

TNF- α inhibitor therapy can improve the immune imbalance of CD4+ T cells and negative regulatory cells but not CD8+ T cells in ankylosing spondylitis

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Abstract

Background: Studies into ankylosing spondylitis (AS) and its relationship with immune function are controversial, and the correlation between the efficacy of TNF- α inhibitor and changes in immune function is unclear.

Methods: A total of 40 immune cells were tested with flow cytometry, and the results of 105 HC (healthy control) subjects, 177 active-stage AS patients, and 23 AS cases before and after 12 weeks of TNF- α inhibitor therapy (Anbainuo) were analyzed.

Results: Compared with the HC group, the proportion of immune cells, such as naïve and central memory CD4+T cells, in AS increased ($p < 0.0001$), but effector memory and terminally differentiated CD4+T cells were decreased ($p < 0.01$ and 0.0001 , respectively). Naïve, central memory, and effector memory CD8+T cells were increased ($p < 0.0001$, 0.001 , and 0.01 , respectively), but terminally differentiated CD8+T cells were decreased ($p < 0.0001$). Th1 cells (helper T cells-1), Tfh1 cells (follicular helper T cells-1), Tc1 cells (cytotoxic T cells-1), and Tregs (regulatory T cells) were lower ($p < 0.01$, 0.05 , 0.0001 , and 0.001 , respectively), but Th17 cells, Tfh17 cells, and Tc cells were higher ($p < 0.001$, 0.0001 and 0.001 , respectively). The proportions of total B cells and class-switched B cells were increased ($p < 0.05$), but non-switched B cells, plasma cells, memory B cells, and immature Bregs (regulatory B cells) were lower ($p < 0.01$, 0.0001 , 0.0001 , and 0.0001 , respectively). After Anbainuo therapy, the percentage of naïve CD4+ T cells had decreased ($p < 0.05$) but Tregs and B10 cells (IL-10-producing regulatory B cells) had increased ($p < 0.01$ and 0.05 , respectively), and the increase in Tregs was positively correlated with the decrease in CRP (C-reactive protein) ($r = 0.489$, $p = 0.018$).

Conclusions: We found that active-stage AS patients have an immunity imbalance involving multiple types of immune cells, including CD4+T cells, CD8+T cells, Th cells, Tfh cells, Tc cells, Tregs, Bregs, and B cells. TNF- α inhibitor Anbainuo can not only help to inhibit disease activity but can also improve the immune imbalance of CD4+ T cells and negative regulatory cells. But CD8 + T cells have not been rescued.

Introduction

As the prototype of spondyloarthritis, ankylosing spondylitis (AS) is a chronic inflammatory disease that affects the spine and sacroiliac joints. The pathogenesis of AS can be attributed to both hereditary and environmental factors [1]. Recent studies have suggested that the onset of AS involves innate immunity and acquired immunity. In the past, many studies have been conducted on regulatory T cells (Tregs) are essential in maintaining immune homeostasis in AS, but an association has not yet been determined. Some studies have suggested that the frequency of Treg cells in AS patients is significantly decreased in comparison to healthy control subjects [2], but there has been no more in-depth verification or functional testing. Some reports have even suggested that the level of Treg cells does not change in AS patients [3]. Studies on acquired immunity to AS have suggested that T-cell-mediated immune regulation may also

play an important role [4, 5]. CD4+ T and CD8+ T cells and their subsets are known to act on the pathogenesis of AS, although many questions remain about the subject [4, 6]. However, there are few detailed reports on the stage of differentiation or subsets of CD4+ T or CD8+ T cells in which AS patients show abnormalities. Role of B cells in autoimmune diseases was once thought to be predominantly limited to the production of autoantibodies. However, B cells are now considered to have other potential functions with immunology and inflammatory diseases. Nevertheless, only limited data are available on the phenotypic and functional status of B cells in AS [7, 8]. The ability of B cells to negatively regulate cellular immune responses and inflammation has been explored, and the concept of regulatory B cells (Breg cells) has emerged. Indeed, defective suppressive functions of Breg cell subsets have been observed in several chronic inflammatory diseases [9, 10], but studies into Breg cell activity in AS patients have been scattered. Some small sample research studies showed regulatory B cells to have a defective function in AS patients but observed no significant change in cell frequency [11].

Given that immune cells in AS have imbalance, manipulating immune cells may be a new therapeutic strategy for treating AS. Therefore, exploring factors that influence immune cells number and function will play a critical role in understanding the pathogenesis of these diseases and identifying new treatment strategies. Tumor necrosis factor- α (TNF- α) inhibitor has been shown to have an anti-inflammatory effect in treating AS [12], but its potential beneficial effects on the bone structure are still challenging to identify and its mechanisms remain unclear. The symptoms would recur in the vast majority of patients if the drug is stopped, it is important to know the type and degree of the alterations in the immune system developing during the anti-TNF therapy. Therefore, we propose that AS in active-stage has a variety of immune cells imbalances and TNF- α inhibitor can improve some immune imbalance attributed to AS but cannot rescue all of the immunophenotype. We conducted a prospective study of T-cell and B-cell subset distribution by identifying the phenotype and also analyzed the changes in the proportion of lymphocyte subsets in order to indirectly understand the differentiation status, degree of failure, and cell activity of various cell subsets. That will help us to further clarify changes of the immune system caused by AS and to explore resistance that could contribute to relapse after treatment.

Materials And Methods

Patients and controls

The Ethics Committee approved this study of the Third Affiliated Hospital of Sun Yat-Sen University, Guangzhou, China. Besides, informed consent was obtained from all patients and all HCs.

All of the AS patients in our study had high disease activity (Ankylosing Spondylitis Disease Activity Score, ASDAS ≥ 1.3), were diagnosed with AS according to the Modified New York Criteria (mNY) [13], and must not have received any biological treatment in the three months leading up to the study. Inclusion criteria for control subjects were: over the age of 18 years; no current or chronic medication intake; and no known disease or condition. Both AS and HC subjects must not have received a vaccination or suffered an infection in the 3 months leading up to the study.

Peripheral blood lymphocytes were tested with flow cytometry. Parameters of disease activity, including ASDAS and C-reactive protein (CRP), were also registered. No statistically significant differences were observed in terms of age or gender between AS patients and HCs in the two comparison phases. In the primary screening phase of the study, 67 AS patients and 50 HCs were included. In the expanded validation phase, 110 AS patients and 55 HCs were included.

TNF- α inhibitor (Anbainuo)-treated AS patients

All patients fulfilling the modified New York criteria [13] who were eligible for TNF- α inhibitor treatment due to persistently high disease activity (ASDAS \geq 1.3) despite treatment with NSAIDs, or who were unable to take NSAIDs due to contraindications, were included in the study. For all AS patients, the clinical and laboratory assessment was the first time they had been treated with TNF- α inhibitors. Exclusion criteria were any significant comorbidity and the use of any drug other than NSAID or a proton pump inhibitor in the 3 months before the study. Active-stage AS patients received a subcutaneous injection of etanercept biosimilar: Anbainuo (Recombinant Human Tumor Necrosis Factor- α Receptor α : IgG Fc Fusion Protein for Injection, Zhejiang Hisun Pharmaceutica, China) [14] 50 mg weekly for 12 weeks. Peripheral blood lymphocytes were tested with flow cytometry on the baseline and after Anbainuo treatment. Parameters of disease activity, including ASDAS, CRP, and Bath Ankylosing Spondylitis Disease Activity Index (BASDAI)[15] before and after treatment, were also recorded.

Blood sampling

Blood samples were taken in the laboratory unit of the Department of Rheumatology and Immunology. After peripheral venous blood sampling, 5ml samples preserved with heparin sodium anticoagulant tubes were used to detect immune cells, and 3ml samples preserved with blood collection tubes without anticoagulant were used to conduct a CRP test. The samples were stored at room temperature (20°C -30°C) for no more than 6 hours before handling.

Lymphocyte immunophenotyping by Flow Cytometry

Immunophenotyping of peripheral blood lymphocytes was quantified by flow cytometry (BD) according to the manufacturer's instructions. Whole blood was added to a panel of the following fluorescently labeled antibodies for incubation (for 15 min at room temperature in the dark): CD3-PerCP-Cy5.5, CD4-APC-H7, CD8-BV510, CD127-BV421, CD25 L-PE, CD45RAL-FITC, CCR7-AF647, CD28-PE-Cy7, CD3-APC-H7, CD4-PE-Cy7, CD8-PerCP-Cy5.5, CD183(CXCR3)-Alexa488, CD196 (CCR6)-BV510, CD185 (CXCR5)-Alexa 647, CD194-BV421, CD279(PD-1)-PE for T cell subsets; and CD45-APC-H7, CD19-PerCP-Cy5.5, CD27-BV421, IgD-BB515, IgM-BV510, CD38L-APC, CD24L-PE, and CD21-PE-Cy7 for B cell subsets.

Supplementary Table 1 details more information about the categorizations of T/B cell surface markers. After red blood cells were lysed (4° C in the dark) and centrifuged, the cells were suspended with phosphate-buffered saline and then analyzed on a BD flow cytometry.

Supplementary Table 1. The 28 T lymphocyte and 12 B lymphocyte subsets [16]

Lymphocyte subsets	Phenotype
T cell	CD3+
Th cell	CD3+CD4+
Tc cell	CD3+CD8+
DP T cell	CD3+CD4+CD8+
Naïve CD4+T cell	CD3+CD4+CD45RA+CCR7+
Terminally Differentiated CD4+T cell	CD3+CD4+CD45RA+CCR7-
Central Memory CD4+T cell	CD3+CD4+CD45RA-CCR7+
Effector Memory CD4+T cell	CD3+CD4+CD45RA-CCR7-
Exhausted CD4+T cell	CD3+CD4+CD28-
Functional CD4+T cell	CD3+CD4+CD28+
Treg cell	CD3+CD4+CD25+CD127-
Naïve CD8+T cell	CD3+CD8+CCR7+CD45RA+
Terminally Differentiated CD8+T cell	CD3+CD8+CCR7-CD45RA+
Central Memory CD8+T cell	CD3+CD8+CCR7+CD45RA-
Effector Memory CD8+T cell	CD3+CD8+CCR7-CD45RA-
Exhausted CD8+T cell	CD3+CD8+CD28-
Tfh cell	CD3+CD4+CXCR5+
Th1 cell	CD3+CD4+CXCR5-CXCR3+CCR4-
Th2 cell	CD3+CD4+CXCR5-CXCR3-CCR4+
Th17 cell	CD3+CD4+CXCR5-CXCR3-CCR4-CCR6+
Tfh1 cell	CD3+CD4+CXCR5+CXCR3+CCR4-
Tfh2 cell	CD3+CD4+CXCR5+CXCR3-CCR4+
Tfh17 cell	CD3+CD4+CXCR5+CXCR3-CCR4-CCR6+
Tc1 cell	CD3+CD8+CXCR5-CXCR3+CCR4-
Tc2 cell	CD3+CD8+CXCR5-CXCR3-CCR4+
Tc17 cell	CD3+CD8+CXCR5-CXCR3-CCR4-CCR6+
Peripheral Th cell	CD3+CD4+CXCR5-PD-1+
Activated Tfh cell	CD3+CD4+CXCR5+PD-1+
B cell	CD3-CD19+
Naïve B cell	CD3-CD19+CD27-IgD+
MZ B cell	CD3-CD19+CD27+IgD+
CD21 ^{low} B cell	CD3-CD19+IgD+CD27- CD38 ^{low} CD21 ^{low}
Pre-Naïve B cell	CD3-CD19+IgM-IgD-CD27-CD38+
Plasma cell	CD3-CD19+IgD-IgM-CD27+CD38+
Class-switched B cell	CD3-CD19+IgD-IgM-CD27+CD38-
B10 cell	CD3-CD19+IgD+IgM+CD27+CD38-CD24+
Memory B cell	CD3-CD19+IgD+IgM+CD27+CD38+CD24+
Non-switched B cell	CD3-CD19+IgD+IgM+CD27+CD38+CD24-
Immature Breg cell	CD3-CD19+IgD+IgM+CD27-CD38+CD24+
Transitional B cell	CD3-CD19+IgM+IgD-CD27-CD38+CD24+

Th cell: Helper T cell, Tc cell: Cytotoxic T lymphocyte, DP T cell: Double Positive T cell, Treg cell: Regulatory T cell, Tfh cell: Follicular helper T cell, Breg cell: Regulatory B cell. CD183: CXCR3, CD185: CXCR5, CD194: CCR4, CD196: CCR6, CD279: PD1.

CRP analysis

CRP was tested by immunoturbidimetry in venous blood serum obtained at the same time.

Statistical analysis

Statistical analysis was carried out using SPSS 23.0 software. The Shapiro-Wilk test was used to test for normality. Reference ranges were calculated using mean \pm standard deviation for normally distributed data and median (Interquartile Range (IQR)) for non-normally distributed data. For data that conformed to a normal distribution, a parameter test was used, and for data that did not conform to a normal distribution, a non-parametric test was used. The difference between the two groups was compared using 2-sided tests for parametric data and the Wilcoxon rank-sum test for nonparametric data. The gender ratio was compared between the control subjects and patients using a Chi-square test. Correlation between clinical parameters and the ratio of lymphocyte subtypes was tested using the parametric Pearson correlation coefficient test or the nonparametric Spearman's rank correlation coefficient test. Statistical significance was considered to exist when $P < 0.05$. A heat map was created using the R package "pheatmap"

Results

Case summary

Supplementary Table 2 compares the characteristics of the AS patients and HCs in two phases, the primary screening phase, and the expanded validation phase.

Supplementary Table 2. Characteristics of AS patients and HCs.

Parameters	Primary Screening Phase			Expanded Validation Phase		
	AS (n=67)	HC (n=50)	P	AS (n=110)	HC (n=55)	P
Age (years)	33 (26-41)	31.5 (28-38)	0.976	31.5 (25-37)	30 (27-37)	0.402
Sex ratio (% male)	80.60	60	0.184	73.64	63.64	0.185
D D (years)	16.75 (11.5-23.75)	-	-	8 (3-14)	-	-
CRP (mg/L)	10.25 (4.18-26.51)	-	-	8.50(2.30-18.10)	-	-
ASDAS-CRP	2.89 \pm 1.13	-	-	2.53 \pm 0.97	-	-

Results are displayed as mean \pm standard deviation for normally distributed data and median (IQR) for non-normally distributed data. AS ankylosing spondylitis. HC, healthy control. DD disease duration. CRP: C-reactive protein. ASDAS: Ankylosing Spondylitis Disease Activity Score.

Primary screening phase

The 28 T lymphocytes and 12 B lymphocytes subsets content was analyzed in the AS and HC groups at both the primary screening phase and expanded validation phase. There was a significant change in the percentage of T lymphocytes and B lymphocytes in the AS patients compared to the HCs. The percentage of Th1 cells (CD3+CD4+CXCR3+CCR4-CXCR5-), Tfh1 cells (CD3+CD4+CXCR3+CCR4-CXCR5+), Tc1 cells (CD3+CD8+CXCR3+CCR4-CXCR5-), Memory B cells (CD3-CD19+CD27+CD24+CD38+IgD+IgM+), and non-switched B cells (CD3-CD19+ CD24-CD27+CD38+IgD+IgM+) were found to be significantly higher in the AS patients. However, the percentage of Th2 cells (CD3+CD4+CXCR3-CCR4+CXCR5-CCR6-), Th17 cells (CD3+CD4+CXCR3-CCR4-CXCR5-CCR6+), Tfh2 cells (CD3+ CD4+CXCR3-CCR4+CXCR5+), and B cells (CD3-CD19+) were found to be significantly lower in the AS patients. The statistical results are listed in Supplementary Table 3 ($p < 0.05$). Representative cytofluorometric gating strategy and detailed examples of analyses for the subsets have been represented in supplementary figure 1.

Supplementary Table 3. Differences in lymphocyte frequencies between AS patients and HCs at the primary screening phase.

Subtype	Frequency (%)		P
	AS (n=67)	HC (n=50)	
Th1 cell	14.06 \pm 6.57	18.08 \pm 8.5	0.01
Th2 cell	14.4 (10.9-18.3)	12.8 (9.35-15.8)	0.02
Th17 cell	1.55 (0.8-2.8)	1.06 (0.65-1.85)	0.01
Tfh1 cell	11.5 (9.57-13.6)	13.1 (11.28-15.7)	0.02
Tfh2 cell	37.75 \pm 6.96	34.47 \pm 7.48	0.02
Tc1 cell	40.62 \pm 18.4	54.362 \pm 12.4	**
B cell	12.86 \pm 5.53	9.92 \pm 4.36	**
Memory B cell	0.25 (0.13-0.79)	0.6 (0.35-1.42)	**
Non-switched B cell	0.05 (0-0.13)	0.1 (0.05-0.22)	**

Results are displayed as mean \pm standard deviation for normally distributed data and median (IQR) for non-normally distributed data. Th cell: Helper T cell. Tfh cell: Follicular helper T cell. Tc cell: Cytotoxic T lymphocyte. (**, $P < 0.01$)

Expanded validation phase

Cluster analyses of immunophenotypic parameters that were differentially expressed in the AS patients and HCs at the expanded validation phase are summarized in Figure 1 (Fig 1). The magnitude of parameter expression is color-coded with red for a relative increase in expression and blue for a relative decrease in expression. Strong cluster separation can be observed between AS and HC. Part of immune cells with a relative increase in expression (upper left quadrant and lower right quadrant) clearly separated from others with a relative decrease in expression (upper right quadrant and lower left quadrant). Supplementary Table 4 shows the statistical results of average frequencies and p-values of a total of 21 immune cells with significant differences between the two groups.

Supplementary Table 4. Differences in lymphocyte frequencies between the AS patients and HCs at the expanded validation phase.

Subtype	Frequency (%)		P-value
	AS (n=110)	HC (n=55)	
Naïve CD4+T cell	29.77±12.33	20.95±10.87	**
Terminally Differentiated CD4+T cell	13.5(6.58-23.6)	23.8(15-31.3)	**
Central Memory CD4+T cell	10.7(7.62-16)	7.96(5.07-10.4)	**
Effector Memory CD4+T cell	42.07±11.24	46.90±10.28	**
Naïve CD8+T cell	35.75(26.28-48.2)	20.5(10.9-32.6)	**
Terminally Differentiated CD8+T cell	27.00±13.90	48.15±14.78	**
Central Memory CD8+T cell	1.21(0.63-2.26)	0.66(0.38-0.94)	**
Effector Memory CD8+T cell	34.58±12.29	28.81±11.15	**
Th1 cell	12.3(8.42-16.65)	16.6(12.7-22.6)	**
Th17 cell	1.82(1.05-2.97)	1.08(0.66-1.89)	**
Tfh1 cell	12.40±4.28	13.85±4.25	0.04
Tfh17 cell	8.1(5.97-10.23)	5.13(3.66-7.33)	**
Tc cell	34.12±8.01	29.55±8.23	**
Tc1 cell	38.56±16.65	54.63±12.72	**
Treg cell	3.54±1.21	5.45±4.58	**
B cell	11.8(8.91-14.93)	9.43(6.47-12.9)	**
Class-switched B cell	7.9(5.63-12.3)	6.1(3.49-8.97)	**
Non-switched B cell	0.06(0.03-0.13)	0.09(0.04-0.22)	0.03
Plasma cell	3.1(1.82-5.50)	6.45(1.74-10.3)	**
Memory B cell	0.31(0.13-0.62)	0.56(0.33-1.39)	**
Immature Breg cell	0.11(0.04-0.27)	0.35(0.12-0.68)	**

Results are displayed as mean ± standard deviation for normal distribution data and median (IQR) for non-normally distributed data. Th cell: Helper T cell. Tfh cell: Follicular helper T cell. Tc cell: Cytotoxic T lymphocyte. Treg cell: Regulatory T cell. (**, P<0.01)

T lymphocyte

The percentage of CD4+ T cells at different stages of differentiation were calculated, and significant differences between the AS patients and HCs are shown in Fig. 2 CCR7+ CD4+T cells including naïve CD4+T cells (CD3+CD4+CD45RA+CCR7+, Fig. 2A), and central memory CD4+T cells (CD3+CD4+CD45RA-CCR7+, Fig. 2C) were significantly increased in the AS group, but CCR7- CD4+T cells including terminally differentiated CD4+T cells (CD3+CD4+CD45RA+CCR7-, Fig. 2B), and effector memory CD4+T cells (CD3+CD4+CD45RA-CCR7-, Fig. 2D) were significantly decreased.

As shown in Fig. 2, the percentage of CD8+ T cells at different stages of differentiation was also calculated. Naïve CD8+T cells (CD3+CD8+CD45RA+CCR7+, Fig. 2E), central memory CD8+T cells (CD3+CD8+CD45RA-CCR7+, Fig. 2G), and effector memory CD8+T cells (CD3+CD8+CD45RA-CCR7-, Fig. 2H) were significantly increased in the AS group, but terminally differentiated CD8+T cells (CD3+CD8+CD45RA+CCR7-, Fig. 2F) were significantly decreased.

Simultaneously, we tested Th cells (Th1 cells, Th2 cells, Th17 cells), Tc cells (Tc1 cells, Tc2 cells, Tc17 cells), and Tfh cells (Tfh1 cells, Tfh2 cells, Tfh17 cells), and the results with significant differences are shown in Fig. 3. The percentage of CXCR3+T cells, including Th1 cells (CD3+CD4+ CXCR3+ CCR4-CXCR5-, Fig. 3A), Tfh1 cells (CD3+CD4+CXCR3+CCR4-CXCR5+, Fig. 3B), and Tc1 cells (CD3+CD8+CXCR3+ CCR4-CXCR5-, Fig. 3C) was found to be significantly lower in the AS group. However, the percentage of CCR6+ helper T cells, such as Th17 cells (CD3+CD4+CXCR3-CCR4-CXCR5-CCR6+, Fig. 3D) and Tfh17 cells (CD3+CD4+CXCR3-CCR4-CXCR5+CCR6+, Fig 3E) were found to be significantly higher. The total Tc cells (CD3+CD8+) (Fig 3. F) were also significantly increased in the AS group.

Regulatory lymphocytes

We compared changes in the ratio in negative regulatory cells, such as Tregs and Bregs, between the AS and HC groups. The percentage of Tregs (CD3+CD4+CD25+CD127-) and immature Bregs (CD3-CD19+CD24+CD27-CD38+IgD+IgM+) was found to be significantly lower in the AS group than in the HC group ($p < 0.0001$). Supplementary Fig. 2 lists the scatter plot results. B10 cells (CD3-CD19+CD24+CD27+CD38-IgD+IgM+) were also shown to be decreased in the AS group, but the difference was not significant.

B lymphocytes

Fig. 4 shows the percentage of B cells at different stages of differentiation. The proportion of total B cells (CD3-CD19+, Fig. 4A) and class-switched B cells (CD3-CD19+CD27+CD38-IgD-IgM-, Fig. 4B) were all significantly increased in the AS group. However, antibody-secreting phenotype B cells (CD19+ CD38+) including non-switched B cells (CD3-CD19+CD27+CD38+CD24-IgD+ IgM+, Fig. 4C), plasma cells (CD3-CD19+CD27+CD38+IgD-IgM-, Fig. 4D), memory B cells (CD3-CD19+CD24 +CD27+CD38+IgD+IgM+, Fig. 4E), and immature Bregs (CD3-CD19+IgD+IgM+ CD27-CD38+CD24+, Fig. 4F) were found to be significantly decreased in the AS group compared to the HC group.

The impact of Anbainuo therapy on lymphocyte subsets in AS

A total of 23 active-phase AS patients were included in this research. The mean age of the patients (M/F:19/4) was 30 years (range, 25-36 years), and mean disease duration was 8 years (range, 4.5-13.00 years) at baseline. Disease activity was indicated by CRP of 12.10 mg/L (range, 2.60-20.90 mg/L), ASDAS of 2.97±1.02, and BASDAI of 4.25±1.37 before treatment. After 12 weeks of Anbainuo therapy, CRP, ASDAS, and BASDAI all decreased significantly after treatment ($p < 0.05$), and the average values were 2.50 mg/L (range, 0.50-8.00 mg/L), 1.37±1.04, and 1.69±1.32, respectively, as shown in Supplementary Table 5.

Supplementary Table 5. Demographic and disease characteristics of AS patients treated with Anbainuo.

	Baseline	After Anbainuo Therapy	P
Demographics			
Sex (M: F)	19:4	-	-
Age (years)	30 (25-36)	-	-
D D (years)	8.00 (4.50-13.00)	-	-
Disease status			
CRP (mg/L)	12.10 (2.60-20.90)	2.50 (0.50-8.00)	0.015
ASDAS-CRP	2.97±1.02	1.37±1.04	**
BASDAI	4.25±1.37	1.69±1.32	**

*Results are displayed as mean ± standard deviation for normal distribution data and median (IQR) for non-normally distributed data. Anbainuo: Recombinant Human Tumor Necrosis Factor-α Receptor 1: IgG Fc Fusion Protein for Injection, made in China. M: male, F: female. DD: disease duration. CRP: C-reactive protein. BASDAI: Bath Ankylosing Disease Activity Index. ASDAS: Ankylosing Spondylitis Disease Activity Score. (**, $P < 0.01$)*

After 12 weeks of Anbainuo therapy, the amount of some lymphocyte subsets in the peripheral blood of the AS patients changed significantly (Fig. 5). Here we compared lymphocytes in 55 HCs and 23 Ambino treated AS patients. CD4+T cells and CD8+T cells were measured at different stages of differentiation, and comparisons were made between the AS and HC groups. As shown in Fig 5, naïve CD4+ T cells (CD3+CD4+CD45RA+CCR7+, Fig. 5A) were decreased and effector memory CD8+ T cells (CD3+CD8+CD45RA-CCR7-, Fig 5D) increased after Anbainuo therapy.

The number of regulatory lymphocytes detected in the blood of the AS patients changed significantly after Anbainuo treatment, with the percentage of Treg cells (CD3+CD4+CD25+CD127-, Fig. 5B) and B10 cells (CD3-CD19+CD24+CD27+ CD38-IgD+IgM+, Fig 5C) increasing significantly but immature Bregs (CD3-CD19+CD24+ CD27CD38+IgD+IgM+, Fig. 5G) decreasing significantly.

Simultaneously, we measured the number of Th cells (Th1 cells, Th2 cells, Th17 cells), Tc cells (Tc1 cells, Tc2 cells, and Tc17 cells), and Tfh cells (Tfh1 cells, Tfh2 cells, and Tfh17 cells) before and after

Anbainuo therapy. As shown in Fig. 5, the proportion of Tc1 cells (CD3+CD8+CXCR3+CCR4-CXCR5-, Fig. 5E) decreased, and the proportion of Tfh17 cells (CD3+CD4+CXCR3-CCR4-CXCR5+ CCR6+, Fig 5F) increased after treatment.

However, apart from immature Bregs and B10 cells, the proportion of various B cell subtypes did not change significantly after treatment with Anbainuo.

Correlation between immune cells and clinical parameters

We analyzed the correlation between the frequency of immune cells and some clinical parameters and found that CD4+T cells and CD8+T cells at different stages of differentiation were associated with disease duration. Disease duration was negatively correlated with the frequency of naïve T cells including Naïve CD4+T cells (CD3+ CD4+CD45RA+CCR7+, Fig. 6A) and naïve CD8+T cells (CD3+CD8+CD45RA+ CCR7+, Fig. 6D). However, disease duration was positively correlated with the proportion of T cells including effector memory CD4+T cells (CD3+CD4+CD45RA-CCR7-, Fig. 6B), exhausted CD4+T cells (CD3+CD4+ CD28- Fig. 6C), terminally differentiated CD8+T cells (CD3+CD8+CD45RA+ CCR7-, Fig. 6.E), and exhausted CD8+T cells (CD3+CD8+CD28-, Fig 6F), at the late stage of differentiation in patients with AS.

When analyzing the correlation between C-reactive protein (CRP) levels and lymphocyte frequency, only the frequency of Tc1 cells (CD3+CD8+CXCR3+CCR4-CXCR5-) was found to be negatively correlated with CRP level ($r = -0.182$, $p = 0.041$).

To understand the correlation between changes in disease status (including CRP, BASDAI, and ASDAS) and changes in lymphocyte frequency after Anbainuo therapy, Spearman's rank correlation analyses showed that the decrease in CRP was positively correlated with the increase in the frequency of Tregs (CD3+CD4+CD25+CD127-) following Anbainuo therapy for 12 weeks ($r = 0.489$, $p = 0.018$).

Discussion

The inflammatory effects seen throughout AS are associated with abnormal immune function. Previous studies have found changes in the immune cells of patients with AS [6, 17, 18]. Our study, which included an extensively validated large sample size, showed that active AS causes abnormalities in the proportion of lymphocyte subsets and that help us to indirectly understand the differentiation status, degree of failure, and cell activity of various cell subsets. Our results, which showed changes in the frequency of Th cells, Tfh cells, Tc cells, and Treg cells in patients with AS were consistent with the findings of past studies [4, 8, 17]. We also made a discovery regarding abnormal changes in CD4+T cells and CD8+T cells at different stages of differentiation of AS, which further suggests that AS patients have an irregular proportion of T cells at different stages of differentiation. Some previous studies based on small sample sizes found only abnormal changes in CD4+T cell subsets at different stages of differentiation but not in CD8+T cell subsets [19].

Furthermore, we also found that disease duration was negatively correlated with the frequency of Naïve T cells but, in the late stage of differentiation, positively correlated with the frequency of T cells. We found the number of naïve T cells to decrease gradually and the number of T cells in the effect stage and exhausting stages to gradually increase, which indicates that the number of T cells rose as the disease persisted. Long-term chronic inflammation can affect the immune function of AS patients through changes in T cell subsets. Among these significantly altered cells, we noticed that CCR7⁺T cells were increased, but CCR7⁻ T cells were decreased. The relevance of the function of cells labeled with CCR7 cells to the pathogenesis of AS deserves further investigation, and the marker CCR7 may be used as a potential indicator of immune status.

In the past, there have been a small number of reports on the abnormal distribution of B cells in the peripheral blood of AS patients [18]. However, the distribution of different subtypes of B cells or different stages of differentiation has not yet been fully revealed. Our study found that AS patients experienced a significant increase in the total of B cells and class-switched B cells but a significant reduction in the proportion of antibody-secreting subtype B cells during the B cell effector phase. At the same time, no significant difference was observed in terms of autoimmune-related changes in the AS patients compared with the HCs. This shows that AS is better defined as an autoinflammatory disease than an autoimmune disease. In the future, potential immune regulation against B cells is likely to benefit patients with AS.

Our results showed that Anbainuo injections could address partial immune function imbalance in AS patients by affecting differentiation and activation of immune cell subtypes, including naïve CD4⁺ T cells, Tregs, and B10 cells. Among these, the decreased proportion of naïve CD4⁺ T cells after Anbainuo treatment was consistent with the findings of past research [20]. Our observation that naïve CD4⁺ T cells decreased but effector memory CD8⁺ T cells increased after Anbainuo treatment may have several possible explanations. For example, migration of T cells precursors from the thymus or release of naïve T cells from the secondary lymphoid organs may have been impaired, but differentiation of memory T cells from naïve cells may have accelerated. This could be one of the mechanisms by which Anbainuo regulates immunity and plays an anti-inflammatory role.

We also find that Anbainuo treatment for AS can increase the proportion of negative regulatory cells such as Tregs and Bregs in inflammation that are significantly lower than normal before treatment. In humans, plasmablasts, immature B cells, B10 cells, B regulatory 1 (Br1) cells, and Granzyme B (GrB)⁺ B cells comprise the identified Breg subsets [21]. In human peripheral blood, Bregs cells contain a high proportion of IL-10-producing Bregs cells, named B10 cells [22], which, via the production of IL-10, suppress TNF- α production by monocytes. We have described the decreased frequency of Tregs, immature Bregs, and B10 cells, in AS patients who had previously received no biological treatment agents. (We found that B10 cells did not display a significant change between AS and HCs). This may suggest that the lack of negative regulatory cells in inflammation may contribute to the pathogenesis of AS. The parallel compensatory increase seen in Tregs and B10 cells after Anbainuo treatment for 12 weeks suggests that TNF- α inhibitor may assist with the recovery of patients' immune tolerance. An

increase in Tregs during long-term anti-TNF therapy for AS has also been noted [20]. Our research further reveals a positive correlation between the reduction in CRP value and a higher frequency of Tregs. However, another small sample size study about Bregs led to the opposite conclusion [7]. Human regulatory plasmablasts are suggested to drive from immature B cells and act through IL-10 production [23, 24]. This may explain our observation that immature Bregs decreased but B10 cells increased after Anbainuo treatment. IL-10 secreted by B10 cells could be used as potential indicators for the therapeutic efficacy of TNF- α inhibitors.

However, Anbainuo is not able to regulate all of the immune imbalances attributed to AS. Our results show that the proportion of effector memory CD8⁺ T cells and Tfh17 cells, which was significantly higher in AS patients (110 AS patients vs. 55 HCs), and the proportion of Tc1 cells, which was significantly lower in AS patients (110 AS patients vs. 55 HCs), have not been rescued after treatment. Initially, we found the frequency of Tc1 cells to be significantly decreased and to be negatively associated with CRP levels. However, after Anbainuo treatment, although the disease activity (ASDAS and BASDAI) of AS patients improved, and the average CRP level decreased, the frequency of Tc1 cells was still lower than the baseline. This may indicate that T cells cannot act as an effective antitumor or antimicrobial defense, even after a substantial period of Anbainuo therapy. More importantly, it also suggests that some immunological abnormality remains despite the effects of Anbainuo. Further study is required to establish better whether the abnormalities seen in the frequency of the CD8⁺ T cells and their functions are related to the pathogenesis of AS, and their value as potential targets for AS therapy is also worthy of attention in future research.

AS we known, AS pathogenesis is affected by the relationship between the host genetics, the intestinal microbiome and the immune response [25].

(1) AS has long been associated with inheritance of the HLA allele B27 [1], and the pathogenic role of HLAB27 remains unclear despite intensive research. The arthritogenic peptide theory proposes that HLAB27 plays a central pathogenic role in the presentation of joint-specific peptides to CD8⁺ cytotoxic T cells. Specific self or environmental peptides are proposed to bind to and be presented by HLA-B27, to activate CD8⁺ cells.

(2) Another major theory for the pathogenesis of HLA-B27 in AS revolves around the ability of HLA-B27 to aberrantly fold to form homodimers [26]. Circulating CD4⁺ T cells expressing the killer cell immunoglobulin receptor (KIR3DL2) after activation, recognize HLA-B27 homodimers, and this recognition is associated with the secretion of large amounts of inflammatory cytokines including high levels of IL-17A, and these cells are polarized towards a Th17 phenotype [27].

(3) Recent genetic and immunological research has highlighted a key role for IL-17A/IL-23 cytokine dysregulation of the Th17 immune pathway in AS [1]. IL-23 signalling through the IL-23 receptor (IL-23R) on CD4⁺ Th cells is required for the differentiation and expansion of Th17 cells. A perpetual activation of both the innate and the adaptive immune systems, in particular of the IL-23/IL-17 axis and of Th1

effector T-cell lineage with the overproduction of TNF- α , are considered to be key steps in the pathogenesis of AS.

(4) TNF- α induces the production of IL-17 and has an impact on the differentiation and activation of various T-cell subtypes, and therefore it has a significant role in shaping the adaptive immune system [28].

(5) One of the most specific pathologic features of AS is inflammation at the enthesis [29]. Mechanical stress and/or infectious stress resulting in overexpression of the IL-17A/IL23 axis with activation of resident enthesal cells that can respond to IL-23 including CD4+T cells, CD8+ T cells, $\gamma\delta$ T cells and other innate immune cells. This leads to production of IL-17A, IL-22, TNF- α and other cytokines that mediate spinal and peripheral inflammation directly or through tissue-resident effector cells [30].

(6) Environmental factors, which may include the microbiome and infections, probably also play a significant role in AS pathogenesis. Bowel inflammation and AS are frequently clinically associated [31]. IL-10 secreted by B10 cells plays a role in the development of intestinal immunity [32].

Our research is basically consistent with the above immunological concepts in the pathogenesis of AS. We found that the proportion of naïve CD4+/CD8+T cells and memory CD4+/CD8+T cells had increased while terminally differentiated CD4+/CD8+T cells had decreased, which just confirmed the role of activation of CD4+/CD8+ T cells in the classic theory of AS pathogenesis. We also found that the proportion of Th17 cells increased while the proportion of Th1 cells decreased, confirming the key role of IL-17A/IL-23 signaling pathway in the pathogenesis of AS. TNF- α inhibitor Anbainuo can not only help to inhibit disease activity and partial immune function imbalance in AS patients by affecting the differentiation and activation of immune cell subtypes of CD4+T cells, but it can also increase the number of negative regulatory cells in inflammation such as Tregs and Bregs. Our finding that B10 cells increased after Anbainuo treatment may provide further support to the hypothesis that AS is caused by interactions between the host gut immune system and the gut microbiome.

This study has a few weak points. It was found that the average onset age of the first stage (primary screening phase) is lower than general AS population [33]. That may be related to the limited sample size of the first stage without artificial screening for demographic characteristics. That may cause some bias especially the result of immune imbalance associated with the course of the disease. However, the onset age of the second stage (expanded validation phase) (22.7 ± 7.85) is consistent with the general AS population. Here we truthfully report the true statistics and plan to further expand the verification study in the future.

Conclusion

Our prospective study, which had a large sample size, found active-stage AS patients to have an immunity imbalance involving multiple immune cells. The pathogenesis of AS sees not only an increase in effector immune cells but also a decrease in negative regulatory immune cells. TNF- α inhibitor

Anbainuo can not only help to inhibit disease activity and partial immune function imbalance in AS patients by affecting the differentiation and activation of immune cell subtypes of CD4+T cells, but it can also increase the number of negative regulatory cells in inflammation. However, Anbainuo is not able to regulate all immune imbalances for patients with AS and CD8+T cells have not been rescued after treatment. The immune cells that cannot be improved in the treatment of TNF- α inhibitor need to be further investigated in future research.

Declarations

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Authors' Contributions

MCY and QL designed experiments strategy, analysed data, and wrote the manuscript. QJW performed experiments. MX analysed data. YFP, OJ and JRG designed the experimental strategy and determined clinical inclusion criteria. YTJ, JQ, LKF, YX, SYC, ZML, YLZ, LDT and MJZ characterised the patients and determined clinical inclusion criteria. All authors read and approved the final manuscript.

Availability of data and materials

The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

The Ethics Committee approved the study of the Third Affiliated Hospital of Sun Yat-Sen University, and informed consent was obtained from all patients.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

- [1]. Simone, D., M.M. Al and P. Bowness, Progress in our understanding of the pathogenesis of ankylosing spondylitis. *Rheumatology (Oxford)*, 2018. 57(suppl_6): p. vi4-vi9.
- [2]. Ye, L., et al., TCR usage, gene expression and function of two distinct FOXP3(+) Treg subsets within CD4(+) CD25(hi) T cells identified by expression of CD39 and CD45RO. *Immunol Cell Biol*, 2016. 94(3): p. 293-305.
- [3]. Miao, J. and P. Zhu, Functional Defects of Treg Cells: New Targets in Rheumatic Diseases, Including Ankylosing Spondylitis. *Curr Rheumatol Rep*, 2018. 20(5): p. 30.
- [4]. Wang, C., et al., T lymphocyte subset imbalances in patients contribute to ankylosing spondylitis. *Exp Ther Med*, 2015. 9(1): p. 250-256.
- [5]. Smith, J.A., Update on ankylosing spondylitis: current concepts in pathogenesis. *Curr Allergy Asthma Rep*, 2015. 15(1): p. 489.
- [6]. Duan, Z., et al., The immune dysfunction in ankylosing spondylitis patients. *Biosci Trends*, 2017. 11(1): p. 69-76.
- [7]. Bautista-Caro, M.B., et al., Increased frequency of circulating CD19+ CD24hi CD38hi B cells with regulatory capacity in patients with Ankylosing spondylitis (AS) naive for biological agents. *PLoS One*, 2017. 12(7): p. e0180726.
- [8]. Long, S., et al., High frequency of circulating follicular helper T cells is correlated with B cell subtypes in patients with ankylosing spondylitis. *Exp Ther Med*, 2018. 15(5): p. 4578-4586.
- [9]. Mavropoulos, A., et al., Breg Cells Are Numerically Decreased and Functionally Impaired in Patients with Systemic Sclerosis. *Arthritis Rheumatol*, 2016. 68(2): p. 494-504.
- [10]. Matsushita, T., et al., Decreased levels of regulatory B cells in patients with systemic sclerosis: association with autoantibody production and disease activity. *Rheumatology (Oxford)*, 2016. 55(2): p. 263-7.
- [11]. Chen, M., et al., Defective Function of CD24(+) CD38(+) Regulatory B Cells in Ankylosing Spondylitis. *DNA Cell Biol*, 2016. 35(2): p. 88-95.
- [12]. Schulte-Wrede, U., et al., An explorative study on deep profiling of peripheral leukocytes to identify predictors for responsiveness to anti-tumour necrosis factor alpha therapies in ankylosing spondylitis: natural killer cells in focus. *Arthritis Res Ther*, 2018. 20(1): p. 191.
- [13]. van der Linden, S., H.A. Valkenburg and A. Cats, Evaluation of diagnostic criteria for ankylosing spondylitis. A proposal for modification of the New York criteria. *Arthritis Rheum*, 1984. 27(4): p. 361-8.

- [14]. Chen, X.X., et al., A randomized, controlled trial of efficacy and safety of Anbainuo, a bio-similar etanercept, for moderate to severe rheumatoid arthritis inadequately responding to methotrexate. *Clin Rheumatol*, 2016. 35(9): p. 2175-83.
- [15]. Zochling, J., Measures of symptoms and disease status in ankylosing spondylitis: Ankylosing Spondylitis Disease Activity Score (ASDAS), Ankylosing Spondylitis Quality of Life Scale (ASQoL), Bath Ankylosing Spondylitis Disease Activity Index (BASDAI), Bath Ankylosing Spondylitis Functional Index (BASFI), Bath Ankylosing Spondylitis Global Score (BAS-G), Bath Ankylosing Spondylitis Metrology Index (BASMI), Dougados Functional Index (DFI), and Health Assessment Questionnaire for the Spondylarthropathies (HAQ-S). *Arthritis Care Res (Hoboken)*, 2011. 63 Suppl 11: p. S47-58.
- [16]. Finak, G., et al., Standardizing Flow Cytometry Immunophenotyping Analysis from the Human ImmunoPhenotyping Consortium. *Sci Rep*, 2016. 6: p. 20686.
- [17]. An, H., et al., The absolute counts of peripheral T lymphocyte subsets in patient with ankylosing spondylitis and the effect of low-dose interleukin-2. *Medicine (Baltimore)*, 2019. 98(15): p. e15094.
- [18]. Niu, X.Y., et al., Peripheral B-cell activation and exhaustion markers in patients with ankylosing spondylitis. *Life Sci*, 2013. 93(18-19): p. 687-92.
- [19]. Sulicka, J., et al., Endothelial dysfunction is independent of inflammation and altered CCR7 T cell expression in patients with ankylosing spondylitis. *Clin Exp Rheumatol*, 2017. 35(5): p. 844-849.
- [20]. Dulic, S., et al., The Impact of Anti-TNF Therapy on CD4+ and CD8+ Cell Subsets in Ankylosing Spondylitis. *Pathobiology*, 2018. 85(3): p. 201-210.
- [21]. Valizadeh, A., et al., Potential role of regulatory B cells in immunological diseases. *Immunol Lett*, 2019. 215: p. 48-59.
- [22]. Mauri, C. and M. Menon, The expanding family of regulatory B cells. *Int Immunol*, 2015. 27(10): p. 479-86.
- [23]. Matsumoto, M., et al., Interleukin-10-producing plasmablasts exert regulatory function in autoimmune inflammation. *Immunity*, 2014. 41(6): p. 1040-51.
- [24]. Shen, P., et al., IL-35-producing B cells are critical regulators of immunity during autoimmune and infectious diseases. *Nature*, 2014. 507(7492): p. 366-370.
- [25]. Li, Z., et al., Epigenetic and gene expression analysis of ankylosing spondylitis-associated loci implicate immune cells and the gut in the disease pathogenesis. *Genes Immun*, 2017. 18(3): p. 135-143.
- [26]. Allen, R.L., et al., Cutting edge: HLA-B27 can form a novel beta 2-microglobulin-free heavy chain homodimer structure. *J Immunol*, 1999. 162(9): p. 5045-8.

- [27]. Bowness, P., et al., Th17 cells expressing KIR3DL2+ and responsive to HLA-B27 homodimers are increased in ankylosing spondylitis. *J Immunol*, 2011. 186(4): p. 2672-80.
- [28]. Croft, M., The TNF family in T cell differentiation and function—unanswered questions and future directions. *Semin Immunol*, 2014. 26(3): p. 183-90.
- [29]. Bridgwood, C., et al., Interleukin-23 pathway at the enthesis: The emerging story of enthesitis in spondyloarthritis. *Immunol Rev*, 2020. 294(1): p. 27-47.
- [30]. Sherlock, J.P., et al., IL-23 induces spondyloarthritis by acting on ROR-gamma⁺ CD3⁺CD4⁺CD8⁻ enthesal resident T cells. *Nat Med*, 2012. 18(7): p. 1069-76.
- [31]. Orchard, T.R., et al., Clinical phenotype is related to HLA genotype in the peripheral arthropathies of inflammatory bowel disease. *Gastroenterology*, 2000. 118(2): p. 274-8.
- [32]. Kamada, N., et al., Role of the gut microbiota in immunity and inflammatory disease. *Nat Rev Immunol*, 2013. 13(5): p. 321-35.
- [33]. Feldtkeller, E., et al., Age at disease onset and diagnosis delay in HLA-B27 negative vs. positive patients with ankylosing spondylitis. *Rheumatol Int*, 2003. 23(2): p. 61-6.

Supplementary Figure Legend

Supplementary Fig 1. Representative gating strategy to identify T lymphocyte (A-N) and B lymphocyte (O-X) subsets. CM: Central Memory, EM: Effector Memory, Th cell: Helper T cell, Tfh cell: Follicular helper T cell, Tc cell: Cytotoxic T cell, EMRA cells: terminal differentiated effector memory cells, Treg cell: Regulatory T cell, Breg cell: Regulatory B cell.

Supplementary Fig 2. The proportion of regulatory lymphocytes is shown for both the AS and HC groups. P-value summary: (*, P<0.05) (**, P<0.01) (***, P<0.001) (****, P<0.0001). Treg cell: Regulatory T cell, Breg cell: Regulatory B cell.

Figures

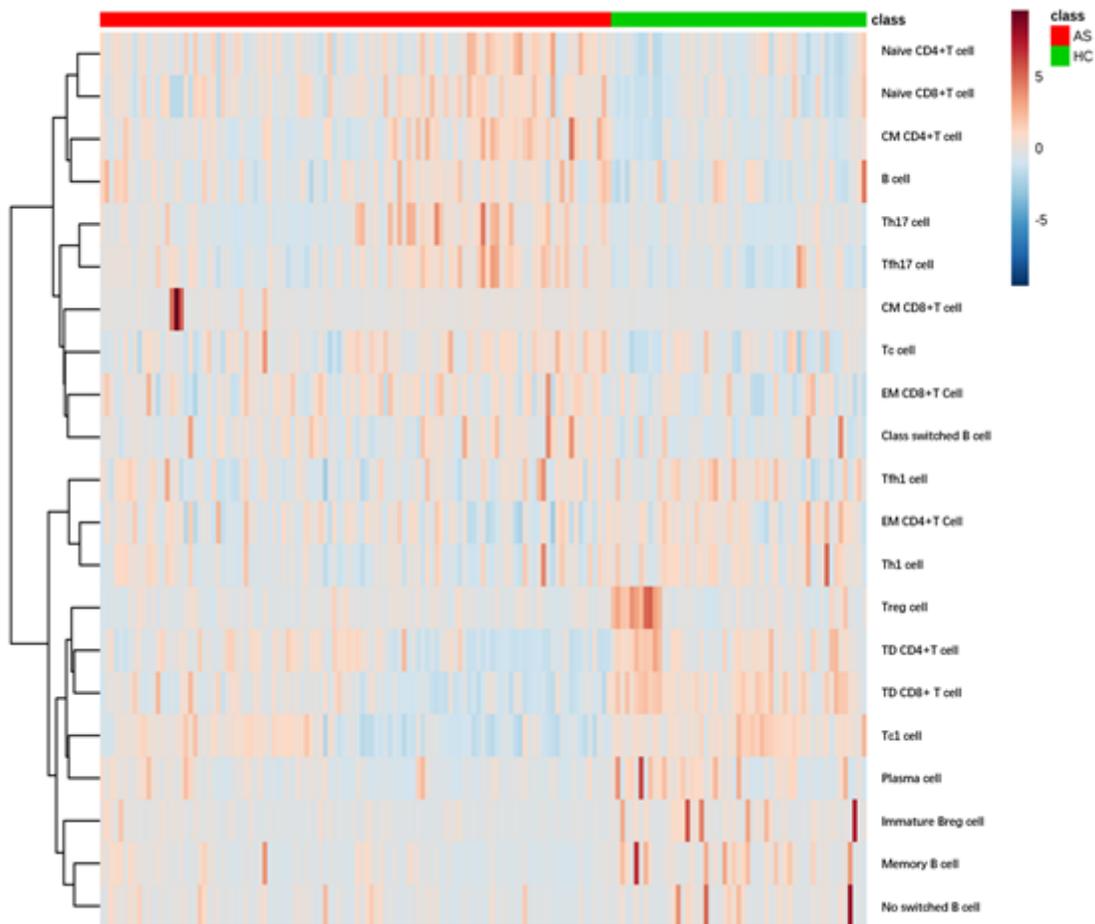


Figure 1

Cluster analyses of immunophenotypic parameters that were differentially expressed in the AS patients and HCs at the expanded validation phase. Each column represents individual AS patient or HC, and the color code in the first line above the graph indicates AS group (Red) or HC group (Green). The rows represent immune cells that are differentially expressed in AS and HC with a p-value<0.05. The magnitude of parameter expression is color-coded with red for a relative increase in expression and blue for a relative decrease in expression. CM CD4+T cell: Central Memory CD4+T cell, EM CD4+T cell: Effector Memory CD4+T cell, CM CD8+T cell: Central Memory CD8+T cell, EM CD8+T cell: Effector Memory CD8+T cell, Th cell: Helper T cell, Tfh cell: Follicular helper T cell, Tc cell: Cytotoxic T lymphocyte, Treg cell: Regulatory T cell, Breg cell: Regulatory B cell

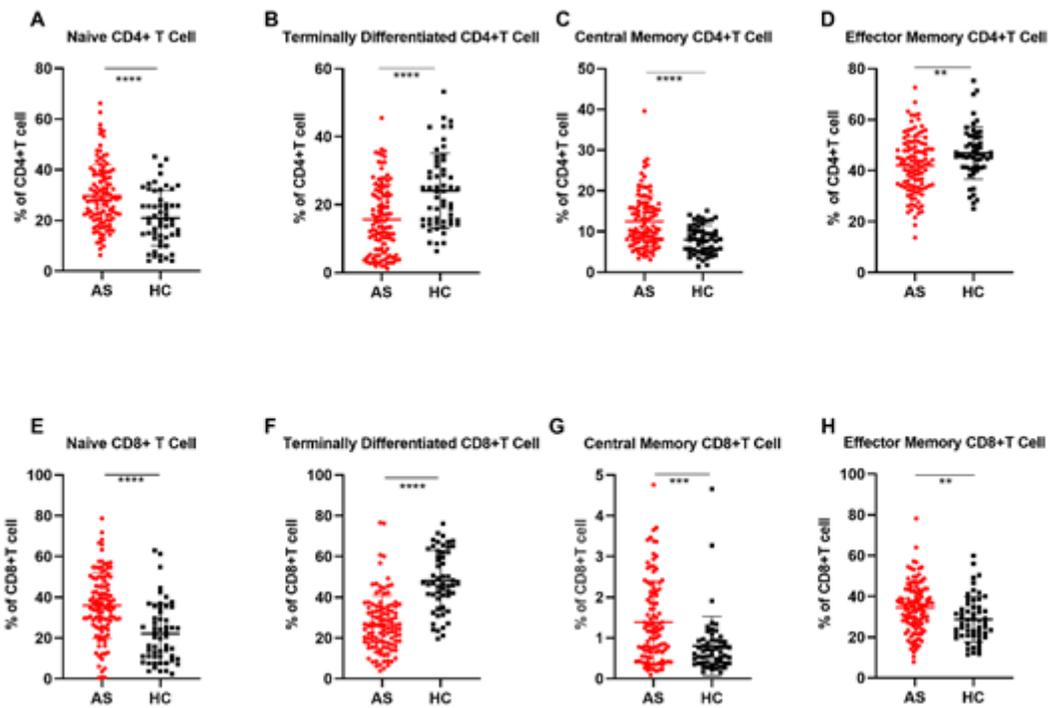


Figure 2

Differences in CD4+ T cells and CD8+ T cells in the AS and HC groups at different stages of differentiation. P-value summary: (*, P<0.05) (**, P<0.01) (***, P<0.001) (****, P<0.0001).

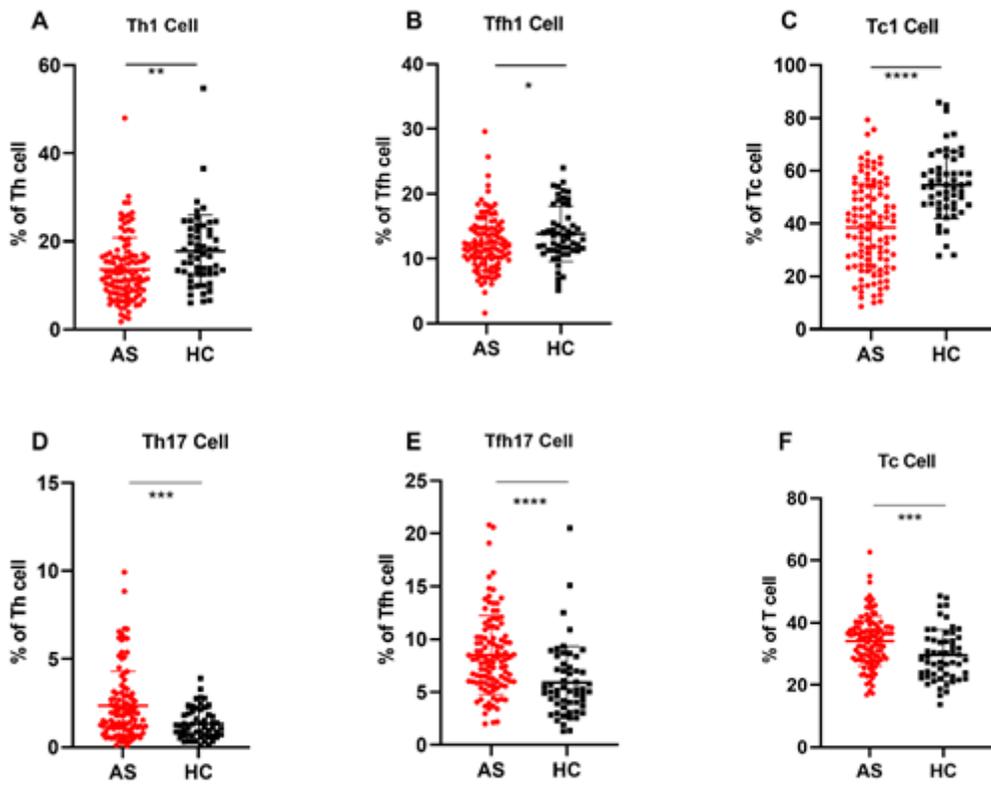


Figure 3

Differences in the percentage of Th cells, Tfh cells, and Tc cells in the AS and HC groups. P-value summary: (*, $P < 0.05$) (**, $P < 0.01$) (***, $P < 0.001$) (****, $P < 0.0001$). Th cell: Helper T cell, Tc cell: Cytotoxic T lymphocyte, Tfh cell: Follicular helper T cell.

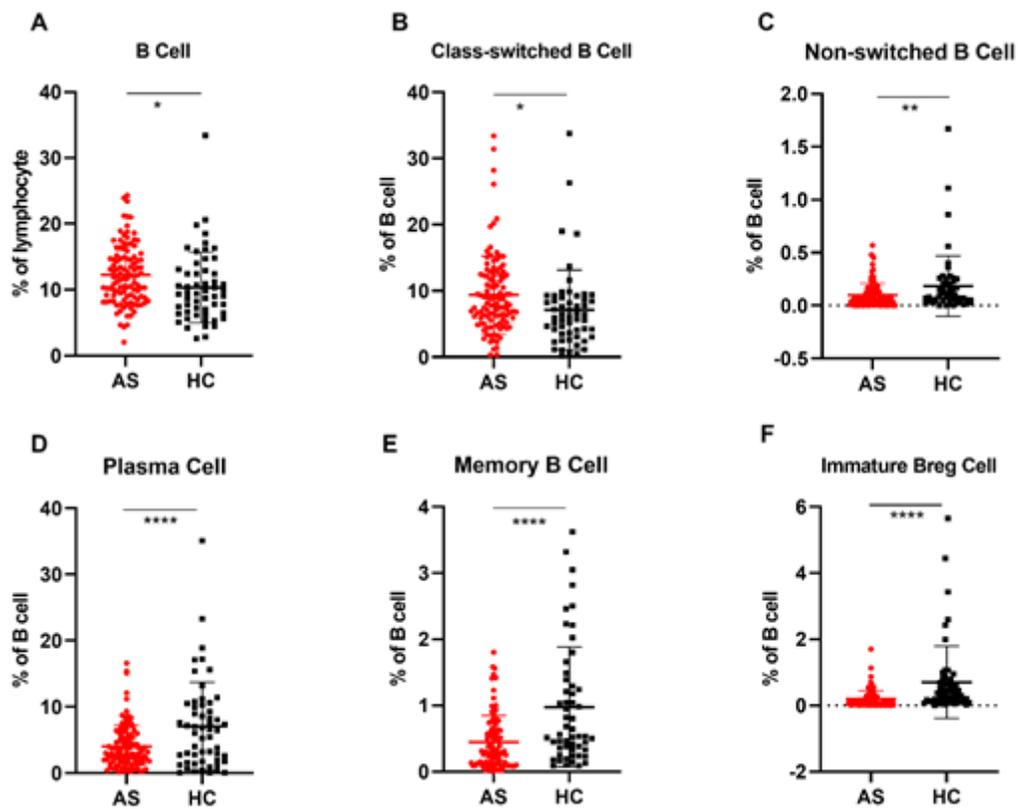


Figure 4

Differences between AS and HCs in B cell percentage. P-value summary: (*, $P < 0.05$) (**, $P < 0.01$) (***, $P < 0.001$) (****, $P < 0.0001$)

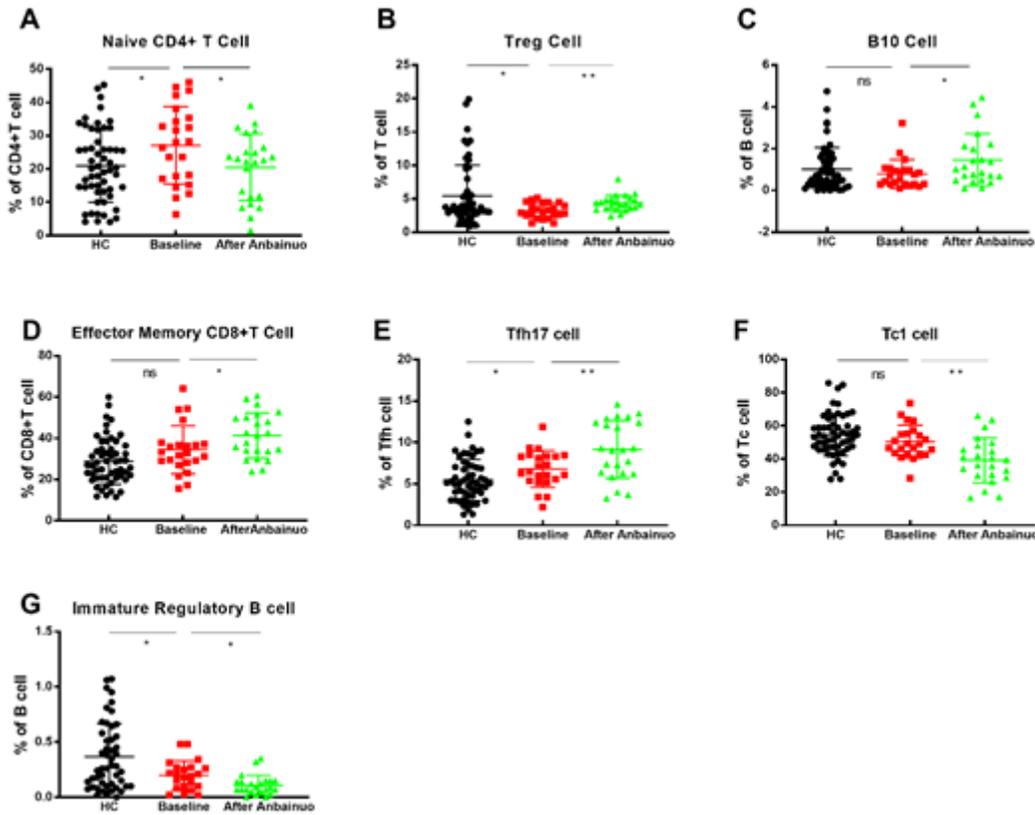


Figure 5

Following Anbainuo treatment, some T cell and B cell subtypes displayed significant changes. Anbainuo: Recombinant Human Tumor Necrosis Factor- α Receptor \times IgG Fc Fusion Protein for Injection, made in China. P-value summary: (*, $P < 0.05$) (**, $P < 0.01$) (***, $P < 0.001$) (****, $P < 0.0001$). Treg cell: Regulatory T cell, Tc cell: Cytotoxic T lymphocyte, Tfh cell: Follicular helper T cell. B10 cell: IL-10 producing regulatory B cell.

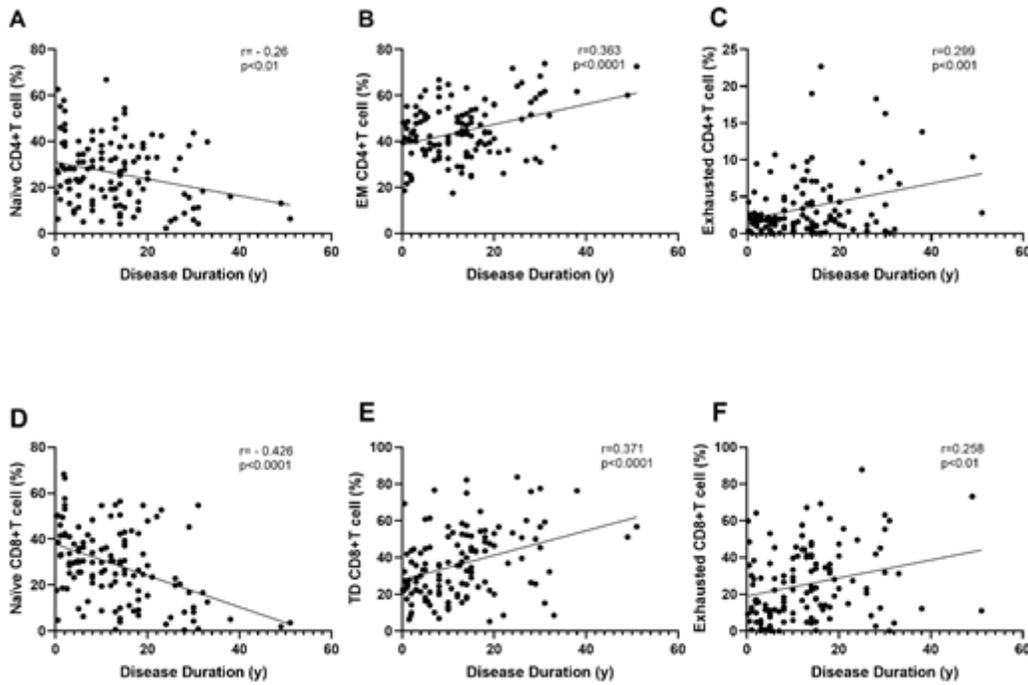


Figure 6

The association between disease duration and different differentiation stages of CD4+T cells and CD8+T cells in patients with AS. EM CD4+T cell: Effector Memory CD4+T cell. TD CD8+T cell: Terminally Differentiated CD8+T cell.

Supplementary Files

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