

Acute toxicity of manganese and its role in regulating testicular enzyme activity of male rats at different time points

Xiaonian Zhu

Guilin Medical University

Yonghua He

Guilin Medical University

Wenxiang Shi

Guilin Medical University

Lin Yang

Guilin Medical University

Yi Sun

Guilin Medical University

Chaoyan Ou (oak009@163.com)

Guilin Medical University https://orcid.org/0000-0003-4104-7721

Research article

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Abstract

Background: In order to provide scientific basis for toxicity evaluation and prevention of manganese, this study aims to investigate acute toxicity and testicular alkaline phosphatase (ALP), acid phosphatase (ACP), lactic dehydrogenase (LDH) in male rats after manganese exposure at different time points.

Methods: Under strict control of light/darkness (12 h:12 h), the experimental animals were injected intraperitoneally with MnCl2·4H2O at different zeitgeber time (ZT) after adaptive feeding for 7 days. The acute toxicity test was performed by Kunming mice treated with manganese once and observed for 14 days. LD50 was calculated by improved Karber method according to the death of animals in each group. Short-term repeated manganese exposure was conducted by continuous 30 mg/kg manganese exposure once a day for 21 days. And then the adult SD male rats were killed to detect activities of testicular ALP, ACP and LDH by ELISA at different ZT points the second day after exposure.

Results: Acute toxic reactions of mice exposed to manganese were varied at different ZT points. The LD50 at ZT2, ZT8, ZT14 and ZT20 were 472.0 mg/kg, 408.2 mg/kg, 303.3 mg/kg and 358.0 mg/kg, respectively. Furthermore, short-term repeated manganese exposure could induce the activity changes of ACP, ALP and LDH in testes at different ZT points. ALP increased at ZT20 while decreased at other time points (P<0.05). ACP only decreased at ZT2 (P<0.05). LDH increased at ZT2 and ZT8 (P<0.05), but decreased at ZT14 and ZT20 (P<0.05). In addition, all the testicular enzymes except ACP had interactions between manganese and exposure time.

Conclusion: The acute toxicity and function injury of male reproductive caused by manganese exposure are varied at different ZT points. The timing of toxic reaction needs to be considered in the toxicity evaluation of manganese.

Background

As a kind of heavy metal, manganese (Mn) is considered to be one of the necessary trace elements for human body. Manganese is involved in many important biochemical reactions as a component of metabolic enzymes or agonists [1]. In addition, manganese is a cofactor of many enzymes and necessary for reproduction. Previous study found that hormone secretion blocked by deficient manganese can cause hypaphrodisia, sterility, inhibition of reproductive function and obstruction of sperm maturation [2]. Excessive manganese can accumulate in testicle to disrupt male reproductive function and destroy the testicular structure [3]. These studies show that both deficient and excessive manganese can lead to reproductive dysfunction. In modern toxicology, the time study is becoming a new research focus for metal toxicity [4]. However, no time study has addressed the direct connection between manganese exposure and testicular enzyme activity in the context of reproductive disease.

The activities of alkaline phosphatase (ALP), acid phosphatase (ACP) and lactic dehydrogenase (LDH) in testicular tissue can be used as biomarkers for the injury of male reproductive and the change of sperm quality. Semen ALP, ACP and LDH are positively related to sperm motility and quantity, showing a

testicular origin of these enzymes [4]. Moreover, the levels of these enzymes in semen indicate the function, integrity and damage of spermatozoa, that is why they are recommended as important biomarkers of sperm quality [5, 6]. Accumulated evidence indicates these enzymes can be regulated by manganese exposure. Eidi A et al. showed that manganese could significantly attenuate the increase of serum ALP to nearly normal levels, exerting a hepatoprotection against CCl4-induced liver injury in rats [5]. Li P et al. assessed the relationships between multiple metals burden in human seminal plasma and semen quality parameters. They found manganese concentration was significantly higher in human with fertility problems than that in normal human, while the ACP activity was significantly higher in normal human [6]. Manganese treatment was reported to significantly decrease the cell viability of SK-N-MC cell and increase the release of LDH [7].

Circadian rhythm is a common biorhythm for physiological and behavioral activities of almost all organisms. The biological clock genes are also expressed in testicles and spermatozoa, suggesting circadian rhythm plays an important role in the regulation of sperm development and male reproduction [8, 9]. Because of circadian rhythm, the body has different sensitivities to the same poison at different times of the day and night [10]. As a result, the toxicity of most exogenous chemicals is related to the exposure time. The toxic reaction of animals to the same dose of chemicals sometimes is of different degrees simply because of different exposure time. It may even lead to a phenomenon called all or none, that highly sensitive at one time point while no reaction at another time point. Therefore, the toxicity of chemicals should be analyzed not only from the dose, but also from the time point of view.

In this study, acute toxicity parameters and testicular enzymes were used as indexes to explore the time toxicity of manganese. The timed acute toxicity and male reproduction in experimental animals exposed to manganese will provide relevant basis for toxicity evaluation, prevention and treatment of manganese.

Methods

Reagents

MnCl₂•4H₂O purity greater than 99% was purchased from Sigma (St. Louis, MO, USA). ACP, ALP and LDH kits were purchased from Lingnan Biological Products (Guangxi, China). BCA kit for protein concentration was purchased from Beyotime Biotechnology (Shanghai, China).

Experimental animals and feeding

The experimental animals used are of SPF grade and provided by the Experimental Animal Center of Guilin Medical University. The conditions of animal feeding room were as follows: light/darkness (12 h:12 h), temperature (24±1)°C, humidity (55±10)%. The experimental animals were free to drink and eat during the whole experiment period. When the experiments were finished, the animals were anaesthetized by intraperitoneal injection of pentobarbital sodium (50 mg/kg) using a 1-mL plastic syringe. After the animals were asleep, they were killed by cervical dislocation. The animals were managed in agreement with the criteria defined in the NIH publication (no. 85-23, revised in 1985), and the experimental protocol

was approved by the Ethics Committee for Animal Care of Guilin Medical University (no. GLMC201603027).

Definition of zeitgeber time (ZT)

The starting time of light is defined as ZT0 (equivalent to 7: 00 of Beijing time) [11]. The natural time is converted to ZT. The exposure time points ZT2, ZT8, ZT14 and ZT20 are equivalent to 9: 00, 15:00, 21:00 and 3: 00 of Beijing time, respectively.

Acute toxicity test

One hundred Kunming mice of 18-22 g were randomly divided into 20 groups after 7 days of adaptive feeding. According to the zeitgeber time points and 0.2 mL/10 g·body weight, the mice were injected intraperitoneally with 250.00 mg/kg, 329.56 mg/kg, 434.45 mg/kg, 572.72 mg/kg and 754.99 mg/kg of $MnCl_2\cdot 4H_2O$. After exposure, the toxicity and the number of dead animals were closely observed and recorded for 14 days. The LD_{50} and 95%CI were calculated with death number of animals according to the improved Karber method.

Short-term repeated toxicity test

Ninety-six healthy adult SD male rats with initial body weight of 251.06 ± 21.52 g were selected for the test. After adaptive feeding for 1 week, animals were randomly divided into 8 groups (4 control groups and 4 manganese exposure groups) and injected intraperitoneally with distilled water or 30 mg/kg MnCl₂·4H₂O at the corresponding ZT points for 21 days. The animals were killed for obtaining testes at different ZT points the second day after exposure. The testes were then stored at -80°C for detecting enzyme activity within 2 weeks.

Detection of ACP, ALP, LDH activity and protein concentration

Testicular tissue (0.3 g) on the same side of rats was added with 2.7 g pre-cooled saline for 10% tissue homogenate by electric homogenizer at 4°C. And then the tissue homogenate was centrifugated at 3000 r/min for 10 min and the supernatant was used to determine the activity of ACP, ALP and LDH by ELISA method on 7170A Automatic Biochemical Analyzer (Olympus, Japan). The protein concentration was determined by BCA kit on 721 Spectrophotometer (Olympus, Japan) and used to adjust the activity of above enzymes.

Statistical analysis

All the data was analyzed by SPSS 16.0 statistical software. Two samples t test and factorial analysis were carried out and P<0.05 was considered as statistically significant.

Results

Toxic reactions of mice in acute toxicity test

The experimental mice had different degrees of poisoning symptoms after MnCl₂•4H₂O exposure, the mild ones appeared listlessness, emaciation, loss of appetite or abolition, curling up, while standing unstable, hair removal and leg edema in the heavy ones. At later stage of the test, the mice fell on the ground, curled claws and toes, had leg stiffness and tremor. With dose of MnCl₂•4H₂O increased, all these reactions aggravated aggravation. The earliest symptom appeared at once upon MnCl₂•4H₂O exposure, and death began as early as 20 min and stopped on the 12th day after MnCl₂•4H₂O exposure. We found the dead mice had dark red of blood, light color of muscle, larger and darker of filling gallbladder and punctate bleeding liver or some khaki-yellow liver. There was no obvious pathological change in other organs.

$\ensuremath{\mathsf{LD}_{50}}$ and 95%Cl of mice exposed to manganese

The death of mice at ZT2, ZT8, ZT14 and ZT20 points was shown in Table 1. As shown in Table 2, the acute toxicity of mice after manganese exposure at ZT14 point was the highest, and the LD_{50} (95%CI) was 303.30 mg/kg (275.30-334.20 mg/kg). The LD_{50} (95%CI) at ZT20 and ZT8 points were 358.01 mg/kg (294.92-434.61 mg/kg) and 399.85 mg/kg (315.42-506.89 mg/kg), respectively. The acute toxicity at ZT2 was the least, and the LD_{50} (95%CI) was 471.95 mg/kg (376.14-592.17 mg/kg).

Effects of manganese exposure on testicular ALP, ACP and LDH of SD rats

As shown in Figure 1, short-term repeated manganese exposure could induce the activity changes of ACP, ALP and LDH in rat testes. However, the changes were varied at different time points. The activity of ALP increased at ZT20 point while decreased at other time points (P<0.05). The activity of ACP was reduced only at ZT2 point (P<0.05), but there was no significant difference in other time points (P>0.05). The activity of LDH increased at ZT2 and ZT8 time points (P<0.05), but decreased at ZT14 and ZT20 time points (P<0.05).

Interactions between testicular enzymes and ZT points after manganese exposure

Above results show that the activities of testicular ACP, ALP and LDH were varied at different ZT points upon manganese exposure, suggesting there may be some interactions between these testicular enzymes and ZT points. As shown in Table 3 and Figure 2, the changes of ALP and LDH in testicular tissue of rats had interactions between manganese and zeitgeber time points (P<0.01), while the interaction between manganese and ZT points of ACP enzyme was not statistically significant (P>0.05).

Discussion

The physiological and behavioral activities of organisms are largely affected by circadian rhythm [10], which leads to the toxic reactions of experimental animals are of varied poisoning degrees due to different exposure time to chemicals. Our results show that even if it is only once of manganese

exposure, the acute toxicities induced by manganese at different ZT points are different. Among them, the acute toxicity at ZT14 is the highest, followed by ZT20 and ZT14, and ZT2 is the least.

Spermatogenesis is a complex process of multitemporal phase, which is closely related to the activity of many enzymes in spermatogenic cells. LDH is the main enzyme that produces energy by glucose metabolism in spermatogenic cells and is one of the enzymes that produce ATP. LDH plays an important role in the metabolic process of sperm survival, movement and fertility, and is a marker predictive of sperm quality [12, 13]. Another key enzyme ACP is mainly distributed in the supporting cells of spermatogonia. It can be used as an index to measure whether spermatogenesis disorder occurs. In patients with clinical ejaculation disorder, ALP from testes and epididymis can be used as a marker to distinguish oligozoospermia from spermatozoa deficiency [14]. In wild boars, there was a correlation between sperm quality and sperm ALP release under maximum or minimum pressure [15]. Wang Meizhen et al. also reported that the increase of LDH enzyme activity was a crucial indicator of energy metabolism weakening and sperm deformity in spermatogenic cells [16]. The activity decrease of ACP enzyme may be caused by weak activity of sertoli cells, which also results in it as an important marker for the decrease of sperm count. This study indicates that manganese exposure can affect the activities of testicular ACP, ALP and LDH. Our results are consistent with those results of Adedara et al. [17], which shows that excessive manganese exposure could decrease the activities of ALP, ACP and LDH in testes.

Because of biorhythm, circadian rhythm implicates in sperm development and production [18]. We found that the activity changes of ALP, ACP and LDH were varied upon the same dose of manganese exposure at different ZT points. ALP increased at ZT20, but decreased at other time points. ACP only decreased at ZT2, and LDH increased at ZT2 and ZT8 while decreased at ZT14 and ZT20. However, the changes of these enzymes were consistent with the decrease of sperm count and motility [19]. Furthermore, factorial analysis showed that the changes of these enzymes in testicular tissue of rats had interactions between manganese and zeitgeber time points except ACP.

Conclusion

In summary, there is a difference of acute toxicity exposed to manganese at different zeitgeber time points. Moreover, the injury of male reproductive system caused by manganese is varied at different zeitgeber time points. In future, it is necessary to consider the timeliness of toxic reaction when evaluating the toxicity of manganese.

Abbreviations

ALP: alkaline phosphatase; ACP: acid phosphatase; LDH: lactic dehydrogenase; ZT: zeitgeber time; Mn: manganese

Declarations

Ethics approval and consent to participate

The animals were managed in agreement with the criteria defined in the NIH publication (no. 85-23, revised in 1985), and the experimental protocol was approved by the Ethics Committee for Animal Care of Guilin Medical University (no. GLMC201603027).

Consent for publication

Not applicable.

Availability of data and materials

All data generated or analysed during this study are included in this published article.

Competing interests

The authors declare that they have no competing interests.

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Authors' contributions

XNZ and CYO made contributions to the conception and design of the study. LY and YHH analyzed the data and wrote the first draft of the article. YS and WXS performed the experiments. All the authors read and approved the final manuscript.

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Not applicable.

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Tables

 $\textbf{Table 1} \ \ \text{Death of mice after intraperitoneal injection of } MnCl_2 \bullet 4H_2O \ \ \text{at different time points}$

Group		Dose		Number of mice	Death number	Death rate (p)	Survival rate (q)	p×q
		mg/kg	logarithm					
Z2	1	250.0	2.3979	5	0	0	1	0
	2	329.6	2.5179	5	2	0.4	0.6	0.24
	3	434.5	2.6379	5	2	0.4	0.6	0.24
	4	572.7	2.7579	5	3	0.6	0.4	0.24
	5	755.0	2.8779	5	4	0.8	0.2	0.16
Z8	1	250.0	2.3979	5	1	0.2	0.8	0.16
	2	329.6	2.5179	5	2	0.4	0.6	0.24
	3	434.5	2.6379	5	3	0.6	0.4	0.24
	4	572.7	2.7579	5	4	0.8	0.2	0.16
	5	755.0	2.8779	5	4	0.8	0.2	0.16
Z14	1	250.0	2.3979	5	0	0	1	0
	2	329.6	2.5179	5	4	0.8	0.2	0.16
	3	434.5	2.6379	5	5	1	0	0
	4	572.7	2.7579	5	5	1	0	0
	5	755.0	2.8779	5	5	1	0	0
Z20	1	250.0	2.3979	5	1	0.2	0.8	0.16
	2	329.6	2.5179	5	2	0.4	0.6	0.24
	3	434.5	2.6379	5	3	0.6	0.4	0.24
	4	572.7	2.7579	5	5	1	0	0
	5	755.0	2.8779	5	5	1	0	0

 $\textbf{Table 2} \quad \text{LD}_{50} \,\, \text{and} \,\,\, 95\% CI \,\, \text{of mice after intraperitoneal injection of } \,\, \text{MnCl}_{2} \, {}^{\bullet}4\text{H}_{2}\text{O} \,\, \text{at different zeitgeber time}$

ZT	LD ₅₀ (mg/kg)	95%CI (mg/kg)		
ZT2	471.95	376.14-592.17		
ZT8	399.85	315.42-506.89		
ZT14	303.32	275.30-334.20		
ZT20	358.01	294.92-434.61		

Table 3 Factorial analysis on the effects of manganese and exposed time points on ALP, ACP and LDH in testes of SD rats

Source	ALP		ACP		LDH	
	F	<i>P</i> -value	F	<i>P</i> -value	F	<i>P</i> -value
Adjusted model	60.125	0.000	6.487	0.000	82.189	0.000
Intercept	4.098E3	0.000	5.339E3	0.000	1.622E4	0.000
Mn	33.438	0.000	1.163	0.293	0.555	0.464
ZT	113.767	0.000	12.363	0.000	64.333	0.000
Mn*ZT	16.072	0.000	2.858	0.060	124.340	0.000

Figures

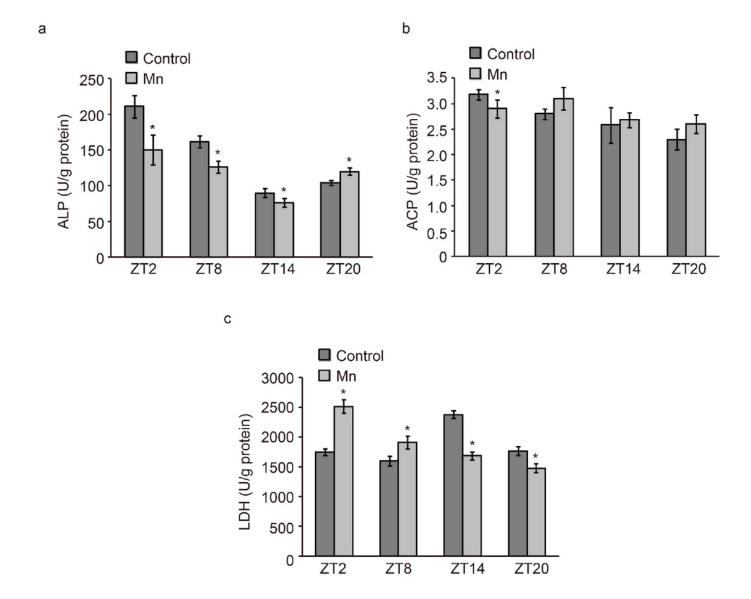


Figure 1

Effects of manganese (Mn) exposure on testicular ALP, ACP and LDH of SD rats at different ZT points. (a) The activity of testicular ALP upon manganese exposure. (b) The activity of testicular ACP upon manganese exposure. (c) The activity of testicular LDH upon manganese exposure. *, P<0.05 is based on the Student t test compared to the control. All results are from three independent experiments.

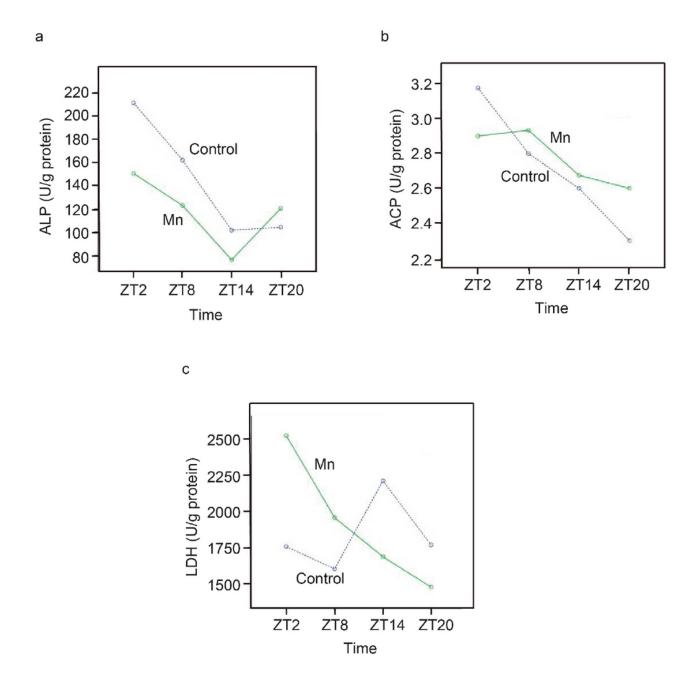


Figure 2

Interactions between testicular enzymes and ZT points after manganese exposure. (a) Interaction between testicular ALP and ZT points upon manganese exposure. (b) Interaction between testicular ACP and ZT points upon manganese exposure. (c) Interaction between testicular LDH and ZT points upon manganese exposure.

Supplementary Files

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