

# Nootropic dipeptide, Noopept can modulate hyperalgesia by effecting on spinal microglia dependent Brain Derived Neurotropic Factor (BDNF) and pro-BDNF expression and apoptosis during persistent inflammation

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#### Research

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# **Abstract**

Background: There are largely unknown associations between changes in pain behavior responses during persistent peripheral inflammation and spinal cell alteration such as apoptosis. Some evidence suggests that microglia and microglia related mediators play notable roles in induction and maintenance of central nervous system pathologies and inflammatory pain. By considering those relationships and microglia related nootrophic factors, such as the Brain Derived Neurotrophic Factor (BDNF) in CNS, we attempted to assess the relationship between microglia dependent BDNF and its precursor with pain behavior through spinal cell apoptosis as well as the effect of Noopept on this relationship.

Methods: Persistent peripheral inflammation was induced by a single subcutaneous injection of Complete Freund's Adjuvant (CFA) on day 0. Thermal hyperalgesia, paw edema, microglial activity, microglia dependent BDNF, pro-BDNF expression, and apoptosis were assessed in different experimental groups by confirmed behavioral and molecular methods on days 0, 7, and 21 of the study.

Results: Our findings revealed hyperalgesia and spinal cell apoptosis significantly increased during the acute phase of CFA-induced inflammation but was then followed by a decrement in the chronic phase of the study. Aligned with these variations in spinal microglial activity, microglia dependent BDNF significantly increased during the acute phase of CFA-induced inflammation. Our results also indicated that daily administration of Noopept (during 21 days of the study) not only caused a significant decrease in hyperalgesia and microglia dependent BDNF expression but also changed the apoptosis process in relation to microglia activity alteration.

Conclusions: It appears that the administration of Noopept can decrease spinal cell apoptosis and hyperalgesia during CFA-induced inflammation due to its direct effects on microglial activity and microglia dependent BDNF and pro-BDNF expression.

# **Background**

In the case of persistent pain states, mounting evidence has illustrated that the release of a variety of inflammatory mediators, such as cytokines and growth factors from neuronal and non-neuronal cells, play a crucial role in the induction of inflammatory pain [1]. Although tremendous efforts have been made to understand the cellular and molecular basis of chronic pain, its physiological aspects remains mostly unknown [2, 3]. Multiple mechanisms contribute to inflammatory pain, each of which is subject to cellular and molecular alterations of nociceptive neurons in the dorsal horn of the spinal cord. These mechanism also underwrite peripheral inflammation, which is accompanied with inflammatory symptoms such as hyperalgesia and edema. Cell apoptosis, as an important consequence of central nervous system (CNS) pathological conditions, is induced by the increment of inflammatory mediators such as cytokines and growth factors [4, 5]. Data from diverse animal models support the idea that apoptosis can induce neuronal sensitization and hyperalgesia [6, 7]. It is noteworthy that these irreversible processes might be accompanied by different nervous system damages such as neuropathic pain [8]. In this regard, Kim et al.

showed that induction of spinal nerve ligation (SNL) in rats resulted in DRG neuronal apoptosis, which precedes pain behaviors such as mechanical allodynia [9]. Microglia, as resident macrophages within the CNS, are involved in mediating inflammatory processes and can release pro-inflammatory mediators such as TNF-α [10]. Like other cells, microglia can produce not only pro/anti-inflammatory mediators and neurotrophic factors such as Brain Derived Neurotrophic Factor (BDNF), but can also play neurotoxic or neuroprotective roles in CNS depending on the released mediators [11, 12]. It has become increasingly clear that microglial cells can powerfully control and modulate pain when they are activated in response to peripheral inflammation [13].

Data generated thus far indicates that BDNF is a neurotrophin with important biologic roles in neuronal survival and differentiation [14]. Also, it has a crucial role in regulating inflammatory pain thresholds and secondary hyperalgesia [15, 16]. Previous studies have shown that BDNF synthesis is greatly increased in different populations of DRG neurons in response to peripheral inflammation, and it also has apoptotic effects on neural cells [17, 18]. Groth and Aanonsen showed that BDNF induces an acute dose-dependent thermal hyperalgesic response and also affects central sensitization in NMDA receptor activation dependent processes [19]. On the other hand, Siuciak et al. presented that midbrain infusion of BDNF can decrease behavioral paw flinch response to subcutaneous formalin injection in both early and late phases of the test [20].

Accumulating evidence supports that Pro-Brain Neurotrophic Factor (pro-BDNF), as a BDNF precursor, binds to the p75 neurotrophin receptor (p75NTR) and exerts the opposing biological functions of mature BDNF. Pro-BDNF plays an active role in regulation of axon pruning, dendritic complexity, and also neuronal apoptosis in the hippocampus. An important piece of the puzzle is missing from the previous studies of BDNF, the microglial activity-dependent pro-BDNF and BDNF relationship with spinal cell apoptosis and pain behaviors during peripheral persistent inflammation [21, 22].

The term nootropics (cognition enhancer) is applied to a group of psychoactive substances which stimulate neuronal functions and enhance cognitive performances such as memory [23, 24]. N-Phenylacetyl-l-prolylglycine ethyl ester (Noopept), a dipeptide analog of piracetam, is one of these synthesized cognition enhancers [25, 26]. There is evidence to suggest that administration of Noopept can significantly suppress peripheral inflammatory responses to carrageenan [27]. Moreover, it has been shown that Noopept treatment can prevent accelerating neuronal death, which may be related to its capacity to block the neurotoxic effects of calcium ions and glutamate and specifically its anti-inflammatory effects. As such, it deserves consideration as a therapeutic target [28, 29].

In light of the principal role of microglia related mediators in inflammatory pain and cell death induction, the identification of neuroprotective and anti-inflammatory roles of Noopept should generate novel therapeutics for the treatment of inflammatory pain. In the context of inflammatory pain, it is also interesting to consider the relationship between spinal cell apoptosis and the pain process. We, therefore, aimed to investigate the relationships between microglia dependent BDNF and pro-BDNF and spinal cell

apoptosis and pain behavioral responses during different stages of CFA-induced inflammatory pain. Moreover, in a second step, we focused on the effects of Noopept administration through this pathway.

# **Methods**

#### Laboratory animals and experimental procedures

All experimental procedures were performed on adult male Wistar rats (200-220 g). The animals were kept under standard temperature, humidity and lighting condition (22±2°C, humidity 60–70%, 12h light/dark cycle) with free access to food and water. All procedures were approved by the Ethics Committee of Shahid Beheshti University of Medical Science (IR.SBMU.MSP.REC.1396.818), and were also in accordance with NIH Guidelines for the Care and Use of Laboratory Animals [5, 29]. The rats were divided into five experimental groups as follows: the (a) CFA group, (b) CFA+saline (vehicle) group, (c) Mineral oil group, (d) CFA+Noopept group, and (e) CFA+minocycline group. According to the study procedure, each group was further divided into 3 subgroups of six animals to receive injections at different time points in the study (days 0, 7, and 21). Similar to our previous studies, a persistent inflammatory pain model was induced by CFA on day zero. The mineral oil group (control group) received a single subcutaneous injection of sterile mineral oil (100  $\mu$ L) without the CFA on day zero. From the first day after inflammatory pain induction, all experimental groups received the daily drug interventions. Behavioral tests, detection of expression of spinal proteins, and a TUNEL assay were done on days 0, 7, and 21 of the study.

## Inflammatory pain induction

CFA-induced inflammatory pain was caused by a single subcutaneous injection of (100µL) heat-killed Mycobacterium tuberculosis suspended in sterile mineral oil (10 mg/ml; CFA; Sigma, St Louis, MO, USA) into the rats' hind paw on day zero under light anesthesia with methoxyisoflurane. The first week after CFA injection was considered the inflammatory phase and the third week was the arthritic phase [30, 31].

### **Drug administration**

In this study, Minocycline (Sigma-Aldrich, Germany), as a microglia activation inhibitor, was diluted in physiological saline (0.9% NaCl) and injected intraperitoneally (i.p.) daily at a dose of 40 ml/kg from the first day after CFA injection to the 21<sup>st</sup> day of the study in the CFA+minocycline group [32]. Noopept (a nootropic dipeptide) was dissolved in saline solution and injected (i.p.) daily at a dose of 5 mg/kg from the first day after CFA injection till day 21 in the CFA+Noopept group [27, 28, 33].

#### Assessment of paw edema

Paw edema assessment was done by measuring paw volume variations during the different stages of the study using a Plethysmometer (model 7141; Ugo Basile, Comerio-Varese, Italy). Briefly, the rats' hind paws were submerged to the tibiotarsal joint in the electrolyte-filled perspex cell of the plethysmometer. The amount of liquid displaced, which is related to the paw volume, was shown on a digital display. Volume measurements were done twice for each paw, and the average was calculated. Edema was considered by measuring the differences between day zero and the other time points, days 7 and 21 of the study [34].

#### Thermal hyperalgesia assessment

Paw withdrawal latency (PWL) was used to measure thermal hyperalgesia by applying the Hargreaves test [35]. For this purpose, the rats were placed in Plexiglass chambers (Ugo Basile, Verse, Italy) and habituated for 15 minutes to the test environment. The heat source was adjusted under the plantar surface of the injected and non-injected hind paws and an infrared light was projected focally, and then PWL was measured. Each paw was tested three times at 5 min intervals and the average value of the withdrawal latency of three consecutive tests was calculated and deducted from the other paw. In the present study, the cut-off time in the absence of a response was 20s.

#### Western blot

After the behavioral tests, western blot was used to examine the amounts of BDNF, pro-BDNF, Iba1, and Caspase-3 expression in the lumbar spinal cord. In this regard, the lumbar part of the spinal cord (L1-L5) was removed and homogenized in a lysis buffer. Protein samples were separated on a sodium dodecyl sulfate—polyacrylamide gel electrophoresis (12% SDS-PAGE) and transferred to an activated Polyvinylidene difluoride (PVDF) membrane (Roche Diagnostics, Indianapolis, IN, USA), then blocked with 5% Bovine serum albumin (BSA) and incubated overnight at 4°C with primary antibodies against BDNF (1:1000, ab205067), pro-BDNF (1:1000, NB100-98754), Iba1 (1:1000, ab178846), and Caspase-3 (1;3000, ab184787). The membrane then was washed three times and incubated for 1h at room temperature with a secondary antibody (1:20000, cell signaling #7074). Finally, the blots were visualized with a chemiluminescence detection system (ECL, Amersham, RPN2235). Band intensity was measured densitometrically using Image J and expressed as the ratio of the intensity of the BDNF, pro-BDNF, Iba1, and Caspase-3 band to  $\beta$ -actin [36].

#### **TUNEL** assay

The apoptotic cells in the lumbar section of the spinal cord were detected by TUNEL staining. Briefly, lumbar segments of the spinal cord were embedded in paraffin after fixing in 4% paraformaldehyde for

24h. Then, five slides from each block were prepared for assessing the apoptotic index by means of the TUNEL technique. After deparaffinization in xylene, the sections were rehydrated with different dilutions of alcohol and treated with  $H_2O_2$  and methanol at room temperature. Afterwards, they were incubated in proteinase K. The TUNEL technique was carried out using a TUNEL-based assay kit (SIGMA, Roche, Germany). The sections were incubated first in horseradish peroxidase (POD) and then in diaminobenzidine (DAB) solution and subsequently counterstained with hematoxylin. Five watch fields were chosen randomly on each section under a microscope (Nikon-E600). Positive brown cells and total cells were counted and analyzed using GraphPad Prism 6 software [37, 38].

#### Statistical analysis

Data were shown as mean±standard error of mean (SEM). The statistical analysis used for the behavioral, cellular, and molecular tests was a one way ANOVA. Statistical comparisons were analyzed by repeated measurements followed by Post hoc Tukey's for evaluation of variants within the groups. An unpaired Student's t test was used to compare the changes of variants on the same days between the two groups. Statistical significance was accepted at the  $p \le 0.05$  level. Statistical analysis were performed using the GraphPad Prism 6.0 program.

# **Results**

Paw volume variation during different stages of CFA-induced inflammatory pain

There were no significant differences in behavioral and molecular parameters between the mineral oil and CFA groups on day zero (data not shown). CFA injection into the rat's hind paw caused a significant increase in paw volume, which persisted until the 21st day of the study. Paw volume showed a significant increase on the 7th and 21st days after CFA injection compared with day zero in the CFA group (p < 0.001). Our data also indicated that paw volume significantly increased on day 21 compared with day 7 after CFA injection in the CFA group (p < 0.001). On the other hand, three weeks administration of Noopept in the CFA + Noopept group significantly reduced paw volume on the 7th (p < 0.001) and 21st (p < 0.001) days of the study compared to the same days in the CFA group. Furthermore, our results revealed that paw volume significantly decreased in the CFA + Noopept group in comparison with the CFA + Minocycline group on days 7 and 21 of the study (p < 0.001). Saline administration, as a vehicle, caused no significant differences in paw volume in comparison with the CFA group at all-time points of the study (Fig. 1).

Thermal hyperalgesia variations during different stages of CFA-induced inflammatory pain

Our results revealed that hyperalgesia noticeably increased on the 7th day after CFA injection compared with day zero (p < 0.001). However, it significantly decreased on day 21 in comparison with day 7 (p < 0.01) in the CFA group, although hyperalgesia on day 21 was still more than day zero (p < 0.001). Long-

term Noopept administration in the CFA + Noopept group significantly reduced thermal hyperalgesia compared with the CFA group on days 7 and 21 (p < 0.001). Moreover, the results illustrated that thermal hyperalgesia significantly decreased in the CFA + Noopept group in comparison with CFA + Minocycline group on days 7 and 21 (p < 0.001). The CFA + Saline (vehicle group) group indicated no significant differences in paw withdrawal latency in comparison to the CFA group at all times of the study (Fig. 2).

Western blot analysis of spinal BDNF expression during different stages of CFA-induced inflammatory pain

Following intraplantar injection of CFA, lumbar spinal BDNF expression demonstrated various changes during the different stages of the study. The results revealed that CFA injection caused significant increase in BDNF expression on day 7 in comparison with day zero (p < 0.001). There was a significant decrease in BDNF expression on day 21 in comparison with day 7 in the CFA group (p < 0.001). Our findings indicated that the administration of Noopept in the CFA + Noopept group caused a significant decrease in spinal BDNF expression on the 7th (p < 0.001) and 21st (p < 0.001) days of study in comparison with the same day in the CFA group. In addition, in comparison with the CFA group, the decrease in BDNF expression in the the CFA + Noopept group was significantly more than what was observed in the CFA + Minocycline group on the 7th (p < 0.001) and 21st (p < 0.001) days of study. The CFA + Saline (vehicle) group indicated no significant differences in spinal BDNF expression in comparison to the CFA group at all-time points of the study (Fig. 3).

Assessment of spinal pro-BDNF expression during different stages of CFA-induced inflammatory pain

Western blotting analysis of pro-BDNF expression in the lumbar part of the spinal cord showed a significant increase on day 7 in comparison with day zero in the CFA group (p < 0.001). On the other hand, spinal pro-BDNF expression in the CFA group considerably decreased on day 21 compared with days zero (p < 0.05) and 7 (p < 0.001). Our findings also indicated that the Noopept treatment in the CFA + Noopept group significantly decreased pro-BDNF expression on day 7 (p < 0.001 compared with the same day in the CFA group. Based on our results, administration of Noopept in the CFA injected rats was significantly more effective in decreasing pro-BDNF expression compared with the CFA + Minocycline and CFA groups during acute phase of the study (p < 0.001). The CFA + Saline group presented no significant differences in spinal pro-BDNF expression in comparison with the CFA group at all time points of the study (Fig. 4).

Spinal microglial activation variations during different stages of CFA-induced inflammatory pain

Following intra plantar injection of CFA, expression of Iba1, as a microglial marker at the lumbar spinal cord, revealed a significant increase on 7th day of the study compared with day zero (p < 0.001). There was significant difference in the Iba1 expression level between day 21 compared with days 0 (p < 0.05) and 7 (p < 0.001) in the CFA group. As shown in Fig. 5, administration of Noopept in the CFA + Noopept group considerably decreased the Iba1 expression level on days 7 (p < 0.001) and 21 (p < 0.05) in comparison with the same days in the CFA group. Furthermore, our findings showed that the decrease in spinal Iba1/ $\beta$ -actin expression level in the CFA + Noopept group was significantly greater than what was

observed in the CFA + Minocycline group on 7th day compared with the CFA group (p < 0.001). The CFA + Saline (vehicle group) indicated no significant differences in spinal lba1 expression in comparison with the CFA group at all times of the study (Fig. 5).

Measurement of spinal apoptotic cells during different stages of CFA-induced inflammatory pain

After CFA injection, the apoptosis index significantly increased in the superficial laminas of the ipsilateral lumbar segment of the spinal cord on day 7 (p < 0.001) in the CFA group compared with day zero. In addition, There was significant decrease in the apoptosis index on day 21 compared with day 0 (p < 0.01). Regarding our findings, the number of apoptotic cells considerably decreased on the 7th (p < 0.001) and 21st (p < 0.01) days in the CFA + Noopept group compared with the same days in the CFA group. As presented in Fig. 6, There was significant decrease in the percentage of apoptotic cells on the 7th (p < 0.05) and 21st (p < 0.05) days in the CFA + Noopept compared with the same days in the CFA + Minocycline group. There was no significant difference in the apoptosis index in the CFA + Saline group in comparison with the CFA group at all times of the study (Fig. 6).

Cleaved spinal caspase-3 variation during different phases of CFA-induced inflammatory pain

Our results revealed that CFA injection caused a significant increase in cleaved caspase-3 levels on day 7 (p < 0.001) compared with day 0 in the CFA group. There was also significant decrease in cleaved caspase-3 levels on 21st day of study compared with day 7 (p < 0.001) in the CFA group. Long-term Noopept treatment in the CFA + Noopept group significantly decreased CFA-induced caspase-3 activity on the 7th day of study (p < 0.05) compared with the same days in the CFA group. Based on our results, administration of Noopept in the CFA + Noopept group more significantly decreased cleaved caspase-3 levels on both the 7th (p < 0.05) and 21st (p < 0.05) days when compared with the same day in the CFA + Minocycline group. The CFA + Saline group presented no significant differences in spinal cleaved caspase-3 levels in comparison with the CFA group in all time points of the study (Fig. 7).

# **Discussion**

The current study was designed to clarify the involvement of microglia dependent BDNF in the alteration of pain behaviors and spinal cell apoptosis. We also examined microglial activity and the spinal level of BDNF in Noopept-treated rats in the context of inflammatory pain and spinal cell apoptosis. Our findings in the first step of the study indicated that microglia dependent BDNF and pro-BDNF expressions aligned with thermal hyperalgesia, and spinal dorsal horn cell apoptosis increased in the CFA injected rats on day 7 but decreased on day 21. Thus, inhibition of microglial activity by minocycline caused a decrement of these variations during different phase of the study.

Large bodies of evidence exist to substantiate that increment of pain sensitivity is initially due to neural function variations, and microglia has a pivotal role in the pathogenesis of pain [13, 39]. In line with the results from our previous studies, this study shows that CFA-induced inflammation caused persistent spinal microglial activation, which occurred in parallel with hyperalgesia increment, and can be inhibited

by Minocycline as a specific microglia inhibitor [30, 40]. It is therefore conceivable that microglia has a vitally important role in the initiation and maintenance of pain, but little attention has been paid to the specific mechanisms of spinal glia cells and their roles in the induction and development of pain behaviors [41, 42].

It is currently well established that, except for certain neurons, activated microglia can release various mediators such as BDNF in response to homeostatic changes [43, 44]. Ulmann et al. demonstrated that following peripheral nerve injury in in vivo and in vitro models, the stimulation of purinergic receptor P2X leads to the release of BDNF from activated microglia [4, 45]. In this study, we found that microglia dependent BDNF and pro-BDNF expressions in the lumbar part of the spinal cord increased on day 7, but decreased on day 21. These results matched with the behavioral outcomes. It is intriguing to note that inhibition of microglia led to a decrement of spinal BDNF and pro-BDNF expression on day 7; however, its effects on BDNF and its precursor on day 21 was not significant. BDNF, a member of the neurotrophin family, is produced following intra and extra cellular processing of its precursor, pro-BDNF [15, 46]. Many studies have reported that pro-BDNF is not an inactive biological molecule and has diverse cellular functions [47, 48]. Numerous data convincingly support the notion that the effects of BDNF and pro-BDNF are, in fact, opposite [21]. It is widely accepted that BDNF promotes neuronal survival through activation of the TrKB receptor, while pro-BDNF induces cell apoptosis through p75 receptor activation [49]. However, there is a dearth of information about the relationship of microglia dependent BDNF and pro-BDNF on the spinal dorsal horn in pain behaviors during inflammatory pain. BDNF primarily synthesized as a larger precursor, pro-BDNF, which undergoes proteolytic processing by proteases [50, 51]. It is noteworthy that the conversion of pro-BDNF to BDNF depends on neuronal activity [52]. Matsumoto et al. have shown that the capacity of a neuron for pro-BDNF processing is limited [53]. In this regard, Yoshimura et al. and Wysokiński have reported that a disturbance in BDNF and pro-BDNF balances may be involved in the pathogenesis of mental disorders [54, 55]. Moreover, in vitro studies have indicated that pro-BDNF enhances apoptosis of basal forebrain neurons and sympathetic neurons [56, 57].

Apoptosis is a crucial physiological process and any defect in apoptotic mechanisms is closely related to different disorders like pain [58, 59]. This study results showed that the number of apoptotic cells that aligned with the spinal cleaved caspase-3 level during the first week after CFA injection significantly increased in the ipsilateral dorsal horn of the spinal cord in comparison to day zero, which shows the changes in apoptosis following CFA injection were also well related to hyperalgesia during various time points of peripheral inflammation [60].

The above finding was consistent with changes in microglia dependent BDNF and pro-BDNF expression during the study. So, it seems that microglial BDNF and pro-BDNF imbalances during different phases of the study, specifically on day 7, may lead to apoptosis more than neuronal survival. Therefore, a greater understanding of the role of the microglia signaling pathways in spinal apoptosis and their interaction with hyperalgesia variations opens up the intriguing possibility of manipulating microglial cells to eliminate or reduce pain.

Our observations in the second step of the study suggested that the long-term administration of Noopept during persistent peripheral inflammation not only decreases the activity of spinal microglia but also decreases microglia dependent BDNF and pro-BDNF expressions in parallel with thermal hyperalgesia and dorsal horn cell apoptosis. Despite the many studies done on the effects of Nootropics, such as Noopept, on cognitive disorders like Attention-Deficit/Hyperactivity Disorder (ADHD), Alzheimer's and Parkinson's diseases [61, 62], the mechanism of the effect of this novel dipeptide on persistent inflammatory pain and its involved central mechanisms remained largely uncharacterized. Our results demonstrated that Noopept treatment was not only effective in reducing pain behaviors but also reduced spinal cell apoptosis, which is related to pain behavior variations in the context of CFA-induced inflammation. It is of interest to note that during this study the administration of Noopept not only reduced spinal Iba1 level, as a microglial activity marker, but also reduced microglia dependent BDNF and pro-BDNF expression levels in the lumbar part of the spinal cord. Although there are considerable studies on the anti-inflammatory effect of Noopept [28, 61], the current study stated the novel dipeptide antihyperalgesic and anti-inflammatory effects were mediated via inhibition of microglia activity and apoptosis. Ostrovskaya et al. showed that Noopept, through normalizing incretin system parameters, had an anti-apoptotic effect on pancreatic β cells in the context of experimental Diabetes [63], and in another study they indicated neuroprotective effects [64]. Moreover, the effects reported in this paper in response to Noopept were significantly more effective than those reported for minocycline administration in the CFA-inflamed groups, which indicates that there are other pathways involved in Noopept antiinflammatory effects which need to be investigated. The results of the present experiment and similar reports dealing with experimental models of inflammatory pain suggest that alleviation of pain symptoms and cell apoptosis associated with long-term Noopept treatment may have been mediated more by decreasing the levels of microglia dependent BDNF than pro-BDNF expression. Data obtained from different part of this study did not rule out the involvement of other pathways, specifically in glia cells.

# Conclusion

The results of this study highlight that an alteration of spinal cell apoptosis and thermal hyperalgesia was linked with the amplification of microglial activity and increment of microglia dependent levels of BDNF and pro-BDNF. Furthermore, the anti-apoptotic and analgesic effects of Noopept may be mediated via decreasing microglial activity and microglia dependent levels of BDNF and pro-BDNF during CFA-induced persistent inflammatory pain. In our opinion, nootrophic peptides administration, such as Noopept, can be considered as a new and important target for inflammatory pain reduction, although this conclusion requires further in-depth research.

# **Declarations**

Ethics approval and consent to participate

All animal procedures were in compliance with the Ethics Committee of Shahid Beheshti University of Medical Science (IR.SBMU.MSP.REC.1396.818), and also NIH Guidelines for the Care and Use of Laboratory Animals.

## **Consent for publication**

Not applicable.

#### Availability of data and materials

Please contact corresponds for data requests.

#### **Competing interests**

The authors declared that they have no competing interests.

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#### **Authors' contributions**

Conceptualization: Dr. Jalal Zaringhalam; Methodology: Dr. Nader Maghsoudi, Dr. Homa manaheji, Dr. Valery Akparov, Dr.Samira Danyali, Dr. Rasoul Ghasemi; Manuscript Preparation: Mona Taghizadeh, Mansoureh Baniasadi, Mola Mohammadi.

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# References

- [1] V. Nazemian, S. Kalanaky, H. Manaheji, E. Houshmandi, M. Mohammadi, J. Zaringhalam, S. Mirjafari, Anti-hyperalgesia effect of nanchelating based nano particle, RAc1, can be mediated via liver hepcidin expression modulation during persistent inflammation, Int. Immunopharmacol. 69 (2019) 337–46. https://doi.org/10.1016/j.intimp.2019.02.003.
- [2] S.B. McMahon, W.B.J. Cafferty, F. Marchand, Immune and glial cell factors as pain mediators and modulators, Exp. Neurol. 192 (2005) 444–62. https://doi.org/10.1016/j.expneurol.2004.11.001

- [3] V. Nazemian, M. Shadnoush, H. Manaheji, J. Zaringhalam, Probiotics and inflammatory pain: a literature review study, Middle East J. Rehabil. Health Stud. 3 (2016) e36087. https://10.17795/mejrh-36087.
- [4] E. K. Joseph, J.D. Levine, Caspase signalling in neuropathic and inflammatory pain in the rat, Eur. J. Neurosci. 20 (2004) 2896–902. https://doi.org/10.1111/j.1460-9568.2004.03750.x
- [5] V. Nazemian, H. Manaheji, A.M. Sharifi, J. Zaringhalam, Long term treatment by mesenchymal stem cells conditioned medium modulates cellular, molecular and behavioral aspects of adjuvant-induced arthritis, Cell. Mol. Biol. (Noisy-le-grand) 64 (2018) 19-26. http://dx.doi.org/10.14715/cmb/2018.64.2.5.
- [6] M. Zimmermann, Pathobiology of neuropathic pain, Eur. J. Pharmacol. 429 (2001) 23–37. https://doi.org/10.1016/S0014-2999(01)01303-6.
- [7] G.T. Whiteside, R. Munglani, Cell death in the superficial dorsal horn in a model of neuropathic pain, J. Neurosci. Res. 64 (2001) 168–73. https://doi.org/10.1002/jnr.1062.
- [8] A. Coutaux, F. Adam, J.C. Willer, D. Le Bars, Hyperalgesia and allodynia: peripheral mechanisms, Jt. Bone Spine. 72 (2005) 359–71. https://doi.org/10.1016/j.jbspin.2004.01.010.
- [9] S.H. Kim, J.S. Nam, D.K. Choi, W.W. Koh, J.H. Suh, J.G. Song, J.W. Shin, J.G. Leem, Tumor Necrosis Factor-alpha and Apoptosis Following Spinal Nerve Ligation Injury in Rats, Korean J. Pain 24 (2011) 185–90. https://doi.org/10.3344/kjp.2011.24.4.185.
- [10] J.M. Zhang, J. An, Cytokines, inflammation, and pain, Int. Anesthesiol. Clin. 45 (2007) 27–37. https://doi.org/10.1097/AIA.0b013e318034194e.
- [11] M.R. Suter, Y.R. Wen, I. Decosterd, R.R. Ji, Do glial cells control pain?, Neuron Glia Biol. 3 (2007) 255–68. https://doi.org/10.1017/S1740925X08000100.

- [12] T. Trang, S. Beggs, M.W. Salter, Brain-derived neurotrophic factor from microglia: a molecular substrate for neuropathic pain, Neuron Glia Biol. 7 (2011) 99–108. https://doi.org/10.1017/S1740925X12000087.
- [13] H. Zhao, A. Alam, Q. Chen, M.A. Eusman, A. Pal, S. Eguchi, L. Wu, D. Ma, The role of microglia in the pathobiology of neuropathic pain development: what do we know?, B.J.A. Br. J. Anaesth. 118 (2017) 504–16. https://doi.org/10.1093/bja/aex006.
- [14] F. Ferrini, Y. De Koninck, Microglia control neuronal network excitability via BDNF signalling, Neural. Plast. 2013 (2013) 429815. http://dx.doi.org/10.1155/2013/429815.
- [15] D.K. Binder, H.E. Scharfman, Brain-derived Neurotrophic Factor, Growth Factors, 22 (2004) 123-131. https://dx.doi.org/10.1080%2F08977190410001723308
- [16] J. Zhao, A. Seereeram, M.A. Nassar, A. Levato, S. Pezet, G. Hathaway, et al., Nociceptor-derived brain-derived neurotrophic factor regulates acute and inflammatory but not neuropathic pain, Mol. Cell. Neurosci. 31 (2006) 539–48. https://doi.org/10.1016/j.mcn.2005.11.008.
- [17] K. Obata, K. Noguchi, BDNF in sensory neurons and chronic pain, Neurosci. Res. 55 (2006) 1–10. https://doi.org/10.1016/j.neures.2006.01.005.
- [18] X.H. Cao, H.S. Byun, S.R. Chen, Y.Q. Cai, H.L. Pan, Reduction in voltage-gated K+ channel activity in primary sensory neurons in painful diabetic neuropathy: role of brain-derived neurotrophic factor, J. Neurochem. 114 (2010) 1460-75. https://doi.org/10.1111/j.1471-4159.2010.06863.x.
- [19] R. Groth, L. Aanonsen, Spinal brain-derived neurotrophic factor (BDNF) produces hyperalgesia in normal mice while antisense directed against either BDNF or trkB, prevent inflammation-induced hyperalgesia, Pain 100 (2002) 171–81. https://doi.org/10.1016/S0304-3959(02)00264-6.

- [20] Siuciak JA, Wong V, Pearsall D, Wiegand SJ, Lindsay RM. BDNF produces analgesia in the formalin test and modifies neuropeptide levels in rat brain and spinal cord areas associated with nociception. Eur J Neurosci. 1995;7(4):663–70. https://dx.doi.org/ 10.1111/j.1460-9568.1995.tb00670.x
- [21] T.V. Ilchibaeva, E.M. Kondaurova, A.S. Tsybko, R.V. Kozhemyakina, N.K. Popova, V.S. Naumenko, Brain-derived neurotrophic factor (BDNF) and its precursor (proBDNF) in genetically defined fear-induced aggression, Behav. Brain Res. 290 (2015) 45–50. https://doi.org/10.1016/j.bbr.2015.04.041.
- [22] C. Luo, X.L. Zhong, F.H. Zhou, J. Li, P. Zhou, J.M. Xu, B. Song, C.Q. Li, X.F. Zhou, R.P. Dai, Peripheral Brain Derived Neurotrophic Factor Precursor Regulates Pain as an Inflammatory Mediator, Sci. Rep. 6 (2016) 27171. https://doi.org/10.1038/srep27171.
- [23] Giurgea, C. E. 1972. "Vers Une Pharmacologie de l'activité Intégrative Du Cerveau. Tentative Du Concept Nootrope En Psychopharmacologie."
- [24] Gudasheva, Tatiana A., Rita U. Ostrovskaya, and Sergey B. Seredenin. "Novel Technologies for Dipeptide Drugs Design and Their Implantation." Current Pharmaceutical Design 24 (2018) 3020–27. https://doi.org/10.2174/1381612824666181008105641.
- [25] N.A. Suliman, C.N. Mat Taib, M.A. Mohd Moklas, M.I. Adenan, M.T. Hidayat Baharuldin, R. Basir, Establishing Natural Nootropics: Recent Molecular Enhancement Influenced by Natural Nootropic, Evidence-Based Complement Altern. Med. 2016 (2016) 1–12. http://dx.doi.org/10.1155/2016/4391375.
- [26] Gudasheva, T. A., T. A. Voronina, R. U. Ostrovskaya, G. G. Rozantsev, N. I. Vasilevich, S. S. Trofimov, E. V. Kravchenko, A. P. Skoldinov, and S. B. Seredenin. "Synthesis and Antiamnesic Activity of a Series of N-Acylprolyl-Containing Dipeptides." European Journal of Medicinal Chemistry 31 (1996) 151–57. https://doi.org/10.1016/0223-5234(96)80448-X.

- [27] S.V. Alekseeva, L.P. Kovalenko, A.V. Tallerova, T.A. Gudasheva, A.D. Durnev AD, An experimental study of the anti-inflammatory action of noopept and its effect on the level of cytokines, Eksp. Klin. Farmakol. 75 (2012) 25–7.
- [28] L.P. Kovalenko, M.G. Miramedova, S.V. Alekseeva, T.A. Gudasheva, R.U. Ostrovskaia, S.B. Seredenin, Anti-inflammatory properties of noopept (dipeptide nootropic agent GVS-111), Eksp. Klin. Farmakol. 65 (2018) 53–5.
- [29] J. Zaringhalam, A. Akbari, A. Zali, H. Manaheji, V. Nazemian, M. Shadnoush, S. Ezzatpanah, Long-Term Treatment by Vitamin B<sub>1</sub> and Reduction of Serum Proinflammatory Cytokines, Hyperalgesia, and Paw Edema in Adjuvant-Induced Arthritis, Basic Clin. Neurosci. 7 (2016) 331-340. https://doi.org/10.15412/J.BCN.03070406.
- [30] V Nazemian, B Nasseri, H. Manaheji, J. Zaringhalam, Effects of mesenchymal stem cells conditioned medium on behavioral aspects of inflammatory arthritic pain induced by CFA adjuvant, J. Cell. Mol. Anesth. 1 (2016) 47-55. https://doi.org/10.22037/jcma.v1i2.11429.
- [31] B. Nasseri, J. Zaringhalam, S. Daniali, H. Manaheji, Z. Abbasnejad, V. Nazemian, Thymulin treatment attenuates inflammatory pain by modulating spinal cellular and molecular signaling pathways, Int. Immunopharmacol. 70 (2019) 225–34. https://doi.org/10.1016/j.intimp.2019.02.042.
- [32] B. Nasseri, V. Nazemian, H. Manaheji, J. Zaringhalam, Microglia are involve in pain related behaviors during the acute and chronic phase of arthritis inflammation, J. Cell. Mol. Anesth. 1 (2016) 137–45. https://doi.org/10.22037/jcma.v1i4.13557.
- [33] S.S. Boyko, V.P. Zherdev, R.V. Shevchenko, Pharmacokinetics of noopept and its active metabolite cycloprolyl glycine in rats, Biomeditsinskaia Khimiia 64 (2018) 455-8. https://doi.org/10.18097/PBMC20186405455.

- [34] J. Zaringhalam, E. Tekieh, H. Manaheji, Z. Akhtari, Cellular events during arthritis-induced hyperalgesia are mediated by Interleukin-6 and p38 MAPK and their effects on the expression of spinal mu-opioid receptors, Rheumatol. Int. 33 (2013) 2291–9. https://doi.org/10.1007/s00296-013-2715-2.
- [35] S. Nazemi, H. Manaheji, J. Zaringhalam, M. Sadeghi, A. Haghparast, Post-injury repeated administrations of minocycline improve the antinociceptive effect of morphine in chronic constriction injury model of neuropathic pain in rat, Pharmacol. Biochem. Behav. 102 (2012) 520–5. https://doi.org/10.1016/j.pbb.2012.07.001.
- [36] J. Zaringhalam, A. Hormozi, E. Tekieh, J. Razavi, R. Khanmohammad, S. Golabi, Serum IL-10 involved in morphine tolerance development during adjuvant-induced arthritis, J. Physiol. Biochem. 70 (2014) 497–507. https://doi.org/10.1007/s13105-014-0330-7.
- [37] X. Hu, X. Zhou, B. He, C. Xu, L. Wu, B. Cui, H. Wen, Z. Lu, H. Jiang, Minocycline protects against myocardial ischemia and reperfusion injury by inhibiting high mobility group box 1 protein in rats, Eur. J. Pharmacol. 638 (2010) 84–9. https://doi.org/10.1016/j.ejphar.2010.03.059.
- [38] Q. Wei, Y. Bian, F. Yu, Q. Zhang, G. Zhang, Y. Li, S. Song, X. Ren, J. Tong, Chronic intermittent hypoxia induces cardiac inflammation and dysfunction in a rat obstructive sleep apnea model, J. Biomed. Res. 30 (2016) 490–5. https://dx.doi.org/10.7555%2FJBR.30.20160110.
- [39] E.D. Milligan, L.R. Watkins, Pathological and protective roles of glia in chronic pain, Nat. Rev. Neurosci. 10 (2009) 23–36. https://doi.org/10.1038/nrn2533.
- [40] H. Zeinali, H. Manaheji, J. Zaringhalam, Z. Bahari, S. Nazemi, M. Sadeghi, Age-related differences in neuropathic pain behavior and spinal microglial activity after L5 spinal nerve ligation in male rats, Basic Clin. Neurosci. 7 (2016) 203–12. https://dx.doi.org/10.15412%2FJ.BCN.03070305.
- [41] R.R. Ji, T. Berta, M. Nedergaard, Glia and pain: Is chronic pain a gliopathy?, Pain 154 (2013) S10–28. https://doi.org/10.1016/j.pain.2013.06.022.

- [42] C. Abbadie, S. Bhangoo, Y. De Koninck, M. Malcangio, S. Melik-Parsadaniantz, F.A. White, Chemokines and pain mechanisms, Brain Res. Rev. 60 (2009) 125–34. https://doi.org/10.1016/j.brainresrev.2008.12.002.
- [43] E. Amy Old, A.K. Clark, M. Malcangio, The Role of Glia in the Spinal Cord in Neuropathic and Inflammatory Pain, Handb. Exp. Pharmacol. 227 (2015) 145-170. https://doi.org/10.1007/978-3-662-46450-2\_8.
- [44] M. Tsuda, K. Inoue, M.W. Salter, Neuropathic pain and spinal microglia: a big problem from molecules in 'small' glia, Trends Neurosci. 28 (2005) 101–7. https://doi.org/10.1016/j.tins.2004.12.002.
- [45] L. Ulmann, J.P. Hatcher, J.P. Hughes, S. Chaumont, P.J. Green, F. Conquet, G.N. Buell, A.J. Reeve, I.P. Chessell, F. Rassendren, Up-Regulation of P2X4 Receptors in Spinal Microglia after Peripheral Nerve Injury Mediates BDNF Release and Neuropathic Pain, J. Neurosci. 28 (2008) 11263–8. https://doi.org/10.1523/JNEUROSCI.2308-08.2008.
- [46] B. Fayard, S. Loeffler, J. Weis, E. Vögelin, A. Krüttgen, The secreted brain-derived neurotrophic factor precursor pro-BDNF binds to TrkB and p75NTR but not to TrkA or TrkC, J. Neurosci. Res. 80 (2005) 18–28. https://doi.org/10.1002/jnr.20432.
- [47] H. Koshimizu, K. Kiyosue, T. Hara, S. Hazama, S. Suzuki, K. Uegaki, et al., Multiple functions of precursor BDNF to CNS neurons: negative regulation of neurite growth, spine formation and cell survival, Mol. Brain. 2 (2009) 27. https://doi.org/10.1186/1756-6606-2-27.
- [48] J. Yang, L.C. Harte-Hargrove, C.J. Siao, T. Marinic, R. Clarke, Q. Ma, et al., proBDNF negatively regulates neuronal remodeling, synaptic transmission, and synaptic plasticity in hippocampus, Cell. Rep. 7 (2014) 796–806. https://doi.org/10.1016/j.celrep.2014.03.040.

- [49] M. Bergami, S. Santi, E. Formaggio, C. Cagnoli, C. Verderio, R. Blum, B. Berninger, M. Matteoli, M.Canossa, Uptake and recycling of pro-BDNF for transmitter-induced secretion by cortical astrocytes, J. Cell. Biol. 183 (2008) 213–21. https://doi.org/10.1083/jcb.200806137.
- [50] Peng S, Wuu J, Mufson EJ, Fahnestock M, Precursor form of brain-derived neurotrophic factor and mature brain-derived neurotrophic factor are decreased in the pre-clinical stages of Alzheimer's disease., J Neurochem. 2005 Jun;93 (6):1412-21. https://doi.org/ 10.1111/j.1471-4159.2005.03135.x
- [51] Fang W, Liao W, Zheng Y, Huang X, Weng X, Fan S, Chen X, Zhang X, Chen J, Xiao S, Thea A, Luan P, Liu J, Neurotropin reduces memory impairment and neuroinflammation via BDNF/NF-κB in a transgenic mouse model of Alzheimer's disease., Am J Transl Res. 2019 Mar;11(3):1541-1554.
- [52] H.S. Je, F. Yang, Y. Ji, G. Nagappan, B.L. Hempstead, B. Lu, Role of pro-brain-derived neurotrophic factor (proBDNF) to mature BDNF conversion in activity-dependent competition at developing neuromuscular synapses, Proc. Natl. Acad. Sci. 109 (2012) 15924–9. https://doi.org/10.1073/pnas.1207767109.
- [53] T. Matsumoto, S. Rauskolb, M. Polack, J. Klose, R. Kolbeck, M. Korte, Y.A. Barde, Biosynthesis and processing of endogenous BDNF: CNS neurons store and secrete BDNF, not pro-BDNF, Nat. Neurosci. 11 (2008) 131–3. https://doi.org/10.1038/nn2038.
- [54] A. Wysokiński, Serum levels of brain-derived neurotrophic factor (BDNF) and neurotrophin-3 (NT-3) in depressed patients with schizophrenia, Nord. J. Psychiatry. 70 (2016) 267–71. https://doi.org/10.3109/08039488.2015.1087592.
- [55] R. Yoshimura, T. Kishi, H. Hori, K. Atake, A. Katsuki, W. Nakano-Umene, A. Ikenouchi-Sugita, N. Iwata, J. Nakamura, Serum proBDNF/BDNF and response to fluvoxamine in drug-naïve first-episode major depressive disorder patients, Ann. Gen. Psychiatry. 13 (2014) 19. https://doi.org/10.1186/1744-859X-13-19.

- [56] M. Volosin, W. Song, R.D. Almeida, D.R. Kaplan, B.L. Hempstead, W.J. Friedman, Interaction of survival and death signaling in basal forebrain neurons: roles of neurotrophins and proneurotrophins, J. Neurosci. 26 (2006) 7756–66. https://doi.org/10.1523/JNEUROSCI.1560-06.2006.
- [57] R.S. Kenchappa, N. Zampieri, M.V. Chao, P.A. Barker, H.K. Teng, B.L. Hempstead, B.D. Carter, Ligand-Dependent Cleavage of the P75 Neurotrophin Receptor Is Necessary for NRIF Nuclear Translocation and Apoptosis in Sympathetic Neurons, Neuron 50 (2006) 219–32. https://doi.org/10.1016/j.neuron.2006.03.011.
- [58] C.J. Woolf, Dissecting out mechanisms responsible for peripheral neuropathic pain: Implications for diagnosis and therapy, Life Sci. 74 (2004) 2605-10. https://doi.org/10.1016/j.lfs.2004.01.003.
- [59] R.S. Hotchkiss, A. Strasser, J.E. McDunn, P.E. Swanson, Cell Death. N. Engl. J. Med. 361 (2009) 1570–83. https://doi.org/10.1056/NEJMra0901217.
- [60] M. Baniasadi, H. Manaheji, N. Maghsoudi, S. Danyali, Z. Zakeri, A. Maghsoudi, et al. Microglial-induced apoptosis is potentially responsible for hyperalgesia variations during CFA-induced inflammation, Inflammopharmacology. 2019. https://doi.org/10.1007/s10787-019-00623-3.
- [61] R.U. Ostrovskaya, Y.V. Vakhitova, U.S. Kuzmina, M.K. Salimgareeva, L.F. Zainullina, T.A. Gudasheva, V.A. Vakhitov, S.B. Seredenin, Neuroprotective effect of novel cognitive enhancer noopept on AD-related cellular model involves the attenuation of apoptosis and tau hyperphosphorylation, J. Biomed. Sci. 21 (2014) 74. https://doi.org/10.1186/s12929-014-0074-2.
- [62] V. Vorobyov, V. Kaptsov, G. Kovalev, F. Sengpiel, Effects of nootropics on the EEG in conscious rats and their modification by glutamatergic inhibitors, Brain Res. Bull. 85 (2011) 123–32. https://doi.org/10.1016/j.brainresbull.2011.02.011.
- [63] Ostrovskaya RU, Zolotov NN, Ozerova IV, Ivanova EA, Kapitsa IG, Taraban KV, Michunskaya AM, Voronina TA, Gudasheva TA, Seredenin SB, Noopept normalizes parameters of the incretin system in rats

with experimental diabetes, Bull Exp Biol Med. 157 (2014) 344-9. https://doi.org/10.1007/s10517-014-2562-5.

[64] Ostrovskaya RU, Romanova GA, Barskov IV, Shanina EV, Gudasheva TA, Victorov IV, Voronina TA, Seredenin SB, Memory restoring and neuroprotective effects of the proline-containing dipeptide, GVS-111, in a photochemical stroke model, Behav Pharmacol. 10 (1999) 549-53.

# **Figures**

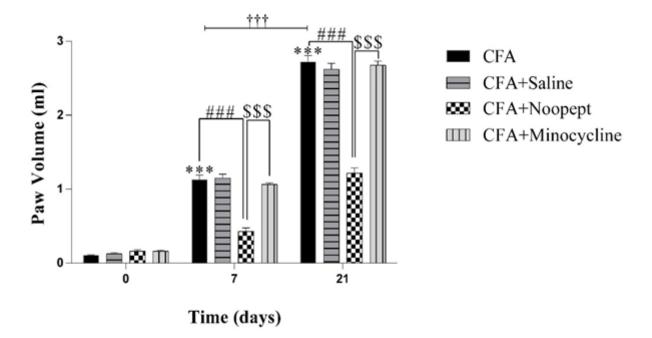


Figure 1

Paw volume variations during different stages of the study. Noopept treatment in CFA group significantly reduced paw edema during 21 days of the study. Data is presented as mean±SEM (n = 6/group). \*\*\*\*P<0.001 for comparison of paw volume variations on days 7 and 21 of the study with day 0 in CFA group. †††P<0.001 for comparison of paw volume variations on day 21 of the study with day 7 in CFA group. ###P<0.001 for comparing of paw volume variations between CFA and CFA+Noopept groups in the same days. \$\$\$P<0.001 for comparing paw volume variations between CFA+Noopept and CFA+Minocycline groups in the same days.

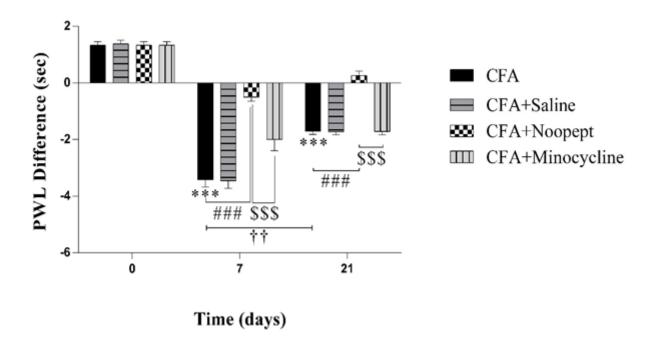


Figure 2

Thermal hyperalgesia varied in the injected paw during different stages of inflammation. Data is presented as mean±SEM (n = 6/group). \*\*\*P<0.001 for illustrating hyperalgesia variations on days 7th and 21st of the study in comparison with day zero in CFA group. ††P<0.01 for comparison of hyperalgesia variations on day 21st of the study with day 7 in CFA group. ###P<0.001 for comparing hyperalgesia variation between CFA and CFA+Noopept. \$\$\$P<0.001 for comparing hyperalgesia variation between CFA+Noopept and CFA+Minocycline treated rats in same days.

Figure 3A:

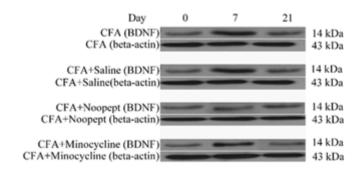


Figure 3B:

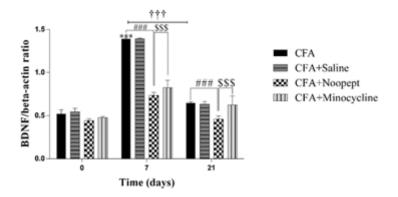


Figure 3

A: Immunoblots of spinal BDNF expression in different experimental groups. All densitometry data were demonstrated as BDNF/\(\beta\)-actin ratio. B: Noopept treatment could significantly decrease spinal BDNF expression on day 7th of the study. Data is presented as mean \(\pm\)SEM (n=6/group). \(\pm\)+\(\pm\)P<0.001 for comparison of spinal BDNF expression on day 21st of the study with day 7 in the CFA group. \(\pm\)##P<0.001 for comparison of spinal BDNF/\(\beta\)-actin protein band intensity between CFA and CFA+Noopept groups. \(\ps\)\$\(\pm\)\$P<0.001 for comparison of spinal BDNF/\(\beta\)-actin protein band intensity between CFA+Noopept and CFA+Minocycline groups.

Figure 4A:

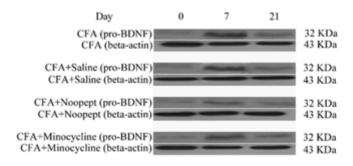


Figure 4B:

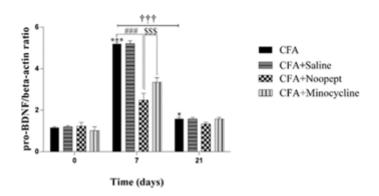


Figure 4

A: Immunoblots of spinal pro-BDNF expression in different experimental groups. All densitometry data were demonstrated as pro-BDNF/ß-actin ratio. B: Noopept administration caused significant decrease in spinal pro-BDNF expression in acute phase of the study. Data is presented as mean±SEM (n=6/group). \* P<0.05 and \*\*\* P<0.001 for comparison of spinal pro-BDNF expression in the CFA group on the 21st and 7th days of the study with day 0. †††P<0.001 for comparison of spinal pro-BDNF expression on day 21th of the study with day 7 in the CFA group. ###P<0.001 for comparison of spinal pro-BDNF/ \$\mathcal{B}\$-actin protein band intensity between CFA+Noopept groups. \$\$\$P<0.001 for comparison of spinal pro-BDNF/\$\mathcal{B}\$-actin protein band intensity between CFA+Noopept and CFA+Minocycline groups.

Figure 5A:

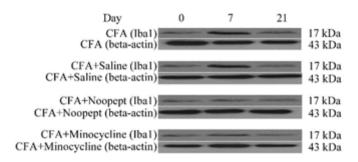


Figure 5B:

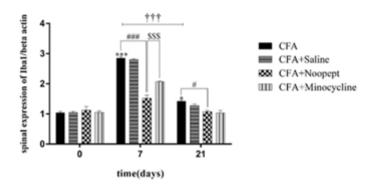


Figure 5

A: Immunoblots of spinal Iba1 expression in different experimental groups during different stages of study. All densitometry data were demonstrated as Iba1/\(\beta\)-actin ratio. B: Long term Noopept administration could change spinal Iba1 expression on days 7 and 21 of the study. Data is presented as mean \(\pm \)SEM (n=6/group). \*P<0.05 and \*\*\*P<0.001 for comparing spinal Iba1 expression on 21th and 7th days of the study with day 0 in the CFA group. †††P<0.001 for comparing spinal Iba1 expression on day 21 with day 7 in the CFA group. #P<0.05 and ### P<0.001 for comparing spinal Iba1/\(\beta\)-actin band intensity between CFA and CFA+Noopept groups. \$\$\$P<0.001 for comparing spinal Iba1/\(\beta\)-actin band intensity between CFA+Noopept and CFA+Minocycline groups.

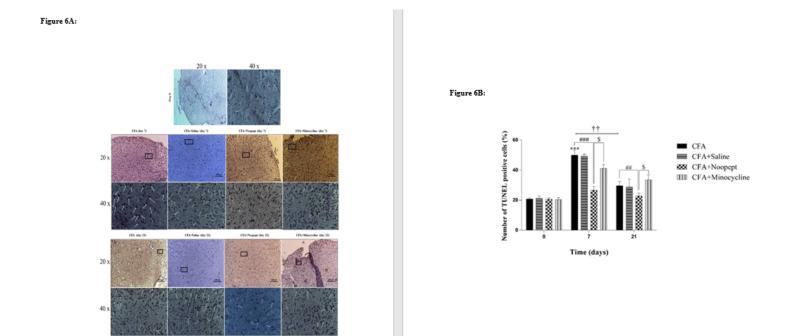


Figure 6

A: Effect of Noopept administration on lumbar spinal cord tissue apoptosis during different phases of inflammatory pain. Arrowheads indicate positive cells. B: Treatment with Noopept following CFA injection could decrease the percentage of TUNEL positive cells in the CFA+Noopept group compared with CFA group. Five non-overlapping zones were randomly selected. The positive cells were counted and an apoptotic index (AI) was calculated depend on following formula: AI (%)=number of TUNEL-positive cells/total number of cells × 100. Data is presented as mean±SEM (n=6 rats/group, per each time point).

\*\*\*P<0.001 for comparison of TUNEL positive cells between days 0 and 7 in the CFA group. ††P<0.01 for comparison of TUNEL positive cells between days 7 and 21 in the CFA group. ##P<0.01 and ###P<0.001 for comparison of TUNEL positive cells between CFA and CFA+Noopept groups. \$P<0.05 for comparison of TUNEL positive cells between CFA and CFA+Noopept groups.

Figure 7A:

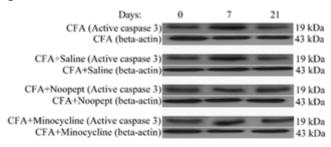


Figure 7B:

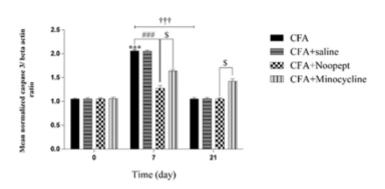


Figure 7

A: Immunoblots of cleavage of spinal caspase-3 in different experimental groups during different stages of study. All densitometry data were demonstrated as Caspase-3 /ß-actin ratio. B: Noopept administration reduced cleavage of spinal caspase-3 following CFA-induced inflammation. Data is presented as mean±SEM (n=6/group). \*\*\*P<0.001: for comparison of cleavage of spinal caspase-3 in the CFA group on the 7th day of the study with day 0. †††P<0.001 for comparing cleavage of spinal caspase-3 on day 21 with day 7 in the CFA group. ###P<0.001 for comparison of cleavage of spinal caspase-3/ B-actin band intensity between CFA and CFA+Noopept groups. \$P<0.05 for comparison of cleavage of spinal caspase-3/B-actin band intensity between CFA+Noopept and CFA+Minocycline groups.