

# Evaluation of *Ferula Assa-Foetida* Accessions for Germination Parameters Under Cold Stratification to Overcome Seed Dormancy and Effect of Media Mixtures on Seedling Growth

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## Research Article

**Keywords:** Seed dormancy, chilling, germination, genotypes, Apiaceae and *Ferula assa-foetida*.

**Posted Date:** December 3rd, 2020

**DOI:** <https://doi.org/10.21203/rs.3.rs-112608/v1>

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1 **Evaluation of *Ferula assa-foetida* accessions for germination parameters**  
2 **under cold stratification to overcome seed dormancy and effect of media**  
3 **mixtures on seedling growth**

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11 **ABSTRACT**

12 *Ferula assa-foetida* L. is a perennial of Apiaceae family having seed dormancy which inhibits  
13 the germination. Oleo-gum resin obtained from the rhizomes of *Ferula assa-foetida* plant has  
14 several medicinal properties and used for the treatment of various diseases, pharmaceutical  
15 industries and in cooking of food in some countries. In present study, three different temperature  
16 treatments (5°, 15° and 25°C) were used to break seed dormancy in six *Ferula assa-foetida*  
17 accessions repeatedly for two years. Also the seedling survival rate with other morphological  
18 parameters like plant height, no. of leaves, leaf width, leaf length, root length and root diameter  
19 were observed on 3 month old seedlings in six different media mixtures during year 2019-20.  
20 The germination ranges from 3.63 (EC968466 at 25°C) to 81.88 percent (EC966538 at 5°C) with  
21 an average of 31.14 percent over all temperatures and genotypes. The mean germination time  
22 was ranged from 19.69 to 42.71 days with an average of 30.14 days. The highest germination  
23 (66.9%) and least mean germination time was observed at 5°C (20.85) which showed that this  
24 crop required a 5°C chilling treatment of about 20 days for breaking seed dormancy. The results  
25 pertaining to seedling survival experiment showed that media mixture of sand, soil, FYM and  
26 cocopeat (1:2:2:1 and 1:1:1:1 ratio) gave higher seedling survival rate (87.66%) and other  
27 morphological traits. It was also observed that the genotype EC966538 and EC968470 were the

28 best performer for overall germination as well as seedling survival parameters and could be used  
29 as base population in future selection and improvement breeding programs.

30 **Keywords:** Seed dormancy, chilling, germination, genotypes, Apiaceae and *Ferula assa-foetida*.

31 *Ferula assa-foetida* L. is a small perennial monocarpic herb belonging to family Apiaceae  
32 (Umbelliferae) and commonly known as "Heeng" in Hindi <sup>1</sup>. The plant height is about 1-1.5  
33 meter in length, large size compound leaves and a large size thick taproot with root hairs is  
34 present and has a pungent smell. This plant is native to Iran and Afghanistan<sup>2,3</sup>. About 170  
35 species are found all over the world and 60 species of *Ferula assa-foetida* are geographically  
36 distributed around North Africa, Central Asia and Europe<sup>4</sup>. This species is distributed at an  
37 elevation between 2000-4000m above mean sea level, with an annual rainfall of 250-350mm <sup>5</sup>.  
38 Two species of genus *Ferula* i.e. *Ferula alliacea* and *Ferula jaeschkeana* are found in India<sup>6</sup>. The  
39 oleo-gum resin is present in the fleshy tap roots and this exudate extracted from the roots is  
40 called asafoetida. *F. assa-foetida* is one of the most significant plant of Iran and Afghanistan<sup>7</sup>  
41 because of high export demand for its oleo-gum resin. It has medicinal use in traditional  
42 medicines <sup>8</sup>. Asafoetida plays an important role in cooking food as a condiment and in medicine  
43 for the treatment of various diseases in India, China, Nepal, Tibet and Iran. The extract of  
44 asafoetida is used to treat various diseases *viz.* Respiratory infections urinary, gastrointestinal and  
45 emmenagogues and also used to diagnose for a snake bite, insect bite and worm infection due to  
46 bite<sup>9,10</sup>. It is also antispasmodic, carminative and stimulant, diuretic, expectorant, anthelmintic  
47 and slightly laxative<sup>11</sup>. The oleo-gum resin of asafoetida contains 25% gum, 62% oleo-resin and  
48 approximately 3-7% essential oil<sup>12</sup>.

49 The main problem with the medicinal plants which are native to arid lands is that they germinate  
50 and grow well within their habitat or native environment, but fails to show good germination or  
51 growth in another environment<sup>13</sup>. For every plant, germination is the most important stage in its  
52 life cycle, which controls changes in its population with major useful consequences<sup>14</sup>. Apiaceae  
53 family shows very poor germination ability because of seed dormancy <sup>15,16</sup>. Seed dormancy is a  
54 state when a viable seed failure to complete germination <sup>17,18</sup>. It is an innate feature of seeds  
55 which regulates germination and completion of the plant life cycle<sup>19,20</sup>. But this period of the  
56 seed dormancy could be cut short by giving stratified cold treatments to the imbibed seeds. *F.*

57 *assa-foetida* seeds are difficult to germinate and take a long time period to germinate seed due to  
58 seed dormancy. Two types of seed dormancy have occurred as primary that are internal and  
59 external. The internal seed dormancy was belonging to physiological dormancy which can be  
60 removed by the chilling, chemical, hormonal and heating treatments but from all the treatments  
61 chilling treatment shows the best result to break seed dormancy<sup>21,22,23</sup>. To break seed dormancy  
62 there are different methods to germinate dormant seeds<sup>24</sup>. Chilling treatment plays a very  
63 important role to break seed dormancy in many species of Apiaceae family<sup>22</sup> and enhances the  
64 germination and speed of germination in dormant seeds.

65 However, germination of *Ferula assa-foetida* does not occur easily because of seed dormancy.  
66 Therefore, a systematic study is necessary to standardize germination parameter for a seed  
67 propagated plant having dormancy. Keeping this in view, the study was undertaken with the  
68 objectives to determine the effect of cold temperature stratification and genotypic variation on  
69 seed dormancy and also to identify suitable media mixture for seedling emergence and growth.

## 70 **Result and Discussion**

71 All the six accessions of *Ferula assa-foetida* were examined for seed viability, germination and  
72 seedling survival under various experiments during 2018 and 2019 (Fig. 1a and 1b) The results  
73 pertaining to seed viability, germination, germination parameters and seedling survival rate are  
74 given as in following headings.

### 75 **Seed viability**

76 Seed viability was examined in both the years in three tests repetitions. The results of analysis of  
77 variance (ANOVA) showed that the accessions were significantly different for seed viability  
78 (Table 2). However, the tests repetitions were found non-significant and suggested that the seed  
79 viability was not degraded over a time period of 18 months. Overall, 72.22 percent of seed  
80 viability (ranged from 56.00 to 92.00%) was observed (Table 3). The average highest seed  
81 viability was observed for EC968470 (84%) and EC966538 (84%), while the lowest one was  
82 observed for EC968469 (60%).

### 83 **Seed Germination**

84 The experiment of seed germination of six *Ferula assa-foetida* accessions were examined in  
85 2018-19 and 2019-20 (Fig. 1b). The germination data of both the years 2018-19 and 2019-20  
86 were analyzed individually as well as pooled after conducted Bartlett test to verify the  
87 homogeneity assumption for analysis of variance (ANOVA). The different temperature  
88 treatments, accessions and their interaction were found highly significant for all the parameters  
89 studied (Table 2). The results pertaining to germination, germination parameters and seedling  
90 survival rate are given as in following headings.

### 91 **Effect of various temperature treatments on germination**

92 The pooled and individual analysis of variance (ANOVA) for both the years 2018 and 2019  
93 showed significant difference ( $p \leq 0.01$ ) for temperature treatments (Table 4). The germination  
94 ranges from 3.63 percent (EC968466 at 25°C) to 81.88 percent (EC966538 at 5°C) with an  
95 average of 31.14 percent over all temperatures and accessions. The average seed germination  
96 percentage for all the accessions was highest (Table 5) at 5°C (66.90%) than 15°C (21.23%) and  
97 25°C (5.30%). The highly significant results of temperature treatments showed that seeds of *F.*  
98 *assa-foetida* require chilling treatment of 5°C for their germination. This indicates physiological  
99 endogenous dormancy in which factors within embryo inhibits seed germination and require  
100 chilling treatment (cold stratification) to initiate germination. This is most common form of seed  
101 dormancy in angiosperm plants<sup>26,43,44</sup>. Chilling temperature generally increase the production of  
102 germination promoting hormones thereby shifting the balance among promoters and inhibitors  
103 towards growth promoters<sup>45,46</sup>. In recent studies, down regulation of ABA and up regulation of  
104 GA content in *Hydysarum scoparium* seeds after cold stratification<sup>47</sup>. It is also important to  
105 obtain further valid information on plant growth promoting hormones for *F. assa-foetida* seed  
106 germination<sup>48</sup>. The significant positive effect of chilling treatment for breaking seed dormancy  
107 was also reported in *F. assa-foetida*<sup>49</sup>, *Ferula gummosa* and *Ferula ovina*<sup>50</sup>. In *Bunium persicum*  
108 species from same Apiaceae family were observed similar results<sup>51</sup>.

### 109 **Effect of genotypes on germination**

110 Genotypes and their interaction with temperature were significantly different ( $p \leq 0.01$ ) for  
111 germination percentage in pooled as well as individual analysis of variance (ANOVA) for both  
112 the years 2018-19 and 2019-20 (Table 4). The maximum germination (81.88%) was observed in

113 EC966538 accession at 5°C, while EC968469 accession showed lowest germination (55.50%) at  
114 the same (5°C) level of temperature. In case of adverse high temperature of 25°C, the accession  
115 EC966538 showed maximum germination (8.13%), while EC968466 had very poor germination  
116 (3.63%) at this adverse temperature (Table 5) and (Fig. 2). The results of genotypic effect on  
117 germination showed that the accession EC966538 has consistently higher germination  
118 percentage among all the accessions in all temperature treatments. Seed dormancy is the major  
119 challenge in *F. assa-foetida* and other *Ferula* species<sup>52,53,54,5</sup> and genetic background of this  
120 accession (EC966538) could be used in future germination improvement breeding programs.

### 121 **Other germination parameters**

122 Seeds of *F. assa-foetida* are not germinated in a single flush, and involvement of multi-level seed  
123 dormancy causes continuous germination up to several days even with some favorable  
124 environment. Hence, germination percentage should not be only the single parameter to access  
125 the germination capability of this crop. Thus, in present study we have observed several other  
126 parameters to identify best accession and environment for good germination. These parameters  
127 are germination index (GI), mean germination time (MGT), mean daily germination (MDG),  
128 coefficient of velocity of germination (CVG), peak value (PV), germination value (GV), days to  
129 25% germination (DG25%), days to 50% germination (DG50%), days to 75% germination  
130 (DG75%), radical length (RDL) and seed vigor index (SVI).

131 The pooled and individual analysis of variance (ANOVA) for both the years 2018 and 2019  
132 showed significant difference ( $p \leq 0.01$ ) for temperature treatments, genotypes and their  
133 interaction for all the other germination parameters studied (Table 4). Mean germination time  
134 (MGT) measures mean time required by any seed sample to initiate and terminate germination.  
135 Lower is the value of MGT, faster a seed lot has germinated<sup>55</sup>. The results pertaining to mean  
136 germination time (MGT) showed that germination was achieved very fast (Table 3) and (Fig. 3)  
137 at 5°C temperature treatment (20.85 days) than at 15°C (29.88) and 25°C (39.71). MGT ranges  
138 from 19.69 days (EC968469 at 5°C) to 42.71 days (EC968466 at 25°C) with an average of 30.14  
139 days over all temperatures and genotypes. The germination rate per day measured as mean daily  
140 germination (MDG) was also highest for 5°C (1.48 seedlings/day) than at 15°C (0.47) and 25°C  
141 (0.12). Over, all the temperature treatments, the accession EC968469 has lowest mean

142 germination time (28.80 days) at 0.59 seedlings per day germination rate. Similarly, the days to  
143 25%, 50% and 75% germination were also found lowest at 5°C (15.19, 18.54 and 24.06 days,  
144 respectively). The accession EC966538 has highest per day germination rate (0.84  
145 seedlings/day). Whereas, mean germination time decreased significantly by increasing  
146 temperature from 15°C to 20°C under constant temperature treatment<sup>56</sup>.

147 Germination index (GI) is a measure of both germination percentage and speed of germination. It  
148 gives maximum weightage to early germinating seeds and less weightage to late germination<sup>55</sup>.  
149 The GI ranged from 0.09 (EC968466 at 25°C) to 3.94 (EC966538 at 5°C) with an average of  
150 1.44 over all temperatures and genotypes. The average germination index for all the genotypes  
151 was highest (Table 5) at 5°C (3.47) than 15°C (0.73) and 25°C (0.14).

152 Coefficient of velocity of germination (CVG) denotes the rapidity of germination and increases  
153 with germination of seeds and time required for their germination is reduced<sup>55</sup>. CVG ranged  
154 from 2.35 (EC968466 at 25°C) to 5.08 (EC968469 and EC968470 at 5°C) with an average of  
155 3.59 over all temperatures and genotypes. Recently, the mean germination time, germination  
156 index and coefficient variation of germination were studied for *Magnolia grandiflora* plant after  
157 cold stratification<sup>40</sup>. The results pertaining to PV, GV and SVI were also in accordance to results  
158 of mean germination time and per day germination rate *i.e.* the temperature treatment of 5°C and  
159 accession EC966538 followed by EC968470 were found best performer (Table 3). Also, the  
160 radical length (Fig. 4 and Fig. 5) and seed vigor index was highest at 5°C (2.55 cm and 1.70,  
161 respectively) followed by 15°C (1.71 cm and 0.36,). However, in overall genotypic effects, the  
162 highest radical length and seed vigor was found for EC968470 (1.87 cm and 0.84, respectively).  
163 These seed germination parameters also studied in some Himalayan leguminous and actinorhizal  
164 plants<sup>57,58</sup>.

### 165 **Seedling survival rate**

166 To check the seedling survival rate, all newly germinated plants were examined in the year 2019-  
167 20 under six different media mixtures (Table 1) for various morphological traits *viz.* plant height  
168 (cm), number of leaves, leaf length (cm), leaf width (cm), root length (cm) and root diameter  
169 (mm). The results of analysis of variance for seedling survival traits showed that all the  
170 genotypes and media mixtures were highly significant ( $p < 0.01$ ) for all traits studied (Table 6).  
171 While, genotype × media interaction effect was non-significant for number of leaves, leaf length

172 and root diameter. It showed that the studied accessions were genetically diverse for seedling  
173 survivals and could be utilized as base population for further selection and breeding programs.

174 The mean performance of different media mixture and genotypes showed that the accession  
175 EC968466 in media M4 and EC968467 in M5 have highest seedling survival (91.50%) (Table 5)  
176 and (Fig. 6 and Fig. 7). Over all media mixtures, M5 has the highest seedling survival rate  
177 (87.66%), followed by M4 (87.08%). Media M4 has highest plant height (21.49 cm), number of  
178 leaves (5.27), leaf length (9.22 cm), leaf width (6.58 cm), root length (11.93 cm) and root  
179 diameter (5.36 mm). It showed that the combination of soil, sand, FYM and cocopeat in 1:2:2:1  
180 ratio was the best mixture to attained maximum survival of *F. assa-foetida* seedlings. While,  
181 media M6, which was only soil has lowest seedling survival rate (46.50) and also found lowest  
182 for all other seedling survival parameters, which indicated that this crop needs a survival media  
183 at initial stage to get good seedling establishment.

184 Genotypic effects on seedling establishment showed that accession EC968467 has highest  
185 survival rate (75.94%) and number of leaves (5.17) but lowest root diameter (4.16 mm).  
186 Accession EC968470 has highest plant height (15.15 cm) and root length (9.67 cm), while  
187 accession EC966538 was best for other leaf parameters *i.e.* number of leaves, leaf length and leaf  
188 width Table 7. It was observed that the accession EC968469 has lowest seedling survival rate  
189 (65.33%), leaf length (6.58 cm), leaf width (4.88 cm) and root length (8.30 cm). vermiculite as  
190 suitable media for seed germination of *Jatropha Curcas*<sup>57</sup>. Some Himalayan leguminous and  
191 actinorhizal plants shows higher germination on moistened filter paper as compared to mixture  
192 of soil and sand<sup>58</sup>.

## 193 **Summary**

194 According to present study, we concluded that the chilling treatment at 5°C is more effective for  
195 dormancy breaking of *Ferula asafoetida* seeds. It shows that dormancy is caused by an inhibiting  
196 chilling in the interior or exterior surface layers of seeds. Further studies are required to explain  
197 the agro-practices to cultivate this endangered plant. Cold treatment at 5°C for about 20 days was  
198 appropriate to breaking seed dormancy and maximum seed germination by. Our finding suggests  
199 that cold treatments are commercial and effortlessly applicable by poor farmers and nursery  
200 manual workers in developing bulk planting material, over costly supplementary technicalities

201 and plant growth regulators (PGR). The results of germination in Laboratory condition can also  
202 be applied to propagation of plants that would help conservation programs within the study area.  
203 But maximum germination of *F. asafoetida* through seed is very difficult and almost very low.  
204 Media mixture of sand, soil, FYM and cocopeat (1:2:2:1 and 1:1:1:1 ratio) gave higher seedling  
205 survival. It was also observed that the accession EC966538 and EC968470 were the best  
206 performer for overall germination as well as seedling survival parameters.

## 207 **Material and methods**

### 208 **Plant material**

209 Six accessions of *Ferula assa-foetida* (Heeng) seeds used in the study were EC966538,  
210 EC968466, EC968467, EC968468, EC968469 and EC968470 procured through National Bureau  
211 of Plant Genetic Resources, New Delhi. The seeds were cleaned with removing the chaff  
212 material and damaged or immature seeds. Healthy and mature seeds separated from the seed lot  
213 were stored under ambient laboratory conditions prior to use in the experiments. Weight of 100  
214 seed was 1.82g. The present study was conducted during 2018-2019 and 2019-2020 in the  
215 laboratory of Agrotechnology Division, CSIR-Institute of Himalayan Bioresource Technology,  
216 Palampur (Himachal Pradesh), India (N 32°6.36546' latitude, E 76°33.52122' longitude at an  
217 altitude of 1310.0 m with average annual rainfall 2493mm and average annual temperature is  
218 19.1°C). All the experiments of germination and seedling survival were conducted in Completely  
219 Randomized Design (CRD) design in with four replications.

### 220 **Seed viability test**

221 Seeds of *F. assa-foetida* were examined for seed viability with the help of Tetrazolium test  
222 (2,3,5-tri-phenyltetrazolium chloride). For that, 1% Tetrazolium solution was prepared by adding  
223 1g 2,3,5-tri-phenyl-2H-tetrazolium chloride (TTC) in 100 ml of doubled distilled water in a  
224 brown bottle, mixed well and confirmed the pH of the solution at 7. Further, four replicates of 25  
225 seeds from each accessions were dissected using a magnifying lens. Then the dissected seeds  
226 with embryo were kept in 1% tetrazolium solution and incubate at room temperature for 24 hours  
227 in dark <sup>19</sup>. Seeds were evaluated on their staining pattern and colour intensity as the bright red  
228 color stained seeds considered as viable while partially or light stained seeds were considered as

229 non-viable <sup>25,26</sup>. The seed viability test was repeated three times in every 6 months during the  
230 study period *i.e.* in 2018, 2019-I and 2019-II.

### 231 **Surface sterilization**

232 Healthy seeds of *F. assa-foetida* selected on the basis of their shape and size were surface  
233 sterilized by pre-washing with tap water for 1h and then soaking the seeds in 1% sodium  
234 hypochlorite solution (NaOCl) with tween-20 (2 drops /100 ml) for 25 min and then washed with  
235 sterilized double distilled water to remove traces of sterilizing agents before putting in petri-  
236 dishes.

### 237 **Seed germination**

238 To overcome the seed dormancy in *F. assa-foetida*, an experiment with different temperatures  
239 and accessions was conducted during 2018-19 and 2019-20. For the stratification treatments,  
240 seeds of six different accessions were kept at three controlled and constant temperature  
241 treatments *i.e.* 5°C, 15°C and 25°C for germination. For germination tests, four replicates of 25  
242 seeds were incubated in petri dishes lined with double layer of sterile Whatman no.1 filter paper  
243 moistened with 5 ml double distilled water. All the petri plates were sealed with parafilm and  
244 seeds were allowed to germinate at 5°C temperature in a cold chamber (Blue star company),  
245 15°C and 25°C temperature in growth room for a time period of 45 days. During the incubation  
246 period, filter papers were kept moist with distilled water. The germination counts were observed  
247 daily from first to last day of maximum seed germination <sup>27,28,29,30</sup>. Seeds at the time of radicle  
248 emergence were considered germinated <sup>31</sup> and an interval 45 days was found enough to get  
249 maximum germination and differentiation of non-dormant seeds from dormant ones <sup>22,32,33</sup>.

### 250 **Germination parameters**

#### 251 **Seed germination**

252 Seed germination (%) was recorded after 45 days of chilling treatment and under different  
253 temperature. The total number of germinated seeds were counted and germination was computed  
254 in percentage using following formula:

255 Germination (%) =  $\frac{\text{Number of seeds germinated}}{\text{Total number of seeds sown}} \times 100$

256 **Mean germination time**

257 Mean germination time (MGT) is considered as an indicator of seedling emergence in field and  
258 calculated by using formula<sup>34</sup>.

259 Mean germination time (Seed per day) =  $\frac{\sum F_i n_i}{N}$

260 Where,  $n_i$  is the number of germinated seeds on  $f_i^{\text{th}}$  day of observation during germination time  
261 (from 0 to 45 days) and N is the total number of germinated seeds.

262 **Coefficient of velocity of germination**

263 Coefficient of velocity of germination (CVG) was computed using following formula<sup>35</sup>.

264 
$$\text{CVG} = \frac{N_1 + N_2 + \dots + N_i}{100 \times (N_1 T_1 + \dots + N_i T_i)}$$

265 Where  $N_i$  is the number of seeds germinated on  $T_i^{\text{th}}$  day of observation from seeding.

266 **Germination index**

267 Germination index (GI) was calculated by using formula<sup>36</sup>.

268 Germination Index (GI) =  $(45 \times N_1) + (44 \times N_2) + \dots + (1 \times N_{45})$

269 Where  $N_1, N_2 \dots N_{45}$  is the number of germinated seeds counted every day till 45<sup>th</sup> day and the  
270 constants (45, 44, ..., 1 etc) are the weights provided.

271 **Mean daily germination**

272 Mean daily germination (MDG) was calculated by using formula<sup>37</sup>.

273 Mean daily germination (MDG) =  $\frac{\text{Total number of germinated seeds}}{\text{Total number of days}}$

274 **Peak value**

275 Peak value (PV) was calculated by using given formula<sup>37</sup>.

276 
$$\text{Peak value (PV)} = \frac{\text{Highest seed germinated}}{\text{number of days}}$$

### 277 **Germination Value**

278 Germination Value (GV) was calculated by using formula<sup>38,39,40</sup>.

279 
$$\text{Germination Value (GV)} = (\sum \text{DGS}/N) \times \text{GP}/10$$

280 Where DGS is the ratio of cumulative germination percentage to the number of days from  
281 seeding, N is days counts from the germination initiation and GP is final germination percentage.

### 282 **Seed vigor index**

283 The seed vigor index (SVI) was computed using following formula<sup>41</sup>.

284 
$$\text{Seed vigor index (V}_i\text{)} = \frac{L_s \times P_g}{100}$$

285 Where V<sub>i</sub> is the vigor index, L<sub>s</sub> is the length of seedling and P<sub>g</sub> is germination percentage.

### 286 **Days to 25%, 50% and 75% germination**

287 Days to 25%, 50% and 75% germination were calculated when 25, 50 and 75 percent  
288 germination of *Ferula assa-foetida* seeds was achieved.

### 289 **Radical length**

290 Radical length was measured with the help of meter scale in centimeter (cm).

### 291 **Media Mixtures**

292 After cold stratification treatment germinating seeds of all the six accessions were transferred to  
293 different media mixtures under glasshouse conditions to study the survival and growth of the  
294 germinating seedlings Table 1.

### 295 **Morphological data collection**

296 After three months, data from different media mixtures on seedling survival and other growth  
297 parameters *viz.* plant height (cm), number of leaves, leaf length (cm), leaf width (cm), root length  
298 (cm) were recorded with the help of geometrical scale and root diameter was measured with the  
299 help of Vernier calliper in millimeter (mm).

300 Seedling survival rate was calculated after three months of transferred seedling in the poly  
301 sleeves using following formula.

302  $\text{Survival rate} = (\text{Total number of survived seedling} / \text{Total number of transferred seedlings}) \times$   
303 100

### 304 **Statistical analysis**

305 Analysis of variance (ANOVA) for all the three experiments *i.e.* seed viability, germination and  
306 seedling survival were computed separately using PROC ANOVA in SAS v9.4<sup>42</sup> by considering  
307 all the variables as fixed effect.

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# Figures

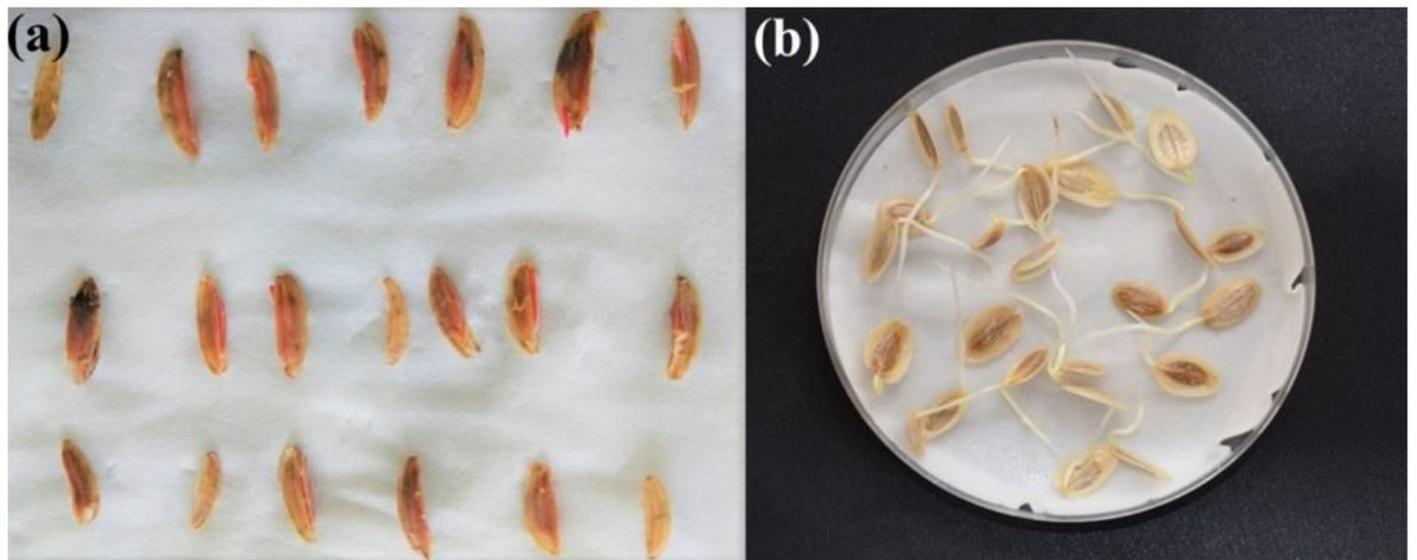


Figure 1

TTC stained seed (a) and germinated seedling (b) of *Ferula assa-foetida*

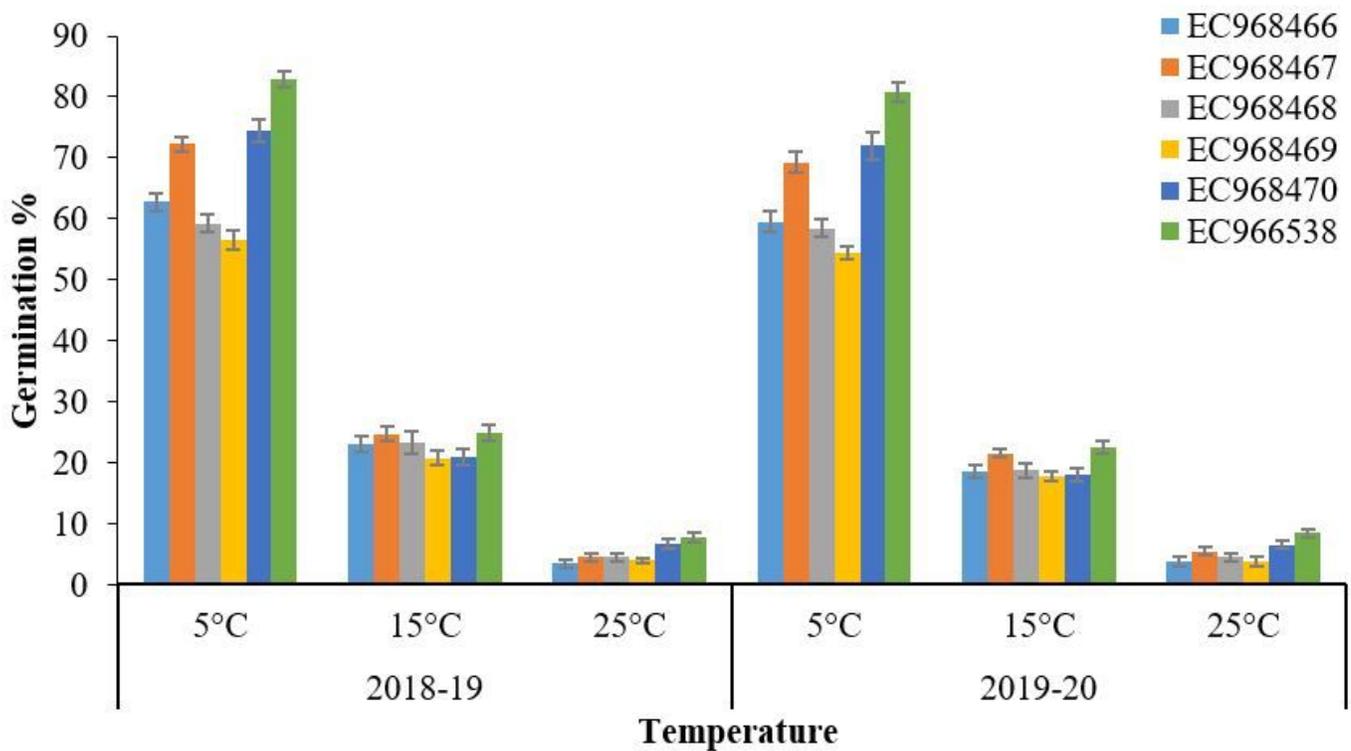


Figure 2

Germination (%) of different accession plotted over different temperature in 2018-19 and 2019-20.

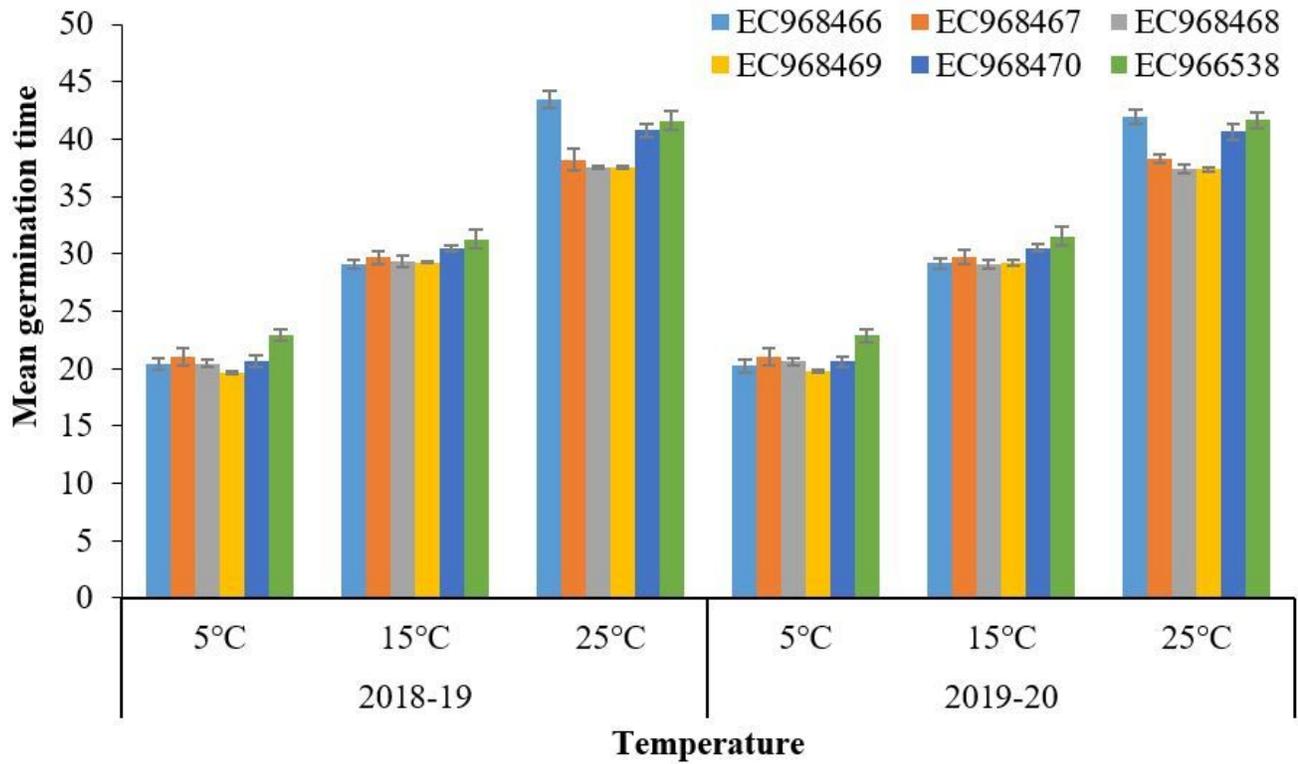


Figure 3

Mean Germination time of different accessions plotted over different temperature in 2018-19 and 2019-20

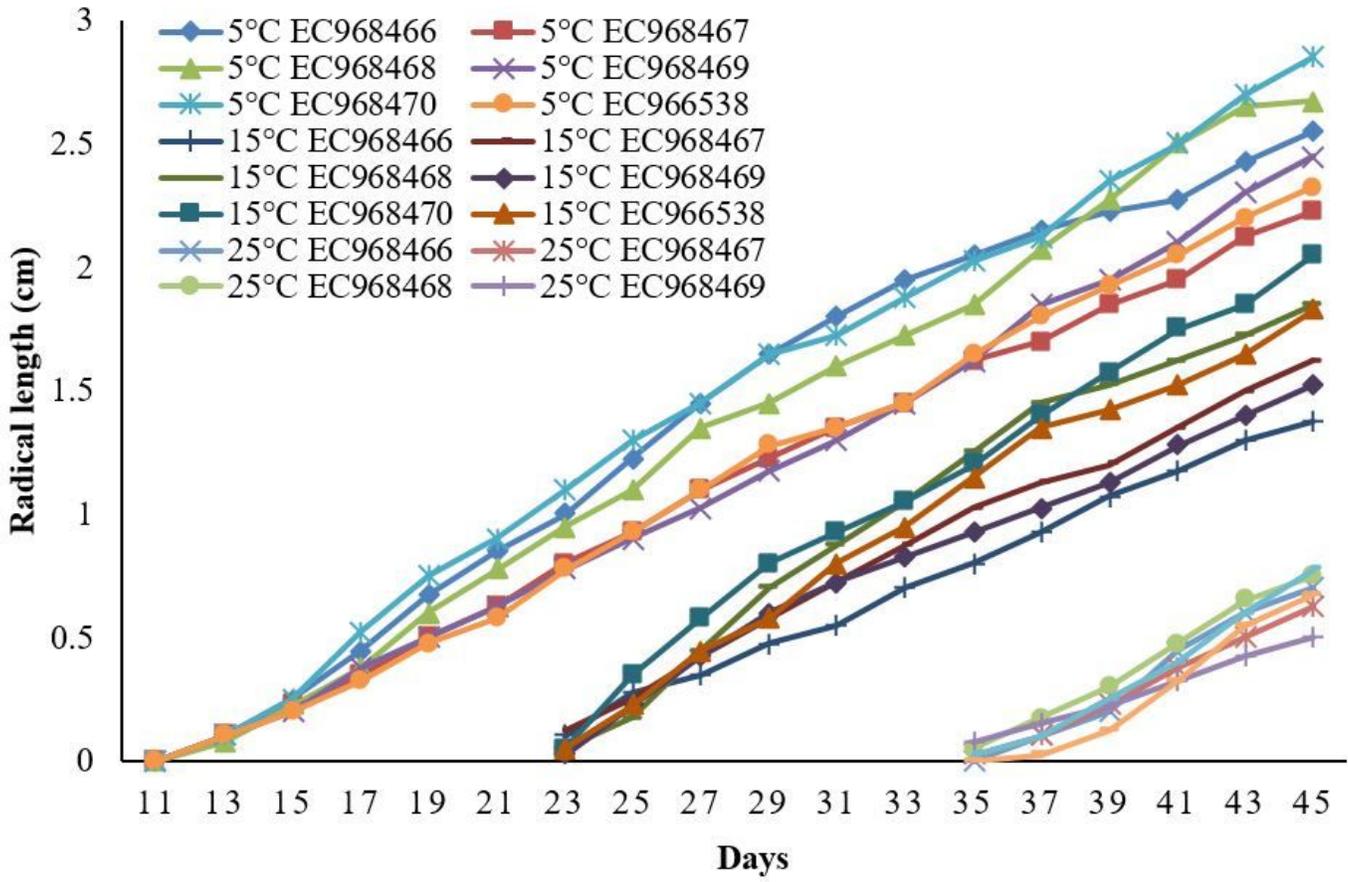


Figure 4

Rate of root emergence per day in year 2018-19.

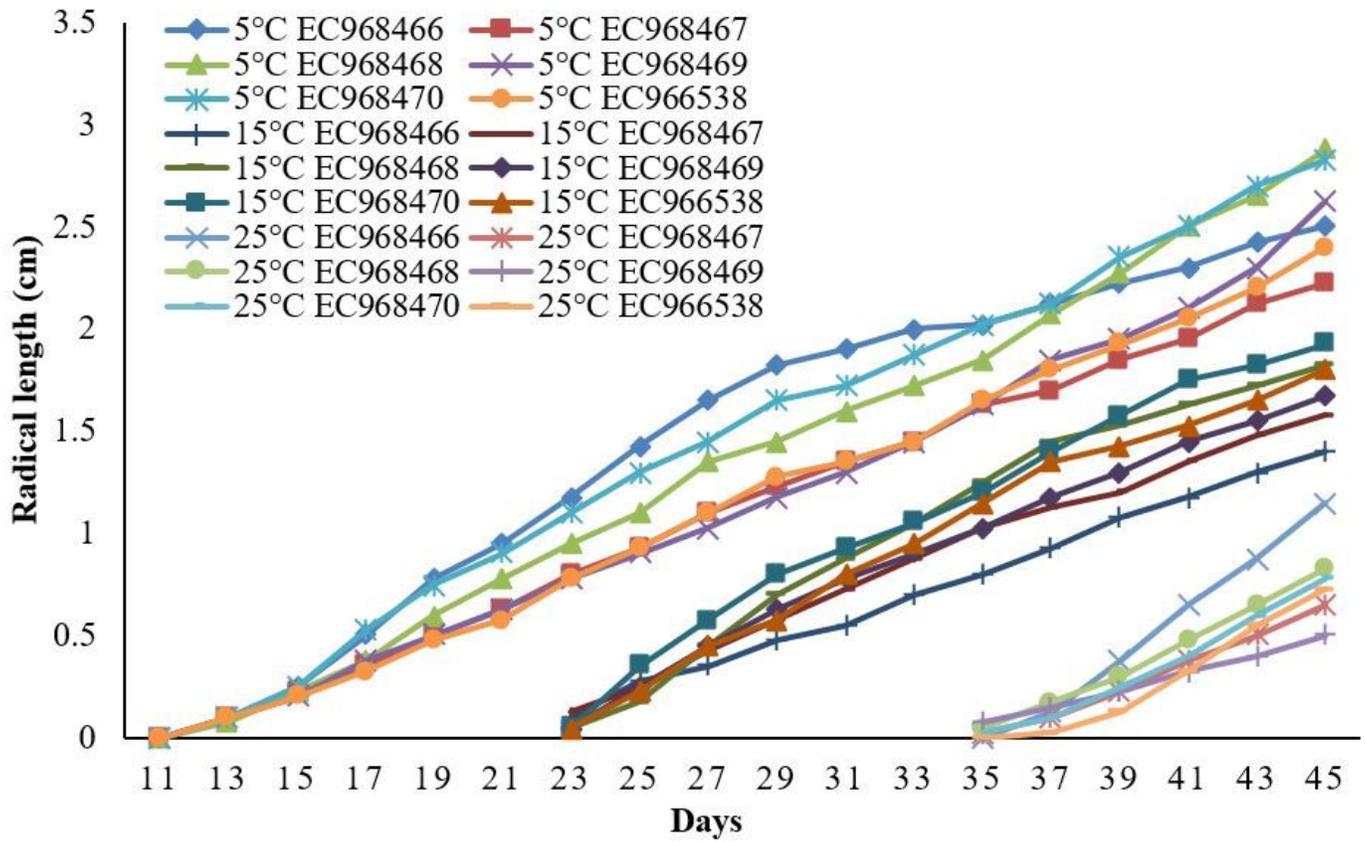
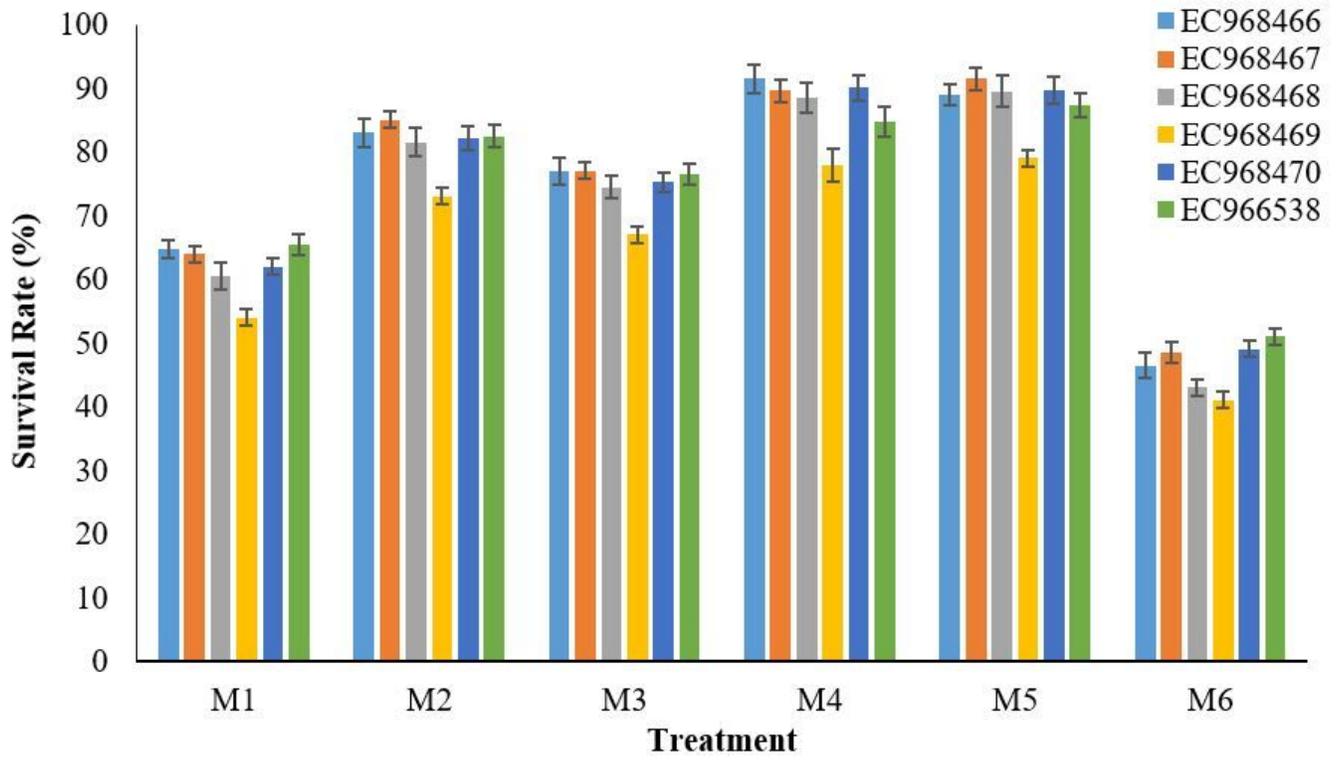


Figure 5

Rate of root emergence per day in year 2019-20.



**Figure 6**

Survival rate of seedling under different media mixture treatments.

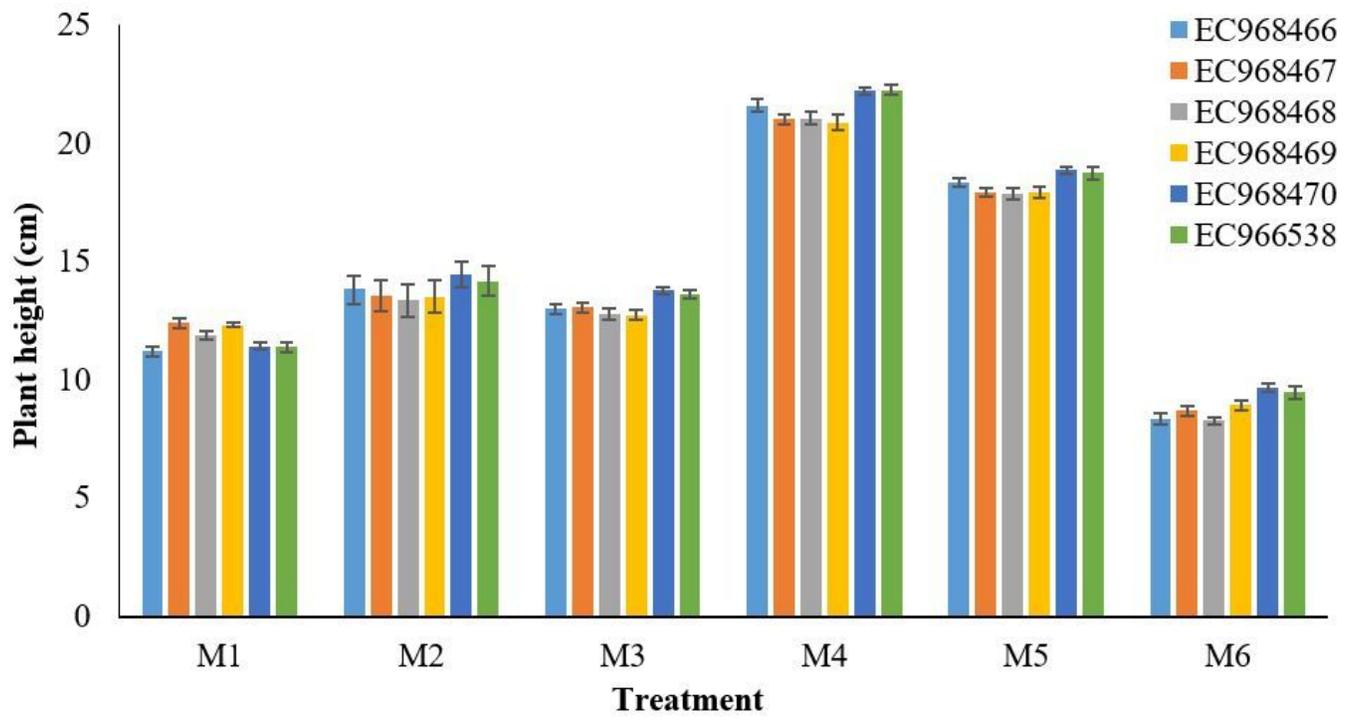


Figure 7

Plant height under different media mixture treatments