

Genotype-by-environment Interaction in Holstein Heifer Fertility Traits using Single-step Genomic Reaction Norm Models

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ABSTRACT

30 **Background:** The effect of heat stress on livestock production is a worldwide
31 issue, where animal performance is influenced by exposure to high environmental
32 temperatures, indicating the existence of possible genotype-by-environment
33 interactions (G×E). The main objectives of this study were to (1) detect the time periods
34 in which heifer fertility traits are more sensitive to the exposure to high environmental
35 temperature and/or humidity, (2) investigate G×E due to heat stress in heifer fertility
36 traits, and (3) identify genomic regions associated with heifer fertility and heat stress in
37 Holstein cattle.

38 **Results:** Phenotypic records for three heifer fertility traits (i.e., age at first
39 calving, interval from first to last service, and conception rate at the first service) were
40 collected, from 2005 to 2018, for 56,998 Holstein heifers raised in 15 herds in the
41 Beijing area (China). By integrating environmental data including hourly air
42 temperature and relative humidity, the critical periods in which the heifers are more
43 sensitive to heat stress were defined as ≤30 days before the first service for age at first
44 calving and interval from first to last service, or 10 days before and ≤ 60 days after

45 the first service for conception rate. Using reaction norm models, significant G×E was
46 detected for all three traits regarding both environmental gradients, proportion of days
47 exceeding heat threshold and minimum temperature humidity index. Through single-
48 step genome-wide association study, *PLAG1*, *AMHR2*, *SP1*, *KRT8*, *KRT18*, *MLH1*, and
49 *EOMES* were suggested as candidate genes for heifer fertility while *HCRTR1*, *AGRP*,
50 *PC*, and *GUCY1B1* were for heat tolerance.

51 **Conclusions:** The critical periods in which reproductive performances of
52 heifers are more sensitive to heat stress are trait-dependent. Thus, detailed analysis
53 should be conducted to determine this particular period for other fertility traits. The
54 considerable magnitude of G×E and sire re-ranking indicates the necessity to consider
55 G×E in breeding schemes. This will enable selection of more heat-tolerant animals with
56 high reproductive efficiency under harsh climatic conditions. The candidate genes
57 identified to be linked with response to heat stress provide a better understanding of the
58 underlying biological mechanisms of heat tolerance in dairy cattle.

59

60 **Keywords:** heifer; heat stress; genotype by environment interaction; reaction norm;
61 single-step GWAS

62

63 **BACKGROUND**

64 In modern dairy cattle, female fertility is of great importance, due to its close
65 relationship with reproductive management, veterinary treatments, involuntary culling
66 and, eventually, the farm profitability [1]. However, as widely emphasized in previous

67 studies [2–4], the low heritability estimates for fertility traits and unfavorable genetic
68 correlations with milk production traits have led to reduced genetic progress for female
69 fertility traits. Moreover, the increase of joint genetic evaluation across farms in various
70 geographical locations emphasizes the role that genotype by environment interactions
71 (G×E) [5] might play, and consequently, the selection for animals (especially bulls) that
72 have progeny with high performance even in challenging environments. Significant
73 G×E for female fertility traits have been detected in several Holstein populations, where
74 the “E” were the production system and grass ratio of feed [6], and herd reproduction
75 level [7]. However, the investigation for other important environmental variations such
76 as the climatic change remain scarce.

77 With global warming and climatic change, heat stress has already become an
78 issue for livestock production in many countries around the world [8]. The temperature
79 and humidity index (THI) is often used as an environmental indicator to assess the
80 condition of heat stress in dairy cattle [9]. It is widely accepted that dairy cows start to
81 experience mild heat stress when THI surpasses 72 [10]. Studies of American Holstein
82 population have shown that heat conditions can lead to 165 kg loss of milk yield
83 annually and 0.4% reduce in milk fat percentage [11, 12], 0.85 kg decrease in feed
84 intake with one unit increase in air temperature [13], and about 15% decrease in
85 conception rate when THI surpasses 72 [14].

86 In summer (from June to August), the average daily THI, in many regions of
87 the world, can exceed 72, indicating that dairy cattle raised in these regions may suffer
88 from mild to severe heat stress [15]. For instance, in Beijing (China), the THI fluctuates

89 substantially within a day, that is, extremely high THI in the afternoon and dramatically
90 reduces to thermoneutral level in the evening). The difference in hourly THI within a
91 day can be up to 30 THI units during the late summer, but the daily average is usually
92 just relatively “mild” (Suppl. File 1). In such case, simply using the daily average of
93 THI may lead to the underestimation of the impact of heat stress. In addition to the
94 timing of a day, for a dairy cow, the timing of its reproductive period may also influence
95 the response to heat stress. Fertility performance may be compromised when an animal
96 experiences heat stress in certain physiological stages. Several studies have
97 demonstrated that the conception rate of dairy cows decreased when they experienced
98 heat stress before and after insemination [16–18], which emphasized the role of critical
99 periods exposed to heat conditions. To the best of our knowledge, no studies have
100 identified the most influential (critical) period for fertility traits due to their complex
101 characteristics. However, this is of utmost value for incorporating G×E models in
102 genetic and genomic evaluations.

103 Reaction norm models (RNM) are widely used to detect G×E when the
104 differences in environments can be measured by a continuous environmental gradient
105 (EG) [5]. In RNM, the breeding value of an animal is partitioned into an environment-
106 independent part (intercept) and an environment-dependent part (slope). The
107 relationship matrix of the RNM can be structured either by pedigree, i.e. pedigree-based
108 best linear unbiased predictor (BLUP), or combining both pedigree and genomic
109 information, i.e. single-step genomic BLUP (ssGBLUP) [19, 20]. On the basis of
110 ssGBLUP, Wang et al. [21] proposed a method termed single-step GWAS (ssGWAS)

111 which can obtain marker effects from estimated breeding values. Markers related with
 112 the intercept and slope could be mapped by applying ssGWAS procedures using RNMs.

113 The main objectives of this study were to: (1) explore the most heat-sensitive
 114 periods regarding heifer fertility (age at first calving – AFC, interval from first to last
 115 service – IFL, and conception rate of first service – CR); (2) detect G×E for heifer
 116 fertility traits using RNMs with pedigree-genomic combined relationship matrix; and
 117 (3) unravel genomic regions contributing to heat tolerance and heifer fertility traits in
 118 high-producing Holstein cattle.

119

120 RESULTS

121 Descriptive statistics

122 The summary statistics for heifer fertility traits are shown in Table 1. Large
 123 phenotypic variation was detected, especially for IFL (coefficient of variation equals to
 124 1.89) and CR (coefficient of variation equals to 0.83). The genetic parameters estimated
 125 using the conventional animal model, which were relatively low, are also provided in
 126 Table 1.

127

128 **Table 1** Descriptive statistics of heifer fertility traits and genetic parameters estimated
 129 using pedigree-based animal models

Trait ^a	N	Mean	SD	CV	Min	Max	σ_a^2	σ_e^2	h^2
AFC	56998	769.05	74.06	0.10	505	1100	794.40 (54.27)	4035.00 (46.35)	0.16 (0.011)
IFL	56998	29.25	55.17	1.89	0	365	190.42 (22.33)	2740.10 (23.82)	0.06 (0.007)
CR	56998	0.59	0.49	0.83	0	1	6.61e-3 (1.11e-3)	2.16e-1 (1.57e-3)	0.03 (0.005)

130 ^a AFC = age at first calving, IFL = interval from first to last service, CR = conception
131 rate of first service.

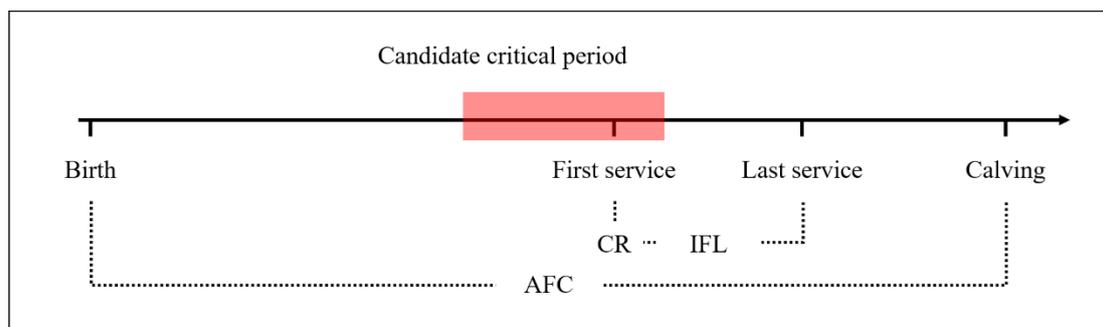
132

133 **Critical period selection for each environmental gradient scenario**

134 Two heat related EGs were used in the current study: 1) the number of days that
135 exceeded the THI threshold in the evaluated critical period (prop-EG); 2) the minimum
136 THI for each day of the candidate period (mTHI-EG). To avoid the underestimation of
137 the heat stress effect, the days in which the hourly THI was higher than 72 for six
138 continuous hours were considered as heat-stress days for prop-EG. Akaike Information
139 Criteria (AIC) [22] was obtained for various time combinations to select the best fit
140 period for each trait.

141 The critical periods (Fig. 1) selected for each trait and EG under scenario one
142 (S1) and scenario two (S2) are listed in Table 2. The same 60 days, from 30 days before
143 the first insemination to 30 days after the first insemination, were chosen as the control
144 period for S1. For S2, critical periods ranged from 30 to 70 days, of which only the
145 period (-90, -30) for IFL was the same for both EGs. Only the critical periods of CR
146 end after the first service (60 or 30 days). The detailed results of AICs for the 19 tested
147 combinations are presented in Supp. File 2.

148



149 **Fig. 1** Reproductive events and the definition of critical period in heifers. The red
 150 rectangle represents the critical period, defining as the time period of which heifers are
 151 likely to suffer from heat stress. AFC = age at first calving, IFL = interval from first to
 152 last service, CR = conception rate of first service.

153

154 **Table 2** The critical periods selected for each trait in different scenarios

EG ^a	Trait ^b	Scenario ^c	Number of days	Period ^d
Prop-EG	AFC	S1	60	(-30, 30)
		S2	60	(-90, -30)
	IFL	S1	60	(-30, 30)
		S2	60	(-90, -30)
	CR	S1	60	(-30, 30)
		S2	70	(-10, 60)
mTHI-EG	AFC	S1	60	(-30, 30)
		S2	30	(-90, -60)
	IFL	S1	60	(-30, 30)
		S2	60	(-90, -30)
	CR	S1	60	(-30, 30)
		S2	40	(-10, 30)

155 ^a prop-EG = the number of days that exceeding the threshold temperature humidity
 156 index in the period; mTHI-EG = minimum temperature humidity index for each day
 157 of the period

158 ^b AFC = age at first calving; IFL = interval from the first to last service; CR =
 159 conception rate of first service

160 ^c S1 = control critical period; S2 = periods selected based on the Akaike's information
 161 criterion

162 ^d Periods were counted based on the first service day; minus means before and plus
 163 means after.

164

165 The definitions of two types of EGs had some overlaps. For example, prop-EG
166 would be recorded as 1 if minimum THI of all the days in critical period were above
167 67.02 (Table 3). To calculate the overlap rate between prop-EG and mTHI-EG, top
168 animals sorted by genomic estimated breeding values (gEBV), with estimation
169 accuracy greater than 0.4 (average accuracy for three traits), were chosen for each trait
170 with regards to each EG. When using the *H* matrix, approximately 75% of the heifers
171 were the same in both scenarios for AFC and IFL, but relatively low (29.63% ~ 65.82%)
172 overlap rates were observed in CR (Table 3). Similar results were found when using
173 the *A* matrix (Supp. File 3).

174

175 **Table 3** The proportions of overlapped top 1% heifers^a from using prop-EG and
176 mTHI-EG as environmental gradients (EGs) by reaction norm models (RNM) with
177 the *H* matrix in Holstein cattle.

EG ^b		AFC ^c		IFL		CR	
Prop-EG	mTHI-EG	S1 ^d	S2	S1	S2	S1	S2
0.2	43.03	76.47%	82.69%	72.92%	64.37%	29.63%	36.63%
0.4	48.29	84.57%	87.35%	81.24%	77.69%	59.60%	51.50%
0.6	52.34	88.24%	82.02%	85.68%	78.91%	65.82%	60.93%
0.8	55.54	83.02%	75.47%	80.69%	75.47%	65.70%	62.93%
1	67.02	82.24%	76.03%	79.69%	74.92%	64.04%	60.27%

178 ^a Heifers were selected based on gEBV and accuracy of estimation (> 0.4)

179 ^b prop-EG = using the number of days that exceeding the threshold temperature
 180 humidity index in the period as EG; mTHI-EG = using the minimum temperature
 181 humidity index of a day of the critical period as EG

182 ^c AFC = age at first calving; IFL = interval from the first to last service; CR =
 183 conception rate of first service

184 ^d S1 = reference period; S2 = periods selected based on the Akaike's information
 185 criterion

186

187 **(Co)variance Components and G×E**

188 The estimates of (co)variance components obtained from RNMs with different
 189 kinship matrices (*A* or *H*) were similar for all traits analyzed. The correlation
 190 coefficients between the intercept and slope for each trait were all negative and ranged
 191 from -0.25 (IFL in S1 of prop-EG) to -0.98 (CR in both S1 and S2 of mTHI-EG) using
 192 the *H* matrix (Table 4). Furthermore, the absolute value of coefficients estimated using
 193 prop-EG were relatively small than those using mTHI-EG, especially for AFC and IFL.
 194 The genetic parameters estimated by *A* matrix are shown in Supp. File 4.

195

196 **Table 4** Variances of the intercept ($\sigma_{a_0}^2$) and slope ($\sigma_{a_1}^2$), the covariance between the
 197 intercept and slope ($\sigma_{a_0a_1}$), residual variance (σ_e^2) and genetic correlation between the
 198 intercept and slope ($r_{a_0a_1}$), with their standard errors in parentheses, estimated using
 199 reaction norm models with *H* matrix in Holstein cattle.

EG ^a	Trait ^b	Scenario ^c	$\sigma_{a_0}^2$	$\sigma_{a_1}^2$	$\sigma_{a_0a_1}$	σ_e^2	$r_{a_0a_1}$
-----------------	--------------------	-----------------------	------------------	------------------	-------------------	--------------	--------------

Prop-EG	AFC	S1	971.97 (62.74)	0.53 (0.05)	-9.62 (1.41)	3413.70 (45.80)	-0.43 (0.02)
		S2	963.72 (60.99)	0.96 (0.06)	-14.05 (1.62)	3203.00 (44.60)	-0.46 (0.01)
	IFL	S1	218.16 (26.82)	0.25 (0.03)	-1.80 (0.68)	2456.90 (25.14)	-0.25 (0.03)
		S2	236.35 (27.59)	0.51 (0.03)	-4.98 (0.87)	2323.10 (24.76)	-0.46 (0.02)
	CR	S1	1.01e-2 (1.60e-3)	1.60e-5 (2.00e-6)	-3.35e-4 (5.20e-5)	1.99e-1 (1.61e-03)	-0.83 (0.05)
		S2	1.12e-2 (1.71e-3)	1.30e-5 (2.00e-6)	-3.23e-4 (4.70e-5)	1.98e-1 (1.62e-3)	-0.84 (0.05)
mTHI-EG	AFC	S1	2735.10 (233.57)	1.37 (0.12)	-51.60 (5.10)	3396.80 (45.06)	-0.84 (0.03)
		S2	3950.90 (271.77)	2.06 (0.12)	-81.46 (5.69)	3194.70 (43.73)	-0.90 (0.03)
	IFL	S1	1073.60 (126.34)	0.78 (0.07)	-26.15 (2.92)	2417.70 (25.10)	-0.90 (0.04)
		S2	1591.40 (143.27)	1.24 (0.08)	-41.85 (3.35)	2309.10 (24.37)	-0.94 (0.03)
	CR	S1	7.78e-2 (9.60e-3)	4.30e-5 (5.00e-6)	-1.81e-3 (2.14e-4)	1.99e-1 (1.58e-3)	-0.98 (0.06)
		S2	8.90e-2 (1.03e-2)	4.50e-5 (5.00e-6)	-1.96e-3 (2.19e-4)	1.98e-1 (1.59e-3)	-0.98 (0.05)

200 ^a prop-EG = the number of days that exceeding the threshold temperature humidity

201 index in the period; mTHI-EG = minimum temperature humidity index for each day

202 of the period

203 ^b AFC = age at first calving; IFL = interval from the first to last service; CR =

204 conception rate of first service

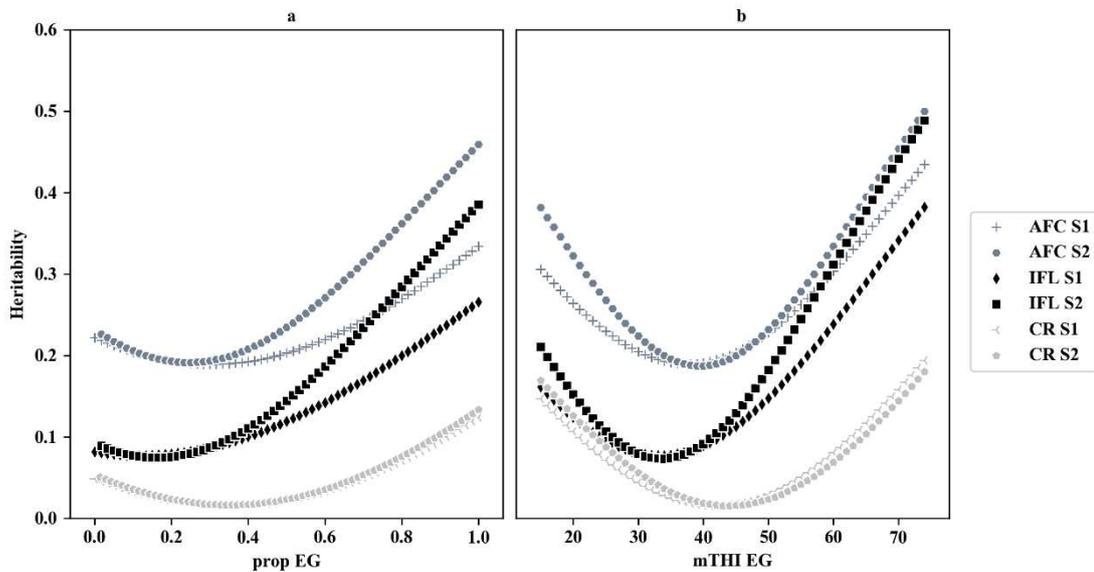
205 ^c S1 = reference period; S2 = periods selected based on the Akaike's information

206 criterion

207

208 Heritabilities estimated from genomic RNM using prop-EG and mTHI-EG are
 209 presented in Fig 2. Generally, AFC had the highest heritability estimates, whereas CR
 210 was the least heritable across all EGs. The shapes of heritability curves were similar
 211 when using different relationship matrices but different across EGs. The curve patterns
 212 were quadratic for mTHI-EG, which indicated that the highest heritabilities were
 213 generally observed in either cold (mTHI-EG < 20) or heat-stress environments (mTHI-
 214 EG > 72). However, the patterns were flatter when prop-EG was used, and the highest
 215 heritabilities only appeared in heat stress conditions. Similar curve patterns were
 216 observed when using the *A* matrix (Suppl File 5).

217

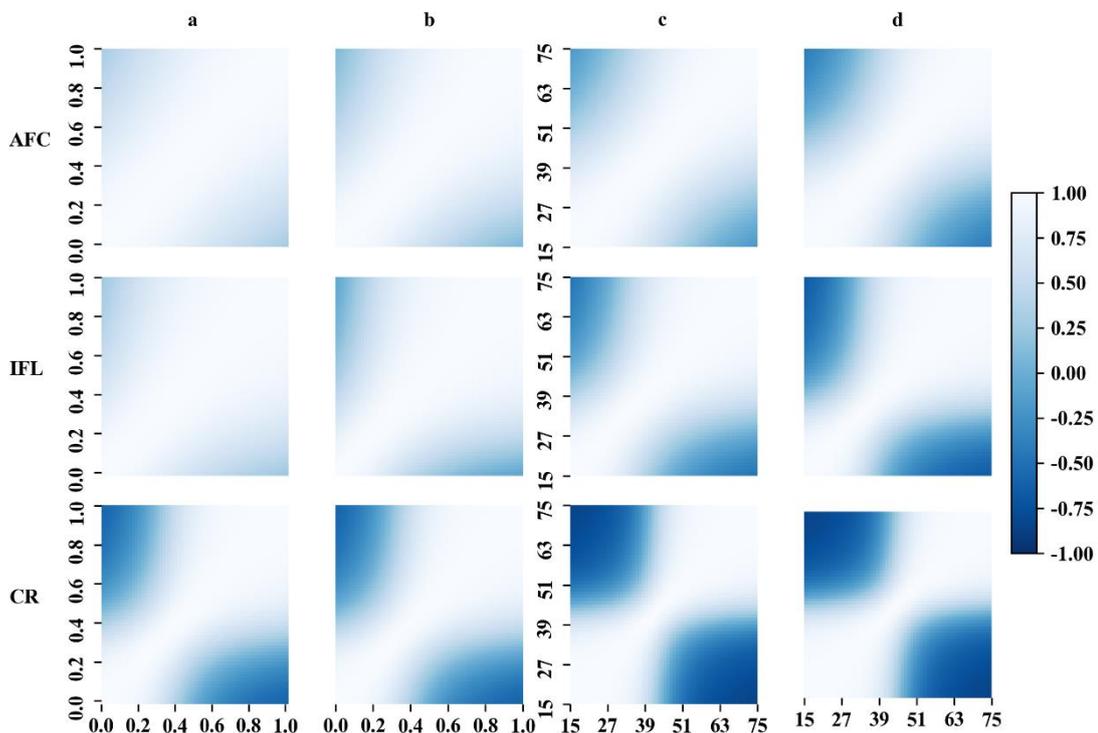


218

219 **Fig. 2** Heritabilities estimated by reaction norm models with the matrix *H* using **a)**
 220 prop-EG or **b)** mTHI-EG as environmental gradient. For **a)**, the x-axis is the proportion
 221 of days exceeding the threshold with a range of 0 to 1; while for **b)**, the x-axis is the
 222 minimum THI with a range of 15 to 75.

223

224 As shown in Table 4, for all traits, the variance of the slope was significantly different
 225 from zero based on a one-tailed test ($P < 0.01$), indicating the existence of $G \times E$. Genetic
 226 correlations between different EGs, from RNM with the H matrix, are shown in Fig. 3.
 227 Generally, the more different the EGs, the less they were correlated. More negative
 228 coefficients of correlation were obtained for AFC and IFL when the mTHI-EG was
 229 evaluated, compared with prop-EG. This is consistent with much stronger correlation
 230 between the intercept and slope being observed when using mTHI-EG as EG compared
 231 to using prop-EG as EG. Similar patterns were also observed when using A matrix
 232 (Suppl File 6).
 233



234
 235 **Fig. 3** Genetic correlations estimated by reaction norm models (RNMs) with the matrix
 236 H . The color indicates the magnitude of the genetic correlation. **a**) Correlations between
 237 different levels of prop-EG estimated from RNM under S1. The x-axis and y-axis are

238 the proportion of days exceeding the threshold, ranging from 0 to 1. **b)** Correlations
 239 between different levels of prop-EG estimated from RNM under S2. The x-axis and y-
 240 axis are the proportion of days exceeding the threshold, ranging from 0 to 1. **c)**
 241 Correlations between different levels of mTHI-EG estimated from RNM under S1. The
 242 x-axis and y-axis are the minimum THI, ranging from 15 to 75. **d)** Correlations between
 243 different levels of mTHI-EG estimated from RNM under S2. The x-axis and y-axis are
 244 the minimum THI, ranging from 15 to 75.

245

246 Among the top sires with more than 20 daughters with phenotypes, the number
 247 of sires overlapping across the two EGs, reflecting the magnitude of the re-ranking of
 248 sires, are listed in Table 5. The number of overlapped sires decreased as the EGs became
 249 more divergent, especially for CR (e.g. from 11 to 1 in S2 of prop-EG). The magnitude
 250 of re-ranking increased when using mTHI-EG (only 3 common sires in all
 251 environmental combinations).

252

253 **Table 5** The number of common animals among the top 50 sires between 2 levels
 254 of EGs

EG ^a	Trait ^b	Scenario ^c	1 vs. 99% ^d	5 vs. 95%	10 vs. 90%	25 vs. 75%
Prop-EG	AFC	S1	18	20	21	28
		S2	13	14	17	28
	IFL	S1	18	20	21	29
		S2	12	14	17	27
	CR	S1	0	0	1	7
		S2	1	1	3	11
mTHI-EG	AFC	S1	5	8	15	25
		S2	3	4	9	24

	IFL	S1	3	5	6	22
		S2	0	0	2	22
	CR	S1	0	0	0	3
		S2	0	0	0	3

255 ^a prop-EG = the number of days that exceeding the threshold temperature humidity
 256 index in the period; mTHI-EG = minimum temperature humidity index for each day
 257 of the period

258 ^b AFC = age at first calving; IFL = interval from the first to last service; CR =
 259 conception rate of first service

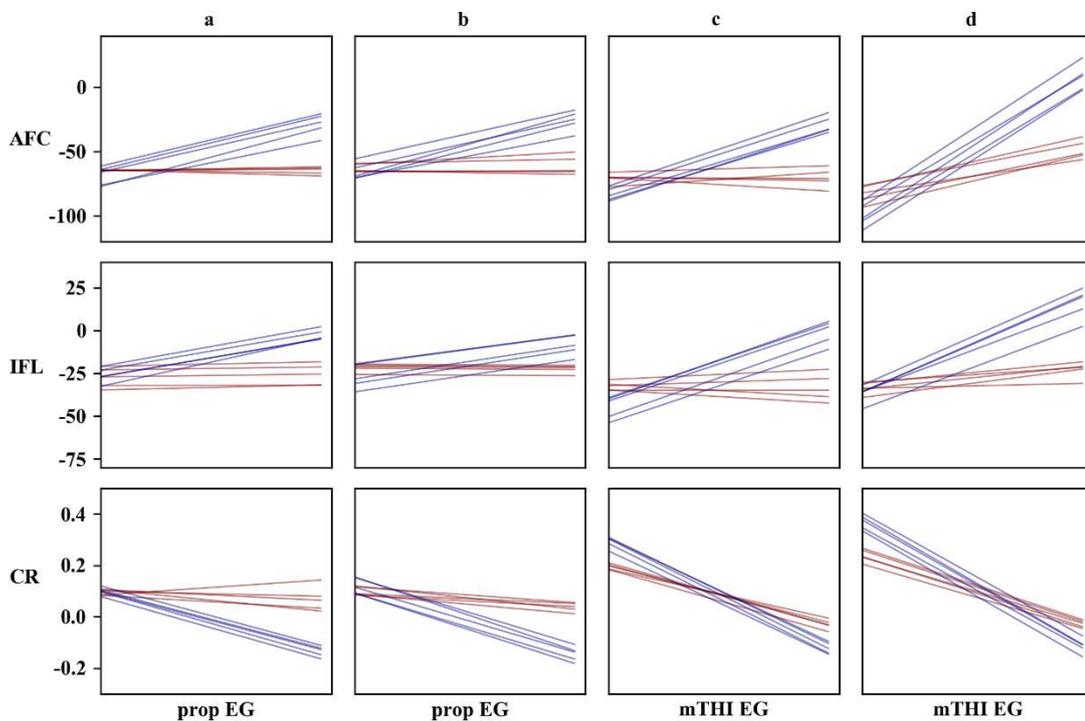
260 ^c S1 = reference period; S2 = periods selected based on the Akaike's information
 261 criterion

262 ^d the number of overlapping animals in the top sires in the 1 and 99%, 5 and 95%, 10
 263 and 90%, and 25 and 75% quantiles of EGs

264

265 We further visualized breeding value re-ranking by plotting gEBV of sires with
 266 the most preferential intercepts (gEBV less than average minus two times standard
 267 deviation for AFC and IFL; gEBV greater than average plus two times standard
 268 deviation for CR) in Fig. 4. The top 5 sires with the flattest slopes (more climatic
 269 resilient) were drawn in red, while the top 5 sires with the steep slopes (more climatic
 270 sensitive) were drawn in blue. In this case, sires that are sensitive to the environments
 271 (blue lines), would perform worse than those had flat slopes (red lines) in heat stressed
 272 conditions. For instance, the gEBV of CR is 0.10 when prop-EG is 0, but the gEBVs
 273 for blue lines decreased to around -0.15 when prop-EG is 1. Meanwhile, the gEBVs of
 274 the red lines were stable along the whole prop-EG (Fig. 4a). This further verified the

275 existence of G×E regarding the change of mTHI-EG and/or prop-EG. Larger changes
 276 were observed for gEBVs when using mTHI-EG. Implementing mTHI-EG, gEBVs of
 277 IFL for two bulls increased from around -50 day in thermoneutral condition to 0 day in
 278 heat stress condition (Fig. 4c-d), which is nearly twice the change as gEBVs using prop-
 279 EG.
 280



281
 282 **Fig. 4** The re-ranking plots for gEBVs of sires. The blue and red lines represent
 283 sensitive and resilient sires, respectively. **a)** Re-ranking plots for three traits estimated
 284 using prop-EG under S1. The x-axis is the proportion of days exceeding the threshold
 285 with a range of 0 to 1 and y-axis is gEBV. **b)** Re-ranking plots for three traits estimated
 286 using prop-EG under S2. The x-axis is the proportion of days exceeding the threshold
 287 with a range of 0 to 1 and y-axis is gEBV. **c)** Re-ranking plots for three traits estimated
 288 using mTHI-EG under S1. The x-axis is the minimum THI with a range of 15 to 75 and

289 y-axis is gEBV. **d)** Re-ranking plots for three traits estimated using mTHI-EG under
290 S2. The x-axis is the minimum THI with a range of 15 to 75 and y-axis is gEBV

291

292 **Single-step genome-wide association analyses**

293 Overall, similar genomic regions were detected to be associated with the same
294 trait when using two scenarios of prop-EG, especially for CR (Figs. 5 and 6). For S1,
295 nine regions were shared for both the intercept and the slope for AFC, among which two
296 (from 26,669,442 to 26,802,092 and from 26,803,676 to 26,880,091 bp) were located
297 in BTA14 and three regions (from 24,762,252 to 25,487,353 bp, from 106,901,044 to
298 106,946,812 bp, and from 106,948,226 to 106,980,536 bp) in BTA5, respectively. The
299 overlapping region that explained the highest average variance (0.92% for the intercept
300 and 2.30% for the slope) was in BTA14 (from 26,803,676 to 26,927,342 bp). Similarly,
301 the same region (from 26,821,555 to 26,899,089 bp), which is one of the four shared
302 genomic windows, explained 1.12% and 0.91% genetic variance for the intercept and
303 slope of IFL, respectively. For CR, 17 regions were in common between using THI and
304 prop-EG variables in RNM, and a narrower region (from 26,819,709 to 26,888,221 bp)
305 in BTA14, which explained 1.83% and 1.72% genetic variance for the intercept and
306 slope, respectively, was located in the same region detected in AFC and IFL.

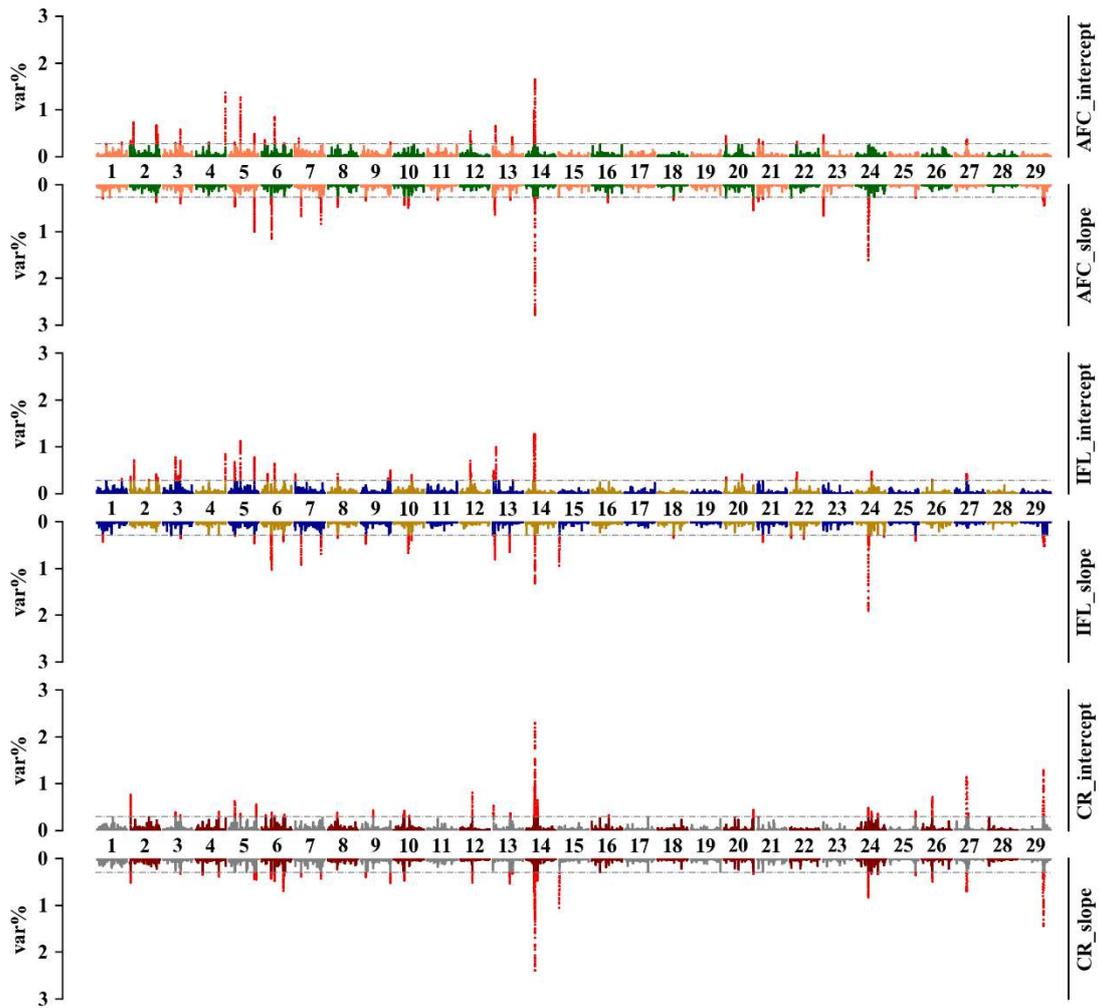
307 The genomic windows explaining the highest variance were not connected for
308 AFC and IFL under S2. However, the area from 26,819,709 to 26,887,021 bp that
309 explained the highest proportion of the total additive genetic variance (2.38% and
310 2.29% for the intercept and slope, respectively) for CR, was still located in BTA14. We

311 detected 21 overlapping genomic windows for CR between two variables, which is
312 more than detected for AFC and IFL (4 and 13, respectively).

313 More shared genomic regions were detected when the same variables (the
314 intercept or slope) of the two scenarios were tested. For AFC and IFL, more than 10
315 genomic areas were connected in the absence of regions explained the most genetic
316 variance. However, the longest shared region in BTA14 was still detected for both the
317 intercept (from 26,819,709 to 26,887,021 bp) and the slope (from 26,821,555 to
318 26,888,221 bp) for CR. Similarly, more than 25 overlapping areas were mapped for
319 each variable of CR.

320 The Manhattan plots of mTHI-EG are provided in Supp. Files 7 and 8. Basically,
321 more shared regions were mapped when using mTHI-EG compared to prop-EG, but
322 the most associated genomic regions for each trait were found to be distributed in
323 different chromosomes. Detailed information for genomic regions is listed in Supp.
324 Files 9 and 10.

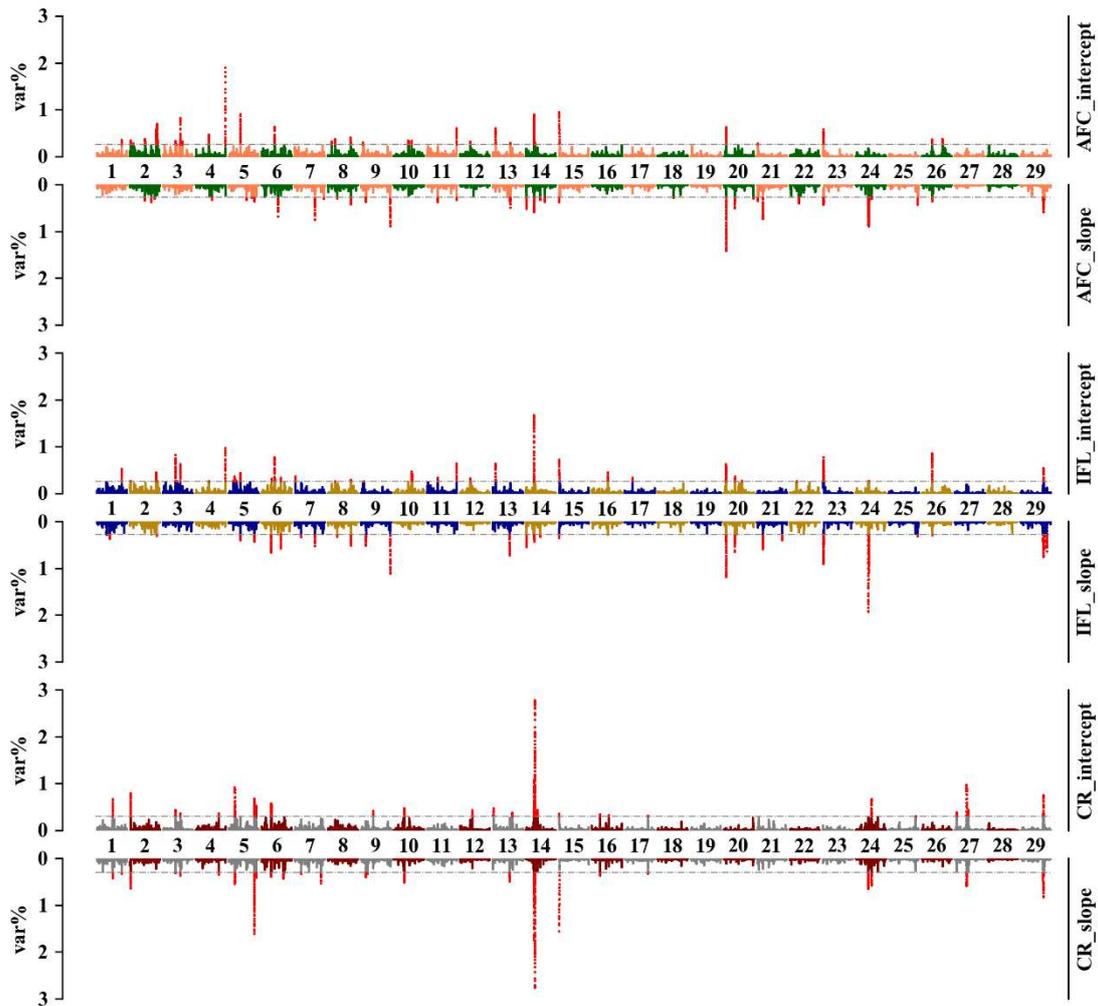
325



326

327 **Fig. 5** Percentages of the intercept and slope genetic variances explained by a sliding
 328 window of 20 SNPs for three traits, which were estimated under scenario one of prop-
 329 EG. The x-axis is autosome segments; the y-axis represents the proportion of explained
 330 variances; the grey horizontal lines are thresholds (top 0.5%) for candidate genomic
 331 regions; different color sets indicate different traits

332



333

334 **Fig. 6** Percentages of the intercept and slope genetic variances explained by a sliding
 335 window of 20 SNPs for three traits, which were estimated under scenario two of prop-
 336 EG. The x-axis is autosome segments; the y-axis represents the proportion of explained
 337 variances; the grey horizontal lines are thresholds (top 0.5%) for candidate genomic
 338 regions; different color sets indicate different traits.

339

340 The mapped positional candidate genes are shown in Table 6 and Supp. Files 9
 341 and 10. Candidate genomic regions of the intercept term were previously linked to
 342 several types of quantitative trait loci (QTL) such as milk kappa-casein percentage,
 343 metabolic body weight, average daily gain, length of productive life, dry-matter intake,

344 conception rate, and pregnancy rate (Supp. Files 9 and 10). Most of the mapped QTLs
345 are associated with production traits, and the rest are associated with reproduction,
346 health and meat/carcass traits. The identified biological processes ($P < 0.05$) related to
347 heifer reproduction were: developmental process involved in reproduction, oocyte
348 maturation, oocyte development, oocyte differentiation, oogenesis, placenta blood
349 vessel development, and embryo development. Two identified pathways were related
350 to stress response: regulation of response to stress and response to oxidative stress.
351 Other pathways included muscle and body development.

352 Candidate genomic regions of slope term have been previously linked to a
353 variety of trait groups, including luteal activity, body weight, stillbirth, and many milk-
354 related QTLs. Similarly, most of the QTLs identified were associated with milk
355 production traits, because most of the QTLs overlapped between the two genetic terms.
356 The reproductive biological processes identified ($P < 0.05$) using the slope term were:
357 reproductive processes, fertilization, sexual reproduction, granulosa cell differentiation,
358 oocyte development, acrosome reaction, oocyte differentiation, and regulation of
359 luteinizing hormone secretion (Supp. Files 9 and 10). Additionally, more potential
360 stress-related pathways were identified such as response to abiotic stimulus, detection
361 of stimulus involved in sensory perception, response to temperature stimulus, response
362 to radiation, negative regulation of saliva secretion, aerobic respiration, and energy
363 derivation by oxidation of organic compounds.

364

365 **Table 6** Genes and QTLs for top genomic regions of two environmental gradients.

EG ^a	Trait ^b	Covariable	Chromosome	Genes	Var%	#QTL	QTL traits
Prop-EG	AFC	slope	BTA14	<i>CLVSI</i>	2.30	1	Milk unglycosylated kappa-casein percentage
	CR	intercept	BTA14	<i>CLVSI</i>	2.29	3	Milk kappa-casein percentage
	CR	slope	BTA14	<i>CLVSI</i>	1.83	1	Milk kappa-casein percentage
	CR	intercept	BTA14	<i>CLVSI</i>	1.72	2	Milk kappa-casein percentage
	AFC	slope	BTA14	<i>LOC112449637</i>	1.49	1	Milk unglycosylated kappa-casein percentage
	CR	slope	BTA5	<i>FKBP4, DDX11</i>	1.20	3	Bovine tuberculosis susceptibility, Milk kappa-casein percentage
mTHI-EG	AFC	intercept	BTA14	<i>CLVSI</i>	2.27	1	Milk glycosylated kappa-casein percentage
	AFC	slope	BTA14	<i>CLVSI</i>	2.03	1	Milk unglycosylated kappa-casein percentage
	AFC	slope	BTA14	<i>LOC112449637</i>	1.80	1	Milk unglycosylated kappa-casein percentage
	CR	intercept	BTA14	<i>CLVSI</i>	1.39	3	Milk kappa-casein percentage
	CR	slope	BTA5	<i>FKBP4, DDX11</i>	1.35	2	Milk kappa-casein percentage

CR	slope	BTA14	<i>CLVSI</i>	1.34	3	Milk kappa-casein percentage
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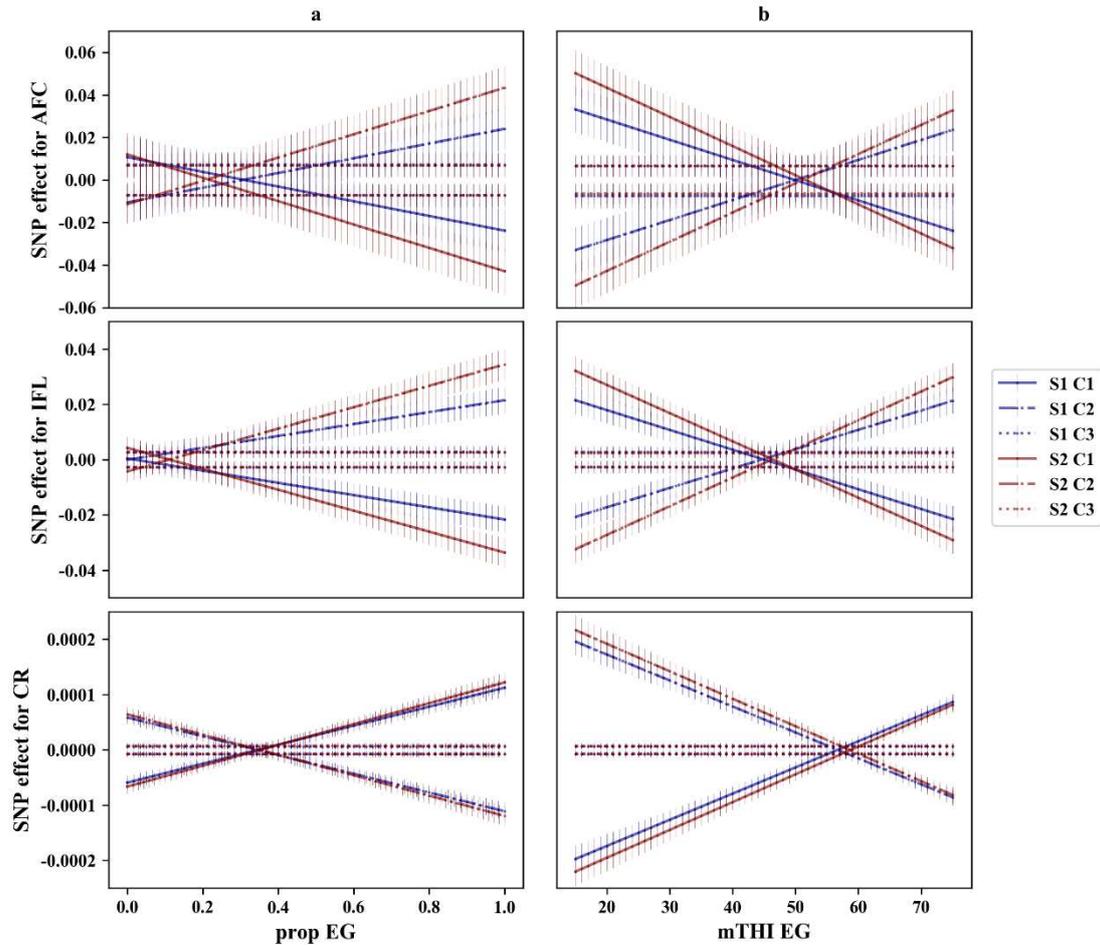
366 ^a prop-EG = the number of days that exceeding the threshold temperature humidity
367 index in the period; mTHI-EG = minimum temperature humidity index for each day
368 of the period

369 ^b AFC = age at first calving; IFL = interval from the first to last service; CR =
370 conception rate of first service

371

372 **Cluster analysis for SNP effect trajectories**

373 The pattern of SNP effects over different EGs of each trait, scenario, and cluster
374 are shown in Fig. 7. The SNP effects remained at a specific level within each trait, and
375 the effects of CR were almost 100 times less than those of AFC and IFL. The magnitude
376 of SNP effects changes was higher for S2 (red lines) in all traits and EGs in C1 and C2,
377 whereas the SNP effects in C3 were similar in each trait and scenario. The cross point
378 of different clusters appeared later when mTHI-EG was used. Furthermore, lower
379 standard deviations were observed for CR in different scenarios and clusters.



380

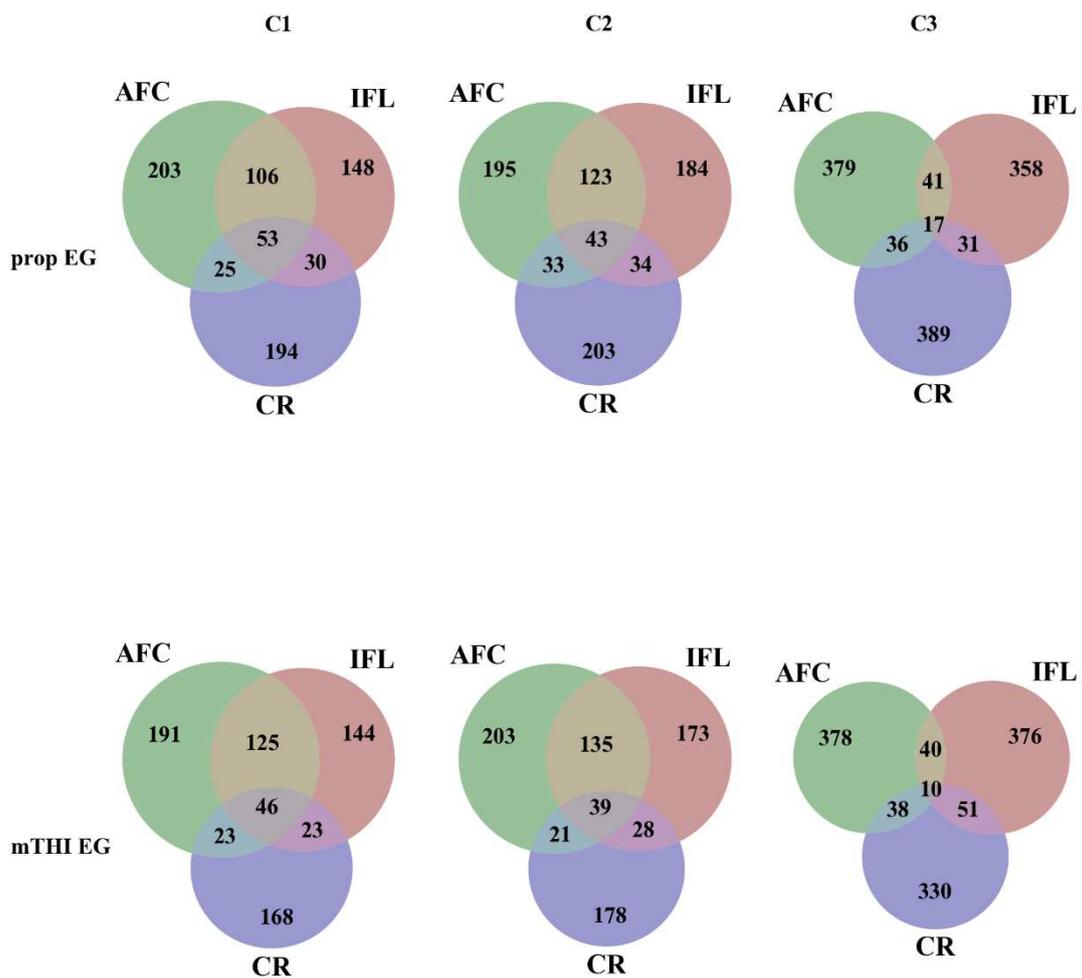
381 **Fig. 7** Trajectories of SNP effects changing over EGs. The x-axis is environmental
 382 gradient; the y-axis represents the SNP effects; vertical bar is the standard deviations
 383 of SNP effects at each level of EG; blue lines indicate scenario one; red lines indicate
 384 scenario two; different color sets indicate different clusters. **a)** Trajectories of SNP
 385 effects changing over prop-EG. The x-axis is the proportion of days exceeding the
 386 threshold with a range of 0 to 1. **b)** Trajectories of SNP effects changing over mTHI-
 387 EG. The x-axis is the minimum THI with a range of 15 to 75.

388

389 Approximately 50 overlapped genes were identified in C1 and C2, whereas 10
 390 or 17 shared genes were detected in C3. However, over 330 positional genes were

391 detected within each trait in C3, which is nearly twice the number of genes mapped in
 392 C1 and C2 (Fig. 8). Additionally, the number of overlapping genes between AFC and
 393 IFL was found to be higher than those shared with CR in C1 and C2. A total of 149
 394 common genes were identified in different clusters, among which 50 genes overlapped
 395 between prop-EG and mTHI-EG.

396



397

398 **Fig. 8** Number of shared candidate genes for each EG in different traits and clusters.
 399 C1 = SNP effects changes in preferential ways (decrease for AFC and IFL; increase for
 400 CR); C2 = SNP effects changes in the opposite ways (increase for AFC and IFL;
 401 decrease for CR); C3 = constant SNP effects over time.

402

403 **DISCUSSION**

404 **Heritabilities estimated from the conventional animal model**

405 The heritability estimates were 0.03 (0.005) for CR, 0.06 (0.007) for IFL and
406 0.16 (0.011) for AFC (Table 1). The heritability of AFC in our study was at a similar
407 level as that estimated previously (0.07–0.24) [23]. For heifer IFL and CR, the
408 heritabilities observed agree with earlier studies in Irish Holstein population [23] and
409 in the same Chinese Holstein populations [4]. The current study showed that heritability
410 estimates for interval traits (AFC and IFL) were generally higher than for the binary
411 trait (CR), which is also consistent with several studies [7, 24, 25].

412

413 **The influence of heat stress on heifer fertility traits**

414 Several studies have demonstrated that heat stress may have adverse effects on
415 heifer fertility in two aspects [26]: (1) follicular development or estrus, and (2)
416 pregnancy. The percentage of Holstein heifers having two follicular waves in the heat-
417 stressed group (33 °C, 60% relative humidity) and thermoneutral group (21 °C, 60%
418 relative humidity) have been shown to be 18% and 91%, respectively; the day of
419 functional luteolysis was delayed for almost 9 days in the heat-stressed group,
420 compared with the controlled group [27]. Sakatani et al. [28] found that the estrus
421 detection rate of non-lactating Japanese Black cows was significantly lower in summer,
422 whereas another study of lactating Holsteins also illustrated that the estrus detection
423 rate decreased when THI exceeded 72 [14]. Follicular development and estrus detection

424 are negatively affected by heat stress, which extends AFC and IFL. Another analysis of
425 20,606 cows also showed that pregnancy rates declined when THI was greater than 72,
426 with a decrease in pregnancy rate of 1.03% per unit increase of THI [29]. In the current
427 study, a dramatic decrease in the month of July was observed for the CR phenotypes
428 (Suppl. File 11), which is consistent with the results of previous studies.

429 The critical periods found under S2 of CR, on the basis of first insemination day,
430 were (-10, 60) and (-10, 30) for prop-EG and mTHI-EG, respectively, which are similar
431 to the values reported in previous studies. Khan et al. [17] explored the effects of heat
432 stress on the pregnancy rate using 1,100 crossbred dairy cows in India and reported that
433 pregnancy rates decreased significantly when cows experienced high THI (>72) within
434 a period of at least 30 days before and after the insemination day. For AFC and IFL,
435 the critical period was earlier before the first service: (-90, -30) or (-90, -60). This may
436 be partly due to the delay of first insemination; that is, follicular development or estrus
437 detection may be compromised by heat stress. A study evaluating the whole cycle of
438 estrus showed that heat stress inhibits the development of the dominant follicle during
439 preovulatory period in heifers [27]. Thus, the onset of estrus for heifers would be
440 delayed to the next cycle due to the delayed development of follicles, and heat stress
441 actually impacts the heifers earlier before the first insemination. Another possible
442 explanation is that heat stress may affect puberty of heifers. The age at puberty is
443 generally 13 months, and the average age at first service in this study was 491.95 days
444 (around 16 months) with a standard error of 72.48 days. This may explain the reason
445 that the boundary of the critical periods for AFC and IFL was found to be 90 days (3

446 months) before the first insemination. However, additional studies are needed to
447 validate this relationship.

448 To the best of our knowledge, the critical periods of heat stress for heifer
449 reproduction traits have not been well defined, and most studies focused solely on
450 pregnancy rate or CR. Amundson et al. [30] detected negative associations of THI with
451 pregnancy rate for *Bos taurus* crossbred cows in all three breeding periods: (0, 21), (0,
452 42) and (0, 60). Another study indicated that a high heat load 3-5 weeks before and 1
453 week after service was associated with reduced CR in cattle [31]. In [18], the CR of
454 lactating dairy cows was negatively affected by heat stress both before and after the day
455 of insemination (-42, 31), with the greatest negative impact in (-21, -1). The results of
456 the current research indicated that the critical period of heat stress is trait-related, and
457 the periods we identified for CR are within the range reported in previous studies, which
458 provides substantial support for the results of AFC and IFL.

459 The overlapping test of the two EGs under two scenarios (Table 3) indicated
460 that prop-EG and mTHI-EG capture different mechanisms. For instance, prop-EG
461 equaled 0.5 in a period of 60 days when the THI of any 30 days in this period exceeded
462 the threshold. However, the average daily THI of these 30 days may be 73 or 78 (more
463 severe heat stress). Similarly, prop-EG equaled 0 when THI did not exceed the
464 threshold for all the days in a period, but these days may occur in autumn or winter
465 (cold stress). Consequently, prop-EG could not assess cold or severe heat conditions as
466 mTHI-EG; it represents the average heat load of heifers, i.e., the proportion of days
467 experiencing heat stress conditions, which would not include all the climatic variation

468 due to the use of a THI threshold. Compared with AFC and IFL, CR may be more
469 sensitive to climatic changes, as it produced fewer overlapping animals between the
470 two EGs.

471

472 **Variance components estimated from RNMs and $G \times E$**

473 We estimated variance components using RNMs with the matrices A and H
474 (Table 4 and Supp. File 4). The genetic variances obtained from conventional RNMs
475 and those from genomic RNMs were similar for all traits and agree with the results of
476 previous studies (e.g., [7, 32]). Heritabilities estimated from RNMs were generally at
477 similar levels, compared with those estimated from conventional animal models (Fig.
478 2). Zhang et al. [7] detected heterogeneities in heritabilities across different EGs for
479 fertility traits from both matrices, which is consistent with our study. A recent study
480 found the genetic variances of production traits varied in different THI levels, and a
481 quadratic curve was observed in the heritabilities of protein yield [8]. In Danish
482 Holstein populations, heterogeneities of heritabilities of several fertility traits were
483 observed across different production levels or grass ratio in feed [6, 33]. Although
484 heritabilities vary across EGs in different analyses, patterns of heritabilities with
485 changes in the environment are predictable. Various environmental indicators and
486 analyzed traits could account for this result. Heterogeneities in heritabilities provide the
487 insight that future genetic evaluations should consider different environmental
488 variances to obtain accurate results. Incorporating heat stress into breeding objectives

489 could be helpful for the correct selection of animals in different environments,
490 especially in countries where climatic conditions are highly variable.

491 Based on the t-test for the variance of the slope, a statistically significant $G \times E$
492 was observed for all traits in this study [5]. Moderate to strong negative correlations
493 were detected between the intercept and slope for each trait. This negative relationship
494 was also found by other researchers who estimated the variance components of cow
495 fertility traits using RNM [6, 7]. Previous studies have suggested that a low correlation
496 between the intercept and slope could indicate re-ranking of animals across different
497 environments [34, 35]. In addition, Liu et al. [6] illustrated that this negative correlation
498 could increase the magnitude of $G \times E$. Collectively, robust animals with preferential
499 intercepts and flat slopes are able to perform well across various environments [36].
500 Based on RNMs, individual breeding values for each THI condition can be
501 appropriately estimated to enable selection of heat tolerant cows. These estimates could
502 be more accurate for bulls whose daughters have different records under different
503 environments. Genetic correlations between different EGs estimated using
504 conventional and genomic RNMs were almost the same (Fig 3 and Supp. File 5). The
505 larger the difference between the two EGs, the less they were correlated, which is
506 consistent with studies using different EGs [6, 7, 37]. This indicates that the re-ranking
507 of sires may occur in different EGs, as indicated in Table 5. The re-ranking was more
508 obvious for CR, compared with the other two traits, indicating that CR has a higher
509 sensitivity under different THI. As mentioned in the previous section, heat stress has
510 adverse effects on follicular development, which is directly associated with conception.

511 Furthermore, we observed that the re-ranking was stronger when using mTHI-EG for
512 AFC and IFL, which validates the hypothesis that mTHI-EG captured more variation
513 in environments than prop-EG.

514 In a breeding scheme for improved heat tolerance, the best approach could
515 involve selecting the best-performing individuals in heat stress conditions, provided
516 that they are not underperforming in thermoneutral conditions. These robust animals
517 can be presented graphically as shown in Fig. 4. For these traits, all animals have
518 consistent intercepts, whereas the gEBVs of heat-tolerant heifers (red lines) did not
519 change dramatically along all environmental conditions. The less the AFC and IFL, the
520 greater the farm profit. All the robust heifers performed better in higher THI conditions.
521 Adverse trends were observed for CR because a higher CR is preferential. The
522 magnitude of re-ranking was higher when using mTHI-EG. This is additional evidence
523 that mTHI-EG is more sensitive than prop-EG.

524

525 **Candidate genomic regions for the intercepts and slopes**

526 We performed ssGWAS to detect candidate genomic regions associated with
527 heifer fertility and heat sensitivity (Figs 5 and 6, Supp. Files 7 and 8). Some identified
528 genomic regions are common between two traits or two variables in the same trait (Supp.
529 Files 9 and 10). Generally, most of these genomic regions explained a small (< 1%)
530 proportion of the total additive genetic variance, indicating that fertility and heat
531 tolerance are largely polygenic traits. However, the functional analysis confirmed that
532 the fertility traits are influenced by heat stress. For example, some of the genomic

533 regions were previously reported to be associated with pathways such as response to
534 abiotic stimulus, detection of stimulus involved in sensory perception, response to
535 temperature stimulus, response to radiation, negative regulation of saliva secretion, and
536 aerobic respiration and energy.

537 As expected, the genomic regions associated with the intercept (average
538 performance in thermoneutral conditions) were linked with several reproductive genes.
539 For example, the overlaps of genomic windows between the intercept of AFC and IFL
540 contained genes such as *AMHR2*, *SPI*, *KRT8*, and *KRT18*. Ilha et al. [38] found that the
541 mRNA expression levels of *AMHR2* decreased in the follicles during follicular
542 deviation, whereas another study also indicated that *AMHR2* plays a role in follicular
543 development by regulating granulosa cells [39]. *KRT18* may be a molecular marker for
544 bovine microfold cells in the follicle-associated epithelium [40]. Together with *KRT8*,
545 these two keratin family genes influence the bovine estrous cycle with regards to luteal
546 cells [41]. *SPI* has been demonstrated to co-express with other regulators to control
547 early placental differentiation [42]. *PLAG1*, which was mapped in the most evident
548 region of BTA14 for AFC and IFL, has been often reported to be associated with growth
549 and reproduction traits in cattle [43–45]. The overlapping genomic regions between
550 AFC and CR contained other growth-related genes, such as *SDC3* [46] and *FABP3* [47].
551 One possible explanation is that most heifers are physiologically immature at the time
552 of first insemination.

553 Various candidate genes associated with heat tolerance (slope term) were also
554 reported in the literature. Several candidate genes are related to cow reproduction, for

555 example, *GPER1*, which has been reported to induce the non-genomic suppression
556 luteinizing hormone secretion in cattle [48], was mapped in both AFC and IFL in the
557 genomic region of BTA25. *RAD51* has been reported to be associated with bovine
558 oocytes meiosis progress [49, 50]. Some candidate genes (e.g., *LAP3*, *GLYCAM1*,
559 *PDE1B*, *MICALL2*, *NPC1*) are related with other economic traits in cattle such as milk
560 production, carcass traits, body weight, body height, and body length [51–55].
561 Additionally, several genes were annotated to be associated with stress response in
562 cattle. *HCRTR1*, which regulates orexin receptor type 1, has been suggested to
563 participate in negative feedback regulation in the adrenal gland [56]. Several studies
564 examined cultured bovine adrenal cells and indicated that the products of *AGRP* would
565 inhibit the cortisol production of adrenal gland [57, 58]. The stress reaction of animals
566 activates the hypothalamic-pituitary-adrenal axis, together with an increase in the
567 cortisol concentration [59]. Thus, *HCRTR1* and *AGRP* may play vital roles in the cattle
568 stress reaction process. White et al. [60] reported that endogenous *PC* expression in
569 bovine primary hepatocytes and kidney epithelial was significantly higher in thermal
570 stress conditions, which indicated that *PC* may contribute to the physiological response
571 to thermal stress. *PC* has been reported to be associated with fatty acids regulations
572 while feeding and thus affect the feed intake of cattle [60, 61].

573 The biological processes identified in functional analyses were mainly
574 reproduction- and stress response-related, which corroborates our findings. The QTLs
575 we found were mainly milk and production associated (Supp. Files 9 and 10), indicating
576 that these economic traits may change as the climate conditions become more extreme.

577 Most of the QTLs overlapped when using different EGs, and many were located in
578 BTA14. Costa et al. [62] reported that the QTLs in BTA14 (24.3 Mb) and BTA24 (23.4
579 Mb) are associated with AFC in Nellore. The genomic regions detected from 23 to 26
580 Mb in BTA14 and BTA24 were most evident in the Manhattan plots among the
581 different traits and EGs. Mota et al. [45] supported that the region of BTA14 plays a
582 key role in the puberty of heifers through growth hormone signaling and may be
583 regulated by *PLAG1*. Another genomic region with high genetic variance is located in
584 BTA29 (from 44.3 to 44.9 Mb), which includes the heat stress-related gene *PC*. Most
585 of the QTLs were shared by the intercept and slope, which is consistent with the strong
586 negative genetic correlations between them. The fertility-related QTLs would be
587 evident in heat stress conditions as well because the change in environments may alter
588 the genetic expression of these regions when a strong G×E exists.

589

590 **Clustered SNP effects and genes**

591 The trajectory of SNP effects over EGs (Fig. 7) suggests a substantial SNP by
592 environment interactions, which has also been reported for reproduction traits in pigs
593 [32] and cattle [45]. The dramatic changes in C1 and C2 in S2 indicates that the critical
594 periods, which were chosen through AIC, may capture more genetic variation. As for
595 CR, half of the periods overlapped in S1 (-30, 30) and S2 (-10, 30 or 60), which may
596 explain the similarity in the trajectories between the two scenarios. For C1 and C2, the
597 variation in SNP effects was greater at high levels of prop-EG (Fig. 7a) than at low or
598 middle levels, which was expected due to the higher genetic variance detected at high

599 prop-EG levels compared to low and middle prop-EG levels (Fig. 2a). Similarly, this
600 explains the greater SNP effects at both high and low levels of mTHI-EG and the late
601 “cross point”, which represents the lowest genetic variance at middle mTHI-EG levels
602 (Fig. 2b). These findings agree with those of Silva et al. [32]. The effects of top SNPs
603 were more constant (lower standard deviation) in CR compared with AFC and IFL.
604 This finding may be due to the relatively low heritability of CR, which causes smaller
605 effect variations in each level of EG.

606 Three SNP clusters were inspected to identify common genes among all the
607 fertility traits (Fig. 8). As expected, more candidate genes were shared between interval
608 traits (AFC and IFL), especially in C1 and C2. The number of overlapping genes among
609 all the traits and EGs were comparable for each cluster. For C3, the top SNPs with slope
610 effects close to 0 were chosen, but the variation was negligible, i.e. SNP effects of the
611 slope ranged from $9.48e-9$ to $1.47E-11$ for all traits. Thus, a slight change in the effect
612 estimates may cause re-ranking of SNPs, which results in a small proportion of shared
613 genes among traits in C3. The trajectories of C1 and C2 (Fig. 7) indicate the existence
614 of SNP by environment interactions, which indicate that some related genes may be
615 activated at specific temperature and humidity levels. Thus, candidate genes that play
616 an important role in SNP effect changes (C1 and C2) are the priority of the current
617 research.

618 Some genes detected in both EGs were reproduction– or milk–related. For
619 instance, *MLHI* is associated with oocyte development [50] and *EOMES* plays a vital
620 role in the early pregnancy stage of ruminants. Sakurai et al. [63] reported that cattle

621 *EOMES* expression increases when conceptuses attach to the uterine epithelium. In a
622 Chinese Holstein population, Han et al. [64] profiled the genetic effect of *ACACB*,
623 which affects milk composition traits, using whole genome re-sequenced data. As
624 production and reproduction traits are genetically related [2], it is reasonable that some
625 production associated genes were also detected in this study.

626 When prop-EG was considered, reproductive genes *NR5A2*, *THBS2* and
627 *PRKCE* were identified. *NR5A2* was mapped in C1, and it has been reported to affected
628 steroidogenic pathways of progesterone production during the luteal phase of the
629 estrous cycle in cattle [65]. *THBS2* and *PRKCE* contain several candidate SNPs in C2,
630 and their potential luteolytic functions were illustrated in previous studies [66, 67].
631 Additionally, *EGFR*, whose expression is related with bovine reproduction stage [68],
632 was detected in both C1 and C2. Wijayagunawardane et al. [69] explored the potential
633 mechanisms responsible for the detrimental effect of heat stress by exposing bovine
634 oviductal epithelial cells to heat stress conditions (40 and 43 °C). The results indicated
635 that *EGFR* could be involved in the regulation of the bovine oviductal
636 microenvironment, but these regulatory mechanisms may be compromised in the
637 presence of heat stress. This indicates that the regulatory functions of detected
638 reproductive (or even milk- and growth-related) genes might be compromised in heat
639 stress conditions. Thus, these altered SNP effects were observed at higher levels of
640 prop- and mTHI-EGs (Fig. 7).

641 When mTHI-EG was used, *GUCY1B1* was identified in C2. Several papers [70,
642 71] have demonstrated that *GUCY1B1* interacts with heat shock protein 90 (*HSP90*),

643 whereas Khan et al. [72] detected *HSPA13* as a differentially expressed gene in heat-
644 stressed bovine granulosa cells. This indicates that the interaction between *GUCY1B*
645 and *HSP90* in cattle may be related to the heat stress response.

646

647 **CONCLUSIONS**

648 In the present study, we analyzed the impacts of heat stress on dairy cattle based
649 on three reproductive traits. The critical periods, which are when heifers may be easily
650 affected by heat stress, were found to be related to the environmental gradient used,
651 centered on the first insemination day. This indicates that detailed analysis for other
652 traits should be applied to derive this period. Genetic parameters suggest significant
653 and considerable magnitude of G×E for all three heifer fertility traits, indicating that
654 breeding values may change under heat stress conditions for these traits. The re-ranking
655 of sires between different environments further demonstrates the effects of G×E on
656 animal breeding. Several reproduction-, growth-, production-, and resilience-related
657 genes and QTLs were identified in the candidate genomic regions affecting fertility
658 traits. Overall, G×E models should be integrated into current dairy cattle breeding
659 schemes to select more climatic resilient animals. The heat stress-related genes or QTLs
660 are important for exploring the mechanisms of heat stress response in dairy cattle.

661

662 **MATERIALS AND METHODS**

663 **Data**

664 Field records of birth, service and calving during 2005 – 2018 for heifers raised
665 in 15 Holstein farms (Sunlon Livestock Development Co. Ltd) in Beijing, China, were
666 collected from the herd management software AfiFarm (AfiFarm,
667 www.afimilk.com.cn). All herds were kept in a free-stall design and included 1,000 to
668 2,000 heifers and the management strategies of these farms are assumed similar. The
669 analyzed heifer traits are AFC (age at first calving), IFL (interval from the first to last
670 service), and CR (conception rate of first service). CR was coded as 1 when there was
671 a confirmed pregnancy after the first service and 0 otherwise. The number of interval
672 days is 0 when a heifer is pregnant after the first service. Further criteria for the data
673 editing included AFC between 500 and 1,100 days and IFL between 0 and 365 days.
674 Records exceeding these thresholds were dropped. Animals which changed herds
675 during the analyzed period were excluded. The number of records for each trait after
676 editing are 56,998 (Table 1). The pedigree was derived from field birth records and
677 each animal was traced back at least three generations. The final pedigree contains
678 181,693 individuals, among which 6,556 are sires.

679 Phenotypes (of daughters) and genotypes were available for 3,731 heifers and
680 537 bulls. All bulls and a subset of 2,379 heifers were genotyped using the GeneSeek
681 Genomic Profiler Bovine 50K Chip, whereas the remaining 1,352 heifers were
682 genotyped using the GeneSeek Genomic Profiler Bovine 150K Chip. In addition, 1,769
683 heifers genotyped with the 150K chip were included in the reference population to
684 improve imputation accuracy. The animals genotyped with the 50K chip were imputed
685 to the 150K using the Beagle v5.0 software [73] with an expected imputation accuracy

686 greater than 0.95. After imputation, SNP markers were filtered by removing markers
687 with minor allele frequency lower than 0.05, missing rate greater than 0.10 and
688 presenting extreme deviation from the Hardy–Weinberg equilibrium ($P \leq 10E-5$).
689 Individuals with genotype call rate lower than 0.90 were dropped and only autosomal
690 markers were retained for this study. After quality control, 111,068 SNPs remained in
691 the dataset.

692 Hourly recorded temperature data of Beijing (all farms are within 30 km from
693 the weather station) during the test years were obtained from the National Oceanic and
694 Atmospheric Administration (www.noaa.gov). Hourly THI was calculated using the
695 following formula [74]:

$$696 \quad \text{THI} = T_{\text{db}} + (0.36T_{\text{dp}}) + 41.2$$

697 where T_{db} is the hourly dry bulb temperature ($^{\circ}\text{C}$) and T_{dp} is the dew point temperature
698 ($^{\circ}\text{C}$). Then, the daily THI was calculated by averaging hourly values.

699

700 **Models**

701 Variance components were obtained using the following single-trait animal
702 model:

$$703 \quad \mathbf{y} = \mathbf{X}\mathbf{b} + \mathbf{Z}\mathbf{a} + \mathbf{e}$$

704 where \mathbf{y} is the phenotype of each heifer for the three fertility traits (AFC, IFL, and CR);
705 \mathbf{X} is an incidence matrix connecting the vector of fixed effects \mathbf{b} (herd-year-month of
706 the first service, service technician and sexual-controlled semen) to \mathbf{y} ; \mathbf{Z} is an incidence

707 matrix connecting \mathbf{a} (additive genetic effect) to \mathbf{y} , and \mathbf{e} is the residual effect. The
 708 following RNM was used to investigate G×E:

$$709 \quad \mathbf{y} = \mathbf{X}\mathbf{b} + \mathbf{Z}_0\mathbf{a}_0 + \mathbf{Z}_1\mathbf{a}_1 + \mathbf{e}$$

710 where \mathbf{Z}_0 is an incidence matrix connecting \mathbf{a}_0 (intercept) to \mathbf{y} , and \mathbf{Z}_1 is an incidence
 711 matrix containing EGs as covariables to connect \mathbf{a}_1 to \mathbf{y} (slope). It was assumed that

$$712 \quad \begin{bmatrix} \mathbf{a}_0 \\ \mathbf{a}_1 \end{bmatrix} \sim N \left(0, \mathbf{K} \otimes \begin{bmatrix} \sigma_{a_0}^2 & \sigma_{a_0 a_1} \\ \sigma_{a_0 a_1} & \sigma_{a_1}^2 \end{bmatrix} \right),$$

713 where \mathbf{K} is \mathbf{A} (numerator relationship matrix) for
 714 pedigree-based BLUP models, or to \mathbf{H} (combined pedigree-genomic relationship
 715 matrix) for ssGBLUP models.

715 The matrix \mathbf{A} was constructed using pedigree data only for conventional BLUP
 716 models, whereas for the ssGBLUP models, the inverse of the \mathbf{H} matrix (\mathbf{H}^{-1}), which
 717 was calculated as follows [20, 75]:

$$718 \quad \mathbf{H}^{-1} = \mathbf{A}^{-1} + \begin{bmatrix} 0 & 0 \\ 0 & \mathbf{G}^{-1} - \mathbf{A}_{22} \end{bmatrix}$$

719 where \mathbf{A}_{22} is the subset of \mathbf{A} for genotyped individuals and \mathbf{G} is the blended genomic
 720 relationship matrix. \mathbf{G} was built using $(1 - \omega)\mathbf{G}_0 + \omega\mathbf{A}_{22}$, where ω is the assumed
 721 weight of the genetic variance not captured by markers and was set to 0.05 according
 722 in previous studies [7, 76]. \mathbf{G}_0 was constructed using the method proposed by [77].
 723 Finally, \mathbf{G} was tuned to have the same scale as \mathbf{A}_{22} [78, 79].

724 The random residual vector \mathbf{e} is assumed to follow $\mathbf{N}(0, \mathbf{I}\sigma_e^2)$, where \mathbf{I} is an
 725 identity matrix. The residual variances were assumed to be the same across different
 726 EGs, based on the results of preliminary analyses, which suggested that residual
 727 variances were similar across different levels of EG. The full data was divided into

728 three subsets based on the percentile of EGs (percentiles of EG of ≤ 0.1 , 0.1–0.3, and
729 ≥ 0.3) to achieve similar sample sizes across subsets.

730 The (co)variance components for RNMs were estimated using the average
731 information REML method implemented in the BLUPF90 programs [80]. The standard
732 errors of (co)variance components were obtained from the average information matrix.
733 The standard errors of heritabilities among different EGs were calculated using the
734 Taylor series expansions [81].

735

736 **Critical period and environmental gradient**

737 To evaluate the impact of heat stress on heifer reproduction, the basic
738 assumption was that heifers experiencing higher THI in a specific period have reduced
739 fertility performance based on critical periods, as defined in Fig 1. The levels of prop-
740 EG range from 0 to 1 according to its definition. For mTHI-EG, the levels of EG are
741 the true minimum THI value, ranging from 15 to 75. Two scenarios were included in
742 each EG to select critical period: 1) Scenario 1 (S1) contained only one reference period
743 of 60 days, which goes from 30 days prior to the first insemination to 30 days after the
744 first insemination [17] for three reproductive traits and two EG schemes; scenario 2
745 (S2): the critical periods were selected based on the AIC of RNMs, which resulted in
746 different critical periods for each trait or EG scheme. A total of 19 combinations were
747 tested in S2, and periods with the lowest AIC were chosen (Supp. File 2). For each trait,
748 two critical periods (S1 and S2) were chosen under each EG (prop-EG and mTHI-EG)
749 to estimate the (co)variance components of the RNMs. The relationships of the two EG

750 schemes were also evaluated by calculating the equivalent values: the proportion of
751 days exceeding threshold THI when the average minimum THI of this period at a
752 certain level. Afterwards, the overlapping heifers were counted for each equivalent
753 environmental condition between two EG schemes.

754

755 **Magnitude of G×E**

756 G×E exists if the variance of the slope was significantly different from zero by
757 using a one-tailed t-test with the significance level of 0.05 [6, 33]. One possible
758 consequence of G×E is the re-ranking of animals across different environments [5];
759 thus, the top 50 sires with at least 20 daughters having phenotypes were selected to
760 assess the magnitude of re-ranking. Re-ranking plots were drawn to show the change
761 pattern of breeding values for sires with the most preferential intercepts or with the
762 lowest slopes.

763

764 **Single-step genome-wide association study (ssGWAS)**

765 The marker effects of the intercept and the slope for all traits were estimated
766 using the ssGWAS method proposed by Wang et al. [21]. The percentage of genetic
767 variance explained by a moving genomic window of 20 adjacent SNPs was obtained,
768 by applying the postGSf90 package [82]. The number of adjacent SNPs were defined
769 based on the level of linkage disequilibrium in this population, following [7]. The top
770 0.5% genomic regions that explained the highest genetic variance of intercept or slope
771 was considered as the candidate genomic regions. Subsequently, candidate genes or

772 QTLs within the candidate genomic regions were annotated based on the ARS-UCD1.2
773 genome (<http://hgdownload.soe.ucsc.edu/goldenPath/bosTau9/bigZips/genes/>) and the
774 Cattle QTL database (<https://www.animalgenome.org/cgi-bin/QTLdb/>). The biological
775 processes of candidate genes were annotated using the PANTHER Classification
776 System [83].

777

778 **Cluster analyses of relevant SNPs**

779 The SNPs were chosen to investigate the trajectories of their effects. Firstly,
780 SNPs were ranked according to the magnitude of their slope effects for each trait, EG,
781 and scenario. Then, three clusters (C) of SNPs were obtained according to the trajectory
782 of their SNP effects across each EG: C1 = SNP effects changes in preferential ways
783 (decrease for AFC and IFL; increase for CR); C2 = SNP effects changes in opposite
784 ways (increase for AFC and IFL; decrease for CR); C3 = constant SNP effects over
785 time. For C1 and C2, the top 0.5% (n=555) SNPs with the highest or the lowest slope
786 effects were selected, whereas for C3, the 0.5% SNPs with slope effects closest to zero
787 were selected. Choosing the top 1% SNPs for trajectory analyses has been implemented
788 in several GWAS studies based on the 50K SNP panel [32, 84]. The results were
789 visualized based on the average and standard deviation of SNP effects in each EG to
790 show the differences among the three clusters. For C3, SNPs were grouped into two
791 categories according to their average effects (lower or higher than 0). Only the genes
792 containing SNPs further confirmed in the cluster analysis and shared among three traits
793 were further annotated for biological functions.

794

795 **ABBREVIATIONS**

796 G × E: Genotype by environment interaction; QTL: Quantitative trait loci; THI:
797 Temperature humidity index; EG: Environmental gradient; RNM: Reaction norm
798 model; BLUP: Best linear unbiased predictor; ssGBLUP: Single-step genomic BLUP;
799 ssGWAS: Single-step genome-wide association study; AFC: Age at first calving; IFL:
800 Interval from first to last service; CR: Conception rate; prop: The proportion of days
801 that exceeded the threshold THI in candidate period; mTHI: minimum THI for each
802 day of the candidate period; AIC: Akaike Information Criteria; gEBV: Genomic
803 estimated breeding value; S: Scenario; C: Cluster.

804

805 **DECLARATIONS**

806 *Ethics approval and consent to participate*

807 Not applicable.

808

809 *Consent for publication*

810 Not applicable.

811

812 *Availability of data and materials*

813 The datasets used and/or analyzed during the current study are available from the
814 corresponding author on reasonable request.

815

816 ***Competing interests***

817 The authors declare that they have no competing interests.

818

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827

828 ***Authors' contributions***

829 RS, LFB, YW designed the study. RS, LFB, AL, HL, ZC analyzed and interpreted
830 data. RS wrote the manuscript. RS, LFB, AL, HM, BD, AVDL substantively revised
831 the manuscript. LL, GG contributed to the access to tools and materials. All authors
832 read and approved the final manuscript.

833

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840

841 **REFERENCES**

842 1. Gonzalez-Recio O, Pérez-Cabal M, Alenda R. Economic Value of Female Fertility
843 and its Relationship with Profit in Spanish Dairy Cattle. *Journal of dairy science*.
844 2004;87:3053–61.

845 2. Veerkamp RF, Beerda B. Genetics and genomics to improve fertility in high
846 producing dairy cows. *Theriogenology*. 2007;68:S266–73.

847 3. Sun C, Madsen P, Lund M, Zhang Y, Nielsen U, Su G. Improvement in genetic
848 evaluation of female fertility in dairy cattle using multiple-trait models including milk
849 production traits. *Journal of animal science*. 2009;88:871–8.

850 4. Liu A, Lund MS, Wang Y, Guo G, Dong G, Madsen P, et al. Variance components
851 and correlations of female fertility traits in Chinese Holstein population. *J Animal Sci*
852 *Biotechnol*. 2017;8:56.

853 5. Falconer DS, Mackay TFC. *Introduction to Quantitative Genetics*. New York:
854 Longman; 1996.

855 6. Liu A, Su G, Höglund J, Zhang Z, Thomasen J, Christiansen I, et al. Genotype by
856 environment interaction for female fertility traits under conventional and organic
857 production systems in Danish Holsteins. *Journal of Dairy Science*. 2019;102:8134–47.

858 7. Zhang Z, Kargo M, Liu A, Thomasen JR, Pan Y, Su G. Genotype-by-environment
859 interaction of fertility traits in Danish Holstein cattle using a single-step genomic
860 reaction norm model. *Heredity*. 2019;123:202–14.

861 8. Cheruiyot EK. Genotype-by-environment (temperature-humidity) interaction of
862 milk production traits in Australian Holstein cattle. 2020;103:17.

863 9. Bohmanova J, Misztal I. Temperature-Humidity Indices as Indicators of Milk
864 Production Losses due to Heat Stress. *Journal of dairy science*. 2007;90:1947–56.

865 10. Armstrong DV. Heat Stress Interaction with Shade and Cooling. *Journal of Dairy*
866 *Science*. 1994;77:2044–50.

867 11. Ravagnolo O, Misztal I, Hoogenboom G. Genetic Component of Heat Stress in
868 Dairy Cattle, Development of Heat Index Function. *Journal of dairy science*.
869 2000;83:2120–5.

- 870 12. Lambertz C, Sanker C, Gauly M. Climatic effects on milk production traits and
871 somatic cell score in lactating Holstein-Friesian cows in different housing systems.
872 *Journal of dairy science*. 2013;97:319-29.
- 873 13. West J, Mullinix B, Bernard J. Effects of Hot, Humid Weather on Milk
874 Temperature, Dry Matter Intake, and Milk Yield of Lactating Dairy Cows. *Journal of*
875 *dairy science*. 2003;86:232-42.
- 876 14. Cartmill JA, El-Zarkouny SZ, Hensley BA, Rozell TG, Smith JF, Stevenson JS.
877 An Alternative AI Breeding Protocol for Dairy Cows Exposed to Elevated Ambient
878 Temperatures before or after Calving or Both. *Journal of Dairy Science*.
879 2001;84:799-806.
- 880 15. Sammad A, Umer S, Shi R, Zhu H, Zhao X, Wang Y. Dairy cow reproduction
881 under the influence of heat stress. *Journal of Animal Physiology and Animal*
882 *Nutrition*. 2020;104:978-86.
- 883 16. Morton JM, Tranter WP, Mayer DG, Jonsson NN. Effects of Environmental Heat
884 on Conception Rates in Lactating Dairy Cows: Critical Periods of Exposure. *Journal*
885 *of Dairy Science*. 2007;90:2271-8.
- 886 17. Khan F, Prasad S, Gupta H. Effect of heat stress on pregnancy rates of crossbred
887 dairy cattle in Terai region of Uttarakhand, India. *Asian Pacific Journal of*
888 *Reproduction*. 2013;2:277-9.
- 889 18. Schüller LK, Burfeind O, Heuwieser W. Impact of heat stress on conception rate
890 of dairy cows in the moderate climate considering different temperature-humidity
891 index thresholds, periods relative to breeding, and heat load indices. *Theriogenology*.
892 2014;81:1050-7.
- 893 19. Misztal I, Legarra A, Aguilar I. Computing procedures for genetic evaluation
894 including phenotypic, full pedigree, and genomic information. *Journal of Dairy*
895 *Science*. 2009;92:4648-55.
- 896 20. Christensen OF, Lund MS. Genomic prediction when some animals are not
897 genotyped. *Genet Sel Evol*. 2010;42:2.
- 898 21. Wang H, Misztal I, Aguilar I, Legarra A, Muir WM. Genome-wide association
899 mapping including phenotypes from relatives without genotypes. *Genet Res*.
900 2012;94:73-83.
- 901 22. Akaike H. A New Look At The Statistical Model Identification. *Automatic*
902 *Control, IEEE Transactions on*. 1975;19:716-23.
- 903 23. Berry DP, Wall E, Pryce JE. Genetics and genomics of reproductive performance
904 in dairy and beef cattle. *Animal*. 2014;8:105-21.

- 905 24. Ghiasi H, Pakdel A, Nejati-Javaremi A, Mehrabani-Yeganeh H, Honarvar M,
906 González-Recio O, et al. Genetic variance components for female fertility in Iranian
907 Holstein cows. *Livestock Science*. 2011;139:277–80.
- 908 25. Berry DP, Kearney JF, Twomey K, Evans RD. Genetics of reproductive
909 performance in seasonal calving dairy cattle production systems. *Irish Journal of*
910 *Agricultural and Food Research*. 2013;52:1–16.
- 911 26. Liu J, Li L, Chen X, Lu Y, Wang D. Effects of heat stress on body temperature,
912 milk production, and reproduction in dairy cows: a novel idea for monitoring and
913 evaluation of heat stress — A review. *Asian-Australas J Anim Sci*. 2019;32:1332–9.
- 914 27. Wilson SJ, Kirby CJ, Koenigsfeld AT, Keisler DH, Lucy MC. Effects of
915 Controlled Heat Stress on Ovarian Function of Dairy Cattle. 2. Heifers. *Journal of*
916 *Dairy Science*. 1998;81:2132–8.
- 917 28. Sakatani M, Takahashi M, Takenouchi N. The efficiency of vaginal temperature
918 measurement for detection of estrus in Japanese Black cows. *Journal of Reproduction*
919 *and Development*. 2016;62:201–7.
- 920 29. Lozano Domínguez RR, Vásquez Peláez CG, Padilla EG. Effect of heat stress and
921 its interaction with other management and productive variables on pregnancy rate in
922 dairy cows in Aguascalientes, Mexico. *Veterinaria Mexico*. 2005;36:245-60.
- 923 30. Amundson JL, Mader TL, Rasby RJ, Hu QS. Environmental effects on pregnancy
924 rate in beef cattle1. *Journal of Animal Science*. 2006;84:3415–20.
- 925 31. Morton JM, Tranter WP, Mayer DG, Jonsson NN. Effects of Environmental Heat
926 on Conception Rates in Lactating Dairy Cows: Critical Periods of Exposure. *Journal*
927 *of Dairy Science*. 2007;90:2271–8.
- 928 32. Silva FF, Mulder HA, Knol EF, Lopes MS, Guimarães SEF, Lopes PS, et al. Sire
929 evaluation for total number born in pigs using a genomic reaction norms approach1.
930 *Journal of Animal Science*. 2014;92:3825–34.
- 931 33. Ismael A, Strandberg E, Berglund B, Kargo M, Fogh A, Løvendahl P. Genotype
932 by environment interaction for activity-based estrus traits in relation to production
933 level for Danish Holstein. *Journal of Dairy Science*. 2016;99:9834–44.
- 934 34. Su G, Madsen P, Lund MS, Sorensen D, Korsgaard IR, Jensen J. Bayesian
935 analysis of the linear reaction norm model with unknown covariates1. *Journal of*
936 *Animal Science*. 2006;84:1651–7.
- 937 35. Santana ML, Eler JP, Cardoso FF, Albuquerque LG, Ferraz JBS. Phenotypic
938 plasticity of composite beef cattle performance using reaction norms model with
939 unknown covariate. *Animal*. 2013;7:202–10.

- 940 36. Kolmodin R, Strandberg E, Madsen P, Jensen J, Jorjani H. Genotype by
941 Environment Interaction in Nordic Dairy Cattle Studied Using Reaction Norms. *Acta*
942 *Agriculturae Scandinavica*, Section A - Animal Science. 2002;52:11–24.
- 943 37. Oliveira DP, Lourenco DAL, Tsuruta S, Misztal I, Santos DJA, de Araújo Neto
944 FR, et al. Reaction norm for yearling weight in beef cattle using single-step genomic
945 evaluation1. *Journal of Animal Science*. 2018;96:27–34.
- 946 38. Ilha GF, Rovani MT, Gasperin BG, Ferreira R, de Macedo MP, Neto OA, et al.
947 Regulation of Anti-Müllerian Hormone and Its Receptor Expression around Follicle
948 Deviation in Cattle. *Reprod Domest Anim*. 2016;51:188–94.
- 949 39. Poole DH, Ocón-Grove OM, Johnson AL. Anti-Müllerian hormone (AMH)
950 receptor type II expression and AMH activity in bovine granulosa cells.
951 *Theriogenology*. 2016;86:1353–60.
- 952 40. Hondo T, Kanaya T, Takakura I, Watanabe H, Takahashi Y, Nagasawa Y, et al.
953 Cytokeratin 18 is a specific marker of bovine intestinal M cell. *Am J Physiol*
954 *Gastrointest Liver Physiol*. 2011;300:G442-453.
- 955 41. Duncan A, Forcina J, Birt A, Townson D. Estrous cycle-dependent changes of Fas
956 expression in the bovine corpus luteum: influence of keratin 8/18 intermediate
957 filaments and cytokines. *Reprod Biol Endocrinol*. 2012;10:90.
- 958 42. Degrelle SA, Murthi P, Evain-Brion D, Fournier T, Hue I. Expression and
959 localization of DLX3, PPARG and SP1 in bovine trophoblast during binucleated
960 cell differentiation. *Placenta*. 2011;32:917–20.
- 961 43. Zhong J-L, Xu J-W, Wang J, Wen Y-F, Niu H, Zheng L, et al. A novel SNP of
962 PLAG1 gene and its association with growth traits in Chinese cattle. *Gene*.
963 2019;689:166–71.
- 964 44. Hou J, Qu K, Jia P, Hanif Q, Zhang J, Chen N, et al. A SNP in PLAG1 is
965 associated with body height trait in Chinese cattle. *Anim Genet*. 2020;51:87–90.
- 966 45. Mota LFM, Lopes FB, Fernandes Júnior GA, Rosa GJM, Magalhães AFB,
967 Carvalheiro R, et al. Genome-wide scan highlights the role of candidate genes on
968 phenotypic plasticity for age at first calving in Nellore heifers. *Sci Rep*.
969 2020;10:6481.
- 970 46. Huang Y-Z, Wang Q, Zhang C-L, Fang X-T, Song E-L, Chen H. Genetic Variants
971 in SDC3 Gene are Significantly Associated with Growth Traits in Two Chinese Beef
972 Cattle Breeds. *Anim Biotechnol*. 2016;27:190–8.
- 973 47. Liang W, Zhang HL, Liu Y, Lu BC, Liu X, Li Q, et al. Investigation of the
974 association of two candidate genes (H-FABP and PSMC1) with growth and carcass
975 traits in Qinchuan beef cattle from China. *Genet Mol Res*. 2014;13:1876–84.

- 976 48. Rudolf FO, Kadokawa H. Cytoplasmic kinases downstream of GPR30 suppress
977 gonadotropin-releasing hormone (GnRH)-induced luteinizing hormone secretion from
978 bovine anterior pituitary cells. *J Reprod Dev.* 2016;62:65–9.
- 979 49. Kujjo LL, Ronningen R, Ross P, Pereira RJG, Rodriguez R, Beyhan Z, et al.
980 RAD51 plays a crucial role in halting cell death program induced by ionizing
981 radiation in bovine oocytes. *Biol Reprod.* 2012;86:76.
- 982 50. Bilotto S, Boni R, Russo GL, Lioi MB. Meiosis progression and donor age affect
983 expression profile of DNA repair genes in bovine oocytes. *Zygote.* 2015;23:11–8.
- 984 51. Ju Z, Zheng X, Huang J, Qi C, Zhang Y, Li J, et al. Functional characterization of
985 genetic polymorphisms identified in the promoter region of the bovine PEPS gene.
986 *DNA Cell Biol.* 2012;31:1038–45.
- 987 52. Pedersen LRL, Nielsen SB, Hansted JG, Petersen TE, Otzen DE, Sørensen ES.
988 PP3 forms stable tetrameric structures through hydrophobic interactions via the C-
989 terminal amphipathic helix and undergoes reversible thermal dissociation and
990 denaturation. *FEBS J.* 2012;279:336–47.
- 991 53. Shin S, Heo J, Yeo J, Lee C, Chung E. Genetic association of phosphodiesterase
992 1B (PDE1B) with carcass traits in Korean cattle. *Mol Biol Rep.* 2012;39:4869–74.
- 993 54. Xu Y, Zhang L, Shi T, Zhou Y, Cai H, Lan X, et al. Copy number variations of
994 MICAL-L2 shaping gene expression contribute to different phenotypes of cattle.
995 *Mamm Genome.* 2013;24:508–16.
- 996 55. Dang Y, Li M, Yang M, Cao X, Lan X, Lei C, et al. Identification of bovine
997 NPC1 gene cSNPs and their effects on body size traits of Qinchuan cattle. *Gene.*
998 2014;540:153–60.
- 999 56. Nemoto T, Toyoshima-Aoyama F, Ueda Y, Ohba T, Yanagita T, Watanabe H, et al.
1000 al. Involvement of the Orexin System in Adrenal Sympathetic Regulation.
1001 *Pharmacology.* 2013;91:250–8.
- 1002 57. Doghman M, Delagrangé P, Blondet A, Berthelon M-C, Durand P, Naville D, et al.
1003 al. Agouti-related protein antagonizes glucocorticoid production induced through
1004 melanocortin 4 receptor activation in bovine adrenal cells: a possible autocrine
1005 control. *Endocrinology.* 2004;145:541–7.
- 1006 58. Doghman M, Delagrangé P, Berthelon M-C, Durand P, Naville D, Bégeot M.
1007 Sustained inhibitory effect of Agouti Related Protein on the ACTH-induced cortisol
1008 production by bovine cultured adrenal cells. *Regul Pept.* 2005;124:215–9.
- 1009 59. Tiwari RV, Parajuli P, Sylvester PW. [gamma]-Tocotrienol-induced endoplasmic
1010 reticulum stress and autophagy act concurrently to promote breast cancer cell death.
1011 *Biochemistry and cell biology = Biochimie et biologie cellulaire.* 2015;93:306.

- 1012 60. White HM, Koser SL, Donkin SS. Regulation of bovine pyruvate carboxylase
1013 mRNA and promoter expression by thermal stress. *J Anim Sci.* 2012;90:2979–87.
- 1014 61. Velez JC, Donkin SS. Feed restriction induces pyruvate carboxylase but not
1015 phosphoenolpyruvate carboxykinase in dairy cows. *J Dairy Sci.* 2005;88:2938–48.
- 1016 62. Costa RB, Camargo GM, Diaz ID, Irano N, Dias MM, Carneiro R, et al.
1017 Genome-wide association study of reproductive traits in Nelore heifers using
1018 Bayesian inference. *Genetics Selection Evolution.* 2015;47:67.
- 1019 63. Sakurai T, Bai H, Bai R, Sato D, Arai M, Okuda K, et al. Down-regulation of
1020 interferon tau gene transcription with a transcription factor, EOMES. *Mol Reprod*
1021 *Dev.* 2013;80:371–83.
- 1022 64. Han B, Liang W, Liu L, Li Y, Sun D. Genetic association of the ACACB gene
1023 with milk yield and composition traits in dairy cattle. *Anim Genet.* 2018;49:169–77.
- 1024 65. Taniguchi H, Komiyama J, Viger RS, Okuda K. The expression of the nuclear
1025 receptors NR5A1 and NR5A2 and transcription factor GATA6 correlates with
1026 steroidogenic gene expression in the bovine corpus luteum. *Mol Reprod Dev.*
1027 2009;76:873–80.
- 1028 66. Goravanahally MP, Sen A, Inskeep EK, Flores JA. PKC epsilon and an increase
1029 in intracellular calcium concentration are necessary for PGF2 alpha to inhibit LH-
1030 stimulated progesterone secretion in cultured bovine steroidogenic luteal cells. *Reprod*
1031 *Biol Endocrinol.* 2007;5:37.
- 1032 67. Berisha B, Schams D, Rodler D, Sinowatz F. Expression pattern of HIF1alpha and
1033 vasohibins during follicle maturation and corpus luteum function in the bovine ovary.
1034 *Reproduction in Domestic Animals.* 2016; doi: 10.1111/rda.12867.
- 1035 68. Sağsöz H, Ketani MA, Saruhan BG. Expression of the erbB/HER receptor family
1036 in the bovine uterus during the sexual cycle and the relation of this family to serum
1037 sex steroids. *Biotech Histochem.* 2012;87:105–16.
- 1038 69. Wijayagunawardane MPB, Hambruch N, Haeger J-D, Pfarrer C. Effect of
1039 epidermal growth factor (EGF) on the phosphorylation of mitogen-activated protein
1040 kinase (MAPK) in the bovine oviduct in vitro: Alteration by heat stress. *J Reprod*
1041 *Dev.* 2015;61:383–9.
- 1042 70. Nedvetsky PI, Meurer S, Opitz N, Nedvetskaya TY, Müller H, Schmidt HHHW.
1043 Heat shock protein 90 regulates stabilization rather than activation of soluble
1044 guanylate cyclase. *FEBS Lett.* 2008;582:327–31.
- 1045 71. Sarkar A, Dai Y, Haque MM, Seeger F, Ghosh A, Garcin ED, et al. Heat Shock
1046 Protein 90 Associates with the Per-Arnt-Sim Domain of Heme-free Soluble Guanylate
1047 Cyclase: Implications for Enzyme Maturation. *J Biol Chem.* 2015;290:21615–28.

- 1048 72. Khan A, Dou J, Wang Y, Jiang X, Zahoor M, Hanpeng L, et al. Evaluation of heat
1049 stress effects on cellular and transcriptional adaptation of bovine granulosa cells.
1050 *Journal of Animal Science and Biotechnology*. 2020; doi: 10.1186/s40104-019-0408-
1051 8.
- 1052 73. Browning BL, Zhou Y, Browning SR. A One-Penny Imputed Genome from Next-
1053 Generation Reference Panels. *The American Journal of Human Genetics*.
1054 2018;103:338–48.
- 1055 74. Yousef MK. *Stress physiology in livestock*. CRC Press; 1985.
- 1056 75. Aguilar I, Misztal I, Johnson DL, Legarra A, Tsuruta S, Lawlor TJ. Hot topic: a
1057 unified approach to utilize phenotypic, full pedigree, and genomic information for
1058 genetic evaluation of Holstein final score. *Journal of Dairy Science*. 2010;93:743–52.
- 1059 76. Gao H, Christensen OF, Madsen P, Nielsen US, Zhang Y, Lund MS, et al.
1060 Comparison on genomic predictions using three GBLUP methods and two single-step
1061 blending methods in the Nordic Holstein population. *Genet Sel Evol*. 2012;44:8.
- 1062 77. VanRaden PM. Efficient Methods to Compute Genomic Predictions. *Journal of*
1063 *Dairy Science*. 2008;91:4414–23.
- 1064 78. Powell JE, Visscher PM, Goddard ME. Reconciling the analysis of IBD and IBS
1065 in complex trait studies. *Nature Reviews Genetics*. 2010;11:800–5.
- 1066 79. Vitezica ZG, Aguilar I, Misztal I, Legarra A. Bias in genomic predictions for
1067 populations under selection. *Genet Res*. 2011;93:357–66.
- 1068 80. Misztal I. Complex Models, More Data: Simpler Programming? *IEEE Internet*
1069 *Computing - INTERNET*. 1999;20.
- 1070 81. Su G, Lund MS, Sorensen D. Selection for litter size at day five to improve litter
1071 size at weaning and piglet survival rate1. *Journal of Animal Science*. 2007;85:1385–
1072 92.
- 1073 82. Aguilar I, Misztal I, Tsuruta S, Legarra A, Wang H. PREGSF90 – POSTGSF90:
1074 Computational Tools for the Implementation of Single-step Genomic Selection and
1075 Genome-wide Association with Ungenotyped Individuals in BLUPF90 Programs.
1076 2014; doi: 10.13140/2.1.4801.5045.
- 1077 83. Thomas P, Campbell M, Kejariwal A, Mi H, Karlak B, Daverman R, et al.
1078 PANTHER: A Library of Protein Families and Subfamilies Indexed by Function.
1079 *Genome research*. 2003;13:2129–41.
- 1080 84. Oliveira HR, Lourenco DAL, Masuda Y, Misztal I, Tsuruta S, Jamrozik J, et al.
1081 Single-step genome-wide association for longitudinal traits of Canadian Ayrshire,
1082 Holstein, and Jersey dairy cattle. *Journal of Dairy Science*. 2019;102:9995–10011.

Figures

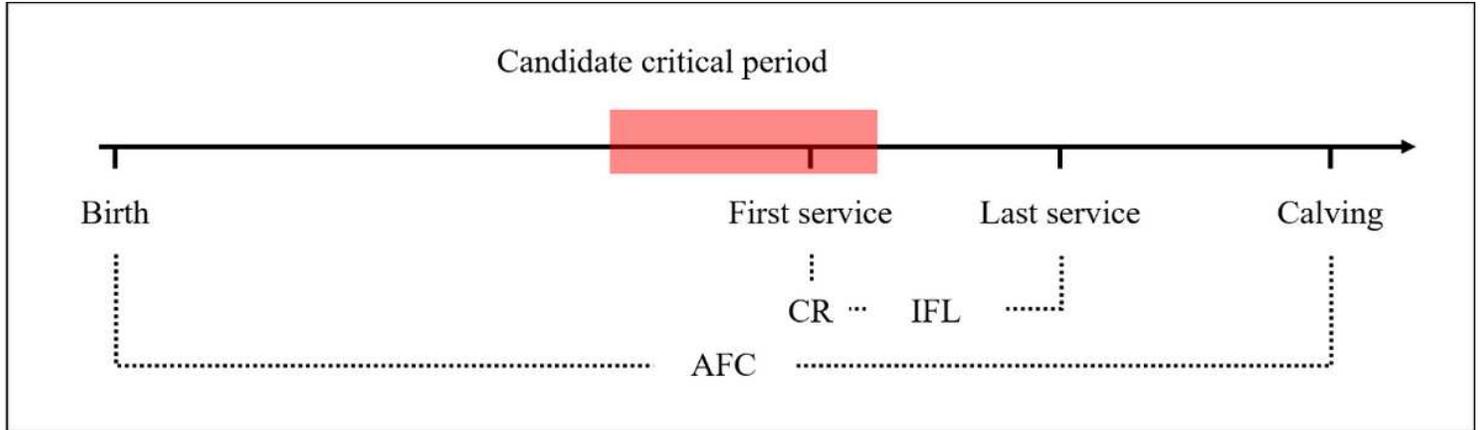


Figure 1

Reproductive events and the definition of critical period in heifers. The red rectangle represents the critical period, defining as the time period of which heifers are likely to suffer from heat stress. AFC = age at first calving, IFL = interval from first to last service, CR = conception rate of first service.

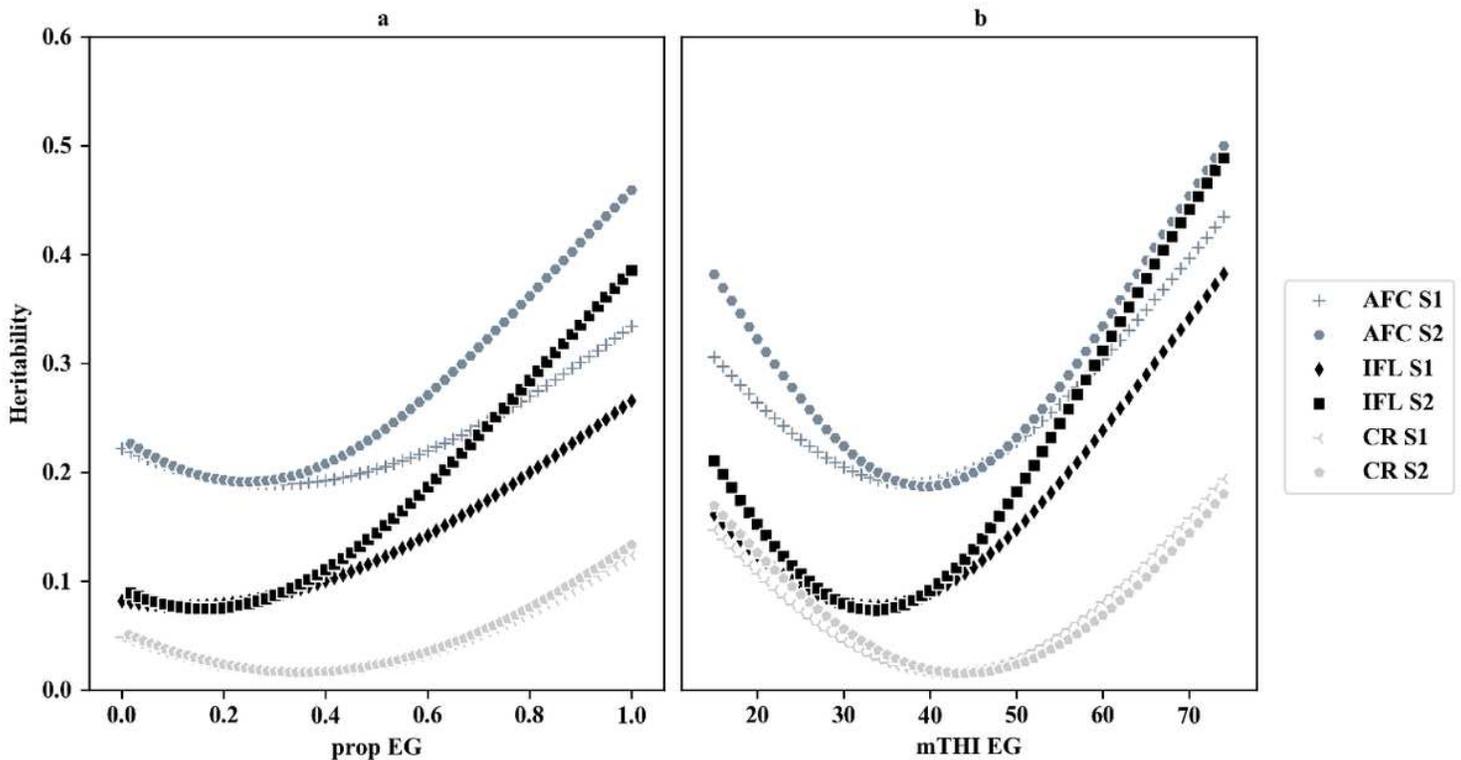


Figure 2

Heritabilities estimated by reaction norm models with the matrix H using a) prop-EG or b) mTHI-EG as environmental gradient. For a), the x-axis is the proportion of days exceeding the threshold with a range of 0 to 1; while for b), the x-axis is the minimum THI with a range of 15 to 75.

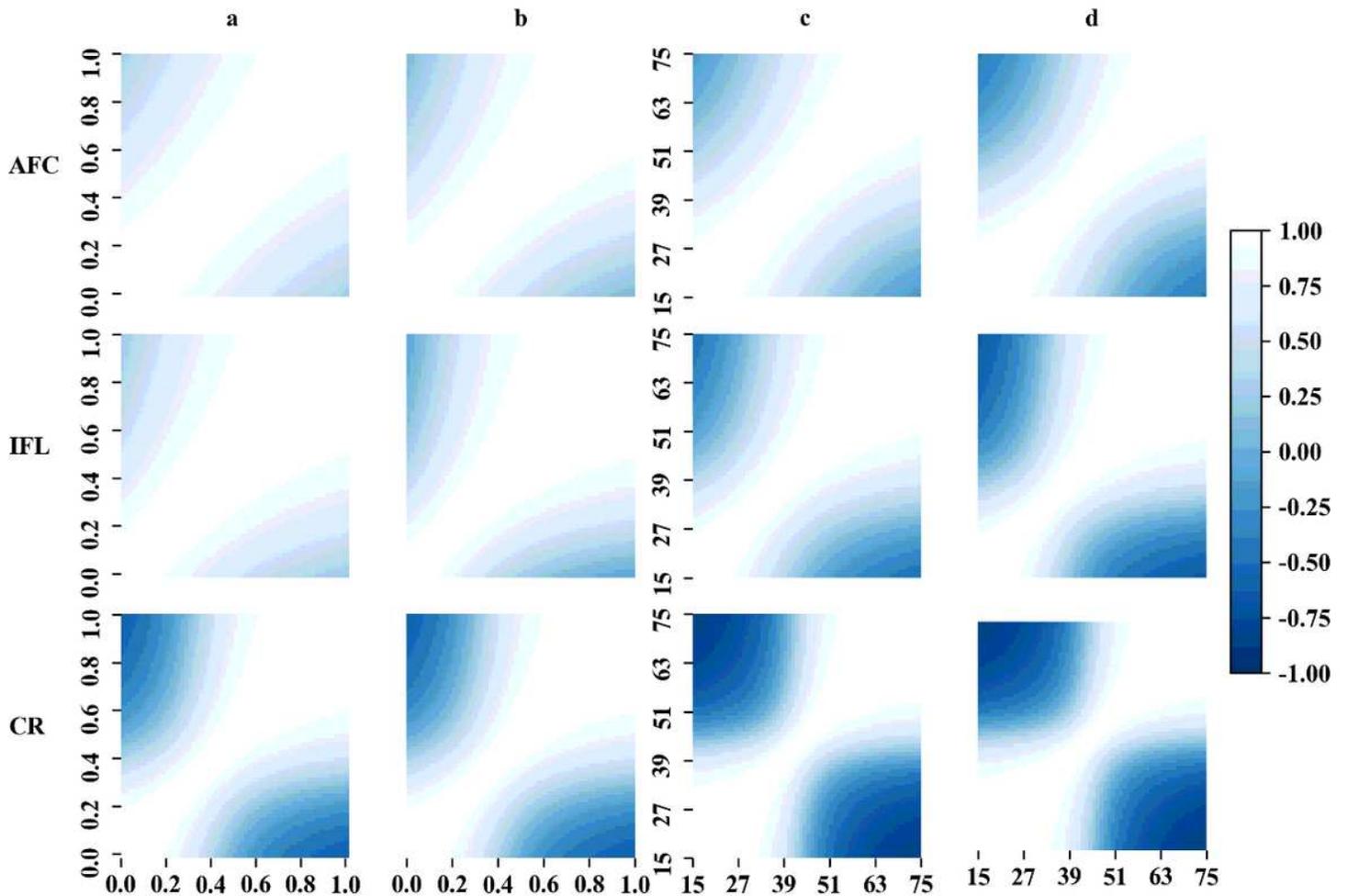


Figure 3

Genetic correlations estimated by reaction norm models (RNMs) with the matrix H. The color indicates the magnitude of the genetic correlation. a) Correlations between different levels of prop-EG estimated from RNM under S1. The x-axis and y-axis are the proportion of days exceeding the threshold, ranging from 0 to 1. b) Correlations between different levels of prop-EG estimated from RNM under S2. The x-axis and y-axis are the proportion of days exceeding the threshold, ranging from 0 to 1. c) Correlations between different levels of mTHI-EG estimated from RNM under S1. The x-axis and y-axis are the minimum THI, ranging from 15 to 75. d) Correlations between different levels of mTHI-EG estimated from RNM under S2. The x-axis and y-axis are the minimum THI, ranging from 15 to 75.

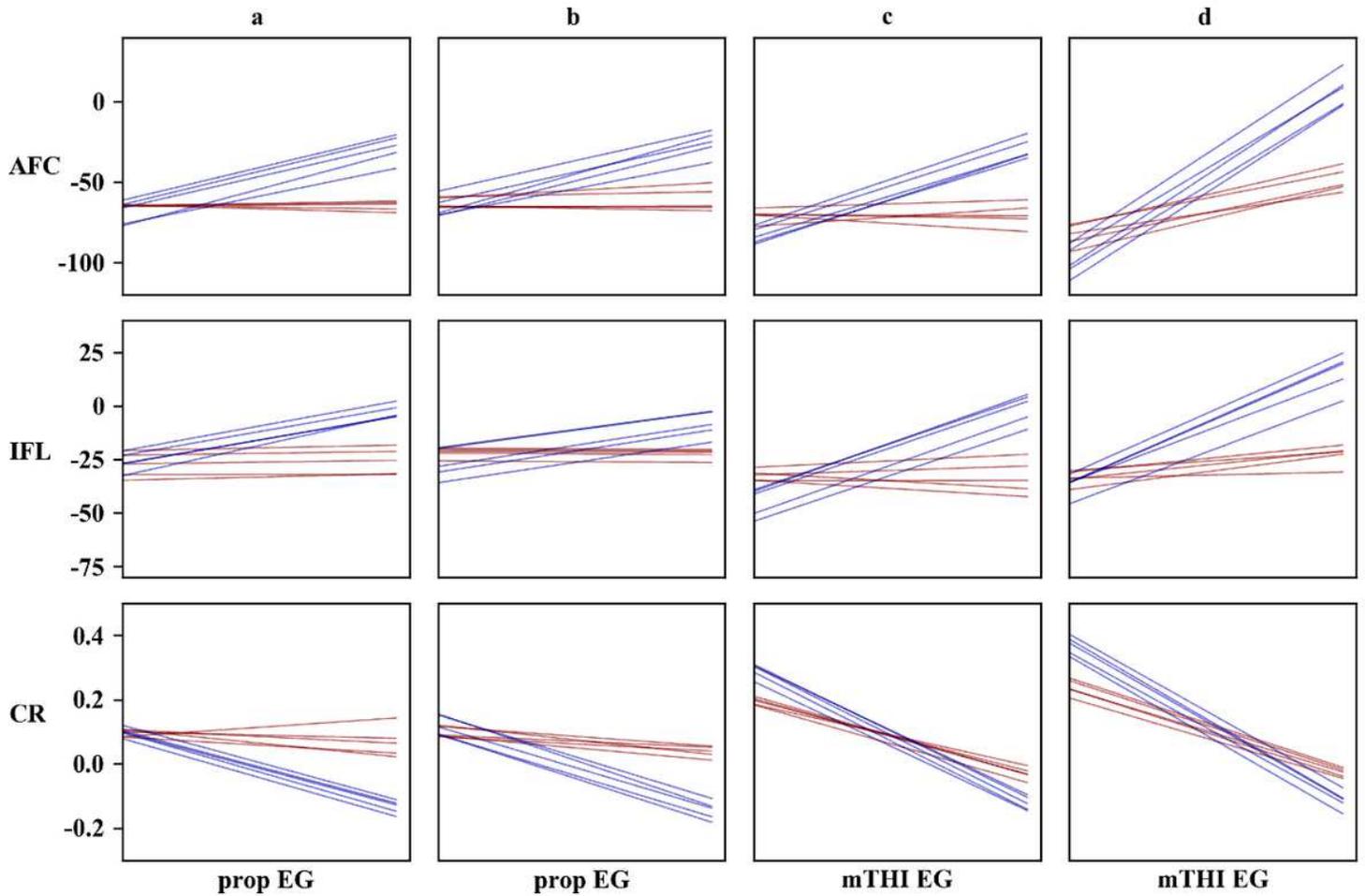


Figure 4

The re-ranking plots for gEBVs of sires. The blue and red lines represent sensitive and resilient sires, respectively. a) Re-ranking plots for three traits estimated using prop-EG under S1. The x-axis is the proportion of days exceeding the threshold with a range of 0 to 1 and y-axis is gEBV. b) Re-ranking plots for three traits estimated using prop-EG under S2. The x-axis is the proportion of days exceeding the threshold with a range of 0 to 1 and y-axis is gEBV. c) Re-ranking plots for three traits estimated using mTHI-EG under S1. The x-axis is the minimum THI with a range of 15 to 75 and y-axis is gEBV. d) Re-ranking plots for three traits estimated using mTHI-EG under S2. The x-axis is the minimum THI with a range of 15 to 75 and y-axis is gEBV

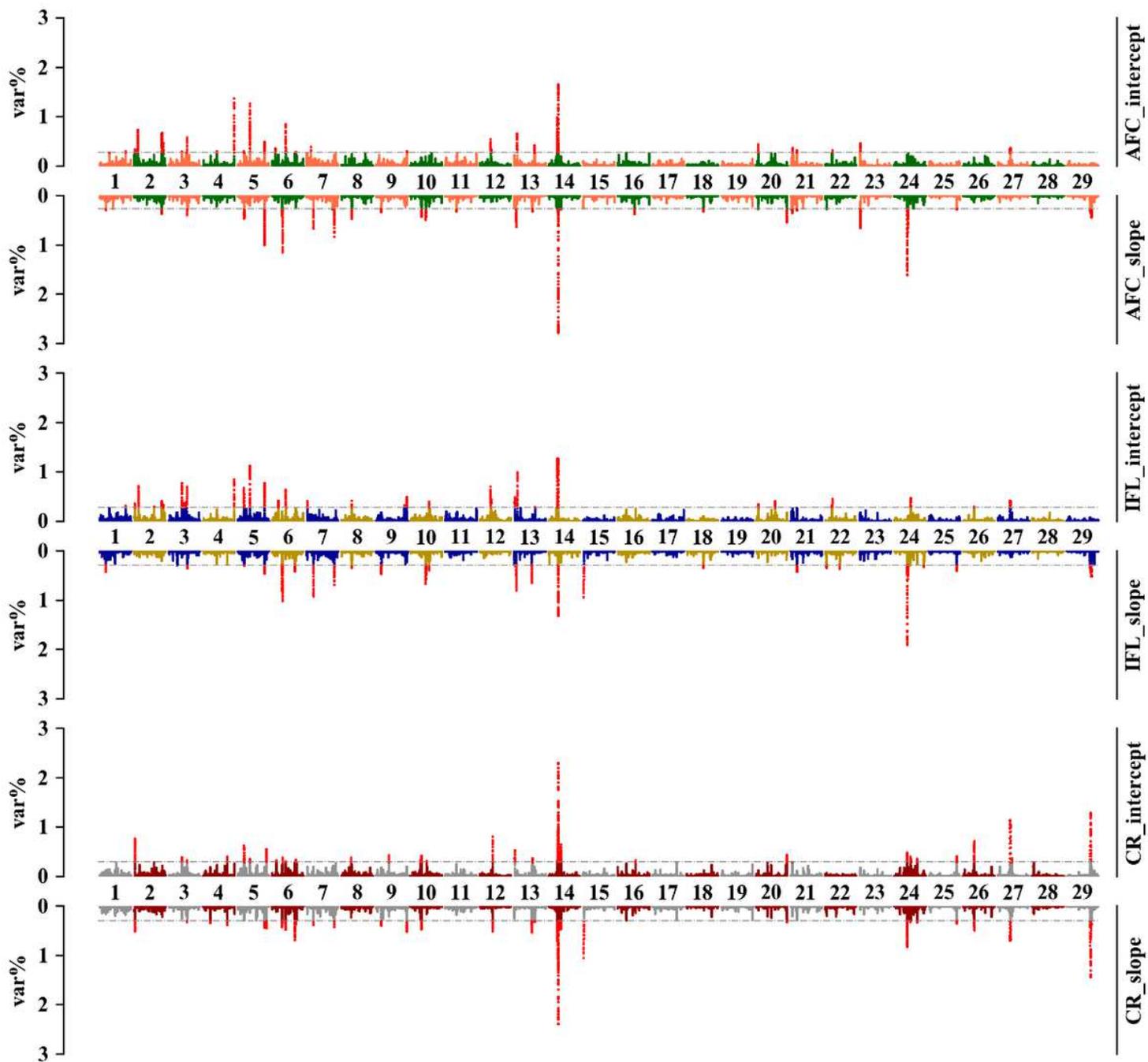


Figure 5

Percentages of the intercept and slope genetic variances explained by a sliding window of 20 SNPs for three traits, which were estimated under scenario one of prop-EG. The x-axis is autosome segments; the y-axis represents the proportion of explained variances; the grey horizontal lines are thresholds (top 0.5%) for candidate genomic regions; different color sets indicate different traits

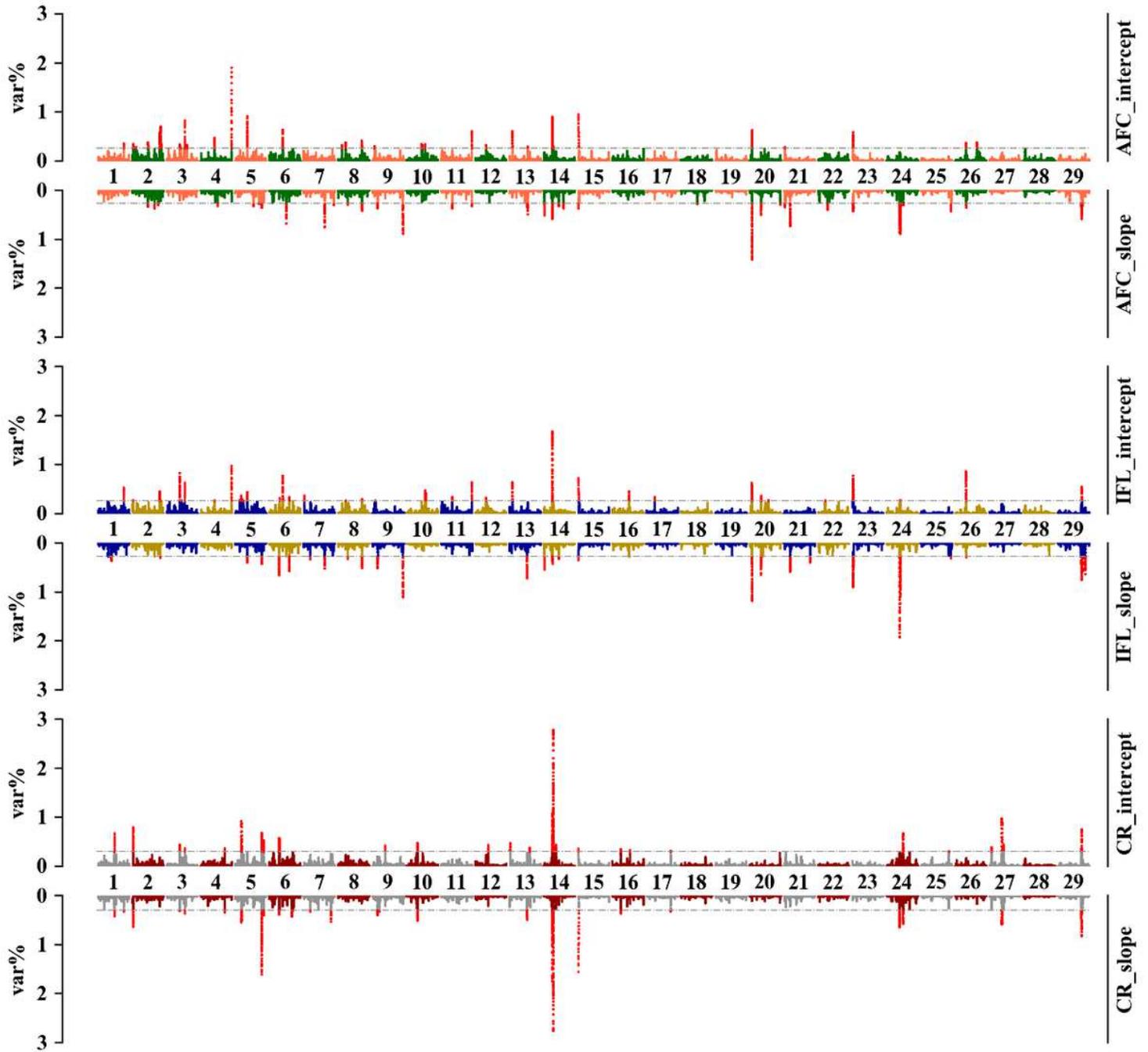


Figure 6

Percentages of the intercept and slope genetic variances explained by a sliding window of 20 SNPs for three traits, which were estimated under scenario two of prop-EG. The x-axis is autosome segments; the y-axis represents the proportion of explained variances; the grey horizontal lines are thresholds (top 0.5%) for candidate genomic regions; different color sets indicate different traits.

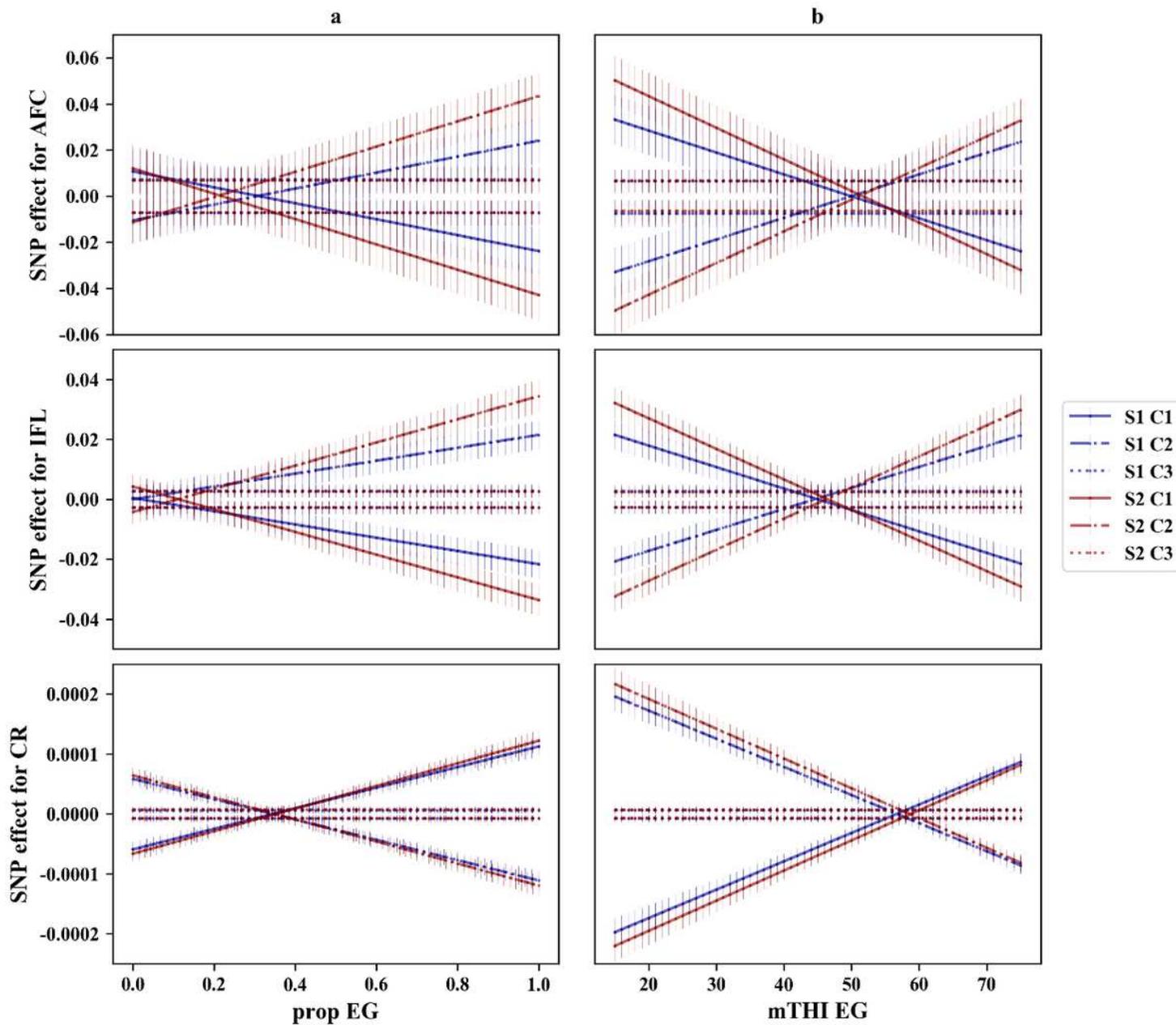


Figure 7

Trajectories of SNP effects changing over EGs. The x-axis is environmental gradient; the y-axis represents the SNP effects; vertical bar is the standard deviations of SNP effects at each level of EG; blue lines indicate scenario one; red lines indicate scenario two; different color sets indicate different clusters. a) Trajectories of SNP effects changing over prop-EG. The x-axis is the proportion of days exceeding the threshold with a range of 0 to 1. b) Trajectories of SNP effects changing over mTHI-EG. The x-axis is the minimum THI with a range of 15 to 75.

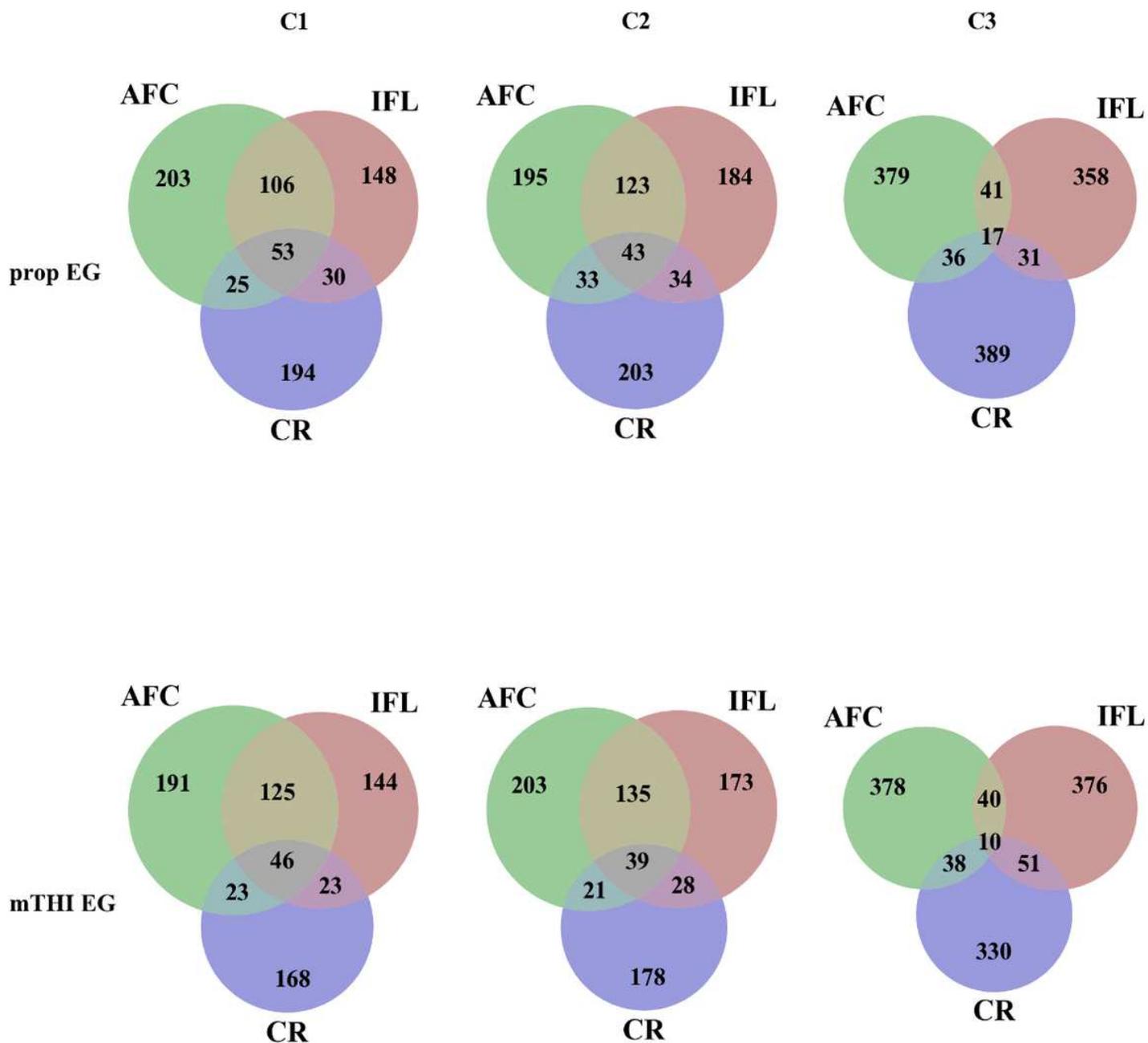


Figure 8

Number of shared candidate genes for each EG in different traits and clusters. C1 = SNP effects changes in preferential ways (decrease for AFC and IFL; increase for CR); C2 = SNP effects changes in the opposite ways (increase for AFC and IFL; decrease for CR); C3 = constant SNP effects over time.

Supplementary Files

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