

Long Non-Coding RNA H19 Expression and Functional Polymorphism rs217727 Are Linked to Increased Ischemic Stroke Risk

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Abstract

Background: Efforts to identify potential biomarkers for the diagnosis of ischemic stroke (IS) are valuable. The H19 gene plays a functional role in increasing the prevalence of IS risk factors. We evaluated the correlation between H19 rs217727 polymorphism and the expression level of H19 lncRNA with susceptibility to IS among the Iranian population.

Methods: Blood samples were collected from IS patients (n = 114) and controls (n = 114). We concentrated on the expression pattern of H19 at different time points (i.e., 0-24, 24-48, and 48-72 hours after stroke). The tetra-primer ARMS-PCR method was applied for DNA genotyping. We used the quantitative real-time PCR to evaluate H19 expression levels.

Results: The rs217727 polymorphism of H19 was related with IS susceptibility in the co-dominant (OR=2.92, 95% CI=0.91-10.92, P=0.04) and recessive models (OR=2.80, 95% CI=0.96-8.15, P=0.04). H19 expression was significantly upregulated in IS and remained high for 72 hours after stroke. ROC curves showed that H19 expression within the first 24 hours from stroke onset might serve as a biomarker for the early diagnosis of IS with 79.49% sensitivity and 80.00% specificity. H19 expression in SVO and LAA patients were 3.74 and 3.34 times higher than the UD subtype, respectively [OR=3.74 95% CL (1.14-12.27) P=0.030 and OR=3.34 95% CL (1.13-9.85) P=0.029].

Conclusion: The rs217727 polymorphism of the H19 is correlated with IS susceptibility, and H19 expression levels were higher in SVO and LAA patients. The upregulation of H19 may be considered as a diagnostic biomarker in IS among the Iranian population.

Background

Stroke is one of the leading causes of death globally (accounting for 5.5 million deaths annually). The high proportion of stroke patients are young adults in Iran, with a higher mortality rate compared to Western countries [1]. Ischemic stroke (IS) accounts for 75% of all stroke cases. IS patients often suffer from disability and loss of productivity, which can cause a heavy socioeconomic burden [2]. Based on different molecular studies, the clarification of genetic biology, and epigenetic can facilitate the prediction and diagnosis of severe strokes [3].

Long non-coding RNAs (lncRNAs) are RNAs that have more than 200 nucleotides and lack protein-coding capacity. Documents have increasingly revealed that dysregulated lncRNA expression levels can lead to various human diseases [4–6]. lncRNAs are highly expressed in neural cells, and there is a strong likelihood that they are involved in brain dysfunction [7–9]. Recent research works have shown that the expression of selected lncRNA changes over time after IS is detected in the blood of patients [10–12]. H19 lncRNA is the primary transcript of H19 locus; after birth, the abnormal expression of lncRNA H19 was presented as a potential diagnostic biomarker in different cancers [13]. Additionally, lncRNA H19 is typically re-expressed in atherosclerotic plaque [13], and it can be activated under hypoxic conditions [14].

LncRNAs H19 has been revealed to affect the pathophysiology of IS by promoting neuroinflammation [15], preventing neurogenesis [16], and increasing the risk of atherosclerosis [17]. Notably, previous studies have suggested that the genetic polymorphisms on lncRNA influence lncRNA expression and, thus, affect disease susceptibility [18, 19]. Gao et al. (2014) reported that rs217727 polymorphism in the H19 gene was correlated with an increased risk of coronary artery disease [20]. Moreover, Ghaedi et al. demonstrated an association between H19 rs217727 polymorphism with type 2 diabetes susceptibility [21]. Coronary artery disease and diabetes are two main risk factors for IS [22, 23]. Zhu et al. noted that H19 rs217727 gene polymorphism can serve as a marker for IS in the northern Chinese population [24]. On the other hand, Haung et al. (2019) reported that the rs217727 polymorphism was not correlated with IS in the southern Chinese population [25]. Perhaps the main reason for this inconsistency is that the genetic heterogeneity in different areas. Thus, more studies are needed to explain the association between rs217727 polymorphism and IS risk in the populations of different countries.

Previous studies suggested that rs217727 polymorphism is related to the pathogenesis of breast cancer, oral squamous cell carcinoma, and type 2 diabetes susceptibility in the Iranian population [21, 26, 27]. Research on the role of lncRNAs in IS pathogenesis is just beginning, and we have performed this study to assess the association between lncRNA H19 expression and its rs217727 polymorphism with susceptibility to IS in the Iranian population. Especially given the importance of the alteration in lncRNA expression after IS over time, we compared the change in the expression level of lncRNA H19 in three different times (0–24, 24–48, 48–72 hours after the onset of symptoms). The importance of this study becomes more apparent due to the high prevalence of IS in the Iranian population [28]. Therefore, our effort to identify a potential diagnostic marker for IS is valuable. In the current study, the diagnostic potential of lncRNA H19 level as a biomarker in IS patients was evaluated by ROC curve analysis and were compared at different times after stroke.

Material And Methods

Study subjects

This case-control study was conducted from August 2018 to August 2019 at Namazi Hospital in Shiraz, a high-volume referral center in the south of Iran. According to the Recognition of Stroke in the Emergency Room (ROSIER) scale, IS was screened as a focal neurological deficit that persisted beyond 24 hours in surviving patients. Then, radiological confirmation was performed via a brain CT or MRI. IS was defined according to international guidelines [29]. Patients with IS aged above 18 years who filled the informed consent were included in the study. Exclusion criteria were primary intracranial hemorrhage, arterial dissection, cerebral vasculitis, cerebral venous thrombosis, hypoperfusion syndromes, iatrogenic stroke, thrombosed intracranial aneurysm, meningitis, thrombotic thrombocytopenic purpura-hemolytic uremic syndrome, Moyamoya disease, cerebral vasoconstriction syndrome, and malignancy. Control participants were a representative sample of the Shiraz population who were randomly selected from neighbor controls at the nearest from the case's place, sex, and age frequency matched with cases. The examined

subjects were aged 32–90 years. Controls with a history of stroke, transient ischemic attack, illness or disorder of the brain, or malignancy were excluded.

The National Institutes of Health Stroke Scale (NIHSS) score was assessed upon admission. Higher scores indicated increased severity. Ethics approval for this research was obtained by the local ethics committee of Kazeroon Branch, Islamic Azad University with the number 97 01 94 -19386.

Collection Of Blood Samples

After obtaining informed consent, peripheral venous blood samples were collected from IS patients 0–24 (n = 39), 24–48 (n = 38), and 48–72 (n = 37) hours after the onset of symptoms, and control subjects in ethylene diamine tetra acetic acid-coated (EDTA) tubes as an anticoagulant.

Snp Genotyping And Measurement Of The Lncrna H19 Levels

DNA was isolated from the blood samples using the DNA Extraction Kit (Favorgen, Taiwan).

To evaluate the purity and concentration of the isolated DNA, a photo nanometer was applied at 230, 260, and 280 nm ed. Polymorphism has been evaluated by a tetra-amplification refractory mutation system-polymerase chain reaction (T-ARMS-PCR) as described previously by Ghapanchi et al. (2020) [30]. Using the Tera-ARMS PCR method in genomic research allows for rapid SNP identification in a reliable (accuracy of 99.9%), and low-cost manner [31]. Sanger sequencing was done on approximately 20% of the random samples and demonstrated no genotyping error.

Total RNA was extracted from fresh blood samples using a total RNA extraction kit (Favorgen, Taiwan) following the manufacturer's instructions. To remove DNA contamination, the isolated RNA was treated with DNase I enzyme. RNA samples were used for cDNA synthesis using the cDNA synthesis Kit (Yektatajhiz, Iran). The relative abundance of the H19 level was measured by quantitative real-time PCR using RealQ Plus 2x Master Mix Green Low ROX™ (Ampliqon, Denmark). Here, we used the Quantstudio 3 Real-Time PCR System (Applied Biosystems, Foster City, USA) with the thermal-cycling settings of 10 min at 95 °C (1 repeat) accompanied by 40 cycles for 15 s at 95 °C and 1 min at 60 °C. Every complete amplification phase was accompanied by a melting phase, for 15 s at 95°C, 30 s at 60°C, and 15 s at 95 C. The following primers were used: for the H19 5'- GCAGACAGTACAGCATCCA – 3' (forward) and 5'- CTCCTGAGAGCTCATTCCTC – 3' (reverse); for the TATA-Binding Protein (TBP, reference gene), 5'- CCCGAAACGCCGAATATAATC-3' (forward) and 5'-TCTGGACTGTTCTTCACTCTTG-3' (reverse).

To normalize the target gene expression, the *TBP* gene was selected. $2^{-\Delta CT}$ was used to calculate the H19 comparative expression level for each individual [32].

Statistical analysis

We evaluated the Hardy-Weinberg equilibrium (HWE) by using a chi-squared (χ^2) test to compare the expected genotype frequencies with the observed ones among the controls. Further, used were unconditional logistic regression analyses to test each SNP's relationship with case/control status under different genetic models, such as dominant, co-dominant, recessive, and over-dominant. Odds ratios (ORs) and their corresponding 95% confidence intervals (95% CI) were scored to assess the strength of association between the risk of IS and H19 polymorphisms. The OR of the H19 expression level was estimated in univariate and multiple logistic regression analyses. Using a multiple logistic regression model, we adjusted for all predictors with ≤ 0.3 of the univariate logistic regression analysis and the most important clinical factors. Comparisons were made between categorical data was assessed by using a chi-square test. The differences between numeric variables were evaluated using an independent two-sample t-test. Correlations were analyzed using the Spearman correlation. Receiver operating characteristic (ROC) curve analysis was carried out to assess whether lncRNAs is a valid biomarker for differentiating IS patients from healthy patients. The analyses were done using the version 19.0 of SPSS software and GraphPad Prism 5.0. A P-value of < 0.05 was regarded as statistically significant.

Results

Clinical characteristics of IS patients and controls

There was no statistically significant difference between the IS patients and the healthy control patients with respect to age, sex, and BMI ($P > 0.05$, Table 1). However, IS patients' levels of hypertension and smoking ($P < 0.001$) diabetes ($P < 0.003$), drinking ($P < 0.005$), and dyslipidemia ($P < 0.026$) were significantly higher than those of the controls (Table 1).

Table 1
Clinical characteristics of study subjects.

Characteristics	Cases (n = 114)	Controls (n = 114)	<i>p</i>
Male, n (%)	68 (59.64)	68 (59.64)	0.999 ^a
Female, n (%)	46 (40.35)	46 (40.35)	
Age (years)	65.88 ± 14.44	65.88 ± 14.44	0.999 ^b
BMI (kg/m ²)	25.80 ± 5.21	26.21 ± 4.51	0.523
NIHSS at admission	63 (55.26)	-	
≤ 6	51 (44.73)		
≥ 7			
Toast of classification cases n (%)			
LAA	34(29.82)		
SVO	26(22.8)		
CE	26(22.8)		
UD	28(24.5)		
Vascular risk factors			
Hypertension, n (%)	65 (57.02)	31 (27.20)	< 0.001 ^a
Diabetes, n (%)	35 (30.7)	16 (14.04)	0.003 ^a
Smoking, n (%)	34 (29.82)	8 (7.02)	< 0.001 ^a
Drinking, n (%)	16 (14.03)	3 (2.63)	0.005 ^a
Dyslipidemia	39 (34.21)	24(21.05)	0.026 ^a
NIHSS: National institutes of health stroke scale; LAA: large-artery atherosclerosis; SVO: small vessel occlusion CE: cardio embolism; and UD: undetermined etiology; BMI: Body Mass Index			
Data were shown as mean ± standard deviation (SD) or as n (%). ^a Chi-square Test; ^b Independent two-sample T-test. P-value of < 0.05 was regarded as statistically significant			

H19 rs217727 polymorphism has a positive association with the risk of IS

We did not find any significant deviations from the Hardy-Weinberg equilibrium for rs217727 polymorphism in the control group. The results confirmed the association between the IS and rs217727 polymorphism in Iranian individuals. We found that the C allele of rs217727 was a significant protective factor against IS in the Iranian populations (C vs. T: OR = 0.62, 95% CI: 0.39–0.99, P = 0.04).

Furthermore, the frequency of TT genotypes of H19 rs217727 was significantly higher in IS patients than in healthy controls (11.4 vs. 4.38%). Moreover, the TT genotype was associated with a 2.92-fold increase in IS risk in the co-dominant model (OR = 2.92, 95% CI = 0.91–10.92, P = 0.04). Also, H19 rs217727 polymorphism was related to a 2.80-fold increase in the risk of IS in the recessive model (TT vs. CC + CT genotypes) (Table 2).

Table 2
Allele and genotype frequencies of *H19* rs217727 for IS patients and the control group.

Inheritance model	rs217727 Polymorphism	Cases (%)	Controls (%)	OR (95% CI)	P
Codominant ^a	Genotype				
	CC	73 (64.03)	82 (71.92)	1	
	CT	28 (24.56)	27 (23.68)	1.16 (0.62–2.15)	0.62
	TT	13 (11.40)	5 (4.38)	2.92 (0.91–10.92)	0.04
Dominant ^b	CC	73 (64.03)	82 (71.92)	1	
	CT + TT	41 (35.96)	32 (28.07)	1.43 (0.82–2.51)	0.20
Recessive ^c	CC + CT	101 (88.59)	109 (95.61)	1	
	TT	13 (11.40)	5 (4.38)	2.80 (0.96–8.15)	0.04
Over-dominant ^d	CC + TT	86 (75.43)	87 (76.31)	1	
	CT	28 (24.56)	27 (23.68)	1.04 (0.57–1.92)	0.87
	Allele				
	T	54 (23.68)	37 (16.22)	1	
	C	174 (76.31)	191 (83.77)	0.62 (0.39–0.99)	0.04
OR, odd ratio; CI, confidence interval.					
^a Co-dominant; major allele homozygotes vs. heterozygotes.					
^b Dominant; major allele homozygotes vs. heterozygotes + minor allele homozygotes.					
^c Recessive; major allele homozygotes + heterozygotes vs. minor allele homozygotes.					
^d Over dominant; major allele homozygotes + minor allele homozygotes vs. heterozygotes.					

H19 expression was significantly upregulated in the peripheral blood of IS Patients

We found that H19 expression was significantly upregulated in IS cases when compared to controls (Fig. 1, A, $P < 0.001$). Further analysis indicated that H19 expression was found more often in IS patients than in the controls 0–24, 24–48, and 48–72 hours after the onset of stroke ($P < 0.001$, $P < 0.05$, $P < 0.01$, respectively). An independent t-test was used to compare the relative H19 expression in peripheral blood of IS cases at each time point with age- and sex-matched controls (Figs. 1B-1D). H19 expression levels in IS patients were compared to levels of controls 0–24 hours (4.51 ± 0.88 vs 0.96 ± 0.12), 24–48 hours (7.12 ± 1.53 vs 2.73 ± 1.19), and 48–72 hours (6.38 ± 1.36 vs 1.84 ± 0.36) after stroke. Interestingly, we observed that upregulated H19 remained high 72 hours after a stroke (Fig. 1E).

H19 expression was associated with TOAST subtypes of IS patients

Patients were assigned to four subtypes according to TOAST classification (Table 1). We found a significant elevation in H19 expression in patients with LAA, SVO, and UD when compared to controls (ANOVA, $P < 0.01$). IS patients with CE showed the lowest levels of H19 without a significant difference with healthy controls (Fig. 2) ($P > 0.05$).

Multiple logistic regression analysis was done to detect the association between TOAST subtypes and H19 expression after adjusting for several confounders (Table 3). We showed that H19 expression levels were significantly associated with TOAST subtypes. H19 expression levels in SVO and LAA patients were 3.74 and 3.34 times higher than the UD subtype, respectively [OR = 3.74 95% CL (1.14–12.27) $P = 0.030$ and OR = 3.34 95% CL (1.13–9.85) $P = 0.029$].

Table 3
The results of univariate and multiple logistic regression analyses

Characteristics	Univariate			Multiplevariate *		
	OR	95% CI	P-Value	OR	95% CI	P-Value
Male sex	1.26	0.59–2.67	0.542	1.36	0.59–3.14	0.466
Age (year)	1.00	0.97–1.02	0.835	0.99	0.96–1.02	0.567
Hypertension	1.80	0.85–3.81	0.125	1.82	0.80–4.11	0.151
Diabetes mellitus	1.14	0.52–2.53	0.743	0.99	0.40–2.47	0.988
Dyslipidemia	1.33	0.61–2.89	0.477	1.35	0.56–3.22	0.502
Cigarette Smoking	1.05	0.47–2.35	0.903			
NIHSS score	1.01	0.95–1.07	0.727	1.02	0.96–1.09	0.505
Alcohol use	1.04	0.36–2.99	0.940			
Type of IS UD	1					
CE	0.95	0.31–2.92	0.933	1.24	0.38–4.06	0.725
SVO	2.88	0.95–8.69	0.061	3.74	1.14–12.27	0.030
LAA	2.91	1.03–8.20	0.044	3.34	1.13–9.85	0.029
BMI	1.02	0.94–1.10	0.664			

*The OR was adjusted for all predictors with ≤ 0.3 of the univariate logistic regression analysis and for the most important clinical factors. NIHSS: National institutes of health stroke scale, BMI: Body Mass Index; LAA: large-artery atherosclerosis; SVO: small vessel occlusion CE: cardio embolism; and UD: undetermined etiology. P-value < 0.05 was regarded as statistically significant.

H19 expression levels in different genotypes of rs217727 polymorphism in IS patients

Further analysis revealed that there were no significant differences in relative H19 expression levels among the three groups of rs217727 (CC, CT and TT) genotypes in case ($F = 0.00$; $df = 2$, $P = 0.999$) and control ($F = 0.14$; $df = 2$, $P = 0.865$) (Fig. 3).

H19 expression level in peripheral blood might serve as a diagnostic biomarker

The ROC curve analysis showed that H19 is a potential biomarker for IS diagnosis cases (Table 4). For the period 0–24 hours after IS, AUC was 0.875 ± 0.039 (95% CI, 0.798–0.952; Fig. 4A); the sensitivity and specificity were 79.49% and 80.00%, respectively. For the period 24–48 hours after IS, AUC was 0.813 ± 0.049 (95% CI, 0.715–0.910; Fig. 4B), and the sensitivity and specificity were 77.5% and 75.00%, respectively. For the period 48–72 hours after IS, AUC was 0.788 ± 0.052 (95% CI, 0.685–0.891; Fig. 4C), and the sensitivity and specificity were 75.68% and 67.57%, respectively. Circulating H19 levels within the

first 24 hours after the onset of a stroke indicated high sensitivity and specificity for the early diagnosis of acute stroke.

Table 4
Prediction of diagnostic value

Parameter	AUC ± SE	95% CI	P	Se (%)	Sp (%)
Diagnostic value					
H19 expression in 0–24 hours	0.875 ± 0.039	0.798–0.952	< 0.0001	79.49%	80.00%
H19 expression in 24–48 hours	0.813 ± 0.049	0.715–0.910	< 0.0001	77.5%	75.00%
H19 expression in 48–72 hours	0.788 ± 0.052	0.685–0.891	< 0.0001	75.68%	67.57%
Abbreviations: AUC; Area Under the Curve; CI, Confidence Interval; Se, sensitivity; Sp, specificity P < 0.05 was considered statistically significant					

Discussion

This research showed the effect of H19 gene polymorphism (rs217727) on IS susceptibility through a case-control study on individuals from southwestern Iran. The findings demonstrated that rs217727 polymorphism was significantly associated with the risk of IS in the co-dominant and recessive models. However, it did not correlate with H19 expression levels in the peripheral blood of Iranian patients. In addition, we observed a significant increase in the lncRNA H19 expression levels of IS patients when compared to healthy controls. This result is consistent with previous studies that reported the upregulation of H19 in the peripheral blood of IS patients [25, 33, 34]. We evaluated H19 expression at three different times (0–24 hours [n = 39], 24–48 hours [n = 38], and 48–72 hours [n = 37] after stroke). We demonstrated that lncRNA H19 expression significantly increased in IS patients and remained high for 72 hours after a stroke. Xiao et al. reported a high expression level of H19 in 40 IS patients within the first three hours after a stroke. This finding was in line with the results of Wang et al. and Haung et al., whose results indicated high levels of lncRNA H19 for the first 24 hours after IS stroke (in 36 and 64 IS patients, respectively) [15, 25].

The results obtained from in vitro and in vivo experiments showed that H19 was upregulated following ischemic reperfusion injuries in the ischemic penumbra of animals, as well as in the neuroblastoma cell after oxygen-glucose deprivation and reperfusion (OGD/R)-induced injury [33, 35, 36]. We have little information about the basic mechanism of lncRNA H19 in IS. Gao et al. (2020) reported that lncRNA H19 could aggravate oxidative stress and apoptosis in I/R tissues and OGD/R-induced cells via a PTEN/Akt signaling pathway, whereas H19 was overexpressed and miR-19a-3p was down-regulated [36]. Recently, Xiao et al. reported neuronal apoptosis (both in vitro and in vivo) was reduced by the inhibition of H19 with siRNA. Also, the inhibition of H19 in ischemic rats significantly decreased brain infarct size, neurological deficit, and neuronal apoptosis [33]. Wang et al. demonstrated that lncRNA H19 caused by activation of autophagy induces cerebral ischemia reperfusion damage [35]. During cerebral ischemia

reperfusion, autophagosome acts as a protective mechanism since it can recover the energy and materials by clean up the damaged organelles [37]. However, excessive autophagy activation plays an important role in the induction of apoptosis, necrosis, and the autophagic death of neurons following cerebral I/R [38]. Furthermore, lncRNA H19 has a critical role in atherosclerosis-related pathophysiology and the development of atherosclerosis [39]. In this regard, Haung et al. reported that the expression of acid phosphatase 5 protein is promoted by the H19 gene. This protein contributes to atherosclerosis and increases the risk of IS [17].

In our patients, the levels of upregulated H19 expression remained high until 72 hours after the onset of symptoms. It may take more time to recover to normal levels. The maximum recovery in the expression of different lncRNAs after stroke can take up to seven days [12]. Another important finding was that circulating H19 levels were positively associated with LAA and SVO subtypes. Atherosclerosis is a predictable risk factor for LAA and SVO subtypes [40]. This is consistent with another observation that showed that H19 is correlated with LAA subtypes of atherosclerotic patients [17]. Many investigations have demonstrated that H19 is involved in the native atherosclerosis via different mechanisms, such as cellular proliferation and apoptosis, adipocyte differentiation, inflammatory response, lipid metabolism, and regulation of angiogenesis [39]. Recent studies have shown that H19 might play a critical role in the onset and progression of atherosclerosis [25, 41]. We found that patients with the CE subtype had the lowest levels of H19 expression. This finding was also reported by Huang et al. (2019) [17].

In the present investigation, lncRNA H19 expression was not positively correlated with NIHSS scores. This outcome is contrary to the findings of Xiao et al. (2019), who found a positive correlation between NIHSS scores and H19 expression three hours after stroke onset [33]. It could be argued that the observed positive association by Xiao et al. was due to blood sampling within the first 3 hours. Meanwhile, in our study, blood samples were collected at three different times. The main reason for this discrepancy can be explained by the assumption that there are optimal blood sampling time windows for estimating a positive association between elevated circulating H19 and NIHSS scores.

A valuable method for early screening of IS is developing a high-sensitive noninvasive blood biomarker. Evidence shows that circulating lncRNAs could be considered an IS biomarker [42, 43]. Elevated H19 expression has been reported in various oncological disorders [13, 27]. Therefore, it can be used as a non-specific biomarker for IS. Subjects with known oncological diseases were excluded from the present study. The current study showed that circulating H19 levels within the first 24 hours after the onset of a stroke have a sensitivity of 79.4% and a specificity of 80%. Therefore, H19 expression could serve as a potential biomarker for the early diagnosis of acute stroke; after 24 hours, its sensitivity and specificity decreased over time.

We showed that the TT genotype of the rs217727 polymorphism was significantly correlated with increased risk of IS. This is in line with the results of Zhu et al. and Wang et al [24, 35]. However, the expression levels of H19 in different genotypes of the rs217727 polymorphism did not show a significant difference. Probably, the rs217727 polymorphism affects the lncRNA H19 expression within the first few

hours after the onset of IS. Therefore, the current study could not detect this issue, or perhaps mutations changed the translational efficiency, thereby leading to changes in lncRNA H19 structure and expression.

In this study, all patients were chosen consecutively from hospitals during the same period with various ethnic groups, and so selection bias could not be avoided. More accurate findings could be observed by using larger sample sizes and by taking blood samples during the first few hours after a stroke to determine whether H19 SNP affects H19 expression and whether NIHSS has a significant correlation with H19 expression level. Although the current study indicates that circulating H19 can be a diagnostic biomarker for IS, further research is required to find reliable blood biomarkers for clinical use.

Conclusion

This study demonstrated that the circulating H19 in the first 24 hours after a stroke might be a diagnostic biomarker for IS. Also, rs217727 polymorphism in the co-dominant and recessive models could be significantly associated with IS risk among the Iranian population. H19 expression was associated with TOAST subtypes in IS patients. However, further large-scale studies are required to confirm our findings and validate the clinical application of this biomarker for IS diagnosis.

Abbreviations

AUC

Area under the curve; BMI:Body mass index; CE:Cardio embolism; CI:Confidence interval; EDTA:Ethylene diamine tetra acetic acid; HWE:Hardy-Weinberg equilibrium; IS:Ischemic stroke; LAA:Large-artery atherosclerosis; lncRNA:Long non coding- RNA; NIHSS:National institutes of health stroke scale; OGD/R:Oxygen-glucose deprivation and reperfusion; OR:Odds ratio; ROC:Receiver operating characteristic; ROSIER:Recognition of stroke in the emergency room; Se:Sensitivity; SD:Standard deviation; Sp:Specificity; SVO:Small vessel occlusion; T-ARMS-PCR:Tetra-amplification refractory mutation system-polymerase chain reaction; TBP:TATA-binding protein; UD:Undetermined etiology.

Declarations

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Authors' contributions

TA- study concept and design, acquisition of data.

MJ- study concept and design, acquisition of data, analysis and interpretation of data, study supervision, drafting/revising the manuscript for content.

MB- study concept and design, acquisition of data drafting/revising the manuscript for content.

AS- study concept and design, revising the manuscript for content.

MD- study concept and design.

RT- analysis and interpretation of data.

AB- study concept and design, acquisition of data, study supervision, drafting/ revising the manuscript for content

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Availability of data and materials

All data analyzed during this study are included in this published article.

Ethics approval and consent to participate

Ethics approval for this research was obtained by the local ethics committee of Kazeroon Branch, Islamic Azad University with the number 97 01 94 -19386.

Consent for publication

Not applicable.

Competing interests

The authors declare that this research was performed in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. Borhani-Haghighi A, Safari R, Heydari ST, Soleimani F, Sharifian M, Kashkuli SY, et al. Hospital mortality associated with stroke in southern Iran. *Iranian journal of medical sciences*. 2013;38 4:314.
2. Ren W, Yang X. Pathophysiology of long non-coding RNAs in ischemic stroke. *Frontiers in molecular neuroscience*. 2018;11:96.
3. Kim SJ, Moon GJ, Bang OY. Biomarkers for stroke. *Journal of stroke*. 2013;15 1:27.
4. López-Urrutia E, Montes LPB, de Guevara Cervantes DL, Pérez-Plasencia C, Campos-Parra AD. Crosstalk between long Non-coding RNAs, Micro-RNAs and mRNAs: Deciphering molecular

- mechanisms of master regulators in cancer. *Frontiers in oncology*. 2019;9.
5. Marques-Rocha JL, Samblas M, Milagro FI, Bressan J, Martinez JA, Marti A. Noncoding RNAs, cytokines, and inflammation-related diseases. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology*. 2015;29 9:3595-611; doi: 10.1096/fj.14-260323.
 6. Tietze L, Kessler SM. The Good, the Bad, the Question-H19 in Hepatocellular Carcinoma. *Cancers*. 2020;12 5; doi: 10.3390/cancers12051261.
 7. Bernard D, Prasanth KV, Tripathi V, Colasse S, Nakamura T, Xuan Z, et al. A long nuclear-retained non-coding RNA regulates synaptogenesis by modulating gene expression. *The EMBO journal*. 2010;29 18:3082-93.
 8. Mercer TR, Dinger ME, Mattick JS. Long non-coding RNAs: insights into functions. *Nature reviews genetics*. 2009;10 3:155-9.
 9. Wapinski O, Chang HY. Long noncoding RNAs and human disease. *Trends in cell biology*. 2011;21 6:354-61.
 10. Xiang Y, Zhang Y, Xia Y, Zhao H, Liu A, Chen Y. LncRNA MEG3 targeting miR-424-5p via MAPK signaling pathway mediates neuronal apoptosis in ischemic stroke. *Aging*. 2020;12 4:3156-74; doi: 10.18632/aging.102790.
 11. Wang Y, Gu XX, Huang HT, Liu CH, Wei YS. A genetic variant in the promoter of lncRNA MALAT1 is related to susceptibility of ischemic stroke. *Lipids in health and disease*. 2020;19 1:57; doi: 10.1186/s12944-020-01236-4.
 12. Zhu W, Tian L, Yue X, Liu J, Fu Y, Yan Y. LncRNA Expression Profiling of Ischemic Stroke During the Transition From the Acute to Subacute Stage. *Frontiers in neurology*. 2019;10:36; doi: 10.3389/fneur.2019.00036.
 13. Sun W, Yang Y, Xu C, Xie Y, Guo J. Roles of long noncoding RNAs in gastric cancer and their clinical applications. *Journal of cancer research and clinical oncology*. 2016;142 11:2231-7; doi: 10.1007/s00432-016-2183-7.
 14. Yoshimizu T, Miroglio A, Ripoche M-A, Gabory A, Vernucci M, Riccio A, et al. The H19 locus acts in vivo as a tumor suppressor. *Proceedings of the National Academy of Sciences*. 2008;105 34:12417-22.
 15. Wang J, Zhao H, Fan Z, Li G, Ma Q, Tao Z, et al. Long noncoding RNA H19 promotes Neuroinflammation in ischemic stroke by driving histone Deacetylase 1-dependent M1 microglial polarization. *Stroke*. 2017;48 8:2211-21.
 16. Wang J, Cao B, Zhao H, Gao Y, Luo Y, Chen Y, et al. Long noncoding RNA H19 prevents neurogenesis in ischemic stroke through p53/Notch1 pathway. *Brain research bulletin*. 2019;150:111-7.
 17. Huang Y, Wang L, Mao Y, Nan G. Long noncoding RNA-H19 contributes to atherosclerosis and induces ischemic stroke via the upregulation of acid phosphatase 5. *Frontiers in neurology*. 2019;10:32.
 18. Shastri BS. SNPs: impact on gene function and phenotype. *Methods Mol Biol*. 2009;578:3-22; doi: 10.1007/978-1-60327-411-1_1.

19. Wapinski O, Chang HY. Long noncoding RNAs and human disease. *Trends Cell Biol.* 2011;21 6:354-61; doi: 10.1016/j.tcb.2011.04.001.
20. Gao W, Zhu M, Wang H, Zhao S, Zhao D, Yang Y, et al. Association of polymorphisms in long non-coding RNA H19 with coronary artery disease risk in a Chinese population. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis.* 2015;772:15-22.
21. Ghaedi H, Zare A, Omrani MD, Doustimotlagh AH, Meshkani R, Alipour S, et al. Genetic variants in long noncoding RNA H19 and MEG3 confer risk of type 2 diabetes in an Iranian population. *Gene.* 2018;675:265-71.
22. Einarson TR, Acs A, Ludwig C, Panton UH. Prevalence of cardiovascular disease in type 2 diabetes: a systematic literature review of scientific evidence from across the world in 2007-2017. *Cardiovascular diabetology.* 2018;17 1:83; doi: 10.1186/s12933-018-0728-6.
23. Battaglini D, Robba C, Lopes da Silva A, Dos Santos Samary C, Leme Silva P, Dal Pizzol F, et al. Brain-heart interaction after acute ischemic stroke. *Critical care.* 2020;24 1:163; doi: 10.1186/s13054-020-02885-8.
24. Zhu R, Liu X, He Z. Long non-coding RNA H19 and MALAT1 gene variants in patients with ischemic stroke in a northern Chinese Han population. *Molecular brain.* 2018;11 1:58.
25. Huang J, Yang J, Li J, Chen Z, Guo X, Huang S, et al. Association of long noncoding RNA H19 polymorphisms with the susceptibility and clinical features of ischemic stroke in southern Chinese Han population. *Metabolic brain disease.* 2019;34 4:1011-21.
26. Ghapanchi J, Ranjbar Z, Mokhtari MJ, Koohpeima F, Derakhshan M, Khademi B, et al. The LncRNA H19 rs217727 Polymorphism Is Associated with Oral Squamous Cell Carcinoma Susceptibility in Iranian Population. *BioMed research international.* 2020;2020:1634252; doi: 10.1155/2020/1634252.
27. Abdollahzadeh S, Ghorbian S. Association of the study between LncRNA-H19 gene polymorphisms with the risk of breast cancer. *Journal of clinical laboratory analysis.* 2019;33 3:e22826; doi: 10.1002/jcla.22826.
28. Daneshfard B, Izadi S, Shariat A, Toudaji MA, Beyzavi Z, Niknam L. Epidemiology of stroke in Shiraz, Iran. *Iranian journal of neurology.* 2015;14 3:158-63.
29. Powers WJ, Rabinstein AA, Ackerson T, Adeoye OM, Bambakidis NC, Becker K, et al. 2018 guidelines for the early management of patients with acute ischemic stroke: a guideline for healthcare professionals from the American Heart Association/American Stroke Association. *stroke.* 2018;49 3:e46-e99.
30. Ghapanchi J, Ranjbar Z, Mokhtari MJ, Koohpeima F, Derakhshan M, Khademi B, et al. The LncRNA H19 rs217727 Polymorphism Is Associated with Oral Squamous Cell Carcinoma Susceptibility in Iranian Population. *BioMed Research International.* 2020;2020.
31. GEORGIANA. DUTA-CORNESCU1 AS-G, NICOLETA. CONSTANTIN1, F. STANCIU1, MICHAELA DOBRE2 DORINA. BANICA3, RODICA. TUDUCE4, P. CRISTEA4,VERONICA. STOIAN1 A comparative study of

- ARMS – PCR and RFLP – PCR as methods for rapid SNP identification. *Romanian Biotechnological Letters*. 2009;14:4845-50
32. Schmittgen TD, Livak KJ. Analyzing real-time PCR data by the comparative C T method. *Nature protocols*. 2008;3 6:1101.
 33. Xiao Z, Qiu Y, Lin Y, Medina R, Zhuang S, Rosenblum JS, et al. Blocking lncRNA H19-miR-19a-Id2 axis attenuates hypoxia/ischemia induced neuronal injury. *Aging*. 2019;11 11:3585-600; doi: 10.18632/aging.101999.
 34. Wang J, Zhao H, Fan Z, Li G, Ma Q, Tao Z, et al. Long Noncoding RNA H19 Promotes Neuroinflammation in Ischemic Stroke by Driving Histone Deacetylase 1-Dependent M1 Microglial Polarization. *Stroke*. 2017;48 8:2211-21; doi: 10.1161/STROKEAHA.117.017387.
 35. Wang J, Cao B, Han D, Sun M, Feng J. Long non-coding RNA H19 induces cerebral ischemia reperfusion injury via activation of autophagy. *Aging and disease*. 2017;8 1:71.
 36. Gao N, Tang H, Gao L, Tu GL, Luo H, Xia Y. LncRNA H19 Aggravates Cerebral Ischemia/Reperfusion Injury by Functioning as a ceRNA for miR-19a-3p to Target PTEN. *Neuroscience*. 2020;437:117-29; doi: 10.1016/j.neuroscience.2020.04.020.
 37. Puyal J, Clarke PG. Targeting autophagy to prevent neonatal stroke damage. *Autophagy*. 2009;5 7:1060-1; doi: 10.4161/auto.5.7.9728.
 38. Wang W, Kang J, Li H, Su J, Wu J, Xu Y, et al. Regulation of endoplasmic reticulum stress in rat cortex by p62/ZIP through the Keap1-Nrf2-ARE signalling pathway after transient focal cerebral ischaemia. *Brain injury*. 2013;27 7-8:924-33; doi: 10.3109/02699052.2013.793397.
 39. Shi X, Wei YT, Li H, Jiang T, Zheng XL, Yin K, et al. Long non-coding RNA H19 in atherosclerosis: what role? *Molecular medicine*. 2020;26 1:72; doi: 10.1186/s10020-020-00196-w.
 40. Bezerra DC, Sharrett AR, Matsushita K, Gottesman RF, Shibata D, Mosley TH, Jr., et al. Risk factors for lacune subtypes in the Atherosclerosis Risk in Communities (ARIC) Study. *Neurology*. 2012;78 2:102-8; doi: 10.1212/WNL.0b013e31823efc42.
 41. Pan JX. LncRNA H19 promotes atherosclerosis by regulating MAPK and NF-κB signaling pathway. *European review for medical and pharmacological sciences*. 2017;21 2:322-8.
 42. Dykstra-Aiello C, Jickling GC, Ander BP, Shroff N, Zhan X, Liu D, et al. Altered expression of long noncoding RNAs in blood after ischemic stroke and proximity to putative stroke risk loci. *Stroke*. 2016;47 12:2896-903.
 43. Deng Q-W, Li S, Wang H, Sun H-L, Zuo L, Gu Z-T, et al. Differential long noncoding RNA expressions in peripheral blood mononuclear cells for detection of acute ischemic stroke. *Clinical Science*. 2018;132 14:1597-614.

Figures

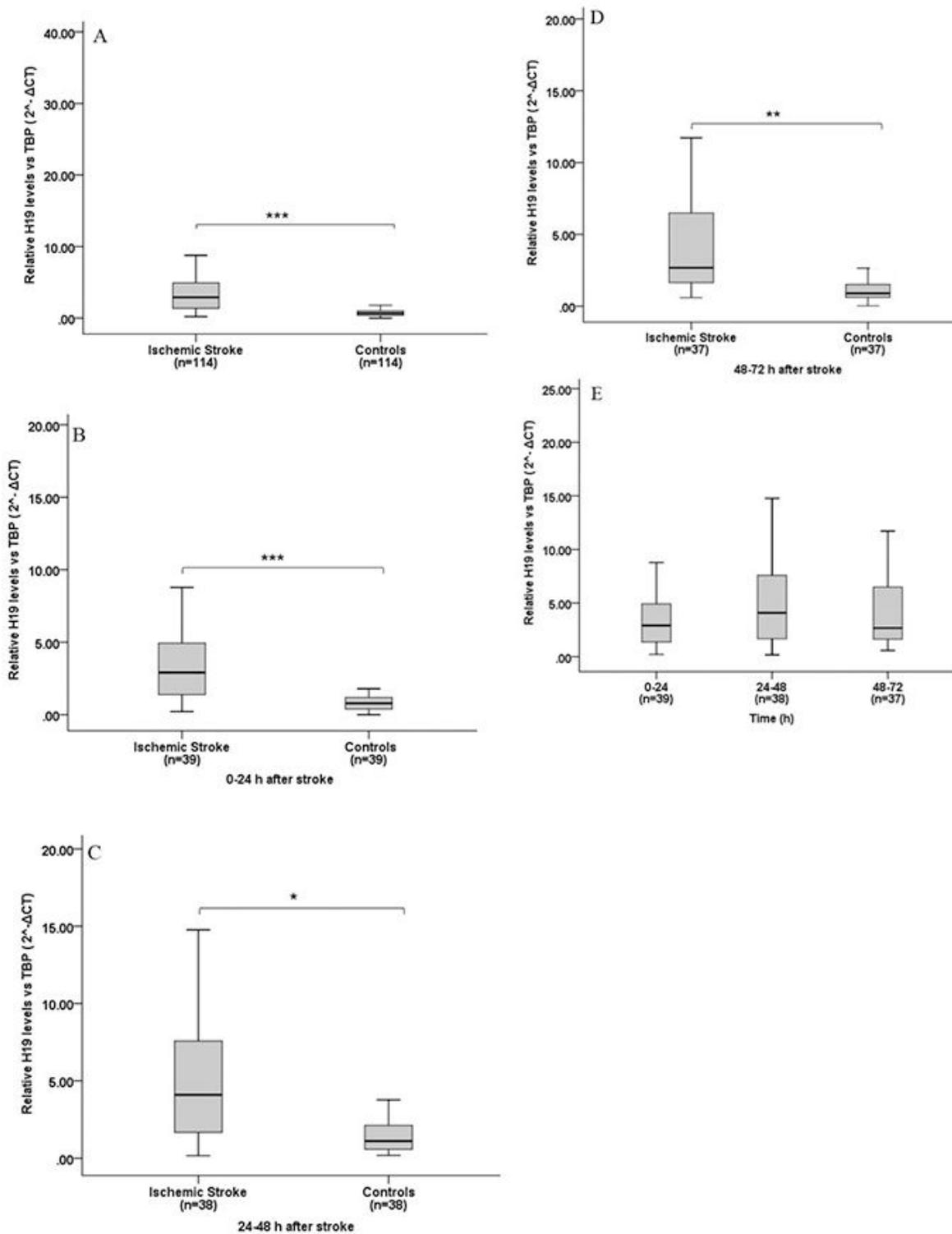


Figure 1

The expression levels of H19 in different time points. H19 expression level in IS was significantly more than those in controls (A). H19 levels in IS patients 0-24 (B), 24-48 (C), 48-72 (D) hours after symptom onset were higher than the controls, while no significant differences were found between 3 time points (E). Results were expressed as mean \pm SE. Significance was set at (*P<0.05, **P<0.01, ***P<0.001).

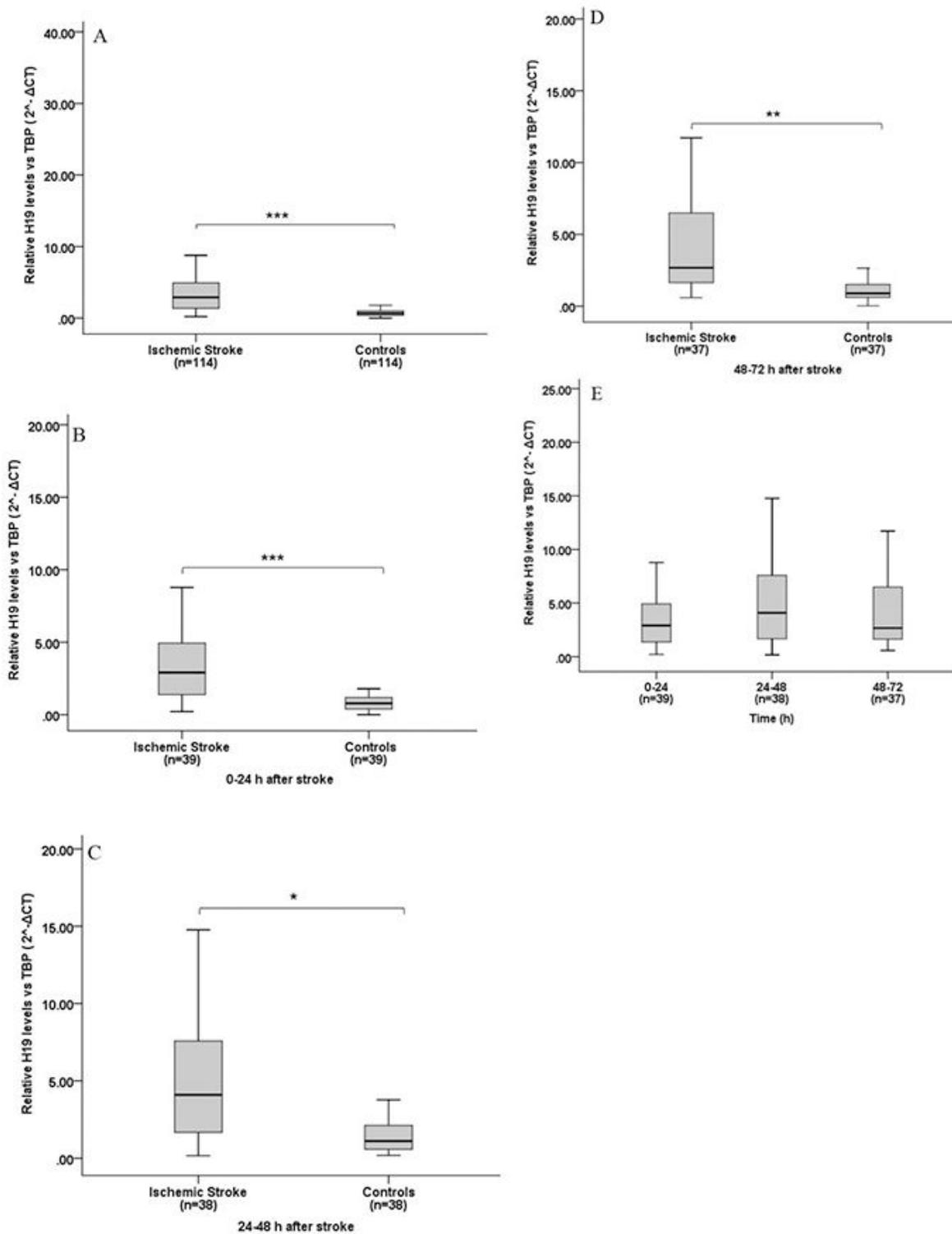


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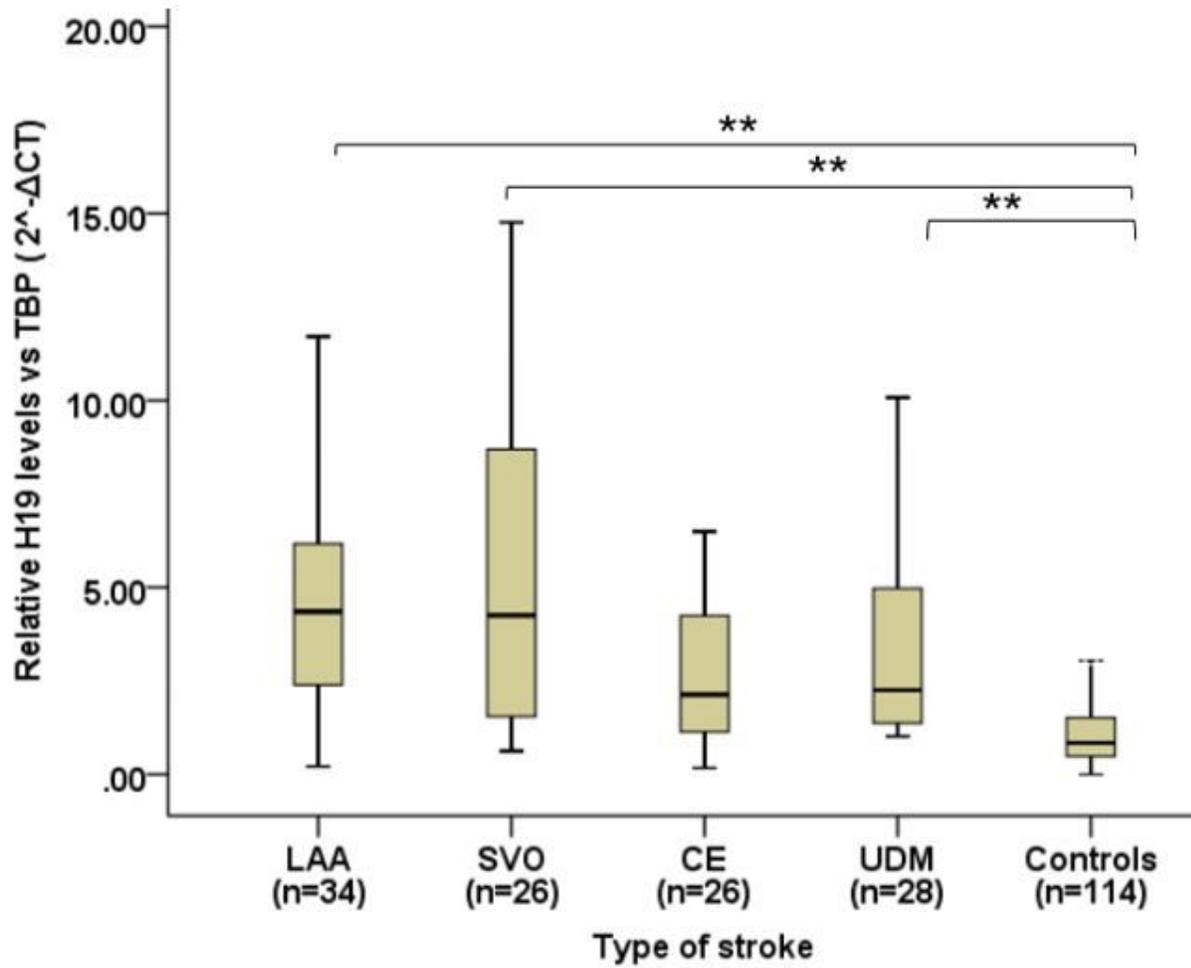


Figure 2

H19 expression was associated with TOAST subtypes of IS patients. H19 levels in LAA, SVO and UD groups were significantly higher than those controls. Data were shown as mean \pm SE. Abbreviations: LAA: large-artery atherosclerosis; SVO: small-vessel occlusion; CE: cardioembolism; UD: undetermined.

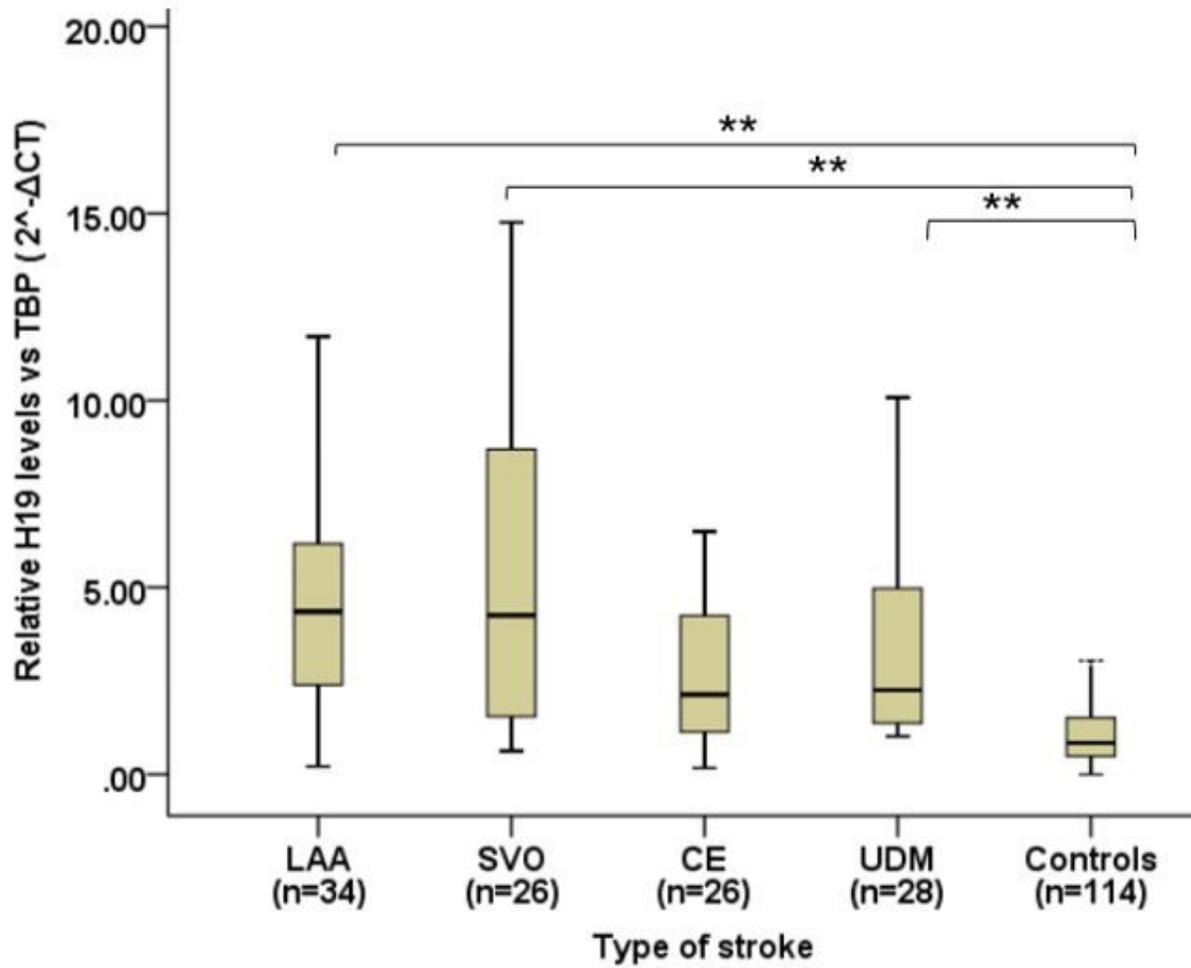


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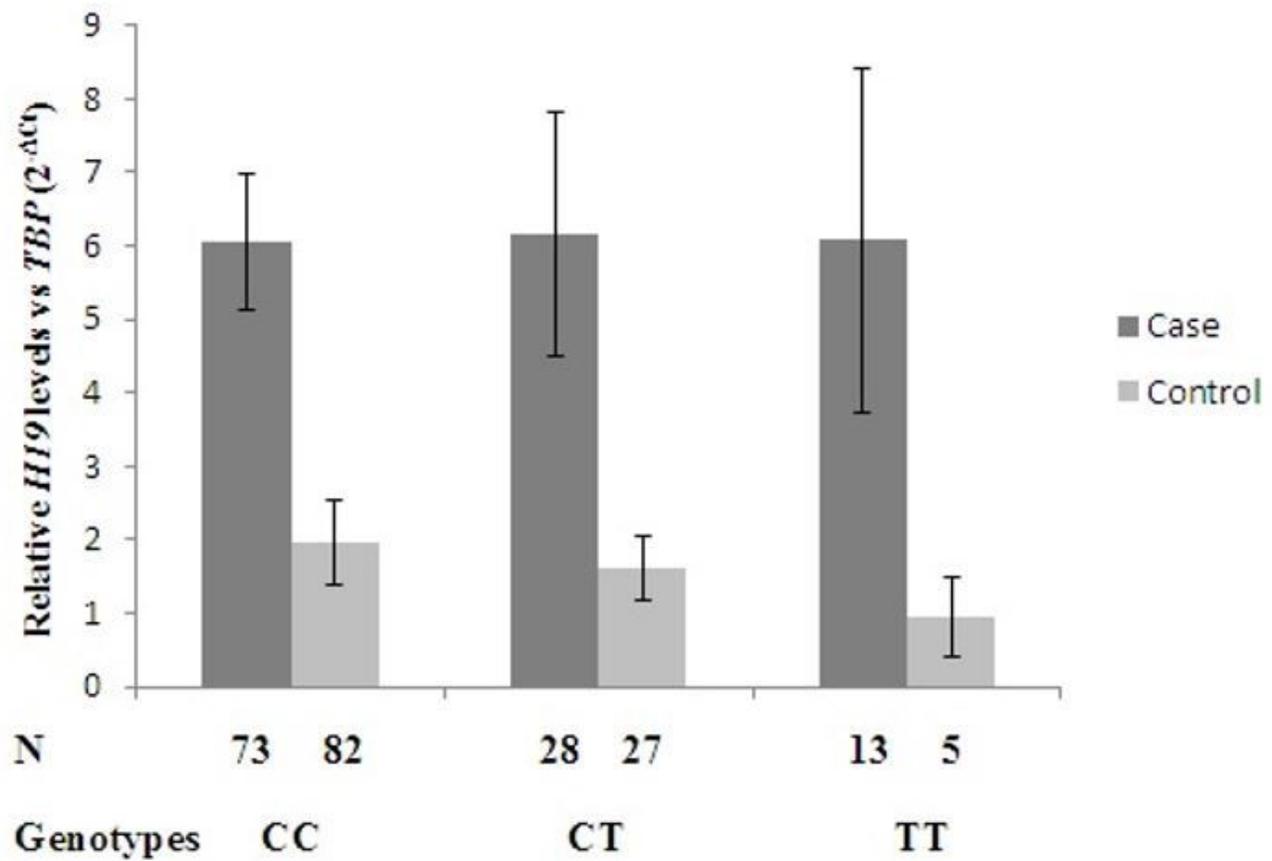


Figure 3

The comparison of relative expression levels of H19 lncRNA between different genotypes of rs217727 polymorphism in cases and controls. Data were shown as mean \pm SE. N shows the number of participants with each genotype.

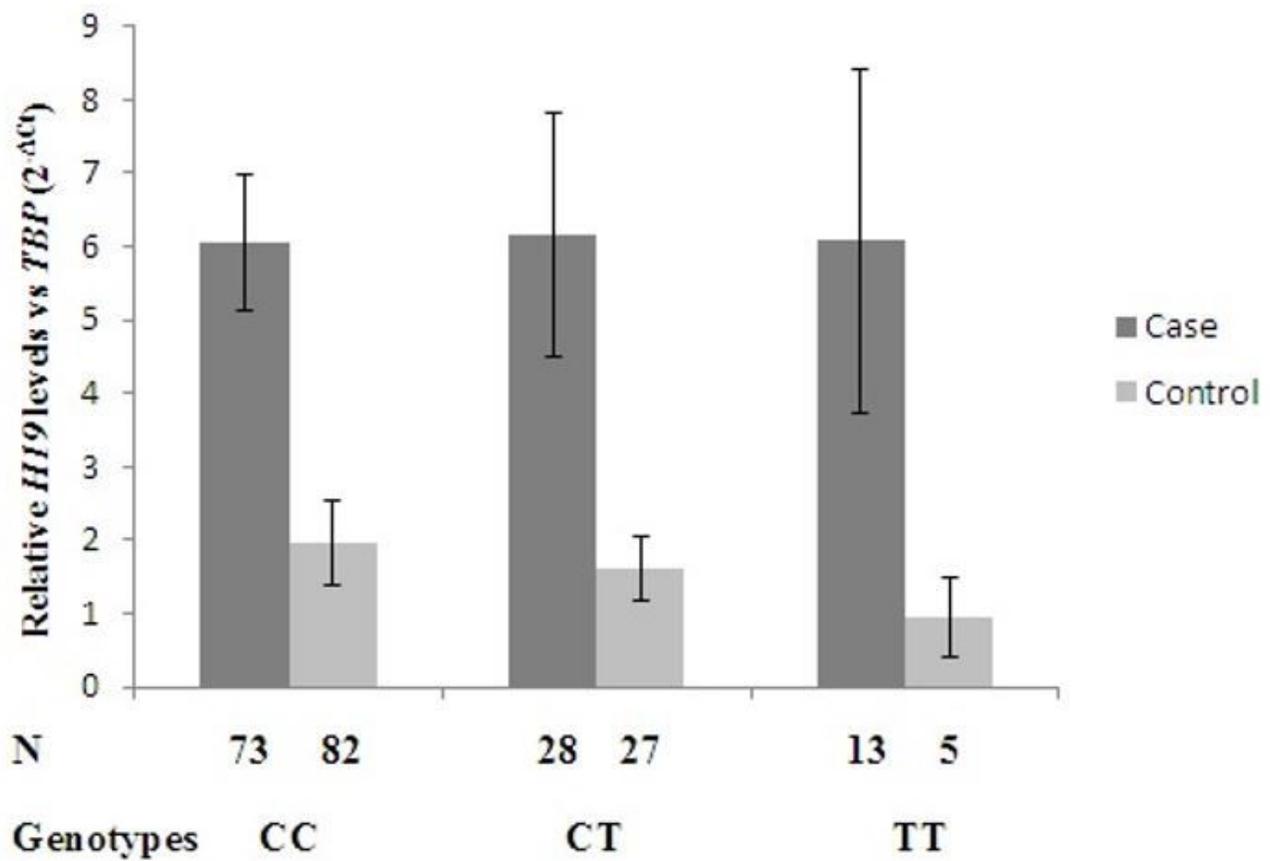


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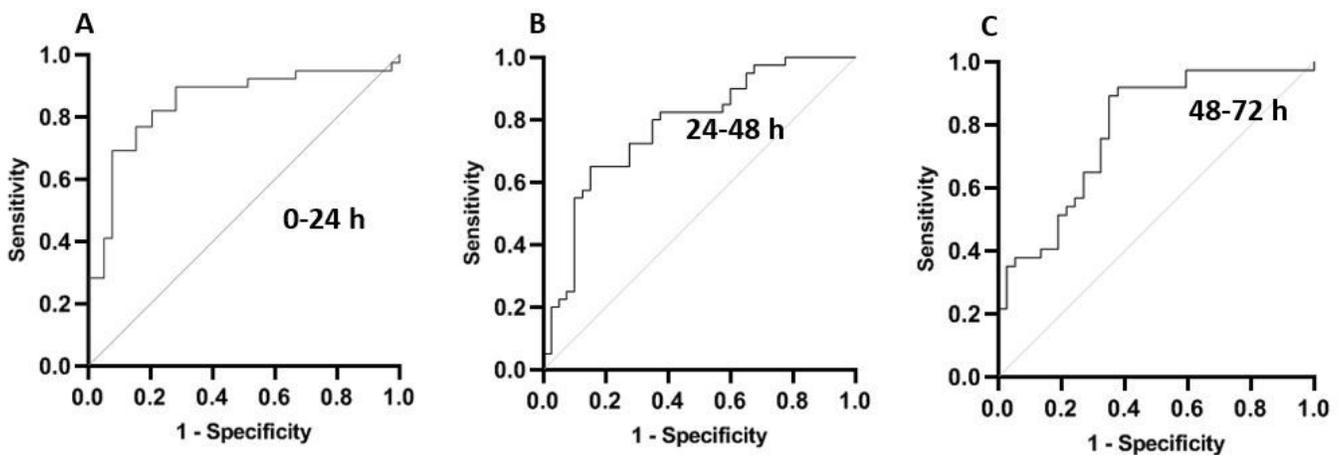


Figure 4

ROC curve analysis showed that H19 was a potential biomarker for IS diagnosis cases from the controls.0- 24 hours (A), 24-48 hours (B), 48-72 hours (C) after IS.

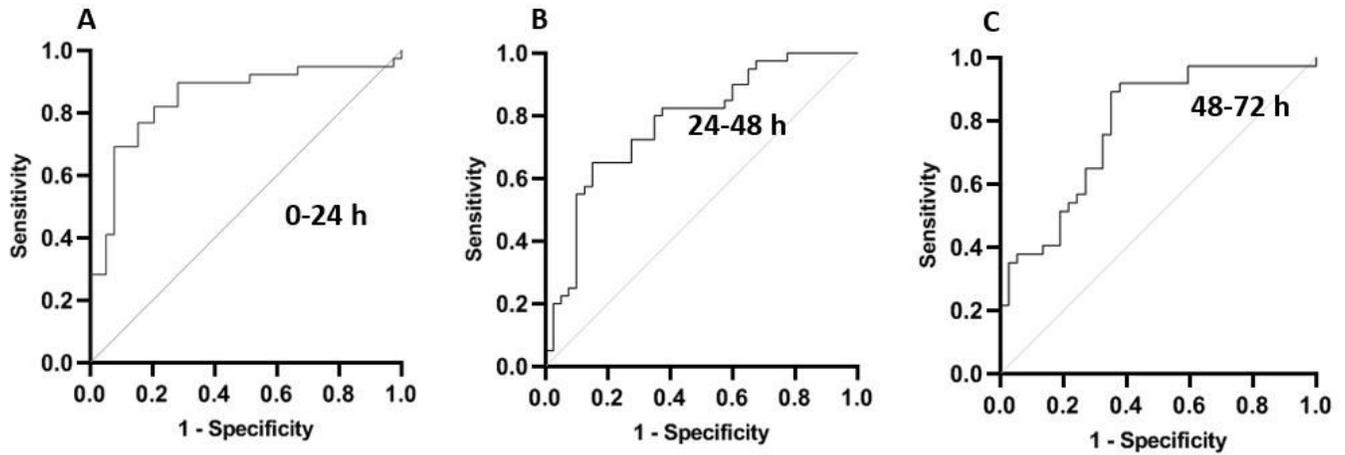


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