

A Novel De Novo Ddx3x Missense Variant in a Female With Brachycephaly and Intellectual Disability: A Case Report

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Case report

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Abstract

Background: *De novo* pathogenic variants in the *DDX3X* gene are reported to account for 1–3% of unexplained intellectual disability (ID) in females, leading to the rare disease known as *DDX3X* syndrome (MRXSSB, OMIM #300958). Besides ID, these patients manifest a variable clinical presentation, which includes neurological and behavioral defects, and abnormal brain MRIs.

Case presentation: We report a 10-year-old girl affected by delayed psychomotor development, delayed myelination, and polymicrogyria (PMG). We identified a novel *de novo* missense mutation in the *DDX3X* gene (c.C625G) by exome sequencing. The *DDX3X* gene encodes a DEAD-box ATP-dependent RNA-helicase broadly implicated in gene expression through regulation of mRNA metabolism. The identified mutation is located just upstream the helicase domain and is suggested to impair the protein activity, thus resulting in the altered translation of *DDX3X*-dependent mRNAs. The proband, presenting with the typical PMG phenotype related to the syndrome, does not show other clinical signs frequently reported in presence of missense *DDX3X* mutations that are associated with a most severe clinical presentation. In addition, she has brachycephaly, never described in female *DDX3X* patients, and macroglossia, that has never been associated with the syndrome.

Conclusions: This case expands the knowledge of *DDX3X* pathogenic variants and the associated *DDX3X* syndrome phenotypic spectrum.

Background

About 1–3% of females with unexplained intellectual disability (ID) have *de novo* pathogenic variants in the X-linked *DDX3X* gene (Xp11.4) [1]. Up to about 300 cases have been identified [2], both females with X-linked dominant inheritance, and very few males showing an X-linked recessive pattern of inheritance (Snijders Blok et al., 2015).

Patients with the rare *DDX3X* syndrome (MRXSSB, OMIM #300958) show a variable clinical presentation with different degrees of ID and/or developmental delay, neurological and behavioral defects, including microcephaly, hypotonia, epilepsy, movement disorders, autism spectrum disorder and aggressiveness. Brain malformations are also reported, and include corpus callosum hypoplasia, ventricular enlargement, and polymicrogyria. Additional clinical features, among which facial dysmorphisms and sensory deficits, may also be present.

To date, all reported pathogenic *DDX3X* variants identified in affected females are *de novo* loss of function, leading to haploinsufficiency and probably to embryonic lethality in males. Most of *DDX3X* mutations reported in males are maternally inherited missense variants. In these families, males showed borderline to severe ID and carrier females were unaffected and expressed a *DDX3X* protein retaining a partial functionality [1]. Several hypotheses have been evaluated to explain the gender-specific pathogenicity, including a skewing X-inactivation pattern, but no definitive conclusions have been drawn. Kellaris et al.[3] proposed that hypomorphic *DDX3X* variants may be viable in hemizygous males and do

not cause clinical phenotype in female carriers. However, few *de novo* *DDX3X* mutations have been identified in males [4]. Although these male patients share many of the clinical features with affected females, some distinct clinical phenotypes can depend on the gender of the patient and on the pathogenicity of the variant.

These findings suggest that *DDX3X* is dosage sensitive and might have a differential activity in females and males. Indeed, Snijders Blok et al.[1] reported a family with recurrent miscarriages and a male viable pregnancy, terminated after the identification of severe congenital anomalies by ultrasound imaging. By exome sequencing (ES) a missense mutation in the *DDX3X* gene was detected in the fetus, suggesting a potential germline mosaicism in the mother.

The *DDX3X* gene encodes a DEAD-box ATP-dependent RNA-helicase broadly implicated in gene expression through regulation of mRNA metabolism [5]. It acts as a translational regulator of target mRNAs with highly structured 5' untranslated regions (UTRs) [6] and it is involved in repeat-associated non-AUG translation [7]. *DDX3X* also takes part in stress response and stress granule assembly, innate immune signaling, mitotic chromosome segregation and it can also exert a role in tumorigenesis [8]. In addition, it is thought to be an essential factor in the RNA-interference pathway [9], and it is a key regulator of the Wnt/ β -catenin pathway [10].

Despite the ubiquitous expression of the *DDX3X* protein, high expression levels are detected in all the cortical layers of the embryonic brain, consistent with its crucial role in cortical development during neurogenesis [10] and, as a consequence, *DDX3X* mutations are reported to impact on neuronal function [4]. In particular, defective neurite outgrowth and neural progenitor differentiation/migration could account for brain malformations and the consequent characteristic clinical phenotypes of the *DDX3X* syndrome [11].

Here, we report a 10-year-old girl with delayed psychomotor development, delayed myelination, bilateral frontal polymicrogyria (PMG) and thin body and splenium of the corpus callosum, suggestive of the *DDX3X* syndrome. WES allowed the identification of the novel *de novo* *DDX3X* mutation c.C625G (p.H209D), thus expanding the number of *DDX3X* pathogenic variants and their associated phenotypic spectrum.

Case Presentation

The proband was referred to our laboratory by the pediatric geneticists of our Institution, Fondazione IRCCS Ca' Granda Ospedale Policlinico (Milan). Appropriate written informed consent was obtained from all family members.

Patient II-3 (Fig. 1A), who is currently 10 years old, is the third-born of healthy parents with a doubtful distant consanguinity (beyond the second generation). The two older siblings (II-1 and II-2, of 16 and 14 years old respectively) are both healthy. Between the first and second pregnancy three miscarriages occurred, but no additional information is available. The family history is negative for genetic conditions.

The patient was born at 38 + 5 gestational weeks, after a normal pregnancy, characterized by regular growth, morphology and fetal movements. She was born by cesarean delivery scheduled for previous cesarean section.

Clinical signs are reported in Table 1. At birth, the child was 2740 g (25th centile), 49 cm long (50th centile), presented an OFC of 34 cm (50th centile) and an Apgar score of 10–10. Absence of skin adnexa (eyelashes, eyebrows and nails) and difficult scarring of the umbilical stump are referred. In addition, a bilateral II-III toe syndactyly and a small left preauricular tag were evident. Ultrasounds revealed normal abdominal and cerebral morphology.

Table 1

Clinical features of II-3 compared to the characteristic clinical signs reported for the *DDX3X* syndrome and relative prevalence.

<i>DDX3X</i> syndrome clinical signs (relative prevalence)	Clinical signs of II-3
Development	
Developmental delay (106/106) [10]	+
Intellectual disability (106/106) [10]	+
Language delay (38/75) [10]	+
Growth	
Failure to thrive (13/44) [13]	+
Short stature (1/6) [13]	-
Microcephaly (39/107) [13]	-
Brachycephaly (3/11) [13]	+
Neurologic/behavioral	
Seizures (24/116) [10]	EEG anomalies
Hypotonia (66/116) [10]	+
Hypertonia/spasticity (9/78) [13]	-
Mixed hypo and hypertonia (31/93) [10]	-
Sleep disturbance (2/6) [13]	+
Movement disorders/spasticity in the legs (22/49) [13]	-
Behavior disorders/autism spectrum disorder/aggression (24/49) [13]	-
Hyperreflexia (9/78) [13]	+
Brain MRI	
Polymicrogyria (11/89) [10]	+ (anterior)
Corpus callosum hypoplasia/agenesis (76/105) [13]	+
Ventricular enlargement (27/105) [13]	-
Key-hole shaped temporal horns (32/89) [10]	-
Colpocephaly (3/89) [10]	-
Delayed myelination/decreased cortical white matter (50/89) [10]	+

Abbreviations: ND: not defined

DDX3X syndrome clinical signs (relative prevalence)	Clinical signs of II-3
Small pons (11/89) [10]	-
Small inferior vermis (6/89) [10]	-
Sensory	
Vision problems (strabismus, coloboma, astigmatism, nystagmus) (29/92) [10]	+
Hearing problems (11/114) [13]	-
Facial dysmorphisms	
Short/down-slanting palpebral fissure length (2/6) [13]	-
Hypertelorism/telecanthus (6/36) [13]	+
Epicanthal folds (1/6) [13]	-
Elongated/flattened/triangular face (9/36) [13]	+
High/broad forehead (8/36) [13]	+
Wide nasal bridge/bulbous tip (9/36) [13]	+
Short/narrow nose, anteverted nares (11/36) [13]	-
Micrognathia (2/6) [13]	+
High arched palate (4/6) [13]	ND
Thin upper lip (4/6) [13]	+
Low set/protruding/wide ears (2/6) [13]	+
Smooth/long philtrum (3/6) [13]	-
Cleft lip/palate (3/44) [13]	-
Macroglossia (ND)	+
Other	
Congenital cardiac defects (13/90) [10]	-
Precocious puberty (11/94) [10]	-
Feeding difficulties (gastro-esophageal reflux/swallowing) (3/6) [13]	+
Joint hyperlaxity (14/44) [13]	-
Scoliosis (15/94) [10]	-

Abbreviations: ND: not defined

<i>DDX3X</i> syndrome clinical signs (relative prevalence)	Clinical signs of II-3
Malformations of the hands (1/6) [13]	+
Skin pigmentation anomalies (16/44) [13]	-
Loss/reduced subcutaneous fat (2/6) [13]	+
<i>Abbreviations: ND: not defined</i>	

In the first months of life, a severe gastro-esophageal reflux with initial weight loss and sleep disturbance was reported, which improved after the introduction of proton-pump inhibitors therapy.

Growth was regular, but psychomotor development was severely delayed, with independent walking and babbling acquired towards 6 years of age.

Evaluation at 6 years old revealed the following cranio-facial dysmorphisms: brachycephaly and a flattened-triangular-asymmetrical face characterized by micrognathia, mild hypertelorism, wide and prominent nose, short philtrum, thin lips and macroglossia. She also presented a slight facial grimacing, large and anteverted ears and small left ear tag. Growth parameters were normal, with a weight of 16.5 kg (10-25th centiles), a height of 111 cm (50-75th centile), head circumference of 50.5 cm (25-50th centile).

Brain MRI showed delayed myelination (improved at the second MRI), bilateral frontal polymicrogyria (PMG) and thin body and splenium of the corpus callosum. Electroencephalogram (EEG) showed sharp-wave anomalies on the rear right regions. She also presents hyperopia and divergent strabismus.

Karyotype, aCGH and mutational analysis of the *ADGRG1* gene (associated to bilateral frontoparietal polymicrogyria) resulted normal. An NGS panel for Rett syndrome (including the *MECP2*, *CDKL5*, *FOXP1*, *MEF2C*, *SCN1A*, *UBE3A*, *PCDH19*, *STXBP1* genes) was also performed, and no pathogenic variants were identified.

Since no other clinical suspicions were hypothesized, the proband and her parents underwent ES on DNA extracted from peripheral blood leukocytes. Trio ES and variants interpretation according to the pedigree and the clinical features allowed the identification of the novel missense variant c.C625G (p.H209D) in exon 7 of the *DDX3X* gene. No other candidate variants were found, and the identified variant was confirmed by Sanger sequencing. The absence of the mutation in the healthy mother confirmed the hypothesis of a *de novo* inheritance (Fig. 1B), as reported for all *DDX3X* mutations found in females [1]. However, considering the occurrence of three miscarriages between the first and the second pregnancy, we cannot exclude the possibility that also the aborted fetuses could have carried the c.C625G mutation as a consequence of a possible germline mosaicism in the mother. This is consistent with the association of *DDX3X* mutations to miscarriage recurrence and aborted male fetuses [1].

The identified *DDX3X* variant is neither reported in ClinVar nor in HGMD Professional 2020.1. To determine the impact of this amino acid substitution, the pathogenic score was evaluated by thirteen

different *in-silico* predictors, and it was predicted to be damaging by nine of them (Supplementary Table 1). This score is consistent with the amino acid substitution from a positively charged histidine to a negatively charged aspartic acid, in a highly conserved residue.

Appropriate written informed consent was obtained from all family members.

Discussion And Conclusions

The novel c.C625G variant involves the amino acidic position 209, which maps 2 bp upstream the helicase ATP binding domain (Fig. 1C). Most reported *DDX3X* mutations map within the two helicase domains impairing the protein helicase activity, with a dominant negative mechanism[10]. The H209D variant, as other missense mutations that fall in proximity of the helicase domain, such as the V206M already reported in a patient with the *DDX3X* syndrome [10], is suggested to impair the protein helicase activity. Deficient *DDX3X* helicase activity results in the enzyme's inability to release the bound RNA after ATP hydrolysis and the altered translation of *DDX3X*-dependent targets, in particular mRNAs containing highly structured 5' UTRs and/or high GC content [6, 12]. This leads to the sequestration of RNAs and RNA binding proteins, and the formation of aberrant ribonucleoprotein (RNP) granules containing newly synthesized proteins and/or stalled polysomes [10]. The defective translation of *DDX3X*-target transcripts has been proposed to lead to impaired neurogenesis, likely affecting embryonic cortical development [10]. This mechanism may also account for the more severe phenotypes associated with missense mutations, compared to nonsense and frameshift (presumed loss of function) lesions, that affect translation of some targets but do not induce granule formation, resulting in a less severe clinical presentation [10]. This pathomechanism may also apply to the H209D mutation here reported, as suggested, not only by the affected protein domain, but also by the PMG phenotype of the proband.

According to the recent studies on genotype-phenotype correlations in the *DDX3X* syndrome and underlying pathogenetic mechanisms, patients with severe cerebral defects, including complete or partial absence of the corpus callosum and PMG (frequently associated to microcephaly), presented missense mutations or single amino acid deletion hotspot mutations, including the nearby V206M (Fig. 1C). These mutations are also associated to more complex clinical presentations, including epilepsy, autism spectrum disorder, severe intellectual disability, and cardiovascular anomalies, not present in patients without PMG. These evidences thus suggest that these variants are likely to lead to a more severe dominant negative phenotype, while loss of function mutations are associated to a milder spectrum of clinical traits with no sign of PMG. Despite the proband presents with PMG, she does not show other clinical traits reported in most severe cases of *DDX3X* syndrome, such as microcephaly, congenital cardiac defects and epilepsy (she only presents EEG anomalies) (Table 1). In addition, she shows reduced subcutaneous fat, already reported in two other patients [13], and she is the first female patient presenting with brachycephaly (which has so far been described only in few males [1]) (Table 1). She also has macroglossia, an additional clinical sign that has never been associated with the *DDX3X* syndrome before (Table 1).

In conclusion, these distinctive clinical features, together with the presence of PMG, but no other signs associated to a severe phenotype, furtherly expand genotype-phenotype correlations of *DDX3X* missense mutations.

This case report emphasizes the clinical utility of WES in ending the diagnostic odyssey of individuals with unexplained ID.

Abbreviations

DDX3X

DEAD-Box Helicase 3 X-Linked; ID:intellectual disability; MRXSSB:Mental retardation, X-linked 102;

PMG:polymicrogyria; ES:exome sequencing; MRI:magnetic resonance imaging;

EEG:electroencephalogram; aCGH:array-Comparative Genomic Hybridization

Declarations

Ethics approval and consent to participate

This work is part of the “CARE: Challenging Approaches to undiagnosed Rare diseases with exome sequencing” Project (Protocol code: PED-CARE-2018).

Appropriate written informed consent was obtained from all family members.

Consent for publication

Appropriate written informed consent was obtained for publication.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Competing interests

The authors declare that they have no competing interests.

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Authors' contributions

GM: WES data analysis and manuscript drafting; JC: bioinformatic analysis; CS: performed WES experiments; OR: WES data analysis and interpretation; FG, EP, SO, DM, PM: clinical evaluation and follow-up of the patient; IC: coordinated clinical data collection; LF, MM: supervised the project and contributed to the interpretation of the results. All authors read and approved the final manuscript.

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Figures

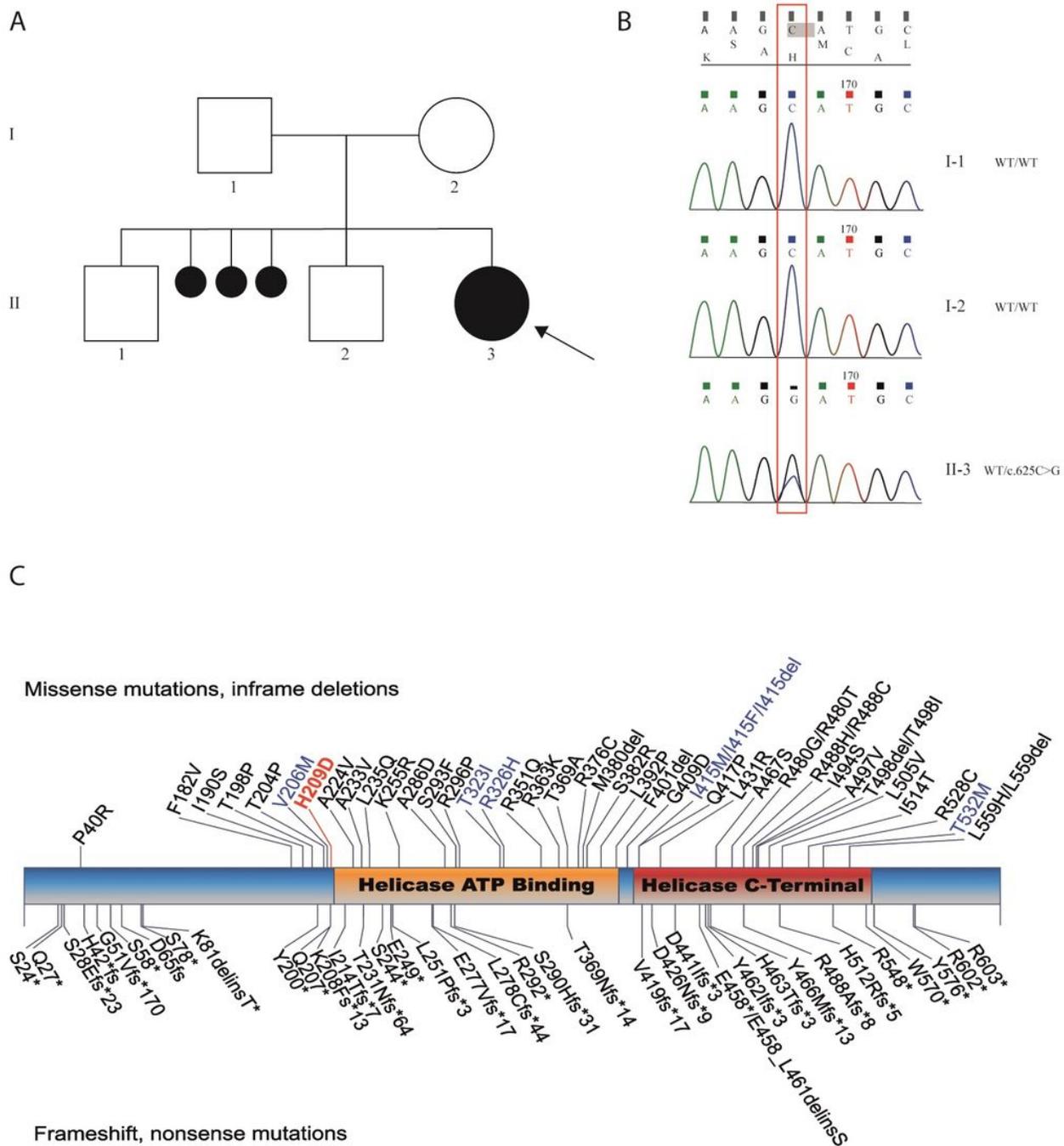


Figure 1

A) Pedigree of the family. B) Sequencing chromatograms of DNA samples from the proband and the parents. The position of the novel DDX3X variant identified is indicated by a red box. The proband is confirmed to be heterozygous for the DDX3X c.625C>G variant, while parents are wild-type. C) Schematic representation of the DDX3X protein domains and mapping of mutations already reported in patients affected by the DDX3X syndrome (adapted from [10]). Missense mutations and in-frame deletions are

reported on top, while frameshift and nonsense mutations are annotated on the bottom. Mutations reported in patients with PMG are displayed in blue. The H209D variant identified in II-3 is reported in red.

Supplementary Files

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