

# Low-cost and easily fabricated ultrasound-guided breast phantom for breast biopsy training

Si Yen Ng

National Cheng Kung University

Yao-Lung Kuo

National Cheng Kung University Hospital

Chi-Lun Lin (✉ [linc@mail.ncku.edu.tw](mailto:linc@mail.ncku.edu.tw))

National Cheng Kung University <https://orcid.org/0000-0003-1873-6078>

---

## Research article

**Keywords:** Training phantom, Ultrasound imaging, Breast biopsy, Haptic feedback, Acoustic properties

**Posted Date:** January 2nd, 2020

**DOI:** <https://doi.org/10.21203/rs.2.19957/v1>

**License:** © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

---

**Version of Record:** A version of this preprint was published at Applied Sciences on August 22nd, 2021.  
See the published version at <https://doi.org/10.3390/app11167728>.

# Abstract

**Background:** We developed an inexpensive and easy-to-fabricate gelatin-based breast biopsy training phantom and evaluated its effectiveness in improving the biopsy skill level and confidence level of residents.

**Methods:** Six residents were requested to evaluate the effectiveness of the double-layered breast phantom, which contained a malignant tumor, two benign tumors, and a cyst. The Young's modulus and acoustic properties of the gelatin tissue phantom and simulated tumors were investigated.

**Results:** The result showed that 83% (n=5) of the participants agreed that the ultrasound image quality produced by the breast phantom was excellent or good. However, only 17% (n=1) of the participants claimed that there was room of improvement for the haptic feedback they received during the placement of core needle into the breast phantom. The mean pre-instructional score was 17% (SD 33%) for all participants. The mean post-instructional score was 83% (SD 33%), giving an overall improvement of 1.32/2 mean scores. The mean needle biopsy skill and confidence levels of the participants substantially increased through simulation training on our breast phantom.

**Conclusions:** The proposed breast phantom can be easily fabricated by a resident for breast biopsy training. Simulation training using our breast phantom can improve a resident's confidence when performing challenging ultrasound-guided breast biopsy. According to the participants' feedback, our breast phantom is sufficiently realistic in terms of ultrasound imaging and haptic feedback during needle insertion; thus, the training outcome can be linked to the performance of residents when they perform a live biopsy.

## Background

Impalpable breast lesions found in breast cancer screening can be diagnosed through percutaneous image-guided breast biopsy. This technique has achieved dramatic improvements in terms of effectiveness and accuracy of diagnosis over the past few decades and is now commonly performed for palpable and nonpalpable lesions. The number of open surgical biopsies has declined due to the high accuracy of this minimally invasive needle biopsy [1].

The human breast is a heterogeneous structure containing different layers of tissues, predominantly fat and glandular tissues. Factors such as age, menstrual cycle, pregnancy, lactation, hormone therapy, and menopause affect the distribution of these various tissues. Pathophysiologic processes such as the tumor development change the intrinsic elasticity of soft tissues [2].

Young's modulus, the modulus of elasticity, has been used in several studies to quantify the mechanical properties of breast tissues [3–8]. These studies have reported that the stiffness of tumor tissue is much higher than that of normal breast tissues. Fibrous, glandular, and tumor tissues have higher Young's moduli than adipose tissue. Ramião et al. summarized the results of mechanical testing of ex vivo breast

tissue: the average Young's modulus is 0.69–24 kPa for normal fat tissue, 0.73–271.8 kPa for normal glandular tissue, 3.4–2162 kPa for tumor tissue (ductal carcinoma in situ), and 10–1366.5 kPa for tumor tissue (invasive ductal carcinoma) [9]. This wide variation in Young's modulus is found both across and within various tissue types. The variation is particularly large in normal fat and fibroglandular tissues.

The acoustic properties of real tissues vary among people and are not constant even within a person's body, giving rise to considerable variation in the literature [10, 11].

Ultrasound-guided breast biopsy is commonly used for tissue diagnoses of sonographically visible breast lesions. Nevertheless, many radiology residents in their graduate residencies have no hands-on experience in performing the procedure. The traditional apprenticeship training method on live patients has been practiced in the medical field instead of simulation training [12]. The Accreditation Council for Graduate Medical Education has strongly encouraged enhancing safety, predictability, and respect for patients by increasing the use of simulation training in graduate medical education [13]. Hence, there is a clinically unmet need for training phantoms.

In numerous residency training programs, turkey breast or gel breast phantoms are commonly used for simulation training of freehand ultrasound-guided breast procedures. Simulation techniques using a gelatin-based phantom are the most commonly described techniques in the literature [12, 14–21]. Pitted olives with pimentos, capers, grapes, peas, potatoes, and strawberries have been used to simulate breast masses for ultrasound-guided core biopsy. Rubber glove fingers filled with water and bath oil beads coated with nail polish have been used to simulate breast cysts for ultrasound-guided fine-needle aspiration. Plastic beads are embedded in the gelatin-based phantom developed by the Nicholson group [22]. Ruschin et al. proposed a gel-based breast phantom consisting of a simulated tumor with realistic imaging properties [23]. The background used in breast tissue simulation was made of ballistic gelatin powder and Metamucil, whereas the simulated tumors were composed of barium sulfate, copper sulfate, Metamucil, and ballistic gelatin. Another new soft tissue-mimicking material for ultrasound-guided breast biopsy training is paraffin-gel wax. Vieira et al. developed a paraffin-gel wax-based phantom embedded with cyst and tumor models [24]. To develop a breast phantom that has more realistic tactility and appearance in ultrasound images, a chicken breast phantom was embedded with poly (vinyl acetate) lesions to mimic the heterogeneity of real breast tissue [25]. In 1982, Madsen et al. produced a breast phantom that closely mimics the acoustic properties and density of real breast tissues [26]. Oil, gelatin, and agar were combined to fabricate the glandular tissue, fat tissue, skin, and Cooper's ligaments. The difference between normal and abnormal breast tissues was mimicked by employing the Young's modulus differences between the oil and gelatin. Finally, in 2006, Madsen et al. suggested dispersing safflower oil in solid aqueous gelatin to allow elastography in a breast phantom [27].

Many phantom users have reported that the existing phantoms do not provide adequate discrimination for basic imaging characteristics (e.g., resolution and contrast resolution). The present methods and test objects remain unable to relate image quality to clinical performance. In that sense, the influence of fat is

largely underestimated. Test objects do not generally show the type of artifacts that normally appear in real tissue nor do they represent complex tissue structures [28].

Ultrasound plays a vital role in breast cancer screening and has been used to distinguish benign lesions from malignant lesions and provide guidance during interventional biopsies. Therefore, the ultrasound operator must be properly trained. This can be implemented by the use of anthropomorphic training phantoms. Commercially available breast phantoms are very expensive and unaffordable by clinics. Although low-cost gelatin-based training phantoms have been proposed, the mechanical properties of their embedded simulated tumors have not been studied. The existing low-cost phantoms all have simple geometry and construction; they do not represent the real breast tissue structure. Therefore, this study aimed to develop a low-cost and easy-to-fabricate gelatin phantom embedded with simulated tumors that provide realistic haptic feedback. Test objects were constructed in three dimensions to obtain a realistic ultrasound image. Additionally, acoustic properties and ultrasound images for our simulated tumors are provided in this study as a reference for future research.

## Methods

### *Breast phantom manufacture*

A double-layered gelatin-based phantom consisting of malignant tumors, benign tumors, and cysts was produced. Gelatin tissue samples were produced by tuning the concentration of gelatin until its Young's modulus was similar to that of fat or glandular tissues. The simulated tumors were chosen on the basis of feedback from an experienced doctor to provide a realistic haptic feedback force during core needle placement.

### *Molds*

Two hemisphere-shaped molds with different diameters, 90 mm (mold A) and 120 mm (mold B), were designed using the computer-aided design software SolidWorks (SolidWorks 2018; Dassault Systèmes SolidWorks Corporation). The molds were manufactured through three-dimensional printing using polylactide.

### *Simulated tumors*

A rubber glove finger filled with colored solution was used to simulate cysts with a diameter of approximately 20 mm. Benign tumors were simulated using regularly shaped dried sweet potato and dried agar konjac with a diameter of 10 mm. The malignant tumor, which is the target of needle biopsy, was simulated using a pickled shallot carved into an irregular shape.

### *Glandular tissue*

Gelatin powder (40 g) was dissolved in 200 mL of 70°C tap water. The solution was mixed at 60 rpm for 5 min. The air bubbles were removed. Red and blue food coloring was added to give the gel solution an

opaque appearance. The gel solution was poured into mold A until it was 90% full to simulate the breast contour. The gel solution was refrigerated at 4°C for 30 min until it reached a semimolten consistency. The simulated tumors (Fig. 1a) were placed shallow in the mold. The gel solution was poured into the mold again until it was fully filled to cover the simulated tumors. The semimolten gel was then refrigerated for another 2 h. The completely solidified glandular tissue was removed from the mold and prepared for use later.

### *Fat tissue*

Gelatin powder (15 g) was dissolved in 300 mL of 70°C tap water. The solution was mixed at 60 rpm for 5 min. Air bubbles were removed. Red and blue food coloring was mixed in to give the gel solution an opaque appearance. A height of 12.5 mm from the base of mold B was marked. The gel solution was poured into mold B up to the marked line (Fig. 1b). The gel solution was refrigerated for 30 min at 4°C. Solidified glandular tissue from mold A was placed on top of the solidified gel in mold B, and space between mold B and glandular tissue was made sure to be uniform (12.5 mm for spacing distance), as depicted in Fig. 1b. Gel solution was poured into the mold again until the uniform space between mold B and the glandular tissue was fully filled. The gel solution was refrigerated for another 4 h until it had completely solidified. A successfully developed multilayered breast phantom embedded with simulated tumors is shown in Fig. 1c.

### *Mechanical property characterization*

Cylindrically shaped gelatin tissue samples (wt. 16.67% and wt. 4.76%) with 76-mm diameter and 25-mm height were fabricated and tested to characterize their mechanical behavior under deformation. The indentation test was conducted at room temperature (24°C) by using MTS INSIGHT-1 to obtain the relationship between the compressive force and displacement. An indenter 12 mm in diameter moved downward with a velocity of 0.5 mm/s until a depth of 6 mm was reached while the force and displacement were recorded. The Young's modulus ( $E$ ) of the gelatin tissue sample was calculated as follows [29]:

### **See Formula 1 in the Supplemental Files**

where  $F$  is the force applied to the indenter at the maximum indentation depth,  $\nu$  is the Poisson's ratio of the sample,  $w$  is the indentation depth,  $a$  is the radius of the indenter, and  $\kappa$  is a nondimensional parameter, which can be defined by a given combination of  $\nu$ ,  $a/h$ , and  $w/h$  ( $h$  is the thickness of sample). In our experimental setup,  $\kappa$  was calculated as 1.3624 according to the parameter table provided in [29], given that the Poisson's ratio of the gelatin tissue sample was approximately 0.5,  $a/h$  was 0.24, and  $w/h$  was 4%.

### *Acoustic property characterization*

The acoustic testing system comprised a pulse receiver (Model 5072PR; OLYMPUS), a three-axis motor stage, a data acquisition card (PXI-5152; National Instruments), and the programming environment

LabView (Fig. 2). A broadband immersion transducer with frequencies centered at 5 MHz was used in this system.

The single transducer worked as both a transmitter and receiver (pulse-echo approach) to measure the speed of sound, attenuation coefficient, and relative backscatter power of the sample. The measurements were conducted in a degassed-water-filled tank, the glass bottom of which acted as a reflector plane. The three-axis motor stage accurately positioned the transducer in the tank to focus on the reflector. The ultrasonic pulse was transmitted through the water to the reflector. The transducer received the reflected pulse, and the data were saved and analyzed in MATLAB R2018b. For each measurement of a sample, two data sets were obtained by scanning the reflector with and without the sample placed between the transducer and reflector (reference data and sample data, respectively). Three measurements were performed on each sample. All acoustic measurements were performed in degassed water at room temperature of 24°C.

The time interval between the peaks of the radio frequency pulses was computed using MATLAB. A Perspex mold was placed inside the water tank, between the transducer and reflector, when the sample was not in place. The following equation was used to calculate the speed of sound of the sample [30]:

### **See Formula 2 in the Supplemental Files**

where  $c_w$  is the speed of sound in water measured by scanning the reflector plane at 1 mm height differences, as illustrated in Figure 2;  $T_m$  and  $T_w$  are the travel times of the pulse signal obtained from the sample and reference data, respectively; and  $t_1$  and  $t_2$  are the travel times of the pulse signal from the transducer to the front and rear faces of the sample, respectively.

The logarithmic difference between the spectra was used to calculate the attenuation coefficient,  $\alpha$  (dB/cm), as shown in Eq. (3) [31], where  $d$  is the thickness of the sample and  $A(f)$  and  $A_o(f)$  are the magnitude spectrum of the sample and reference at frequency  $f$ , respectively.

### **See Formula 3 in the Supplemental Files**

A backscattered radio frequency (RF) signal was captured between the front face of the sample and reflector plane for backscatter power measurement. Welch's method was used to estimate the spectral power density and obtain the normalized distribution of power per unit frequency to the total received power in the backscattered RF signal. The relative backscatter power of the sample at frequency  $f$ , was calculated using the reference power spectrum,  $P_o(f)$ , of the reference RF signal as follows:

### **See Formula 4 in the Supplemental Files**

#### ***Breast phantom validation workshop***

A workshop was organized to evaluate the effectiveness of the developed phantom in improving participants' core needle biopsy skill (Fig. 3a). High-frequency linear probes, core needles, and a gelatin-

based breast phantom (Fig. 3b) were prepared. Six participants with different experience levels were invited to perform preinstructional biopsy. An experienced doctor then delivered a hands-on tutorial to the participants. The participants were given 10 min to practice the biopsy as many times as desired. A postinstructional biopsy was then performed by the participants after their practice session.

At the beginning of the workshop, the participants' levels of experience with ultrasound-guided core needle biopsy were assessed through three questions, resulting in a score range of 0–7. The participants were then categorized as inexperienced (score of 0), moderately experienced (1–4), or experienced (5–7).

The effectiveness of the gelatin-based breast phantom at improving the participants' confidence in performing challenging ultrasound-guided core needle biopsy was assessed through a self-assessment questionnaire. Before the simulation training, each participant completed a questionnaire scored using a 10-point Likert scale to assess his or her confidence in performing the ultrasound-guided breast biopsy without causing chest wall injury.

During the practical session, a participant's skill level of performing core needle biopsy was evaluated by comparing their preinstructional and postinstructional scores. A total of 2 points could be awarded for each needle biopsy operation if the biopsy was found to be a success in ultrasound imaging (+1) and a sufficient portion of the simulated malignant tumor had been retrieved in the biopsy needle (+1). However, a through-and-through puncture or penetration of the phantom backing was considered a chest wall hit, which resulted in the deduction of 1 point (–1). The participants were encouraged to perform as many practice biopsies as necessary before and after instruction. The percentage score was the total score of the participant divided by the highest possible score based on the number of biopsies performed. The percentage scores of participants before and after instruction were compared. The percentage mean scores of preinstructional and postinstructional tests were plotted on standard distribution curves.

After the simulation training, each participant repeated the questionnaire to assess their confidence in performing the ultrasound-guided breast biopsy. The satisfaction of the participants with the gelatin-based breast phantom was evaluated. Feedback was collected regarding the ultrasound image quality produced by the phantom and haptic feedback of the phantom that the participants received during core needle placement. Through this survey, the aspects of the phantom that residents valued the most could be better understood.

## Results

### Mechanical and acoustic property characterization

The mechanical and acoustic properties of the gelatin-based phantom and its simulated tumors were investigated. Gelatin tissue samples with wt. 16.7% and wt. 4.76% were fabricated to simulate glandular tissue and fat tissue, respectively. Force and displacement graphs for gelatin tissue samples were acquired from indentation tests performed at room temperature (24 °C). The Young's modulus of the glandular and fat gelatin tissue samples were 15.4 and 2.3 kPa, respectively, which fall reasonably in the

ranges for real normal fibroglandular tissue and normal fat tissue, as listed in Table 1 [9]. Also, the difference in stiffness between these two types of tissues could be distinguished. The speed of sound, attenuation coefficient, and relative backscatter power of each sample were measured at a frequency of 5 MHz at room temperature. The range of acoustic properties discovered in the samples is displayed in Table 2.

Table 1

Mechanical and acoustic properties for fat tissue, glandular tissue, and tumor from literature review [9]

Breast tissue composition	Young's modulus (kPa)	Speed of sound	Attenuation coefficient
Normal fat tissue	0.69-24	1553 ± 35	2.0 ± 0.7 at 7 MHz
Normal fibroglandular tissue	0.73–271.8	1479 ± 32	0.6 ± 0.1 at 7 MHz
Malignant lesions	6.41 ± 2.86	1550 ± 35	1.0 ± 0.2 at 7 MHz

Table 2

Experimental results of acoustic properties

Sample	Speed of sound (m/s)	Attenuation (dB/cm)	Integral backscattering (dB)
Glandular tissue	1684.26 ± 36.16	2.69 ± 0.07	3.11 ± 0.18
Fat tissue	1534.49 ± 1.22	2.34 ± 0.03	0.49 ± 0.10
Rubber glove with colored solution	1506.50 ± 6.07	4.50 ± 0.49	-0.46 ± 0.70
Dried sweet potato	1475.50 ± 17.82	4.12 ± 0.20	12.62 ± 0.48
Dried agar Konjac	1428.49 ± 1.81	3.46 ± 0.54	3.04 ± 0.26
Shallot	1503.83 ± 1.19	6.98 ± 0.11	7.21 ± 0.15

## Breast phantom validation workshop

Single-layered gelatin-based phantoms (glandular tissue embedded with simulated tumors) were evaluated by six participants in the validation workshop. All the participants completed the pretraining and posttraining questionnaires. The participants ranged from trainees to experienced doctors, with varying levels of experience in ultrasound-guided biopsy: 50% (n = 3) of the participants were inexperienced, 33% (n = 2) were moderately experienced, and the remaining 17% (n = 1) were experienced.

The mean preinstructional score of the participants was 17% (SD 33%). The mean postinstructional score was 83% (SD 33%) in which an overall improvement of 1.32 out of the total 2 was achieved (Fig. 4).

The largest improvement was achieved by the moderately experience group, with mean improvement of 2 out of the total 2. No chest wall hits were found in any group in the preinstructional and postinstructional tests.

Overall, the confidence level of all participants in the workshop substantially increased. The highest percentage of improvement in confidence level was obtained by the moderately experienced group (30%). A comparison between the pretraining and posttraining evaluation scores is shown in Fig. 5.

The results of the survey (Table 3) showed that, overall, the developed phantom satisfied by the participants. Furthermore, 83% (n = 5) of the participants agreed that the ultrasound image quality produced by the breast phantom was excellent or good. Example ultrasound images of the tumors embedded in the gelatin-based phantom are shown in Fig. 6. However, only 17% (n = 1) of the participants claimed that there was room of improvement for the haptic feedback they received during the placement of core needle into the breast phantom. In this survey, the aspects of the breast phantom that the participants valued the most were investigated. The most valuable aspects were ultrasound image quality (25%), haptic feedback (25%), ease of use (25%), cost (15%), and design (10%).

Table 3  
Survey on single-layered gelatin-based breast phantom

Survey (Single-layered phantom)	Number of respondents (total n = 6)				
	Excellent	Good	Neutral	Poor	Very poor
1. Overall how satisfied would you rate the developed phantom?	2 (33%)	3 (50%)	1 (17%)	-	-
2. How was the ultrasound image quality produced by the phantom?	1 (17%)	4 (66%)	1 (17%)	-	-
3. Was the haptic feedback you received during the placement of core needle into the phantom was a realistic feel?	1 (17%)	2 (33%)	2 (33%)	1 (17%)	-

A further study on the multilayered gelatin-based phantom (fat tissue, glandular tissue embedded with simulated tumors) was conducted (Table 4). All participants agreed that the ultrasound image produced by the multilayered breast phantom was similar to that of actual breast fat tissue. Moreover, all the participants also claimed that the haptic feedback they received during placement of the core needle into the multilayered gelatin-based phantom was more realistic than that for the single-layered phantom.

Table 4  
Survey on multilayered gelatin-based breast phantom

Survey (Multilayered phantom)	Number of respondents (total n = 6)	
	Yes	No
1. Was the ultrasound image produced by the multilayered phantom similar to the fat tissue for real breast?	6 (100%)	-
2. Was the haptic feedback of multilayered phantom you received during the placement of core needle into it was more realistic than the single-layered gelatin based phantom?	6 (100%)	-

## Discussion

The learning curve for performing ultrasound-guided breast biopsy could be overcome by performing simulation training using a breast phantom. Considerable improvements were achieved in the moderately experienced and inexperienced groups through training using the developed breast phantom. Compared with the preinstructional performance, the postinstructional score revealed a 4.88-fold improvement (Fig. 4), with the largest improvement achieved in the moderately experienced group. The experienced group, comprising one participant who had prior training in ultrasound-guided biopsy and frequently performed office-based ultrasound in his or her practice, already had the highest percentage score in the preinstructional test, and this was unchanged in the postinstructional test.

The breast-phantom-based training was well received by the participants and significantly increased their confidence in performing the ultrasound-guided breast biopsy. The outcome could be seen in the improved percentage mean scores of the postinstructional test for the moderately experienced and inexperienced groups.

Overall, the gelatin-based breast phantom developed in this study satisfied the participants. The Young's modulus of the phantom tissue samples measured from the indentation test fell reasonably in the reported ranges of the Young's modulus for real normal fibroglandular tissue and normal fat tissue [9]. However, our survey suggested that there should be a larger difference in haptic feedback between benign and malignant tumors, which can be understood as the Young's moduli in the literature having large variation.

The participants could differentiate between the shapes of cysts and benign and malignant tumors in ultrasound images. The multilayered gelatin-based breast phantom produced an ultrasound image similar to that produced by the fat tissue in real breasts. Artifacts caused by the boundary between the fat and glandular tissues in a real breast were successfully duplicated in our multilayered gelatin-based

phantom (Fig. 6e). Acoustic properties and ultrasound images for our simulated tumors have been provided as a reference for future studies.

Numerous low-cost DIY breast phantoms have been proposed. Turkey breast embedded with peas and gel-based phantoms are the most commonly used models in ultrasound-guided breast biopsy simulation training [12, 14–23, 25]. Although turkey breast can be easily obtained from the supermarket, its preparation process, such as cleaning and thawing, can be troublesome and is not user friendly. Different types of materials have been used to simulate tumors. Aspects such as shape, brightness, and contrast of the embedded simulator in ultrasound images are prioritized when selecting a material, and the haptic force produced by the embedded simulator during needle insertion may be overlooked. We were also unable to find a low-cost DIY breast phantom that contains all three types of simulators: a benign tumor, malignant tumor, and cyst. Moreover, we only found one breast phantom, proposed by Madsen et al., in which different layers of tissue are mimicked, but the lack of simulated tumors makes this phantom unsuitable for breast biopsy training [26]. Our gelatin-based breast phantom is a low-cost and multilayered breast phantom embedded with simulated benign tumors, malignant tumors, and cysts for breast biopsy simulation training. The total cost is approximately USD 1.59. The price of our training phantom is thus extremely low compared with that of the commercially available ultrasound-guided breast phantoms, which is approximately USD 157.35.

Environmental temperature control is crucial to maintaining the stability of a gelatin-based phantom. The Young's modulus of a gelatin-based phantom decreases as the temperature increases. The indentation tests were conducted at 24 °C, and the results showed that the Young's modulus of the components in the phantom fell reasonably within the reported ranges. The temperature of the breast phantoms used in the validation workshop could not be ideally controlled. Each phantom was kept in a cooler until being offered to a participant to operate. In this scenario, the temperature of each phantom was measured as 19 °C immediately before the operation. Based on the results of the indentation tests and validation, we recommend that users handle the proposed gelatin-based phantom at 19 °C–24 °C. A gelatin sample with a low concentration, such as a fat-tissue-mimicking sample, will start losing its original form when the sample temperature exceeds 25 °C.

This study certainly has limitations. Our participants were few and from a single academic center. Moreover, some facets of real-world experience were not reproduced in our simulation. For instance, the orientation of the patient cannot be adjusted as easily as that of the small training phantom during the biopsy procedure. A gap between the improved performance of participants' biopsy skill level after simulation training and that in the clinical setting may therefore appear. However, our findings can serve as a basis for proving that an unmet clinical need exists, and training involving our breast phantom is efficient in improving the confidence and skill level of residents at performing needle biopsy. A follow-up evaluation must be performed to determine whether the training outcome is related to the performance of the participants in the real clinical setting.

## Conclusions

This study provides a protocol for developing a low-cost and easy-to-fabricate multilayered gelatin-based breast phantom. The mechanical properties of the gelatin phantom and embedded simulated tumors were studied to ensure that realistic needle-insertion haptic feedback is provided. The gelatin-based phantom enabled residents to improve their confidence level in performing ultrasound-guided needle biopsy. Simulation training on the breast phantom can enable residents to master the needle biopsy skill before advancing to performing procedures on patients. Complications can be reduced through training, especially when the ultrasound-guided biopsy procedure is performed by an inexperienced resident. Acoustic properties and ultrasound images for our simulated tumors are provided as a reference for future study.

## **Declarations**

### **Ethics approval and consent to participate**

The consent to participate was informed verbally to all participants on the day of the breast phantom validation workshop. The workshop was to collect user feedback of operating needle biopsy on the developed breast phantom. The need for consent is waived by the IRB in Taiwan (Document No. 8800-4-04-008: educational assessment, teaching skills, or effectiveness evaluations conducted in a general teaching environment by anonymous or unrecognizable means).

### **Consent for publication**

Not applicable.

### **Availability of data and materials**

The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

### **Competing interests**

The authors declare that they have no competing interests.

### **Funding**

This research is supported by the Ministry of Science and Technology under funding no. 106-2221-E-006-049.

### **Authors' contributions**

SY contributed to the research ideation, summarized the relevant literature, studied and manufactured the breast phantoms, conducted engineering tests and user surveys, and completed writing the major part of this paper. CL initiated the research idea, led the discussions on developing methodologies and evaluating results, made strategic plans and decisions, supervised research progress and provided

guidance, collaborated on and reviewed the paper writing. YL contributed his medical expertise in research ideation and regular discussions, provided user feedback and advice for the design improvement of the breast phantom, helped planned and organized the surveys, and collaborated on and reviewed the paper writing.

## Acknowledgment

The authors would like to thank residents from National Cheng Kung University Hospital who participated in the breast phantom validation workshop.

## Abbreviations

1. Radio frequency (RF)

## References

1. O'Flynn EAM, Wilson ARM, Michell MJ. Image-guided breast biopsy: state-of-the-art. *Clin Radiol*. 2010;65(4):259-70.
2. Bogomoletz WV. Elastosis in breast cancer. *Pathol Annu*. 1986;21:347-66.
3. Jurvelin JS, Buschmann MD, Hunziker EB. Optical and mechanical determination of poisson's ratio of adult bovine humeral articular cartilage. *J Biomech*. 1997;30(3):235-41.
4. Krouskop TA, Wheeler TM, Kallel F, Garra BS, Hall T. Elastic moduli of breast and prostate tissues under compression. *Ultrason Imaging*. 1998;20(4):260-74.
5. Wellman P, Howe RD, Dalton E, Kern KA. Breast tissue stiffness in compression is correlated to histological diagnosis. Harvard BioRobotics Laboratory Technical Report; 1999. p. 1-15.
6. Abbas S, Judit Z, Donald P. Elastic moduli of normal and pathological human breast tissues: an inversion-technique-based investigation of 169 samples. *Phys Med Biol*. 2007;52(6):1565.
7. Unlu MZ, Krol A, Magri A, Mandel JA, Lee W, Baum KG, et al. Computerized method for nonrigid MR-to-PET breast-image registration. *Comput Biol Med*. 2010;40(1):37-53.
8. Cox TR, Epler JT. Remodeling and homeostasis of the extracellular matrix: implications for fibrotic diseases and cancer. *Dis Model Mech*. 2011;4(2):165-78.
9. Ramião NG, Martins PS, Rynkevic R, Fernandes AA, Barroso M, Santos DC. Biomechanical properties of breast tissue, a state-of-the-art review. *Biomech Model Mechanobiol* 2016;15(5):1307-23.
10. Jones RM. Tissue substitutes, phantoms and computational modelling in medical ultrasound-ICRU (Report 61). *Health Phys*. 2000;78(3):344.

11. John C. The coronio-apically varying ultrasonic velocity in human hard dental tissues. *J e Acoust Soc Am.* 2004;116(1):545-56.
12. Sutcliffe J, Hardman RL, Dornbluth NC, Kist KA. A novel technique for teaching challenging ultrasound-guided breast procedures to radiology residents. *J Ultrasound Med.* 2013;32(10):1845-54.
13. Philibert I, Leach, DC. Simulation and rehearsal ACGME bulletin. Accreditation Council for Graduate Medical Education. 2005.
14. Silver B, Metzger TS, Matalon TA. A simple phantom for learning needle placement for sonographically guided biopsy. *Am J Roentgenol.* 1990;154(4):847-8.
15. Bude RO, Adler RS. An easily made, low-cost, tissue-like ultrasound phantom material. *J Clin Ultrasound.* 1995;23(4):271-3.
16. Georgian-Smith D, Shiels WE. From the RSNA refresher courses: freehand interventional sonography in the breast: basic principles and clinical applications. *Radiographics.* 1996;16(1):149-61.
17. Harvey JA, Moran RE, Hamer MM, DeAngelis GA, Omary RA. Evaluation of a turkey-breast phantom for teaching freehand, US-guided core-needle breast biopsy. *Acad Radiol.* 1997;4(8):565-9.
18. Sisney GA, Hunt KA. A low-cost gelatin phantom for learning sonographically guided interventional breast radiology techniques. *Am J Roentgenol.* 1998;171(1):65-6.
19. Hassard MK, McCurdy LI, Williams JC, Downey DB. Training module to teach ultrasound-guided breast biopsy skills to residents improves accuracy. *Can Assoc Radiol J.* 2003;54(3):155-9.
20. Morehouse H, Thaker HP, Persaud C. Addition of metamucil to gelatin for a realistic breast biopsy phantom. *J Ultrasound Med.* 2007;26(8):1123-6.
21. Meng K, Lipson JA. Utilizing a PACS-integrated ultrasound-guided breast biopsy simulation exercise to reinforce the ACR practice guideline for ultrasound-guided percutaneous breast interventional procedures during radiology residency. *Acad Radiol.* 2011;18(10):1324-8.
22. Nicholson RA, Crofton M. Training phantom for ultrasound guided biopsy. *Br J Radiol.* 1997;70(830):192-4.
23. Ruschin M, Davidson SRH, Phounsy W, Yoo TS, Chin L, Pignol J-P, et al. Technical Note: Multipurpose CT, ultrasound, and MRI breast phantom for use in radiotherapy and minimally invasive interventions. *Med Phys.* 2016;43(5):2508-14.
24. Vieira SL, Pavan TZ, Junior JE, Carneiro AAO. Paraffin-gel tissue-mimicking material for ultrasound-guided needle biopsy phantom. *Ultrasound Med Biol.* 2013;39(12):2477-84.

25. Smith WL, Surry KJM, Kumar A, McCurdy L, Downey DB, Fenster A. Comparison of core needle breast biopsy techniques: freehand versus three-dimensional US guidance. *Acad Radiol.* 2002;9(5):541-50.
26. Madsen EL, Zagzebsk JA, Frank GR. An anthropomorphic ultrasound breast phantom containing intermediate-sized scatterers. *Ultrasound Med Biol.* 1982;8(4):381-92.
27. Madsen EL, Hobson MA, Frank GR, Shi H, Jiang J, Hall TJ, et al. Anthropomorphic breast phantoms for testing elastography systems. *Ultrasound Med Biol.* 2006;32(6):857-74.
28. Shaw A, Hekkenberg R. Standards to support performance evaluation for diagnostic ultrasound imaging equipment. National Physical Laboratory; 2007.
29. Zhang M, Zheng YP, Mak AFT. Estimating the effective Young's modulus of soft tissues from indentation tests—nonlinear finite element analysis of effects of friction and large deformation. *Med Eng Phys.* 1997;19(6):512-7.
30. Shung KK. *Diagnostic ultrasound: imaging and blood flow measurements.* 2005
31. Technical Standards Committee AIUM. *Methods for specifying acoustic properties of tissue mimicking phantoms and objects.* Laurel, MD: American Institute of Ultrasound in Medicine; 1995.

## Figures



(a)



(b)



(c)

**Figure 1**

(a) Rubber glove finger filled with colored solution, dried sweet potato, dried agar konjac and carved shallot were used to simulate tumors (from left to right); (b) Gel solution was poured until the line mark (indicated by black hollow arrow) located at a height of 12.5 mm from the base (left) and spacing distance of 12.5 mm between the solidified glandular tissue and mold B (right); (c) Gelatin-based phantom which consists two layers with different stiffness and embedded with simulated tumors

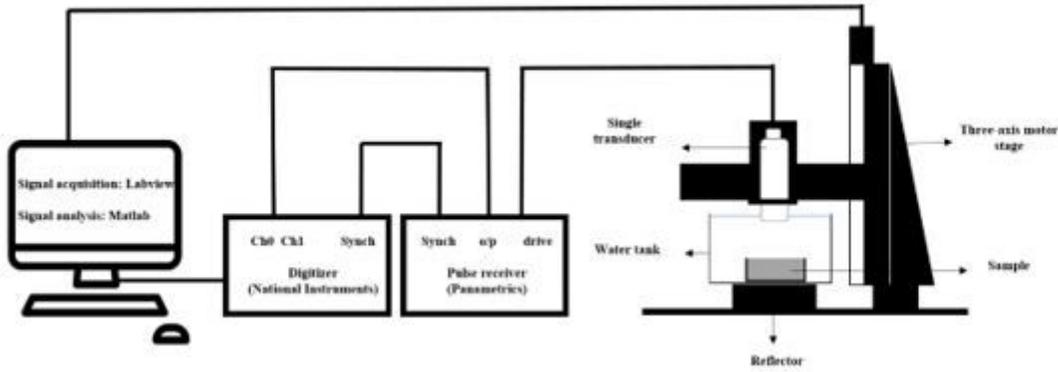


Figure 2

Scanning acoustic microscope (SAMA) system



Figure 3

(a) Workshop for evaluating the efficiency of the phantom in improving core needle biopsy skill of participant. (b) Single-layered gelatin-based phantom (glandular tissue embedded with simulated tumors)

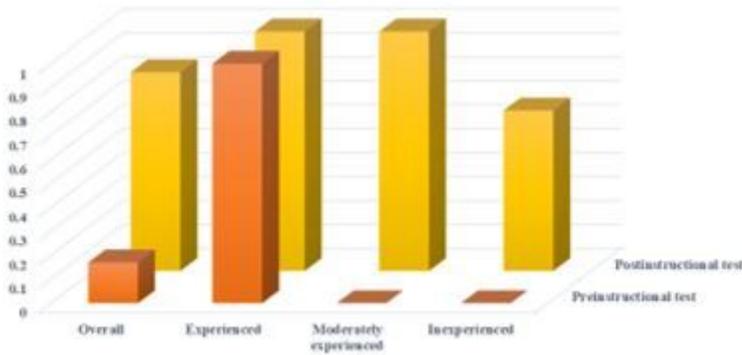
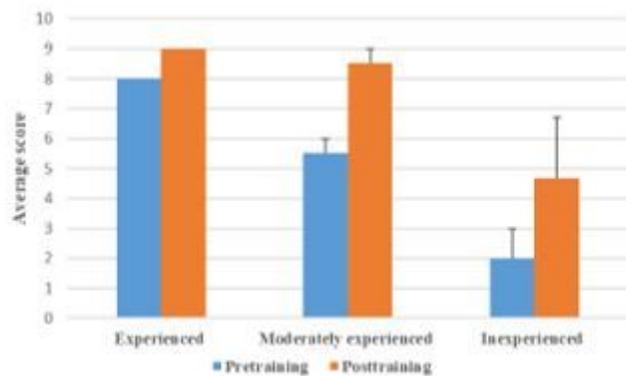


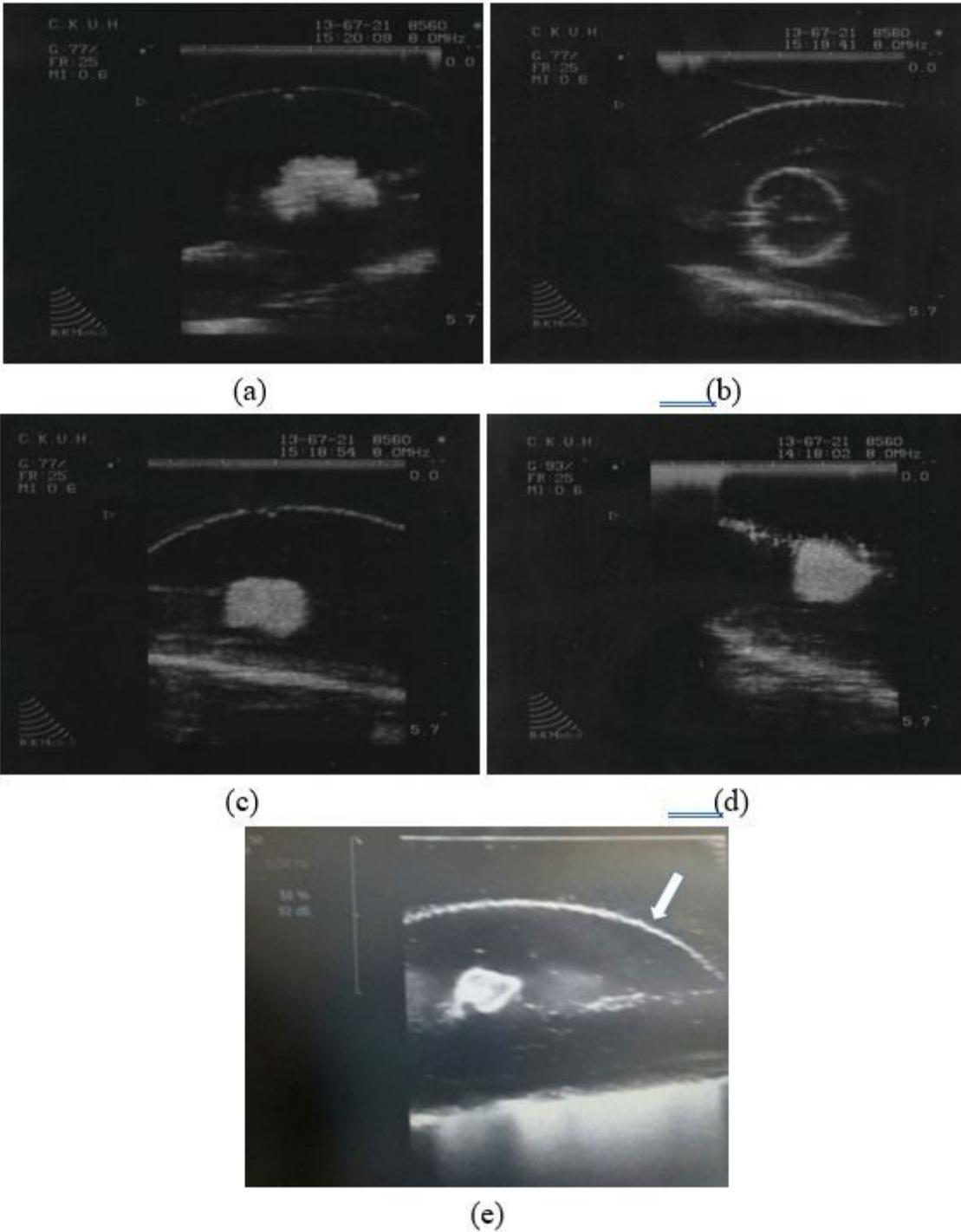
Figure 4

Pre-instructional and post-instructional test results



**Figure 5**

Survey results for pre-training and post-training confidence level of the participants (n = 6) who participated in the US-guided core needle biopsy



**Figure 6**

Ultrasound images for embedded tumors and artefact in gelatin-based phantom. (a) Malignant tumor (Shallot); (b) Cyst (Rubber glove finger filled with colored solution); (c) Benign tumor (Dried agar konjac); (d) Benign tumor (Dried sweet potato); (e) Artefact caused by the boundary between the fat and glandular tissues is indicated by the white arrow

## Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [formula2.JPG](#)
- [formula4.JPG](#)
- [formula3.JPG](#)
- [formula1.JPG](#)