The evolutionary landscape of primate longevity

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35

Abstract:

Is it possible to slow the rate of aging, or do biological constraints limit its plasticity? We test this ‘invariant rate of aging’ hypothesis with an unprecedented collection of 39 human and nonhuman primate datasets across seven genera. We first recapitulate, in nonhuman primates, the highly regular relationship between life expectancy and lifespan equality seen in humans. We next demonstrate that variation in the rate of aging within genera is orders of magnitude smaller than variation in pre-adult and age-independent mortality. Finally, we demonstrate that changes in the rate of aging, but not other mortality parameters, produce striking, species-atypical changes in mortality patterns. Our results support the invariant rate of aging hypothesis, implying biological constraints on how much the human rate of aging can be slowed.

Introduction:

The highest recorded human life expectancy has increased since the mid-1800s by approximately 3 months per year¹. These gains have resulted from shifting the majority of deaths from early to later and later ages, rather than from slowing the rate at which mortality increases with age (i.e., the ‘rate of aging’)². Further substantial extensions of human longevity will depend on whether it is possible to slow the rate of aging or otherwise reduce late life mortality. Consequently, the nature of biological constraints on aging is a central problem in the health sciences and, because of its implications for demographic patterns, is also of long-standing interest in ecology and evolutionary biology.

Across species, rates of aging are strongly correlated with other aspects of the life history—pre-adult mortality, age at first reproduction, birth rate, metabolic rate and generation time—as well as with morphological traits such as body size and growth rate^{3,4}. These correlations suggest that aging evolves in concert with a suite of other traits, which may produce constraints on the rate of aging within species. Indeed, researchers have long hypothesized that the rate of aging is relatively fixed within species, not only in humans but also other animals⁵⁻⁷.

This ‘invariant rate of aging’ hypothesis has received mixed support. Several studies have documented a strong phylogenetic signal in the rate of aging across multiple species of birds and mammals, suggesting strong biological constraints and little within-species variance in this rate^{7,8}. Furthermore, Bronikowski and colleagues⁹ observed greater variation in initial adult mortality than in the rate of aging across several populations of baboons. On the other hand, across multiple mammal species, measurable differences in the rate of aging have been documented between populations in different environments (e.g., zoo versus wild¹⁰).

Understanding the nature and extent of biological constraints on the rate of aging and other aspects of age-specific mortality patterns is critical for identifying possible targets of intervention to extend human lifespans, and for understanding the evolutionary forces that have shaped lifespans within and across species. Although no consensus has been reached about the invariant rate of aging hypothesis, further evidence that biological constraints may shape human aging comes from the remarkably consistent relationship between life expectancy at birth (e_0) and lifespan equality (ϵ_0) in an extremely diverse set of human populations^{11,12}. While life expectancy at birth (a measure of the ‘pace’ of mortality¹³) describes the average lifespan in a

population, lifespan equality (a measure of the ‘shape’ of mortality ¹³) describes the spread in the distribution of ages at death in a population (see also ^{14,15}). Lifespan equality is highly correlated with other measures of the distribution of ages at death, such as the coefficient of variation and the Gini coefficient, often used to measure economic inequality ¹¹. The distribution of ages at death tells us whether the risk of death is evenly distributed across the range of observed lifespans, or is concentrated around certain ages. For instance, if deaths are evenly distributed across age classes or show multiple modes, the result is high lifespan variance and low lifespan equality, while if deaths are concentrated at the tail-end of the lifespan distribution (as in most developed nations), the result is low lifespan variance and high lifespan equality. The extremely tight positive relationship between life expectancy (e_0) and lifespan equality (ϵ_0) across an enormous range of human populations indicates strong but poorly understood constraints underlying variation in human mortality ^{2,11}.

Understanding the biological constraints on aging requires mortality data for multiple populations of nonhuman species, as well as for humans. However, data from multiple populations of nonhuman animals are rarely available, making it difficult to unveil the forces underlying mortality differences within versus between species. The challenge is particularly acute for long-lived species, including nonhuman primates, the closest relatives of humans. Nonetheless, these are precisely the species that will shed most light on how biological constraints have shaped the evolution of aging within the lineage leading to humans.

To better understand biological constraints on aging, we sought to answer two questions. First, is the highly regular linear relationship between life expectancy and lifespan equality in humans also evident in other primates? Second, if so, do biological constraints on aging underlie this highly regular relationship? To address these questions, we assembled an unprecedented dataset on age-specific mortality rates in multiple populations of several different primate genera. Our combined dataset includes data from both wild and captive primate populations. The data from wild populations consist of individual-based birth and death data on males and females from 17 continuous long-term studies of wild primate populations representing 6 genera distributed across the order Primates, and include Old World monkeys (2 genera), New World monkeys (1 genus), great apes (2 genera, both African), and an indriid (1 genus, endemic to Madagascar) (Supplementary Table S1). For those same genera we obtained individual-based birth and death data from 13 species in zoos from Species360’s Zoological Information Management System (ZIMS) ¹⁶ (see Methods, Supplementary Table S1). We also included data on a 7th primate genus, *Homo*, using male and female human mortality data from nine of the human datasets studied by Colchero and colleagues ¹¹, specifically populations that had not benefited from modern advances in public health, medicine and standards of living, which allowed us to carry out the most salient comparisons with nonhuman primates. The Human Mortality Database ¹⁷ yielded life tables for **1)** Sweden from 1751-1759, **2)** Sweden in 1773, **3)** Sweden from 1850-1859, **4)** and Iceland in 1882. We included additional human life tables for **5)** England from 1600-1725 ¹⁸, **6)** Trinidad from 1813-1815 ¹⁹, **7)** Ukraine in 1933 ²⁰ and two hunter gatherer populations, **8)** the Hadza, based on data collected between 1985 and 2000 ²¹ and **9)** the Ache during the pre-contact period of 1900-1978 ²². In the aggregate, our 39 combined datasets (17 wild and 13 zoo nonhuman primates, and 9 human populations; Supplementary Table S1) comprise a taxonomically diverse sample of primates and represent considerable environmental variability within genera, maximizing the probability of detecting variation in aging.

To understand potential constraints on primate aging, we compared age-specific changes in the risk of death across multiple populations of each genus. The age-specific risk of death, often described by a hazard rate, is the basic building block of the distribution of ages at death, and therefore determines both life expectancy and lifespan equality for a population. Among most mammal species, the risk of death is high in infancy, rapidly declines during the immature period, remains relatively low until early adulthood and then rises with age as a result of senescence. This pattern can be described mathematically by the five-parameter Siler function²³, given by

$$\mu(x) = \exp(a_0 - a_1x) + c + \exp(b_0 + b_1x), \text{ for } x \geq 0$$

where a_0 , a_1 , c , b_0 , b_1 are mortality parameters, each of which governs different stages of the age-specific mortality. In short, parameters a_0 and a_1 drive infant and juvenile mortality, c is commonly described as the age-independent mortality, and b_0 and b_1 control senescent mortality. Parameters a_0 , c and b_0 are scale parameters, while a_1 determines the speed of decline in infant and juvenile mortality and b_1 determines the rate of increase in adult and senescent mortality, analogous to the rate of senescence or rate of aging. We first fitted Siler models of age-specific mortality for males and females for each of the 30 non-human primate populations (Methods, Supplementary Tables S1 and S2). We then examined how each of the five Siler parameters varied within and between the genera (Supplementary Figs. S1 and S2). We also calculated sex-specific values for life expectancy at birth (e_0) and lifespan equality (ϵ_0) in each population, and used these values to examine the relationship between life expectancy and lifespan equality within each genus (Supplementary Table S3). We conducted genus-level rather than species-level analyses because restricting ourselves to the species level severely restricted the availability of individual-based datasets (e.g., among guenons, only one or two individual-based datasets were available for each species, while examining the genus provided five such datasets).

Results

Age-specific mortality across populations and life expectancy-lifespan equality relation. Our regression analyses yielded clear linear relationships between e_0 and ϵ_0 within each primate genus, mirroring the relationship observed within humans (Fig. 1A, and B and Fig S3). This pattern emerged despite considerable variation in age-specific mortality, in the distribution of ages at death, and in the Siler mortality parameters among populations of each genus (Supplementary Figs. S1-S4, Supplementary Table S2). The slopes of these regression lines were statistically significant (i.e., p -value < 0.05) in 5 of 7 genus-level datasets for females and in 4 of 7 for males (Fig. 1A, 1B, Supplementary Table S4); the regression lines did not reach statistical significance in analyses that included relatively few populations or that included small or heavily censored datasets. The slopes of the regression lines were statistically significantly different than the slope of the line for humans in female sifaka, baboons, guenons, and gorillas, and in male guenons, gorillas, and chimpanzees.

Drivers of the linear relationship between life expectancy and lifespan equality. Having confirmed that the relationship between life expectancy and lifespan equality is linear and highly regular within other primate genera, as it is in humans, we next sought possible causes for this regularity. Specifically, we asked which Siler mortality parameters best explain variation among populations in life expectancy and lifespan equality, and therefore which have a disproportionately large effect on the slopes of the regression lines. To pursue this question, we

initially conducted a sensitivity analysis by simulating independent changes in each of the Siler mortality parameters (Fig. 1C) and graphically examining the effects of these changes on the life expectancy-lifespan equality relationships. Specifically, we varied one Siler parameter at a time within each genus, keeping the other four Siler parameters constant at the value found at the midpoint of the regression line.

This approach produced striking results: within each genus, simulated variation in pre-adult mortality (captured by Siler parameters a_0 and a_1) and in age-independent mortality (Siler parameter c) all produced lines of similar direction to the observed regression lines (Fig. 1D). That is, within the observed range of e_0 values, changes in these three Siler parameters resulted in ε_0 similar to the observed range. Therefore, consistent with theory and with the long-understood effect of averting early deaths, observed variation in life expectancy and lifespan equality within each primate genus appears to be largely accounted for by variation in the pattern of early deaths, and very little by actuarial senescence.

In stark contrast, simulated variation in the rate-of-aging parameter (Siler parameter b_1) produced lines with conspicuously different direction from the observed regression lines. Specifically, changing b_1 moved the life expectancy - lifespan equality values away from the regression lines (Fig. 1D).

Sensitivity of life expectancy and lifespan equality to mortality parameters. These findings led us to postulate that, while variation in early deaths is the primary cause of observed variation in life expectancy and lifespan equality within each genus, changes in the rate of aging in one or more populations in a genus could shift those populations towards the lines of other genera. To further investigate this possibility, we derived mathematical functions for the sensitivity of life expectancy and lifespan equality to changes in any given mortality parameter (see Supplementary Text). These sensitivity functions allowed us to obtain precise measures of the amount of change in life expectancy and lifespan equality for a unit change in any given mortality parameter at any point in the life expectancy-lifespan equality landscape (including along each of the regression lines).

The resulting vectors of change (Fig. 2A) are consistent with our graphical exploration, and they also revealed the relative magnitudes of changes that each mortality parameter produces in the life expectancy-lifespan equality landscape (Fig. 2B). Specifically, a unit change in the rate of aging parameter b_1 shifts the life expectancy and lifespan equality values in a direction almost perpendicular to the regression lines, and the magnitude of that change is disproportionately large compared to the other four parameters. We then calculated the degree of collinearity (how parallel versus perpendicular two vectors are) between the seven genera-specific regression lines for females and the vectors of change for each parameter. We found that the two parameters that govern infant mortality, a_0 and a_1 , and the age-independent parameter c , produce vectors of change that are almost parallel to the regression lines. In contrast, Siler parameter b_0 produces vectors that are intermediate between parallel and perpendicular, while the rate-of-aging parameter, b_1 , produces vectors that are almost perpendicular to the regression lines (Fig. 2C). In short, changes in pre-adult mortality and in age-independent mortality tend to move a population along the regression line typical of its genus. In contrast, changes in the aging parameters, b_0 and particularly b_1 , will shift a population away from this line, into the space occupied by other genera in the landscape.

Amount of change in each mortality parameter along the genus lines. If variation in pre-adult and age-independent mortality parameters account for most of the within-genus differences in life expectancy and lifespan equality, we expect the parameters that control infant and age-independent mortality to be much more highly sensitive to perturbations of e_0 and a_0 than the parameters that control adult and senescent mortality, particularly b_1 . To test these expectations, we quantified the relative change in each parameter along each genus line by calculating the partial derivatives of the log-transformed parameter with respect to changes in e_0 and a_0 ; see Supplementary Text). We then calculated path integrals of these sensitivities along each genus line in order to quantify the total amount of change in each parameter for all seven genera. We found that, in agreement with our previous results, in all cases the parameters that govern infant and age-independent mortality changed orders of magnitude more than those that drive adult and senescent mortality (Fig. 3).

Discussion

Our results provide the most comprehensive support to date for the idea that observed variation in mortality patterns among populations of a given genus is driven largely by changes in pre-adult mortality: previous support for this idea comes from studies of just one or a few species, typically including humans or primarily captive animal populations^{2,5,8,10}. Notably, recent research on human populations² shows that increases in life expectancy can occur not just through decreases in pre-adult mortality but also through decreases in adult mortality, specifically through reductions in the b_0 parameter. This possibility is supported by our result that the vectors of change for Siler parameter b_0 produced by our sensitivity analysis are markedly less colinear with our genus-specific regression lines than the vectors of change for the pre-adult mortality parameters (Fig. 2C).

More strikingly, our results provide fresh insight into the ‘invariant rate of aging’ hypothesis. In support of that hypothesis, we find that, within primate genera, rates of aging (captured by Siler parameter b_1) do indeed vary across populations, but along each genus line they vary orders of magnitude less than other mortality parameters. Further, our results illustrate that, within any given genus, large changes in the rate of aging would shift a population across the life expectancy-lifespan equality landscape to a position closer to other genera. This result supports the ‘invariant rate of aging’ hypothesis, although it does not rule out heterogeneity among individuals within a population in rate of aging. More importantly, it implicates changes in the rate of aging as a likely source of variation in lifespan between distantly related taxa⁶.

Furthermore, by considering populations exposed to a wide range of environmental conditions—from high predation and low resource availability, to unconstrained resources and veterinary care in zoos—our results have implications both for life history theory and for conservation. Life history theory predicts that among species with slow life histories (i.e., long lifespans, small litters and delayed maturity), adult survival should be buffered from environmental variability, while juvenile survival is expected to vary widely in response to the environment²⁴⁻²⁷. Our findings support this buffering hypothesis, in that the most dramatic observed changes in life expectancy occur because of changes in juvenile survival, while changes in adult or senescent survival account for relatively little of the observed variation within each genus.

Importantly, sufficient demographic information to understand and predict population dynamics exists for less than 1.5% of extant vertebrate species²⁸. By unravelling the interdependence of mortality parameters within a species or genus, we can contribute to filling these glaring demographic knowledge gaps and further our understanding of the ecology and evolution of a wide range of animal species, as well as the conservation of species worldwide.

Finally, on the question of whether humans can slow our own rate of aging, our findings support the idea that environmentally-influenced infant and age-independent mortality improvements were the central contributor to the decades-long trend towards longer human life expectancies and greater lifespan equality, when life expectancies and lifespan equality were low². Since the middle of the 20th century, however, declines in the baseline level of adult mortality, b_0 , have played an increasingly important role^{2,6}. As we show here, improvements in the environment are unlikely to translate into a substantial reduction in the rate of aging, or in the dramatic increase in lifespan that would result from such a change. It remains to be seen if future advances in medicine can overcome the biological constraints that we have identified here, and achieve what evolution has not.

Methods

Data for non-human primates. We obtained 30 datasets for six genera of non-human primates: sifaka (*Propithecus spp*), gracile capuchin monkey (*Cebus spp*), guenon (*Cercopithecus spp*), baboon (*Papio spp*), gorilla (*Gorilla spp*), and chimpanzee (*Pan troglodytes*) (Extended Data Table 1). Of these, 17 datasets correspond to long-term projects in the wild, while 13 were contributed by the non-profit Species360 from the Zoological Information Management System (ZIMS, Data Use Approval Number RR5-2019)¹⁶, which is the most extensive database of life history information for animals under human care.

Survival analysis. To estimate age-specific survival for all the wild populations of non-human primates, we modified the Bayesian model developed by Colchero et al¹¹ and Barthold et al²⁹. This model is particularly appropriate for primate studies that follow individuals continuously within a study area and when individuals of one or both sexes can permanently leave the study area (out-migration), while other individuals can join the study population from other areas (in-migration). Thus, it allowed us to make inferences on age-specific survival (or mortality) and on the age at out-migration.

We define a random variable X for ages at death, with observations $x \geq 0$. The model requires defining a hazards rate or mortality function, given here by the Siler function²³, of the form

$$\mu(x) = \exp(a_0 - a_1x) + c + \exp(b_0 + b_1x), \quad (1)$$

where $\theta = [a_0, a_1, c, b_0, b_1]$ is a vector of parameters to be estimated, and where $a_0, b_0 \in \mathbb{R}$ and $a_1, c, b_1 \geq 0$. From the mortality model in Eq. (1) the cumulative survival function can be calculated as $S(x) = \int_0^x -\mu(t)dt$, while the probability density function of ages at death is given by $f(x) = \mu(x)S(x)$ for $x \geq 0$.

For all species we studied, individuals of one or both sexes often leave their natal groups to join other neighboring groups in a process commonly identified as natal dispersal. For some species, individuals who have undergone natal dispersal can then disperse additional times, described as secondary dispersal. Although dispersal within monitored groups (i.e. those belonging to the study area) does not affect the estimation of mortality, the fate of individuals that permanently leave the study area to join unmonitored groups can be mistaken for possible

death. We identify this process as “out-migration”, which we classify as natal or immigrant out-migration, the first for natal and the second for secondary dispersals to unmonitored groups. This distinction is particularly relevant because not all out-migrations are identified as such, and therefore the fate of some individuals is unknown after their last detection. For these individuals we define a latent out-migration state at the time they were last detected, given by the random variable indicator O , with observations $o_{ij} = 0, 1$, where $o_{ij} = 1$ if individual i out-migrated and $o_{ij} = 0$ otherwise, and where $j = 1$ denotes natal out-migration and $j = 2$ for immigrant out-migration. For known out-migrations, we automatically assign $o_{ij} = 1$. The model therefore estimates the Bernoulli probability of out-migration, π_j , such that $O_{ij} \sim \text{Bern}(\pi_j)$. Those individuals assigned as exhibiting out-migration, as well as known emigrants and immigrants, contribute to the estimation of the distribution of ages at out-migration. Here, we define a gamma-distributed random variable V for ages at out-migration, with realizations $v \geq 0$, where $V_j | O_j = 1 \sim \text{Gam}(\gamma_{j1}, \gamma_{j2})$ and where $\gamma_{j1}, \gamma_{j2} > 0$ are parameters to be estimated with j defined as above. The probability density function for the gamma distribution is $g_V(x - v_j | \gamma_{j1}, \gamma_{j2})$ for $x \geq 0$, where v_j is the minimum age at natal or immigrant out-migration.

In addition, since not all individuals have known birth dates, the model samples the unknown births b_i as $x_{il} = t_{il} - b_i$, where t_{il} is the time of last detection for individual i . The likelihood is then defined as

$$p(x_{il}, x_{if} | \boldsymbol{\theta}, \boldsymbol{\gamma}_1, \boldsymbol{\gamma}_2, \pi_j, o_{ij}) = \begin{cases} \frac{f(x_{il})}{s(x_{if})} (1 - \pi_j) & \text{if } o_{ij} = 0 \\ \frac{s(x_{il})}{s(x_{if})} \pi_j g_V(x_{il} - v_j) & \text{if } o_{ij} = 1 \end{cases}, \quad (2)$$

where x_{if} is the age at first detection, given by $x_{if} = t_{if} - b_i$, with t_{if} as the corresponding time of first detection. The parameter vectors $\boldsymbol{\gamma}_1$ and $\boldsymbol{\gamma}_2$ are for natal and immigrant out-migration, respectively. In other words, individuals with $o_{ij} = 0$ are assumed to have died shortly after the last detection, while those with $o_{ij} = 1$ are censored and contribute to the estimation of the distribution of ages at out-migration. The full Bayesian posterior is then given by

$$p(\boldsymbol{\theta}, \boldsymbol{\gamma}_1, \boldsymbol{\gamma}_2, \boldsymbol{\pi}, \mathbf{b}_u, \mathbf{o}_u, \mathbf{v}_{u1}, \mathbf{v}_{u2} | \mathbf{b}_k, \mathbf{o}_k, \mathbf{t}_f, \mathbf{t}_l) \propto p(\mathbf{x}_l, \mathbf{x}_f | \boldsymbol{\theta}, \boldsymbol{\gamma}_1, \boldsymbol{\gamma}_2, \boldsymbol{\pi}, \mathbf{d}) \times p(\boldsymbol{\theta})p(\boldsymbol{\gamma}_1)p(\boldsymbol{\gamma}_2)p(\boldsymbol{\pi}), \quad (3)$$

where the first term on the right-hand-side of Eq. (3) is the likelihood in Eq. (2), and the following terms are the priors for the unknown parameters. The vector $\boldsymbol{\pi} = [\pi_1, \pi_2]$ is the vector of probabilities of out-migration while the subscripts u and k refer to unknown and known, respectively.

Following Colchero et al ¹¹, we used published data, expert information and an agent-based model to estimate the mortality and out-migration prior parameters for each population. We assumed a normal (or truncated normal distribution depending on the parameter’s support) for all the parameters. We used vague priors for the mortality and natal out-migration parameters ($\text{sd} = 10$), and informative priors for the immigrant out-migration parameters ($\text{sd} = 0.5$). We ran six MCMC parallel chains for 25 000 iterations each with a burn-in of 5 000 iterations for each population, and assessed convergence using potential scale reduction factor (32).

For the zoo data we used a simplified version of the model described above, which omitted all parts that related to out-migration. In order to produce Supplementary Figs. S1 and S2, we used the same method as for the zoo data on the human life tables. To achieve this, we created an individual level dataset from the l_x column of each population, and then fitted the Siler model to this simulated data. It is important to note that the Siler model might not provide the best fit to human data, in part due to the late life mortality plateau common among human

populations³⁰. It is therefore likely that the values of the mortality parameter b_1 we report in Supplementary Table S2 for the human populations are under-estimated. Nonetheless, and for the purposes of our analyses, the Siler fits to the human populations we considered here are reasonable (Supplementary Fig. S6) and we can therefore confidently state that the limitations of the Siler model do not affect the generality of our results.

Estimation of life expectancy and lifespan equality. We calculated life expectancy at birth as

$$e = \int_0^\infty S(t|\hat{\theta})dt, \quad (4)$$

where $S_X(x)$ is the cumulative survival function as defined above and where $\hat{\theta}$ is the vector of mortality parameters calculated as the mean of the conditional posterior densities from the survival analysis described above. We calculated the lifespan inequality^{15,31}, H , as

$$H = -\frac{1}{e} \int_0^\infty S(t|\hat{\theta}) \log[S(t|\hat{\theta})]dt, \quad (5)$$

Following Colchero et al¹¹, we defined lifespan equality as $\varepsilon_0 = -\log(H)$. We calculated both measures for each of the study populations, and performed weighted least squares regressions for each genus, with weights given by the reciprocal of the standard error of the estimated life expectancies.

Sensitivities of life expectancy and lifespan equality to mortality parameters. For simplicity and since we are calculating both measures from birth, we use hereafter $e = e_0$ and $\varepsilon = \varepsilon_0$. We derived the functions for the sensitivity of life expectancy and lifespan equality to changes in mortality parameters, where the hazard rate is described by a continuous function of age as in Eq. (1). The sensitivity of life expectancy to a given mortality parameter, $\theta \in \boldsymbol{\theta}$, is given by

$$\frac{\partial e}{\partial \theta} = e_\theta = \int_0^\infty S_\theta dx, \quad (6)$$

where $S_\theta = \frac{\partial}{\partial \theta} S_X(x)$ is the first partial derivative of the cumulative survival with respect to the mortality parameter θ . The sensitivity of lifespan equality to changes in parameter θ is given by

$$\frac{\partial \varepsilon}{\partial \theta} = \varepsilon_\theta = \frac{1}{e} [e_\theta (1 + H^{-1}) - H^{-1} \int_0^\infty S_\theta U dx], \quad (7)$$

where $U = \int_0^x \mu(t)dt$ is the cumulative hazards and H is the life table inequality defined in Eq. (5) (for full derivation see Supplementary Methods). From the results in Eqs. (6) and (7), we calculated the vectors of change (gradient vectors) at any point $\langle e_j, \varepsilon_j \rangle$ of the life expectancy-lifespan equality landscape, as a function of each of the Siler mortality parameters (See Fig. 2A,B).

To quantify the amount of change of each parameter along the genus lines, we derived the sensitivities of a given mortality parameter θ to changes in life expectancy and lifespan equality, namely $\frac{\partial \theta}{\partial e} = \frac{1}{e_\theta}$ for $e_\theta \neq 0$, and $\frac{\partial \theta}{\partial \varepsilon} = \frac{1}{\varepsilon_\theta}$ for $\varepsilon_\theta \neq 0$. With these sensitivities we calculated the gradient vector

$$\nabla \theta = \langle \frac{\partial \theta}{\partial e}, \frac{\partial \theta}{\partial \varepsilon} \rangle, \quad (8)$$

for any parameter at any point along the genus lines. Since we found a linear relationship between life expectancy and lifespan equality for every genus studied here, we calculated the relative amount of change of each parameter along the genus line by numerically solving the path integral

$$\Theta_j = \int_{C_j} \nabla g(\theta) d\mathbf{r}, \quad (9)$$

where $g(\theta) = \log(\theta)$, $d\mathbf{r} = \langle d\epsilon, d\epsilon \rangle$ and the integral subscript C_j represents the linear path from the genus j line. In short, the path integral Θ_j provides a measure of the relative change in parameter θ along the genus line (Fig. 3). To allow comparisons between all genera, we scaled the values of each path integral by the length of each line.

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Author contributions

F.C. contributed Conceptualization, Methodology, Formal Analysis, Visualization, Writing-Original and Writing-Review/editing, Project Administration. F.V. contributed Methodology, Writing - Review/editing. J.M.A. contributed Methodology, Writing - review/editing. J.W.V. contributed Conceptualization, Writing – Review/editing, Funding Acquisition. S.C.A contributed Conceptualization, Methodology, Resources, Writing-Original and Writing-Review/editing, Visualization, Project Administration, Funding Acquisition. All other authors: Resources, Methodology, Writing - Review/editing. F.C. and S.C.A contributed equally to this work.

Competing Interests

The authors declare no competing interests.

Additional Information

Supplementary information is available at ...

Correspondence and requests for materials should be addressed to F.C. or S.C.A

Data Availability: Data underlying the analyses here are available in the Dryad data repository, URL: <https://doi.org/10.5061/dryad.4b8gthtb4>

Fig. 1.

Fig. 1 The life expectancy–lifespan equality landscape for seven genera of primates for A) females and B) males. Each genus is characterized by a relatively constrained relationship between life expectancy and lifespan equality, and thus a distinct regression line. The type of line (e.g. continuous, dashed, or dotted) depicts three levels for the p -values of the slopes (how significantly different from 0 they are), while the shaded polygons show the 95% confidence intervals of the regressions. Panel C) shows the relationship between the Siler mortality parameters and the resulting mortality function, given by the equation $\mu(x) = \exp(a_0 - a_1 x) + c + \exp(b_0 + b_1 x)$, where infant and juvenile mortality are controlled by parameters a_0 and a_1 , age-independent mortality is captured by c , and senescent mortality is captured by b_0 (initial adult mortality) and b_1 (rate of aging). Panel D) shows how gradual changes in in each Siler mortality parameter modify the life expectancy and lifespan equality values (thick purple line). The green line corresponds to the regression line for female chimpanzees, shown for reference to illustrate the general trends among all genus lines. The purple curves show the changes in life expectancy and lifespan equality after varying individual Siler parameters while holding the other parameters constant. Note the striking change in life expectancy and lifespan equality that would result from changes in the aging parameters, particularly b_1 . See Supplementary Fig. S3 for plots that include individual points for each population.

Fig. 2.

Fig. 2 Sensitivities of life expectancy and lifespan equality to changes in mortality parameters.

A) Using the female chimpanzee line as an example, vectors depict the sensitivity at the mid-point of the genus line. Each vector depicts the direction and magnitude of change in life expectancy and lifespan equality for a unit change in the corresponding Siler mortality parameter. The vectors for c and b_1 are particularly large, represented by broken lines. B)

Gradient field of sensitivities of life expectancy and lifespan equality to changes in each mortality parameter, showing the direction of change any population would experience for a given change in the parameter, from any starting point in the landscape. The green chimpanzee line is provided for reference. Each sensitivity vector (bright purple) can be interpreted as those in A, but calculated from different points on the landscape. C) Boxplots representing the values of the seven collinearity values (one for each genus) for each of the Siler parameters. Collinearity is calculated between the mid-point of the genus line and the sensitivity vector for each parameter; a value of 1 would imply that the vector is parallel, a value of 0 would imply that it is perpendicular. Note the relatively large collinearity values for a_0 , a_1 , and c and the relatively small value for b_1 . The horizontal black line in each boxplot shows the median.

Fig. 3.

Fig. 3 Relative magnitude of change of each parameter along the genus lines. Pre-adult and age-independent mortality parameters (a_0 , a_1 , and c) vary several orders of magnitude more, within each genus, than the aging parameters (b_0 and b_1). Values were calculated by numerically solving the path integral in Eq. (9) (see Material and Methods and Supplementary Text) for each parameter along each genus line. The y-axes were scaled by the logarithm base 10 to improve interpretability.

Figures

Figure 1

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