Effect of Stimulation of Subthalamic Nucleus on beta Oscillations and Thalamus Tremor Activity in a Computational Model of Parkinson's Disease

Seyed-Mojtaba Alavi

Shahid Rajaee Teacher Training University

Amin Mirzaei

Insilico Biotechnology AG

Alireza Valizadeh

Institute for Advanced Studies in Basic Sciences

Reza Ebrahimpour (✉ rebrahimpour@stru.ac.ir)

Shahid Rajaee Teacher Training University

Research Article

Keywords: Subthalamic Nucleus, Globus Pallidus, Parkinson's Disease, Deep Brain Stimulation, Beta Oscillations, Tremor, Computational Model

Posted Date: December 16th, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-125807/v1>

License: This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Effect of Stimulation of Subthalamic Nucleus on beta Oscillations and Thalamus Tremor Activity in a Computational Model of Parkinson's Disease

Seyed Mojtaba Alavi^{1,2}, Amin Mirzaei³, Alireza Valizadeh⁴, and Reza Ebrahimpour^{1,2,*}

¹Faculty of Computer Engineering, Shahid Rajaee Teacher Training University, Tehran, Iran

²School of Cognitive Sciences (SCS), Institute for Research in Fundamental Sciences (IPM), Tehran, Iran

³Insilico Biotechnology AG, Stuttgart, Germany

⁴Department of Physics, Institute for Advance Studies in Basic Sciences (IASBS), Zanjan, Iran

*rebrahimpour@sru.ac.ir, ebrahimpour@ipm.ir

ABSTRACT

Parkinson's disease (PD) is associated with abnormal β band oscillations (13-30 Hz) in the cortico-basal ganglia circuits. Abnormally increased striato-pallidal inhibition and strengthening the synaptic coupling between subthalamic nucleus (STN) and globus pallidus externa (GPe), due to the loss of dopamine, are accounted as the potential sources of β oscillations in the basal ganglia. Deep brain stimulation (DBS) of the basal ganglia subregions is known as a way to reduce the pathological β oscillations and motor deficits related to PD. Despite the success of the DBS, its underlying mechanism is poorly understood and, there is controversy about the inhibitory or excitatory role of the DBS in the literature. Here, we utilized a computational network model of basal ganglia which consists STN, GPe, globus pallidus interna (GPi), and thalamus neuronal population. This model can capture healthy and pathological β oscillations as what has been observed in experimental studies. Using this model, we investigated the effect of DBS to understand whether its effect is excitatory or inhibitory. Our results show that the excitatory DBS (EDBS) is able to quench the pathological synchrony and β oscillations, while, applying inhibitory DBS (IDBS) failed to quench the PD signs. In addition, the EDBS ameliorated the thalamic activity related to tremor in the model, while, the IDBS outperformed. However, with the help of the model results, we conclude that the effect of the DBS on its target is excitatory.

keywords: Subthalamic Nucleus, Globus Pallidus, Parkinson's Disease, Deep Brain Stimulation, Beta Oscillations, Tremor, Computational Model

Introduction

Parkinson's disease (PD) is resulted from malfunctioning of basal ganglia (BG;¹⁻³). This malfunctioning is following degeneration of dopaminergic neurons in pars compacta section of the substantia nigra (SNc;^{2,4}). Rigidity, bradykinesia, tremor, and postural instability are common signs of PD^{5,6}. In addition, this disorder is associated with excessive synchronization and abnormally β band (13-30 Hz) oscillations in subregions of BG⁶⁻¹⁰ and enhanced β oscillations are known as the biomarker of the PD¹¹⁻¹³. However, the source of the oscillations is still under debate. Several studies have suggested the subthalamo-pallidal circuit as a main source of the generation of the β oscillations in experimental and computational studies^{7,8,14-24}. While, the induction of the β oscillations from cortex to the BG has also been claimed^{6,25,26}. Furthermore, deep brain stimulation (DBS) of the BG subregions, mainly the subthalamic nucleus (STN), is a standard approach to treating PD²⁷⁻³³. Although the DBS quenches the β oscillations and improves the PD motor symptoms, its underlying mechanism is poorly understood³⁴⁻³⁷ and, there is a controversy between the excitatory and inhibitory role of the DBS.

Reduction of firing rate of the stimulated neuronal area has been observed in human^{38,39} and monkeys⁴⁰ with PD which remarks the inhibitory role of the DBS. Moreover, several mechanisms suggested to explain the inhibitory role of the DBS such as depolarization block^{36,41}, inactivation of voltage-gated currents⁴²⁻⁴⁴, and activation of inhibitory afferents^{38,40,45-50}. In a study has been suggested the effect of DBS is inhibitory⁴⁵. its authors showed the GPi neurons were inhibited during stimulation, while the neurons were inhibited using GABA blocker. On the other hand, the excitatory role of the DBS has also been suggested by several studies^{48,51}. Applying DBS on internal segment of globus pallidus (GPi) reduces firing rates of thalamic neurons which are inhibited by the GPi⁵². Also, applying the DBS on STN neurons (the excitatory neuronal population) increases firing rate of GPi, globus pallidus externa (GPe), and substantia nigra pars reticulata (SNr) of human and animal with PD⁵³⁻⁵⁵ which support the excitatory effect of the DBS.

Computational studies have also explored the effect(s) of the DBS with inhibitory and excitatory pulses on the models of BG with PD signs. In⁵⁶ the BG has been modeled by leaky integrated and fire (LIF) neurons which can generate β oscillations in PD condition, and have shown that the PD β oscillations quenched when the model exposed to the inhibitory DBS. While, other computational studies which are based on Hodgkin-Huxley type neurons, used excitatory DBS to suppress PD like oscillations^{18,57-62}. To investigate whether the effect of the DBS on its target is inhibitory or excitatory, in the current study, we used a computational model based on a variation of the model proposed in¹⁸ that resulted in β oscillations in PD condition. In our model the β oscillations are generated by the subthalamo-pallidal circuit and we have shown that the DBS with excitatory pulses quenches them (and with inhibitory pulses not). In addition, since the activity of the thalamus is correlated with resting tremor in limb^{63,64}, we studied the thalamus activity in the model during the PD condition. We tested the effect of DBS on thalamus activity related to resting tremor in the model and it is shown the DBS with excitatory pulses ameliorate the tremor while the inhibitory DBS intensify the signs of tremor.

Materials and methods

Structure of the network model

The network model consists of STN, GPe, GPi, and thalamus. Each neuronal population includes 20 Hodgkin-Huxley type neurons. The basic network model structure is similar to¹⁸ and¹⁷. the network model structure resembles the sparse pattern of connectivity⁶⁵ proposed in¹⁷. STN excites GPe and GPi while receiving inhibitory input from GPe. Similar to¹⁸ each STN neuron receives inhibitory input from two GPe neurons. In¹⁸ each GPe neuron was receiving excitatory input from three STN neurons, while in our simulation, each GPe neuron receives excitatory input from one STN neuron. Similar to¹⁸ each GPi neuron receives excitatory input from one STN neuron. In addition, each GPi neuron receives inhibitory input from two GPe neurons. In our simulation each thalamus neuron receives inhibitory input from one GPi neuron, while in¹⁸ each thalamus neuron was receiving inhibitory input from eight GPi neurons. See Figure 1 for more details on the network connectivity and structure.

Neuron and synapse model

Membrane potential of the STN, GPe, and GPi neurons in the network model were computed using the following differential equations:

$$C_m v' = -I_L - I_{Na} - I_K - I_{Ca} - I_T - I_{AHP} - I_{pre \rightarrow post} + I_{app}, \quad (1)$$

$$X' = \frac{\phi_X(X_\infty(v) - X)}{\tau_X(v)}, \quad (2)$$

where I_L , I_{Na} , I_K , I_{Ca} , I_T , and I_{AHP} are the leak, sodium, potassium, high threshold calcium, low threshold calcium, and after hyper polarization currents, respectively. I_{app} is the external current applied to the neurons (i.e. the DBS current). $I_{pre \rightarrow post}$ is synaptic current from the presynaptic to the postsynaptic neuron. X represents gating channels such as potassium channels (n), opening (m) and closing (h) sodium channels, and low threshold calcium channels (r). The $\tau_X(v)$ in equation 2 is defined as follows:

$$\tau_X(v) = \tau_X^0 + \frac{\tau_X^1}{1 + e^{-(v - \theta_X^T)}}. \quad (3)$$

While, in the GPe and GPi neurons the $\tau_X(v)$ is constant and equal to τ_r . The ionic currents used in equation 1 were computed as follows:

$$I_L = g_L(v - E_L), \quad (4)$$

$$I_{Na} = g_{Na} m_\infty^3(v) h(v - E_{Na}), \quad (5)$$

$$I_K = g_K n^4(v - E_K), \quad (6)$$

$$I_{Ca} = g_{Ca} s_\infty^2(v)(v - E_{Ca}), \quad (7)$$

$$I_T = g_T a_\infty^3(v) b_\infty^2(r)(v - E_{Ca}). \quad (8)$$

In the equations 4 to 8 the $X = n, h$ is the ionic gating channel variables (h for closing sodium channel and n for potassium). In these equations the $X_\infty = m, a, r$ or s is the steady state of the ionic gating channels (m for opening sodium channel, a for T-type and s for L-type calcium channel) and is computed by the equation 9.

$$X_\infty(v) = \frac{1}{1 + e^{-(v - \theta_X)/\sigma_X}}, \quad (9)$$

Figure 1. Network model structure and finding appropriate DBS currents. (A) Schematic of the network model. The DBS is considered to be either inhibitory or excitatory input to the STN. (B) Details of the network connectivity. The i th STN neuron excites the i th GPe and GPi neurons. The i th GPe neuron inhibits the $(i-1)$ th and $(i+1)$ th STN, GPi, and GPe neurons. Each GPi neuron inhibits its corresponding thalamus neuron. (C) The STN mean firing rates for IDBS and EDBS shown versus the amplitude of the stimulation current. Inset is zoomed-in presentation of the results in the rectangle. Each point in the plots is averaged over 20 trials. The IDBS curve crosses the EDBS curve at 126.57 and 147.36 pA points.

But, the function $b_\infty(r)$ used in 8 is computed with different equation:

$$b_\infty(r) = \frac{1}{1 + e^{(r-\theta_b)/\sigma_b}} - \frac{1}{e^{-\theta_b/\sigma_b}}. \quad (10)$$

The after hyper-polarization (AHP) current used in equation 1 (I_{AHP}) is

$$I_{AHP} = g_{AHP}(v - E_K)([Ca]/([Ca] + k_1)), \quad (11)$$

where the $[Ca]$ is the intra-cellular calcium concentration:

$$[Ca]' = \varepsilon(-I_{Ca} - I_T - k_{Ca}[Ca]). \quad (12)$$

The parameters and their values of STN, GPe, and GPi neurons are presented in table 1 to 3.

Table 1. Parameters and their corresponding values of STN neurons. The stars indicate transition from healthy to PD.

Parameter	Value	Parameter	Value	Parameter	Value	Parameter	Value
g_L	2.25	τ_h^0	1	θ_a	-63	σ_n	8
g_{Na}	30	τ_n^0	1	θ_b	0.4	σ_r	-2
g_K	40	τ_r^0	40	θ_s	-39	σ_a	7.8
g_T	0.5	ϕ_h	5	θ_h^τ	-57	σ_b	-0.1
g_{Ca}	0.5	ϕ_n	5	θ_n^τ	-80	σ_s	8
g_{AHP}	9	ϕ_r	2	θ_r^τ	68	σ_h^τ	-3
E_L	-60	k_1	15	θ^H	-39	$\theta_X \tau_n$	-26
E_{Na}	55	k_{Ca}	22.5	θ	20	σ_r^τ	-2.2
E_K	-80	e	3×10^{-5}	α	2	σ^H	8
E_{Ca}	140	θ_m	-30	$g_{GPe \rightarrow STN}$	$2.2 \mapsto 7^*$	β	0.08
τ_h^1	500	θ_h	-39	$E_{GPe \rightarrow STN}$	-85	I_{app}	$8.4 \mapsto 3^*$
τ_h^1	100	θ_n	-32	σ_m	15	C_m	1
θ_r^1	17.5	θ_r	-67	σ_h	-3.1		

Membrane potential of thalamus neurons in the network model is computed using the following differential equations:

$$C_m v' = -I_L - I_{Na} - I_K - I_T - I_{GPi \rightarrow Th} + I_{SMC}, \quad (13)$$

$$h' = \frac{h_\infty(v) - h}{\tau_h(v)}, \quad (14)$$

$$r' = \frac{r_\infty(v) - r}{\tau_r(v)}, \quad (15)$$

where I_L , I_{Na} , I_K , and I_T are the leak, sodium, potassium, and low threshold calcium currents, respectively. $I_{GPi \rightarrow Th}$ is the synaptic current from a GPi neuron to a thalamus neuron in the network model. The I_{SMC} represents cortico-thalamic sensorimotor pulses applied to the thalamus neurons. The equations 16 to 19 compute the ionic currents used in equation 13.

$$I_L = g_L(v_{Th} - E_L), \quad (16)$$

$$I_{Na} = g_{Na} m_\infty^3(v_{Th}) h_{Th}(v_{Th} - E_{Na}), \quad (17)$$

$$I_K = g_K [0.75(1 - h_{Th})]^4 [v_{Th} - E_K], \quad (18)$$

$$I_T = g_T p_\infty^2(v_{Th}) r_{Th}(v_{Th} - E_T) m \quad (19)$$

Table 2. Parameters and their corresponding values of GPe neurons. The stars indicate transition from healthy to PD.

Parameter	Value	Parameter	Value	Parameter	Value	Parameter	Value
g_L	0.1	τ_n^0	0.05	θ_s	-35	σ_r	-2
g_{Na}	120	τ_r	30	θ_h^τ	-40	σ_a	2
g_K	30	ϕ_h	0.135	θ_n^τ	-40	σ_s	2
g_T	0.5	ϕ_n	0.165	θ^H	-57	σ_h^τ	-12
g_{Ca}	0.15	ϕ_r	1	θ	30	σ_n^τ	-12
g_{AHP}	30	k_1	30	α	5	σ^H	2
E_L	-55	k_{Ca}	2.4	$g_{STN \rightarrow GPe}$	$0.01 \mapsto 0.55^*$	β	0.14
E_{Na}	55	e	0.0055	$E_{STN \rightarrow GPe}$	0	I_{app}	$5.9 \mapsto 0.5^*$
E_K	-80	θ_m	-37	$g_{GPe \rightarrow GPe}$	$0.01 \mapsto 0.9^*$	C_m	1
E_{Ca}	120	θ_h	-58	$E_{GPe \rightarrow GPe}$	-100		
τ_h^1	0.27	θ_n	-50	σ_m	10		
τ_n^1	0.27	θ_r	-70	σ_h	-12		
θ_h^0	0.05	θ_a	-57	σ_n	14		

and the functions used in equations 14 to 19 are computed as follows:

$$h_\infty(v_{Th}) = \frac{1}{1 + e^{(v_{Th} + 41)/4}}, \quad (20)$$

$$r_\infty(v_{Th}) = \frac{1}{1 + e^{(v_{Th} + 48)/4}}, \quad (21)$$

$$\tau_h(v_{Th}) = \frac{1}{a_h(v_{Th}) + b_h(v_{Th})}, \quad (22)$$

$$\tau_r(v_{Th}) = 28 + e^{-(v_{Th} + 25)/10.5}, \quad (23)$$

$$m_\infty(v_{Th}) = \frac{1}{1 + e^{-(v_{Th} + 37)/7}}, \quad (24)$$

$$p_\infty(v_{Th}) = \frac{1}{1 + e^{-(v_{Th} + 60)/6.2}}, \quad (25)$$

which the $a_h(v_{Th})$ and $b_h(v_{Th})$ are

$$a_h(v_{Th}) = 0.128e^{-(v_{Th} + 46)/18}, \quad (26)$$

$$b_h(v_{Th}) = \frac{4}{1 + e^{-(v_{Th} + 23)/5}}. \quad (27)$$

The parameters and their values of thalamus neurons are presented in table 4.

In addition to modification of the network structure we modified the network model parameters compared to the Terman et al. (2004) as follows: for the STN neurons g_{Na} was decreased from 37.5 to $30nS/\mu m^2$. The g_K was decreased from 45 to $40nS/\mu m^2$. The value of ϕ was taken from Park et al. (2011; i.e. $\phi_n = \phi_h = 5$, $\phi_r = 2$). The value of ε was considered to be $3 \times 10^{-5}ms^{-1}$. The $g_{GPe \rightarrow STN}$ and I_{app} of the STN in the healthy state is $2.2nS/\mu m^2$ and $8.4pA/\mu m^2$, respectively. In the PD state of the network model these two parameters were changed to $7nS/\mu m^2$ ^{266–69} and $3pA/\mu m^2$ ²⁷⁰, respectively.

Table 3. Parameters and their corresponding values of GPi neurons. The stars indicate transition from healthy to PD.

Parameter	Value	Parameter	Value	Parameter	Value	Parameter	Value
g_L	0.1	τ_n^0	0.05	θ_s	-35	σ_r	-2
g_{Na}	120	τ_r	30	θ_h^τ	-40	σ_a	2
g_K	30	ϕ_h	0.1	θ_n^τ	-40	σ_s	2
g_T	0.5	ϕ_n	0.135	θ^H	-57	σ_h^τ	-12
g_{Ca}	0.15	ϕ_r	1	θ	30	σ_n^τ	-12
g_{AHP}	30	k_1	30	α	5	σ^H	2
E_L	-55	k_{Ca}	2.4	$g_{STN \rightarrow GPe}$	0.005 $\mapsto$ 1.1*	β	0.14
E_{Na}	55	e	0.0055	$E_{STN \rightarrow GPe}$	0	I_{app}	7.7 $\mapsto$ 4*
E_K	-80	θ_m	-37	$g_{GPe \rightarrow GPe}$	0.01 $\mapsto$ 1.9*	C_m	1
E_{Ca}	120	θ_h	-58	$E_{GPe \rightarrow GPe}$	-100		
τ_h^1	0.27	θ_n	-50	σ_m	10		
τ_n^1	0.27	θ_r	-70	σ_h	-12		
θ_h^0	0.05	θ_a	-57	σ_n	14		

Table 4. Thalamus parameters and the corresponding values

Parameter	Value
g_L	0.05
E_L	-70
g_{Na}	3
E_{Na}	50
g_T	5
E_T	0
$g_{GPi \rightarrow Th}$	0.05
$E_{GPi \rightarrow Th}$	-85
θ^H	-57
σ^H	2

To simulate GPe neurons in the network model, we set $\phi_h = 0.135$, $\phi_n = 0.165$, $\phi_r = 1$, and $\varepsilon = 0.0055$ (similar to Park et al., 2011). The $g_{GPe \rightarrow GPe}$, $g_{STN \rightarrow GPe}$, and I_{app} of GPe in the healthy state were 0.01, 0.01 nS/ μm^2 , and 5.9 pA/ μm^2 , respectively. To simulate the PD state of the network model, these parameters were changed to 0.9, 0.55 nS/ μm^2 ^{66–69}, and 0.5 pA/ μm^2 , respectively. Note that in the PD state of the network model I_{app} of the GPe and GPi decreases leading to less activity of the GPe

and GPi neurons due to the increasing striatal inhibition (explained in⁵⁶) in the network model. Parameters of the GPi neurons are similar to the GPe neurons with the difference that for the GPi neurons, $\phi_h = 0.1$ and $\phi_n = 0.135$. The $g_{GPe \rightarrow GPi}$, $g_{STN \rightarrow GPi}$, and I_{app} of GPi in the healthy state were 0.01, $0.005 \text{ nS}/\mu\text{m}^2$, and $7.7 \text{ pA}/\mu\text{m}^2$. To simulate the PD state of the network model, these parameters similar to GPe neurons were changed to 1.9, $1.1 \text{ nS}/\mu\text{m}^2$, and $4 \text{ pA}/\mu\text{m}^2$, respectively. Parameters of the thalamic neurons are the same as in¹⁸ with the difference that in our network model $g_{GPi \rightarrow Th}$ was $0.05 \text{ nS}/\mu\text{m}^2$. These modifications moved our network model activity more close to the experimental results.

The synaptic model used here is of conductance based type similar to the model used in^{17,18,61,62,71,72}. The synaptic currents used in equations 1 and 13 are computed as follows:

$$I_{pre \rightarrow post} = g_{pre \rightarrow post} (v - E_{pre \rightarrow post}) \sum s_j, \quad (28)$$

where the j is the index of presynaptic neuron. The parameter s in equation 28 is

$$s' = \alpha H_\infty(v_{pre} - \theta_{pre})(1 - s) - \beta_{pre} s, \quad (29)$$

where the $H_\infty(v)$ as follows:

$$H_\infty(v) = \frac{1}{1 + e^{-(v - \theta^H)/\sigma^H}}. \quad (30)$$

population firing rate

To compute the time resolved population firing rate for each neuronal population in the network model we used 10 milliseconds sliding window and shifted with steps of 1 millisecond over the entire simulation time while for each step we counted the number of spikes for all neurons in the population and converted it to spikes per second (i.e. to Hz).

Sensorimotor and DBS pulses

The sensorimotor and DBS pulses are simulated using the following equation:

$$I_{\text{pulse}} = A \times H\left(\sin\left(\frac{2\pi f t}{1000}\right)\right) \times \left(1 - H\left(\sin\left(\frac{2\pi f(t + \delta)}{1000}\right)\right)\right) \quad (31)$$

where A , f , t , and δ are pulse amplitude, frequency, time (in ms), and pulse duration (in ms), respectively. The $H(\cdot)$ is the Heaviside function. To find an appropriate value for the A for the excitatory and inhibitory DBS, we computed the STN mean firing rate (over 20 trials) by varying $A = 50 \text{ pA}/\mu\text{m}^2$ to $300 \text{ pA}/\mu\text{m}^2$ (Figure 1-C), then we chose the constraint the firing rate of the STN in excitatory DBS (EDBS) and inhibitory DBS (IDBS) to be equal in the cases. Therefore, to simulate EDBS, we set $A = 126.57 \text{ pA}/\mu\text{m}^2$ and $147.36 \text{ pA}/\mu\text{m}^2$, $f = 150 \text{ Hz}$, and $\delta = 0.1 \text{ ms}$. Parameter settings for IDBS were the same as EDBS, with the difference that $A = -126.57 \text{ pA}/\mu\text{m}^2$ and $-147.36 \text{ pA}/\mu\text{m}^2$. The DBS parameters are adapted from^{10,54,73–77}. For sensorimotor pulses (i.e. cortico-thalamic input), $A = 4.5 \text{ pA}/\mu\text{m}^2$, $f = 20 \text{ Hz}$, and $\delta = 5 \text{ ms}$. To generate irregular pulses we used Poisson process in equation 31.

Thalamus fidelity

Thalamus neurons in the network model show four types of responses to the cortico-thalamic input pulse: 1) Correct spike: a single thalamic spike in response to a cortico-thalamic input pulse. 2) Missed spike: refers to the case when there is no thalamic spike in response to a cortico-thalamic input spike. 3) Extra spike: refers to the case when a thalamic neuron shows more than one spike in response to a cortico-thalamic input pulse. 4) Undesired spike: occurs when a thalamic neuron spikes while there is no cortico-thalamic input spike. According to the four response types of the thalamus neurons to a single cortical pulse, the thalamus fidelity is computed using the following equation:

$$\text{thalamus fidelity} = 1 - \frac{N_M + N_E + N_U}{N_{\text{exp}}} \quad (32)$$

where the N_M , N_E , and N_U are the number of missed spikes, extra spikes, and undesired spikes, respectively. The N_{exp} is the number of expected thalamic spikes due to cortico-thalamic pulses. Since each cortical pulse is given to all thalamus neurons in the network model, we expect to observe that each thalamus neuron, relaying the cortico-thalamic pulse, emits a spike in response to the cortico-thalamic pulse. Therefore, the number of expected thalamic spikes in response to the cortical inputs (i.e. N_{exp}) equals to the number of thalamus neurons multiplied by the number of cortico-thalamic pulses^{18,78}.

Synchrony index

We used Fano Factor (FF) to measure synchronous spiking activity for each neuronal population in the network model. To compute FF we used the following equation:

$$FF = \frac{Var(PFR)}{E(PFR)} \quad (33)$$

where the $Var(.)$ and $E(.)$ are the variance and mean of the population firing rate (PFR), respectively. Higher FF values represent more synchrony in the spiking activity of a neuronal population in the network model^{56,79}.

Mean power spectral density

The power spectral density (PSD) of the population firing rates was computed using the Welch's method in python 2.7 (i.e. using `scipy.signal.welch` python package;⁸⁰). The sampling rate and the segment length were set to 1000 Hz and 1000 data points, respectively. Other parameters required for the `scipy.signal.welch` function were set to the predefined default values (see <https://docs.scipy.org/doc/scipy-0.14.0/reference/generated/scipy.signal.welch.html>). The mean power spectral density was computed by averaging over 50 simulations.

Oscillation index

Oscillation index was computed by dividing the area under the curve of a PSD in the β frequency range (i.e. between 13 to 30 Hz) by the area under the curve for the whole frequency range (i.e. from 1 to 500 Hz).

Tremor frequency

We assumed that the extra spikes of the thalamus neurons during the resting state of the network model (i.e. when there is no cortico-thalamic sensorimotor pulses) are related to limb tremor. So, to compute the tremor frequency, we measured the mean firing rate of thalamus neurons (in Hz), over the whole simulation period, during the resting state.

Simulation

The simulations were implemented in python 2.7. All differential equations were solved using `odeint` from SciPy library⁸⁰ with 0.05 ms time resolution (see <https://docs.scipy.org/doc/scipy-0.18.1/reference/generated/scipy.integrate.odeint.html>). To reduce the simulation time, we performed parallel programming using python message passing interface (MPI) in a cluster computing with 30 core processors (Intel 3.2 GHz). To avoid the initial transient network model responses, we did not consider the first 250 ms of each simulation in our analysis.

Results

The network model captures features of the healthy and PD BG

The activity of different BG regions in the healthy state is non-oscillatory and desynchronized^{3,81–83}. This feature is captured in our network model. Similar to the experimental results, the STN spiking activity in the healthy network model is asynchronous irregular (Figure 2A, top panel; the same for the GPe and GPi spiking activity, data not shown). This is also reflected in the STN population activity in the healthy state of the network model (Figure 2B, top panel). The STN population activity in the healthy network model is non-oscillatory. This leads to a flat PSD of the STN population firing rate in the healthy state (Figure 2C). Altogether, these results indicate that the healthy activity of the network model is irregular, inline with the experimental studies showing that the BG activity in the healthy state is non-oscillatory.

The STN, GPe, and GPi mean population firing rates, in the healthy state of the network model, are 19.4 ± 1.1 Hz, 45.47 ± 1.2 Hz and 56.52 ± 2 Hz, respectively which match the previously reported experimental values^{83,84}.

In the PD state, the network model neurons show synchronized bursts of spiking activities in the β frequency range (see the middle panels in Figures 2A and 2B). This also matches experimental studies which indicate synchronized β band oscillatory spiking activities in the BG as a hallmark of PD^{7–10,84–98}.

To bring the network model from the healthy to the PD state, we followed three steps. First, the I_{app} applied to the GPe and to the GPi neurons (Equation 1) was decreased from $5.9 \text{ pA}/\mu\text{m}^2$ and $7.7 \text{ pA}/\mu\text{m}^2$ (healthy state) to $0.5 \text{ pA}/\mu\text{m}^2$ and $4 \text{ pA}/\mu\text{m}^2$ (PD state), respectively. This leads to reduction in the activity of the GPe neurons (39.1 ± 0.8 Hz; independent two-tailed t-test, $p < 0.001$) in the PD state of the network model, compared to the healthy state (Figure 2D). This is inline with the experimental studies indicating that the GPe firing activity decreases during PD^{83,99,100}. However, despite decreasing the I_{app} applied to the GPi neurons, the GPi firing rate increases (64.9 ± 0.86 Hz; independent two-tailed t-test, $p < 0.001$) compared to the healthy state (Figure 2D;^{99,101,102}). The reason is that the lower GPe activity during the PD state of the network

model disinhibits the GPi neurons. Thereby, the GPi population firing rate in the PD network model increases compared to the healthy state (Figure 2D).

Second, the I_{app} applied to the STN neurons, representing cortico-subthalamic input, in the network model (Equation 1) was decreased from $8.4 \text{ pA}/\mu\text{m}^2$ (healthy state) to $3 \text{ pA}/\mu\text{m}^2$ (PD state). Such a change in the network model is inline with the experimental studies showing that the cortical activity decreases in PD⁷⁰ which can lead to less cortico-subthalamic drive, due to direct cortico-subthalamic connectivity^{103–105}. Note that despite decreasing the I_{app} of the STN neurons in the PD state of the network model, the STN activity increases ($27.9 \pm 1.5 \text{ Hz}$; independent two-tailed t-test, $p < 0.001$) compared to the healthy state (Figure 2D; ^{84,85}). The reason for this is STN disinhibition due to reduction in the activity of the GPe units in the PD state of the network model (Figure 2D).

Figure 2. EDBS is able to quench PD-like β oscillations and to improve thalamus fidelity in the network model. (A) Membrane potential of three STN neurons in the network model in the healthy state (top), PD state (middle), and during regular EDBS (bottom). The black vertical thick lines indicate 50 mV. (B) Time resolved population firing rate of the STN neurons in the Healthy state (top), PD state (middle), and during regular EDBS. (C) Mean power spectrum (average of 50 trials) of the STN time resolved population firing rate in the healthy state (light green), PD state (brown), and during regular (cyan) and irregular (light blue) EDBS. (D-F) Population mean firing rate (D), Fano factor (E), and oscillation index (F) of the STN, GPe, and GPi in the network model (error bars show standard deviation; color codes correspond to C). (G) Thalamus fidelity in the healthy state, PD state, and during regular and irregular EDBS.

Third, the synaptic connectivity in the subthalamo-pallidal circuit (STN to GPe and GPe to STN synapses) were strengthened in the PD state, compared to the healthy state (see materials and methods). Such subthalamo-pallidal synaptic strengthening is inline with the experimental studies showing that both STN to GPe and GPe to STN synapses are strengthened during PD^{66–69}.

Applying these three changes brings the network model from the healthy state to the PD state. STN neurons of the PD network model show synchronized bursts of spiking activity in the β frequency range (Figure 2A; the GPe and GPi neurons show the same behavior). The STN PD-like β band oscillations are also observed in the STN population firing rate (Figures 2B, 2C) as well as in the GPe and GPi population activities (Figures 2E, 2F). In addition, we measured the burst rate of GPi neurons and we found a significant increase in PD state compared to the healthy ($p < 0.001$, independent two-tailed t-test; Figure 4A) which matches with the previous experimental⁵⁴ and computational studies⁶¹.

To check whether the thalamus neurons are able to process the cortical inputs (i.e. cortico-thalamic motor commands), we measured thalamus fidelity in the healthy and PD states of the network model. Our results indicate that thalamus fidelity (see materials and methods) to the cortical motor commands decreases in the PD state of the network model compared to the

healthy state (Figure 2G). This result is inline with the previous computational studies^{18,59,60,78,106} indicating reduction of the thalamus fidelity during PD.

All in all, our network model can reproduce features of the experimental data for both healthy and PD state of the BG. Mainly, the network model show asynchronous irregular spiking activity in the healthy state and synchronous β band oscillatory activity in the PD state. In addition, thalamus fidelity decreases in the PD state of the network model compared to the healthy state.

Effects of EDBS and IDBS on the network model

The STN high frequency DBS has therapeutic effects on PD signs such as reducing the pathological β oscillations in the cortico-BG loop^{33,36,74,107–109}, and improving PD-related motor symptoms^{27–32,110–112}. However, the mechanism(s) underlying STN DBS is yet unknown. To understand whether the STN DBS therapeutic effects are due to excitation of STN or inhibition of STN, we applied EDBS (i.e. excitatory DBS) and IDBS (i.e. inhibitory DBS) to the STN neurons in the network model and investigated the effect(s) of each DBS type. We tested which DBS type can quench the PD-like β oscillations in the network model. Furthermore, we also tested which DBS type can improve the thalamus fidelity to the cortical motor commands. Here, we used 126.57pA and 147.36pA current amplitudes for both types of DBS. The STN mean firing rate is approximately the same in these two amplitudes (Figure 2 and 3D; not significant, independent two-tailed t-test; see materials and methods). Due to the similarity of results we reported the DBSs with 147.36pA current except for the Figure 5.

EDBS effects

Applying the EDBS (see materials and methods) to the STN neurons in the network model quenches the PD-like β oscillations in the STN (and in the other regions included in the network model; Figures 2A–C, bottom panels). We applied two EDBS types to the PD network model (i.e. regular and irregular EDBS; regular DBS is rectangular pulses with constant phase and the irregular DBS is similar to the regular with random phases; see materials and methods for more details on each EDBS type). To quantify the performance of regular and irregular EDBS on the PD-like β oscillations in the network model, we measured the Fano factor and the oscillation index of the STN, GPe, and GPi population firing rates while the STN was stimulated. For regular EDBS, both Fano factor and oscillation index dramatically decreased compared to the PD state ($P < 0.001$, independent two-tailed t-test), for all regions measured (i.e. STN, GPe, and GPi; Figures 2E, F). We also compared the performance of the regular and irregular EDBS on improving the thalamus fidelity in the network model. We found that both regular and irregular EDBS increased the thalamus fidelity (almost to the healthy level; Figure 2G). In addition, we found that the burst rate of the GPi neurons significantly decreased ($P < 0.001$, independent two-tailed t-test) in both types of EDBS state compared to the PD and also to the healthy (Figure 4A) which matches the previous experimental⁵⁴ and computational studies⁶¹.

All in all, this result indicates that applying regular/irregular EDBS to the STN neurons in the network model, quenches the PD-like β oscillations and improves the thalamus fidelity to the cortical motor command.

IDBS effects

Thus far, we showed that whether regular or irregular high frequency EDBS is able to quench PD-like β oscillations and improve the thalamus fidelity in the network model. Next, we tested whether high frequency inhibition of the STN neurons (IDBS) has the same effects as the EDBS in the network model. To do that, we stimulated the STN neurons in the pathological network model by high frequency inhibitory pulses (see materials and methods) and measured the PSD, Fano factor, and oscillation index of the STN population firing rate as well as the thalamus fidelity in the network model (Figure 3). Our simulation results show that regular and irregular IDBS (unlike the EDBS) fail to quench PD-like β oscillations and to improve the thalamus fidelity (Figure 3). Although the burst rate of the GPi neurons in regular and irregular IDBS slightly decreases ($p < 0.001$, independent two-tailed t-test) compared to the PD state, the GPi burst rate were still above the healthy state when the network model exposed to regular and irregular IDBS (Figure 4A). However, comparing the performance of EDBS and IDBS in the network model reveals that EDBS outperforms IDBS in improving PD signs (i.e. quenching PD-like β oscillations and improving the thalamus fidelity) in the network model. In the following, we explain why IDBS fails to improve the PD signs in the network model.

IDBS can not treat the pathological STN rebound bursting activity in the network model

We showed that only EDBS (and not IDBS) is able to quench the PD-like β oscillations in the network model (Compare Figure 2 with Figure 3). The reason why IDBS fails to quench the PD-like β oscillations in the network model is rebound bursting activity of the STN neurons due to IDBS. In line with the experimental studies^{87,113,114}, the STN neurons in the network model show rebound bursts of spiking activity when the strong inhibition finished. As the IDBS is a barrage of inhibitory inputs to the STN neurons in the network model (see materials and methods), the STN neurons react to it by rebound bursts of spiking activities (Figure 5C). STN rebound bursts lead to increase in the GPe spiking activity through subthalamo-pallidal excitatory pathway. Then, the higher GPe activity gives rise to inhibition of the STN neurons which resulted in rebound bursting due the T-type calcium current (Figure 5C). Such a mechanism retains the PD-like β oscillations in the network model during IDBS.

Figure 3. IDBS fails to quench PD-like β oscillations and to improve thalamus fidelity in the network model. (A) Membrane potential of three STN neurons in the network model in the PD state (top), during regular IDBS (middle), and during irregular IDBS (bottom). The black vertical thick lines indicate 50 mV. (B) Time resolved population firing rate of the STN neurons in the PD state (top), during regular IDBS (middle), and during irregular IDBS. (C) Mean power spectrum (average of 50 trials) of the STN time resolved population firing rate in the healthy state (light green), PD state (brown), and during regular (cyan) and irregular (light blue) IDBS. (D-F) Population mean firing rate (D), Fano factor (E), and oscillation index (F) of the STN, GPe, and GPi in the network model (error bars show standard deviation; color codes correspond to C). (G) Thalamus fidelity in the healthy state, PD state, and during regular and irregular IDBS.

Therefore, the reason why IDBS fails to quench the PD-like β oscillations in the network model is propagation of the STN rebound bursts through the subthalamo-pallidal loop.

During EDBS, two behaviors can occur depending on the current amplitude. First, for 147.36 pA , the STN neurons in the network model do not show rebound bursting activity (Figures 2, and 5A). The reason is that the STN spiking activity is driven by the EDBS pulses, counteracts with the pallido-subthalamic inhibition, thereby no STN rebound bursting activity can happen due to the non-sufficient T-type calcium current. Second, for 126.57 pA , the EDBS extends the burst durations and the inter-burst intervals which resulting in quenching the PD-like β oscillations (Figure 5B).

Tremor-like spiking activity of the thalamus

Tremor signs in healthy and PD state of the network model

So far, using our simulation results, we showed that the STN EDBS (and not IDBS) can quench pathological β oscillations emerging in the PD state of the network model. Next, we investigated the effect(s) of each stimulation scenario (i.e. EDBS or IDBS) on the resting state activity of the healthy and the PD network model. To simulate the resting state of the network model, we excluded the cortico-thalamic sensorimotor pulses (i.e. by setting the I_{SMC} in the equation 13 to zero for both healthy and PD network model simulations; see materials and methods). Our simulation results indicate that the thalamus neurons in the healthy network model, do not show any spiking activity during the resting state (Figure 6A). However, in the PD network model, the thalamus neurons show the mean spiking activity of $8.24 \pm 1\text{ Hz}$ during the resting state (Figure 7C). We considered this as resting state tremor-like activity in the network model because of three reasons. First, such thalamus activity is specific to the PD state of the network model (compare Figures 6A and 7C). Second, according to the experimental studies^{115–117} the tremor is driven by the pathological thalamus activity in PD. Third, the resting state pathological thalamus activity in the network model approximately is 8 Hz which matches the tremor frequency reported in the experimental studies^{63,64,88,118–121}. In the following, we explain why such tremor-like thalamus spiking activity emerges only in the PD network model (and not in

Figure 4. Burst profile of GPi neurons. (A) Burst rate of GPi neurons in healthy, PD, regular EDBS, irregular EDBS, regular IDBS and irregular IDBS states. (B and C) Mean burst duration (in msec) and mean number of spikes per burst in healthy, PD, regular/irregular EDBS, and regular/irregular IDBS states. The error bars show standard deviation.

the healthy network model).

The thalamus neurons in the network model receive inhibitory inputs only from the GPi neurons (Figure 1A; note that the cortico-thalamic input is set to zero during the resting state). Thereby, the resting state thalamus activity in the network model is driven by inhibitory inputs from the GPi. Our resting state simulation results show that the healthy GPi spiking activity is tonic (Figure 6B, bottom panel). Such tonic GPi spiking activity or weak bursting activity (burst with lower number of spikes; Figure 6B, two bottom panels; and also see Figure 4B and C) is not strong enough to sufficiently flow the T-type calcium current of the thalamus neurons (see the T-Current in Figure 5B). This leads to no thalamus spiking activity during the resting state and in the healthy condition of the network model (Figure 5B, top panel).

STN EDBS improves the thalamus tremor-like activity in the network model

Next, we investigated the effect(s) of EDBS and IDBS on the tremor-like thalamus activity in the network model. Mainly, we tested which DBS type (i.e. EDBS or IDBS) can reduce the resting state tremor-like thalamus spiking activity in the PD network model. Our simulation results show that applying EDBS to the STN neurons in the network model leads to a better

Figure 5. The role of inhibitory rebound bursting in STN neurons during EDBS and IDBS. (A) Membrane potential of an STN neuron (top), and its corresponding T-type calcium current (bottom; see materials and methods) during EDBS with current of 147.36 pA . (B) The same as A, EDBS with current of 126.57 pA . (C) The same as A, during IDBS.

Figure 6. Tremor frequency profile in healthy and PD state. (A) Membrane potential of 10 thalamus neurons in the healthy network model during resting state. The orange vertical thick line indicates 50 mV. (B) From top to bottom, membrane potential, low threshold T-type calcium current, synaptic inhibitory input from the connected GPi and membrane potential of the corresponding GPi neuron (see Materials and methods) in the healthy state of the network model. (C, D) The same as A and B for PD state of the network model.

performance as compared to applying the IDBS (Figure 6). In other words, STN EDBS reduces the tremor-like thalamus spiking activity nearly to the healthy state while STN IDBS fails to do so (Figure 7A-C). In the following, we explain why STN EDBS outperforms STN IDBS in improving the tremor-like thalamus activity in the network model.

In line with previous experimental and computational studies^{54,61}, GPi neurons in the PD state of our network model switch between long lasting hyperactivity and no activity states when the STN is stimulated by the EDBS (Figure 7D, bottom panel and also see the burst duration and number of spikes per burst in Figure 4B and C). During the GPi hyperactivity periods, thalamus neurons are strongly inhibited and thereby do not show tremor-like spiking activity anymore (Figure 7D). However, a single thalamus spike occurs at the end of the GPi activity (Figure 7D, top panel; also see Figure 7A). This single thalamus spike is due to after-hyperpolarization increase in the T-type calcium current of the thalamus neuron receiving the GPi strong inhibitory input (Figure 7D).

However, when the STN is exposed to the IDBS, the GPi neurons in the network model show regular oscillatory bursts of spiking activity in (Figure 7E, bottom panel). These strong GPi bursts (similar property to PD state; see Figure 4B and C) cause sufficient flow of the T-type calcium current of the thalamus neurons in the network model (see the T current fluctuations in Figure 7E) which consequently, leads to tremor-like thalamus spiking activity.

Altogether, our simulation results indicate that the EDBS outperforms the IDBS, not only in quenching the PD-like β oscillations in the network model (Figure 2), but also in reducing the tremor-like spiking activity of the thalamus neurons nearly to the healthy state (Figure 7C).

Figure 7. Tremor frequency profile of PD state during EDBS and IDBS. (A, B) Membrane potential of 10 thalamus neurons when STN is exposed to regular EDBS (A) and regular IDBS (B) during the resting state of the network model. The orange vertical thick lines indicate 50 mV. (C) Tremor-like frequency of thalamus in the healthy and PD states and when the STN is exposed to regular/irregular EDBS and regular/irregular IDBS (error bars show standard deviation). (D) From top to bottom, membrane potential, low threshold T-type calcium current, synaptic inhibitory input from the connected GPi and membrane potential of the corresponding GPi neuron (see Materials and methods) when STN is exposed to EDBS in the network model. (E) The same as D, when STN is exposed to the IDBS.

Discussion

Permanent and excessive β oscillations in BG is the hallmark of the PD^{11–13}. The DBS quenches β oscillations and improves motor symptoms related to PD^{27–33,36,74,107–112}. But, the mechanism of the DBS is poorly understood. In the present study, we investigated the generation of the pathological β oscillations in the BG by neural modeling and the effects of the DBS in different scenarios. We showed that the subthalamo-pallidal circuit is the potential source of the generation of the β oscillations based on the computational network model. Our findings confirm the rebound burst activity of the STN neurons is the key reason to generation of the β oscillations as what has been suggested in^{17,23}. Then we investigated the role of the DBS on quenching the β oscillations. We applied inhibitory and excitatory DBS on STN neurons to compare their effect with those observed in experimental studies. Thereby, we tuned current amplitude of each DBS types to have equal STN firing rates. The results show that the EDBS can counteract the pallido-subthalamic inhibition and therefore, the rebound burst of the STN neurons were controlled (Figure 5). Also, EDBS can extend the duration of STN bursts which results in quenching the β oscillations (Figure 5). While, the IDBS is not able to quench the β oscillations due to its ineffectiveness in the suppression of rebound burst activity of STN neurons and in the extension of burst duration. Our results suggest that effect of DBS on STN neurons is excitatory. Our simulation results also help to clarify the relationship between tremor and the activity of the basal ganglia and thalamus. We showed that the postinhibitory rebound activity of the thalamus neurons due to the strong inhibition of the GPi neurons is the potential reason of the resting state tremor. Like other signs of PD, the tremor activity of thalamus was quenched by EDBS while the IDBS was worsened it.

In the following, we discuss about the network model, the generation of β oscillations, the tremor, and the role of DBS in improving the PD signs in detail.

Network model

Our BG model has been created using Hodgkin-Huxley type neuron model that can generate neural behavior in detail (i.e. ion currents). Previous computational BG studies^{17,18,57–62,72,122} except^{24,123} which were created by Hodgkin-Huxley type neurons, were not reported β oscillations in PD state. In these studies, one of changes to set the model in PD state is strengthening subthalamo-pallidal synapses, based on the experimental results^{66–69}. The PD β oscillations also was reported by the model proposed in⁵⁶ which was created using LIF neuron model. This model moves to PD state only by increasing striato-pallidal spiking activity (without changing any synaptic strength). They show the PD β oscillations might arise from the network effects, without taking into account the details of neuronal dynamics. While,²³ used integrate-and-fire-or-burst neuron in their network model for subthalamo-pallidal circuit. Their model moves to PD state by changing in synaptic strength and input spikes in subthalamo-pallidal circuit and represents the PD β oscillations. In this model, the rebound burst activity of the STN and GPe neurons played an important role in the generation of the β oscillations. The PD β oscillations has also reported in the firing rate models^{20,124–126}. These models cannot be used for single neuron study in the network. By contrast, the role of the calcium bursting can not be demonstrated in these models. However, the modified BG model in present study was able to generate the β oscillations in PD state. Also, the other properties of BG model (i.e. firing rates and changes in firing rates when moving from the healthy to the PD state) were matched the experimental observations. these modifications were made by changing in some intrinsic and connectivity neuronal properties (see materials and methods).

Generation of beta oscillations

Computational and experimental studies have implicated the subthalamo-pallidal circuit is the potential source of the β oscillations in PD^{7,8,14–24,78}. Moreover, the induction of β oscillations from cortex into the BG has been claimed by^{6,25,26}. However, more detailed models for the generation of the β oscillations, the rebound burst activity of the STN neurons due to the inhibition of the GPe neurons in the BG has been considered^{15,16,114}. Somehow, mentioned hypothesize is confirmed when the motor symptom of patients with PD suppressed after receiving T-type calcium channel blocker such as Zonisamide¹²⁷. Also, the computational studies such as^{17,18,23,24,123} confirmed this hypothesis. While, the other studies demonstrate that the excessive inhibition on inhibitory population (such as striato-pallidal inhibition) and/or excessive excitation on excitatory population (such as cortico-subthalamic) result in β oscillations^{20,56,72,128,129}. Our findings confirm the role of post-inhibitory rebound bursts of STN neurons in generation of the β oscillations.

The role of the DBS is excitatory

The DBS improves PD-related motor symptoms^{27–32,110–112} and PD-related neuronal behavior in BG^{10–13,33,36,74,107–109}. In our network model, we quantified the PD-related signs by β oscillations, thalamus fidelity, and tremor frequency (see below and materials and methods). Here, we applied two scenarios of the DBS in our network model to see which satisfy our expectation about the amelioration of PD signs with DBS. Our findings suggest that the EDBS improves the PD neuronal behavior and the motor symptoms while the IDBS worsened them. Although the previous studies^{38–41,43–47,49,50,130} indicate that the role of the DBS on its target is inhibitory, in^{48,51,53–55} has been demonstrated the opposite role in accordance with our

findings. On the other hand, the computational study in⁵⁶ has claimed the excitation of excitatory and/or inhibition of inhibitory populations leads the network to oscillations (in this case is β oscillations). Therefore, the pathological β oscillations have been quenched using high frequency inhibitory spike trains on STN, that is not consistent with EDBS in our network. still, there is opposite evidence for this study's results: the initiation of movement accompanied with increasing cortical activity^{131–134} and the STN neurons are excited by cortex^{103–105} which quenches briefly the β oscillations in patients with PD^{8,135,136} and in a computational model proposed in²⁴. Moreover, the neuronal bursting activity were not investigated in that study, while, in¹³⁷ a computational model was proposed that the neuronal bursting activities have been investigated. This study showed the variety of STN burst range can affect on β oscillations. In detail, the low burst rate of STN neurons quenched the β oscillations while the high STN burst rate generated the β oscillations with a little shift in frequency. Indeed, the intrinsic behavior of STN neurons in non-pathological state are bursty and the higher burst rate of BG nuclei in PD condition are also shown (¹³⁸; and see also Figure 4 for GPi) which is consistent with the results that reported in¹³⁷. The consistency of this study and our network model is justified with considering that 1-the STN burst rate in our network model corresponds to high STN burst rate of the model in¹³⁷ and 2-with considering the STN burst rate when it exposes to EDBS corresponds to low STN burst rate in that study. Consequently, it seems to the therapeutic effects of DBS acts by excitation of the STN.

In addition, in^{37,139,140} it is shown that the DBS inhibits the STN neurons while it induces spike in their axons. In this hypothesis, despite the inhibition of STN neurons, their post-synaptic neurons (GPe and GPi) receive glutamate. So, inducing excitatory pulses in STN neurons in the network model matches with this hypothesis.

In addition, our results suggested that the post inhibitory rebound burst of the STN neurons is the main cause of the β oscillations. The EDBS by counteracting the pallidal inhibition in the network model eliminates the cause of the rebound bursting of the STN neurons. While, the IDBS helps the pallidal inhibition that resulted in more rebound burst activity of the STN neurons in the network model. Altogether, based on our network model results we conclude that the role of the DBS on its target is excitatory (and not inhibitory).

Tremor

The tremor in patients with PD occurs when they are at rest. The frequency of the resting tremor in PD state is reported 3 to 8 Hz^{64,88,118–121} and it correlates with thalamus neuronal activity^{115–117}. Previous computational studies^{17,18,57} demonstrated tremor activity by single neuron spiking activity in the tremor frequency in BG populations such as STN. While, in⁸⁸ the correlation between high frequency activity of STN and tremor has been shown. In a computational study the tremor activity in the PD state has been represented by synaptic input from the GPi to the thalamus neurons in tremor frequency⁶². However, the resting state was not simulated in mentioned computational studies. In our network model, we simulated resting state with interrupting sensorimotor command pulses to the thalamus neurons. Our results, show the extra thalamic spikes in tremor frequency in PD state (and not in healthy state). This result was consistent with previous experimental tremor-related thalamic activity observations. The main reason of extra spiking activity of thalamus in PD state is post inhibitory rebound spiking activity due to the strong inhibitory input from the GPi. The T-type calcium current of the thalamus neurons plays an important role in the rebound spiking activity.

However, the tremor-related thalamus activities were quenched when the network model exposed to the EDBS, while the tremor activities were increased when the network model exposed to the IDBS. This finding, again, confirmed that the role of the DBS is excitatory. Note that, we assigned the extra spiking activity of the thalamus neurons in the network model during the PD state to the involuntary tremor activity of the limbs, which was consistent with tremor frequency. By applying the EDBS on STN neurons, the tremor frequency was quenched, while in⁷⁶ has been reported by applying STN DBS the frequency of limb activity was increased and the amplitude of the limb movement was suppressed. In fact, the relation between the amplitude of the limb tremor and thalamus activity was not clarified in our network model.

Conclusion

We utilized a computational model of the BG to investigate the underlying mechanisms of the DBS. With the help of the model we concluded that first, the rebound burst activity related to T-type calcium current of the neurons has a key role for the generation of β oscillations. Second, we found that the role of the STN DBS is excitatory (and not inhibitory), because, the excitation of the STN neurons suppressed their rebound burst. Third, again, the rebound burst of thalamus neurons gave rise to the generation of resting tremor. Next, with exposing the STN neuron to high frequency excitation the thalamus tremor activity has quenched, while, with exposing the SNT neurons to high frequency inhibition it has been worsened. In summary, based on our model, we conclude that the role of the DBS is excitatory on its target.

Acknowledgements

We are grateful to Prof Dr Arvind Kumar and Dr Jyotika Bahuguna for their help and comments. This work was partially supported by the Shahid Rajae Teacher Training University.

References

1. Squire, L. R. *Fundamental Neuroscience*. ClinicalKey 2012 (Elsevier/Academic Press, 2013).
2. Galvan, A., Devergnas, A. & Wichmann, T. Alterations in neuronal activity in basal ganglia-thalamocortical circuits in the parkinsonian state. *Front. Neuroanat.* **9**, 5, DOI: [10.3389/fnana.2015.00005](https://doi.org/10.3389/fnana.2015.00005) (2015).
3. Miller, W. C. & DeLong, M. R. Parkinsonian symptomatology. An anatomical and physiological analysis. *Annals New York Acad. Sci.* **515**, 287–302, DOI: [10.1111/j.1749-6632.1988.tb32998.x](https://doi.org/10.1111/j.1749-6632.1988.tb32998.x) (1988).
4. Bernheimer, H., Birkmayer, W., Hornykiewicz, O., Jellinger, K. & Seitelberger, F. Brain dopamine and the syndromes of Parkinson and Huntington Clinical, morphological and neurochemical correlations. *J. Neurol. Sci.* **20**, 415–455, DOI: [10.1016/0022-510X\(73\)90175-5](https://doi.org/10.1016/0022-510X(73)90175-5) (1973).
5. Jankovic, J. Parkinson's disease: clinical features and diagnosis. *J. Neurol. Neurosurg. & Psychiatry* **79**, 368–376, DOI: [10.1136/jnnp.2007.131045](https://doi.org/10.1136/jnnp.2007.131045) (2008).
6. Hammond, C., Bergman, H. & Brown, P. Pathological synchronization in Parkinson's disease: networks, models and treatments. *Trends Neurosci.* **30**, 357–364, DOI: [10.1016/j.tins.2007.05.004](https://doi.org/10.1016/j.tins.2007.05.004) (2007).
7. Brown, P. *et al.* Dopamine dependency of oscillations between subthalamic nucleus and pallidum in Parkinson's disease. *J. Neurosci.* **21**, 1033–1038, DOI: [21/3/1033\[pil\]](https://doi.org/10.1523/JNEUROSCI.1913-18.2018) (2001).
8. Brown, P. & Williams, D. Basal ganglia local field potential activity: Character and functional significance in the human. *Clin. Neurophysiol.* **116**, 2510–2519, DOI: [10.1016/j.clinph.2005.05.009](https://doi.org/10.1016/j.clinph.2005.05.009) (2005).
9. Levy, R. *et al.* Dependence of subthalamic nucleus oscillations on movement and dopamine in Parkinson's disease. *Brain* **125**, 1196–1209, DOI: [10.1093/brain/awf128](https://doi.org/10.1093/brain/awf128) (2002).
10. Holt, A. B. *et al.* Phase-Dependent Suppression of Beta Oscillations in Parkinson's Disease Patients. *The J. Neurosci.* **39**, 1119–1134, DOI: [10.1523/JNEUROSCI.1913-18.2018](https://doi.org/10.1523/JNEUROSCI.1913-18.2018) (2019).
11. Rosin, B. *et al.* Closed-loop deep brain stimulation is superior in ameliorating parkinsonism. *Neuron* **72**, 370–384, DOI: [10.1016/j.neuron.2011.08.023](https://doi.org/10.1016/j.neuron.2011.08.023) (2011).
12. Little, S. *et al.* Adaptive deep brain stimulation in advanced Parkinson disease. *Annals Neurol.* **74**, 449–457, DOI: [10.1002/ana.23951](https://doi.org/10.1002/ana.23951) (2013).
13. Johnson, L. A. *et al.* Closed-Loop Deep Brain Stimulation Effects on Parkinsonian Motor Symptoms in a Non-Human Primate – Is Beta Enough? *Brain Stimul.* **9**, 892–896, DOI: [10.1016/j.brs.2016.06.051](https://doi.org/10.1016/j.brs.2016.06.051) (2016). [15334406](https://doi.org/10.1016/j.brs.2016.06.051).
14. Magill, P. J., Bolam, J. P. & Bevan, M. D. Dopamine regulates the impact of the cerebral cortex on the subthalamic nucleus-globus pallidus network. *Neuroscience* **106**, 313–330, DOI: [10.1016/S0306-4522\(01\)00281-0](https://doi.org/10.1016/S0306-4522(01)00281-0) (2001).
15. Plenz, D. & Kital, S. T. A basal ganglia pacemaker formed by the subthalamic nucleus and external globus pallidus. *Nature* **400**, 677–682, DOI: [10.1038/23281](https://doi.org/10.1038/23281) (1999).
16. Bevan, M. D., Magill, P. J., Terman, D., Bolam, J. P. & Wilson, C. J. Move to the rhythm: Oscillations in the subthalamic nucleus-external globus pallidus network. *Trends Neurosci.* **25**, 525–531, DOI: [10.1016/S0166-2236\(02\)02235-X](https://doi.org/10.1016/S0166-2236(02)02235-X) (2002).
17. Terman, D. & Rubin, J. E. Activity patterns in a model for the subthalamopallidal network of the basal ganglia. *The J. neuroscience : official journal Soc. for Neurosci.* **22**, 2963–2976, DOI: [20026266](https://doi.org/10.1523/JNEUROSCI.4199-08.2008) (2002).
18. Terman, D. & Rubin, J. E. High Frequency Stimulation of the Subthalamic Nucleus Eliminates Pathological Thalamic Rhythmicity in a Computational Model. *J. Comput. Neurosci.* 211–235 (2004).
19. Mallet, N. *et al.* Parkinsonian beta oscillations in the external globus pallidus and their relationship with subthalamic nucleus activity. *The J. neuroscience : official journal Soc. for Neurosci.* **28**, 14245–14258, DOI: [10.1523/JNEUROSCI.4199-08.2008](https://doi.org/10.1523/JNEUROSCI.4199-08.2008) (2008).
20. Pavlides, A., Hogan, S. J. & Bogacz, R. Computational Models Describing Possible Mechanisms for Generation of Excessive Beta Oscillations in Parkinson's Disease. *PLoS Comput. Biol.* **11**, 1–29, DOI: [10.1371/journal.pcbi.1004609](https://doi.org/10.1371/journal.pcbi.1004609) (2015).
21. Tachibana, Y., Iwamuro, H., Kita, H., Takada, M. & Nambu, A. Subthalamo-pallidal interactions underlying parkinsonian neuronal oscillations in the primate basal ganglia. *Eur. J. Neurosci.* **34**, 1470–1484, DOI: [10.1111/j.1460-9568.2011.07865.x](https://doi.org/10.1111/j.1460-9568.2011.07865.x) (2011).
22. Mirzaei, A. *et al.* Sensorimotor Processing in the Basal Ganglia Leads to Transient Beta Oscillations during Behavior. *The J. Neurosci.* **37**, 11220–11232, DOI: [10.1523/JNEUROSCI.1289-17.2017](https://doi.org/10.1523/JNEUROSCI.1289-17.2017) (2017).

23. Wei, W., Rubin, J. E. & Wang, X.-J. Role of the Indirect Pathway of the Basal Ganglia in Perceptual Decision Making. *The J. Neurosci.* **35**, 4052–4064, DOI: [10.1523/JNEUROSCI.3611-14.2015](https://doi.org/10.1523/JNEUROSCI.3611-14.2015) (2015).
24. Shouno, O., Tachibana, Y., Nambu, A. & Doya, K. Computational Model of Recurrent Subthalamo-Pallidal Circuit for Generation of Parkinsonian Oscillations. *Front. Neuroanat.* **11**, 1–15, DOI: [10.3389/fnana.2017.00021](https://doi.org/10.3389/fnana.2017.00021) (2017).
25. Goldberg, J. A. Spike Synchronization in the Cortex-Basal Ganglia Networks of Parkinsonian Primates Reflects Global Dynamics of the Local Field Potentials. *J. Neurosci.* **24**, 6003–6010, DOI: [10.1523/JNEUROSCI.4848-03.2004](https://doi.org/10.1523/JNEUROSCI.4848-03.2004) (2004).
26. Brown, P. Abnormal oscillatory synchronisation in the motor system leads to impaired movement. *Curr. Opin. Neurobiol.* **17**, 656–664, DOI: [10.1016/j.conb.2007.12.001](https://doi.org/10.1016/j.conb.2007.12.001) (2007).
27. Abeso, J. A. & Olanow, C. W. Deep-Brain Stimulation of the Subthalamic Nucleus or the Pars Interna of the Globus Pallidus in Parkinson's Disease. *New Engl. J. Medicine* **345**, 956–963, DOI: [10.1056/NEJMoa000827](https://doi.org/10.1056/NEJMoa000827) (2001).
28. Groiss, S. J., Wojtecki, L., Südmeyer, M. & Schnitzler, A. Deep brain stimulation in Parkinson's disease. *Ther. advances neurological disorders* **2**, 20–8, DOI: [10.1177/1756285609339382](https://doi.org/10.1177/1756285609339382) (2009).
29. Olanow, C. W., Brin, M. F. & Obeso, J. A. The role of deep brain stimulation as a surgical treatment for Parkinson's disease. *Neurology* **55**, S60–6 (2000).
30. Pollak, P. *et al.* Treatment results: Parkinson's disease. *Mov. Disord.* **17**, S75–S83, DOI: [10.1002/mds.10146](https://doi.org/10.1002/mds.10146) (2002).
31. Wichmann, T. & DeLong, M. R. Deep Brain Stimulation for Neurologic and Neuropsychiatric Disorders. *Neuron* **52**, 197–204, DOI: [10.1016/j.neuron.2006.09.022](https://doi.org/10.1016/j.neuron.2006.09.022) (2006).
32. Weaver, F. M. *et al.* Bilateral Deep Brain Stimulation vs Best Medical Therapy for Patients With Advanced Parkinson Disease. *The J. Am. Med. Assoc.* **301**, 301(1): 63–73, DOI: [10.1001/jama.2008.929](https://doi.org/10.1001/jama.2008.929) (2009).
33. Giannicola, G. *et al.* The effects of levodopa and ongoing deep brain stimulation on subthalamic beta oscillations in Parkinson's disease. *Exp. Neurol.* **226**, 120–127, DOI: [10.1016/j.expneurol.2010.08.011](https://doi.org/10.1016/j.expneurol.2010.08.011) (2010).
34. Nambu, A. Seven problems on the basal ganglia. *Curr. Opin. Neurobiol.* **18**, 595–604, DOI: [10.1016/j.conb.2008.11.001](https://doi.org/10.1016/j.conb.2008.11.001) (2008).
35. Benabid, A. L. Deep brain stimulation for Parkinson's disease. *Curr. Opin. Neurobiol.* **13**, 696–706, DOI: [10.1016/j.conb.2003.11.001](https://doi.org/10.1016/j.conb.2003.11.001) (2003).
36. Kringelbach, M. L., Jenkinson, N., Owen, S. L. F. & Aziz, T. Z. Translational principles of deep brain stimulation. *Nat. reviews. Neurosci.* **8**, 623–635, DOI: [10.1038/nrn2196](https://doi.org/10.1038/nrn2196) (2007).
37. McIntyre, C. C., Savasta, M., Walter, B. L. & Vitek, J. L. How does deep brain stimulation work? Present understanding and future questions. *J. clinical neurophysiology : official publication Am. Electroencephalogr. Soc.* **21**, 40–50, DOI: [10.1097/00004691-200401000-00006](https://doi.org/10.1097/00004691-200401000-00006) (2004). NIHMS150003.
38. Dostrovsky, J. O. *et al.* Microstimulation-induced inhibition of neuronal firing in human globus pallidus. *J. neurophysiology* **84**, 570–574 (2000).
39. Lafreniere-Roula, M. *et al.* High-frequency microstimulation in human globus pallidus and substantia nigra. *Exp. Brain Res.* **205**, 251–261, DOI: [10.1007/s00221-010-2362-8](https://doi.org/10.1007/s00221-010-2362-8) (2010).
40. Boraud, T., Bezard, E., Bioulac, B. & Gross, C. High frequency stimulation of the internal Globus Pallidus (GPi) simultaneously improves parkinsonian symptoms and reduces the firing frequency of GPi neurons in the MPTP-treated monkey. *Neurosci. Lett.* **215**, 17–20, DOI: [10.1016/S0304-3940\(96\)12943-8](https://doi.org/10.1016/S0304-3940(96)12943-8) (1996).
41. Chiken, S. & Nambu, A. Mechanism of Deep Brain Stimulation. *The Neurosci.* **22**, 313–322, DOI: [10.1177/1073858415581986](https://doi.org/10.1177/1073858415581986) (2016).
42. Beurrier, C., Bioulac, B., Audin, J. & Hammond, C. High-frequency stimulation produces a transient blockade of voltage-gated currents in subthalamic neurons. *J Neurophysiol* **85**, 1351–1356 (2001).
43. Do, M. T. H. & Bean, B. P. Subthreshold sodium currents and pacemaking of subthalamic neurons: Modulation by slow inactivation. *Neuron* **39**, 109–120, DOI: [10.1016/S0896-6273\(03\)00360-X](https://doi.org/10.1016/S0896-6273(03)00360-X) (2003).
44. Shin, D. S. *et al.* High frequency stimulation or elevated K⁺ depresses neuronal activity in the rat entopeduncular nucleus. *Neuroscience* **149**, 68–86, DOI: [10.1016/j.neuroscience.2007.06.055](https://doi.org/10.1016/j.neuroscience.2007.06.055) (2007).
45. Chiken, S. & Nambu, A. High-Frequency Pallidal Stimulation Disrupts Information Flow through the Pallidum by GABAergic Inhibition. *J. Neurosci.* **33**, 2268–2280, DOI: [10.1523/JNEUROSCI.4144-11.2013](https://doi.org/10.1523/JNEUROSCI.4144-11.2013) (2013).

46. Deniau, J. M., Degos, B., Bosch, C. & Maurice, N. Deep brain stimulation mechanisms: Beyond the concept of local functional inhibition. *Eur. J. Neurosci.* **32**, 1080–1091, DOI: [10.1111/j.1460-9568.2010.07413.x](https://doi.org/10.1111/j.1460-9568.2010.07413.x) (2010).
47. Dostrovsky, J. O. & Lozano, A. M. Mechanisms of deep brain stimulation. *Mov. Disord.* **17**, S63–S68, DOI: [10.1002/mds.10143](https://doi.org/10.1002/mds.10143) (2002).
48. Johnson, M. D. & McIntyre, C. C. Quantifying the Neural Elements Activated and Inhibited by Globus Pallidus Deep Brain Stimulation. *J. Neurophysiol.* **100**, 2549–2563, DOI: [10.1152/jn.90372.2008](https://doi.org/10.1152/jn.90372.2008) (2008).
49. Liu, Y., Postupna, N., Falkenberg, J. & Anderson, M. E. High frequency deep brain stimulation: What are the therapeutic mechanisms? *Neurosci. Biobehav. Rev.* **32**, 343–351, DOI: [10.1016/j.neubiorev.2006.10.007](https://doi.org/10.1016/j.neubiorev.2006.10.007) (2008).
50. Meissner, W. *et al.* Subthalamic high frequency stimulation resets subthalamic firing and reduces abnormal oscillations. *Brain* **128**, 2372–2382, DOI: [10.1093/brain/awh616](https://doi.org/10.1093/brain/awh616) (2005).
51. McCairn, K. W. & Turner, R. S. Deep Brain Stimulation of the Globus Pallidus Internus in the Parkinsonian Primate: Local Entrainment and Suppression of Low-Frequency Oscillations. *J. Neurophysiol.* **101**, 1941–1960, DOI: [10.1152/jn.91092.2008](https://doi.org/10.1152/jn.91092.2008) (2009).
52. Anderson, M. E. Effects of High-Frequency Stimulation in the Internal Globus Pallidus on the Activity of Thalamic Neurons in the Awake Monkey. *J. Neurophysiol.* **89**, 1150–1160, DOI: [10.1152/jn.00475.2002](https://doi.org/10.1152/jn.00475.2002) (2003).
53. Galati, S. *et al.* Biochemical and electrophysiological changes of substantia nigra pars reticulata driven by subthalamic stimulation in patients with Parkinson's disease. *Eur. J. Neurosci.* **23**, 2923–2928, DOI: [10.1111/j.1460-9568.2006.04816.x](https://doi.org/10.1111/j.1460-9568.2006.04816.x) (2006).
54. Hashimoto, T., Elder, C. M., Okun, M. S., Patrick, S. K. & Vitek, J. L. Stimulation of the subthalamic nucleus changes the firing pattern of pallidal neurons. *The J. neuroscience : official journal Soc. for Neurosci.* **23**, 1916–1923, DOI: [10.1523/JNEUROSCI.2355-03.2003](https://doi.org/10.1523/JNEUROSCI.2355-03.2003) (2003).
55. Reese, R. *et al.* Subthalamic deep brain stimulation increases pallidal firing rate and regularity. *Exp. Neurol.* **229**, 517–521, DOI: [10.1016/j.expneurol.2011.01.020](https://doi.org/10.1016/j.expneurol.2011.01.020) (2011).
56. Kumar, A., Cardanobile, S., Rotter, S. & Aertsen, A. The role of inhibition in generating and controlling Parkinson's disease oscillations in the Basal Ganglia. *Front. systems neuroscience* **5**, 86, DOI: [10.3389/fnsys.2011.00086](https://doi.org/10.3389/fnsys.2011.00086) (2011).
57. Feng, X.-j., Greenwald, B., Rabitz, H., Shea-Brown, E. & Kosut, R. Toward closed-loop optimization of deep brain stimulation for Parkinson's disease: concepts and lessons from a computational model. *J. Neural Eng.* **4**, L14 (2007).
58. Summerson, S. R., Aazhang, B. & Kemere, C. Investigating irregularly patterned deep brain stimulation signal design using biophysical models. *Front. Comput. Neurosci.* **9**, 1–10, DOI: [10.3389/fncom.2015.00078](https://doi.org/10.3389/fncom.2015.00078) (2015).
59. Agarwal, R. & Sarma, S. V. The effects of DBS patterns on basal ganglia activity and thalamic relay: A computational study. *J. Comput. Neurosci.* **33**, 151–167, DOI: [10.1007/s10827-011-0379-z](https://doi.org/10.1007/s10827-011-0379-z) (2012).
60. Schiff, S. J. Towards model-based control of Parkinson's disease. *Philos. Transactions Royal Soc. A: Math. Phys. Eng. Sci.* **368**, 2269–2308, DOI: [10.1098/rsta.2010.0050](https://doi.org/10.1098/rsta.2010.0050) (2010).
61. Hahn, P. J. & McIntyre, C. C. Modeling shifts in the rate and pattern of subthalamopallidal network activity during deep brain stimulation. **28**, 425–441, DOI: [10.1007/s10827-010-0225-8](https://doi.org/10.1007/s10827-010-0225-8). Modeling (2011).
62. Liu, C. *et al.* Closed-Loop Control of Tremor-Predominant Parkinsonian State Based on Parameter Estimation. *IEEE Transactions on Neural Syst. Rehabil. Eng.* **24**, 1109–1121, DOI: [10.1109/TNSRE.2016.2535358](https://doi.org/10.1109/TNSRE.2016.2535358) (2016).
63. Hua, S. E., Lenz, F. a., Zirh, T. a., Reich, S. G. & Dougherty, P. M. Thalamic neuronal activity correlated with essential tremor. *J. neurology, neurosurgery, psychiatry* **64**, 273–276, DOI: [10.1136/jnnp.64.2.273](https://doi.org/10.1136/jnnp.64.2.273) (1998).
64. Lenz, F. A. *et al.* Single unit analysis of the human ventral thalamic nuclear group: correlation of thalamic "tremor cells" with the 3-6 Hz component of parkinsonian tremor. *The J. neuroscience : official journal Soc. for Neurosci.* **8**, 754–764 (1988).
65. Sato, F., Parent, M., Levesque, M. & Parent, A. Axonal branching pattern of neurons of the subthalamic nucleus in primates. *J. Comp. Neurol.* **424**, 142–152, DOI: [10.1002/1096-9861\(20000814\)424:1<142::AID-CNE10>3.0.CO;2-8](https://doi.org/10.1002/1096-9861(20000814)424:1<142::AID-CNE10>3.0.CO;2-8) (2000).
66. Shen, K.-Z. & Johnson, S. W. Dopamine depletion alters responses to glutamate and GABA in the rat subthalamic nucleus. *Neuroreport* **16**, 171–4, DOI: [10.1097/00001756-200502080-00021](https://doi.org/10.1097/00001756-200502080-00021) (2005).
67. Ogura, M. & Kita, H. Dynorphin exerts both postsynaptic and presynaptic effects in the Globus pallidus of the rat. *J. neurophysiology* **83**, 3366–3376 (2000).

68. Cooper, A. J. & Stanford, I. M. Dopamine D2 receptor mediated presynaptic inhibition of striatopallidal GABAA IPSCs in vitro. *Neuropharmacology* **41**, 62–71, DOI: [10.1016/S0028-3908\(01\)00038-7](https://doi.org/10.1016/S0028-3908(01)00038-7) (2001).
69. Baufreton, J. & Bevan, M. D. D2-like dopamine receptor-mediated modulation of activity-dependent plasticity at GABAergic synapses in the subthalamic nucleus. *J Physiol* **586**, 2121–2142, DOI: [10.1113/jphysiol.2008.151118](https://doi.org/10.1113/jphysiol.2008.151118) (2008).
70. Doudet, D. J., Gross, C., Arluison, M. & Bioulac, B. Modifications of precentral cortex discharge and EMG activity in monkeys with MPTP-induced lesions of DA nigral neurons. *Exp. Brain Res.* **80**, 177–188, DOI: [10.1007/BF00228859](https://doi.org/10.1007/BF00228859) (1990).
71. Park, C., Worth, R. M. & Rubchinsky, L. L. Neural dynamics in Parkinsonian brain: The boundary between synchronized and nonsynchronized dynamics. *Phys. Rev. E - Stat. Nonlinear, Soft Matter Phys.* **83**, 1–4, DOI: [10.1103/PhysRevE.83.042901](https://doi.org/10.1103/PhysRevE.83.042901) (2011).
72. Ahn, S., Zuber, S. E., Worth, R. M. & Rubchinsky, L. L. Synchronized Beta-Band Oscillations in a Model of the Globus Pallidus-Subthalamic Nucleus Network under External Input. *Front. Comput. Neurosci.* **10**, 1–12, DOI: [10.3389/fncom.2016.00134](https://doi.org/10.3389/fncom.2016.00134) (2016).
73. Agnesi, F., Muralidharan, A., Baker, K. B., Vitek, J. L. & Johnson, M. D. Fidelity of frequency and phase entrainment of circuit-level spike activity during DBS. *J. Neurophysiol.* **114**, 825–834, DOI: [10.1152/jn.00259.2015](https://doi.org/10.1152/jn.00259.2015) (2015).
74. Wingeier, B. *et al.* Intra-operative STN DBS attenuates the prominent beta rhythm in the STN in Parkinson's disease. *Exp. Neurol.* **197**, 244–251, DOI: [10.1016/j.expneurol.2005.09.016](https://doi.org/10.1016/j.expneurol.2005.09.016) (2006).
75. Wagenbreth, C. *et al.* Deep brain stimulation of the subthalamic nucleus modulates reward processing and action selection in Parkinson patients. *J. Neurol.* **262**, 1541–1547, DOI: [10.1007/s00415-015-7749-9](https://doi.org/10.1007/s00415-015-7749-9) (2015).
76. Sturman, M. M., Vaillancourt, D. E., Metman, L. V., Bakay, R. A. E. & Corcos, D. M. Effects of subthalamic nucleus stimulation and medication on resting and postural tremor in Parkinson's disease. *Brain* **127**, 2131–2143, DOI: [10.1093/brain/awh237](https://doi.org/10.1093/brain/awh237) (2004).
77. James Anderson, C. *et al.* Subthalamic deep brain stimulation reduces pathological information transmission to the thalamus in a rat model of parkinsonism. *frontiers Neural Circuits* **9**, 1–11, DOI: [10.3389/fncir.2015.00031](https://doi.org/10.3389/fncir.2015.00031) (2015).
78. So, R. Q., Kent, A. R. & Grill, W. M. Relative contributions of local cell and passing fiber activation and silencing to changes in thalamic fidelity during deep brain stimulation and lesioning: A computational modeling study. *J. Comput. Neurosci.* **32**, 499–519, DOI: [10.1007/s10827-011-0366-4](https://doi.org/10.1007/s10827-011-0366-4) (2012).
79. Kumar, A., Schrader, S., Aertsen, A. & Rotter, S. The High-Conductance State of Cortical Networks. *Neural Comput.* **20**, 1–43, DOI: [10.1162/neco.2008.20.1.1](https://doi.org/10.1162/neco.2008.20.1.1) (2008).
80. Jones, E., Oliphant, T., Peterson, P. *et al.* SciPy: Open source scientific tools for Python (2001). [Online; accessed 22/10/2017].
81. Brown, P. *et al.* Oscillatory Local Field Potentials Recorded from the Subthalamic Nucleus of the Alert Rat. *Exp. Neurol.* **177**, 581–585, DOI: [10.1006/exnr.2002.7984](https://doi.org/10.1006/exnr.2002.7984) (2002).
82. Raz, A., Vaadia, E. & Bergman, H. Firing patterns and correlations of spontaneous discharge of pallidal neurons in the normal and the tremulous 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine vervet model of parkinsonism. *The J. neuroscience : official journal Soc. for Neurosci.* **20**, 8559–71, DOI: [10.22/8559\[pil\]](https://doi.org/10.22/8559[pil]) (2000).
83. Pan, H. S. & Walters, J. R. Unilateral lesion of the nigrostriatal pathway decreases the firing rate and alters the firing pattern of globus pallidus neurons in the rat. *Synapse* **2**, 650–656, DOI: [10.1002/syn.890020612](https://doi.org/10.1002/syn.890020612) (1988).
84. Bergman, H., Wichmann, T., Karmon, B. & DeLong, M. R. The primate subthalamic nucleus. II. Neuronal activity in the MPTP model of parkinsonism. *J. neurophysiology* **72**, 507–20 (1994).
85. Hassani, O. K., Mouroux, M. & Féger, J. Increased subthalamic neuronal activity after nigral dopaminergic lesion independent of disinhibition via the globus pallidus. *Neuroscience* **72**, 105–115, DOI: [10.1016/0306-4522\(95\)00535-8](https://doi.org/10.1016/0306-4522(95)00535-8) (1996).
86. Bergman, H. *et al.* Physiological aspects of information processing in the basal ganglia of normal and parkinsonian monkeys. *Trends Neurosci.* **21**, 32–37 (1998).
87. Beurrier, C., Congar, P., Bioulac, B. & Hammond, C. Subthalamic nucleus neurons switch from single-spike activity to burst-firing mode. *The J. neuroscience : official journal Soc. for Neurosci.* **19**, 599–609 (1999).
88. Levy, R., Hutchison, W. D., Lozano, A. M. & Dostrovsky, J. O. High-frequency synchronization of neuronal activity in the subthalamic nucleus of parkinsonian patients with limb tremor. *J. Neurosci.* **20**, 7766–7775, DOI: [10.1523/JNEUROSCI.2070-00.2000](https://doi.org/10.1523/JNEUROSCI.2070-00.2000) (2000).

89. Magnin, M., Morel, A. & Jeanmond, D. Single-unit analysis of the pallidum, thalamus and subthalamic nucleus in {Parkinson} patients. *Neuroscience* **96**, 549–564 (2000).
90. Benazzouz, A. *et al.* Intraoperative microrecordings of the subthalamic nucleus in Parkinson's disease. *Mov. Disord.* **17**, 145–149, DOI: [10.1002/mds.10156](https://doi.org/10.1002/mds.10156) (2002).
91. Weinberger, M. *et al.* Beta oscillatory activity in the subthalamic nucleus and its relation to dopaminergic response in Parkinson's disease. *J. neurophysiology* **96**, 3248–56, DOI: [10.1152/jn.00697.2006](https://doi.org/10.1152/jn.00697.2006) (2006).
92. Wilson, C. L. *et al.* Subthalamic nucleus neurones in slices from 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-lesioned mice show irregular, dopamine-reversible firing pattern changes, but without synchronous activity. *Neuroscience* **143**, 565–572, DOI: [10.1016/j.neuroscience.2006.07.051](https://doi.org/10.1016/j.neuroscience.2006.07.051) (2006).
93. Wichmann, T., Bergman, H. & DeLong, M. R. The primate subthalamic nucleus. III. Changes in motor behavior and neuronal activity in the internal pallidum induced by subthalamic inactivation in the MPTP model of parkinsonism. *J. Neurophysiol.* **72**, 521–530 (1994).
94. Park, C. *et al.* Fine Temporal Structure of Beta Oscillations Synchronization in Subthalamic Nucleus in Parkinson ' s Disease Fine Temporal Structure of Beta Oscillations Synchronization in Subthalamic Nucleus in Parkinson ' s Disease. 2707–2716, DOI: [10.1152/jn.00724.2009](https://doi.org/10.1152/jn.00724.2009) (2010).
95. Beck, M. H. *et al.* Short- and long-term dopamine depletion causes enhanced beta oscillations in the cortico-basal ganglia loop of parkinsonian rats. *Exp. Neurol.* **286**, 124–136, DOI: [10.1016/j.expneurol.2016.10.005](https://doi.org/10.1016/j.expneurol.2016.10.005) (2016).
96. Kühn, A. A. *et al.* The relationship between local field potential and neuronal discharge in the subthalamic nucleus of patients with Parkinson's disease. *Exp. Neurol.* **194**, 212–220, DOI: [10.1016/j.expneurol.2005.02.010](https://doi.org/10.1016/j.expneurol.2005.02.010) (2005).
97. Cassidy, M. *et al.* Movement-related changes in synchronization in the human basal ganglia. *Brain* **125**, 1235–1246 (2002).
98. Sharott, A. *et al.* Spatio-temporal dynamics of cortical drive to human subthalamic nucleus neurons in Parkinson's disease. *Neurobiol. Dis.* **112**, 49–62, DOI: [10.1016/j.nbd.2018.01.001](https://doi.org/10.1016/j.nbd.2018.01.001) (2018).
99. Miller, W. C. & DeLong, M. R. *Altered Tonic Activity of Neurons in the Globus Pallidus and Subthalamic Nucleus in the Primate MPTP Model of Parkinsonism* (Springer US, Boston, MA, 1987).
100. Sterio, D. *et al.* Neurophysiological Properties of Palhdal Neurons in Palanson ' s Disease. 586–591, DOI: [10.1002/ana.410350512](https://doi.org/10.1002/ana.410350512) (1994).
101. Hutchison, W. D. *et al.* Differential neuronal activity in segments of globus pallidus in Parkinson's disease patients. *Neuroreport* **5**, 1533–7, DOI: [7948856](https://doi.org/10.1002/ana.410350512) (1994).
102. Wichmann, T. *et al.* Comparison of MPTP-induced changes in spontaneous neuronal discharge in the internal pallidal segment and in the substantia nigra pars reticulata in primates. *Exp. Brain Res.* **125**, 397–409, DOI: [10.1007/s002210050696](https://doi.org/10.1007/s002210050696) (1999).
103. Nambu, A., Tokuno, H. & Takada, M. Functional significance of the cortico-subthalamo-pallidal 'hyperdirect' pathway. *Neurosci. Res.* **43**, 111–117, DOI: [10.1016/S0168-0102\(02\)00027-5](https://doi.org/10.1016/S0168-0102(02)00027-5) (2002).
104. Parent, M. & Parent, A. Single-axon tracing study of corticostriatal projections arising from primary motor cortex in primates. *J. Comp. Neurol.* **496**, 202–213, DOI: [10.1002/cne.20925](https://doi.org/10.1002/cne.20925) (2006).
105. Kita, T. & Kita, H. The Subthalamic Nucleus Is One of Multiple Innervation Sites for Long-Range Corticofugal Axons: A Single-Axon Tracing Study in the Rat. *J. Neurosci.* **32**, 5990–5999, DOI: [10.1523/JNEUROSCI.5717-11.2012](https://doi.org/10.1523/JNEUROSCI.5717-11.2012) (2012). [NIHMS150003](https://doi.org/10.1523/JNEUROSCI.5717-11.2012).
106. Guo, Y., Rubin, J. E., McIntyre, C. C., Vitek, J. L. & Terman, D. Thalamocortical Relay Fidelity Varies Across Subthalamic Nucleus Deep Brain Stimulation Protocols in a Data-Driven Computational Model. *J. Neurophysiol.* **99**, 1477–1492, DOI: [10.1152/jn.01080.2007](https://doi.org/10.1152/jn.01080.2007) (2008).
107. Engel, A. K. & Fries, P. Beta-band oscillations-signalling the status quo? *Curr. Opin. Neurobiol.* **20**, 156–165, DOI: [10.1016/j.conb.2010.02.015](https://doi.org/10.1016/j.conb.2010.02.015) (2010).
108. Tinkhauser, G. *et al.* The modulatory effect of adaptive deep brain stimulation on beta bursts in Parkinson's disease. *Brain* **140**, 1053–1067, DOI: [10.1093/brain/awx010](https://doi.org/10.1093/brain/awx010) (2017). [1611.06654](https://doi.org/10.1093/brain/awx010).
109. de Hemptinne, C. *et al.* Therapeutic deep brain stimulation reduces cortical phase-amplitude coupling in Parkinson's disease. *Nat. Neurosci.* **18**, 779–786, DOI: [10.1038/nn.3997](https://doi.org/10.1038/nn.3997) (2015). [arXiv:1011.1669v3](https://arxiv.org/abs/1011.1669v3).

110. Kuhn, A. A. *et al.* High-Frequency Stimulation of the Subthalamic Nucleus Suppresses Oscillatory Activity in Patients with Parkinson's Disease in Parallel with Improvement in Motor Performance. *J. Neurosci.* **28**, 6165–6173, DOI: [10.1523/JNEUROSCI.0282-08.2008](https://doi.org/10.1523/JNEUROSCI.0282-08.2008) (2008).
111. Ray, N. J. *et al.* Local field potential beta activity in the subthalamic nucleus of patients with Parkinson's disease is associated with improvements in bradykinesia after dopamine and deep brain stimulation. *Exp. Neurol.* **213**, 108–113, DOI: [10.1016/j.expneurol.2008.05.008](https://doi.org/10.1016/j.expneurol.2008.05.008) (2008).
112. Zaidel, A., Spivak, A., Grieb, B., Bergman, H. & Israel, Z. Subthalamic span of β oscillations predicts deep brain stimulation efficacy for patients with Parkinson's disease. *Brain* **133**, 2007–2021, DOI: [10.1093/brain/awq144](https://doi.org/10.1093/brain/awq144) (2010).
113. Bevan, M. D. & Wilson, C. J. Mechanisms underlying spontaneous oscillation and rhythmic firing in rat subthalamic neurons. *The J. neuroscience : official journal Soc. for Neurosci.* **19**, 7617–7628, DOI: [10.3389/fnsys.2011.00083](https://doi.org/10.3389/fnsys.2011.00083) (1999).
114. Bevan, M. D., Wilson, C. J., Bolam, J. P. & Magill, P. J. Equilibrium potential of GABA(A) current and implications for rebound burst firing in rat subthalamic neurons in vitro. *J. neurophysiology* **83**, 3169–3172 (2000).
115. Jasper, H. H. Thalamic unit involved in somatic sensation and voluntary and involuntary movements in man. *The Thalamus* 365–390 (1966).
116. OHYE, C. Rhythmic discharges related to tremor in humans and monkeys. in. *Abnorm. neuronal discharges* 37–48 (1978).
117. Lamarre, Y. & Joffroy, A. J. Experimental tremor in monkey-activity of thalamic and precentral cortical-neurons in the absence of peripheral feedback. *Can. J. Neurol. Sci.* **6**, 93 (1979).
118. Marsden, J. F. Coherence between cerebellar thalamus, cortex and muscle in man: Cerebellar thalamus interactions. *Brain* **123**, 1459–1470, DOI: [10.1093/brain/123.7.1459](https://doi.org/10.1093/brain/123.7.1459) (2000).
119. Zirh, T. A., Lenz, F. A., Reich, S. G. & Dougherty, P. M. Patterns of bursting occurring in thalamic cells during parkinsonian tremor. *Neuroscience* **83**, 107–121, DOI: [10.1016/S0306-4522\(97\)00295-9](https://doi.org/10.1016/S0306-4522(97)00295-9) (1998).
120. Liu, X. *et al.* The oscillatory activity in the Parkinsonian subthalamic nucleus investigated using the macro-electrodes for deep brain stimulation. *Clin. Neurophysiol.* **113**, 1667–1672, DOI: [10.1016/S1388-2457\(02\)00256-0](https://doi.org/10.1016/S1388-2457(02)00256-0) (2002).
121. Hua, S. *et al.* The role of the thalamus and basal ganglia in parkinsonian tremor. *Mov. disorders : official journal Mov. Disord. Soc.* **13 Suppl 3**, 40–42, DOI: [10.1002/mds.870131307](https://doi.org/10.1002/mds.870131307) (1998).
122. Best, J., Park, C., Terman, D. & Wilson, C. Transitions between irregular and rhythmic firing patterns in excitatory-inhibitory neuronal networks. *J. Comput. Neurosci.* **23**, 217–235, DOI: [10.1007/s10827-007-0029-7](https://doi.org/10.1007/s10827-007-0029-7) (2007).
123. Kumaravelu, K., Brocker, D. T. & Grill, W. M. A biophysical model of the cortex-basal ganglia-thalamus network in the 6-OHDA lesioned rat model of Parkinson's disease. *J. Comput. Neurosci.* **40**, 207–229, DOI: [10.1007/s10827-016-0593-9](https://doi.org/10.1007/s10827-016-0593-9) (2016).
124. Holgado, A. J. N., Terry, J. R. & Bogacz, R. Conditions for the Generation of Beta Oscillations in the Subthalamic Nucleus-Globus Pallidus Network. *J. Neurosci.* **30**, 12340–12352, DOI: [10.1523/JNEUROSCI.0817-10.2010](https://doi.org/10.1523/JNEUROSCI.0817-10.2010) (2010).
125. Pavlides, A., John Hogan, S. & Bogacz, R. Improved conditions for the generation of beta oscillations in the subthalamic nucleus-globus pallidus network. *Eur. J. Neurosci.* **36**, 2229–2239, DOI: [10.1111/j.1460-9568.2012.08105.x](https://doi.org/10.1111/j.1460-9568.2012.08105.x) (2012).
126. Pasillas-Lépine, W. Delay-induced oscillations in Wilson and Cowan's model: An analysis of the subthalamo-pallidal feedback loop in healthy and parkinsonian subjects. *Biol. Cybern.* **107**, 289–308, DOI: [10.1007/s00422-013-0549-3](https://doi.org/10.1007/s00422-013-0549-3) (2013).
127. Bermejo, P. E. & Anciones, B. A review of the use of zonisamide in Parkinson's disease. *Ther. advances neurological disorders* **2**, 313–7, DOI: [10.1177/1756285609338501](https://doi.org/10.1177/1756285609338501) (2009).
128. Gillies, Willshaw, D. & Li, Z. Subthalamic-pallidal interactions are critical in determining normal and abnormal functioning of the basal ganglia. *Proc. Royal Soc. Lond. B: Biol. Sci.* **269**, 545–551, DOI: [10.1098/rspb.2001.1817](https://doi.org/10.1098/rspb.2001.1817) (2002).
129. Nevado-Holgado, A. J., Mallet, N., Magill, P. J. & Bogacz, R. Effective connectivity of the subthalamic nucleus-globus pallidus network during Parkinsonian oscillations. *The J. physiology* **592**, 1429–1455 (2014).
130. Filali, M., Hutchison, W. D., Palter, V. N., Lozano, A. M. & Dostrovsky, J. O. Stimulation-induced inhibition of neuronal firing in human subthalamic nucleus. *Exp. Brain Res.* **156**, 274–281, DOI: [10.1007/s00221-003-1784-y](https://doi.org/10.1007/s00221-003-1784-y) (2004).
131. Turner, R. S. & DeLong, M. R. Corticostriatal activity in primary motor cortex of the macaque. *The J. neuroscience : official journal Soc. for Neurosci.* **20**, 7096–108, DOI: [20/18/7096\[pil\]](https://doi.org/10.1523/JNEUROSCI.2018-00.2000) (2000).

132. Pasquereau, B. & Turner, R. S. Primary motor cortex of the parkinsonian monkey: Differential effects on the spontaneous activity of pyramidal tract-type neurons. *Cereb. Cortex* **21**, 1362–1378, DOI: [10.1093/cercor/bhq217](https://doi.org/10.1093/cercor/bhq217) (2011).
133. Pfurtscheller, G. & Lopes Da Silva, F. H. Event-related EEG/MEG synchronization and desynchronization: Basic principles. *Clin. Neurophysiol.* **110**, 1842–1857, DOI: [10.1016/S1388-2457\(99\)00141-8](https://doi.org/10.1016/S1388-2457(99)00141-8) (1999). [S1388-2457\(99\)00141-8](https://doi.org/10.1016/S1388-2457(99)00141-8).
134. Toma, K. Movement Rate Effect on Activation and Functional Coupling of Motor Cortical Areas. *J. Neurophysiol.* **88**, 3377–3385, DOI: [10.1152/jn.00281.2002](https://doi.org/10.1152/jn.00281.2002) (2002).
135. Amirnovin, R. Visually Guided Movements Suppress Subthalamic Oscillations in Parkinson's Disease Patients. *J. Neurosci.* **24**, 11302–11306, DOI: [10.1523/JNEUROSCI.3242-04.2004](https://doi.org/10.1523/JNEUROSCI.3242-04.2004) (2004).
136. Little, S. & Brown, P. The functional role of beta oscillations in Parkinson's disease. *Park. Relat. Disord.* **20**, S44–S48, DOI: [10.1016/S1353-8020\(13\)70013-0](https://doi.org/10.1016/S1353-8020(13)70013-0) (2014).
137. Bahuguna, J., Sahasranamam, A. & Kumar, A. Uncoupling the roles of firing rates and spike bursts in shaping the STN-GPe beta band oscillations. *PLOS Comput. Biol.* **16**, 1–31, DOI: [10.1371/journal.pcbi.1007748](https://doi.org/10.1371/journal.pcbi.1007748) (2020).
138. Wichmann, T. Neuronal Firing Before and After Burst Discharges in the Monkey Basal Ganglia Is Predictably Patterned in the Normal State and Altered in Parkinsonism. *J. Neurophysiol.* **95**, 2120–2133, DOI: [10.1152/jn.01013.2005](https://doi.org/10.1152/jn.01013.2005) (2005).
139. McIntyre, C. C., Grill, W. M., Sherman, D. L. & Thakor, N. V. Cellular Effects of Deep Brain Stimulation : Model-Based Analysis of Activation and Inhibition Cellular Effects of Deep Brain Stimulation : Model-Based Analysis of Activation and Inhibition. *J. Neurophysiol.* **91**, 1457–1469, DOI: [10.1152/jn.00989.2003](https://doi.org/10.1152/jn.00989.2003) (2004).
140. McIntyre, C. C., Savasta, M., Kerkerian-Le Goff, L. & Vitek, J. L. Uncovering the mechanism(s) of action of deep brain stimulation: Activation, inhibition, or both. *Clin. Neurophysiol.* **115**, 1239–1248, DOI: [10.1016/j.clinph.2003.12.024](https://doi.org/10.1016/j.clinph.2003.12.024) (2004).

Figures

Figure 1

Network model structure and finding appropriate DBS currents. (A) Schematic of the network model. The DBS is considered to be either inhibitory or excitatory input to the STN. (B) Details of the network connectivity. The i th STN neuron excites the i th GPe and GPi neurons. The i th GPe neuron inhibits the $(i-$

1)th and (i+1)th STN, GPi, and GPe neurons. Each GPi neuron inhibits its corresponding thalamus neuron. (C) The STN mean firing rates for IDBS and EDBS shown versus the amplitude of the stimulation current. Inset is zoomed-in presentation of the results in the rectangle. Each point in the plots is averaged over 20 trials. The IDBS curve crosses the EDBS curve at 126.57 and 147.36 pA points.

Figure 2

EDBS is able to quench PD-like b oscillations and to improve thalamus fidelity in the network model. (A) Membrane potential of three STN neurons in the network model in the healthy state (top), PD state (middle), and during regular EDBS (bottom). The black vertical thick lines indicate 50 mV. (B) Time resolved population firing rate of the STN neurons in the Healthy state (top), PD state (middle), and during regular EDBS. (C) Mean power spectrum (average of 50 trials) of the STN time resolved population firing rate in the healthy state (light green), PD state (brown), and during regular (cyan) and irregular (light blue) EDBS. (D-F) Population mean firing rate (D), Fano factor (E), and oscillation index (F) of the STN, GPe, and GPi in the network model (error bars show standard deviation; color codes correspond to C). (G) Thalamus fidelity in the healthy state, PD state, and during regular and irregular EDBS.

Figure 3

IDBS fails to quench PD-like oscillations and to improve thalamus fidelity in the network model. (A) Membrane potential of three STN neurons in the network model in the PD state (top), during regular IDBS (middle), and during irregular IDBS (bottom). The black vertical thick lines indicate 50 mV. (B) Time resolved population firing rate of the STN neurons in the PD state (top), during regular IDBS (middle), and during irregular IDBS. (C) Mean power spectrum (average of 50 trials) of the STN time resolved population firing rate in the healthy state (light green), PD state (brown), and during regular (cyan) and irregular (light blue) IDBS. (D-F) Population mean firing rate (D), Fano factor (E), and oscillation index (F) of the STN, GPe, and GPi in the network model (error bars show standard deviation; color codes correspond to C). (G) Thalamus fidelity in the healthy state, PD state, and during regular and irregular IDBS.

Figure 4

Burst profile of GPi neurons. (A) Burst rate of GPi neurons in healthy, PD, regular EDBS, irregular EDBS, regular IDBS and irregular IDBS states. (B and C) Mean burst duration (in msec) and mean number of spikes per burst in healthy, PD, regular/irregular EDBS, and regular/irregular IDBS states. The error bars show standard deviation.

Figure 5

The role of inhibitory rebound bursting in STN neurons during EDBS and IDBS. (A) Membrane potential of an STN neuron (top), and its corresponding T-type calcium current (bottom; see materials and methods) during EDBS with current of 147:36pA. (B) The same as A, EDBS with current of 126:57pA. (C) The same as A, during IDBS.

Figure 6

Tremor frequency profile in healthy and PD state. (A) Membrane potential of 10 thalamus neurons in the healthy network model during resting state. The orange vertical thick line indicates 50 mV. (B) From top to bottom, membrane potential, low threshold T-type calcium current, synaptic inhibitory input from the connected GPi and membrane potential of the corresponding GPi neuron (see Materials and methods) in the healthy state of the network model. (C, D) The same as A and B for PD state of the network model.

Figure 7

Tremor frequency profile of PD state during EDBS and IDBS. (A, B) Membrane potential of 10 thalamus neurons when STN is exposed to regular EDBS (A) and regular IDBS (B) during the resting state of the network model. The orange vertical thick lines indicate 50 mV. (C) Tremor-like frequency of thalamus in the healthy and PD states and when the STN is exposed to regular/irregular EDBS and regular/irregular IDBS (error bars show standard deviation). (D) From top to bottom, membrane potential, low threshold T-type calcium current, synaptic inhibitory input from the connected GPi and membrane potential of the corresponding GPi neuron (see Materials and methods) when STN is exposed to EDBS in the network model. (E) The same as D, when STN is exposed to the IDBS.