

MiR-21-3p promotes hepatocellular carcinoma progression through regulating *Smad7/Yap1*.

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Abstract

Background Hepatocellular carcinoma (HCC) remains the global burden due to its high prevalence and mortality. Emerging evidence confirms that microRNAs (miRNAs) play a vital role in cancer initiation and progression. MiRNAs are widely involved in the regulation of signaling pathways by targeting downstream genes. MiR-21-3p as a liver-enriched miRNA has not been fully investigated. Abnormal activation of TGF- β transduction pathway promoted by deletion of *Smad7* matters since HCC occurrence. While the relation between miR-21-3p and *Smad7* has not yet been confirmed. We aimed to explore the influence of miR-21-3p on HCC initiation and progression by targeting *Smad7* and further facilitating the expression of *Yap1*.

Methods MicroRNA (miRNA) microarray analysis was performed for miRNA screening. Dual-luciferase assay was adopted for target verifying. The expressions of miRNA and related genes were quantified by qRT-PCR, western blotting, and immunohistochemical staining. Flow cytometry and transwell assay were used to discover cell apoptosis, invasion and metastasis abilities. Rat models were established to explore the axis's role in hepatocarcinogenesis. Bioinformatics analysis was performed for analyzing clinical significance.

Results MiR-21-3p was significantly increased in HCC, indicating a poor overall survival (OS) rate. High miR-21-3p was associated with advanced stages ($P=0.029$), especially T stages ($P=0.026$). Low *Smad7*/high *Yap1* was verified in HCCs and rat models. *Smad7* was proved to be the direct target of miR-21-3p. MiR-21-3p's effect on tumor malignant phenotypes and promotion of *Yap1* could be partly reversed through transfecting *Smad7*. Overexpressed *Yap1* promoted the downstream effector connective tissue growth factor (CTGF). Co-survival analysis indicated that lower miR-21-3p/higher *Smad7* ($P=0.0494$) and lower miR-21-3p/lower *Yap1* group ($P=0.0379$) patients had better OS rates. GSVA analysis of miR-21-3p and *Smad7* related gene sets displayed strong relation with TGF- β signaling pathway in HCC.

Conclusions MiR-21-3p promotes HCC migration and invasion via directly inhibiting *Smad7* and further improving the expression of *Yap1*.

Introduction

With a rising global incidence and the leading cause of cancer-related death status, hepatocellular carcinoma (HCC) is still lacking effective treatment (1). More than half of HCC patients are already in advanced stages of liver cancer when first diagnosed (2). For those patients with unresectable HCC, sorafenib and lenvatinib as multi-kinases inhibitors are considered as systemic efficiency drugs (3), while their performance on prolonging patient survival rate is far from satisfied (4). It is time to explore specific mechanisms of occurrence and progression of HCC to find effective clinical treatments.

MicroRNAs (miRNAs), the small endogenous non-coding ribonucleic acid molecular family, are mainly involved in RNA silencing and regulation of gene expressions through complementary combination with

the 3'-UTR region of genes (5). MiRNAs are considered to be biomarkers of diagnosis and prognosis since discovered (6). MiR-21-3p was demonstrated to inhibit cell apoptosis rate in liver cancer stem cells via directly regulating PTEN (7). Our miRNA microarray analysis results displayed that miR-21-3p is significantly upregulated in HCC. Previous studies confirmed that miR-21-3p could influence hepatoma cell growth through directly down-regulating adenosyltransferases 2A and 2B (MAT2A/MAT2B) (8). However, more important targets wait to be identified to elucidate the indispensable role of miR-21-3p in HCC.

Smad7 (I-Smad) is the negative regulator of the TGF- β signaling transduction pathway through binding TGF- β Receptor α (T β R α) and interfering with the recruitment of *Smad2/Smad3* (R-smad), blocking functional Smad complex from interacting with DNA in the nucleus (9,10). As a "double-edged sword" involving in maintaining cell normal proliferation and differentiation, the TGF- β signaling pathway plays an irreplaceable role in tumor formation and progression (11). Dysfunction of *Smad7* and excessive activation of the TGF- β signaling pathway were commonly seen in various kinds of cancers(12,13). T Feng's team has verified that the deletion of *Smad7* could accelerate the progress of HCC in mice models through activating the STAT3 signaling pathway (14). Loss-of function of *Smad7* promoted HCC cell proliferation, accelerated G1-S transition and reduced cell apoptosis in vivo (15). The role of *Smad7*'s abnormal deletion in HCC needs more depth researches.

Yap1 (Yes-associated protein 1) is the "star nuclear effector" of the Hippo pathway, which consists of a series of kinases (16). *Yap1* is in hyperactive status and is considered as an oncogene in several kinds of solid tumors, including liver cancer (17). The first discovery of the crosstalk between Hippo and the TGF- β signaling pathway was that cytoplasmic *Yap1* protein helped inhibit part of the signal transduction of TGF- β in COS-7 cell (a CV-1 African green monkey fibroblast cell line by transformation with a mutant strain of Simian Virus 40 (SV40)) via enhancing *Smad7* binding to T β R α , and *Yap1* was demonstrated to be a novel *Smad7*-interacting protein (18). Nevertheless, the close relationship between *Smad7* and *Yap1* in HCC remains unclear.

The present study described the overexpression of miR-21-3p in human HCC. We first proposed that MiR-21-3p could facilitate the expression of *Yap1* and promote malignant phenotypes progression via directly targeting *Smad7* in HCC.

Methods

Clinical tissues and cell culture

Ten paired HCC tissues and corresponding background livers (BLs) were obtained from HCC patients undergoing partial hepatectomy between January 2019 and December 2020. After surgical resection, the specimens were immediately snap-frozen in liquid nitrogen. RNA-later (#AM7201, Thermofisher, USA) was added to the cryogenic vials to prevent RNA degradation for subsequent extraction. The collection and usage of these samples were followed ethical and institutional guidelines and implement it after organizing informed consent from donors. The study was approved by the local Research Ethics

Committee at Zhongnan Hospital of Wuhan University (Approval No.2018078). Cell lines (L02, Huh7, HepG2, HCCL-M3) involved in this experiment were purchased from Stem Cell Bank, Chinese Academy of Sciences. Mixture of Dulbecco's Modified Eagle's Medium (Gibco, USA) and 10% fetal bovine serum (Gibco, Thermo Fisher, USA) and 1% penicillin-streptomycin (Gibco, USA) was used to culture cells. All cells were grown in humidified 5% CO₂ at 37°C.

MiRNA microarray analysis

The microarray data adopted for analysis have been described in detail before (19). Data have been uploaded in NCBI's Gene Expression Omnibus (GEO) public database <http://www.ncbi.nlm.nih.gov/geo/>, accession number, GSE20077).

RNA extraction, Reverse transcription and quantitative real-time PCR

Total RNA extraction from tissues and cells were using Triol Reagent (Invitrogen, USA) following the instructions. The quality and concentration of RNA were detected by Nanodrop 2000 (ThermoFisher, USA). cDNA was obtained through a reverse transcription reaction with kits (Toyobo, Japan). The expression of miRNAs and genes was examined using UltraSYBR Mixture (CWbio, Beijing, China) on the CFX96Touch Real-Time PCR system (Biorad, USA). The mRNA levels of miR-21-3p and genes were normalized to U6 (Ribobio, Guangzhou, China) and *GAPDH*, respectively. The relative expression ratio was calculated with the $2^{-\Delta\Delta Cq}$ method. Primers used listed here:

Smad7: 5'-TTCAGGACCAAACGATCTGCG-3'(sense);

5'-GATGGTGGTGACCTTTGGCAC-3'(antisense);

Yap1: 5'-GAACTCGGCTTCAGGTCCTC-3'(sense);

5'-GGTTCATGGCAAAACGAGGG-3'(antisense);

LATS2:5'-TGGAATGCCAACAATGTAGCG-3' (sense);

5'-ATTATCACTCTCTCCAGGGGCG-3' (antisense);

CTGF: 5'-AGAGGGCTGTCCGGCG-3' (sense)

5'-CACAGGAGCTGGTGTTCAT-3' (antisense);

GAPDH: 5'-AGAAGGCTGGGGCTCATTTG-3'(sense);

5'-GCAGGAGGCATTGCTGATGAT-3'(antisense).

MiRNA related reagents, plasmids and cell transfection

The precursor and inhibitor of miR-21-3p, mimic-NC and inhibitor-NC were obtained from Ribobio (Guangzhou, China). Inhibitors of miR-21-3p were the complementary strand of miR-21-3p and was dedicated to compete the binding sites between miR-21-3p and target genes, but would not decrease the mRNA expression level of miRNAs. The final concentration of these reagents was 50 nM. *Smad7* expression vector (P-*Smad7*) was constructed using GV141 vector by Genechem (Shanghai, China) and plasmids-NC (P-NC) served as an experimental control. All transfection experiments were performed using Lipofectamine 2000 (Invitrogen, Carlsbad, CA) following the guidance.

Western blot

Proteins collection using lysis buffer (Byotime, China) containing protease inhibitor (PMSF, 1:100). Centrifuged at 15000rpm, 20min at 4°C and then transferring supernatant to new tubes. Concentrations of protein were detected using a BCA protein kit (Byotime, China). Mixing lysis buffer with protein loading buffer, denaturing protein at 100°C for 5min. An equal amount of proteins were separated by SDS-polyacrylamide gels and then transferred to polyvinylidene fluoride (PVDF) membranes (Millipore, USA). Blocking for 2h with 5% skimmed milk. The membrane was incubated with primary antibodies overnight at 4°C. Then reacted with second antibodies for 1.5h at room temperature. The protein bands were captured by enhanced chemiluminescence (ECL) kit (Servicebio, Wuhan, China) using GENESys (Synoptics Ltd, China). Anti-*Smad7* (#25840-1-AP), anti-*Yap1* (#14074), anti-E-cad (#3195), anti-N-cad (#131161), anti-Vimentin (#5741), anti-LATS2 (#5888) and anti-CTGF (#86641) were purchased from CST (USA). Anti-Bcl2 (#ab182858), anti-Bax (#ab32503) and anti-GAPDH (#ab8245) were purchased from Abcam. Goat anti-mouse IgG-horseradish peroxidase (#A23220) and goat anti-mouse IgG-horseradish peroxidase (#A25012) were come from Abbkine.

Dual-Luciferase reporter assay

PmiR-GLO-*Smad7* vector was constructed, which consisted of predicted binding sites that were mutated and ligated between the PmeI and XbaI restriction enzymes sites of pmirGLO Dual-Luciferase miRNA Target Reporter Vector (#E1300, Promega, USA). *Smad7* 3'UTR region contains two putative binding sites for miR-21-3p with the seed regions at 1428-1434 and 1445-1451. The strand of mutant 3 (Mut 3) mutated two binding sites simultaneously. The wild-type (WT) and mutant *Smad7* 3'-UTR luciferase reporter plasmids (0.25µg/ well) were co-transfected into cell huh-7 with miR-21-3p mimics or miR-NC (50nM, 0.15µl/ well) respectively. Dual-Glo Luciferase Assay System (#E2920, Promega, USA) was employed to check firefly luciferase and renilla luciferase activity after 24h transfection, respectively. Firefly luciferase activity was normalized to renilla luciferase activity. Data were collected using Enspire 2300 (PerkinElmer). Six repetitions per group were calculated.

Cell apoptosis assay and flow cytometry

A total of 4×10^5 cells were transfected with miR-21-3p mimics with/or *Smad7* plasmids. After 24h, collecting (including dead cells in the culture medium) and processing cells according to the instructions of Annexin V-FITC/PI Double stain apoptosis detection kit (#4101-2, Bestbio, Shanghai, China). Collecting

the fluorescence intensity on the flow cytometer to check the apoptotic rate (Cytoflex Beckman, China) immediately.

Cell invasion and metastasis assay

The procedure for processing cell Huh-7 was the same as the cell apoptosis experiment. Huh-7 was harvested at 24h post-transfections. Suspending cells with 5% FBS culture medium and seeding 5×10^4 cells each upper chamber (6.5 mm. in diameter, 8.0 μ m pore size, Corning, USA). The lower chamber was filled with 600 μ l medium (15% FBS). Cells were fixed with 4% paraformaldehyde 20min at room temperature, stained with 0.1% crystal violet (Sigma-Aldrich, USA). The upper cells were wiped using cotton swabs. Selecting 100 \times and 200 \times fields and taking photos randomly under an inverted microscope (Olympus IX3). The difference between cell invasion and migration assay was that in the latter one upper chamber needed to be pretreated with 0.3% Matrigel matrix (#356234, Corning, USA), 37°C, 4h. Three independent experiments were performed.

Animal experiments

The establishment of Wistar rat models were detailed described before, and grouping of these rats were according to the Metavir score system (20,21). Here was brief introduction. Wistar rats (male, 7-8 weeks old, 200-220g) were purchased from Beijing Vital River Laboratory Animal Technology Co., Ltd. All animal handling and experimental procedures were approved by the Animal Care and Use Committee of Wuhan University following the Animal Experiment/Animal Biosafety Level-III Laboratory Guidelines. Rats were injected 40% carbon tetrachloride (CCl₄) dissolved in maize oil (1.5ml/kg) every twice a week. At 8, 14, 18 weeks, liver fibrosis, cirrhosis and liver cancer were formed, respectively. Hematoxylin-eosin (HE) staining and masson staining which is more conducive to observe the degree of fiber crosslinking of rat models were displayed in Figure S1A.

Immunohistochemistry (IHC)

Immunohistochemical staining of patient tissues and animal tissues was performed by Servicebio Company, Wuhan, China, and photographed by an inverted microscope (Olympus IX3). Positively stained cells were counted in at least five fields from each area with 400 \times magnifications. Primary antibodies listed here:

Anti-*Smad7*: (#25840-1-AP, Proteintech): 1:200; anti-*Yap1* (#14074, CST): 1:400.

Immunofluorescence (IF)

The distribution of microfilaments and cytoskeleton was clearly shown through fluorescently labeled phalloidin staining. Cells were cultured in soft and still matrixes 24h. Fixed with 4% paraformaldehyde 20min at room temperature. Phalloidin dilution rate: 1:500. Incubation at 37°C for 1h. Nuclear staining with DAPI for 5min. Images were processed with an inverted microscope (Olympus IX3).

Gene set variation analysis (GSVA) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways analysis with miR-21-3p/Smad7 expression

GSVA was a useful tool provided by R (3.5.2) to explore the potential biological process and enriched KEGG pathways of research objects according to its high or low expression. The results were presented in the form of intuitive volcano maps and specific heat maps. Gene terms with $|\log FC| \geq 0.1$ and $P < 0.05$ were considered statistically significant. The KEGG gene sets (c2.cp.kegg.v6.2.symbols.gmt) downloaded from the Molecular Signatures Database–MsigDB (<http://www.broad.mit.edu/gsea/msigdb/index.jsp>) were used for enrichment analysis.

Bioinformatics data

MicroRNA Target Prediction Database (miRDB), PicTar and TargetScan databases were used for searching miRNA targets and predicting binding sites. Cohort data including 376 HCC patients RNA-seq and clinical characteristics were downloaded from The Cancer Genome Atlas (TCGA) database (<https://portal.gdc.cancer.gov/>). Data with incomplete clinical information were excluded during analysis.

Statistical analysis

Data were presented here as mean \pm standard deviation (SD) and analyzed by the Student's t-test or one-way ANOVA. Quantitative data were representative of three experiments. Wilcoxon signed-rank test and Wilcoxon rank-sum test was adopted to analyze gene expressions in paired and non-paired tissue samples. The relationships between disease stages and gene expressions was checked by Wilcoxon rank-sum test. Prognosis analysis was performed using Kaplan-Meier method and univariate Cox regression. All bioinformatics statistical analyses and plots were produced using R (v.3.5.2). $P < 0.05$ was considered statistically significant.

Results

MiR-21-3p was upregulated in HCC and its clinical significance in HCC based on bioinformatics analysis

Based on our previous chip analysis results, miR-21-3p was significantly upregulated in HCC compared to normal liver tissues (NLTs), which had statistical significance ($P=0.028$) (Table 1). To further verify the results of chip analysis, the mRNA expression of miR-21-3p was examined in human HCC tissue samples and Wistar rat models, respectively. Results showed that miR-21-3p was significantly enriched at HCC tissues compared to background livers (BLs) (Fig.1A (a)). Its upregulation was also observed in rats with late-stage fibrosis and cirrhosis (Fig.1A (b)). Compared to human relative normal cell L02, miR-21-3p was increased in liver cancer cell lines (Huh-7, Hep-G2, HCCL-M3) (Fig.1A (c)). Considering the well expression consistency in human tissues, rat liver disease models and cell lines, miR-21-3p was selected for subsequent analysis. To explore the clinical significance of the miR-21-3p in human HCCs, the Cancer Genome Atlas (TCGA) data, including 376 HCC patients, whose survival status was available, were analyzed. We found that patients with high miR-21-3p expression were significantly correlated with

advanced clinical stages ($P = 0.029$), especially in T staging ($P = 0.026$), while no noticeable difference was observed in age and gender. (Figure 1B). Higher miR-21-3p expression level was correlated to shorter ten-year overall survival (OS) time (log-rank $P=0.026$) (Fig. 1C). GSEA analysis of miR-21-3p displayed that high expression of miR-21-3p was mainly enriched at 20 pathways in HCC (Fig.1D). In particular, the top three gene sets related were nitrogen metabolism, fatty acid metabolism and primary bile acid biosynthesis, severally (Fig.1E). KEGG pathway analysis of miR-21-3p targets revealed that the downstream targets were strongly linked to metabolic pathways, Hippo signaling road and TGF-beta signaling pathway (Table 2). The intersections of miR-21-3p potential targets from three authoritative databases (TargetScan, PicTar, miRDB) were *Smad7*, HBP1, FBXO11, respectively (Fig.1F). Of these, *Smad7* earned the highest score (Table 3), while the potential relationship between miR-21-3p and *Smad7* waited to be rectified.

Table 1. Different expressions of miRNAs in all 3 cases of HCC tissues and normal liver tissues. (Top fifteen listed here)

#Term	LogFC		
	(HCC vs NLT)	P-value	Corrected P-Value
hsa-miR-663	12.00859435	0.006642752	0.025159423
hsa-miR-452	10.42475206	0.044754151	0.096860769
hsa-miR-576-3p	8.892785445	0.085150937	0.129003669
hsa-miR-224	8.397356424	0.002498379	0.018925218
hsa-miR-96	5.635504649	0.04323621	0.096860769
hsa-miR-216a	3.536217114	0.1519771	0.191871088
hsa-miR-222	3.425619069	0.031592274	0.095724591
hsa-miR-551b	2.443388558	0.360760502	0.364368107
hsa-miR-500*	2.359130217	0.07579151	0.127790049
hsa-miR-500	2.31112225	0.189840985	0.205435066
hsa-miR-505*	2.216287725	0.108599824	0.149571576
hsa-miR-21-3p	2.188741196	0.005914881	0.027790049
hsa-miR-34b*	2.013273752	0.005016725	0.025159423
hsa-miR-18a	1.985718798	0.189563246	0.205435066
hsa-miR-34a	1.440184224	0.002272378	0.018925218

Footnotes: LogFC: Log fold change; HCC: hepatocellular carcinoma; NLT: normal liver tissue

Table 2. The Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis of miR-21-3p downstream targets. (Top twelve listed here)

#Term	ID	Input number	Background number	P-Value	Corrected P-Value
Metabolic pathways	hsa01100	28	1243	0.0068	0.0303
Pathways in cancer	hsa05200	14	397	0.001	0.0110
MAPK signaling pathway	hsa04010	12	255	0.0003	0.0063
PI3K-Akt signaling pathway	hsa04151	12	342	0.0030	0.0173
cGMP-PKG signaling pathway	hsa04022	11	167	2.89E- 05	0.0016
Ubiquitin mediated proteolysis	hsa04120	10	137	2.93E- 05	0.0016
Insulin signaling pathway	hsa04910	10	139	3.29E- 05	0.0016
Hippo signaling pathway	hsa04390	10	154	7.42E- 05	0.0022
cAMP signaling pathway	hsa04024	10	199	0.0005	0.0085
HTLV-I infection	hsa05166	10	259	0.0034	0.01838
TGF-beta signaling pathway	hsa04350	6	84	0.0013	0.0110
mTOR signaling pathway	hsa04150	7	154	0.0060	0.0269

Table 3. Intersections of potential targets of miR-21-3p based on analysis of three online databases

Target Gene	miRDB		
	TargetScan total context score	target score	PicTar score
<i>Smad7</i>	-0.39	92	2.15
<i>HBP1</i>	-0.15	53	2.22
<i>FBXO11</i>	-0.23	58	2.79

***Smad7* as the direct target of miR-21-3p was decreased in HCC**

To further verify our analysis, expressions of *Smad7* in human tissue samples and rat models were checked. Compared to BLs, *Smad7* was obviously decreased in HCCs (9/10) (Fig.2A (a)). Also, the lighter brownish staining was observed in HCCs compared to BLs (Fig.2B (a)). Along with the progressing of liver disease in rat models, a gradually descending trend was observed in *Smad7*'s protein expression level (Fig.2A (b)), and immunohistochemical staining showed the same trend (Fig.2B (b)). Considering the apparent up-regulation of miR-21-3p and down-regulation of *Smad7* in HCC, Dual-Luciferase assay was done in Huh-7 cells to confirm the linear regulation relation between miR-21-3p and *Smad7*. Prediction websites presented there were two binding sites in the 3'-UTR region of *Smad7* with high scores. In the group of co-transfecting miR-21-3p and reporter vector containing wild-type (WT) *Smad7*, luciferase activity was significantly inhibited, while in the group containing mutation *Smad7* reporter vectors (mutation-3 (Mut3) mutating two sites simultaneously), the inhibition efficiency was no statistical significance between miR-NC and miR-21-3p group (Fig.2C). The above results indicated that the two prediction sites might both exert the effect on inhibiting *Smad7*.

Overexpression of *Smad7* partly abrogates the progressive tumor effect of miR-21-3p on cell malignant phenotypes in HCC

Mimics and inhibitors of miR-21-3p were transfected into Huh-7 and Hep-G2 cells to gain and lose its function separately. Plasmids of *Smad7* (P-*Smad7*) was constructed and transfected into Huh-7 and Hep-G2 to overexpress *Smad7*, and Plasmid-NC (P-NC) served as the negative control. *Smad7* expression was promoted after transfecting miR-21-3p inhibitors and decreased following transfecting miR-21-3p mimics in Huh-7 (Fig.3A (a)) and Hep-G2 cells (Fig.3A (b)). To investigate the effect of the miR-21-3p/ *Smad7* on tumor cell malignant phenotypes in HCC, biomarkers related to cell apoptosis, migration and invasion were examined. Pro-apoptotic protein Bax and epithelial signature protein E-cadherin (E-cad) were upregulated while anti-apoptotic protein Bcl-2, mesenchymal characteristic protein N-cadherin (N-cad) and Vimentin were down-regulated after transfecting miR-21-3p inhibitors or *Smad7* plasmids in Huh-7 and Hep-G2 respectively (Fig.3A-B). The effect of miR-21-3p on cell malignant biomarkers was partly attenuated by co-transfecting *Smad7* in both two cell lines (Fig.3B). Flow cytometry results displayed that the early apoptotic rate stained green was increased by transfecting *Smad7* ($P < 0.01$) and decreased by up-regulating miR-21-3p ($P < 0.05$) (Fig.4A). The migration (Fig.4B (a-b)) and invasion abilities (Fig.4B (c-d)) enhanced by transfecting miR-21-3p mimics were partly reversed by co-transfecting *Smad7*.

MiR-21-3p could enhance the expression of *Yap1* partly via down-regulating *Smad7*

The KEGG pathway enrichment analysis displayed that miR-21-3p was significantly related to the Hippo signaling pathway (Table 2). Besides, a previous study confirmed that *Yap1* could enhance the binding of *Smad7* to T β R (18). To further investigate the potential link among, miR-21-3p, *Smad7* and *Yap1* in HCC, the expression of *Yap1* was detected in HCC tissues and rat models. Compared to BLs, *Yap1* was enriched at human HCC tissues (8/10) (Fig.5A), and advanced stages of liver fibrosis and cirrhosis compared to F0 (F: fibrosis) (Fig.2A (b)). The darker brownish staining in immunohistochemistry images

was observed in HCC tissues (Fig.5B (a)) and the stage of F3 and F4 in rats compared to F0 (Fig.5B (b)). In both Huh-7 and Hep-G2 cells, *Yap1*'s expression was promoted through enhancing miR-21-3p and inhibited when cells were transfecting miR-21-3p inhibitors or up-regulating *Smad7* (Fig.5C (a,b)). In protein level, the promotion of expression of *Yap1* was partly attenuated by co-transfecting *Smad7* (Fig.5C (b)). However, in the mRNA level, the influence of miR-21-3p and *Smad7* on *Yap1* had no statistical significance (Fig 5C (c,d)). Large tumor suppressor 2 (*LATS2*) as the upstream regulator of *Yap1* was decreased by up-regulating *Smad7* (Fig.5C(b), S1(A-B)). Besides, mRNA (Fig.S2A-B) and protein (Fig.5C(b)) expression trend of connective tissue growth factor (*CTGF*) which was proven to be the direct nuclear target of nuclear *Yap1* was consistent with *Yap1* here (22) (Fig.5C (b)). Therefore, we deduced that miR-21-3p could promote the expression of *Yap1* partly through downregulating *Smad7*.

The clinical significance of *Smad7*/*Yap1* based on bioinformatics analysis

Data about 376 HCC patients was downloaded from the Cancer Genome Atlas (TCGA) database. Wilcoxon rank-sum test results displayed that *Smad7* was decreased ($P=4.578e-04$) and *Yap1* ($P=3.6e-05$) was increased in HCC compared to adjacent normal liver tissues (Fig.6A). Combining RNA-seq data with patients' clinical traits, low *Smad7*/ high *Yap1* was related to low grading and staging (Fig.6B). Co-survival analysis results indicated that Lower miR-21-3p/Higher *Smad7* ($P=0.049$) and lower miR-21-3p/lower *Yap1* ($P=0.038$) were related to a better five-year OS rate (Fig.6C). GSEA volcano map showed that *Smad7* was mainly involved in 18 pathways (Fig.6D). Of note, TGF- β signaling pathway, Notch signaling pathway, adherens junction were most relevant in HCC (Fig.6E).

Discussion

HCC as the second leading cause of cancer-related death worldwide lacks efficient treatments. Abnormal expression of miRNAs played a critical role in cancer occurrence and progress including hepatocarcinogenesis. Previous reports focused on introducing the guide strand miR-21-5p's function in HCC, few researches about the passenger strand miR-21-3p. Until recently, reports on the independent function of miR-21-3p gradually increased, which brought us a new understanding of this microRNA (23,24). The independent function of miR-21-3p includes it could be used as a signature in predicting the survival rate of triple-negative breast cancer (25). Besides, it's overexpression could facilitate the pulmonary metastasis by influencing the junction between tumor and stromal cells (26). In our study, we investigated miR-21-3p's role in the occurrence and progression of HCC. MiR-21-3p was increased in HCC tissue samples compared to adjacent background livers ($P<0.001$). Further, based on the analysis of mRNA expression level of miR-21-3p in rat models, miR-21-3p may be involved in not only HCC progression but also hepatocarcinogenesis. KEGG pathway analysis of miR-21-3p's targets indicated that it was strongly related to TGF- β signaling transduction pathway and Hippo signaling pathway. Besides, the abnormal overexpression of miR-21-3p facilitated the malignant phenotypes of liver cancer cells.

Chronic liver diseases including liver fibrosis, cirrhosis are strongly associated with HCC (27). One in three patients with cirrhosis will develop HCC in their life (28). Dysregulation of TGF- β signaling transduction

pathway was observed and testified in liver fibrosis and cirrhosis (29). Enhancing evidence demonstrated that miRNAs were involved in the process of liver fibrosis and cirrhosis through directly targeting Smad proteins (30). Our results highlighted the role of *Smad7* in HCC. The absence of *Smad7* was widely seen in HCC tissues compared to BLs, and had a gradually descending trend along with the progression of liver disease in rat models. Besides, deletion of *Smad7* showed a good consistency in tissues and cells in HCC. We confirmed that *Smad7* was the direct target of miR-21-3p in HCC. Restoring *Smad7* could impair the migration and invasion abilities which were enhanced by miR-21-3p of liver cancer cell lines Huh-7 and Hep-G2, promote cell early apoptosis. During GSEA analysis, *Smad7* was mainly involved in TGF- β signaling transduction pathway in HCC. Therefore, in the occurrence and development of HCC, the deletion of *Smad7* really matters.

Previous gene sets analysis of miR-21-3p's targets confirmed that it had strong relation with Hippo signaling pathway. *Yap1* was obviously enriched at HCC tissues compared to BLs. In advanced stages of liver disease in rat models, it's expression was higher than normal liver tissue. Our experiments explored that overexpression of miR-21-3p could facilitate the expression of *Yap1*. The ascending of *Yap1* via overexpression of miR-21-3p could be partly reversed by transfecting *Smad7* in the protein level. Increased *Yap1* enhanced the expression of CTGF. Upstream regulator *LATS2* was the core serine/threonine kinase of the Hippo signaling transduction pathway. Once the Hippo was activated, phosphorylation of *LATS2* could be facilitated and then further phosphorylating *Yap1*, resulting in *Yap1* inactivation, and there is a negative feedback between *Yap1* and *LATS2* (31). Expression of *LATS2* in overexpression of *Smad7* group was downregulated while surged through transfecting miR-21-3p. This indicated that the up-regulation of *Yap1* was not caused by inactivation of *LATS2* but loss function of *Smad7*. Further, increased *Yap1* weakened the negative influence of *LATS2*. Vitro experiments effect in Huh-7 cells was more pronounced than Hep-G2 cells which was correlated to the basic expressions of miR-21-3p and *Smad7* in cells (Fig.1A(a), S1B). We proposed that overexpression *Smad7* could cut the expression of *Yap1* in the protein level, not in the mRNA level. Here we concluded the complete axis in Figure 7.

Besides, recent studies suggested that not only *Yap1* can be regulated by a series of microRNAs, but nuclear *Yap1/TAZ* (*Yap1*'s paralog) could influence the conversion of pri-miRNA into pre-miRNA (32). Combining our results, we highly proposed that in the axis of miR-21-3p-*Smad7/ Yap1*, there is a possibility that overexpression of nuclear *Yap1* could in turn affect the expression of miR-21-3p. Previous studies confirmed that *Yap1* as a mechano-transduction effector translocated from cytoplasm to nucleus when cells were shifted from soft to stiff matrices (33-35). Meanwhile, during the hepatocarcinogenesis, liver fibrosis and cirrhosis are typical examples of extracellular matrix (ECM) sclerosis (36), which are the "boosters" of tumor metastasis. Although there are great differences between people, the stiff of liver cancer tissue (55kpa) is almost about ten times that of normal liver tissue (4kpa) detected by Fibroscan (37). A reasonable and also surprising discovery was that about twofold up-regulation of miR-21-3p was found in three cell lines (Huh-7, Hep-G2, Hep-3B) when cells were cultured on stiffer substrates (40kpa) (Fig.S3A). Besides, the nuclear *Yap1* in cells cultured on stiff gels was increased (Fig.S3B). Immunofluorescence results presented the deformation of cytoskeleton and changes of cell polarity in

the stiffer matrixes (Fig.S3C). This indicated that there might be a positive loop between miR-21-3p and *Yap1* during the progression of HCC. However, how did cells sense the mechanical signals and then transfer them into molecular signals, even changing the level miRNAs remained unclear. Our team believed that there was still an interesting story waiting to be figured out and we're working on it.

Conclusions

Our results revealed the oncogenic role of miR-21-3p in HCC. MiR-21-3p-*Smad7/ Yap1* axis was identified and might be potential therapeutic targets for HCC diagnose and treatments.

List Of Abbreviations

HCC: Hepatocellular carcinoma

MiRNAs: microRNAs

OS: overall survival

CTGF: connective tissue growth effector

Yap1: Yes-associated protein 1

Adenosyltransferases 2A/2B: MAT2A/MAT2B

TβR: TGF-β Receptor

GSVA: Gene set variation analysis

TCGA: The Cancer Genome Atlas

SD: standard deviation

NLT: normal liver tissues

BL: background liver

WT: wild type

ECM: extracellular matrix

Declarations

Ethics approval and consent to participate

Human tissue samples used here were approved by the local Research Ethics Committee at Zhongnan Hospital of Wuhan University (Approval No.2018078). All animal handling and experimental procedures were approved by the Animal Care and Use Committee of Wuhan University following the Animal Experiment/Animal Biosafety Level-III Laboratory Guidelines.

Consent for publication Data

All authors have been approved this publication.

Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Competing interests

The authors declare that they have no competing interests.

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Authors' contributions

HYH designed and wrote this article. HYH and WD did the experiments. YML, WC and LJ revised this manuscript and gave useful suggestions. ZQ, LJ supervised this manuscript. CY funded this article and gave useful instructions. All authors read and approved the final manuscript.

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References

1. Craig AJ, von Felden J, Garcia-Lezana T, Sarcognato S, Villanueva A. Tumour evolution in hepatocellular carcinoma. *Nat Rev Gastroenterol Hepatol* 2020;17:139-52. doi:10.1038/s41575-019-0229-4
2. Yang JD, Hainaut P, Gores GJ, Amadou A, Plymoth A, Roberts LR. A global view of hepatocellular carcinoma: trends, risk, prevention and management. *Nat Rev Gastro Hepat* 2019;16:589-604. doi:10.1038/s41575-019-0186-y

3. Couri T, Pillai A. Goals and targets for personalized therapy for HCC. *Hepatol Int* 2019;13:125-37. doi:10.1007/s12072-018-9919-1
4. Kudo M, Finn RS, Qin S, Han KH, Ikeda K, Piscaglia F, et al. Lenvatinib versus sorafenib in first-line treatment of patients with unresectable hepatocellular carcinoma: a randomized phase 3 non-inferiority trial. *Lancet* 2018;391:1163-73. doi:10.1016/S0140-6736(18)30207-1
5. Tutar L, Tutar E, Tutar Y. MicroRNAs and cancer; an overview. *Curr Pharm Biotechnol* 2014;15:430-7. doi:10.2174/1389201015666140519095304
6. Lin SB, Gregory RI. MicroRNA biogenesis pathways in cancer. *Nature Reviews Cancer* 2015;15:321-33. doi:10.1038/nrc3932
7. Zhu Y, Tang H, Zhang L, Gong L, Wu G, Ni J, et al. Suppression of miR-21-3p enhances TRAIL-mediated apoptosis in liver cancer stem cells by suppressing the PI3K/Akt/Bad cascade via regulating PTEN. *Cancer Manag Res* 2019;11:955-68. doi:10.2147/CMAR.S183328
8. Slapeta J, Linares MC. Combined amplicon pyrosequencing assays reveal presence of the apicomplexan "type-N" (cf. *Gemmocystis cylindrus*) and *Chromera velia* on the Great Barrier Reef, Australia. *PLoS One* 2013;8:e76095. doi:10.1371/journal.pone.0076095
9. Hata A, Chen YG. TGF-beta Signaling from Receptors to Smads. *Cold Spring Harb Perspect Biol* 2016;8. doi:10.1101/cshperspect.a022061
10. Yan X, Liao H, Cheng M, Shi X, Lin X, Feng XH, et al. Smad7 Protein Interacts with Receptor-regulated Smads (R-Smads) to Inhibit Transforming Growth Factor-beta (TGF-beta)/Smad Signaling. *J Biol Chem* 2016;291:382-92. doi:10.1074/jbc.M115.694281
11. Batlle E, Massague J. Transforming Growth Factor-beta Signaling in Immunity and Cancer. *Immunity* 2019;50:924-40. doi:10.1016/j.immuni.2019.03.024
12. Zhou Q, Zhou Q, Liu Q, He Z, Yan Y, Lin J, et al. PRL-3 facilitates Hepatocellular Carcinoma progression by co-amplifying with and activating FAK. *Theranostics* 2020;10:10345-59. doi:10.7150/thno.42069
13. Vivekanandhan S, Mukhopadhyay D. Genetic status of KRAS influences Transforming Growth Factor-beta (TGF-beta) signaling: An insight into Neuropilin-1 (NRP1) mediated tumorigenesis. *Semin Cancer Biol* 2019;54:72-9. doi:10.1016/j.semcancer.2018.01.014
14. Feng T, Dzieran J, Yuan X, Dropmann A, Maass T, Teufel A, et al. Hepatocyte-specific Smad7 deletion accelerates DEN-induced HCC via activation of STAT3 signaling in mice. *Oncogenesis* 2017;6:e294. doi:10.1038/oncsis.2016.85
15. Wang J, Zhao J, Chu ES, Mok MT, Go MY, Man K, et al. Inhibitory role of Smad7 in hepatocarcinogenesis in mice and in vitro. *J Pathol* 2013;230:441-52. doi:10.1002/path.4206
16. Raj N, Bam R. Reciprocal Crosstalk Between YAP1/Hippo Pathway and the p53 Family Proteins: Mechanisms and Outcomes in Cancer. *Front Cell Dev Biol* 2019;7:159. doi:10.3389/fcell.2019.00159
17. Tschaharganeh DF, Chen X, Latzko P, Malz M, Gaida MM, Felix K, et al. Yes-associated protein up-regulates Jagged-1 and activates the Notch pathway in human hepatocellular carcinoma. *Gastroenterology* 2013;144:1530-42 e12. doi:10.1053/j.gastro.2013.02.009

18. Ferrigno O, Lallemand F, Verrecchia F, L'Hoste S, Camonis J, Atfi A, et al. Yes-associated protein (YAP65) interacts with Smad7 and potentiates its inhibitory activity against TGF-beta/Smad signaling. *Oncogene* 2002;21:4879-84. doi:10.1038/sj.onc.1205623
19. He XX, Chang Y, Meng FY, Wang MY, Xie QH, Tang F, et al. MicroRNA-375 targets AEG-1 in hepatocellular carcinoma and suppresses liver cancer cell growth in vitro and in vivo. *Oncogene* 2012;31:3357-69. doi:10.1038/onc.2011.500
20. Knodell RG, Ishak KG, Black WC, Chen TS, Craig R, Kaplowitz N, et al. Formulation and application of a numerical scoring system for assessing histological activity in asymptomatic chronic active hepatitis. *Hepatology* 1981;1:431-5. doi:10.1002/hep.1840010511
21. Wang C, Luo J, Chen Z, Ye M, Hong Y, Liu J, et al. MiR-375 Impairs the Invasive Capabilities of Hepatoma Cells by Targeting HIF1alpha Under Hypoxia. *Dig Dis Sci* 2020. doi:10.1007/s10620-020-06202-9
22. Kang W, Huang T, Zhou Y, Zhang J, Lung RWM, Tong JHM, et al. miR-375 is involved in Hippo pathway by targeting YAP1/TEAD4-CTGF axis in gastric carcinogenesis. *Cell Death Dis* 2018;9:92. doi:10.1038/s41419-017-0134-0
23. Calsina B, Castro-Vega LJ, Torres-Perez R, Inglada-Perez L, Curras-Freixes M, Roldan-Romero JM, et al. Integrative multi-omics analysis identifies a prognostic miRNA signature and a targetable miR-21-3p/TSC2/mTOR axis in metastatic pheochromocytoma/paraganglioma. *Theranostics* 2019;9:4946-58. doi:10.7150/thno.35458
24. Moscetti I, Cannistraro S, Bizzarri AR. Probing direct interaction of oncomiR-21-3p with the tumor suppressor p53 by fluorescence, FRET and atomic force spectroscopy. *Arch Biochem Biophys* 2019;671:35-41. doi:10.1016/j.abb.2019.05.026
25. Wu X, Ding M, Lin J. Three-microRNA expression signature predicts survival in triple-negative breast cancer. *Oncol Lett* 2020;19:301-8. doi:10.3892/ol.2019.11118
26. Ju Q, Zhao L, Gao J, Zhou L, Xu Y, Sun Y, et al. Mutant p53 increases exosome-mediated transfer of miR-21-3p and miR-769-3p to promote pulmonary metastasis. *Chin J Cancer Res* 2019;31:533-46. doi:10.21147/j.issn.1000-9604.2019.03.15
27. Kanda T, Goto T, Hirotsu Y, Moriyama M, Omata M. Molecular Mechanisms Driving Progression of Liver Cirrhosis towards Hepatocellular Carcinoma in Chronic Hepatitis B and C Infections: A Review. *Int J Mol Sci* 2019;20. doi:10.3390/ijms20061358
28. El-Serag HB. Hepatocellular carcinoma. *N Engl J Med* 2011;365:1118-27. doi:10.1056/NEJMra1001683
29. Xu F, Liu C, Zhou D, Zhang L. TGF-beta/SMAD Pathway and Its Regulation in Hepatic Fibrosis. *J Histochem Cytochem* 2016;64:157-67. doi:10.1369/0022155415627681
30. Yang YZ, Zhao XJ, Xu HJ, Wang SC, Pan Y, Wang SJ, et al. Magnesium isoglycyrrhizinate ameliorates high fructose-induced liver fibrosis in rat by increasing miR-375-3p to suppress JAK2/STAT3 pathway and TGF-beta1/Smad signaling. *Acta Pharmacol Sin* 2019;40:879-94. doi:10.1038/s41401-018-0194-4

31. Taha Z, Janse van Rensburg HJ, Yang X. The Hippo Pathway: Immunity and Cancer. *Cancers (Basel)* 2018;10. doi:10.3390/cancers10040094
32. Han Y. Analysis of the role of the Hippo pathway in cancer. *J Transl Med* 2019;17:116. doi:10.1186/s12967-019-1869-4
33. Burnette FS, Flick GJ, Jr. Comparison of Three Assays for Peroxidase in the Blue Crab (*Callinectes sapidus*). *J Food Prot* 1977;40:854-6. doi:10.4315/0362-028X-40.12.854
34. Halder G, Dupont S, Piccolo S. Transduction of mechanical and cytoskeletal cues by YAP and TAZ. *Nat Rev Mol Cell Biol* 2012;13:591-600. doi:10.1038/nrm3416
35. Tse JR, Engler AJ. Preparation of hydrogel substrates with tunable mechanical properties. *Curr Protoc Cell Biol* 2010;Chapter 10:Unit 10 6. doi:10.1002/0471143030.cb1016s47
36. Zeng Z, Guan L, An P, Sun S, O'Brien SJ, Winkler CA, et al. A population-based study to investigate host genetic factors associated with hepatitis B infection and pathogenesis in the Chinese population. *BMC Infect Dis* 2008;8:1. doi:10.1186/1471-2334-8-1
37. Masuzaki R, Tateishi R, Yoshida H, Sato T, Ohki T, Goto T, et al. Assessing liver tumor stiffness by transient elastography. *Hepatol Int* 2007;1:394-7. doi:10.1007/s12072-007-9012-7

Figures

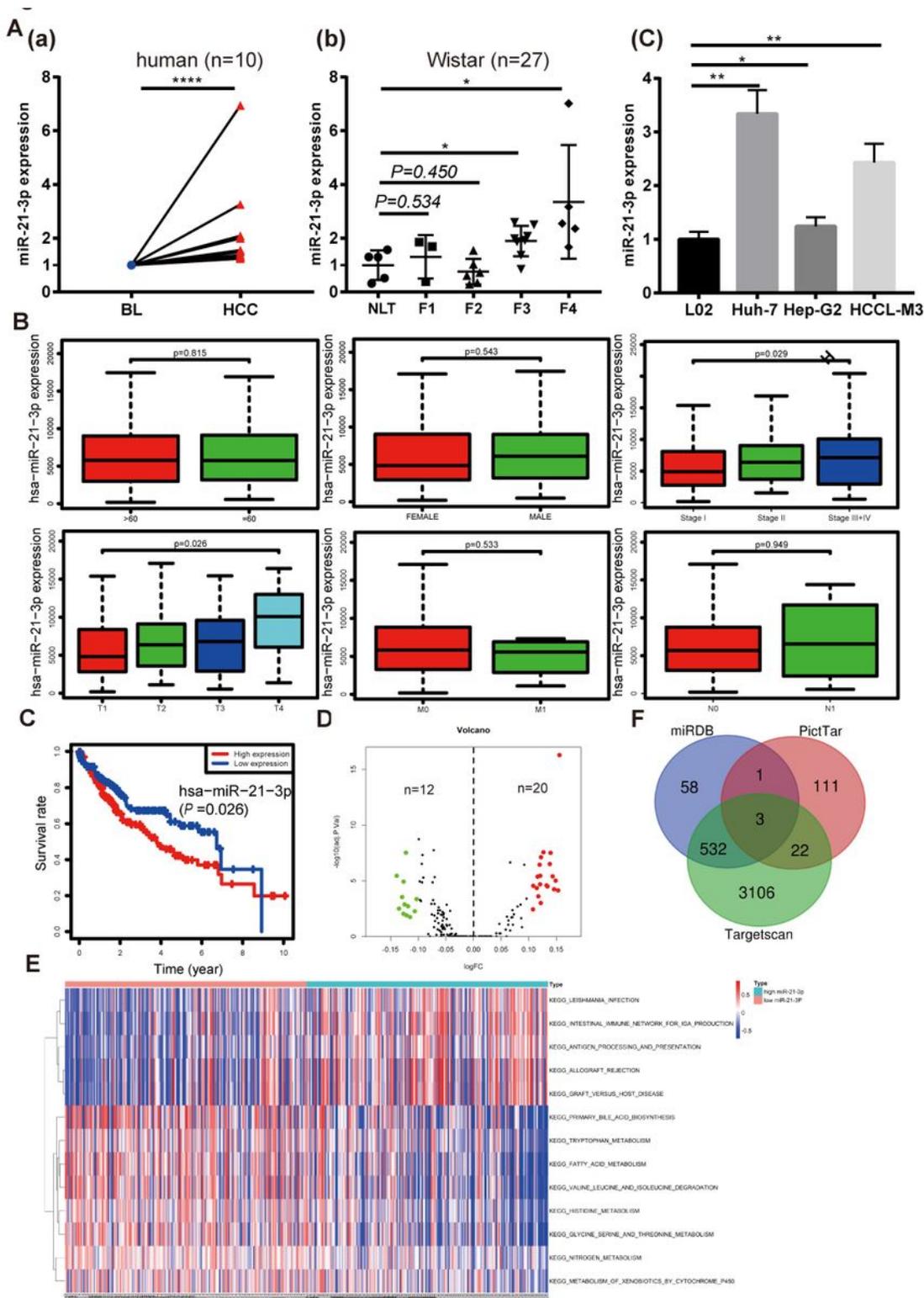


Figure 1

miR-21-3p is upregulated and its clinical significance in hepatocellular carcinoma (HCC). (A) TaqMan quantitative RT-PCR was performed to analyze microRNA-21-3p (miR-21-3p) expressions. 10 pairs of Human HCCs and their counterpart background livers (BLs) were examined (A-a); Normal, fibrosis and cirrhosis tissues from rat models were also analyzed (A-b). MiR-21-3p expression in three cell lines (L02, Huh-7 and Hep-G2) (A-c). Each dot indicates the expression level of an individual case, calculated by the

2- $\Delta\Delta$ Ct method. (B) Association of miR-21-3p expression level with clinical characteristics (age, gender, stage, TNM staging, respectively). (C) Kaplan-Meier curves representing the relation between miR-21-3p and the percent overall survival (OS) rate in HCC patients in the TCGA data set (n=376). Statistical significance between miRNA expression and OS was determined by the Log-rank test. (D) Volcano map of the gene set variation analysis (GSVA) for microarray statistics according to the high and low expression of miR-21-3p. (E) Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways of miR-21-3p based on the gene set variation analysis of The Cancer Genome Atlas (TCGA) data in HCC. Significant terms were described in the heat-map (top-ten listed). (F) Venn diagram of intersections of three databases (miRDB, Targetscan, PicTar) of predicted downstream target genes of miR-21-3p. *P \leq 0.05, ** P \leq 0.01, **** P \leq 0.0001.

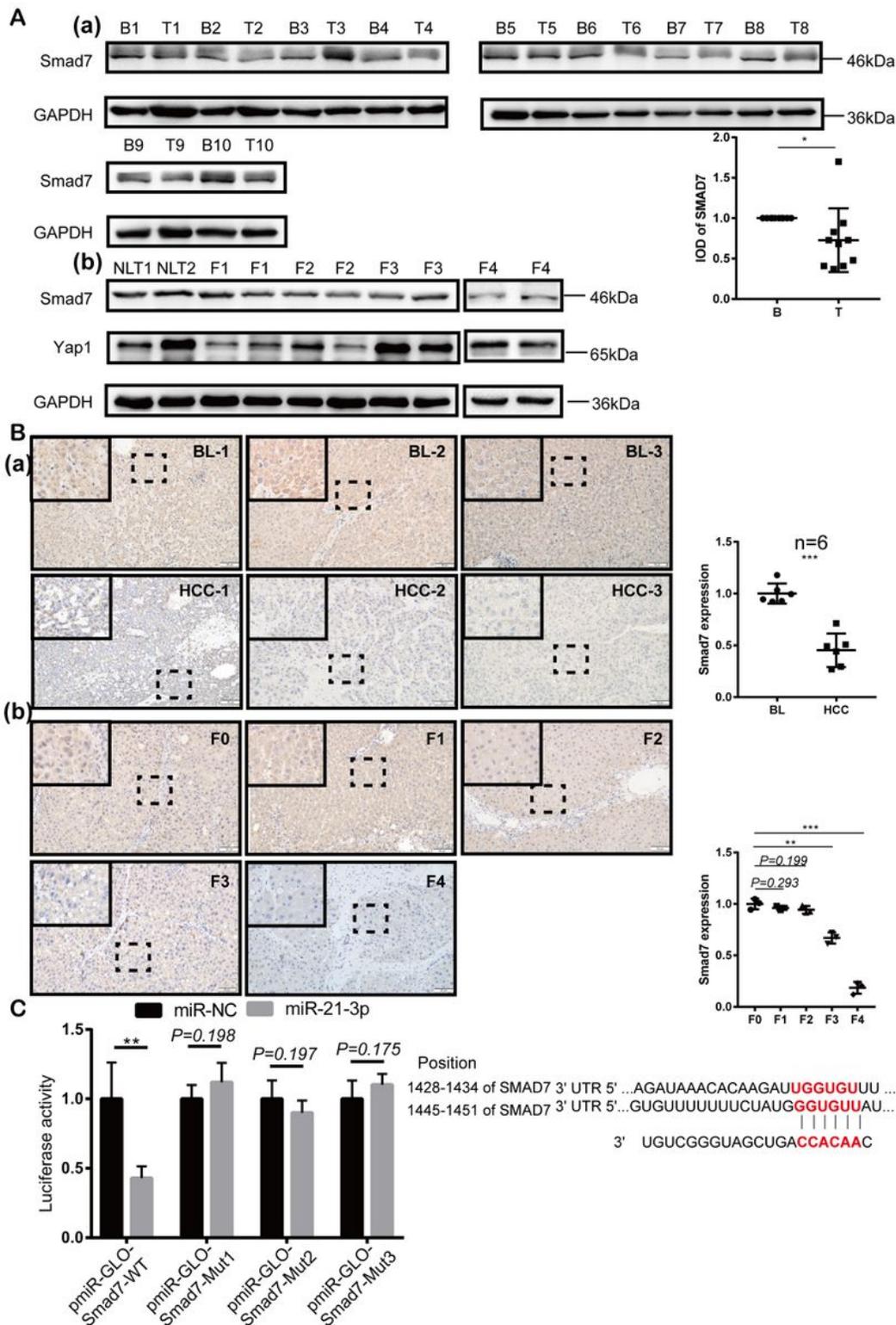


Figure 2

Smad7 was the direct target of miR-21-3p and was decreased in HCCs. (A) Western blotting and density photograph was performed to show Smad7 expressions in human HCCs (T: tumor) and their BLs (B: background) (a), different stages of rat hepatocarcinogenesis, including normal, fibrosis and cirrhosis (b). (B) Immunohistochemistry (IHC) staining images to show Smad7's expressions in HCCs and liver tissues at different stages of rat hepatocarcinogenesis (magnification 400×). Six paired human samples and

three rats each group were calculated during statistical analysis. (C) The base pairing complement suggested the two putative miR-21-3p binding sites at 3'UTR of Yap1 predicted by TargetScan. PmiR-GLO-Smad7 vector was co-transfected with mimic-21 or mimic-NC; wild-type (WT); mutation (Mut). Mut1-2 represented the two predicted binding sites, respectively. Mut-3 meant that two binding sites were mutated. Luciferase activity assay of pmiR-GLO-Smad7-3'UTR cotransfection with miR-21-3p or NC mimics. Firefly luciferase activity was measured 24h after transfection and normalized to renilla luciferase activity. All data for miR-NC groups was set as 1. Data were presented as the mean \pm SEM of the three separate triplicate experiments. ** $P < 0.01$.

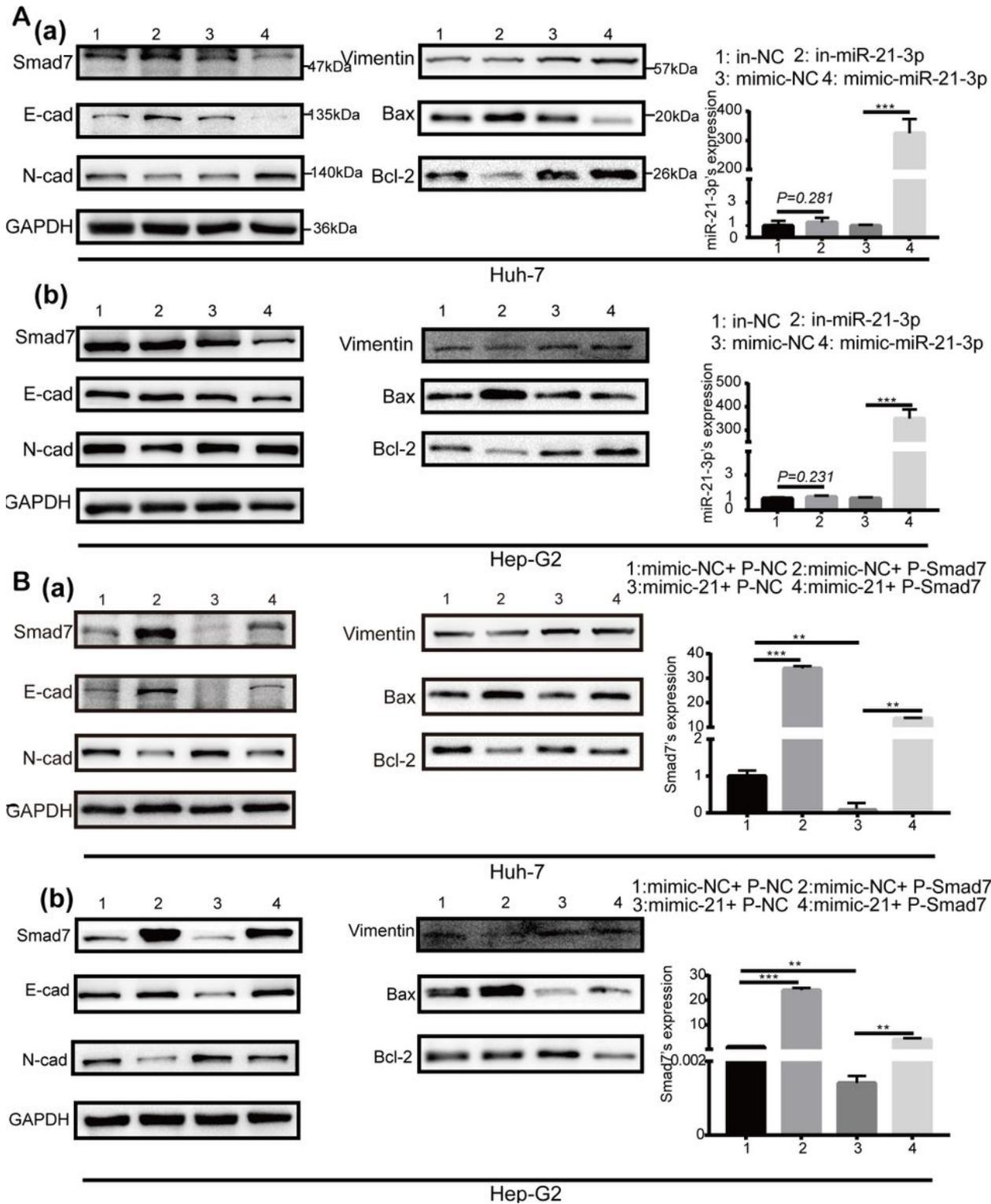


Figure 3

Variations of cell phenotypes biomarkers after transfecting Smad7 with/ or miR-21-3p mimics. (A) Western blotting showed apoptotic protein Bax and Bcl-2, Epithelial-mesenchymal transition (EMT) protein E-cadherin, N-cadherin and Vimentin expression after 48h transfection with miR-21-3p inhibitors with/ or mimics in Huh-7 cells (a) and Hep-G2 cells respectively (b). Transfection efficiency listed beside. (B) Plasmids-Smad7 (P-Smad7) was constructed to overexpress Smad7 and plasmids-NC (P-NC) served as

control. Transfection efficiency of Smad7 listed beside. Cell apoptosis (Bcl-2, Bax) and EMT biomarkers (E-cadherin, N-cadherin, Vimentin) were checked after co-transfecting P-Smad7 and/or miR-21-3p mimics in Huh-7 (a) and Hep-G2 cells (b), photo density in group1 (mimic-NC+ P-NC) was set as 1. Data were presented as the mean \pm SEM of the three separate triplicate experiments. ** $P < 0.01$, *** $P < 0.001$.

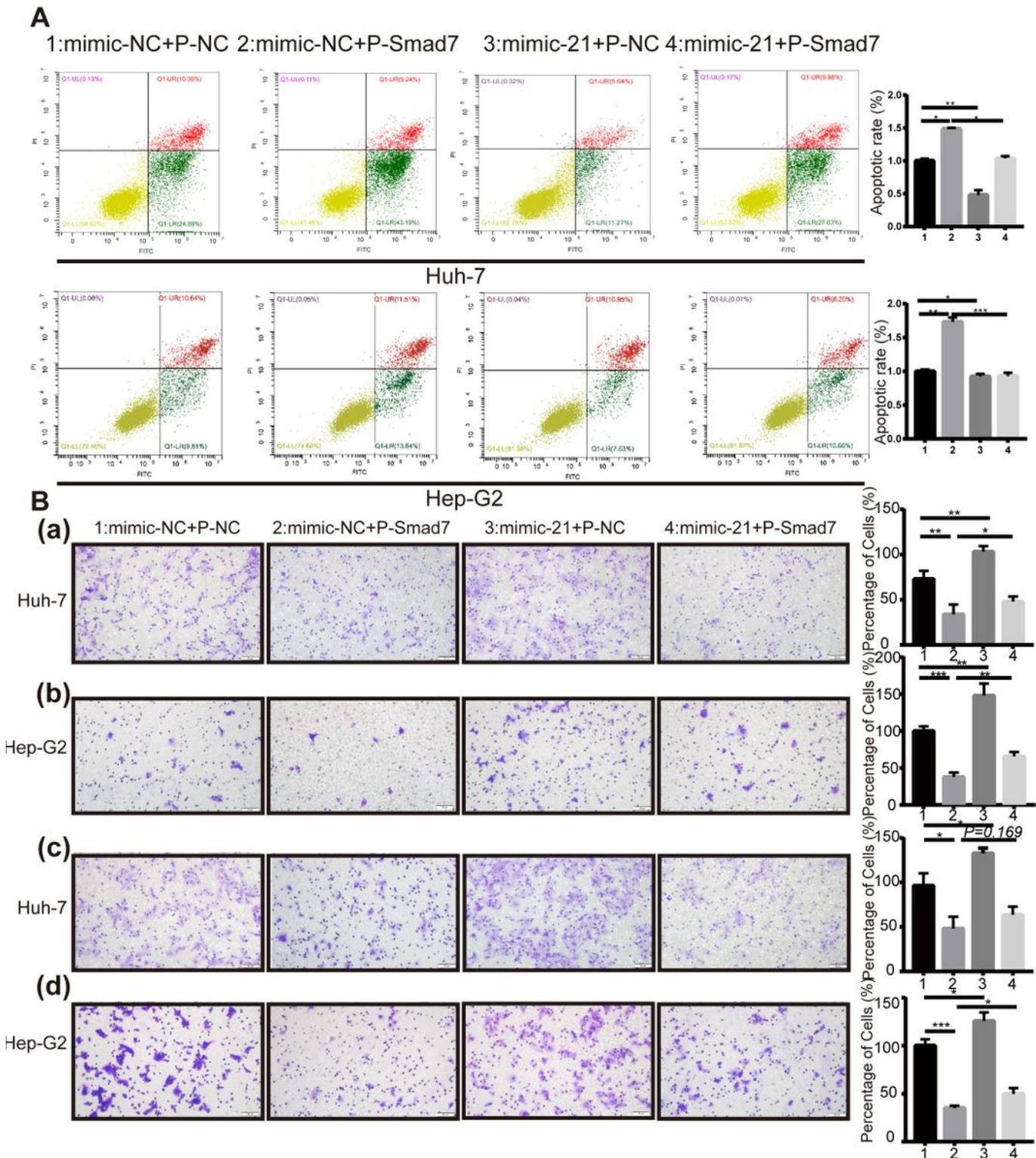


Figure 4

The effect of miR-21-3p on liver cancer cell malignant phenotypes could be partly reversed via overexpression of Smad7. (A) Flow cytometry was performed to analyze cell apoptosis index of Huh-7 and Hep-G2 cells after transfection. Cells were co-stained with Annexin FITC/PI, Annexin FITC+/PI- cells considered as early apoptotic cells were stained green. Normal, late apoptosis and necrosis cells were shown in yellow, red and pink, respectively. Data were shown as mean \pm SD from three independent experiments. (B) Transwell assay detected the migration (a, b) and invasion (c, d) ability of Huh-7 and Hep-G2 cells after transfection separately. * $P < 0.05$, ** $P < 0.01$; *** $P < 0.001$.

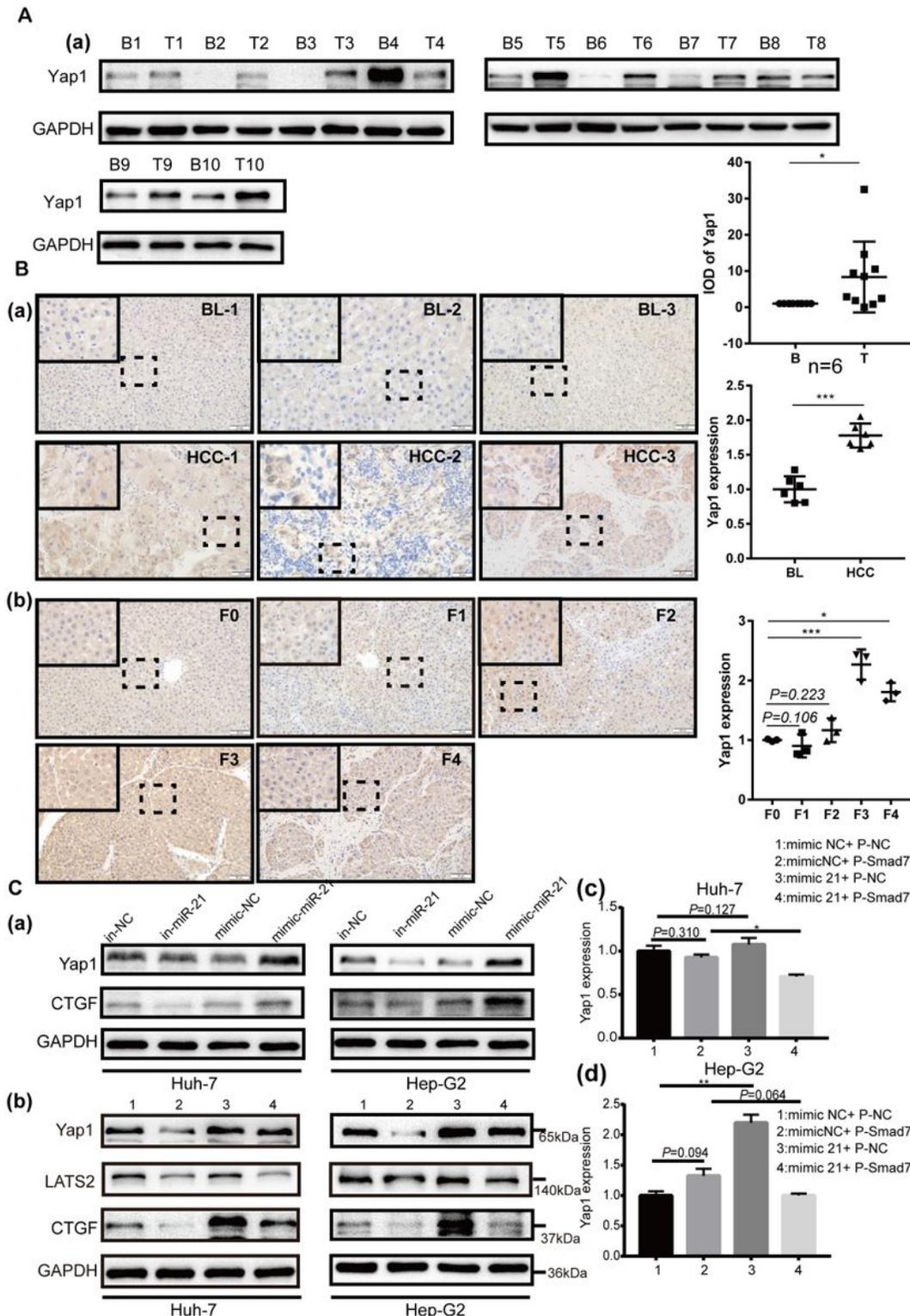


Figure 5

Yap1 was up-regulated in HCCs, and could be facilitated by miR-21-3p via inhibiting Smad7 (A) Western blotting was performed to show Yap1's expressions in human HCCs (T: tumor) and their BLs (B: background). (B) Expression of Yap1 expression in HCCs, BLs (a) and different liver disease stages in rat models (b) was checked through IHC staining assay (magnification 400×). Six paired human samples and three rats each group were calculated during statistical analysis. (C) Western blotting of Yap1 after 48h transfection with miR-21-3p inhibitors or mimics in Huh-7 cells and Hep-G2 cells (a), the protein expression level (b) and mRNA expression level (c,d) of Yap1,CTGF, LATS2 after transfecting plasmids Smad7 with/or miR-21-3p mimics.

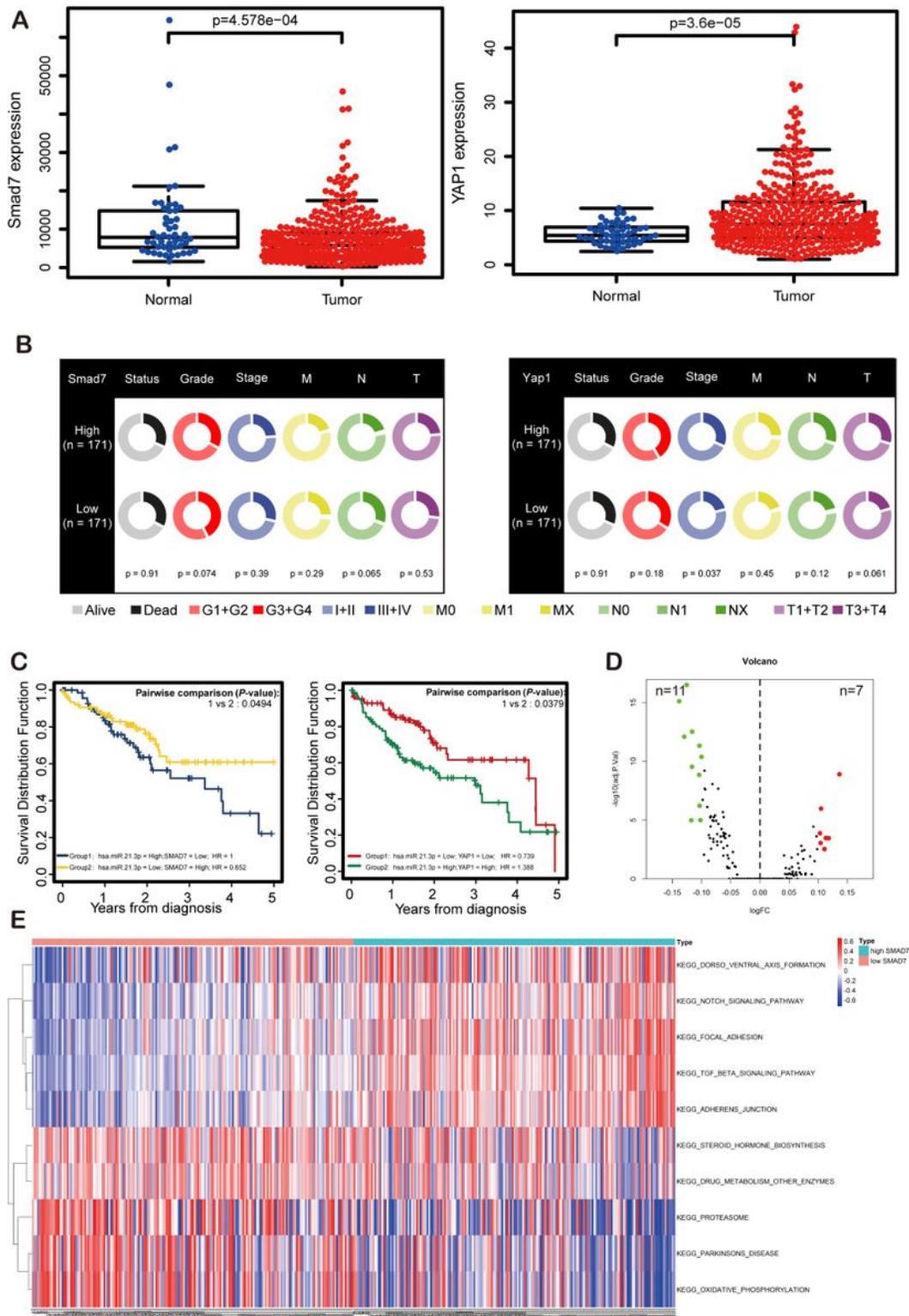


Figure 6

The clinical significance of Smad7/Yap1 in HCCs based on bioinformatics analysis. (A) Discrepancies of Smad7 (a) and Yap1 (b) expression levels in liver tumor tissues (Tumor) and non-paired relative normal samples (Normal) in the TCGA database were presented. (B) Relationship between the expression of Smad7, Yap1 and liver cancer clinicopathological terms using Wilcoxon rank-sum test. Data with incomplete clinical traits were excluded during analysis. (C) Kaplan-Meier curves representing the percent

overall survival (OS) in HCC patients based on miR-21-3p /Smad7 or miR-21-3p/Yap1 expression levels in TCGA data set (n=376). Statistical significance between miRNA/mRNA expression and OS was determined by the Log-rank test. Smad7-related KEGG pathways based on the gene set variation analysis of The Cancer Genome Atlas data. Shown in the volcano map (D) and heat map (E) (top ten including representative terms), respectively.

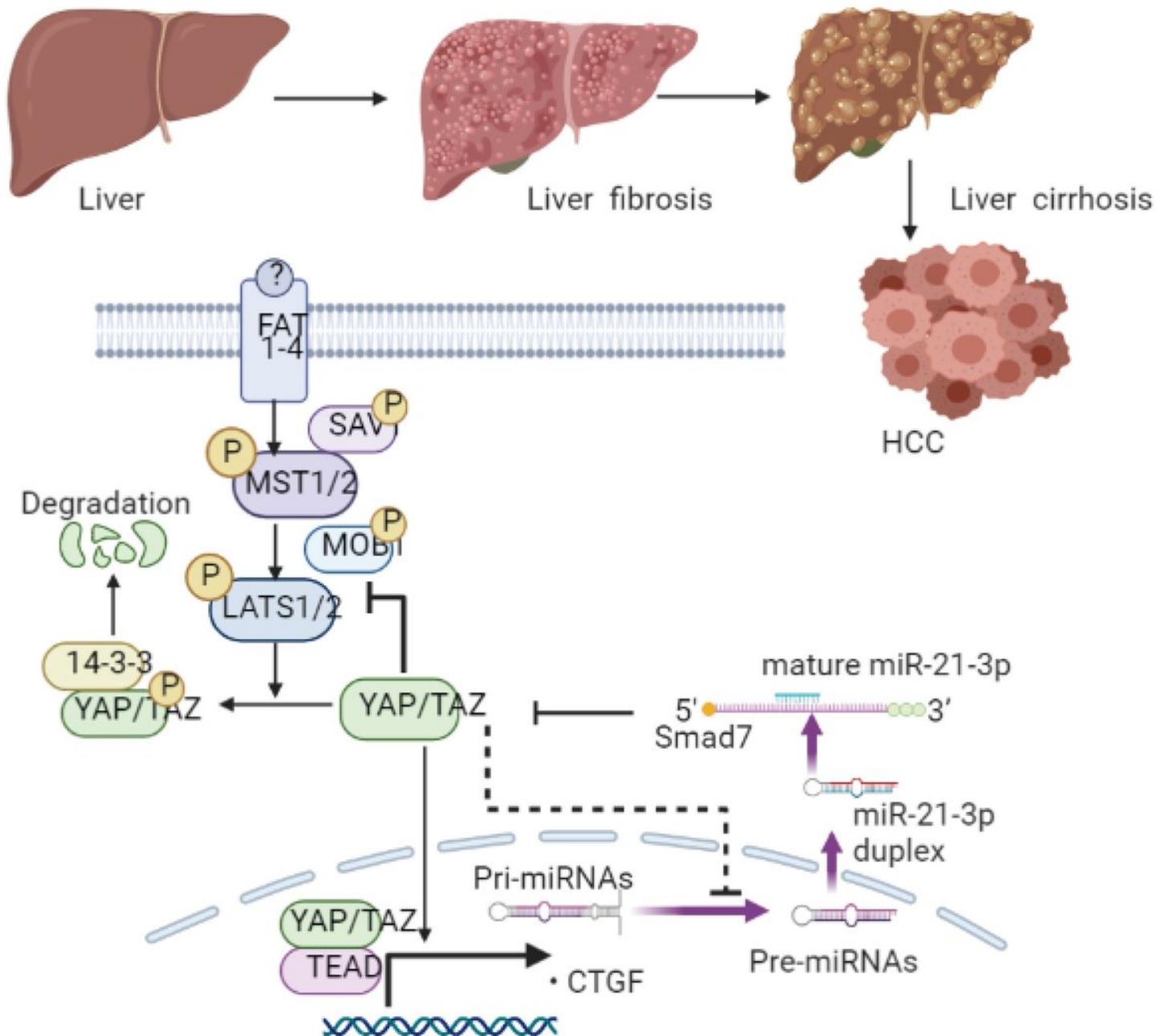


Figure 7

Graph abstract. Formation of most hepatocellular carcinoma experiences the slow process of liver fibrosis and cirrhosis. During this journey, Dysfunction of the Hippo signaling road and the TGF-beta

transduction pathway are commonly seen. Once Hippo is activated, phosphorylation LATS2 will phosphorylate Yap1 and leads to its degradation. Overexpressed miR-21-3p directly silences the expression of Smad7 and further promotes the expression of Yap1. Up-regulation of Yap1 translocates into nuclear and facilitates the downstream gene connective tissue growth factor (CTGF). This graph was drawn with online tool biorender gallery.

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