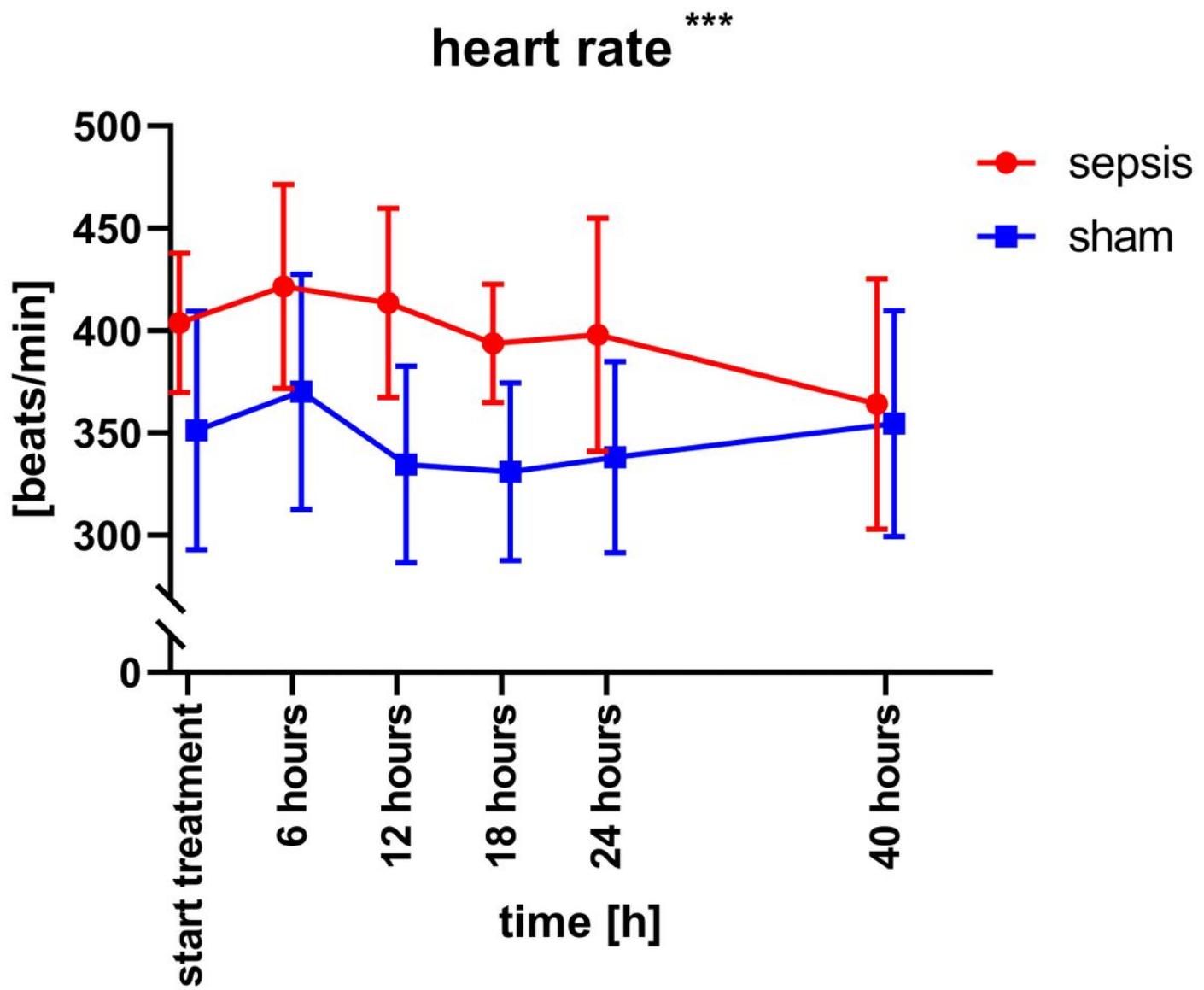


Figure 2

Heart rate Time course of heart rate und septic (red) and sham (blue) animals. Note that measurement of heart rate began 16 hours after sepsis induction /sham surgery, when treatment with surgery and meropenem started. The difference between the groups is significant ($p= 0.005$)

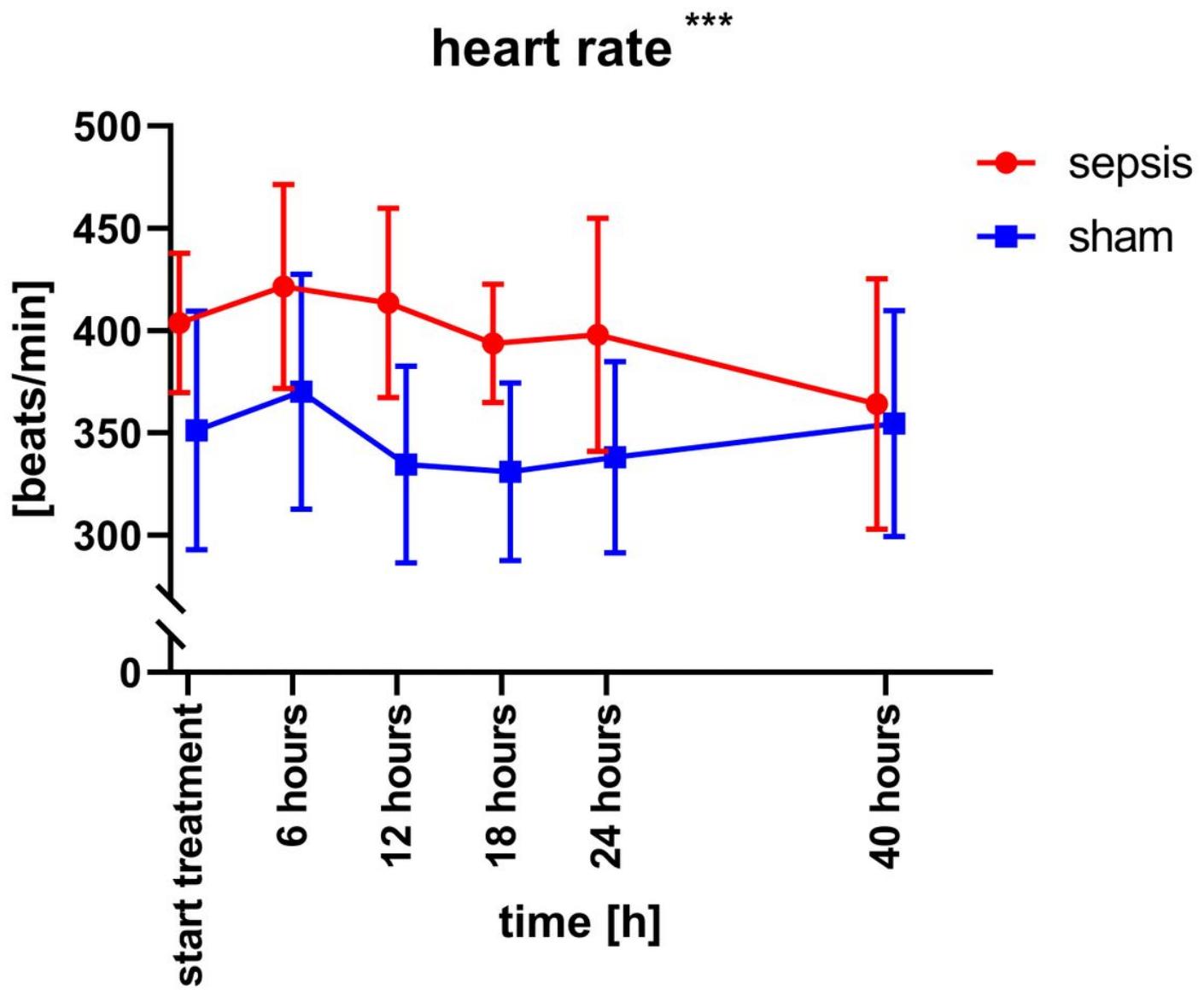


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