

# Pod pepper vein yellows virus, a new recombinant polerovirus infecting *Capsicum frutescens* in Yunnan province, China

**Kuangjie Zhao**

Ningbo University

**Yueyan Yin**

Yunnan Agricultural University

**Mengying Hua**

Ningbo University

**Shaoxiang Wang**

Wenshan Academy of Agricultural Sciences

**Xiaohan Mo**

Yunnan Academy of Tobacco Agricultural Sciences

**Enping Yuan**

Wenshan Academy of Agricultural Sciences

**Hongying Zheng**

Ningbo University

**Lin Lin**

Ningbo University

**Hairu Chen**

Yunnan Agricultural University

**Yuwen Lu**

Ningbo University

**Jianping Chen**

Ningbo University

**Jiejun Peng** (✉ [pengjiejun@yeah.net](mailto:pengjiejun@yeah.net))

Ningbo University <https://orcid.org/0000-0003-2594-9776>

**Fei Yan**

Ningbo University

---

## Short report

**Keywords:** pepper vein yellows virus, recombination, Polerovirus, readthrough domain

**Posted Date:** January 26th, 2021

**DOI:** <https://doi.org/10.21203/rs.3.rs-103456/v2>

**License:**  This work is licensed under a Creative Commons Attribution 4.0 International License. [Read Full License](#)

---

**Version of Record:** A version of this preprint was published on February 23rd, 2021. See the published version at <https://doi.org/10.1186/s12985-021-01511-5>.

## Abstract

Pepper vein yellows viruses (PeVYV) are phloem-restricted viruses in the genus *Polerovirus*, family *Luteoviridae*. Typical viral symptoms of PeVYV including interveinal yellowing of leaves and upward leaf curling were observed in pod pepper plants (*Capsicum frutescens*) growing in Wenshan city, Yunnan province, China. The complete genome sequence of a virus from a sample of these plants was determined by next-generation sequencing and RT-PCR. Pod pepper vein yellows virus (PoPeVYV) (MT188667) has a genome of 6015 nucleotides, and the characteristic genome organization of a member of the genus *Polerovirus*. In the 5' half of its genome (encoding P0 to P4), PoPeVYV is most similar (93.1% nt identity) to PeVYV-3 (*Pepper vein yellows virus 3*) (KP326573) but diverges greatly in the 3'-part encoding P5, where it is most similar (91.7% nt identity) to tobacco vein distorting virus (TVDV, EF529624) suggesting a recombinant origin. Recombination analysis predicted a single recombination event affecting nucleotide positions 4126 to 5192 nt, with PeVYV-3 as the major parent but with the region 4126-5192 nt derived from TVDV as the minor parent. A full-length clone of PoPeVYV was constructed and shown to be infectious in *C. frutescens* by RT-PCR and the presence of icosahedral viral particles.

## Background

Pepper vein yellows viruses (PeVYV) are phloem-restricted viruses in the genus *Polerovirus*, family *Luteoviridae* and are currently classified into six species (International Committee on Taxonomy of Viruses [ICTV] 2019 release), named *Pepper vein yellows virus 1* to *6* [1-6]. They have 86.2-94.6% nucleotide identity between them. Polerovirus genomes have seven open reading frames (ORF0 to ORF5 and ORF3a), putatively encoding proteins P0 to P5 and P3a [6]. Recombination is an important source of genetic variability in viruses, particularly for viruses possessing an RNA genome. PeVYVs have quite close nucleotide similarities to tobacco vein distorting virus (TVDV) in the 5' half of their genomes, and are considered to have arisen from recombination between TVDV and other poleroviruses [4, 7, 8]. We here report a new recombinant of PeVYV with high identity to PeVYV-3 in the 5' half of its genome and to TVDV in the 3' part.

## Main Text

Pod pepper (*Capsicum frutescens*) is widely planted in China, especially around Wenshan city, Yunnan province, and viral diseases have now also become a major threat to pepper production in Yunnan. During July 2019, 89 pepper leaf samples were collected from three different fields in Wenshan. These samples were collected from Da Longshu village (a, 45 samples), Bai Shiyan village (b, 28 samples) and Da Shuduan village (c, 16 samples). All had typical viral symptoms of interveinal leaf yellowing and fruit discoloration (Fig. S1). These leaves (0.4 g per sample) were mixed into three pooled samples by origin, and then sent to the Central Laboratory of Zhejiang Academy of Agricultural Sciences (Hangzhou, China) for Next-generation RNA-Seq sequencing (NGS). A total amount of 1 µg RNA per sample was used as input material for the RNA sample preparations. RNA integrity was checked by Agilent 2100 Bioanalyzer (Agilent Technologies). The TruSeq RNA Sample Preparation Kit (Illumina) was used to construct cDNA libraries according to the manufacturer's instructions. An Illumina NovaSeq 6000 platform with PE150 bp and CLC Genomics Workbench 20 (QIAGEN) was used for sequencing and data analysis. A total of 36,430,754 (a: 17,322,690, b: 11,735,694 and c: 7,372,370) paired-end reads were obtained, and 432,848 contigs (a: 143,689, b: 113,012 and c: 176,147) were generated *de novo* and compared with sequences in the GenBank nt using BLASTn, and 7 contigs were identified with E-values of zero. Contig\_62 was 5992 nt long and had high identities (>87.5%) to the genome of PeVYV-3 (*Pepper vein yellows virus 3*) (KP326573;[3]) and the other contigs were matched with Chilli ringspot virus (ChiRSV, 3 contigs), Chilli vein mottle virus (ChiVMV, 1 contig), Tomato zonate spot virus (TZSV, 1 contig) and Cucumber mosaic virus (CMV, 1 contig).

In order to verify the virus sequences, total RNA was isolated from each sample using TRIzol™ Reagent (Invitrogen) in compliance with the manufacturer's instructions. Reverse transcription (RT) polymerase chain reaction (PCR) analyses were performed using the ReverTra Ace™ qPCR RT Master Mix (Toyobo) and KOD-plus-Neo (Toyobo) following the manufacturer's protocol. RT was performed at 42°C for 60 min with random primers followed by 72°C for 10 min. The cycling conditions for the subsequent PCR were: 98°C 3min, and then 35 cycles of 98°C for 30s, 55°C for 30s, 68°C for 1kb/min; and 68°C for 10min. Primers designed from Contig\_62 were used to amplify a coat protein (CP) fragment of the virus (PeVYV-CP) (Table. S1). Fragments of the expected size (621bp) were obtained in 58 of the 89 symptomatic samples but not from healthy plants (raised from seed in the laboratory). After adding dATP at the 3'-terminal of the PCR products using 1 µL Taq™ (TaKaRa) at 72°C for 10min, they were cloned into pGEM-T Easy vector (Promega) and sequenced commercially (Sangon Biotech). The sequences were aligned using MUSCLE, and pairwise nucleotide sequence comparisons were done using the SDT (Species Demarcation Tool) v1.2 program[9, 10]. The 116 amplicons were 93.0-98.4% identical to PeVYV-3, and so we tentatively designated this virus in pod pepper as Pod Pepper vein yellows virus (PoPeVYV).

In order to acquire the full-length sequence of PoPeVYV in pod pepper, 5' and 3'RACE reactions were then performed to obtain the complete terminal sequences. In brief, the 3' end of the viral RNA was polyadenylated using Poly(A) Tailing Kit (Invitrogen) and first-strand

cDNA was synthesized using M4T primer (Table. S1). The 3' terminal end was PCR amplified from the cDNA using M4 and 3'-RACE-F (Tables. S1). Similarly, the 5' end of the cDNA, after purification by treatment with Gel extraction Kit (Omega) was polyadenylated using the Poly(A) Tailing Kit. The purified cDNA/RNA heterocomplex was ligated with ZHM1 using T4 RNA ligase (Thermo Scientific) and PCR amplified using ZHM2 and 5' RACE-R (Table. S1; Fig. 1A and B). To avoid errors in sequence assembly, the whole viral sequence was then amplified using two overlapping sections with the primer pairs PoPeVYV-1 and PoPeVYV-2 (Table. S1; Fig. 1C). RT-PCR products (expected sizes 4614 bp and 2824bp) were obtained using the methods of KOD-plus-Neo (Toyobo) and the two products (which overlapped by 1423 bp) were cloned into the pGEM-T Easy Vector (Promega) and sequenced. We obtained 14 amplicons of PoPeVYV-1 and 4 amplicons of PoPeVYV-2, and found that 12 amplicons of PoPeVYV-1 exactly matched the overlapping region of PoPeVYV-2. The complete sequence was 6015 nt long (GenBank accession number: MT188667).

PoPeVYV has a genome organization characteristic of members of the genus *Polerovirus*, with seven predicted genes encoding proteins P0 to P5 and P3a [6] (Fig. 1A). Over its entire genome our new virus is related (87.8% nucleotide identity) to Tobacco vein distorting virus (TVDV, accession EF529624) and had 85.0% nt identity to the (Chinese) PeVYV-3 (accession KP300822) (Table. 1). However, the 5' half (nts 1-4251) of PoPeVYV has 93.1% nt identity to the corresponding region of PeVYV-3, and 87.7% nt identity to that of TVDV. The 3' half (nts 4252-6015) of PoPeVYV has only 64.1% nt identity with PeVYV-3, but 91.7% nt identity with TVDV (Fig. S2). These results indicated that PoPeVYV in pod pepper might be a recombined virus with PeVYV-3 and TVDV as parents.

Poleroviruses are prone to recombination among themselves or with viruses belonging to other genera and the relationships between PoPeVYV, the previously described PeVYV isolates and TVDV suggests that there has been a recombination event affecting the 3'-end of the genome. This was confirmed using a variety of methods on the RDP4 recombinant platform [11]. A single recombination event affecting nucleotide positions 4126-5952, with TVDV and PeVYV-3 as the respective minor and major parents was consistently identified using GENECONV (P value of 1.035 E-93), RDP (8.410 E-85), BootScan (7.726 E-80), MaxChi (4.294 E-35), Chimera (7.719 E-05), SiScan (3.341 E-68), and Phylpro (2.331 E-15) (Fig. 2). Alignment of amino acid sequences of the PoPeVYV proteins with those of the six species of PeVYVs and TVDV also indicated a recombination event (Table. 1). Phylogenetic trees were constructed from the complete genome sequences, 5'- half or 3'- half of other poleroviruses and an enamovirus as an outgroup (Fig. S3). PoPeVYV, PeVYVs and TVDV formed an independent group separate from the other poleroviruses. The analysis confirmed that the genome and 3' half of PoPeVYV was similar to TVDV, but that the 5' half of PoPeVYV was similar to the PeVYVs (Fig. S3). This is reflected in the phylogenetic analysis of the amino acid sequences of the separate gene products [12]: the P0 of PoPeVYV is most similar to PeVYV-1/4, P1/P2/P3/P4 are closely related to other PeVYVs but the RTD is most similar to TVDV (Fig. 3). However, proteins encoded by the 5' half of the genome (P0 to P4) had the highest identity to those of PeVYV-3 and PeVYV-6, whereas proteins translated from the 3' half (P5/RTD) were more closely related to those of TVDV (Table. 1, Fig. 3). The various PeVYVs recognized usually have distinct P0 protein sequences and the topology of the P0 phylogenetic tree may depict the ongoing selection for a protein exhibiting better RNA silencing suppression capacity after a recent host jump [4, 7, 13]. The P0 of PoPeVYV has 77.5-87.1% amino acid identity and 85.2-90.5% nucleotide identity to that of the other PeVYVs, and 76.3% amino acid identity (81.3% nucleotide identity) to that of TVDV (Table. 1). In the P0 phylogenetic tree PoPeVYV was most similar to PeVYV-1 and PeVYV-4 and was clearly more similar to the PeVYVs than to TVDV (Fig. 3). These results suggest that PoPeVYV is a new recombinant polerovirus.

An infectious clone of the virus was constructed for further investigation. RT-PCR was performed using KOD-plus-Neo (Toyobo) following the manufacturer's protocol. To generate infectious clones, the CloneExpress MultiS One Step Cloning Kit (Vazyme) was used for homologous recombination. Two overlapped PCR products were amplified with primers (Inf-PoPeVYV-1/2), and recombined with the linearized binary vector pCB301-MD (a modified version of pCB301 [14]), which includes the double 35S promoter and nopaline synthase terminator (NOS) (Fig.S4)[15, 16]. In this way, the full-length PoPeVYV cDNA was inserted between an upstream 35S promoter and a downstream hepatitis delta virus (HDV) ribozyme and NOS terminator in the binary vector to construct pCB-PoPeVYV. The PoPeVYV cDNA was ligated with 3'-terminal of 35S promoter without any extra nucleotide (Fig. S5)[15, 16]. This clone was transformed into *Agrobacterium tumefaciens* which was then delivered to *C. frutescens* plantlets by infiltration. There was mild upward leaf curling 45 days after inoculation (Fig. 4A), and RT-PCR using primers to detect the coat protein gene in the newly-emerged non-inoculated leaves showed that viral RNA was present and had spread systemically in all the inoculated plants (12/12) but not in the controls (Fig. 4B). Virions were purified from *C. frutescens* leaves using the method described previously [17]. Isometric particles about 25nm in diameter were observed in the purified preparation from the inoculated plants (Fig. 4C) but not from the controls. RT-PCR and subsequent sequencing confirmed that the virus in the symptomatic (systemic) leaves had the same sequence as the PoPeVYV cDNA clone inoculated (data not show). These results demonstrate the infectivity of the full-length PoPeVYV to *C. frutescens*.

In fields of cultivated pod pepper in Wenshan, we had observed severe yellowing and curling symptoms with small leaves (Fig. S1). Those symptoms were more severe than we observed in plants inoculated with the infectious clone of PoPeVYV by *Agrobacterium*

*tumefaciens* which also did not have the obvious vein yellowing usually caused by PeVYVs. Previous studies have shown that the P0 proteins of poleroviruses display variable RNA silencing suppression activity, and induce distinct symptoms [18, 19]. The P0 of PoPeVYV has only 77.5-87.1% amino acid identity to the other PeVYVs, which could account for the distinct symptoms but the field plants were also infected with other viruses (PeVYVs, ChiVMV, ChiRSV, CMV *etc*) as frequently happens in the field and which has also been reported in Taiwan and Thailand [20, 21]. The severe viral symptoms in the field may therefore be a synergistic effect of mixed infection.

Recombination is an important source of genetic variability in viruses, particularly for viruses possessing an RNA genome. PeVYVs have higher identities to TVDV in the 5' half of the genome and are considered to be recombinants of TVDV and other poleroviruses [4, 7, 8]. The new recombinant identified here has higher identity to TVDV at the 3' part of genome, indicating a different sort of recombinant event. The 5' half of the PoPeVYV genome shares high identity with PeVYV-3, which was reported from Hunan province in China [3]. The 3' half is much more distant from PeVYV-3 but is more homologous with that of TVDV. TVDV cause a devastating tobacco disease in many tobacco producing areas include Hunan, Guizhou and Yunnan provinces in China [22, 23]. Additionally, PeVYV has been reported to infect tobacco in Guizhou province suggesting that co-infection of tobacco might have provided an opportunity for recombination [24]. Recombination poses a problem for classification. The currently-recommended species demarcation criteria in the family *Luteoviridae* suggest that different species should have >10% difference in amino acid sequence identity in any gene product from their closest relative. The P0 and P5 proteins of PoPeVYV have respectively 12.9-23.7% and 10.3-70.5% difference in amino acid identity to those of PeVYV1-6 (Table. 1). If applied here, those criteria suggest that PoPeVYV could be representative of a distinct species.

## Conclusions

A full-length sequence of Pod pepper vein yellows virus (MT188667) was determined. Alignment and recombination analysis predicted a single recombination event with PeVYV-3 as the major parent but with the region 4126-5192 nt derived from TVDV as the minor parent. PoPeVYV is a new recombinant polerovirus infecting *C. frutescens* in Yunnan province, China.

## Abbreviations

nt: Nucleotide; RACE: Rapid amplification of cDNA ends; RT-PCR: Reverse transcription polymerase chain reaction; PoPeVYV: Pod Pepper vein yellows virus; PeVYV: Pepper vein yellows virus; TVDV: Tobacco vein distorting virus

## Declarations

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Availability of data and materials

The complete genome sequences of PoPeVYV were submitted to the GenBank (<http://www.ncbi.nlm.nih.gov/genbank/>) and the accession number is MT188667.

### Competing interests

The authors declare that they have no conflict of interest.

### Funding

This work was financially supported by Ningbo city Natural Science Foundation of China (2019A610403), Chinese Agriculture Research System (CARS-24-C-04). This work also was supported by grant-in-aid from State Key Laboratory for Managing Biotic and Chemical Threats to the Quality and Safety of Agroproducts (KF20190107) and K. C.Wong education foundation.

### Authors' contributions

FY, JP conceived and designed the experiments. SW, EY, LL collected the samples. XM, KZ, YY, LL and MH performed the experiments. HC, YL and JC analyzed the data. KZ, YY, JP and FY wrote the paper. All authors read and approved the final manuscript.

## Acknowledgements

We thank Professor Mike Adams for manuscript correction.

## Authors' information

<sup>1</sup>. State Key Laboratory for Managing Biotic and Chemical Threats to the Quality and Safety of Agroproducts, Institute of Plant Virology, Ningbo University, Ningbo 315211, Zhejiang, China. <sup>2</sup>. Institute of Alpine Economic Plants, Yunnan Academy of Agricultural Sciences, Lijiang 674100, Yunnan, China. <sup>3</sup>. Department of Plant Protection, Yunnan Agricultural University, Kunming 650201, Yunnan, China. <sup>4</sup>. Wenshan Academy of Agricultural Sciences, Wenshan 663000, Yunnan, China. <sup>5</sup>. Yunnan Academy of Tobacco Agricultural Sciences, Kunming 650021, Yunnan, China.

## References

1. Dombrovsky A, Glanz E, Lachman O, et al: The complete genomic sequence of pepper yellow leaf curl virus (PYLCV) and its implications for our understanding of evolution dynamics in the genus polerovirus. *PLoS One* 2013, 8:e70722.
2. Knierim D, Tsai WS, Kenyon L: Analysis of sequences from field samples reveals the presence of the recently described pepper vein yellows virus (genus Polerovirus) in six additional countries. *Arch Virol* 2013, 158:1337-1341.
3. Liu M, Liu X, Li X, et al: Complete genome sequence of a Chinese isolate of pepper vein yellows virus and evolutionary analysis based on the CP, MP and RdRp coding regions. *Arch Virol* 2016, 161:677-683.
4. Lotos L, Olmos A, Orfanidou C, et al: Insights Into the Etiology of Polerovirus-Induced Pepper Yellows Disease. *Phytopathology* 2017, 107:1567-1576.
5. Maina S, Edwards OR, Jones RA: First Complete Genome Sequence of Pepper vein yellows virus from Australia. *Genome Announc* 2016, 4.
6. Murakami R, Nakashima N, Hinomoto N, et al: The genome sequence of pepper vein yellows virus (family Luteoviridae, genus Polerovirus). *Archives of Virology* 2011, 156:921-923.
7. Ghosh S, Kanakala S, Lebedev G, et al: Transmission of a New Polerovirus Infecting Pepper by the Whitefly *Bemisia tabaci*. *J Virol* 2019, 93.
8. Kamran A, Lotos L, Amer MA, et al: Characterization of Pepper leafroll chlorosis virus, a New Polerovirus Causing Yellowing Disease of Bell Pepper in Saudi Arabia. *Plant Disease* 2018, 102:318-326.
9. Edgar RC: MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res* 2004, 32:1792-1797.
10. Muhire BM, Varsani A, Martin DP: SDT: a virus classification tool based on pairwise sequence alignment and identity calculation. *PLoS One* 2014, 9:e108277.
11. Martin DP, Murrell B, Golden M, et al: RDP4: Detection and analysis of recombination patterns in virus genomes. *Virus Evol* 2015, 1:vev003.
12. Kumar S, Stecher G, Li M, et al: MEGA X: Molecular Evolutionary Genetics Analysis across Computing Platforms. *Mol Biol Evol* 2018, 35:1547-1549.
13. Koeda S, Homma K, Kamitani M, et al: Pepper vein yellows virus 9: a novel polerovirus isolated from chili pepper in Indonesia. *Arch Virol* 2020, 165:3017-3021.
14. Xiang C, Han P, Lutziger I, et al: A mini binary vector series for plant transformation. *Plant Mol Biol* 1999, 40:711-717.
15. Shi BJ, Ding SW, Symons RH: Plasmid vector for cloning infectious cDNAs from plant RNA viruses: high infectivity of cDNA clones of tomato aspermy cucumovirus. *J Gen Virol* 1997, 78 ( Pt 5):1181-1185.
16. Scholthof HB: Rapid delivery of foreign genes into plants by direct rub-inoculation with intact plasmid DNA of a tomato bushy stunt virus gene vector. *J Virol* 1999, 73:7823-7829.
17. Mo X-H, Chen Z-B, Chen J-P: Molecular identification and phylogenetic analysis of a viral RNA associated with the Chinese tobacco bushy top disease complex. *Annals of Applied Biology* 2011, 158:188-193.
18. Kozłowska-Makulska A, Guilley H, Szyndel MS, et al: P0 proteins of European beet-infecting poleroviruses display variable RNA silencing suppression activity. *J Gen Virol* 2010, 91:1082-1091.

19. Almasi R, Miller WA, Ziegler-Graff V: Mild and severe cereal yellow dwarf viruses differ in silencing suppressor efficiency of the P0 protein. *Virus Res* 2015, 208:199-206.
20. Laprom A, Nilthong S, Chukeatirote E: Incidence of viruses infecting pepper in Thailand. *Biomol Concepts* 2019, 10:184-193.
21. Cheng YH, Deng TC, Chen CC, et al: First Report of Pepper mottle virus in Bell Pepper in Taiwan. *Plant Dis* 2011, 95:617.
22. Mo X-H, Qin X-Y, Tan Z-X, et al: First Report of Tobacco Bushy Top Disease in China. *Plant Disease* 2002, 86:74-74.
23. Liu F, Tan G, Li X, et al: Simultaneous detection of four causal agents of tobacco bushy top disease by a multiplex one-step RT-PCR. *Journal of Virological Methods* 2014, 205:99-103.
24. Wang LS, He QC, Chen XJ, et al: First Report of Pepper vein yellows virus Infecting Tobacco (*Nicotiana tabacum*) Naturally in China. *Plant Disease* 2017, 101:1556.

## Tables

Tabel. 1 Identity percentages between the genomes of PoPeVYV and with those of closely related species

	5'NCR		P0		P1		P2		iNCR		P3a		P3		P4		RTD		3'NCR	Full genome
	nt	aa	nt	aa	nt	aa	nt	aa	nt	aa	nt	aa	nt	aa	nt	aa	nt	aa	nt	nt
PoPeVYV	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100
PeVYV-1	<b>98.0</b>	83.5	88.9	89.8	91.9	93.4	93.3	<b>96.3</b>	97.8	97.8	94.2	93.1	89.7	94.7	49.5	55.8	92.4	84.6	84.6	84.6
PeVYV-2	<b>98.0</b>	83.9	88.9	89.9	92.1	94.7	93.8	92.5	91.1	92.0	93.1	92.8	87.6	93.3	65.9	68.9	88.0	87.0	87.0	87.0
PeVYV-3	<b>98.0</b>	<b>87.1</b>	<b>90.5</b>	<b>90.8</b>	<b>92.6</b>	93.9	92.7	95.1	<b>100</b>	<b>97.8</b>	<b>94.2</b>	<b>94.7</b>	90.4	94.7	49.7	60.6	91.6	85.0	85.0	85.0
PeVYV-4	66.0	85.1	89.2	90.1	91.5	94.4	93.7	92.5	95.6	94.9	92.7	92.6	91.0	94.5	49.1	55.8	92.4	84.1	84.1	84.1
PeVYV-5	96.0	77.5	85.2	87.2	90.3	95.0	93.5	92.5	93.3	94.9	94.2	94.4	<b>92.9</b>	<b>95.8</b>	51.8	57.2	<b>92.7</b>	84.1	84.1	84.1
PeVYV-6	96.0	85.1	89.6	90.5	92.9	<b>95.0</b>	<b>94.1</b>	90.0	95.6	94.2	94.7	94.2	92.3	95.8	51.8	58.4	<b>92.7</b>	85.3	85.3	85.3
TVDV	81.6	76.3	81.3	77.2	83.0	90.8	88.6	84.8	95.6	87.7	89.8	92.2	84.6	90.9	<b>89.7</b>	<b>91.9</b>	88.6	<b>87.8</b>	87.8	87.8

R = noncoding region, iNCR = intergenic NCR, aa = amino acid, and nt = nucleotide. Highest percentages are underlined and in bold.

VYV-1 (*Pepper vein yellows virus-1*, AB594828), PeVYV-2 (*Pepper vein yellows virus-2*, HM439608), PeVYV-3 (*Pepper vein yellows virus-3*, KP300822), PeVYV-4 (*Pepper vein yellows virus-4*, KU999109), PeVYV-5 (*Pepper vein yellows virus-5*, 523072), PeVYV-6 (*Pepper vein yellows virus-6*, LT559483), TVDV (*Tobacco vein distorting virus*, EF529624), PoPeVYV (*Pepper vein yellows virus*, MT188667)

## Figures

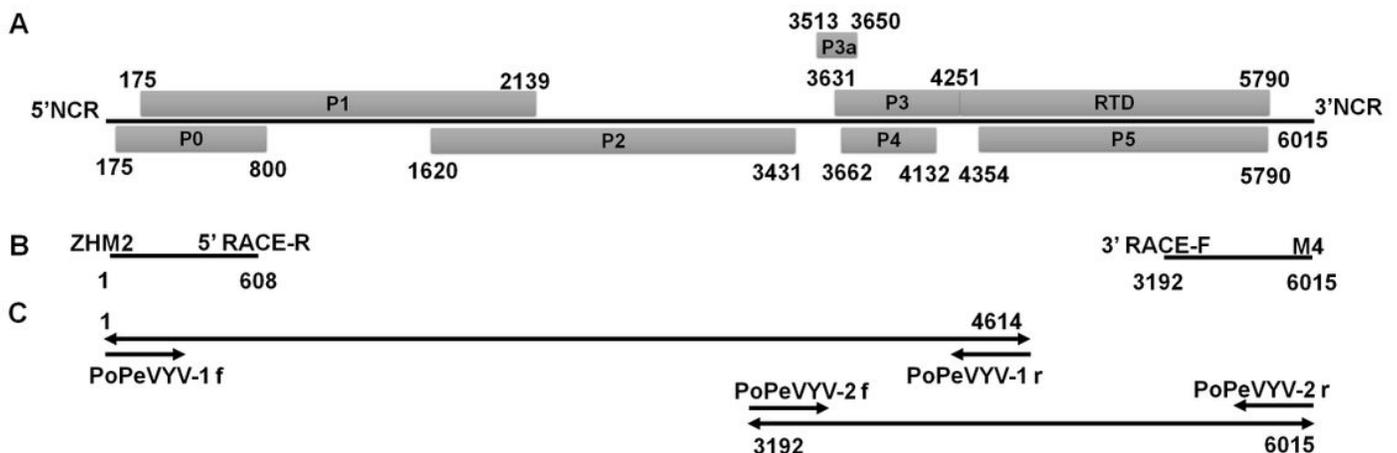
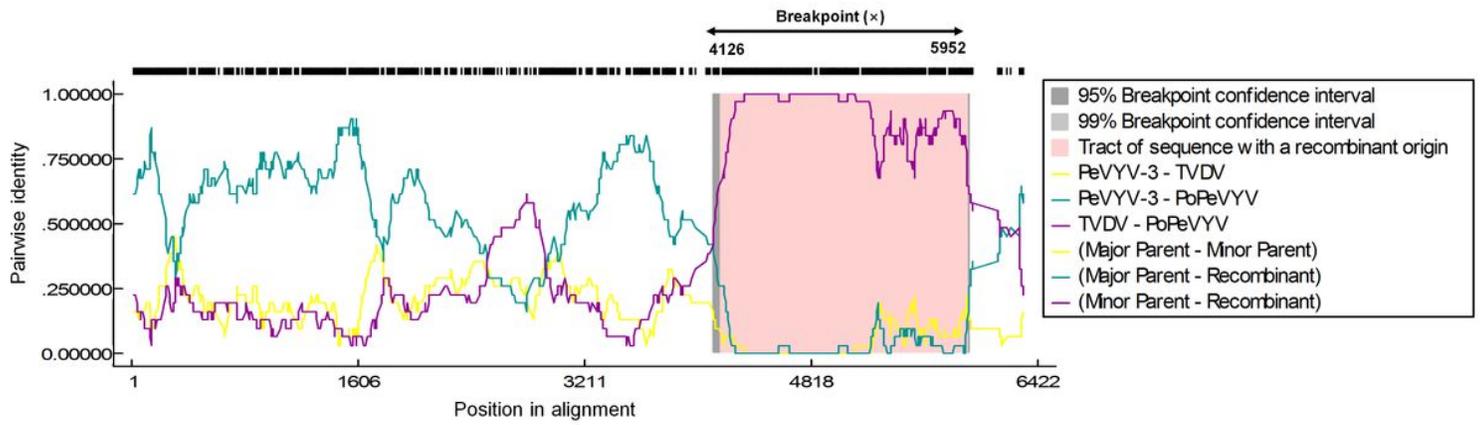


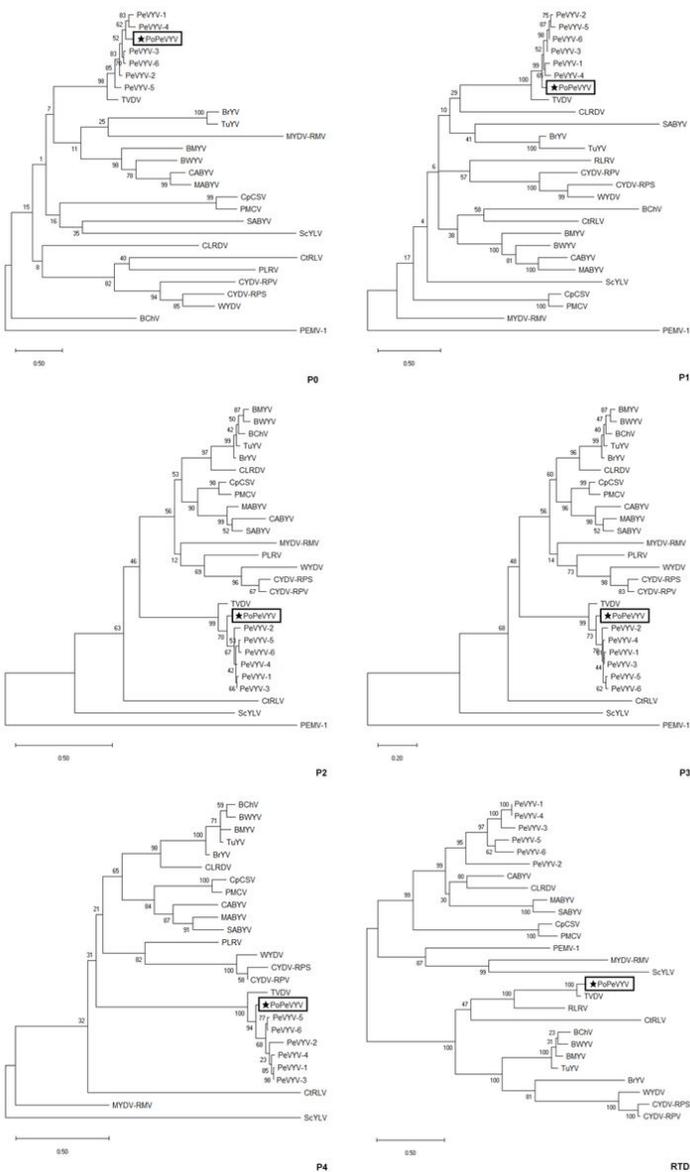
Figure 1

(A) Genome organization PoPeVYV. P1-P2 fusion protein is expressed as a fusion protein with P1 through a translational -1 frameshift, and encodes the RdRp (nts 175-1,620, 1,620-3,431); P3a is expressed by initiation at an AUA codon located at position 3,513; P3-P5 protein is translated via a readthrough of the termination codon at the end of P3 (nts 3,631-4,249, 4,252-5,790). (B-C) Diagram showing how the full-length sequence was constructed by overlap extension RT-PCR.



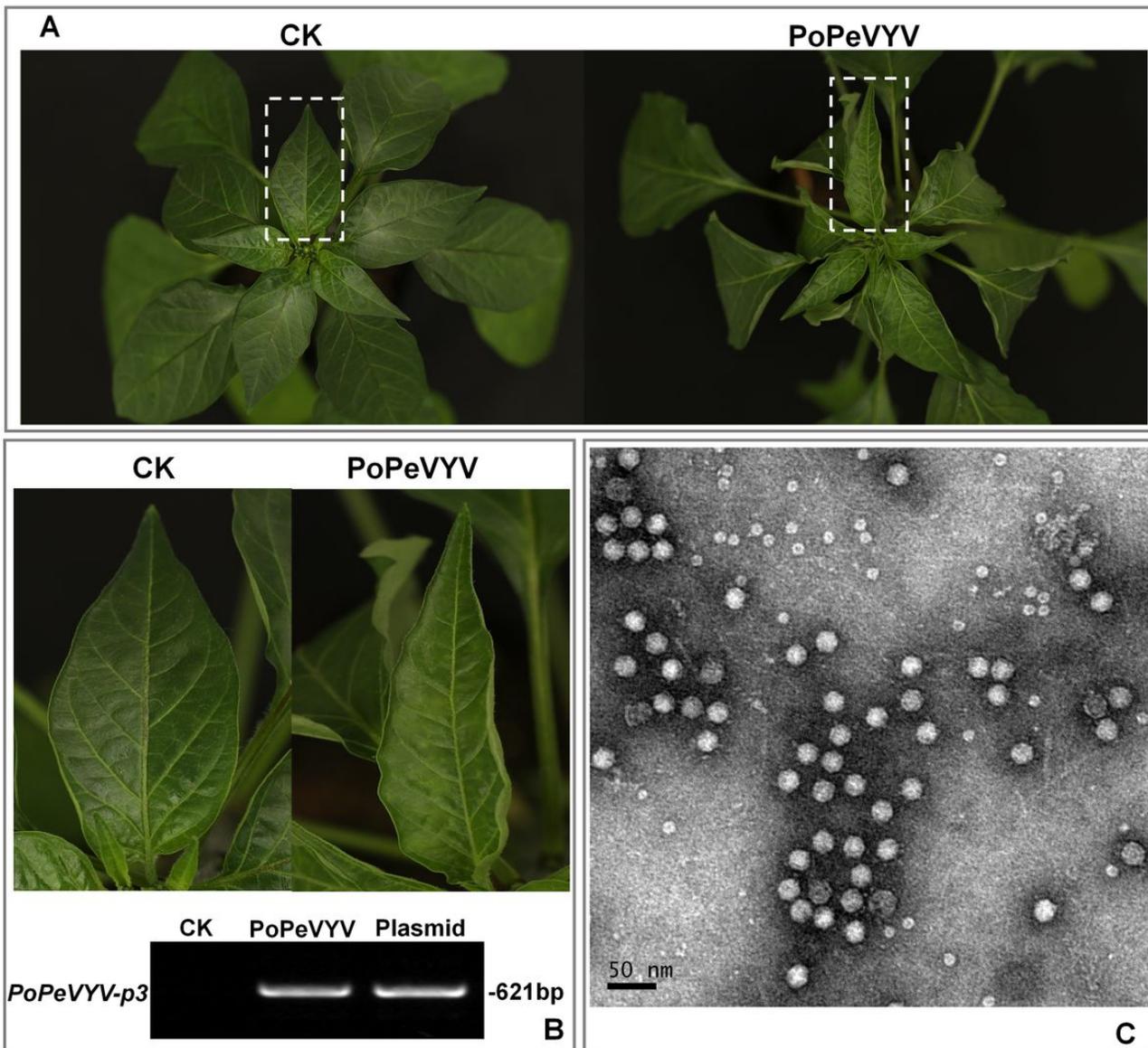
**Figure 2**

Recombination (RDP) and genetic map of PoPeVYV showing major (PeVYV-3) and minor (TVDV) parents and the transition in the genome sequence at the breakpoint (X; 4126-5952 bp); the recombinant region is highlighted in pink. Multiple nucleotide sequences were aligned using MUSCLE, and then analyzed using RDP4 with default parameters.



**Figure 3**

Maximum Likelihood phylogenetic trees constructed using MEGA X, showing the relationship of PoPeVYV to other members of the family Luteoviridae using amino acid sequences of the different gene products. Except for P4, the enamovirus Pea enation mosaic virus 1 (PEMV-1, NC\_003629.1) was used as an outgroup. Numbers on branches are bootstrap support values (1,000 replicates). The models employed were LG+G for P0, JTT+G for P1, LG+G+F for P2, JTT+G for P3, JTT+G+I for P4 and WAG+G for RTD, respectively. The virus abbreviations and accession numbers are as follows: BChV (Beet chlorosis virus, AF352024), BMYV (Beet mild yellowing virus, DQ132996), BrYV (Brassica yellows virus, KF015269), BWYV (Beet western yellows virus, AF473561), CABYV (Cucurbit aphid-borne yellows virus, JQ700305), CLRDV (Cotton leafroll dwarf virus, NC\_014545), CpCSV (Chickpea chlorotic stunt virus, AY956384), CtRLV (Carrot red leaf virus, AY695933), CYDV-RPS (Cereal yellow dwarf virus-RPS, AF235168), CYDV-RPV (Cereal yellow dwarf virus-RPV, EF521927), MABYV (Melon aphid-borne yellows, JQ700307), MYDV-RMV (Maize yellow dwarf virus-RMV, NC\_021484), PeVYV-1 (Pepper vein yellows virus-1, AB594828), PeVYV-2 (Pepper vein yellows virus-2, HM439608), PeVYV-3 (Pepper vein yellows virus-3, KP300822), PeVYV-4 (Pepper vein yellows virus-4, KU999109), PeVYV-5 (Pepper vein yellows virus-5, KY523072), PeVYV-6 (Pepper vein yellows virus-6, LT559483), PLRV (Potato leafroll virus, AF453390), PMCV (Pea mild chlorosis virus, JF507725), SABYV (Suakwa aphid-borne yellows virus, JQ700308), ScYLV (Sugarcane yellow leaf virus, JF925154), TuYV (Tumip yellows virus, X13063), TVDV (Tobacco vein distorting virus, EF529624), and WYDV (Wheat yellow dwarf virus, FM865413).



**Figure 4**

Symptoms caused by PoPeVYV infectious clone in *C. frutescens* (A) Phenotype of *C. frutescens* plants agroinfiltrated with PoPeVYV infectious clone or empty agrobacterium (CK) 45 days post infiltration. (B) RT-PCR confirming the presence of viral RNA in systemic leaves of inoculated plants. A 621-bp fragment of the p3 was amplified, with a plasmid containing the p3 as a positive control. (C) Virions purified from leaves infected by the PoPeVYV infectious clone and observed by TEM.

## Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [8.Fig.S1.jpg](#)
- [9.Fig.S2.jpg](#)
- [10.Fig.S3.jpg](#)
- [11.Fig.S4.jpg](#)
- [12.Fig.S5.jpg](#)
- [3.TableS1.docx](#)
- [13.RDP4files.rar](#)
- [14.PoPeVYVMT188667.txt](#)