

# Novel Liquid Mycofungicide: An Ideal Approach in Protecting Tomato from the Damping-off Disease in Nursery

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## Research article

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# Abstract

**Background:** A novel liquid formulation of *Trichoderma asperellum* was prepared and its effectiveness was assessed by different methods for the management of tomato damping-off in the nursery. The experiments were conducted for two consecutive years in a randomized block design with three replications.

**Results:** *T. asperellum* formulation improved seed germination and management of tomato damping-off when compared with control. All the treatments performed better over control, in the first year, the antagonist's application enhanced seed germination up to 75.75 percent, however, it was 60.13 percent in control. There was 10.93 to 20.38 percent seedling mortality due to damping off which was comparatively lower than the control (26.98%). A similar trend of seed germination and disease incidence was observed in the second year. In addition to managing the disease, the antagonist certainly promoted the vegetative growth which was reflected as increased shoot and root length in comparison to control during both years. During the first year, shoot length ranged 10.90 – 12.85 cm as compared to the control (8.72 cm) and root length ranged from 3.21 to 3.65 cm which was greater than the control. Almost a similar trend in the vegetative growth parameters of seedling was observed during the second year.

**Conclusion:** The present investigation showed that the tested antagonist's formulation could efficiently managed the tomato damping-off as well as encouraged the vegetative growth of seedlings which ultimately ensured better and healthy seedling. And this formulation can successfully used through different methods to take care of tomato damping off.

## Background

Tomato (*Solanum lycopersicum* L.) is an imperative constituent of human food basket as a good source of vitamins and minerals. It is consumed as a fresh, vegetable as well as processed products such as juice, ketchup, sauce, canned fruits, puree, paste, etc. India occupied about 789 thousand ha under tomato with a production volume of approximately 19759 thousand MT. It is also an important crop in Haryana with a 30115 ha area and production 643548 MT [1].

Having a healthy and disease-free nursery is the foremost inevitability in determining profitable crop production. Soil inhabitant fungal phytopathogens is a major challenge in the nursery raising. Species of *Pythium*, *Phytophthora*, *Rhizoctonia*, *Fusarium*, etc cause various soil-borne diseases such as damping off, root rot, wilt, etc [2, 3, 4]. Damping-off is a global problem of seedlings in the nursery, which may cause 5–80% seedling mortality of different crops [5]. Usage of synthetic fungicides had been a preferred approach since long back in managing the soil-borne fungal phytopathogens. However, issues related to human and soil health, environmental hazards, and disturbed ecosystem [6, 7, 8, 9] pressurized the agriculturists to switch over the plant protection measures towards the safer and eco-friendly approaches [10]. Biological control agents could be an ideal approach for diseases management [11, 12, 13] as they have no chance to get into the food chain and hence safe to human beings, animals, and plants [14].

*Trichoderma* is the most widely applied biological tool in plant protection and prospective to minimize the usage of synthetic fungicides in the agriculture sector [15]. It naturally presents in a variety of soils, decaying roots, litter, wood, bark, and other plant materials [16,17]. It alone contributes up to ninety percent of total antagonistic fungi [18] and has positive effects on the plants [19] and efficient to take care of diseases of agricultural, horticultural, ornamental crops, etc [20, 21] caused by phytopathogenic fungi [22], bacteria, and viruses [23] through various modes of actions [24]. It produces several enzymes for the degradation of the cell wall and numerous volatile metabolites [25, 26] for the control of phytopathogens. As of date, about 180 secondary metabolites (antibiotics) are characterized which inhibit or impose a lethal effect on the phytopathogens [27, 28]. Its rhizospheric application strengthens the plants to fight against aerial fungal, bacterial, and viral diseases [18] due to elicitors production and resistance induction [29]. *T. viride*, *T. harzianum*, *T. longibrachiatum*, and *T. koningii* produce volatile antibiotics [30, 31]. This antagonist creates unfavorable conditions for the phytopathogenic fungi through acidification of the environment [18].

It stimulates plant growth, persuades yield, vitamin production, boosts up nitrogen, and phosphorus availability as well as nutrients mobilization [32]. The antagonist produces zeaxanthin and gibberellins to excel in the seed germination. It also produces different acids to enhance the availability of phosphorus and other micronutrients to the plants [29]. It stimulates the local and systemic resistance in plants [33, 34, 35, 36].

It enhanced the concentration of enzymes and other important proteins for plant resistance development [37, 38]. Indigenous isolates of concerned regions performed better in that particular region [33]. Effectiveness of *T. harzianum* proven in controlling root rot (*Phytophthora capsici*) of pepper [39].

*T. harzianum* and *T. virens* found potential in managing the pink rot of potato and stem rot of tomato, boosted up potato tuber yield and survival of tomato seedlings [40]. Tomato, lettuce, pepper, bean, and tobacco showed resistance against *B. cinerea* owing to the application of *T. harzianum* [41]. It also induced the defense system in cucumber plants [38]. It can be used for soil application [42], seed treatment [43] mixing with farmyard manure [44], and vermicompost. It is capable of controlling the seed and soil-borne diseases of vegetables in the nursery [45]. Its 254 species are documented in the literature [46]; nevertheless, a few species have scope for control of phytopathogens [47]. Though *T. asperellum* is effective against numerous phytopathogens [48, 49] however, comparatively lesser attention has been paid for its agricultural application [50, 51].

Integration of *Trichoderma* sp in plant protection strategies could reduce the dose of synthetic fungicides and improve plant health [52] which is environment friendly as well as cost-effective [53]. Keeping into consideration the disease managing potency and plant growth promotional activity of *Trichoderma* spp, the present study was planned to test its bioefficacy against the damping-off of the tomato seedlings caused by *Fusarium* sp in the nursery.

## Results

The investigation revealed that the liquid formulation of the antagonist could improve seed germination as well as reduce the damping-off in comparison with control in the nursery when it was applied in different ways, however, soil application, enriched vermicompost, and FYM found more promising.

In the first year, the seed germinated from 63.88 to 88.75 as compared to 60.13 percent in control and the seedling showed 10.93 to 20.38 percent damping off which was less than the control (26.98%) as indicated in Table 2. Likewise, in the second year, the seed was germinated from 70.83 to 89.50 percent when the antagonist was applied in various ways. The seedlings suffered from damping off disease lesser (8.38–15.33%) as compared to the control bed (20.21%) as evident from Table 3 and Fig. 2.

The antagonist positively supported the vegetative growth of the seedlings in terms of higher shoot and root length in comparison with control during both years (Fig. 2). During the first year, shoot length ranged 10.90–12.85 cm as compared to the control (8.72 cm), nevertheless, root length was observed in the range of 3.21 to 3.91 cm which was more than the control (2.70 cm). Almost a similar trend in the vegetative growth parameters of seedling was noted during the second year (Figs. 3 and 4). The maximum and minimum temperatures during 2018-19 ranged from 17.1 to 21.0 °C and 1.9 to 7.7 °C, respectively. Morning relative humidity was 90.3–99.1 and during the evening it was 48.5–65.7 (Fig. 5). During 2019 -20, the maximum and minimum temperatures ranged from 11.9 to 22.8 and 2.6 to 8.3 °C. Relative humidity during the morning was recorded 88.0 to 100 and during the evening it was 46.0–82.0 percent (Fig. 6).

## Discussion

*Trichoderma* spp found effective in controlling the Fusarium wilt of tomato, watermelon, and muskmelon in the greenhouse and field [54]. Sowing of tomato seeds in *T. harzianum* amended soil led a significant decrease in the incidence of Fusarium crown and root rot disease caused by *F. oxysporum* f. sp. *radicis-lycopersici* [55]. Researchers also assessed this antagonist for the control of this disease of vegetable crops through various application methods and noted similar findings [45].

Seed treatment, seedbed treatment, and enrichment of farmyard manure with *T. viride* and *T. harzianum* could offer a better damping-off solution of cauliflower under field conditions and improved seedling vegetative growth and health [56]. Such treatments were found effective in terms of reduced disease and improved seedling strength and health of tomato and chilli plants [57]. Tomato seed treatment with *T. asperellum* taken care of *P. aphanidermatum* [58]. Seed treatment with a consortium of *T. harzianum* significantly declined the damping-off incidence and enhanced the shoot length, chlorophyll content, and yield [59]. Inoculation of tomato seed and nursery soil with *T. harzianum* spore suspension laid increment in the shoot as well as root length, diameter, and weight when compared to the control. There was a noticeable increase in leaf number, leaf area, chlorophyll content [60].

Application of *T. asperellum* in field conditions, managed phytopathogenic *Fusarium* sp, and significantly increased the vegetative growth in respect of root length, shoot length, plant weight as well as chlorophyll

content. It also improved total phenol, peroxidase, polyphenoloxidase, and phenylalanine ammonium lyase activity which developed plants resistance against the pathogen [61]. Pre-sowing application of *T. viride* (WP) in nursery beds of tomato showed a reduction in disease incidence. Nevertheless, growth parameters showed a positive impact [62]. Soil application of *T. viride* efficiently controlled the pre- and post-emergence damping-off of tomato (*P. aphanidermatum*) and improved vegetative growth of the plant [63]. *T. harzianum* and *T. virens* treated tomato seed revealed up to cent percent seed germination and increased root and shoot length [64].

*Trichoderma* amended compost could effectively suppress root disease causal fungal phytopathogens viz., *F. oxysporum*, *P. debaryanum*, *P. aphanidermatum*, *R. solani* in tomato and persuaded seed germination, seedling height, and overall biomass of seedling [65]. *T. harzianum* enriched organic matter could reduce the tomato damping-off intensity and increase seedling growth [66]. Concurrent application of *T. harzianum* as seed, seedbed, and soil treatment performed better than its soil application in terms of disease management and better growth parameters of eggplant and tomato. Vermicompost enriched with antagonist increased seed germination, promoted seedling vegetative growth, and minimized damping-off incidence [67]. The disease control and plant growth-promoting property of *T. asperellum* were established against *R. solani* in cucumber plant [68]. Inoculation of *Trichoderma* in tomato nursery and field could improve plant health in terms of increased the shoot weight, fruit size, yield [69]. Furrow application of *T. harzianum* in the field conditions could reduce black scurf incidence in organically grown potatoes [70]. A few *Trichoderma* strains showed good control of *P. ultimum* in pea and plant growth promotional property in terms of increased shoot weight, root weight, root length number of lateral roots [71].

## Methods

### Experimental location

The nursery trials were carried out at the research farm at google map position: 29.153843 NS Latitude and 75.694118 EW Longitude, Department of vegetable science, Chaudhary Charan Singh Haryana Agricultural University, Hisar for two consecutive years during 2018-19 and 2019-20.

### Preparation of mycofungicide formulation

The liquid formulation (10% Aqueous Suspension) of *T. asperellum* (KBN-29, accession number ITCC-7764) was prepared following a slightly modified method [72, 58]. The pure culture of the antagonist was inoculated into potato dextrose agar (HiMedia India Pvt. Ltd.) plates, followed by proper sealing with parafilm under aseptic conditions and incubated at  $26 \pm 2^{\circ}\text{C}$  for 5 days in a BOD incubator. Ten milliliters double-distilled sterilized water was added to each plate and fungal biomass (conidia and mycelia) was scrapped with the help of scrapper. The harvested biomass was further diluted with water and a conidial

concentration of approximately  $2 \times 10^9$  per ml was determined through serial dilution plate inoculation technique (Fig. 1). The prepared formulation was kept in the refrigerator and used within a month.

## Source of plants

The locally cultivated tomato var. Selection-7, released by the Department of Vegetable Science, Chaudhary Charan Singh Haryana Agricultural University was used for experimentation. Seed of this variety was obtained from the Vegetable seed store of this Department.

## Assessment of different application methods against damping off

The developed formulation was assessed as a seed treatment, soil application, drenching, enriched FYM, and vermicompost. The antagonist treated seed was dried under shade. In soil application, the formulation was sprayed uniformly in to plot before 4-5 hr. It was mixed with FYM and vermicompost before 15 days of its field application and kept covered under shade. Carbendazim was taken as a standard check in the experiment. There were seven treatments in three replications (Table 1). Raised seedbeds of 90 cm x 45 cm dimensions were prepared in mid-December, 2018, and 2019 (Fig. 2). Two hundred tomato seeds per treatment were sown manually. After 15 days damping-off incidence was recorded. Then drenching of antagonist and carbendazim was done in respective treatments and again after two weeks, disease observation was recorded in comparison with control. After about 45 days of nursery raising, plant growth in terms of shoot and root length (cm) was measured from ten randomly sampled seedlings per treatment.

## Data analysis

The experimental data were statistically analyzed through the online package 'OPSTAT' of the institute.

## Weather conditions during the study period

Weather data of temperature, relative humidity, wind speed, sunshine hours, and rainfall were collected from the Department of Agrometeorology, College of Agriculture, Chaudhary Charan Singh Haryana Agricultural University, Hisar.

## Conclusion

The present investigation showed that the tested antagonist's formulation could efficiently managed the tomato damping-off as well as encouraged the vegetative growth of seedlings which ultimately ensured

better and healthy seedling. And this formulation can successfully used through different methods to take care of tomato damping off.

## Abbreviations

TSA- *Trichoderma* soil application, TD- *Trichoderma* drenching, TST- *Trichoderma* seed treatment, TE-FYM- *Trichoderma* enriched FYM, CarbD-Carbendazim drenching, TEV- *Trichoderma* enriched vermicompost

## Declarations

## Ethics approval and consent to participate

The study was conducted with tomato plant species those are abundant in the ecosystem hence do not require ethical approval.

## Consent for publication

The authors agree to publish this paper. The data has not been published partially or completely in any other journal.

## Availability of data and materials

The datasets generated and analyzed during the current study are not publicly available due to privacy issues but are available from the corresponding author on reasonable request.

## Competing interests

The authors declare that there is no conflict of interest regarding the publication of this paper. It is declared that the authors have no competing interests.

## Funding

There is no financial support for this purpose from any funding agency at this time; it was carried out as an in-house study. However, the BMC (waivers) Plant Biology Journal considered for review process without processing charge.

## Authors' contributions

Kishor Chand Kumhar generated the idea of the study, prepared mycofungicide formulation, decided treatments, recorded disease incidence, analyzed, and presented data in tables and graphs. Kuldeep Kumar arranged seed, raised nursery, and recorded data on growth parameters. Indu Arora has taken care of the nursery during the study period. Arun Kumar Bhatia and Vinod Kumar Batra timely monitored and suggested the required follow up action.

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## References

1. Anonymous. Horticultural Statistics at a glance. India; 2018.
2. Srinon W, Chuncheon K. Efficacies of antagonistic fungi against Fusarium wilt disease of cucumber and tomato and the assay of its enzyme activity. *J Agric ....* 2006;;191–201.
3. Hooda KS, Joshi D, Dhar S, Bhatt JC. Management of damping-off of tomato with botanicals and bio-products in North Western Himalayas. *Indian J Hortic.* 2011;68:219–23.
4. Jiskani MM. Studies on the control of tomato damping-off disease caused by *Rhizoctonia solani* Kühn studies on the control of tomato damping-off. *Pak J Bot.* 2014; December 2007.
5. Lamichhane JR, Dürr C, Schwanck AA, Robin MH, Sarthou JP, Cellier V, et al. Integrated management of damping-off diseases. A review. *Agronomy for Sustainable Development.* 2017;37.
6. Sarwar M. The Killer Chemicals as Controller of Agriculture Insect Pests: The Conventional Insecticides. *Int J Chem Biomol Sci.* 2015;1:141–7.
7. Nicolopoulou-Stamati P, Maipas S, Kotampasi C, Stamatis P, Hens L. Chemical Pesticides and Human Health: The Urgent Need for a New Concept in Agriculture. *Front Public Heal.* 2016;4.
8. García-García CR, Parrón T, Requena M, Alarcón R, Tsatsakis AM, Hernández AF. Occupational pesticide exposure and adverse health effects at the clinical, hematological and biochemical level. *Life Sci.* 2016;145:274–83.
9. Shammi M, Sultana A, Hasan N, Mostafizur Rahman M, Saiful Islam M, Bodrud-Doza M, et al. Pesticide exposures towards health and environmental hazard in Bangladesh: A case study on farmers' perception. *J Saudi Soc Agric Sci.* 2020;19:161–73.
10. Rosenzweig C, Iglesias A, Yang XB, Epstein P, Chivian E. Climate Change and Extreme Weather Events; Implications for Food Production, Plant Diseases, and Pests. *Glob Chang Hum Heal.* 2001;2:90–104.
11. Vos CM, Yang Y, De Coninck B, Cammue BPA. Fungal (-like) biocontrol organisms in tomato disease control. *Biological Control.* 2014;74:65–81.



12. Thakur N, Tripathi A. Biological Management of Damping-Off, Buckeye Rot and Fusarial Wilt of Tomato (cv. Solan Lalima) under Mid-Hill Conditions of Himachal Pradesh. *Agric Sci.* 2015;06:535–44.
13. Mbuthia LW, Muriithi Kiirika L, Afolayan G, Henning VA, Kiirika LM. Interactive effects of arbuscular mycorrhiza fungi *Glomus intraradices* and *Trichoderma harzianum* against *Fusarium* wilt of tomato. *Int J Biosci.* 2019;15:251–68.
14. Monte E, Llobell. *Trichoderma* in organic agriculture. *Proc V World Avocado Congr.* 2003;:725–33.
15. Sayeed Akhtar M, Siddiqui ZA. Biocontrol of a root-rot disease complex of chickpea by *Glomus intraradices*, *Rhizobium* sp. and *Pseudomonas straita*. *Crop Prot.* 2008;27:410–7.
16. Brewer D, Calder FW, Macintyre TM, Taylor A. Ovine ill-thrift in Nova Scotia I. The possible regulation of the rumen flora in sheep by the fungal flora of permanent pasture. *J Agric Sci.* 1971;76:465–77.
17. Danielson RM, Davey CB. The abundance of *Trichoderma* propagules and the distribution of species in forest soils. *Soil Biol Biochem.* 1973;5:485–94.
18. Benítez T, Rincón AM, Limón MC, Codón AC. Biocontrol mechanisms of *Trichoderma* strains. *International Microbiology.* 2004;7:249–60.
19. Hermosa R, Viterbo A, Chet I, Monte E. Plant-beneficial effects of *Trichoderma* and of its genes. *Microbiology.* 2012;158:17–25.
20. Harman GE. Myths and dogmas of biocontrol: Changes in perceptions derived from research on *Trichoderma harzianum* T-22. *Plant Disease.* 2000;84:377–93.
21. Smoliška U, Kowalska B, Oskiera M. The effectivity of trichoderma strains in the protection of cucumber and lettuce against *rhizoctonia solani*. *Veg Crop Res Bull.* 2007;67:81–93.
22. Lewis JA, Lumsden RD. Biocontrol of damping-off of greenhouse-grown crops caused by *Rhizoctonia solani* with a formulation of *Trichoderma* spp. *Crop Prot.* 2001;20:49–56.
23. Hanson LE, Howell CR. Biocontrol efficacy and other characteristics of protoplast fusants between *Trichoderma koningii* and *T. Virens*. *Mycol Res.* 2002;106:321–8.
24. Pliego C, Ramos C, de Vicente A, Cazorla FM. Screening for candidate bacterial biocontrol agents against soilborne fungal plant pathogens. *Plant and Soil.* 2011;340:505–20.
25. Strakowska J, Błaszczyk L, Chełkowski J. The significance of cellulolytic enzymes produced by *Trichoderma* in opportunistic lifestyle of this fungus. *Journal of Basic Microbiology.* 2014;54 SUPPL.1.
26. Jeleń H, Błaszczyk L, Chełkowski J, Rogowicz K, Strakowska J. Formation of 6-n-pentyl-2H-pyran-2-one (6-PAP) and other volatiles by different *Trichoderma* species. *Mycol Prog.* 2014;13:589–600.
27. Gams W, Bissett J. Morphology and Identification of *Trichoderma*. In: *Trichoderma & Gliocladium, Enzymes, biological control and commercial applications.* 1998. p. 1 –34.
28. Reino JL, Guerrero RF, Hernández-Galán R, Collado IG. Secondary metabolites from species of the biocontrol agent *Trichoderma*. *Phytochemistry Reviews.* 2008;7:89–123.

29. Harman GE, Howell CR, Viterbo A, Chet I, Lorito M. Trichoderma species - Opportunistic, avirulent plant symbionts. *Nature Reviews Microbiology*. 2004;2:43–56.
30. Corley DG, Miller-Wideman M, Durley RC. Isolation and structure of harzianum a: A new trichothecene from trichoderma harzianum. *J Nat Prod*. 1994;57:422–5.
31. Gallo A, Mulè G, Favilla M, Altomare C. Isolation and characterisation of a trichodiene synthase homologous gene in Trichoderma harzianum. *Physiol Mol Plant Pathol*. 2004;65:11–20.
32. Błaszczyk L, Siwulski M, Sobieralski K, Lisiecka J, Jędrzycka M. Trichoderma spp. - Application and prospects for use in organic farming and industry. *Journal of Plant Protection Research*. 2014;54:309–17.
33. Howell CR. Mechanisms employed by Trichoderma species in the biological control of plant diseases: The history and evolution of current concepts. *Plant Disease*. 2003;87:4–10.
34. Khan J, Ooka JJ, Miller SA, Madden L V., Hoitink HAJ. Systemic resistance induced by Trichoderma hamatum 382 in cucumber against phytophthora crown rot and leaf blight. *Plant Dis*. 2004;88:280–6.
35. Brunner K, Zeilinger S, Ciliento R, Woo SL, Lorito M, Kubicek CP, et al. Improvement of the fungal biocontrol agent Trichoderma atroviride to enhance both antagonism and induction of plant systemic disease resistance. *Appl Environ Microbiol*. 2005;71:3959–65.
36. Woo SL, Lorito M. Exploiting the interactions between fungal antagonists, pathogens and the plant for biocontrol. *NATO Secur through Sci Ser A Chem Biol*. 2007;:107–30.
37. Shores M, Yedidia I, Chet I. Involvement of jasmonic acid/ethylene signaling pathway in the systemic resistance induced in cucumber by Trichoderma asperellum T203. *Phytopathology*. 2005;95:76–84.
38. Yedidia I, Benhamou N, Chet I. Induction of defense responses in cucumber plants (*Cucumis sativus* L.) by the Biocontrol agent Trichoderma harzianum. *Appl Environ Microbiol*. 1999;65:1061–70.
39. Ezziyyani M, Requena ME, Egea-Gilabert C, Candela ME. Biological control of Phytophthora root rot of pepper using Trichoderma harzianum and Streptomyces rochei in combination. *J Phytopathol*. 2007;155:342–9.
40. Etebarian HR, Scott ES, Wicks TJ. Trichoderma harzianum T39 and T. virens DAR 74290 as potential biological control agents for Phytophthora erythroseptica. *Eur J Plant Pathol*. 2000;106:329–37.
41. De Meyer G, Bigirimana J, Elad Y, Höfte M. Induced systemic resistance in Trichoderma harzianum T39 biocontrol of Botrytis cinerea. *Eur J Plant Pathol*. 1998;104:279–86.
42. Barari H. Biocontrol of Tomato Fusarium wilt by Trichoderma Species under in vitro and in vivo Conditions. *Cercet Agron Mold*. 2016;49:91–8.
43. Jayaraj J, Radhakrishnan N V., Velazhahan R. Development of formulations of Trichoderma harzianum strain M1 for control of damping-off of tomato caused by Pythium aphanidermatum. *Arch Phytopathol Plant Prot*. 2006;39:1–8.

44. Khan MR, Haque Z, Rasool F, Salati K, Khan U, Mohiddin FA, et al. Management of root-rot disease complex of mungbean caused by *Macrophomina phaseolina* and *Rhizoctonia solani* through soil application of *Trichoderma* spp. *Crop Prot.* 2019;119:24–9.
45. Nirmalkar VK, Tiwari RKS, Singh S. Efficacy of bio-agents against damping off in solanaceous crops under nursery conditions. *Int J Plant Prot.* 2018;11:1–9.
46. Bissett J, Gams W, Jaklitsch W, Samuels GJ. Accepted *Trichoderma* names in the year 2015. *IMA Fungus.* 2015;6:263–95.
47. Hermosa MR, Grondona I, Iturriaga EA, Diaz-Minguez JM, Castro C, Monte E, et al. Molecular characterization and identification of biocontrol isolates of *Trichoderma* spp. *Appl Environ Microbiol.* 2000;66:1890–8.
48. Baiyee B, Ito S ichi, Sunpapao A. *Trichoderma asperellum* T1 mediated antifungal activity and induced defense response against leaf spot fungi in lettuce (*Lactuca sativa* L.). *Physiol Mol Plant Pathol.* 2019;106:96–101.
49. Veenstra A, Rafudeen MS, Murray SL. *Trichoderma asperellum* isolated from African maize seed directly inhibits *Fusarium verticillioides* growth in vitro. *Eur J Plant Pathol.* 2019;153:279–83.
50. Marcello CM, Steindorff AS, da Silva SP, Silva R do N, Mendes Bataus LA, Ulhoa CJ. Expression analysis of the  $\text{exo-}\beta\text{-1,3}$ -glucanase from the mycoparasitic fungus *Trichoderma asperellum*. *Microbiol Res.* 2010;165:75–81.
51. Wu Q, Zhang L, Xia H, Yu C, Dou K, Li Y, et al. Omics for understanding synergistic action of validamycin A and *Trichoderma asperellum* GDFS1009 against maize sheath blight pathogen. *Sci Rep.* 2017;7.
52. Monte E. Understanding *Trichoderma*: between biotechnology and microbial ecology. *International microbiology: the official journal of the Spanish Society for Microbiology.* 2001;4:1–4.
53. Dubey SC, Suresh M, Singh B. Evaluation of *Trichoderma* species against *Fusarium oxysporum* f. sp. *ciceris* for integrated management of chickpea wilt. *Biol Control.* 2007;40:118–27.
54. Larkin RP, Fravel DR. Efficacy of various fungal and bacterial biocontrol organisms for control of fusarium wilt of tomato. *Plant Dis.* 1998;82:1022–8.
55. Datnoff LE, Nemecek S, Pernezny K. Biological control of fusarium crown and root rot of tomato in Florida using *Trichoderma harzianum* and *Glomus intraradices*. *Biol Control.* 1995;5:427–31.
56. Shabir-U-Rehman, Lawrence R, Kumar EJ, Badri ZA. Comparative efficacy of *Trichoderma viride*, *T. harzianum* and carbendazim against damping-off disease of cauliflower caused by *Rhizoctonia solani* Kuehn. *J Biopestic.* 2012;5:23–7.
57. Nazir B, Simon S, Das S, Soma R. Comparative efficacy of *Trichoderma Viride* and *T. Harzianum* in management of *Pythium Apanidermatum* and *Rhizoctonia Solani* causing root-rot and damping-off diseases. *J Plant Dis Sci.* 2011;6:60–2.
58. Kipngeno P, Losenge T, Maina N, Kahangi E, Juma P. Efficacy of *Bacillus subtilis* and *Trichoderma asperellum* against *Pythium aphanidermatum* in tomatoes. *Biol Control.* 2015;90:92–5.

59. Singh SP, Singh HB, Singh DK, Rakshit A. Trichoderma-mediated enhancement of nutrient uptake and reduction in incidence of *Rhizoctonia solani* in tomato. *Egypt J Biol.* 2014;16:29.
60. Azarmi R, Hajieghrari B, Giglou A. Effect of trichoderma isolates on tomato seedling growth response and nutrient uptake. *African J Biotechnol.* 2011;10:5850–5.
61. Patel S, Saraf M. Biocontrol efficacy of *Trichoderma asperellum* MSST against tomato wilting by *Fusarium oxysporum* f. sp. *lycopersici*. *Arch Phytopathol Plant Prot.* 2017;50:228–38.
62. Jataraf J, Radhakrim NV, Hannk P, Sakoof R. Biocontrol of tomato damping-off caused by *Pythium aphanidermatum*. *Biocontrol.* 2005;15:55 – 65.
63. Manoranjitham SK, Prakasam V, Rajappan K, Amutha G. Effect of two antagonists on damping off disease of tomato. *Indian Phytopathol.* 2000;53:441–443.
64. Sharma KK, Zaidi NW, Singh US. Effect of biological seed treatment on seed germination and growth promotion of paddy, tomato and mustard. *Vegetos.* 2012;25:375–86.
65. Dukare AS, Prasanna R, Chandra Dubey S, Nain L, Chaudhary V, Singh R, et al. Evaluating novel microbe amended composts as biocontrol agents in tomato. *Crop Prot.* 2011;30:436–42.
66. Kalay AM, Tuhumury GN, Pesireron N, Talaharuruson A. Control of Damping off and Increased Growth of Tomato Seeds by Utilizing *Trichoderma harzianum* Based on Solid Organic Materials. *Agrologia.* 2019;8.
67. Uddin MM, Akhtar N, Faruq AN. Effect of trichoderma *harzianum* and some selected soil amendments on damping-off disease of eggplant and tomato. 1; *Sci Found.* 2009;7:117–26.
68. Ryu J, Jin R, Kim Y, Lee H, Kim K. Biocontrol of Damping-Off ( *Rhizoctonia solani* ) in Cucumber by *Trichoderma asperellum* T-5. *Korean J Soil Sci Fertil.* 2006;39:185–94.
69. Nzanza B, Marais D, Soundy P. Response of tomato (*Solanum lycopersicum* L.) to nursery inoculation with *Trichoderma harzianum* and arbuscular mycorrhizal fungi under field conditions. *Acta Agric Scand Sect B Soil Plant Sci.* 2012;62:209–15.
70. Tsrer L, Barak R, Sneh B. Biological control of black scurf on potato under organic management. *Crop Prot.* 2001;20:145–50.
71. Naseby DC, Pascual JA, Lynch JM. Effect of biocontrol strains of *Trichoderma* on plant growth, *Pythium ultimum* populations, soil microbial communities and soil enzyme activities. *J Appl Microbiol.* 2000;88:161–9.
72. Patel R, Patel D. Screening of *Trichoderma* and antagonistic analysis of a Potential Strain of *Trichoderma* for Production of a Bioformulation. *Int J Sci Res Publ.* 2014;4:1–6.

## Tables

Table 1  
Dose and methods of application for the management of  
tomato damping-off

Treatment	Description	Dose
T1	Soil application of <i>T. asperellum</i>	1 ml /l
T2	Soil drenching of <i>T. asperellum</i>	1 ml /l
T3	Seed treatment <i>T. asperellum</i>	1 ml /kg
T4	FYM <i>T. asperellum</i>	250 ml / kg
T5	Vermicompost <i>T. asperellum</i>	250 ml / kg
T6	Carbendazim drenching	2.5 g /l
T7	Untreated control	-

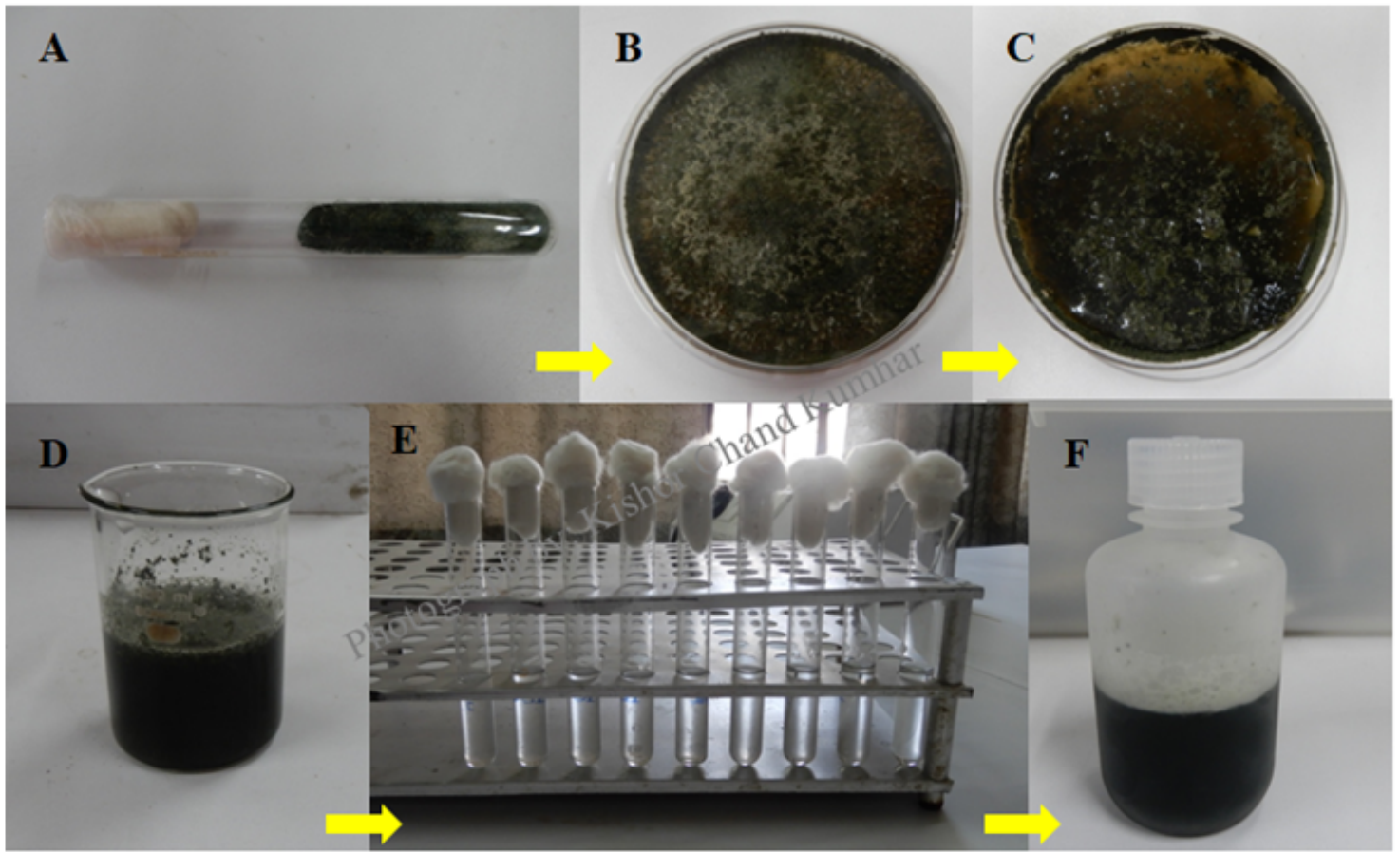
Table 2  
Evaluation of *Trichoderma asperellum* for management of seedling disease of tomato under  
nursery (2018-19)

Treatment	Germination (%)	Damping-off incidence (%)
T1-Soil application of <i>T. asperellum</i>	75.75 (60.51 ± 1.00)	11.37 (19.64 ± 0.92)
T2-Soil drenching of <i>T. asperellum</i>	63.88 (53.10 ± 2.08)	20.38 (26.78 ± 1.16)
T3-Seed treatment <i>T. asperellum</i>	65.50 (54.03 ± 1.23)	19.19 (25.86 ± 1.59)
T4-FYM <i>T. asperellum</i>	72.00 (58.06 ± 1.03)	13.36 (21.43 ± 0.42)
T5-Vermicompost <i>T. asperellum</i>	88.75 (70.86 ± 2.69)	10.93 (19.21 ± 1.14)
T6-Carbendazim drenching	73.00 (58.79 ± 2.10)	12.08 (20.03 ± 2.22)
T7-Untreated control	60.13 (50.85 ± 1.54)	26.98 (31.22 ± 1.49)
CD (p < 0.05)	5.27	3.55
<b>*Average of 3 replications, figures in parenthesis is angular transformed values ± SE</b>		

Table 3  
Bioefficacy of *Trichoderma* liquid formulation against tomato damping-off in the nursery at the vegetable farm (2019-20)

Treatment	Germination (%)	Damping-off incidence (%)
T1-Soil application of <i>T. asperellum</i>	83.17 (65.86 ± 1.89)	8.38 (16.72 ± 1.50)
T2-Soil drenching of <i>T. asperellum</i>	70.83 ( 57.34 ± 1.83)	15.33 (23.03 ± 0.74)
T3-Seed treatment <i>T. asperellum</i>	71.00 (57.48 ± 2.34)	14.43 (22.28 ± 1.08)
T4-FYM <i>T. asperellum</i>	75.67 (60.45 ± 1.16)	11.68 (19.95 ± 0.53)
T5-Vermicompost <i>T. asperellum</i>	89.50 (71.19 ± 1.64)	7.82 (16.09 ± 1.71)
T6-Carbendazim drenching	81.00 (64.21 ± 1.65)	8.54 (16.90 ± 1.26)
T7-Untreated control	66.83 (54.83 ± 1.24)	20.21 (26.69 ± 0.58)
C.D. (p < 0.05)	3.52	3.78
<b>*Average of 3 replications, figures in parenthesis is angular transformed values ± SE</b>		

## Figures



**Figure 1**

Steps of product formulation: A-Pure culture slant, B- Culturing on PDA plate, C- Biomass scrapping, D- Biomass collection, E-Serial dilution and F- Liquid formulation

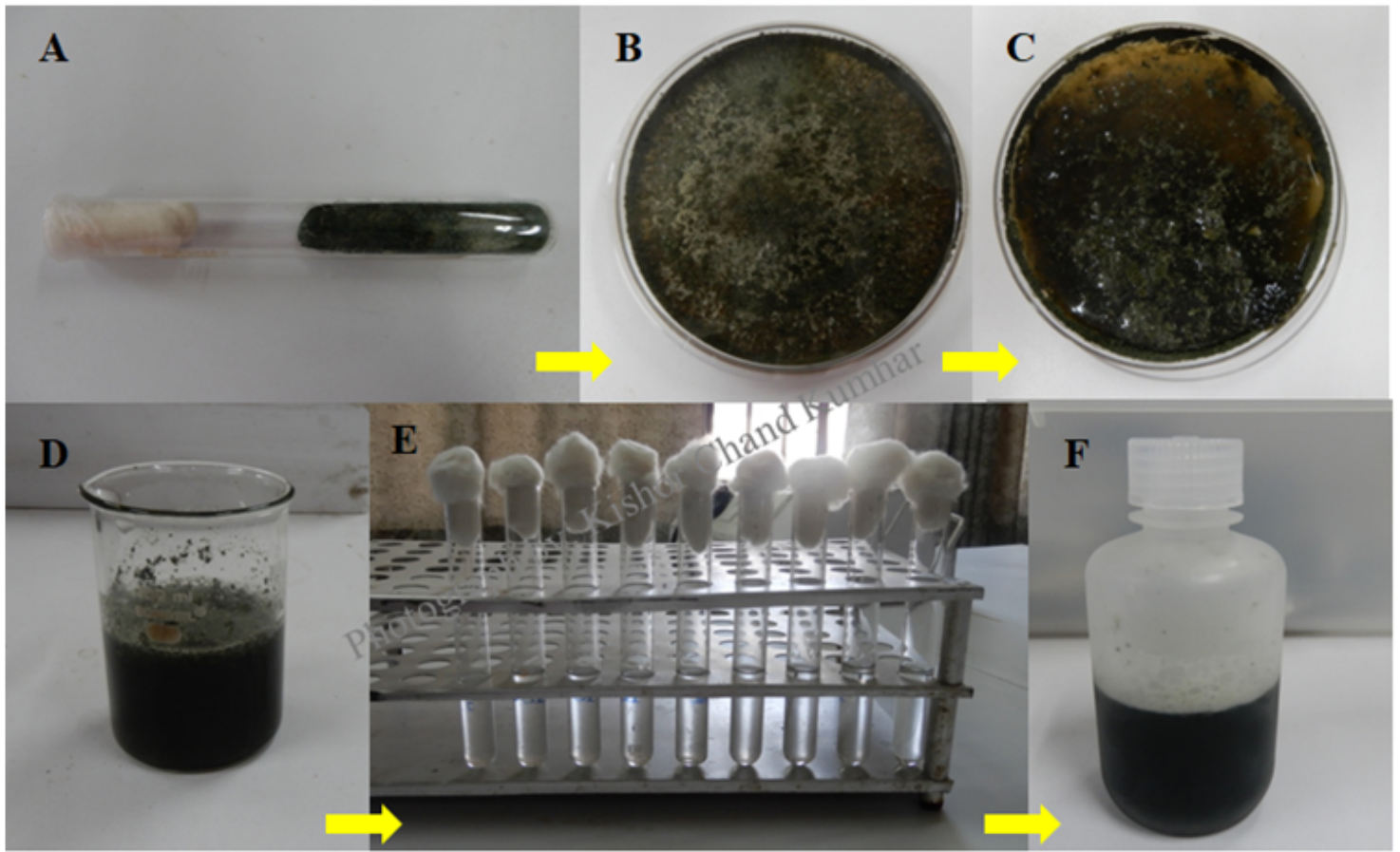


Figure 1

Steps of product formulation: A-Pure culture slant, B- Culturing on PDA plate, C- Biomass scrapping, D- Biomass collection, E-Serial dilution and F- Liquid formulation



Figure 2



Performance of *T. asperellum* in tomato nursery: A- post emergence damping off, B-vegetative growth (arrow indicating control bed)



Figure 2

Performance of *T. asperellum* in tomato nursery: A- post emergence damping off, B-vegetative growth (arrow indicating control bed)

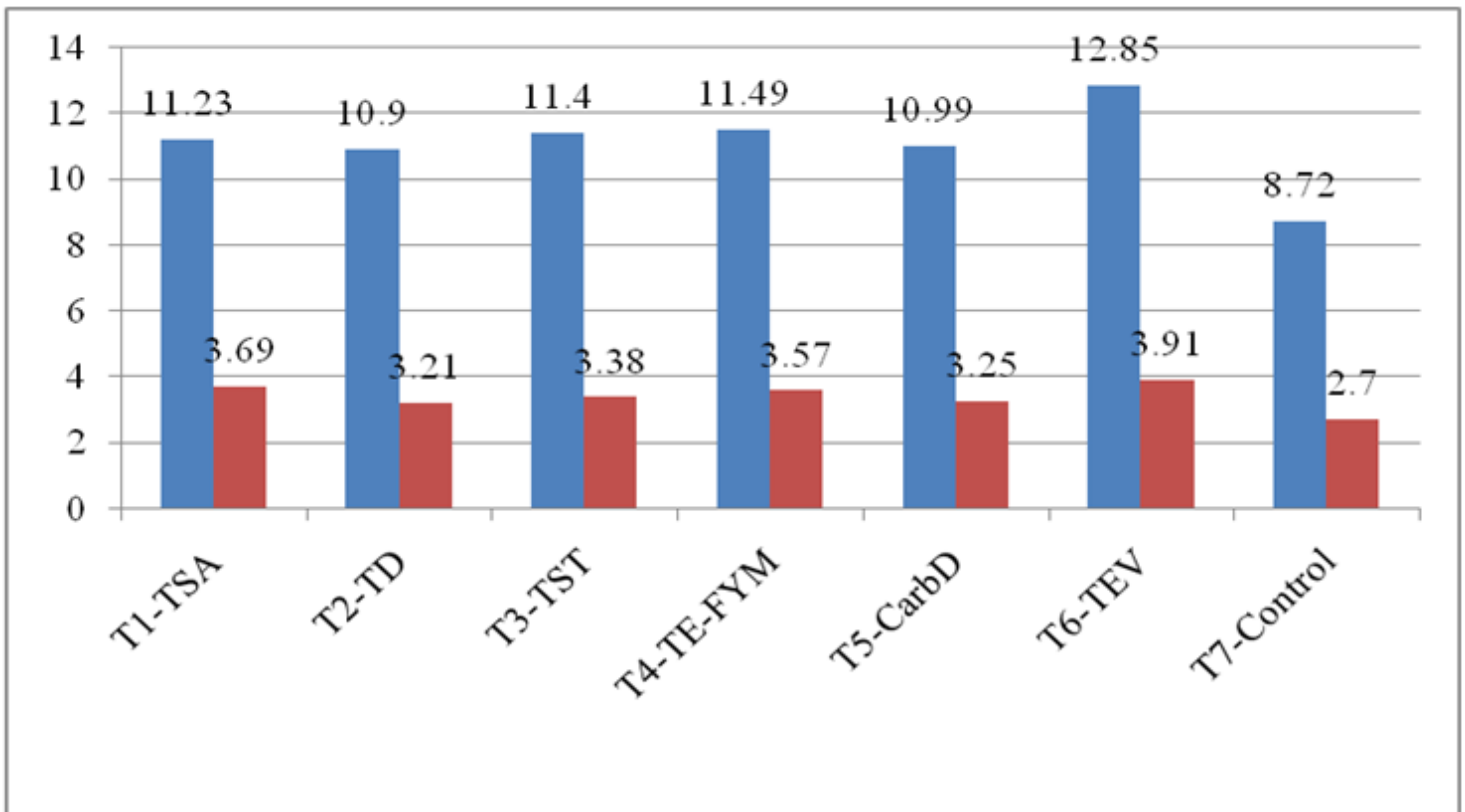
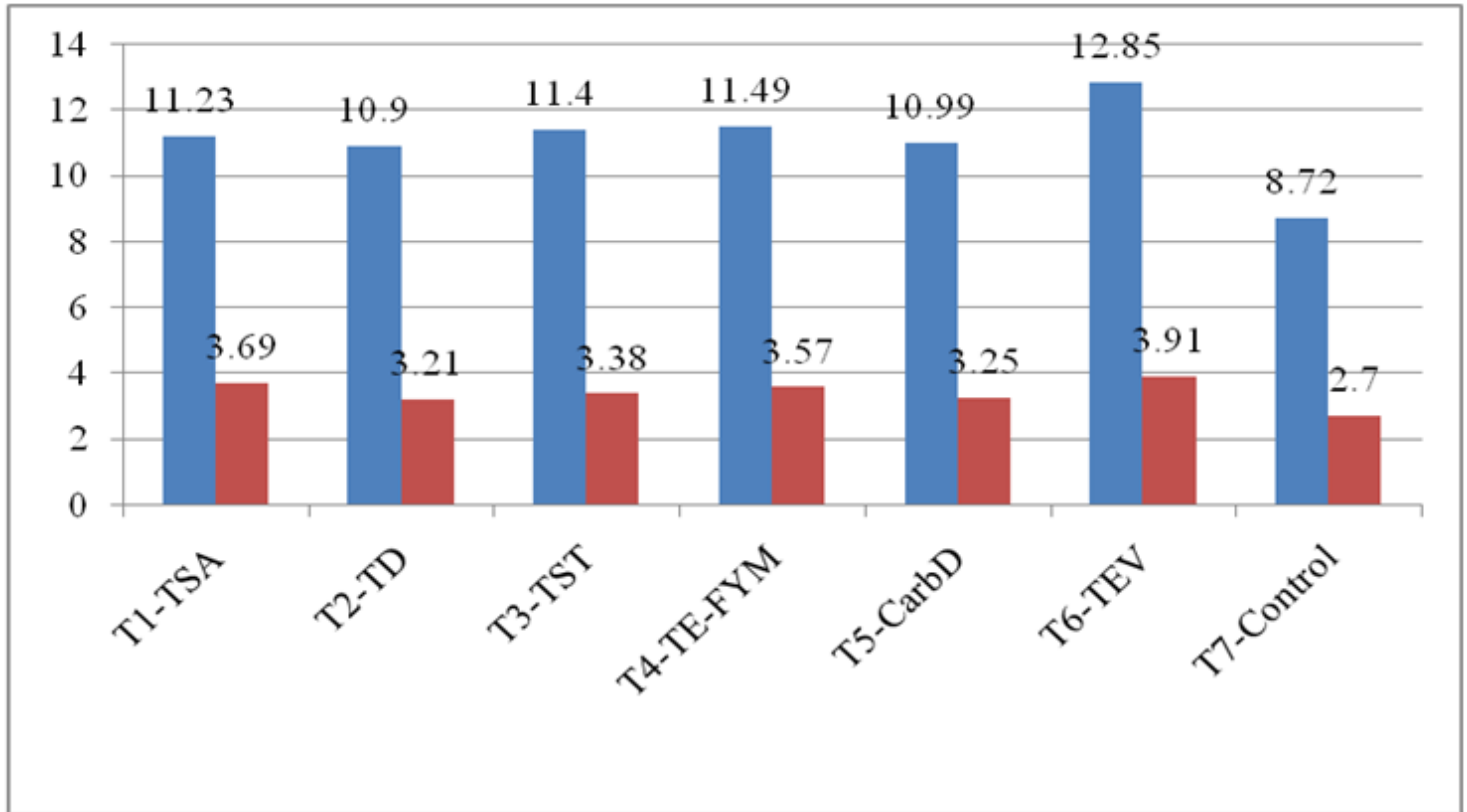


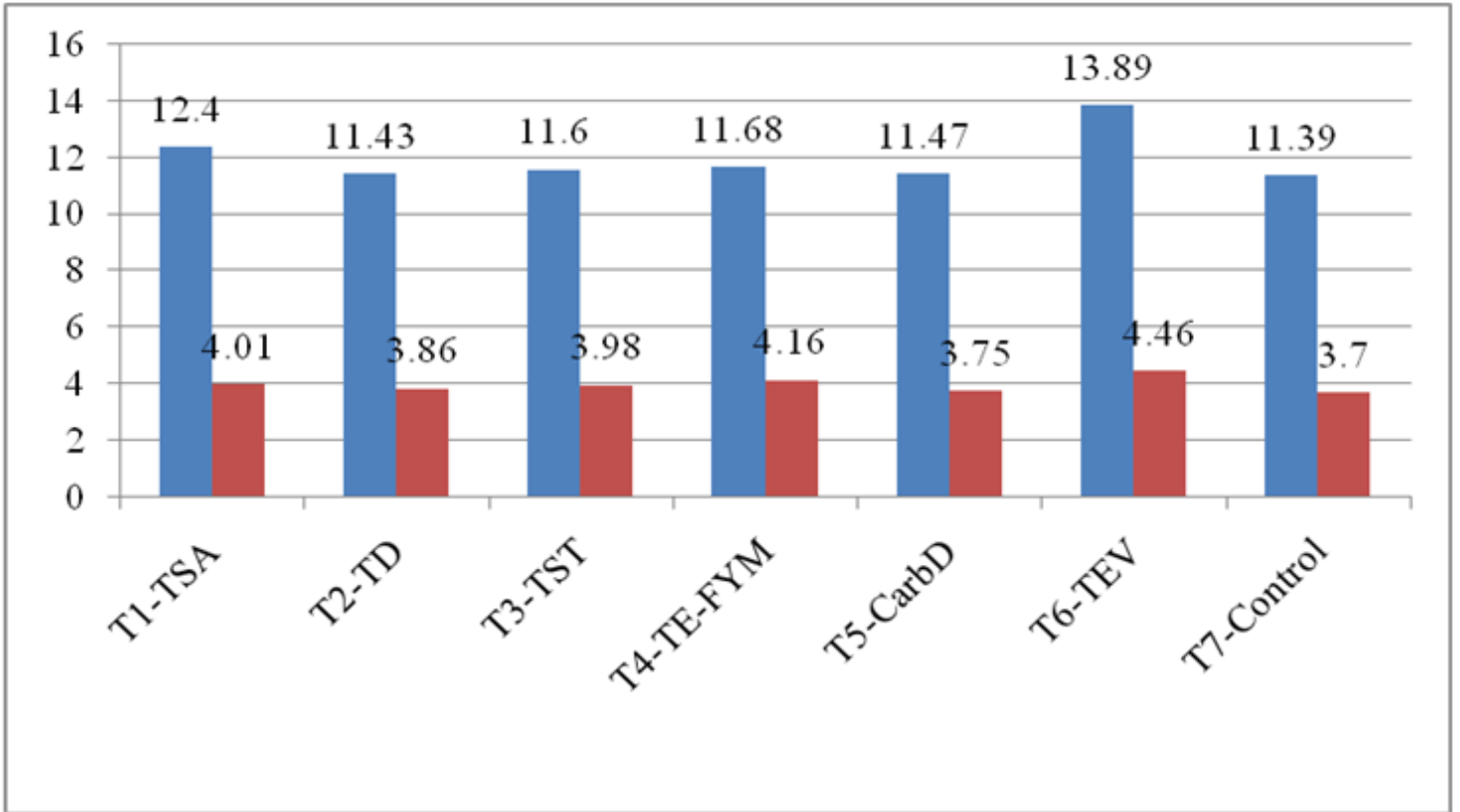
Figure 3

Effect of Trichoderma formulation on shoot and root growth of tomato in nursery (2018-19) Legends: Shoot length (cm) Root length (cm)



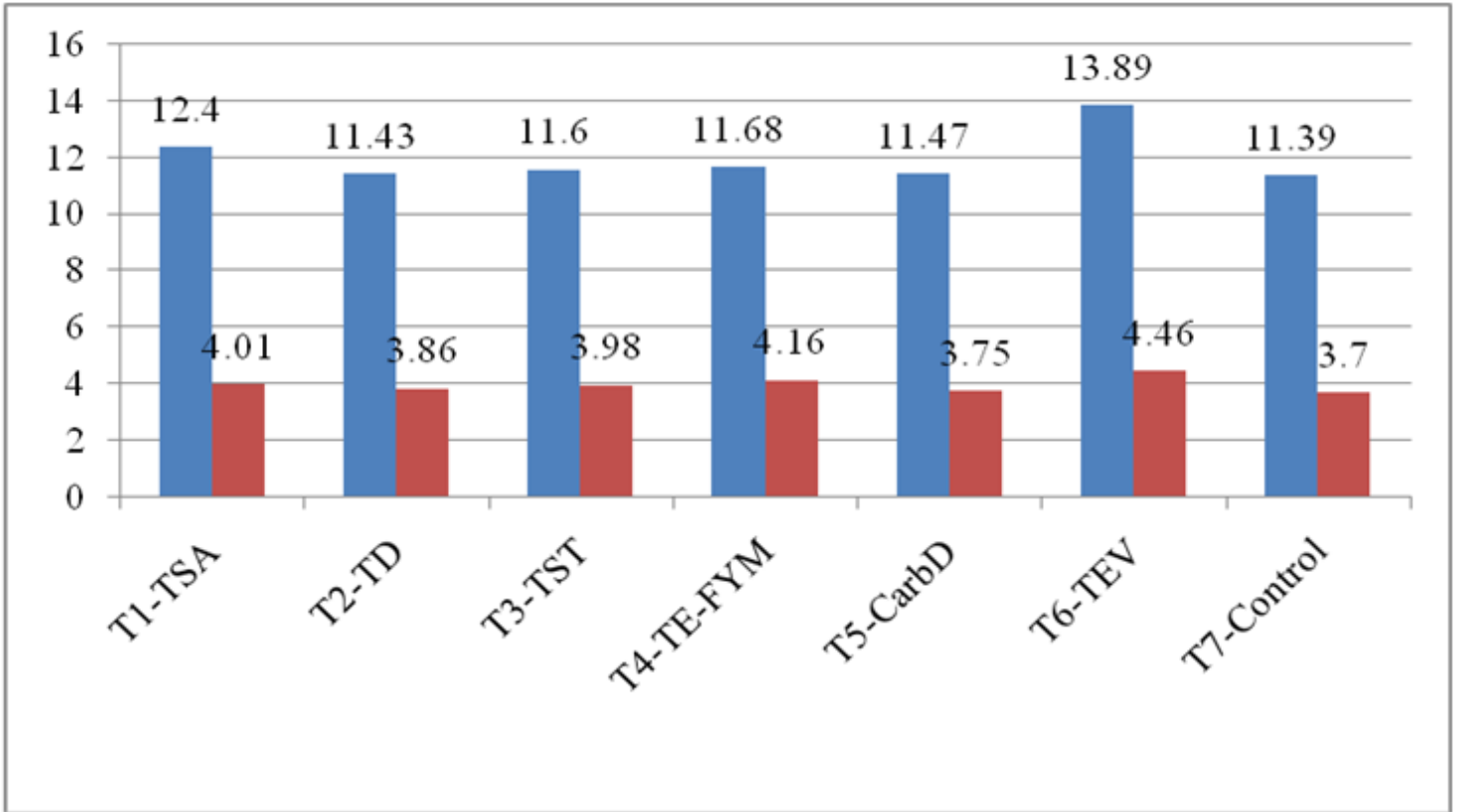
**Figure 3**

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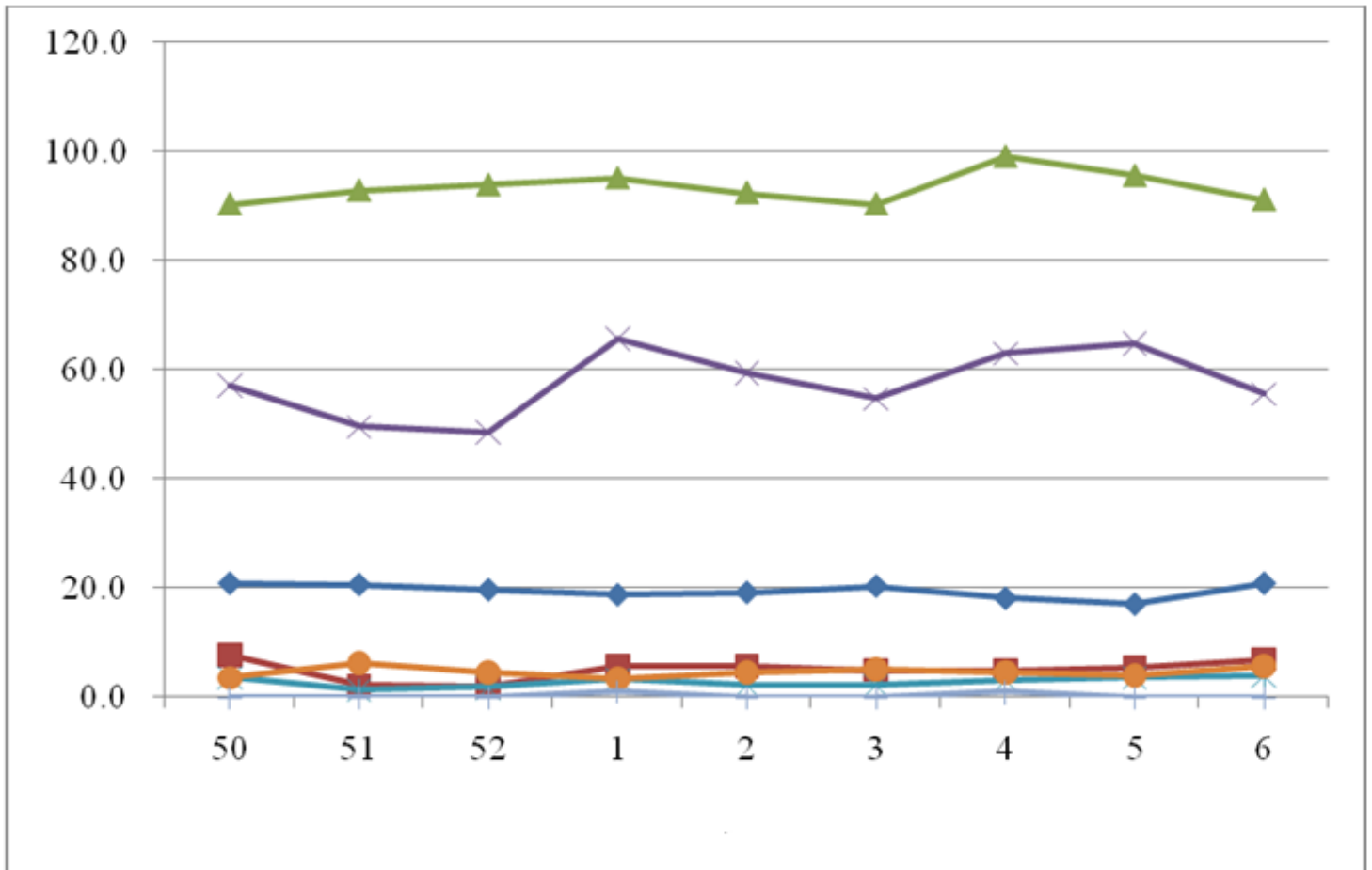
**Figure 4**

Effect of Trichoderma formulation on shoot and root growth of tomato in nursery (2019-20) Shoot length (cm) Root length (cm)



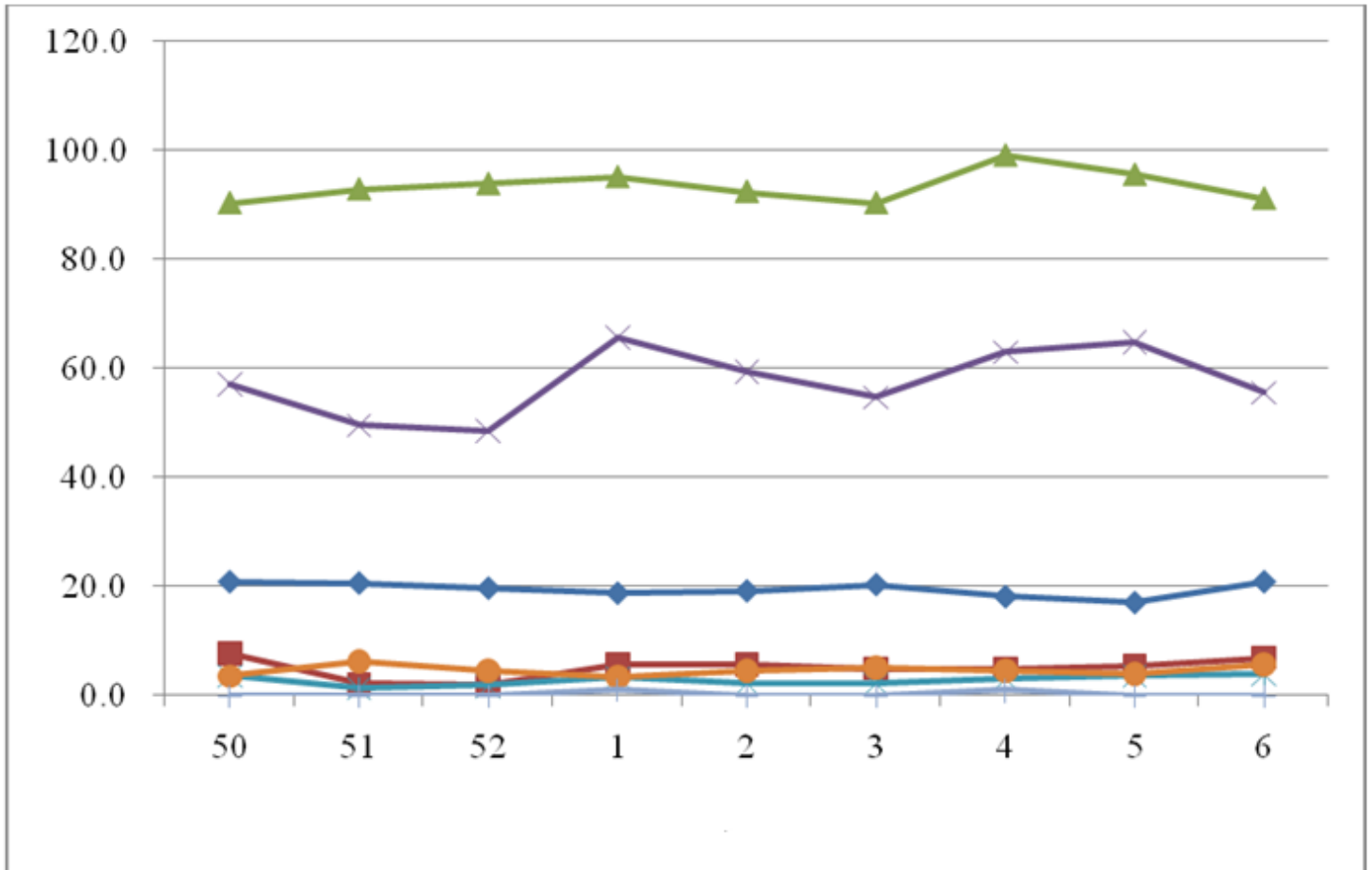
**Figure 4**

Effect of Trichoderma formulation on shoot and root growth of tomato in nursery (2019-20) Shoot length (cm) Root length (cm)



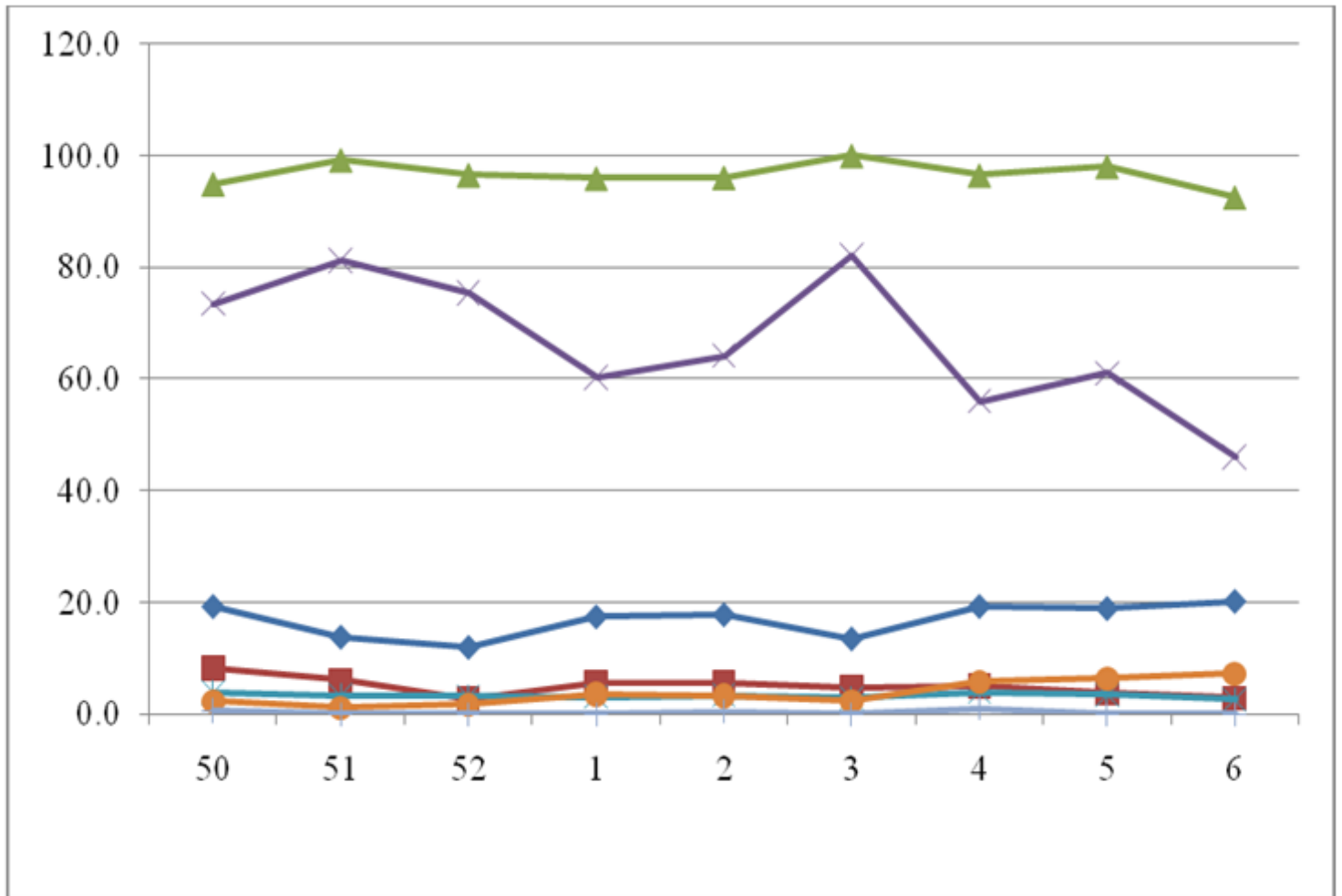
**Figure 5**

Weather conditions during first year (2018-19)



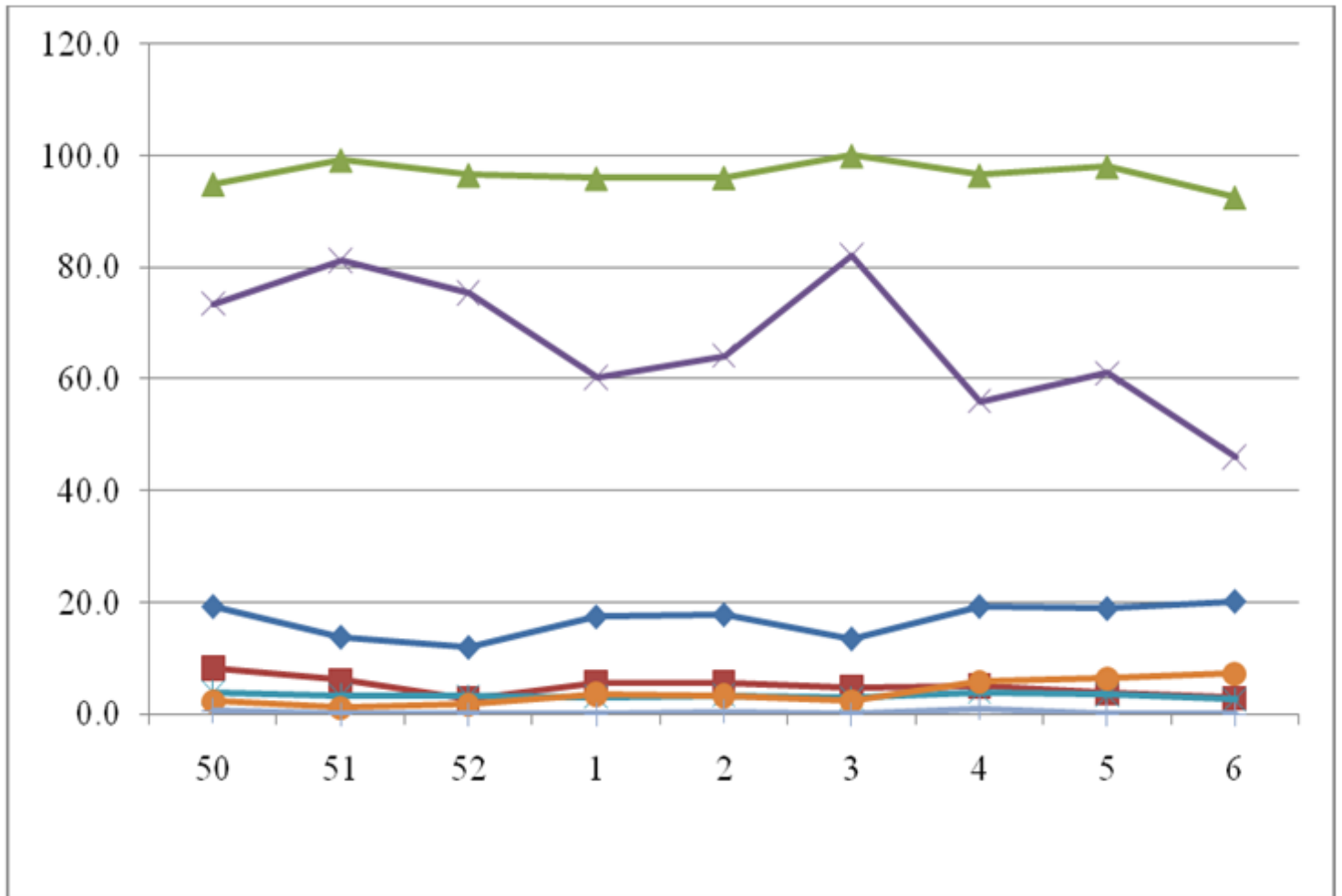
**Figure 5**

Weather conditions during first year (2018-19)



**Figure 6**

Weather conditions during second year (2019-20)



**Figure 6**

Weather conditions during second year (2019-20)