Overexpressed P-S6 Associates with Lymph Node Metastasis and Predicts Poor Prognosis in Non-Small Cell Lung Cancer.

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Abstract

Background

Ribosomal protein S6 (S6), a downstream effect media of the AKT/mTOR pathway, not only is a part of 40S small subunit of eukaryotic ribosome, but also involves in protein synthesis and cell proliferation during cancer development.

Methods

In present study, we explore the association between phosphorylated S6 (p-S6) protein expression and clinicopathological features as well as prognostic implications in NSCLC. P-S6 was detected in tissue microarrays (TMAs) containing 350 NSCLC, 53 non-cancerous lung tissues (Non-CLT), and 88 cases of matched metastatic lymph node lesions via immunohistochemistry (IHC). Transwell assays and wound healing assay were used to assess the effects of p-S6 inhibition on NSCLC cell metastasis.

Results

The p-S6 expression in NSCLC was more evident than that in Non-CLT ($p < 0.05$). Compared to NSCLC patients who have no lymph node metastasis (LNM), those with LNM had higher p-S6 expression ($p = 0.001$). Regardless of lung squamous cell carcinoma (SCC) or adenocarcinoma (ADC), p-S6 was increased obviously in metastatic lymph nodes compared with matched primary cancers ($p = 0.001$, $p = 0.022$, respectively). Inhibition of p-S6 decreased the metastasis ability of NSCLC cells. In addition, p-S6 was an independent predicted marker for LNM in patients with NSCLC ($p < 0.001$). According to survival analysis, patients with highly expressed p-S6 had a lower survival rate compared with that with lower expression ($p = 0.013$). P-S6 is an unfavorable independent prognostic factor for NSCLC patients ($p = 0.011$).

Conclusion

increased expression of p-S6 is not only a novel predictive biomarker of LNM but also poor prognosis in NSCLC.

Background

Lung cancer is a clinical malignant tumor with highest incidence, with about 2 million new lung cancer cases annually, and its mortality rate is also the highest worldwide [1]. 80–90% of lung cancer are diagnosed as non-small cell lung cancer (NSCLC), which is mainly classified into adenocarcinoma (ADC) and squamous cell carcinoma (SCC) [2]. Lung cancer patients' survival varies depending on lymph node metastasis (LNM), clinical stage, geographic region, and other factors. The 5-year survival of NSCLC patients in clinical stageⅠis about 57%, which is far more than that of patients with stage Ⅳ (that is less than 5%) [3]. Unfortunately, more than half of the NSCLC patients are diagnosed in advanced clinical stages, often accompanied by LNM and distant metastasis, which are important reasons for the poor

prognosis [3, 4]. Therefore, it is an urgent need to find new biomarker for early detection of LNM of NSCLC and predict the prognosis.

Recently, the protein kinase B (AKT)/mammalian target of rapamycin (mTOR) signal pathway is over-activated in numerous tumors, including NSCLC, and associated with tumor angiogenesis, invasion, metastasis and so on [5–7]. Ribosomal protein S6 (S6), a part of 40S small subunit of eukaryotic ribosome, is a famous downstream effect media of the AKT/mTOR pathway [8, 9]. As the first ribosomal protein proved to undergo inducible phosphorylation, S6 is mostly induced by activated 70kDa S6 Kinases (S6K) at five phosphorylation sites (Ser235, Ser236, Ser240, Ser244, Ser247), mainly Ser236 [9–12]. Phosphorylated S6 (p-S6) plays a crucial part in protein synthesis, cell size control and cell proliferation as increasing the affinity of the ribosome and mRNAs and improving the efficiency of protein translation [13]. In addition, p-S6 is also the key effector of mTOR in regulating cell size, whose decreased expression results in smaller cell size and reflects the growth defect [14, 15]. Overexpression of p-S6 is found in various solid tumors, such as gastric cancer, glioblastomas, and renal cell carcinomas (RCCs), and associated with poor prognosis [16–18]. S6 phosphorylation was even associated with malignant potential and glucose metabolism of intraductal papillary mucinous neoplasm of the pancreas [19]. In addition, the phosphorylation of S6 is considered to contribute to acquired resistance to MAPK pathway inhibitors in cancers, suggesting that p-S6 plays an essential role in the mechanism of anti-cancer drugs [20, 21].

However, the prognostic implications of p-S6 are poorly understood in NSCLC. In this present research, we have measured the level of p-S6 expression in tissue microarrays (TMAs) of non-cancerous lung tissues (Non-CLT) and NSCLC by immunohistochemistry (IHC) and decreased p-S6 expression in NSCLC cell lines with mTOR inhibitor (RAD001), to estimate not only the association between p-S6 and clinicopathological/prognostic features of NSCLC patients, but also the alteration of NSCLC cell migration and invasion ability.

Material And Methods

Patients and tissue samples

All tissue specimens which were collected by surgical resection acquired from the Thoracic Surgery Department of the Second Xiangya Hospital of Central South University (CSU) between 2003 and 2013. Approval of all protocols were obtained from the Institutional Human Experiment and Ethics Committee of the Second Xiangya Hospital of CSU (Changsha, China) (approval No. S039/2011), and approval of all protocols were obtained. 350 cases of NSCLC, 53 cases of Non-CLT, and 88 cases of matched metastatic lymph node lesions were included in this research. Patients in the study had definite histological diagnosis on the basis of 2015 WHO Classification of lung cancer [22], and were comprehensively staged as per the Eighth Edition Lung Cancer Stage Classification [23]. Complete clinicopathological data obtained from medical records were available for all patients (Supplementary Table).

Cells lines and reagents

Two human NSCLC cell lines, A549 and SPC-A1, were from the Cell Bank, Chinese Academy of Sciences (Shanghai, China) and those were grown in 37°C, 5% CO₂ incubator. Both cell lines were culture in RPMI-1640 (BI, Israel) medium comprising 10% fetal bovine serum (BI, Israel). The mTOR inhibitor Everolimus (RAD001) was procured from Selleckchem, USA. Rabbit antibodies against p-S6 (S235/236) (#4857) were obtained from Cell Signaling Technology, USA. Mouse antibodies against S6 (cat no. 66886-1-Ig) and β -Actin (cat no. 66009-1-Ig) were purchased from proteintech, USA. Peroxidase-conjugated Affinipure Goat anti-Rabbit IgG (cat no. HG-SAR00002a) and anti-Mouse IgG (cat no. YG-SAM00001a) were purchased from HonorGene, China.

Construction of the tissue microarrays

As previously described, tissue microarrays (TMAs) were constructed using paraffin-embedded tissue of NSCLC and Non-CLT [24]. The perforation diameter of each sample was 0.6 mm. For TMAs of NSCLC, each case included two tumor cores, of which 88 cases each also contained two matched metastatic lymph node cores. For TMAs of Non-CLT, two cores per case was included. The average score of the two cores was regarded as the final score for each case.

Immunohistochemistry and scores

IHC for p-S6 in TMAs was carried out and the conditions of antibody staining were adjusted as described previously [25]. Briefly, primary antibody for p-S6 (1:800 of Ser235/236) was applied to detect its expression level. The positive control slide and negative control slide were included in each experiment. The IgG isotype-matched antibody was applied as negative contrast to confirm the antibody specificity.

Immunoreactivity was assessed semi-quantitatively by two independent pathologists. The staining intensity score was multiplied by percentage score of positive expressed tumor cells as the total score. Specifically, staining intensity was scored as 0 to 3 (0 for negative; 1 for mild expression; 2 for moderate expression; 3 for strong expression). The percentage score was identified as 0 to 4 in view of positive cytoplasmic staining: 0 = 0%; 1 = 1%-25%, 2 = 26%-50%, 3 = 51%-75%, 4 = 76%-100%. The range of total score was from 0 to 7. Cutoff level which determined according to overall survival (OS) of NSCLC patients was 2. Expression of p-S6 in tumor cells of which total score is more than 2 was considered high expression and others were regarded as low expression. Agreement between the two assessors is 96%, borderline and ambiguous cases are resolved through discussion.

Western blotting

As previously described [26], preparation of protein lysates and western blotting analysis were performed. The antibodies were used as follows: 1:1000 of Ser235/236 for anti-p-S6 and 1:5000 for anti- β -Actin.

Wound healing assay

1*10⁵ SPC-A1 and A549 cells were seeded and then treated with DMSO or 5 nmol/L RAD001 at about 60% confluence and cultured until 90%. Serum free RPMI-1640 was used to replace the culture medium and wounds were scratched with the tip of 10 μ l pipette in each well. The size of wounds was observed

using inverted microscope (Leica, Germany) and images were captured at 0h, 24h and 48h. The data presented were repeated in triplicate. Wound healing percentage (%) = $(1 - \text{scratch area at } t / \text{scratch area at } 0h) * 100\%$ (t means 24h or 48h).

Transwell assays

24-well chambers (Costar 3422; Corning, USA) were used to perform transwell migration assays. A549 and SPC-A1 cells were treated with DMSO or 5 nmol/L RAD001 for 24h at 37°C. Then 5×10^4 cells were extracted from each group and suspended in 200 μ l of serum-free RPMI-1640 to the upper chamber, while 10% FBS RPMI-1640 was added to the lower room. The cells could migrate through the membrane for 12h to 48h. The membrane was fixed for 20 min in 4% paraformaldehyde solution and dyed with crystal violet for 20min at room temperature. The count of cells under the cell membrane was calculated. The implementation of invasion assay protocol be analogous to that of migration assay, except that Matrigel was coated on the upper cavity and the incubation time was 1 h (356234, Corning, USA).

Statistical analysis

The difference expression pattern of p-S6 between Non-CLT and NSCLC, as well as the association between p-S6 and clinicopathological features were analyzed by Chi-square (χ^2) test. The difference between the metastatic lymph nodes and matched primary tumors was also analyzed by Chi-square(χ^2) test. Multi-logistic regression method was performed to identify whether p-S6 is an independent marker for LNM of NSCLC. Survival rate curve was evaluated by Kaplan-Meier analysis, and comparisons were analyzed via log-rank test. Furthermore, cox proportional hazards model was performed for determining independent prognostic markers. The deviation between control and RAD001 groups was analyzed using one way t-test. All these statistical analyses were put into effect with SPSS Statistics software (version 24). Numerical data were presented as mean $\pm$ SEM. $P < 0.05$ (Two-sided) indicates that the result is statistically significant.

Results

P-S6 was significantly overexpressed in NSCLC

Firstly, we detected the intracellular localization and the level of p-S6 expression by IHC in a total of 403 samples, of which 350 were NSCLC. The results indicated that p-S6 was mainly observed in the cell cytoplasm of lung SCC, ADC and Non-CLT (Figure 1). Quantification of p-S6 expression showed that its percentage of high expression was 30.5% (47/154) in lung SCC, 62.2% (122/196) in lung ADC and 15.1% (8/53) in Non-CLT, respectively. As shown in Supplementary Figure 1, compared with Non-CLT, the level of p-S6 protein was obviously increased both in lung SCC and ADC ($p = 0.031$, $p < 0.001$, respectively).

Association between p-S6 and clinicopathological features

As data showed in Table 1, p-S6 in female was higher than that in male ($p < 0.001$). Patients with lung ADC had obviously higher p-S6 expression level than patients with SCC ($p < 0.001$). Overexpressed p-S6

was significantly correlated with LNM ($p = 0.001$). Moreover, patients with high p-S6 expression level suffered a lower overall survival (OS) rate than that with low expression ($p = 0.005$). However, there was not significantly association between p-S6 expression and other features, such as clinical stages, pathological grade and age (all $p > 0.05$).

Table 1

Analysis of the association between the expression level of p-S6 and clinicopathological features of NSCLC (n=350)

Clinicopathological features (n)	p-S6			
	N	High (%)	Low (%)	P-value
Age(years)				
≤50	95	48(50.5)	47(49.5)	0.632
≥50	255	121(47.5)	134(52.5)	
Gender				
Male	266	114(42.9)	152(57.1)	0.000*
Female	84	55(65.5)	29(34.5)	
Clinical stage				
Stage I-II	151	75(49.7)	76(50.3)	0.519
Stage III	199	93(46.7)	106(53.3)	
LN status				
LNM	210	117(55.7)	93(44.3)	0.001*
No LNM	140	52(37.1)	88(62.9)	
Histological type				
SCC	154	47(30.5)	107(69.5)	0.000*
ADC	196	122(62.2)	74(37.8)	
Pathological grade				
Well/ Moderate	152	77(50.7)	75(49.3)	0.452
Poor	198	92(46.5)	106(53.5)	
Survival status				
Alive	187	77(41.2)	110(58.8)	0.005*
Dead	163	92(56.4)	71(43.6)	
*Chi-square test, statistically significant difference (<i>P</i> < 0.05). Abbreviations: ADC, Adenocarcinoma, SCC, squamous cell carcinoma; LNM, lymph node metastasis.				

Impact of p-S6 expression on LNM in NSCLC

Among the 350 NSCLC patients, 210 cases had LNM and 140 cases were free from LNM. NSCLC patients with LNM had obviously higher p-S6 expression compared with those without LNM ($p = 0.001$, Table 1). We also evaluated the level of p-S6 in primary NSCLC tissues and its matched LNM lesions. As shown in Figure 2, regardless of lung SCC or ADC, the percentage of high p-S6 expression is significantly higher in metastatic lymph node lesions with compared with the matched primary cancer ($p = 0.001$, $p = 0.022$, respectively). To determine whether p-S6 expression was the independent predicted parameter for LNM in NSCLC, a multivariate logistic regression analysis was carried out. As mentioned in Table 2, highly expressed p-S6 ($p = 0.001$), clinical stages ($p < 0.001$), and age ($p = 0.010$) were notably correlated with LNM status of NSCLC patients. These results indicated that overexpressed p-S6 is an independent factor for LNM of NSCLC regardless of other parameters.

Table 2. Multivariate logistic analysis of LNM factors in NSCLC patients

Variables	95.0% CI for Exp (B)						
	B	S.E.	Wald	P-value	Exp(B)	Lower	Upper
p-S6 expression	0.921	0.270	11.644	0.001*	2.512	1.480	4.262
Clinical stages	1.957	0.264	54.968	0.000*	7.068	4.220	11.880
Histological type	-0.026	0.276	0.009	0.925	0.974	0.567	1.674
Pathological grade	-0.051	0.256	0.039	0.843	0.951	0.575	1.571
Age	-0.759	0.293	6.688	0.010*	0.468	0.263	0.832
Gender	0.607	0.324	3.516	0.061	1.835	0.973	3.462

Abbreviations: S.E., standard error; Exp(B), odds ratio;CI, confidence interval; LNM, lymph node metastasis.* $P < 0.05$.

Inhibition of p-S6 reduced the cell migration and invasion of NSCLC

The alteration of cell migration and invasion ability of NSCLC were assessed after inhibition of p-S6 expression level inNSCLC cell lines (A549and SPC-A1).Firstly, we evaluated the inhibition efficiency by Western blotting after the cell lines were treated with RAD001. Figure 3 showed that p-S6 expression was significantly reduced after a 24 h treatment with RAD001 at 5 nmol/L.Secondly, to detect the effect of p-S6 on the cell migration, wound healing and transwell migration experiments were conducted. Figure 4 showed that compared with the control group, the wound healing percentage of RAD001 group was decreased by 10.19% and 20.44% at 24h and 48h in A549 cells ($p = 0.018$, $p = 0.001$, respectively), by 5.90% and 7.55% in SPC-A1 cells ($p = 0.028$, $p = 0.048$, respectively), respectively. Figure 5A revealed that the inhibition of p-S6 expression inA549 and SPC-A1 cell lines decreased the migration ability by transwell migration assay (both $p < 0.001$). Moreover, the cell invasion assay demonstrated that p-S6 inhibition in NSCLC cell lines reduced the number of invaded cells (both $p < 0.001$) (Figure 5 B). Together,

these results revealed that inhibition of p-S6 expression could reduce the cell migration and invasion of NSCLC.

Impact of p-S6 expression on patients' prognosis

In univariate survival analysis, survival rate was estimated by Kaplan-Meier method in 350 NSCLC patients, and comparison was performed via log-rank test. NSCLC patients with highly expressed p-S6 had a poor prognosis compared to those with lowly expressed p-S6 ($p = 0.013$) (Figure 6A & Table 3). The OS rate of NSCLC patients with advanced clinical stage (stage III) was lower relative to those with early stages (stage I-II) ($p < 0.001$) (Figure 6B & Table 3). Of note, patients without LNM have higher OS than those with LNM ($p < 0.001$) (Figure 6C & Table 3). Meanwhile, compared with NSCLC patients with well and moderate pathological grade, lower OS could be seen in those with poor differentiation ($p = 0.003$) (Figure 6D & Table 3).

Table 3

Summary of univariate/multivariate analysis for overall survival in patients with NSCLC (n=350)

Variables	Univariate analysis			Multivariate analysis		
	Average survival time (SE)	95%CI	P-value	Exp (B)	95.0%CI	P-value
p-S6						
High expression	44.561(3.050)	38.583-50.540	0.013*	1.549	1.105-2.173	0.011*
Low expression	67.500(5.336)	57.041-77.958				
Clinical stages						
Stage $\geq$ II	68.200(4.195)	59.977-76.423	0.000*	1.980	1.356-2.893	0.000*
Stage I	47.862(3.834)	40.347-55.378				
LN status						
LNM	44.969(3.201)	38.695-51.243	0.000*	1.609	1.098-2.360	0.015*
No LNM	74.932(5.919)	63.330-86.533				
Histological type						
SCC	63.464(5.825)	52.046-74.882	0.240	1.137	0.806-1.603	0.464
ADC	50.588(3.367)	43.988-57.188				
Pathological grade						
Well/moderated	58.662(3.509)	51.784-65.541	0.003*	1.517	1.094-2.104	0.013*
Poor	54.226(4.164)	46.066-62.387				
Age						

Abbreviations: CI, confidence interval; Exp(B), odds ratio; SE, standard error; LNM, lymph node metastasis;

* $P < 0.05$.

Variables	Univariate analysis			Multivariate analysis		
	Average survival time (SE)	95%CI	P-value	Exp (B)	95.0%CI	P-value
≤50	45.395(3.633)	38.274-52.515	0.454	1.014	0.718-1.430	0.939
≥50	61.199(4.064)	53.234-69.165				
Gender						
Female	54.566(4.467)	45.810-63.321	0.210	0.718	0.484-1.064	0.099
Male	57.363(3.886)	49.747-64.978				
Abbreviations: CI, confidence interval; Exp(B), odds ratio; SE, standard error; LNM, lymph node metastasis;						
* $P < 0.05$.						

To further investigate whether the overexpressed p-S6 was an independent prognostic index for NSCLC patients, Cox regression analysis was carried out. The results shown in Table 3 indicated that up-regulated p-S6 might be a poor prognostic marker for patients with NSCLC ($p = 0.011$), as are clinical stage ($p < 0.001$), LNM status ($p = 0.015$) and pathological grade ($p = 0.013$). Besides, age, gender, histological type had no significant effect on the prognosis of patients with NSCLC.

Discussion

In this study, we demonstrated that the level of p-S6 protein expression in NSCLC was significantly higher than that in Non-CLT. The percentage of high p-S6 expression was obviously increased in patients with LNM than that without LNM. Inhibition of p-S6 expression reduced the invasion and migration ability of NSCLC cells. Our analysis indicated that NSCLC patients with increased p-S6 expression had a lower rate of survival than that with low expression ones. Taking together, our results imply that p-S6 may play a significant part in the progression of NSCLC and aberrant expression of p-S6 might be a novel prognostic marker for NSCLC. Increasing evidence has shown that mTOR/S6K/S6 pathway plays a crucial role in p53-mediated tumor inhibition [27, 28]. p-S6, the key downstream effector of mTOR/S6K pathway, is involved in the occurrence and development of many cancers [9]. Its expression is ascendant in numerous tumors, such as RCCs, pancreatic cancer, and esophageal squamous cell carcinoma [18, 27, 29]. The phosphorylation of S6 can attenuate the autophagy induced by damage-regulated autophagy modulator 1 (DRAM1) and p-S6 is a requirement for AKT-driven malignant transformation of pancreatic islet β cells [28, 30]. Due to the oncogenic role of p-S6, decreasing its expression could potentially provide a clue to find new ideas for the targeted treatment of cancers. For example, the suppression of p-S6 can block the further inhibition of the therapeutic mTOR inhibitor everolimus on protein synthesis and proliferation of

RCCs cells[18]. Knockdown of S6K1 gene can obviously decrease the expression of cyclin D, leading to the decline of survival ability of esophageal cancer cells[29].

Activating invasion and metastasis is one of biological capabilities of cancers[31]. Our study showed that NSCLC patients with LNM had higher p-S6 expression. Furthermore, the expression level of p-S6 in metastatic lymph node lesion was higher than that in matched primary lesion. Multivariate analysis indicated that the high p-S6 expression could be an independent predicted marker for LNM in patients with NSCLC. To further clarify the metastatic ability of p-S6 in NSCLC, we inhibited p-S6 expression in A549 and SPC-A1 cell lines and found that down regulation of p-S6 weakened the migration and invasion ability of NSCLC cells. Taking together, these data suggest that p-S6 might play a major part in promoting invasion and metastasis of NSCLC. Similar to our discoveries, previous studies have reported that overexpressed p-S6 is positively related to LNM in RCCs, colorectal cancer, and epithelial ovarian cancer[18, 32, 33]. On the other hand, inhibition of p-S6 expression or S6 gene knockdown can significantly suppress the cell invasion and migration of several kinds of human cancers, such as esophageal cancer and colorectal cancer cells[29, 34]. All the mentioned above indicate p-S6 positively affects cell invasion and migration. Despite the number of studies describing the aberrant expression of p-S6 in tumors activating invasion and metastasis, the concrete biological mechanism is still unclear. One study report that the phosphorylation defect of S6 inhibits the phosphorylation of paxillin, a focal adhesion protein, leading to inhibit the formation of local adhesion[29]. In this current study, we have little knowledge of how the aberrant expression of p-S6 involved in the LNM of NSCLC, further explorations for the intrinsic mechanism are required in the future.

Our results indicated that the OS rate of NSCLC patients with highly expressed p-S6 was obviously lower than that of those with low level of p-S6. Multivariate analysis showed that high expressed p-S6 was an independently prognostic indicator in NSCLC patients, which seem to favor the oncogenic role of p-S6. Previous studies on RCCs, gastric cancer, and glioblastomas also identified p-S6 as a novel poor prognosis biomarker[16-18]. Up to now, there are many inhibitors that can prevent S6 phosphorylation. Studies have reported that inhibition of p-S6 can significantly reduce tumor growth, which is important for effective response to treatment of triple negative breast cancer[35, 36]. In addition, mTOR inhibitor everolimus can effectively inhibit the level of p-S6, thereby reversing the resistance of *HER2*-mutant cancers to neratinib and exerting anti-tumor effects[37]. These further suggest that p-S6 may be a powerful biomarker in tumors and may provide novel strategy in targeted therapy of NSCLC.

Conclusions

In summary, a significant overexpression of p-S6 was found in NSCLC. Inhibition of p-S6 expression could weaken the migration and invasion ability of NSCLC cells and aberrant expression level of p-S6 might be an independent predictor for LNM of NSCLC patients. In addition, overexpressed p-S6 may be a novel poor prognostic biomarker for NSCLC patients.

Abbreviations

NSCLC: Non-small cell lung cancer;ADC: Adenocarcinoma; SCC:Squamous cell carcinoma; LNM: Lymph node metastasis; AKT: The protein kinase B; mTOR: Mammalian target of rapamycin;S6: Ribosomal protein S6; S6K: 70kDa S6 Kinases; p-S6: Phosphorylated S6; RCCs: Renal cell carcinomas; TMAs: Tissue microarrays; Non-CLT: Non-cancerous lung tissues;

Declarations

Ethics approval and consent to participate

In this study, all procedures performed on research samples were confirmed by the ethical standards of the Institute Ethics Committee of the Second Xiangya Hospital of Central South University (approval No. S039/2011) and in accordance with the 1964 Helsinki declaration and its later amendments of comparable ethical standards. Informed consent was obtained from all subjects or their legal guardian.

Consent for publication

Not applicable

Availability of data and materials

All data generated or analyzed during this study are included in this published article (and its supplementary information files). The datasets generated and analysed during the current study are available in the figshare repository, <https://doi.org/10.6084/m9.figshare.16901161.v2>.

Competing interests

The authors declare that they have no conflicts of interest.

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Authors' contributions

Qiuyuan Wen and Songqing Fan conceived and designed the study. Yaoxiang Tang and Jiadi Luo performed the experiments. Yaoxiang Tang, Yin Zhou, and Hongjing Zang wrote the paper. Yang Yang and Sile Liu analyzed the data. Qiuyuan Wen, Hongmei Zheng, and Jian Ma reviewed and edited the manuscript. All authors read and approved the manuscript and agree to take responsibility and be accountable for contents of the article.

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References

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin*. 2018;68(6):394-424. doi: 10.3322/caac.21492
2. Duma N, Santana-Davila R, Molina JR. Non-Small Cell Lung Cancer: Epidemiology, Screening, Diagnosis, and Treatment. *Mayo Clin Proc*. 2019;94(8):1623-40. doi: 10.1016/j.mayocp.2019.01.013
3. Miller KD, Nogueira L, Mariotto AB, Rowland JH, Yabroff KR, Alfano CM, et al. Cancer treatment and survivorship statistics, 2019. *CA Cancer J Clin*. 2019;69(5):363-85. doi: 10.3322/caac.21565
4. Woodard GA, Jones KD, Jablons DM. Lung Cancer Staging and Prognosis. *Cancer Treat Res*. 2016;170:47-75. doi: 10.1007/978-3-319-40389-2_3
5. Wen Q, Wang W, Luo J, Chu S, Chen L, Xu L, et al. CGP57380 enhances efficacy of RAD001 in non-small cell lung cancer through abrogating mTOR inhibition-induced phosphorylation of eIF4E and activating mitochondrial apoptotic pathway. *OncoTarget*. 2016;7(19):27787-801. doi: 10.18632/oncotarget.8497
6. Lu J, Zang H, Zheng H, Zhan Y, Yang Y, Zhang Y, et al. Overexpression of p-Akt, p-mTOR and p-eIF4E proteins associates with metastasis and unfavorable prognosis in non-small cell lung cancer. *PLoS One*. 2020;15(2):e0227768. doi: 10.1371/journal.pone.0227768
7. Aoki M, Fujishita T. Oncogenic Roles of the PI3K/AKT/mTOR Axis. *Curr Top Microbiol Immunol*. 2017;407:153-89. doi: 10.1007/82_2017_6
8. Wool IG, Chan YL, Gluck A. Structure and evolution of mammalian ribosomal proteins. *Biochem Cell Biol*. 1995;73(11-12):933-47. doi: 10.1139/o95-101
9. Ruvinsky I, Meyuhas O. Ribosomal protein S6 phosphorylation: from protein synthesis to cell size. *Trends Biochem Sci*. 2006;31(6):342-8. doi: 10.1016/j.tibs.2006.04.003
10. Gressner AM, Wool IG. The phosphorylation of liver ribosomal proteins in vivo. Evidence that only a single small subunit protein (S6) is phosphorylated. *J Biol Chem*. 1974;249(21):6917-25.
11. Bandi HR, Ferrari S, Krieg J, Meyer HE, Thomas G. Identification of 40 S ribosomal protein S6 phosphorylation sites in Swiss mouse 3T3 fibroblasts stimulated with serum. *J Biol Chem*. 1993;268(6):4530-3.
12. Wettenhall RE, Erikson E, Maller JL. Ordered multisite phosphorylation of *Xenopus* ribosomal protein S6 by S6 kinase II. *J Biol Chem*. 1992;267(13):9021-7.
13. Meyuhas O. Ribosomal Protein S6 Phosphorylation: Four Decades of Research. *Int Rev Cell Mol Biol*. 2015;320:41-73. doi: 10.1016/bs.ircmb.2015.07.006
14. Ruvinsky I, Sharon N, Lerer T, Cohen H, Stolovich-Rain M, Nir T, et al. Ribosomal protein S6 phosphorylation is a determinant of cell size and glucose homeostasis. *Genes Dev*. 2005;19(18):2199-211. doi: 10.1101/gad.351605

15. Ruvinsky I, Katz M, Drazan A, Gielchinsky Y, Saada A, Freedman N, et al. Mice deficient in ribosomal protein S6 phosphorylation suffer from muscle weakness that reflects a growth defect and energy deficit. *PLoS One*. 2009;4(5):e5618. doi: 10.1371/journal.pone.0005618
16. Zheng Z, Zheng Y, Zhang M, Wang J, Yu G, Fang W. Reciprocal expression of p-AMPK α and p-S6 is strongly associated with the prognosis of gastric cancer. *Tumour Biol*. 2016;37(4):4803-11. doi: 10.1007/s13277-015-4193-5
17. Machado LE, Alvarenga AW, da Silva FF, Roffe M, Begnami MD, Torres LFB, et al. Overexpression of mTOR and p(240-244)S6 in IDH1 Wild-Type Human Glioblastomas Is Predictive of Low Survival. *J Histochem Cytochem*. 2018;66(6):403-14. doi: 10.1369/0022155417750838
18. Knoll M, Macher-Goeppinger S, Kopitz J, Duensing S, Pahernik S, Hohenfellner M, et al. The ribosomal protein S6 in renal cell carcinoma: functional relevance and potential as biomarker. *OncoTarget*. 2016;7(1):418-32. doi: 10.18632/oncotarget.6225
19. Hirashita T, Hirashita Y, Iwashita Y, Endo Y, Kiyonaga M, Matsumoto S, et al. S6 ribosomal protein phosphorylation is associated with malignancy of intraductal papillary mucinous neoplasm of the pancreas. *Ann Gastroenterol Surg*. 2020;4(5):571-9. doi: 10.1002/ags3.12367
20. Hirashita Y, Tsukamoto Y, Yanagihara K, Fumoto S, Hijiya N, Nakada C, et al. Reduced phosphorylation of ribosomal protein S6 is associated with sensitivity to MEK inhibition in gastric cancer cells. *Cancer Sci*. 2016;107(12):1919-28. doi: 10.1111/cas.13094
21. Gao M-z, Wang H-b, Chen X-l, Cao W-t, Fu L, Li Y, et al. Aberrant modulation of ribosomal protein S6 phosphorylation confers acquired resistance to MAPK pathway inhibitors in BRAF-mutant melanoma. *Acta Pharmacol Sin*. 2018;40(2):268-78. doi: 10.1038/s41401-018-0020-z
22. Marx A, Chan JK, Coindre JM, Detterbeck F, Girard N, Harris NL, et al. The 2015 World Health Organization Classification of Tumors of the Thymus: Continuity and Changes. *J Thorac Oncol*. 2015;10(10):1383-95. doi: 10.1097/JTO.0000000000000654
23. Detterbeck FC, Boffa DJ, Kim AW, Tanoue LT. The Eighth Edition Lung Cancer Stage Classification. *Chest*. 2017;151(1):193-203. doi: 10.1016/j.chest.2016.10.010
24. Fan SQ, Ma J, Zhou J, Xiong W, Xiao BY, Zhang WL, et al. Differential expression of Epstein-Barr virus-encoded RNA and several tumor-related genes in various types of nasopharyngeal epithelial lesions and nasopharyngeal carcinoma using tissue microarray analysis. *Hum Pathol*. 2006;37(5):593-605. doi: 10.1016/j.humpath.2006.01.010
25. Wen Q, Wang W, Chu S, Luo J, Chen L, Xie G, et al. Flot-2 Expression Correlates with EGFR Levels and Poor Prognosis in Surgically Resected Non-Small Cell Lung Cancer. *PLoS One*. 2015;10(7):e0132190. doi: 10.1371/journal.pone.0132190
26. Liu S, Zang H, Zheng H, Wang W, Wen Q, Zhan Y, et al. miR-4634 augments the anti-tumor effects of RAD001 and associates well with clinical prognosis of non-small cell lung cancer. *Sci Rep*. 2020;10(1):13079. doi: 10.1038/s41598-020-70157-0
27. Khalaileh A, Drazan A, Khatib A, Apel R, Swisa A, Kidess-Bassir N, et al. Phosphorylation of ribosomal protein S6 attenuates DNA damage and tumor suppression during development of

- pancreatic cancer. *Cancer Res.* 2013;73(6):1811-20. doi: 10.1158/0008-5472.CAN-12-2014
28. Lu T, Zhu Z, Wu J, She H, Han R, Xu H, et al. DRAM1 regulates autophagy and cell proliferation via inhibition of the phosphoinositide 3-kinase-Akt-mTOR-ribosomal protein S6 pathway. *Cell Commun Signal.* 2019;17(1):28. doi: 10.1186/s12964-019-0341-7
29. Kim SH, Jang YH, Chau GC, Pyo S, Um SH. Prognostic significance and function of phosphorylated ribosomal protein S6 in esophageal squamous cell carcinoma. *Mod Pathol.* 2013;26(3):327-35. doi: 10.1038/modpathol.2012.161
30. Wittenberg AD, Azar S, Klochendler A, Stolovich-Rain M, Avraham S, Birnbaum L, et al. Phosphorylated Ribosomal Protein S6 Is Required for Akt-Driven Hyperplasia and Malignant Transformation, but Not for Hypertrophy, Aneuploidy and Hyperfunction of Pancreatic beta-Cells. *PLoS One.* 2016;11(2):e0149995. doi: 10.1371/journal.pone.0149995
31. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell.* 2011;144(5):646-74. doi: 10.1016/j.cell.2011.02.013
32. Lai EY, Chen ZG, Zhou X, Fan XR, Wang H, Lai PL, et al. DEPTOR expression negatively correlates with mTORC1 activity and tumor progression in colorectal cancer. *Asian Pac J Cancer Prev.* 2014;15(11):4589-94. doi: 10.7314/apjcp.2014.15.11.4589
33. Liu Z, Yun R, Yu X, Hu H, Huang G, Tan B, et al. Overexpression of Notch3 and pS6 Is Associated with Poor Prognosis in Human Ovarian Epithelial Cancer. *Mediators Inflamm.* 2016;2016:5953498. doi: 10.1155/2016/5953498
34. Shiratori H, Kawai K, Okada M, Nozawa H, Hata K, Tanaka T, et al. Metastatic role of mammalian target of rapamycin signaling activation by chemoradiotherapy in advanced rectal cancer. *Cancer Sci.* 2020;111(4):1291-302. doi: 10.1111/cas.14332
35. Wen W, Marcinkowski E, Luyimbazi D, Luu T, Xing Q, Yan J, et al. Eribulin Synergistically Increases Anti-Tumor Activity of an mTOR Inhibitor by Inhibiting pAKT/pS6K/pS6 in Triple Negative Breast Cancer. *Cells.* 2019;8(9):1010. doi: 10.3390/cells8091010
36. Yuan Y, Wen W, Yost SE, Xing Q, Yan J, Han ES, et al. Combination therapy with BYL719 and LEE011 is synergistic and causes a greater suppression of p-S6 in triple negative breast cancer. *Sci Rep.* 2019;9(1):7509. doi: 10.1038/s41598-019-43429-7
37. Sudhan DR, Guerrero-Zotano A, Won H, Ericsson PG, Servetto A, Huerta-Rosario M, et al. Hyperactivation of TORC1 Drives Resistance to the Pan-HER Tyrosine Kinase Inhibitor Neratinib in HER2-Mutant Cancers. *Cancer Cell.* 2020;37(2):258-9. doi: 10.1016/j.ccell.2020.01.010

Figures

Figure 1

Evaluation of p-S6 expression in lung SCC, lung ADC and Non-CLT

P-S6 was strongly positive in cell cytoplasm in lung SCC (A) and lung ADC (B). In Non-CLT (C), p-S6 staining was negative. (200×, IHC, DAB staining)

Figure 2

Comparison of p-S6 expression in metastatic NSCLC and matched primary cancer.

(A) P-S6 was strongly positive expressed in cell cytoplasm in primary lung SCC (A-a), primary lung ADC (A-b) and mild positive staining in its matched LNM lesions (A-c, A-d). (B) Whether it is lung SCC or lung ADC, the expression level of p-S6 in metastatic cancer foci is significantly higher than that in primary cancer foci ($p=0.001$, $p=0.022$, respectively).

Figure 3

Effect of RAD001 on expression of p-S6 protein in NSCLC cell lines(A549 and SPC-A1)

NSCLC Cells were treated with DMSO or RAD001 (5 nmol/L) for 24 h. In A549 and SPC-A1, compared with Control group, p-S6 protein in RAD001 groups were significantly decreased. Original blots are presented inSupplementary Figure 2.

Figure 4

p-S6 inhibition reduced wound healing of NSCLC cell lines(A549 and SPC-A1)

Low expression of p-S6 decreased the wound healing percentage of A549 and SPC-A1. * $p < 0.05$ and ** $p < 0.01$, Control vs. RAD001. (200 $\times$)

Figure 5

Effects of p-S6 inhibition on migration and invasion of NSCLC cell lines(A549 and SPC-A1)

A549 and SPC-A1 cells were treated with 5 nmol/L RAD001 and were subjected to transwell assay to detect cell migration and invasion activity. (A) Results showed that migration of cell lines, A549 and SPC-A1, decreased following treatment with RAD001. (B) Inhibition of p-S6 decreased the invasion ability of cell lines A549 and SPC-A1. *** $p < 0.001$, Control vs. RAD001. (200 $\times$)

Figure 6

Kaplan-Meier survival estimates in patients with lung SCC and ADC

(A) NSCLC patients with highly expressed p-S6 protein showed worse OS rate in comparison to patients with low p-S6 expression level ($p = 0.013$). (B) Compared to NSCLC patients in stage I-II, patients with stage III had worse OS rates ($p < 0.001$). (C) NSCLC patients with LNM owned poor prognosis compared with patients without LNM ($p < 0.001$). (D) NSCLC patients whose differentiation was poor had lower OS rate than those with well and moderate differentiation ($p = 0.003$).

Supplementary Files

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