

The Role of Eco-Friendly Soil Amendments Co-Application on the Performance of Lingrain (*Linum Usitatissimum* L.) Under Late-Season Irrigation Limitation

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Abstract

This study's main purpose was to investigate the probable amelioration of limited irrigation conditions by soil amendments for lingrain plants. The experiment was accomplished as a completely randomized factorial design along with three replications. The first factor was green manure (without (Gc) and with *Trifolium pratense* (Gr)), the second factor consisted of *Rhizophagus irregularis* mycorrhiza (Fm), vermicompost (Fv), both of mycorrhiza and vermicompost (Fm+v) and none of them (Fc), and also the third factor was irrigation regime (full irrigation and late-season water limitation). Green manure, vermicompost and mycorrhiza single-use enhanced the plant performance under water limitation conditions in comparison with the control. However, in the presence of vermicompost, along with green manure or mycorrhiza developed a positive synergistic effect on most of the traits. Combining green manure with the dual fertilizer (Fm+v) resulted in the vermicompost and mycorrhiza synergistic effects, especially under limited irrigation. Consequently, the triple fertilizer (Gr×Fm+v) experienced the highest amount of LRWC, root colonization, leaf nitrogen, chlorophyll a, chlorophyll b, carotenoids, antioxidant enzymes activity, grain yield and oil yield, which would lead to more resistance of plants to limited irrigation conditions.

Introduction

Climate change and water scarcity have sparked drought as the most critical environmental stress that affect the crops growth, development and productivity in all over the world⁹¹. Along with the increase in greenhouse gas emissions under the constant global climate change, an increase in the drought intensity and incidence is expected soon. Approximately half of the agricultural lands in all over the world are constantly endangered to water shortages, resulting in a 50% reduction in grain yield²³. Drought stress can seriously disturb secondary metabolites concentration, also along with the morphological and physiological traits^{24,64}. Plants could create different mechanisms in order to defend against water stress, for instance: root development increasing for more water absorption, increasing the free radical scavenging enzymes activity, and osmolytes accumulation for moderating the osmotic pressure^{40,95}. Plant resistance to drought can also be increased using other external methods, for instance; by different soil amendments of which the eco-friendly treatments (for example mycorrhizal inoculation and organic fertilizer like vermicompost and green manure) have great importance. Organic fertilizers increase the products yield and quality by soil structure improving and nutrient availability increasing⁷. using nutrients achieved from organic sources helps maintaining the organic matter balance along with improving the chemical, physical and biological properties of the soil⁶⁰. Green manure plays a significant role in nutrients supplying for crops, decreasing dependency on chemical fertilizers, enhancing the crops yield, enriching the cultivated fields ecological environment, and lowering the soil destruction and contamination⁸⁶.

Vermicompost, derived from organic wastes of the earthworm, has nutrients that are readily absorbable by a plant. Vermicompost as a nutrient-rich and nature-friendly substance has several potential applications as soil conditioners²⁰. It stimulates microbial growth and activity and increases the soil nutrient mineralization. It increases fertility and the soil quality and contains plant growth stimulating agents (like vitamins, hormones, and enzymes), which help the plant grow better⁹⁶. Arbuscular mycorrhizal fungi (AMF) are omnipresent in land-dwelling ecosystems and indicates symbiotic interactions with many plant species roots⁵¹. The AMF can protect plants against the biotic and abiotic stresses by different mechanisms such as increasing the nutrient absorption, photosynthetic activity, osmolytes accumulation, antioxidant enzymes production, and also improving the rhizosphere environment⁹³. The methods used by the AM fungi for increasing the host plants-water relationships are not clear enough, but this may be caused by the water cumulative absorption through exterior hyphae, the adjustment of the stomatal system, the increased antioxidant enzymes activity and nutrients uptake mainly phosphorus¹.

Lingrain is one of the earliest identified cultivated plants, and since ancient times, it has been extensively cultivated for its fine, cellulose-rich, bast fibres and its oil⁴⁸. Lingrain oil is a superb dietary supplement being rich in omega-3 fatty acids and

α -linolenic acid. The oil is also benefited in the industrial raw materials production⁵⁸. Although, the worldwide lin grain harvested area and its production went down from 1993 to 2014, the oil yield increased at a rate of 1.72%. Hence it was cultivated on 2.3 million hectares of lands in all over the world with 560000 tons of oil production²².

There is often a severe shortage of water supply for irrigation of crops in the late growing season in most semi-arid countries, including Iran, which coincides with mid and late-summer. The significant effect of various soil amendments, including organic and biologic fertilizers, has separately been reported on improvement of different plants performances under water deficit stress. Since Iranian and most semi-arid regions soils are poor in organic matter, and few available studies were conducted on the simultaneous integrated use of organic and biologic fertilizers, this research purpose was to investigate the effect of single and combined use of these eco-friendly fertilizers for possible mitigation of lin grain performance under late-season irrigation limitation.

Results

Leaf Relative water content (LRWC)

LRWC considerably decreased in water-limited condition. However, Green manure noticeably increased the LRWC in mycorrhizal plants (Fm and Fm + v). In both irrigation conditions, single and combined use of vermicompost and mycorrhiza had the ability of increasing the LRWC significantly, in comparison with the control, with the highest increase (92.56%) belonging to the Fm + v as shown in Table 3.

Colonization

Sole and dual usage of mycorrhiza and vermicompost (Fv, Fm and Fm + v) would lead to more colonization in comparison with non-fertilized treatment (Fc) (Table 3). Green manure usage increased colonization in all fertilizer treatments except Fc in both irrigation and mycorrhizal plants in water limited condition. The greatest colonization was achieved in the combination of mycorrhiza and vermicompost (Fm + v) that were co-applied with green manure as indicated in Table 3. Moreover, limited- irrigation significantly declined the colonization in all fertilizer treatments (Table 3).

Leaf N

Water limitation decrease the levels of leaf nitrogen. The single and dual usage of vermicompost and mycorrhiza would result in increased leaf nitrogen in both irrigation conditions in comparison with Fc. Application of green manure and mycorrhiza significantly elevated leaf nitrogen in both irrigation conditions except fv. Green manure combination with mycorrhiza and vermicompost could generate the highest leaf nitrogen percentage as indicated in Table 3.

Leaf P

Water inadequacy could reduce the leaf P concentrations. Under normal irrigation conditions, single and double fertilizers (Fm, Fv and Fm + v) brought approximately a higher percentage of leaf P in comparison with control (Fc), but there was no significant difference amongst the fertilizer treatments (Table 5).

Although, dual usage of mycorrhiza and vermicompost (Fm + v) accumulated more P in the leaf, but it indicated no significant difference with the single applications of mycorrhiza and vermicompost (Fm and Fv) in water-limited condition (Table 5). In second year leaf P increased significantly than first year (Table 6).

Leaf K

Water stress strongly decreased the leaf K content in all fertilizer treatments. Leaf K percentage was higher in second year in comparison of first year (Table 5). Under normal irrigation conditions Fm + v) were more effective in leaf K content increasing in comparison with single vermicompost and mycorrhiza. However, they were not significantly different from

each other under limited irrigation conditions. On the other hand, they all produced more leaf K in comparison with the control treatment (Fc).

Photosynthetic pigments

Water limitation significantly reduced chlorophyll a, chlorophyll b, and carotenoids contents. Green manure application soared chlorophyll a content in both irrigation condition. Also, Fm + v had the highest content of chlorophyll a in all conditions, after that Fm and Fv were in the second and third place, respectively. In addition, the lowest chlorophyll a quantity belonged to Gc × Fc (Table 3).

Green manure increased the amount of chlorophyll b, and single use of mycorrhiza and vermicompost could not affect the chlorophyll b under full irrigation conditions, but it mycorrhizal plants resulted in a significant increase of chlorophyll b content under limited-irrigation (Table 3). Using green manure significantly increased the level of carotenoids in Fm + v in both full irrigation and late-season water stress. In both conditions, the combined treatment of vermicompost and mycorrhiza produced the highest carotenoids content as shown in Table 3.

Proline

At this stage, Water limitation significantly increased. green manure didn't change the proline concentration except (fm + v) in full irrigation regime. The single and dual application of vermicompost and mycorrhiza increased the proline content in water-limited conditions, also Fm + v indicated the highest proline accumulation under water limitation. Also the single usage of fertilizers had same effect on proline concentration in both irrigation regimes (Table 5).

Total soluble sugar

Total soluble sugar concentration enhanced in second year in comparison of first year in both irrigation regimes (Table 6). Application solo (fm) and (fv) didn't have significant effect on TSS concentration in full irrigation regime. Also fm + v decreased TSS concentration in this condition. All fertilizer sources increased TSS concentration in irrigation cut-off condition. So that the dual fertilizer treatment (Fm + v) produced more TSS than the others under irrigation cut-off condition (Table 5).

Glycine betaine (GB)

Water limitation considerably enhanced the levels of GB in grain leaves. Although, the application of vermicompost and mycorrhiza increased GB content under limited-irrigation circumstance, but the highest GB was obtained from dual fertilizer treatment (Fm + v). Additionally, under full irrigation, only mycorrhizal fertilizers (fm, fm + v) led to an increase in GB content (Table 5).

Hydrogen peroxide

Drought stress considerably increased the level of leaf hydrogen peroxide. However, using fertilizer treatments caused a significant decline in its amount, in comparison with the control (Fc). Dual fertilizer (Fm + v) brought approximately the highest reduction percentage for hydrogen peroxide. The mycorrhiza-containing treatments more effectively reduced the amount of hydrogen peroxide compared to vermicompost (Table 5). Green manure usage had significant effect on H₂O₂ concentration in all treatments except (Fv) in both irrigation regimes (Fig. 1).

Malondialdehyde (MDA)

Water limitation enormously increased the levels of MDA, which was as a result of lipid peroxidation increasing. However, single and dual usage of the fertilizers resulted in a remarkable drop in MDA content compared to the control (Fc). Dual fertilizer (Fm + v) brought approximately the maximum reduction for MDA. The mycorrhizal treatments have the ability of decreasing the level of MDA more effectively, in comparison with vermicompost. Also, the single application of green

manure was helpful in the MDA level reducing. Green manure usage decreased MDA level only in mycorrhizal treatments in irrigation cut-off irrigation (Table 4).

Catalase (CAT)

Water limitation increased CAT activity. Incorporation of green manure under water limitation increased the activity of CAT enzyme in all treatments (Fig. 2). By applying mycorrhiza, with or without vermicompost, indicated a higher influence on the enzyme activity under water limitation, as well. Furthermore, the highest activities of CAT (8.71, 8.61. mmol min⁻¹/g FW) were observed in mycorrhizal plants (Table 5).

Ascorbate peroxidase (APX)

Water deficit elevated the APX activity. Green manure increased the activity only under water limitation condition in all treatments except (fm). Furthermore, vermicompost and mycorrhiza, single or combined, had a more stimulatory effect on the enzyme activity under water limitation. The mycorrhiza and vermicompost combination caused the highest activity of APX (Table 4)

Superoxide dismutase (SOD)

In both years, SOD activity was significantly increased by water limitation as shown in Table 6. The application of green manure enhanced the activity of SOD in the limited-irrigation situation (Fig. 3). The synergistic effects of the co-application and the separate usages of mycorrhiza were more than that of vermicompost under limited-irrigation (Table 5).

Guaicole peroxidase (GPOX)

Water deficit elevated the Guaicole peroxidase activity. Green manure increased the enzyme activity in all treatments except vermicompost in full irrigation regime. Furthermore, enzyme activity was equal when using mycorrhiza under both irrigation conditions. The mycorrhiza and vermicompost combination caused the highest activity of GPOX (Table 4)

Grain yield

Limited irrigation significantly reduced the grain yield compared to full irrigation; however, the application of green manure increased grain yield relative to non-application of green manure in this condition. Although, co-application of green manure with vermicompost and mycorrhiza had no influence on grain yield in full irrigation. Green manure application under complete irrigation situation produced the maximum grain yield (4862 kg ha⁻¹) in Fm + v treatment. In addition, using green manure reduced the adverse effects of water restriction on grain yield, and the presence of mycorrhiza and/or vermicompost will additionally mitigate these adverse effects (Table 4).

Oil yield

The sole and conjoint usage of mycorrhiza and vermicompost along with co-application of green manure under full and limited irrigation would result in an increase in oil yield during comparing them with their corresponding controls (Table 4).

Using mycorrhiza was more effective oil yield in increasing compared to vermicompost, but the dual fertilizer produced the highest oil yield in both irrigation regimes with or without green manure. However, adding green manure to them produced more oil yield than control (Gc × Fc). Furthermore, the green manure inclusion to Fm + v produced the maximum level of oil yield (1602.66 kg h⁻¹) under full irrigation condition as displayed in Table 6.

Discussion

Water deficit significantly reduced the leaf relative water contents (LRWC)⁵⁹. By Considering the high water absorption efficiency of the soil by peripheral hyphae³⁶, using different types of mycorrhizal fungi could cause a substantial rise in the water status of inoculated plants, which is in association with non- mycorrhizal plants, under water- limited conditions⁹⁷.

Using organic fertilizers improves the water potential and maintains leaf turgor pressure, which is in agreement with this study result¹². With respect to the green manure and vermicompost roles in soil organic matter and water content enhancement, dual-fertilized plants (Fm + v) indicated high percentage of LRWC in green manured plots.

This study results demonstrated that water stress inhibited mycorrhizal colonization in linrain roots. These results are in agreement with earlier findings, proposing that mycorrhization will be decreased at low levels of soil moisture⁷⁹. AMF are obligate biotrophic fungi, consequently very slow germination of spores and expansion of the hyphae or their complete inhibition under drought conditions could elucidate the drought adverse consequences on colonization³². Therefore, lower colonization can occur because of two reasons; the carbon obtainability reducing from host plants and also the fungal spore germination reducing in limited water conditions⁹². Up to the best of our knowledge, organic matter can affect the activity and frequency of soil microorganisms like AMF, which can increase the plant nutrients⁵³. Vermicompost promotes the mycorrhiza colonization in the roots^{39,49,80}. The highest percentage of AMF colonization (66%) in *Ocimum basilicum* roots was observed in using vermicompost + *G. intraradices* combination in comparison with control⁸⁹. Therefore, the combination of vermicompost, mycorrhiza and green manure exhibit high root colonization under full irrigation condition in this research.

Nutrients absorption directly relates to the soil water status, thus the nutrient flow from soil to root decreases in moisture shortage conditions⁵. With respect to better solubility of phosphorus in irrigated soil, the phosphorus concentration of the leaves indicates a reduction along with water restriction, which is probably caused by reduced solubility, mobility (mass flow or propagation), transfer between the roots and branches, and also the absorption of P under drought conditions^{43,77}. Mycorrhizal inoculation can increase the infection percentage and absorption range in the extramatrical mycelium, and after that increase the nitrogen absorption rate and phosphorus and improve the nutrition in plants⁶³. However, inoculated plants under limited irrigation contained less nitrogen and phosphorus content, which may be due to drought constraints in mycelium development and reduced absorption range, in comparison with infected plants in well-watered condition⁵².

Green manure will increase the soil holding-capacity for water and nutrients content and mobility in the upper layer of soil, by providing organic matter. Plants can use these elements, and therefore increase their growth and productivity⁸². Moreover, there is an extensive record on the green manure advantages for nitrogen nutrition of succeeding crops^{21,45,68}. Mineralized nitrogen from leguminous plant residues can provide considerable quantities of N to a subsequent crop⁷³. Green manure effect on phosphorous availability could be caused from its P uptake and unavailable inorganic P changing to more available organic forms to the subsequent crop⁴². Furthermore, biodegradation of green manure remainders can produce bicarbonate (H_2CO_3) that can provide higher bio-available P for the plants by dissolving mineral P of the soil⁷⁸.

Vermicompost provided high nutrition by balancing the nutrients, due to the fact that it contains all macro and some micro-nutrients. In regard with our results, vermicompost treatments increased the K uptake of *Setaria* grass⁷⁵ and *Lilium* plant⁶². Moreover, vermicompost increased N percentage of spinach⁶⁹ and leaf P content of corn (Gutiérrez-Miceli *et al.* 2008). Also, the available form of N (nitrate) in vermicompost is more than traditional composted manure⁸⁴.

Water limitation reduced the leaf chlorophyll, and also carotenoid concentrations. One of the common symptoms of oxidative stress under drought conditions is the chlorophyll content reduction, which may be due to photo-oxidation and the pigment degradation^{2,66}. This study results indicated that the amount of carotenoids, chlorophyll b and a significantly decreased under water limitation. These result are not consistent with several studies reported that carotenoid increased under water deficit conditions^{6,9}.

Earlier researches^{9,46,47} approved our results that vermicompost application in soil brought about a remarkable growth in carotenoids content under water deficit condition. mycorrhizal inoculation resulted in the maintenance of a very high chlorophyll content relative to non-mycorrhizal plants in those plants subjected to late-season water limitation. This study

results are in agreement with those reported by (Metwally *et al.* 2019)⁵⁹. Using green manure can enhance the nitrogen availability, which can be used by linrain in the chlorophyll production. Chlorophyll plays a crucial role in photosynthesis, such that increased chlorophyll formation will improve photosynthetic reactions and finally raise plant growth and production. Also, Subaedah and Aladin (2016)⁸² clarified the enhancement of chlorophyll content by using of green manure in maize, which is in parallel with our findings.

Amassing proline under stress conditions is identified as one of the plant remedies for osmotic regulation improving in the leaves. Proline plays a leading role during stress in the plant. In addition to a great osmolyte, it acts as a metal chelator, a signalling molecule and an antioxidant molecule⁸¹. Having a higher capacity for osmotic regulation is one of those plants characteristic with higher drought tolerance. In this study, AMF-treated plants indicated an enhanced accumulation of proline in water-limited condition. These results are in agreement with Gheisari Zardak *et al.* (2018)²⁹ and Tyagi *et al.* (2017)⁸⁷ findings. Consequently, AM plants can be physiologically more tolerant against drought and indicate more osmotic regulation during exposing to stress conditions. Chinsamy *et al.* (2013)¹⁷ demonstrated a substantial elevation in proline concentration within tomato grainlings under salinity stress by the use of vermicompost. Also, The same effect was observed with vermicompost in pomegranate¹⁴ and rice grainlings (García *et al.* 2012)²⁸ under water limitation stress.

The linrain plants exposure to drought stress could increase the TSS concentration. Results of a study accomplished by Masoudi-Sadaghiani *et al.* (2011)⁵⁷ and Vijayalakshmi *et al.* (2012)⁹⁰ have indicated that water constraints can elevate the concentration of TSS in the leaves. Increasing the TSS concentrations occurred in those plants that were treated with one of Fm and Fv, along with their combination in both irrigation regimes. However, under limited irrigation condition, TSS increasing was remarkably higher in mycorrhizal treatments and the presence of green manure. AMF enhance TSS and electrolyte concentrations in host plants. Furthermore, carbohydrates increasing in mycorrhizal plants may be attributed to increased photosynthetic carbon and enzymes activation reduction. Other researchers have reported increased carbohydrate in AM plants^{3,94}. Furthermore, Tejada *et al.* (2008)⁸⁶ reported that soluble carbohydrate contents of maize plants were the highest in *Trifolium pratense* L. amended soils. The increment of leaf TSS of plants treated with vermicompost was also reported by Tejada and Benítez (2011) and Salehi *et al.* (2016)^{76,85}.

The main role of the GB could be considered as protecting the plasma membrane integrity from drought stress injury and contribution to osmotic regulation. Dual application of mycorrhiza and vermicompost increased the GB content under the water-limited conditions, and this indicating that the fertilizers could alleviate drought stress impacts. In addition, GB accumulation in cotton has been reported⁵⁴ as an effective compatible solution under drought stress. Moreover, Hashem *et al.* (2015)⁴¹ reported that enhanced GB accumulation leads to better growth and salt tolerance, and after that improvement in the performance of photosynthetic traits.

Lower accumulation of H₂O₂ provides proof that increasing the antioxidant enzymes activity bring about the removal of ROS and the protection of host plants against oxidative stress²⁶. A higher concentration of MDA in leaves, which can indicate the amount of peroxidation of membrane lipids, may be accompanied by higher H₂O₂ accumulation in drought stress plants³⁰. Additionally, fertilized plants (Fm and Fv) indicated less MDA than control that indicates the fertilizers involvements in ROS metabolism^{14,61,72}. In addition, mycorrhiza inoculation reduced the H₂O₂ and MDA concentration in foxtail millet leaves in drought³³. In consistent with our results, Bidabadi *et al.* (2017)¹⁴ demonstrated a reduction in H₂O₂ content in vermicompost application. Moreover, using green manure would resulted in MDA and H₂O₂ concentrations reduction, and also along with that, enhancement of the catalase and peroxidase activities in water limitation. It appeared that green manure could impede H₂O₂ accumulation and lipid peroxidation in plants by increasing the water content of the soil. Limited-irrigation causes oxidative stress by increasing the production of ROS in plant cells, and oxidative damage is also eased by antioxidant enzymes like APX, CAT, and SOD^{74,83}. Amongst anti-oxidants enzymes, SOD and CAT are recognized as the most frequently used detoxification enzymes, which play a major role in controlling excessive ROS, in collaboration with involved enzymes of the ascorbate–glutathione cycle^{10,15,38}. In this study, the triple application of

mycorrhiza, vermicompost and green manure increased the CAT activity in both irrigation conditions, consequently, they can facilitate quick sweeping of ROS especially H₂O₂, so that metabolic reactions are less affected. Earlier results confirmed that the mycorrhizal and vermicompost-applied plants indicated considerably higher enzyme activities in stress condition^{13,47,61,67}.

Applying the green manure in soils at high doses increased maize grain yield in comparison with the control soil⁵⁰. Similar results were obtained with the green manure application in wheat grain yield by Gruter *et al.* (2017)³⁷.

Vermicompost along with biofertilizers-treated plots indicated a substantial increase in essential oil content, essential oil yield and biomass of *Ocimum basilicum* in comparison with control⁸⁹. Similarly, Doan *et al.* (2015)¹⁹ and Goswami *et al.* (2017)³⁴ reported an increase in grain yield by using vermicompost. Mycorrhizal symbiosis enhanced the grain and oil yields in comparison with non-mycorrhizal plants under both irrigation conditions. The results demonstrated that mycorrhizal colonization was a momentous factor for lingrain development, especially under limited water condition with respect to playing a significant role in nutrient and water absorption in water-limited plants⁴.

Conclusions

These results demonstrated that water limitation could reduce the pigments contents, nutrient contents, LRWC, colonization percentage, and (grain and oil) yields. However, it is responsible for a significant increase in H₂O₂, MDA, osmolytes and antioxidant enzymes activities in this condition. With respect to these results, the inclusion of vermicompost increased the lingrain performance for most of the investigated traits in comparison with the control (Fc). Although, Fv had no advantage compared mycorrhizal fertilizers (Fm and Fm + v), but its presence along with green manure or mycorrhiza could develop a positive synergistic effect on these traits.

Moreover, adding green manure to vermicompost treatments (Gr × Fv) resulted in a significant increase of colonization percentage, oil yield in comparison with Gc × Fv. Also, Gr × Fv significantly increased the APX activity and grain yield under drought stress conditions. However, it caused no significant difference in non-stressed condition, in comparison with single vermicompost treatment (Gc × Fv). In other words, green manure increased the vermicompost efficacy under stress conditions. Also, adding the green manure could result in a remarkable grain yield superiority of Fm + v in comparison with Fm. In the green manure absence, these two fertilizer treatments effect were similar in full irrigation condition. Consequently, combining green manure with the dual fertilizer (Fm + v) led to the synergistic effects of vermicompost and mycorrhiza.

In drought stress conditions, adding the green manure improved mycorrhiza efficiency on elevating RWC, leaf nitrogen, chlorophyll a, chlorophyll b, grain yield, oil yield and antioxidant enzymes activity (catalase, ascorbate peroxidase, superoxide dismutase and Guaiacol peroxidase),

All fertilizer treatments (FV, Fm and Fm + v) imposed less hydrogen peroxide and malondialdehyde on lingrain in both irrigation conditions in comparison with control. The lowest amount of hydrogen peroxide and malondialdehyde was achieved from the application of dual fertilizer under normal irrigation conditions followed by mycorrhiza and vermicompost fertilizers, and also the highest amount was achieved from non-fertilized control and under the limited irrigation conditions .

The co-application of eco-friendly soil amendments would result in the profitable lingrain production under water limitation. Therefore, as a result the triple fertilizer (Gr × Fm + v) experienced the highest amount of LRWC, leaf nitrogen, chlorophyll a, chlorophyll b, carotenoids, grain yield and oil yield. In addition, GrFm + v enjoyed the maximum quantity of and also antioxidant enzymes activity under drought stress conditions. Consequently, it can be concluded that the combined usage of organic and biological fertilizers (green manure, vermicompost, and mycorrhiza) can be also applied as a solution in order to maintain the lingrain plant performance under the water-limited condition.

Materials And Methods

Experimental design

Field experiments were accomplished on 2017 and 2018 at the Saatlou Station (44° N, 45°10'95" E), Urmia Agricultural Research Centre in Iran. The average of rainfall was 17.5 mm and 32.07 mm during the linrain growing season in 2017 and 2018, respectively. This experiment was a 3³ factorial with respect to randomized complete block design with three replications. Factors included (1) irrigation regimes (full irrigation (Iw), limited irrigation (Is) (irrigation cut-off from flowering stage), (2) green manure (no green manure (Gc), *Trifolium pratense* as green manure (Gr) and (3) mycorrhiza inoculation ((*Rhizophagus irregularis*) (Fm) 300 g m⁻²), vermicompost (Fv) (5 t/ha), their combination (mycorrhiza + vermicompost (Fm + v)) and no fertilizer (Fc).

Soil amendments

The soil texture was salty loam with a pH of 7.8. Other soil physical and chemical properties are indicated in Table 1.

Red clover (*Trifolium pratense*) was grained on August 2016 and 2017. Also, fresh Gr was manually incorporated into 20 cm depth of surface soil in March 2017 and 2018, at the flowering stage.

Mycorrhizal inoculum included a blend of sterile sand, mycorrhizal hyphae and spores (30 spores g⁻¹ inoculum), and colonized fragments of canola root that were isolated from an AMF community of a canola farm by plant protection department of Agricultural and Natural Resources Research Centre, West Azerbaijan province, Urmia, Iran. Vermicompost was prepared with respect to the method suggested by Ayyobi *et al.* (2013)⁸. Chemical properties of the vermicompost that was used in these experiments are tabulated in Table 2.

Plant culture

By passing a month from green manure integration, the land was manually prepared. At first and second year, the measured dry biomass production by green manure was 4.72 t ha⁻¹ and 4.95 t ha⁻¹, respectively. Also, inoculum (300 g m⁻²) was incorporated into the soil of grainbed before the grain sowing. The exact amounts of vermicompost were manually spread onto the soil surface and incorporated into the top 20 cm of the soil. Linrain Grains were achieved from the Agricultural and Natural Resources Research Centre of West Azerbaijan province, Urmia, Iran. The grains were sown in 8 rows on 21 April 2017 and 24 April 2018, respectively (with two cm intra-row and 25 cm inter-row spaces). The full irrigation treatment included five irrigations (after sowing, at the 4-leaf stage, at stem elongation, at flowering and capsule formation stages), and limited irrigation treatment (irrigation cut-off from flowering stage) had three irrigations (after sowing, at 4-leaf and stem elongation stages). The irrigation water amount in each plot was measured by the use of a water meter. Moreover, the average humidity at the surface and depths of 20 and 40 cm of soil was determined using a soil moisture meter (EXTECH M0750) in each plot before each irrigation. After that, with respect to the lack of moisture under the field capacity point, the amount of water that was required for each irrigation was calculated by following equation:

$$V_n = (F_c - \theta) \times (A \times h),$$

Where, V_n , the volume of irrigation water (m³), F_c moisture content at the field capacity point (volume), θ soil moisture (volume), A area of the plot (m²) and h , Effective depth of linrain root (m).

In terms of the above equation, the amounts of water used for irrigation were 2346 and 1420 m³/ha (for the first year), and 1750 and 600 m³/ha (for the second year) for full and limited irrigation, respectively.

All the practices were manually handled in this experiment, in order to prevent interference with all materials.

Parameters measured

Leaf relative water content (LRWC):

Leaf relative water content was assayed in leaf samples with respect to García-Mata and Lamattina (2001)²⁷.

AMF Colonization

The mycorrhizal root colonization percentage was determined for each plot in five roots. In order to achieve this goal, roots were cleared by 10% KOH at 90 °C for 10 min, and after that were stained in 0.05% lactic acid-glycerol-Trypan Blue⁷⁰. The mycorrhizal colonization was estimated with respect to Giovannetti and Mosse (1980)³¹ gridline intersect method.

Nutrient content

The leaf samples were dried in an oven for 72 h at 70 °C, and after that, were grounded using an electric mill. The Kjeldahl method was used for Nitrogen measurement⁶⁵. Spectrophotometry and flame photometry methods were applied in order to determine P and K contents, respectively.

Photosynthetic pigment content assay

Approximately 10 ml of acetone 80% was added to extracts of fresh leaf tissue samples gradually (0.2 g). After that, it was centrifuged for 10 min at 400 rpm and the absorbance was recorded using a spectrophotometer at 645, 663, and 470 nm. Chlorophyll and carotenoids were obtained based on the following equations:

$$\text{chlorophyll a} = (19.3 \times A_{633} - 0.86 \times A_{645}) \times V/100W$$

$$\text{chlorophyll b} = (19.3 \times A_{645} - 3.6 \times A_{663}) \times V/100W$$

$$\text{carotenoid} = (1000 \times A_{470} - 1.82 \times \text{Chlorophyll a} - 85.02 \times \text{chlorophyll b})$$

Proline

Leaf fresh tissue (1 g) was used for leaf proline extracting in sulfosalicylic acid 5% (w/v). Leaf proline was determined using spectrophotometric analysis at 520 nm of the ninhydrin reaction, with respect to (Bates *et al.* 1973)¹¹.

Total soluble sugars (TSS)

Leaf total soluble sugar was measured by the anthrone method⁵⁶. Also, the TSS concentration was determined using the glucose standard curve.

Hydrogen peroxide (H₂O₂)

The fresh leaf tissue (0.5 g) was grounded in a pre-chilled mortar with 5 mL of 0.1% (w/v) TCA. After that, this homogenate was centrifuged at 12000 × g for 15 min at 4 °C. also, 0.5 ml of the supernatant was added to 0.5 ml of 10 mM potassium phosphate buffer (pH 7.0) and 1 ml of 1 M KI and the OD of the suspension was read at 390 nm⁸⁸.

Malondialdehyde (MDA)

Malondialdehyde was measured with respect to the thiobarbituric acid (TBA) reaction, as it was explained by Heath and Packer (1968)⁴⁴. Fresh leaf tissues (0.1 g) were frozen in liquid nitrogen, and homogenized in 5 ml of 0.1% trichloroacetic acid (TCA). The homogenates were transferred into tubes and centrifuged at 3600 rpm for 10 min at room temperature. Two millilitres of this extract was added to 5 ml 0.5% TBA. The mixture was incubated for 30 minutes at 95 ° C, and then was immediately placed on ice for 5 minutes and centrifuged again at 10000 × g for 10 minutes, respectively. The supernatant absorbance was determined at 532 and 600, nm (A₅₃₂ and A₆₀₀). The MDA concentration was calculated due to following equation:

$$[(A_{532} - A_{600})/155] \times 100$$

Glycine betaine (GB)

Glycine betaine analysis was performed with respect to the method of Grieve and Grattan (1983)³⁵.

Catalase (CAT), Ascorbate peroxidase (APX) and Superoxide dismutase (SOD)

The fresh leaf tissues (5.0 g) were macerated in 10 mL potassium phosphate buffer [pH 7.8; 50 mM], by the use of an ice-cooled sterilized pestle and mortar. After that, the homogenates were centrifuged for 20 min at 4 °C at 12000 g. The supernatant was applied to the antioxidant enzymes activity assay.

Catalase activity was assayed in regard with Maehly and Chance (1954)⁵⁵. 50 ml enzyme extract was added to 2.5 ml of 50 mM phosphate buffer (pH 7.4), and 0.1 ml of 1% hydrogen peroxide in the ice bath, in order to determine CAT activity. The H₂O₂ content reduction was obtained at 240 nm for 1 min.

Ascorbate peroxidase activity was determined by absorption reducing at 290 nm due to ascorbate oxidation in 3 min, with respect to Chen and Asada (1989)¹⁶. The reaction mixture involved 50 mM potassium phosphate buffer (pH 7.0), 0.1 mM EDTA, 0.5 mM ascorbate, 1.54 mM hydrogen peroxide, and 50 ml enzyme extract. Superoxide dismutase activity was measured regarding Dhindsa *et al.* (1981)¹⁸ method in terms of its ability to inhibit the nitro blue tetrazolium reduction by superoxide radicals generated by photochemical reactions. The 3 ml reaction mixture contained 50 mM phosphate buffer, pH 7–8, 13 mM methionine, 75 µM NBT, 2 µM riboflavin, 0–1 mM EDTA, and 0–50 µl enzyme extract. NBT solution, riboflavin, EDTA and 1.0 ml DDW were incubated in light under 15 W fluorescent lamps for 15 min. The blank reaction was processed also for 15 min in darkness. The samples and controls absorbance was measured at 560 nm. Guaiacol peroxidase was measured regarding²⁵.

Grain Oil Extraction

Soxhlet technique was utilized in order to extract grain oil⁷¹. Oil yield was calculated by the following formula:

Oil yield = % oil X seed yield

Grain yield

At plant maturity, above-ground biomass was harvested from 1 m² of each experimental plot by ignoring marginal effects. Each plot Grains were separated and weighed in order to measure grain yield.

Statistical analysis

Combined factorial analysis of variance (three factors) including those gathered data for about two years (with one year as a random factor) was performed using SAS software and by applying the general linear model (GLM) procedure to establish these three factors main and interactive influences. After that, this model was approved by checking the residues normality and homogeneity (using the Bartlett test). Also, No data transformation was performed. The mean values were compared using the protected least significant difference (PLSD) at $P \leq 0.05$, due to the reason that the treatment factors were qualitative.

Declarations

Conflicts of Interest

The authors declare no conflicts of interest.

References

1. Abd_Allah, E.F., Hashem, A., Alqarawi, A.A., Bahkali, A.H., & Alwhibi, M.S. Enhancing growth performance and systemic acquired resistance of medicinal plant *Sesbania sesban* (L.) Merr using arbuscular mycorrhizal fungi under salt stress. *Saudi. J. Biol. Sci.* **22**, 274–283. (2015).
2. Abdelmoneim, T., Moussa, T.A., Almaghrabi, O., Alzahrani, H.S., & Abdelbagi, I. Increasing plant tolerance to drought stress by inoculation with arbuscular mycorrhizal fungi. *Life. Sci. J.* **11**, 10–17. (2014).
3. Amiri, R., Nikbakht, A., Rahimmalek, M., & Hosseini, H. Variation in the essential oil composition, antioxidant capacity, and physiological characteristics of *Pelargonium graveolens* L. inoculated with two species of mycorrhizal fungi under water deficit conditions. *J. Plant. Grow. Reg.* **36**, 502–515. (2017).
4. Ansari, A., Razmjoo, J., & Karimmojeni, H. Mycorrhizal colonization and seed treatment with salicylic acid to improve physiological traits and tolerance of flaxseed (*Linum usitatissimum* L.) plants grown under drought stress. *Acta. Physiol. Plantarum.* **38**, 34. (2016).
5. Arndt, S., Clifford, S., Wanek, W., Jones, H., & Popp, M. Physiological and morphological adaptations of the fruit tree *Ziziphus rotundifolia* in response to progressive drought stress. *Tree. Physiol.* **21**, 705–715. (2001).
6. Ashraf, M., & Harris, P. Photosynthesis under stressful environments: an overview. *Photosynthetica.* **51**, 163–190. (2013).
7. Ayyobi, H., & Peyvast, G.A. The effects of cow manure vermicompost and municipal solid waste compost on peppermint (*Mentha piperita* L.) in Torbat-e-Jam and Rasht regions of Iran. *Int. J. Rec. Org. Was. Agric.* **3**, 147–153. (2014)
8. Ayyobi, H., Peyvast, G.A., & Olfati, J.A. Effect of vermicompost and vermicompost extract on oil yield and quality of peppermint (*Mentha piperita* L.). *J. Agric. Sci.* **58**, 51–60. (2013)
9. Baghbani-Arani, A., Modarres-Sanavy, S.A.M., Mashhadi-Akbar-Boojar, M., & Mokhtassi-Bidgoli, A. Towards improving the agronomic performance, chlorophyll fluorescence parameters and pigments in fenugreek using zeolite and vermicompost under deficit water stress. *Ind. Cro. Pro.* **109**, 346–357. (2017)
10. Bai, T., Li, C., Ma, F., Feng, F., & Shu, H. Responses of growth and antioxidant system to root-zone hypoxia stress in two *Malus* species. *Plant. Soil.* **327**, 95–105. (2010)
11. Bates, L.S., Waldren, R.P., & Teare, I. Rapid determination of free proline for water-stress studies. *Plant. Soil* **39**, 205–207. (1973)
12. Beykhhormizi, A., Abrishamchi, P., Ganjeali, A., & Parsa, M. Effect of vermicompost on some morphological, physiological and biochemical traits of bean (*Phaseolus vulgaris* L.) under salinity stress. *J. Plant. Nut.* **39**, 883–893. (2016)
13. Bidabadi, S.S., Afazel, M., Poodeh, S.D. The effect of vermicompost leachate on morphological, physiological and biochemical indices of *Stevia rebaudiana* Bertoni in a soilless culture system. *Int. J. Rec. Org. Was. Agric.* **5**, 251–262. (2016)
14. Bidabadi, S.S., Dehghanipoodeh, S., & Wright, G.C. Vermicompost leachate reduces some negative effects of salt stress in pomegranate. *Int. J. Rec. Org. Was. Agric.* **6**, 255–263. (2017)
15. Boaretto, L.F., Carvalho, G., Borgo, L., Creste, S., Landell, M.G., Mazzafera, P., & Azevedo, R.A. Water stress reveals differential antioxidant responses of tolerant and non-tolerant sugarcane genotypes. *Plant. Physiol. Biochem.* **74**, 165–175. (2014)
16. Chen, G.X., & Asada, K. Ascorbate peroxidase in tea leaves: occurrence of two isozymes and the differences in their enzymatic and molecular properties. *Plant. Cell. Physiol.* **30**, 987–998. (1989).
17. Chinsamy, M., Kulkarni, M.G., & Van Staden, J. Garden-waste-vermicompost leachate alleviates salinity stress in tomato seedlings by mobilizing salt tolerance mechanisms. *Plant. Gro. Reg.* **71**, 41–47. (2013)
18. Dhindsa, R.S., PLUMB-DHINDSA, P., & THORPE, T.A. Leaf Senescence: Correlated with Increased Levels of Membrane Permeability and Lipid Peroxidation, and Decreased Levels of Superoxide Dismutase and Catalase. *J. Experi. Bot.* **32**,

- 93–101. (1981).
19. Doan, T.T., Henry-des-Tureaux, T., Rumpel, C., Janeau, J.L., & Jouquet, P. Impact of compost, vermicompost and biochar on soil fertility, maize yield and soil erosion in Northern Vietnam: a three year mesocosm experiment. *Sci. Total. Env.* **514**, 147–154. (2015).
20. Dos Santos Marques, C.T., Gama, E.V.S., da Silva, F., Teles, S., Caiafa, A.N., & Lucchese, A.M. Improvement of biomass and essential oil production of *Lippia alba* (Mill) NE Brown with green manures in succession. *Ind. Crops. Pro.* **112**, 113–118. (2018).
21. Fageria, N.K. Green Manuring in Crop Production. *J. Plant. Nut.* **30**, 691–719. (2007).
22. FAO. 'FAOSTAT.' Available at <http://www.fao.org/faostat/en/#data/QC> [Accessed 26 Jun]. (2014).
23. Farooq, M., Gogoi, N., Barthakur, S., Baroowa, B., Bharadwaj, N., Alghamdi, S.S., & Siddique, K. Drought stress in grain legumes during reproduction and grain filling. *J. Agr. Crop.Sci.* **203**, 81–102. (2017).
24. Fathi, H., Imani, A., Amiri, M.E., Hajilou, J., & Nikbakht, J. Response of Almond Genotypes/Cultivars Grafted on GN15 'Garnem' Rootstock in Deficit-Irrigation Stress Conditions. *J. Nut.* **8**, 123–135. (2017).
25. Fielding, J., & Hall, J. A biochemical and cytochemical study of peroxidase activity in roots of *Pisum sativum*: I. a comparison of DAB-peroxidase and guaiacol-peroxidase with particular emphasis on the properties of cell wall activity. *J. Exp. Bot.* **29**, 969–981. (1978).
26. Fouad, M.O., Essahibi, A., Benhiba, L., & Qaddoury, A. Effectiveness of arbuscular mycorrhizal fungi in the protection of olive plants against oxidative stress induced by drought. *Spa. J. Agric. Res.* **12**, 763–771. (2014).
27. García-Mata, C., & Lamattina, L. Nitric oxide induces stomatal closure and enhances the adaptive plant responses against drought stress. *Plant. Physio.* **126**, 1196–1204. (2001).
28. García, A.C., Berbara, R.L.L., Farías, L.P., Izquierdo, F.G., Hernández, O.L., Campos, R.H., & Castro, R.N. Humic acids of vermicompost as an ecological pathway to increase resistance of rice seedlings to water stress. *Afr. J. Biotech.* **11**, 3125–3134. (2012).
29. Gheisari Zardak, S., Movahhedi Dehnavi, M., Salehi, A., Gholamhoseini, M. Effects of using arbuscular mycorrhizal fungi to alleviate drought stress on the physiological traits and essential oil yield of fennel. *Rhiz.* **6**, 31–38. (2018).
30. Gill, S.S., & Tuteja, N. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant. Physio. Biochem.* **48**, 909–930. (2010).
31. Giovannetti, M., & Mosse, B. An evaluation of techniques for measuring vesicular arbuscular mycorrhizal infection in roots. *New. phytolo.* **84**, 489–500. (1980).
32. Gong, M., Tang, M., Chen, H., Zhang, Q., & Feng, X. Effects of two *Glomus* species on the growth and physiological performance of *Sophora davidii* seedlings under water stress. *New. For.* **44**, 399–408. (2013).
33. Gong, M., You, X., & Zhang, Q. Effects of *Glomus* intraradices on the growth and reactive oxygen metabolism of foxtail millet under drought. *Ann.Micro.* **65**, 595–602. (2015).
34. Goswami, L., Nath, A., Sutradhar, S., Bhattacharya, S.S., Kalamdhad, A., Vellingiri, K., & Kim, K.H. Application of drum compost and vermicompost to improve soil health, growth, and yield parameters for tomato and cabbage plants. *J. Env. Manag.* **200**, 243–252. (2017).
35. Grieve, C., & Grattan, S. Rapid assay for determination of water soluble quaternary ammonium compounds. *Plant. Soil.* **70**, 303–307. (1983).
36. Grümberg, B.C., Urcelay, C., Shroeder, M.A., Vargas-Gil, S., & Luna, C.M. The role of inoculum identity in drought stress mitigation by arbuscular mycorrhizal fungi in soybean. *Biol. Fer. Soils.* **51**, 1–10. (2015).
37. Gruter, R., Costerousse, B., Bertoni, A., Mayer, J., Thonar, C., Frossard, E., Schulin, R., & Tandy, S. Green manure and long-term fertilization effects on soil zinc and cadmium availability and uptake by wheat (*Triticum aestivum* L.) at different growth stages. *Sci. Total. Env.* **599–600**, 1330–1343. (2017).

38. Guo, W., Chen, R., Gong, Z., Yin, Y., Ahmed, S., & He, Y. Exogenous abscisic acid increases antioxidant enzymes and related gene expression in pepper (*Capsicum annuum*) leaves subjected to chilling stress. *Gen. Molec. Res.* **11**, 4063–4080. (2012).
39. Gutierrez-Miceli, F.A., Moguel-Zamudio, B., Abud-Archila, M., Gutierrez-Oliva, V.F., & Dendooven, L. Sheep manure vermicompost supplemented with a native diazotrophic bacteria and mycorrhizas for maize cultivation. *Bior. Tech.* **99**, 7020–6. (2008).
40. Hasanuzzaman, M., Anee, T.I., Bhuiyan, T.F., Nahar, K., & Fujita, M. Emerging role of osmolytes in enhancing abiotic stress tolerance in rice. In 'Advances in Rice Research for Abiotic Stress Tolerance. Elsevier. pp. 677–708.(2019).
41. Hashem, A., Abd_Allah, E.F., Alqarawi, A.A., Aldubise, A., & Egamberdieva, D. Arbuscular mycorrhizal fungi enhances salinity tolerance of *Panicum turgidum* Forssk by altering photosynthetic and antioxidant pathways. *J. Plant. Int.* **10**, 230–242. (2015).
42. Havlin, J.L., Tisdale, S.L., Nelson, W.L., & Beaton, J.D. Soil fertility and fertilizers. *Pearson. Edu. Ind.* (2016).
43. He, M., & Dijkstra, F.A. Drought effect on plant nitrogen and phosphorus: a meta-analysis. *New. Phytol.* **204**, 924–31. (2014).
44. Heath, R.L., & Packer, L. Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. *Arch. Biochem. Bbioph.* **125**, 189–198. (1968).
45. Herridge, D.F., Peoples, M.B., & Boddey, R.M. Global inputs of biological nitrogen fixation in agricultural systems. *Plant. Soil. Biol.* **311**, 1–18. (2008).
46. Hosseinzadeh, S.R., & Ahmadpour, R. Evaluation of vermicompost fertilizer application on growth, nutrient uptake and photosynthetic pigments of lentil (*Lens culinaris* Medik.) under moisture deficiency conditions. *J. Plant. Nut.* **41**, 1276–1284. (2018).
47. Hosseinzadeh, S.R., Amiri, H., & Ismaili, A. Nutrition and biochemical responses of chickpea (*Cicer arietinum* L.) to vermicompost fertilizer and water deficit stress. *J. Plant. Nut.* **40**, 2259–2268. (2017).
48. Huis, R., Hawkins, S., & Neutelings, G. Selection of reference genes for quantitative gene expression normalization in flax (*Linum usitatissimum* L.). *BMC. Plant. Biol.* **10**, 71. (2010).
49. Jan, B., Ali, A., Wahid, F., Shah, S.N.M., Khan, A., & Khan, F. Effect of arbuscular mycorrhiza fungal inoculation with compost on yield and phosphorous uptake of berseem in alkaline calcareous soil. *Amer. J. Plant. Sci.* **5**, 1359. (2014).
50. Kanatas, P., Travlos, I., Kakabouki, I., Papastylianou, P., & Gazoulis, I. Yield of organically grown maize hybrids as affected by two green manure crops in Greece. *Chil.J. Agric. Res.* **80**, 334–341. (2020).
51. Kara, Z., Arslan, D., Güler, M., & Güler, Ş. Inoculation of arbuscular mycorrhizal fungi and application of micronized calcite to olive plant: Effects on some biochemical constituents of olive fruit and oil. *Sci. Hortic.* **185**, 219–227. (2015).
52. Kong, J., Pei, Z., Du, M., Sun, G., & Zhang, X. Effects of arbuscular mycorrhizal fungi on the drought resistance of the mining area repair plant Sainfoin. *Int. J. Min. Sci. Tech.* **24**, 485–489. (2014).
53. Lehmann, A., Veresoglou, S.D., Leifheit, E..F, & Rillig, M.C. Arbuscular mycorrhizal influence on zinc nutrition in crop plants—a meta-analysis. *Soil. Biol. Biochem.* **69**, 123–131. (2014).
54. Lv, S., Yang, A., Zhang, K., Wang, L., & Zhang, J. Increase of glycinebetaine synthesis improves drought tolerance in cotton. *Molec. Breed.* **20**, 233–248. (2007).
55. Maehly, A., & Chance, B. Methods of biochemical analysis. by Glick D., *Interscience, New York* 454. (1954).
56. Mahadevan, A., & Sridhar, R. Enzymes of infected plants and parasites. *Methods in Physiological Plant Pathology. Sivakami. Publi. Mad. Ind.* 79–80. (1986).
57. Masoudi-Sadaghiani, F., Babak, A.M., Zardoshti, M.R., Hassan, R.S.M., & Tavakoli, A. Response of proline, soluble sugars, photosynthetic pigments and antioxidant enzymes in potato (*Solanum tuberosum* L.) to different irrigation regimes in greenhouse condition. *Aus. J. Crop. Sci.* **5**, 55. (2011).

58. McKenzie, R.R., & Deyholos, M.K. Effects of plant growth regulator treatments on stem vascular tissue development in linseed (*Linum usitatissimum* L.). *Ind. Crops. Pro.* **34**, 1119–1127. (2011).
59. Metwally, A., Azooz, M., Nafady, N., & El-Enany, A. Arbuscular mycorrhizal symbiosis alleviates drought stress imposed on wheat plants (*Triticum Aestivum* L.). *Appl. Eco. Env. R Res.* **17**, 13713–13727. (2019).
60. Mir, M., Hassan, G., Mir, A., Hassan, A., & Sulaimani, M. Effects of bio-organics and chemical fertilizers on nutrient availability and biological properties of pomegranate orchard soil. *Afr. J. Agric. Res.* **8**, 4623–4627. (2013)
61. Mo, Y., Wang, Y., Yang, R., Zheng, J., Liu, C., Li, H., Ma, J., Zhang, Y., Wei, C., & Zhang, X. Regulation of Plant Growth, Photosynthesis, Antioxidation and Osmosis by an Arbuscular Mycorrhizal Fungus in Watermelon Seedlings under Well-Watered and Drought Conditions. *Front. Plant. Sci.* **7**, 644. (2016).
62. Moghadam, A.R.L., Ardebili, Z.O., & Saidi, F. Vermicompost induced changes in growth and development of *Lilium Asiatic* hybrid var. Navona. *Afr. J. Agric. Res.* **7**, 2609–2621. (2012).
63. Mollavali, M., Perner, H., Rohn, S., Riehle, P., Hanschen, F.S., & Schwarz, D. Nitrogen form and mycorrhizal inoculation amount and timing affect flavonol biosynthesis in onion (*Allium cepa* L.). *Myco.* **28**, 59–70. (2018).
64. Munibah Afzal, G.S., Ilyas, M., Jan, S.S.A., & Jan, S.A. 2. Impact of climate change on crop adaptation: current challenges and future perspectives. *Pure. Appl. Biol. (PAB)* **7**, 965–972. (2018).
65. Nelson, D., & Sommers, L. Determination of Total Nitrogen in Plant Material 1. *Agr. J.* **65**, 109–112. (1973)
66. Oraki, H., & Aghaalikhana, M. Effect of water deficit stress on proline contents, soluble sugars, chlorophyll and grain yield of sunflower (*Helianthus annuus* L.) hybrids. *Afr. J. Biotech.* **11**, 164–168. (2012).
67. Pedranzani, H., Rodriguez-Rivera, M., Gutierrez, M., Porcel, R., Hause, B., & Ruiz-Lozano, J.M. Arbuscular mycorrhizal symbiosis regulates physiology and performance of *Digitaria eriantha* plants subjected to abiotic stresses by modulating antioxidant and jasmonate levels. *Myco.* **26**, 141–52. (2016).
68. Peoples, M., Brockwell, J., Herridge, D., Rochester, I., Alves, B., Urquiaga, S., Boddey, R., Dakora, F., Bhattarai, S., Maskey, S. The contributions of nitrogen-fixing crop legumes to the productivity of agricultural systems. *Sym.* **48**, 1–17. (2009).
69. Peyvast, G., Olfati, J., Madeni, S., & Forghani, A. Effect of vermicompost on the growth and yield of spinach (*Spinacia oleracea* L.). *J. Food. Agricu. Env.* **6**, 110. (2008).
70. Phillips, J.M., & Hayman, D. Improved procedures for clearing roots and staining parasitic and vesicular-arbuscular mycorrhizal fungi for rapid assessment of infection. *Tran. Bri. Mycolo. Soc.* **55**, 158–161. (1970).
71. Pomeranz, Y. 'Food analysis: theory and practice. *Spr. Sci. Busi. Med.* (2013).
72. Quiroga, G., Erice, G., Aroca, R., Chaumont, F., & Ruiz-Lozano, J.M. Enhanced drought stress tolerance by the arbuscular mycorrhizal symbiosis in a drought-sensitive maize cultivar is related to a broader and differential regulation of host plant aquaporins than in a drought-tolerant cultivar. *Fro. Plan.t Sci.* **8**, 1056. (2017).
73. Roper, M.M., Milroy, S.P., & Poole, M.L. Green and brown manures in dryland wheat production systems in mediterranean-type environments. *Adv. Agron.* **Vol. 117 pp.** 275–313. (2012).
74. Ruiz-Sánchez, M., Aroca, R., Muñoz, Y., Polón, R., & Ruiz-Lozano, J.M. The arbuscular mycorrhizal symbiosis enhances the photosynthetic efficiency and the antioxidative response of rice plants subjected to drought stress. *J. Plant. Physiol.* **167**, 862–869. (2010).
75. Sabrina, D.T., Hanafi, M.M., Gandahi, A.W., Muda Mohamed, M.T., & Abdul Aziz, N.A. Effect of mixed organic-inorganic fertilizer on growth and phosphorus uptake of setaria grass (*Setaria splendida*). *Aus. J. Cro. Sci.* **7**, 75. (2013).
76. Salehi, A., Tasdighi, H., & Gholamhoseini, M. Evaluation of proline, chlorophyll, soluble sugar content and uptake of nutrients in the German chamomile (*Matricaria chamomilla* L.) under drought stress and organic fertilizer treatments. *Asi. Pac.J. Tro. Biomed.* **6**, 886–891. (2016).
77. Sawers, R.J., Gutjahr, C., & Paszkowski, U. Cereal mycorrhiza: an ancient symbiosis in modern agriculture. *Trends. Plant. Sci.* **13**, 93–7. (2008).

78. Sharpley, A., & Smith, S. Mineralization and leaching of phosphorus from soil incubated with surface-applied and incorporated crop residue. *J. Env. Qua.* **18**, 101–105. (1989).
79. Shukla, A., Kumar, A., Jha, A., Salunkhe, O., & Vyas, D. Soil moisture levels affect mycorrhization during early stages of development of agroforestry plants. *Biol. Fer. Soils.* **49**, 545–554. (2013).
80. Sivakumar, N. Effect of edaphic factors and seasonal variation on spore density and root colonization of arbuscular mycorrhizal fungi in sugarcane fields. *Ann. Micro.* **63**, 151–160. (2013).
81. Song, H., Li, Y., Zhou, L., Xu, Z., & Zhou, G. Maize leaf functional responses to drought episode and rewatering. *Agric. For. Meteorol.* **249**, 57–70. (2018).
82. Subaedah, S., & Aladin, A. Fertilization of Nitrogen, Phosphor and Application of Green Manure of *Crotalaria Juncea* in Increasing Yield of Maize in Marginal Dry Land. *Agric. Agric. Sci. Pro.* **9**, 20–25. (2016).
83. Talbi, S., Romero-Puertas, M.C., Hernández, A., Terrón, L., Ferchichi, A., & Sandalio, L.M. Drought tolerance in a Saharian plant *Oudneya africana*: role of antioxidant defences. *Env. Exp. Bot.* **111**, 114–126. (2015).
84. Taleshi, K., Shokoh-Far, A., Rafiee, M., Noormahamadi, G., & Sakinejhad, T. Effect of vermicompost and nitrogen levels on yield and yield component of safflower (*Carthamus tinctorius* L.) under late season drought stress. *Int. J. Agr. Plant. Pro.* **2**, 15–22. (2011).
85. Tejada, M., & Benítez, C. Organic amendment based on vermicompost and compost: differences on soil properties and maize yield. *Was. Man. Res.* **29**, 1185–1196. (2011).
86. Tejada, M., Gonzalez, J., García-Martínez, A., & Parrado, J. Effects of different green manures on soil biological properties and maize yield. *Biore. Tech.* **99**, 1758–1767. (2008).
87. Tyagi, J., Varma, A., & Pudake, R.N. Evaluation of comparative effects of arbuscular mycorrhiza (*Rhizophagus intraradices*) and endophyte (*Piriformospora indica*) association with finger millet (*Eleusine coracana*) under drought stress. *Eur.J. Soil. Biol.* **81**, 1–10. (2017).
88. Velikova, V., Yordanov, I., & Edreva, A. Oxidative stress and some antioxidant systems in acid rain-treated bean plants: protective role of exogenous polyamines. *Plant. Sci.* **151**, 59–66. (2000).
89. Verma, S.K., Pankaj, U., Khan, K., Singh, R., & Verma, R.K. Bioinoculants and Vermicompost Improve *Ocimum basilicum* Yield and Soil Health in a Sustainable Production System. *Clean. Soil. Air. Wat.* **44**, 686–693. (2016).
90. Vijayalakshmi, T., Varalaxmi, Y., Jainender, S., Yadav, S., Vanaja, M., Jyothilakshmi, N., & Maheswari, M. Physiological and biochemical basis of water-deficit stress tolerance in pearl millet hybrid and parents. *Ame.J. Plant. Sci.* **3**, 1730. (2012).
91. Wu, H.H., Zou, Y.N., Rahman, M.M., Ni, Q.D., & Wu, Q.S. Mycorrhizas alter sucrose and proline metabolism in trifoliolate orange exposed to drought stress. *Sci.Rep.* **7**, 42389. (2017).
92. Wu, Q-S, Srivastava, AK, Zou, Y-N (2013) AMF-induced tolerance to drought stress in citrus: a review. *Scientia Horticulturae* **164**, 77–87.
93. Yin, N., Zhang, Z., Wang, L., & Qian, K. Variations in organic carbon, aggregation, and enzyme activities of gangue-fly ash-reconstructed soils with sludge and arbuscular mycorrhizal fungi during 6-year reclamation. *Env. Sci.Poll. Res.* **23**, 17840–17849. (2016)
94. Yooyongwech, S., Phaukinsang, N., Cha-um, S., & Supaibulwatana, K. Arbuscular mycorrhiza improved growth performance in *Macadamia tetraphylla* L. grown under water deficit stress involves soluble sugar and proline accumulation. *Plan.t Gro. Reg.* **69**, 285–293. (2013).
95. Zhang, H.H., Xu, N., Teng, Z.Y., Wang, J.R., Ma, S., Wu, X, Li, X., & Sun, G.Y. 2-Cys Prx plays a critical role in scavenging H₂O₂ and protecting photosynthetic function in leaves of tobacco seedlings under drought stress. *J.Plant. Int.* **14**, 119–128. (2019).
96. Zhang, Z., Wang, H., Zhu, J., Suneethi, S., & Zheng, J. Swine manure vermicomposting via housefly larvae (*Musca domestica*): the dynamics of biochemical and microbial features. *Bior. Tec.* **118**, 563–71. (2012).

97. Zhao, R., Guo, W., Bi, N., Guo, J., Wang, L., Zhao, J., & Zhang, J. Arbuscular mycorrhizal fungi affect the growth, nutrient uptake and water status of maize (*Zea mays* L.) grown in two types of coal mine spoils under drought stress. *App. Soil. Eco.* **88**, 41–49. (2015).

Tables

Table 1. Soil physicochemical properties

Characteristic	Saturation extract	pH	EC	P	K	N	Organic carbon	Sand	Silt	Clay	Texture
			(dS/m)	(mg/kg)	(mg/k)	(%)	(%)	(%)	(%)	(%)	
0-30 cm	42	7.8	0.58	7.1	184	0.09	0.91	23	42	25	Silty loam

Table 2. Vermicompost properties

Properties	pH	EC (dS/m)	Organic matter (%)	Nitrogen (%)	P (%)	K (%)	Mg (%)	Ca (mg/kg)	Cu (mg/kg)	Zn (mg/kg)
Vermicompost	7.64	5.22	12.3	1.3	0.88	0.67	1.1	8.75	3.14	46.76

Table 3. Comparison between 2-year mean values of physiological traits of lingrain affected by green manure × mycorrhiza × vermicompost under full and limited irrigation

Where I_w (full irrigation), I_s (limited irrigation), G_c (no green manure), G_r (green manure), F_c (no fertilizer), F_m (mycorrhiza inoculation), F_v (vermicompost application) and F_{m+v} (combination of mycorrhiza and vermicompost). Means followed by the same letter in each column are not significantly different

Treatment	Leaf Relative water content (%)	Colonization (%)	Leaf N % on dry matter	Leaf k % on dry matter	Chlorophyll a (mg/g FW)	Chlorophyll b (mg/g FW)	Carotenoids (mg/g FW)
$I_W \times G_c \times F_c$	66.64 ^f	7.50 ^{ij}	2.42 ^{fg}	0.18 ^h	2.19 ^h	1.68 ^c	1.87 ^h
$I_W \times G_c \times F_m$	79.66 ^c	56.25 ^c	3.04 ^c	0.25 ^{cd}	3.35 ^d	2.08 ^b	2.9 ^d
$I_W \times G_c \times F_v$	75.04 ^d	13.03 ^g	2.77 ^{de}	0.25 ^{cd}	2.71 ^f	1.96 ^b	2.67 ^e
$I_W \times G_c \times F_{m+v}$	83.71 ^b	58.25 ^c	3.22 ^b	0.28 ^{ab}	4.42 ^b	2.11 ^b	3.83 ^b
$I_S \times G_c \times F_c$	51.35 ^h	4.94 ^k	1.89 ^j	0.9 ⁱ	1.19 ^m	1.06 ^g	1.02 ^l
$I_S \times G_c \times F_m$	63.16 ^g	42.55 ^e	2.3 ^g	0.21 ^{ef}	1.66 ^j	1.54 ^{cd}	1.63 ⁱ
$I_S \times G_c \times F_v$	62.73 ^g	9.40 ^{hi}	2.41 ^{fg}	0.22 ^{ef}	1.34 ^{kl}	1.31 ^{ef}	1.24 ^k
$I_S \times G_c \times F_{m+v}$	65.15 ^{fg}	52.91 ^c	2.41 ^{fg}	0.22 ^{ef}	1.83 ⁱ	1.6 ^{cd}	1.85 ^h
$I_W \times G_r \times F_c$	73.69 ^d	8.29 ^{ij}	2.49 ^f	0.20 ^{fg}	2.41 ^g	1.94 ^b	2.16 ^g
$I_W \times G_r \times F_m$	81.98 ^{bc}	67.40 ^b	3.27 ^b	0.27 ^{abc}	3.68 ^c	2.1 ^b	3.17 ^c
$I_W \times G_r \times F_v$	75.56 ^d	18.39 ^f	2.88 ^b	0.26 ^{bc}	2.98 ^d	1.96 ^b	2.7 ^e
$I_W \times G_r \times F_{m+v}$	92.56 ^a	71.08 ^a	3.38 ^a	0.29 ^a	4.86 ^a	2.44 ^a	4.37 ^a
$I_S \times G_r \times F_c$	53.74 ^h	5.92 ^{jk}	2.17 ^h	0.18 ^h	1.23 ^{ml}	1.22 ^{fg}	1.13 ^{kl}
$I_S \times G_r \times F_m$	71.08 ^e	44.04 ^e	2.79 ^f	0.23 ^{de}	2.46 ^g	1.96 ^b	2.29 ^g
$I_S \times G_r \times F_v$	62.96 ^g	11.57 ^{gh}	2.41 ^e	0.23 ^{de}	1.43 ^k	1.46 ^{de}	1.44 ^j
$I_S \times G_r \times F_{m+v}$	72.95 ^{de}	56.58 ^c	2.90 ^{de}	0.23 ^{de}	2.71 ^f	1.98 ^b	2.46 ^f

Table.4 Comparison between 2-year mean values of physiological traits of lingrain affected by green manure \times mycorrhiza \times vermicompost under full and limited irrigation

Where I_W (full irrigation), I_S (limited irrigation), G_c (no green manure), G_r (green manure), F_c (no fertilizer), F_m (mycorrhiza inoculation), F_v (vermicompost application) and F_{m+v} (combination of mycorrhiza and vermicompost). Means followed by the same letter in each column are not significantly different

Treatment	Glycine betaine ($\mu\text{mol/g}$ FW)	Hydrogen peroxide ($\mu\text{mol/g}$ FW)	Malondialdehyde ($\mu\text{mol/g}$ FW)	Ascorbate Peroxidase activity ($\text{mmol min}^{-1}/\text{g}$ FW)	Guaicole peroxidase activity ($\text{mmol min}^{-1}/\text{g}$ FW)	Grain yield (kg h^{-1})	Oil yield (kg h^{-1})
$I_w \times G_c \times F_c$	19.20 ^{gh}	11.02 ^e	31.80 ^{fg}	1.35 ^j	0.65 ^g	3298 ^{gh}	870.12 ^{gh}
$I_w \times G_c \times F_m$	13.05 ^j	7.32 ^g	24.13 ^{hi}	3.43 ^{fg}	2.38 ^e	4228 ^{bc}	1209.92 ^d
$I_w \times G_c \times F_v$	18.27 ^h	7.67 ^g	26.22 ^h	2.78 ⁱ	2.36 ^c	3995 ^{cd}	1102.8 ^{ef}
$I_w \times G_c \times F_{m+v}$	11.99 ^j	5.65 ^h	16.93 ^j	3.24 ^{gh}	3.45 ^c	4456 ^b	1287.72 ^c
$I_s \times G_c \times F_c$	21.95 ^e	19.02 ^a	70.05 ^a	3.97 ^e	1.75 ^f	2050 ^j	452.09 ^j
$I_s \times G_c \times F_m$	26.67 ^{cd}	12.18 ^{cd}	36.56 ^d	5.51 ^b	2.59 ^{de}	3233 ^{gh}	842.91 ^h
$I_s \times G_c \times F_v$	25.47 ^d	13.05 ^c	43.45 ^c	4.8 ^d	2.47 ^e	2350 ⁱ	604.20 ⁱ
$I_s \times G_c \times F_{m+v}$	33.93 ^a	10.80 ^e	34.26 ^e	5.06 ^c	3.69 ^{bc}	3237 ^{gh}	868.69 ^g
$I_w \times G_r \times F_c$	20.40 ^{fg}	9.67 ^f	29.69 ^g	1.71 ^j	3.24 ^c	3463 ^{fg}	1095.92 ^{ef}
$I_w \times G_r \times F_m$	19.81 ^{fg}	5.74 ^h	23.94 ⁱ	3.81 ^{ef}	4.08 ^b	4359 ^b	1422.83 ^b
$I_w \times G_r \times F_v$	20.98 ^{ef}	7.93 ^g	24.10 ^{hi}	2.93 ^h	3.1 ^{cd}	3999 ^{cd}	1290.68 ^c
$I_w \times G_r \times F_{m+v}$	16.03 ⁱ	5.52 ^h	16.40 ^j	3.37 ^{fgh}	4.24 ^b	4862 ^a	1602.66 ^a
$I_s \times G_r \times F_c$	21.93 ^e	17.10 ^b	59.19 ^a	4.48 ^d	3.37 ^c	2312 ⁱ	637.43 ⁱ
$I_s \times G_r \times F_m$	28.41 ^b	11.37 ^{de}	33.21 ^{ef}	5.92 ^b	4.13 ^b	3587 ^{ef}	1052.97 ^f
$I_s \times G_r \times F_v$	27.48 ^{bc}	12.45 ^c	42.04 ^c	5.61 ^c	4.06 ^b	3187 ^h	928.2 ^g
$I_s \times G_r \times F_{m+v}$	34.84 ^a	9.85 ^e	31.14 ^{fg}	7.15 ^a	5.27 ^a	3806 ^d	1129.08 ^e

Table 5. Comparison amongst 2-year mean values of physio-biochemical traits, leaf P, Total soluble sugars, Catalase and Superoxide Dismutase activity affected by mycorrhiza \times vermicompost under full and limited irrigation

Where I_w (full irrigation), I_s (limited irrigation), F_c (no fertilizer), F_m (mycorrhiza inoculation), F_v (vermicompost application) and F_{m+v} (combination of mycorrhiza and vermicompost). Means that were followed by the same letter in each column, are not significantly different

Treatment	Leaf P % on dry matter	Proline ($\mu\text{mol/g}$ FW)	Total soluble sugars ($\mu\text{mol/g}$ FW)	Glycine betaine ($\mu\text{mol/g}$ FW)	Hydrogen peroxide ($\mu\text{mol/g}$ FW)	Catalase activity ($\text{mmol min}^{-1}/\text{g FW}$)	Superoxide Dismutase activity (Units/g FW)
$I_w \times F_c$	0.47 ^d	23.04 ^d	22.78 ^e	19.80 ^{cd}		4.76 ^f	19.14 ^e
$I_w \times F_m$	0.65 ^{ab}	18.45 ^e	15.93 ^f	16.43 ^e		7.89 ^b	23.21 ^c
$I_w \times F_v$	0.64 ^b	18.63 ^e	15.26 ^f	19.62 ^d		7.23 ^d	21.71 ^d
$I_w \times F_{m+v}$	0.67 ^a	12.68 ^f	10.89 ^g	15.82 ^e		7.71 ^{bc}	23.21 ^c
$I_s \times F_c$	0.25 ^f	25.09 ^c	27.85 ^d	21.94 ^c		5.33 ^e	23.72 ^c
$I_s \times F_m$	0.44 ^{de}	27.80 ^b	39.10 ^b	27.54 ^b		8.61 ^a	32.91 ^a
$I_s \times F_v$	0.42 ^e	27.42 ^b	33.74 ^c	26.47 ^b		7.42 ^{cd}	29.08 ^b
$I_s \times F_{m+v}$	0.53 ^c	36.47 ^a	44.12 ^a	34.39 ^a		8.71 ^a	33.22 ^a

Table 6. Comparison amongst 2-year mean values of physio-biochemical traits, leaf P, Total soluble sugars , Superoxide Dismutase activity under full and limited irrigation

Where Y_1 (first year), Y_2 (second year), I_w (full irrigation), I_s (limited irrigation),. Means that were followed by the same letter in each column, are not significantly different

Treatment	Leaf P % on dry matter	Total soluble sugars ($\mu\text{mol/g}$ FW)	Superoxide Dismutase activity (Units/g FW)
$Y_{1 \times I_w}$	0.57 ^b	15.72 ^d	21.6 ^c
$Y_{1 \times I_c}$	0.37 ^d	35.10 ^b	22.03 ^c
$Y_{2 \times I_w}$	0.66 ^a	16.71 ^c	28.86 ^b
$Y_{2 \times I_c}$	0.45 ^c	37.31 ^a	30.6 ^a

Figures

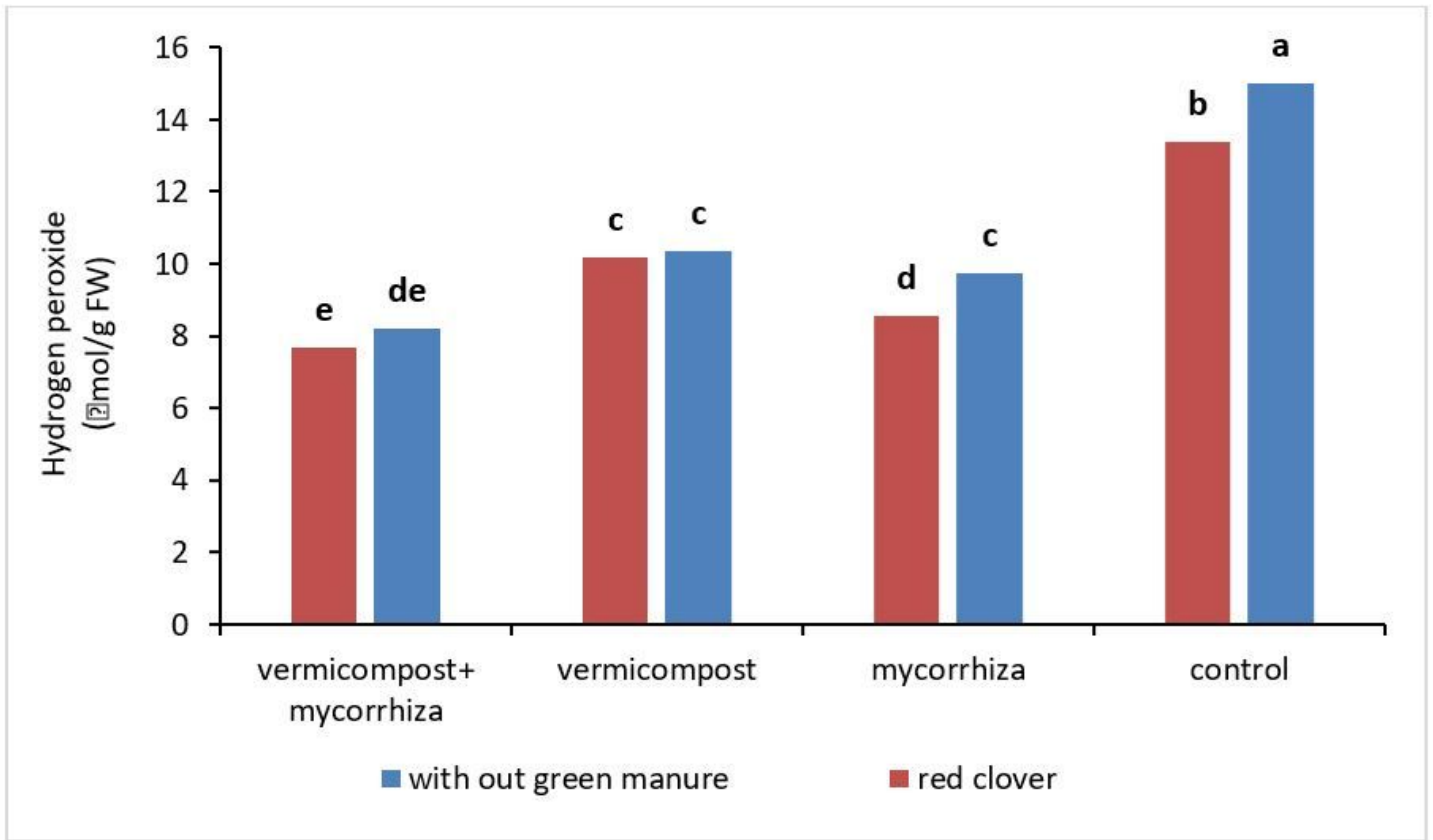


Figure 1

Interaction effects of green manure * fertilizers on Hydrogen peroxide concentration (Means followed by the same letter are not significantly different)

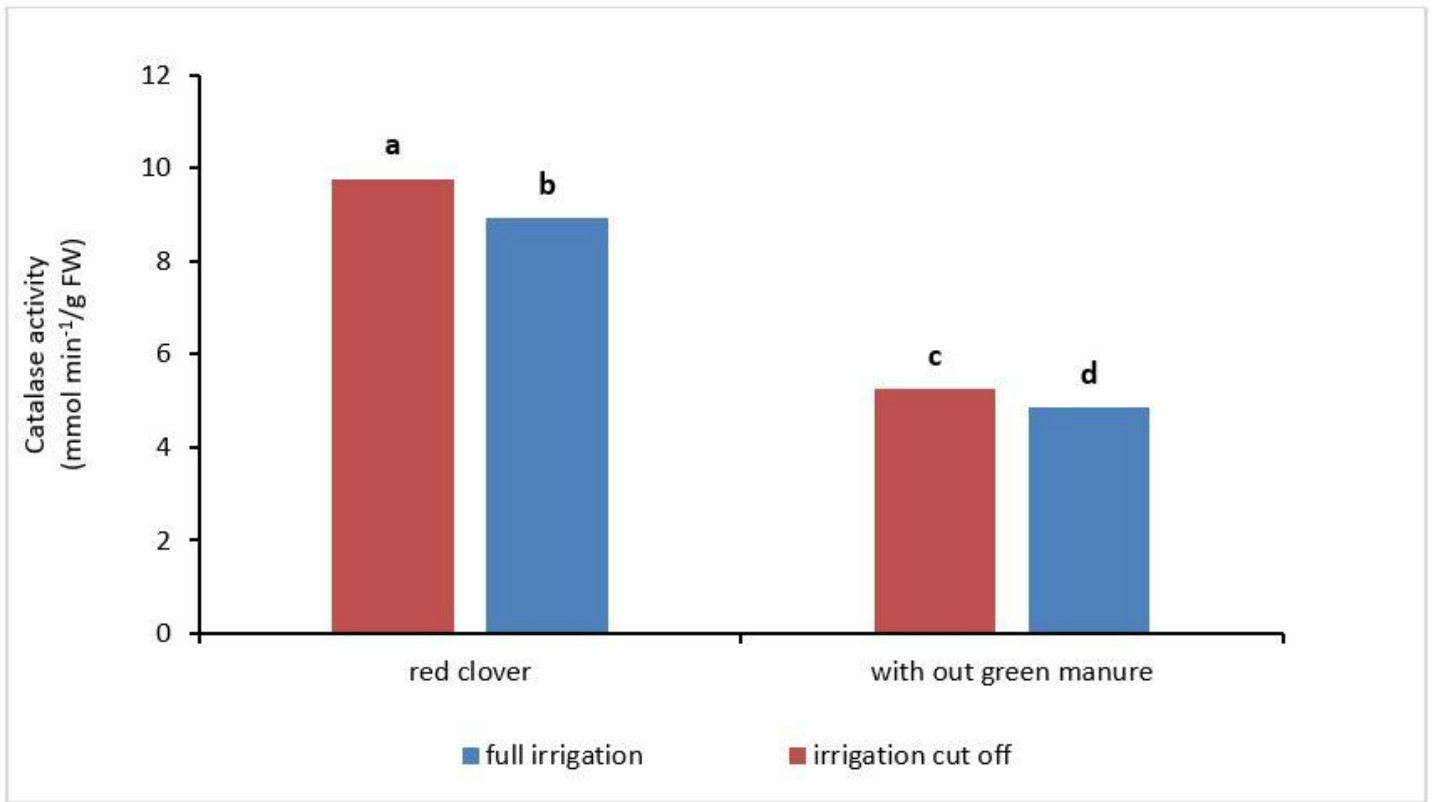


Figure 2

Interaction of green manure and irrigation regimes on catalase activity.

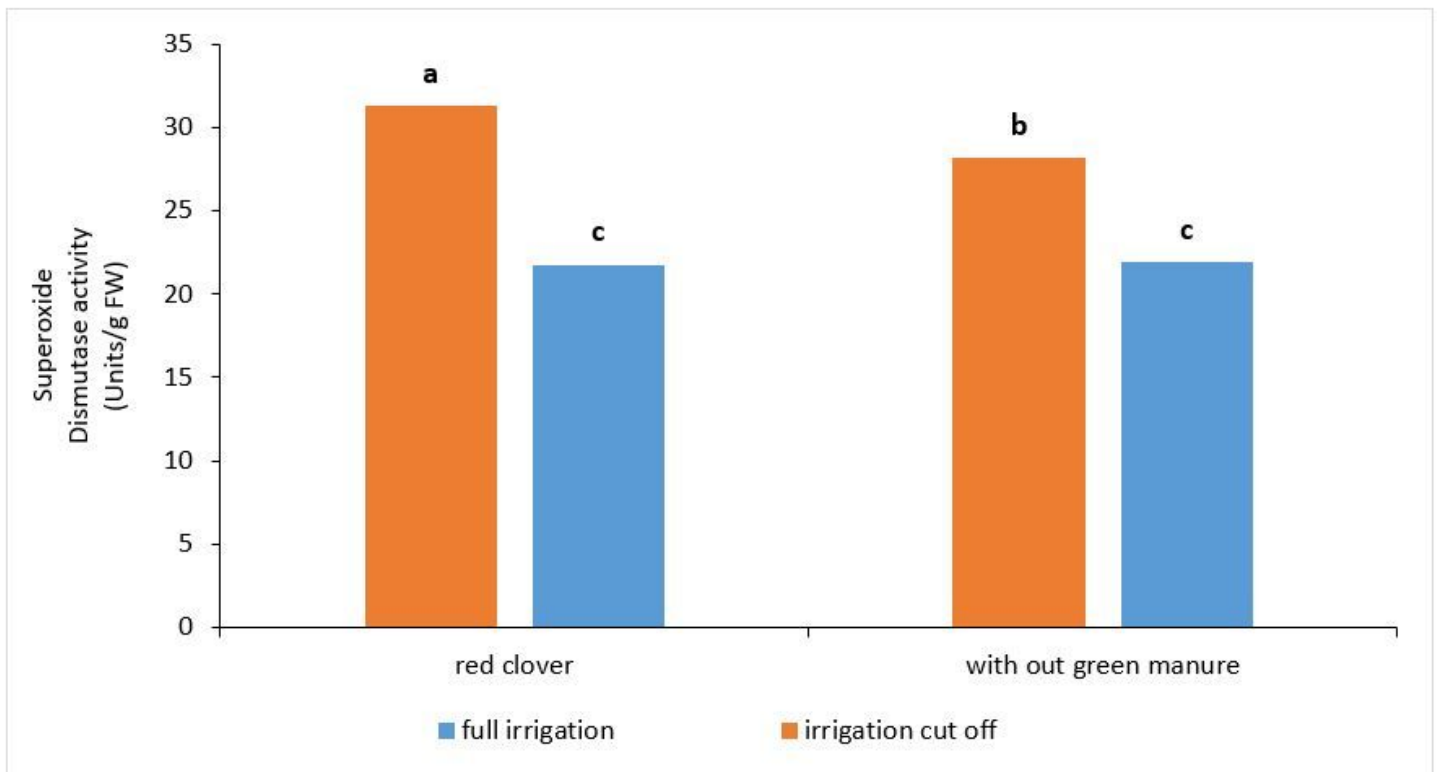


Figure 3

Interaction of green manure and irrigation regimes on Superoxide Dismutase activity.