

S100A8/A9 Acts as Prognostic Indicators and Promotes Migration and Invasion via p38 MAPK Pathway in Nasopharyngeal Carcinoma

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Abstract

Background Nasopharyngeal carcinoma (NPC) is one type of malignancy associated with migration and invasion through a currently unclear mechanism. We previously discovered S100A8/A9 levels were roughly elevated in the plasma of NPC patients as the promising biomarkers. However, their expressions and underlying functions in NPC tissues are still unknown.

Methods In the present study, we analyzed 49 NPC tissues and 20 chronic pharyngitis (CP) tissues. Immunohistochemical staining was performed in different tissues and analyzed by mann-whitney U test statistically. Transwell migration and invasion experiments were further performed to determine S100A8/A9 effects on NPC.

Results Our results showed that S100A8/A9 in NPC tissues were significantly higher than those in CP tissues, closely associated with NPC clinical stages. Intriguingly, exogenous S100A8/A9 protein stimulation could dramatically enhance NPC migration and invasion abilities. In addition, p38 MAPK pathway blockade could diminish the migration and invasion of NPC cells stimulated by S100A8/A9 proteins. The downstream tumor invasion and migration associated proteins (e.g. MMP7) were also elevated in NPC tissues, consistent with S100A8/A9 overexpression.

Conclusions Taken together, our present findings suggest that the secreted soluble inflammatory factors S100A8/A9 might promote cancer migration and invasion via p38 MAPK signaling pathway along with invasion/migration associated proteins overexpression in the tumor microenvironment of NPC. This may shed light on the mechanism understanding of NPC prognosis and provide more novel clues for NPC diagnosis and therapy.

1. Introduction

Nasopharyngeal carcinoma (NPC) is the most common otorhinolaryngological tumor type originated from the upper skin. [1, 2] In 2018, the new incidence was estimated as 130,000 cases from the latest global cancer surveillance report. [3] NPC is well known as a cancer type characterized by distinct ethnic and regional specificity. [4] Endemic to China, this malignancy represents a variable occurrence rate ranging from the high incidence (Southern China) to a low rate (Northern China), while it is a rare disease among Caucasians. [4] Southern China (e.g. Guangdong, Guangxi) suffers a very high-risk of NPC, where the incidence ratio is much higher than the world's. [5, 6] Non-keratinized low differentiation NPC with high degree of malignancy is the main pathological type in clinical. [7] As high as 60–70% of the newly diagnosed NPC patients have already developed to local advanced lesions. [8, 9] Along with the development of modern radiotherapy and chemotherapy, the initial treatment response rate currently reaches 90.9% for these NPC patients with advanced stages (III and IV stage), [10] and the five-year survival rate is 72.3–86%. [11–13] However, NPC often has a high incidence of recurrence and distant metastasis, especially for those patients with advanced stages. As for them, the five-year incidence is 20–30% [14, 15] and the ten-year incidence is 30–40%. [16] What is worse, the treatment response rate decreased to 65.8–66.7%, [17, 18] and median survival time is just 14.0-27.2 months. [17, 19] These biological characteristics and the abundant peripheral lymphoid tissue involvement make NPC more prone to the metastasis and invasion compared with the other head and neck tumor types. The recurrence and distant metastasis are causes of treatment failure for NPC patients. Consequently, the majority of patients succumb to the effects of tumor metastasis rather than to the primary lesion. To better promote NPC prognosis and provide a rationale for novel therapies, it was absolutely essential to discover more promising biomarkers of NPC and unravel their underlying molecular mechanisms.

We previously developed a high-flux proteomic classification system to provide the highly accurate and innovative approach for NPC detection and diagnosis. [20] In the recent study, the plasma proteins from 244 NPC patients (II stage: 36 cases; III stage: 104 cases; IV stage: 104 cases) and 104 healthy donors were screened to discover potential novel NPC biomarkers using the techniques of isobaric tags for relative and absolute quantitation (iTRAQ) and liquid chromatography tandem mass spectrometry (LC-MS/MS). These NPC patients did not receive any treatment before diagnosis and were excluded if they had infection, diabetes, hypertension, autoimmune inflammation and other diseases. The findings showed that both of the S100A8 and S100A9 protein levels in the plasma of these NPC patients diagnosed at any different clinical stages were obviously higher than those in healthy donors, which suggested that S100A8/A9 may be the potential plasma biomarkers for NPC diagnosis. [21]

S100A8 (Calgranulin A, MRP8) and S100A9 (Calgranulin B, MRP14) are a pair of calcium binding proteins in S100 protein family, which often form a heterodimer complex in a calcium-dependent manner. They have the amino terminal ef-1 and carboxyl terminal ef-2 hand domains with low molecular weight (Mr 14000 and 13000, respectively) and display important functions in immunity. [22, 23] Tumor microenvironment (TME) is closely related to tumor occurrence and metastasis. The interactions between soluble factors in microenvironment and tumor cells play important roles in cancer development. [24–27] S100A8/A9 is a pair of secreted soluble inflammatory factors. Their main function is to drive a strong chemotaxis effect on the aggregation, adhesion and migration of white blood cells, as well as amplify the local inflammatory effects in microenvironment. [28, 29] To date, the expression of S100A8/A9 proteins in tumor tissues and their roles in microenvironment is still unknown for NPC.

In this study, we would uncover the expression status of S100A8 and S100A9 proteins in NPC tissues and further reveal a molecular basis for their effects on NPC cell proliferation, migration and invasion, which might provide a rationale for NPC prognosis and novel treatment.

2. Materials And Methods

2.1 Paraffin tissue samples

Paraffin embedded tissue samples with detailed pathological diagnosis information were collected from the pathology department of the affiliated first hospital, Guangxi medical university from January 2013 to June 2014 and the pathology department of the affiliated cancer hospital, Guangxi medical university from March 2012 to April 2013. Through careful inquiry of medical record information, only these NPC samples were included considered that the patients have not received any radiotherapy, chemotherapy or targeted therapy before diagnosis, and have no interference of other diseases including

infection, hypertension, diabetes, or autoimmune inflammation. Professional pathologists confirmed the NPC tissues were non-keratinizing undifferentiated carcinoma, and the degree of tissue infiltration (TI), lymph node metastasis (LNM) and clinical stage (CS) were based on the criteria of the Union for International Cancer Control (UICC, 8th Edition). The structure of NPC tissue samples was clear. The relevant study was conducted in accordance with the Declaration of Helsinki, and the protocol was approved by the patients' informed consents and the medical ethics committee of Guangxi medical university (No.050312).

2.2 Cell culture and cell proliferation

Human NPC cell lines CNE1 (high differentiation), CNE2 (low differentiation) and 6-10B (low tumorigenesis and metastasis) were all purchased from Cell Bank of Xiangya Central Laboratory, Hunan province, China. All the cell lines were cultured in RPMI-1640 medium (HyClone Company, USA) supplemented with 10% fetal bovine serum (Gibco Company, US), penicillin and streptomycin at 37 °C, 5% CO₂. The CCK8 assay was performed to test the cell proliferation, according to the manufacturer's instructions.

2.3 S100A8/A9 solution preparation

The S100A8 Recombinant Protein (Human, Amresco, USA) and S100A9 Recombinant Protein (Human, Amresco, USA) were mixed in a 1:1 ratio and placed at 4 °C for 1 h to form the complex (S100A8/A9).

2.4 Immunohistochemistry

The expression levels of S100A8 and S100A9 in NPC tissues and CP tissues were detected by immunohistochemistry (IHC). Tissue sections (0.6 μm) were used for dewaxing, hydration and antigen repair. S100A8 monoclonal antibody (1:250, Rabbit, Abcam, USA) and S100A9 monoclonal antibody (1:350, Rabbit, Abcam, USA) were used for the primary antibody incubation; PV-9000 reagent was used for secondary antibody incubation (Beijing zhongshanjinjiao company, China); DAB solution was used for color rendering (Beijing zhongshanjinjiao company, China). After section preparation, inverted microscope (Olympus, Japan) was used for sample observation. Five fields (200×) were taken from the top, bottom, left, right and middle, respectively for each sample slide. The software of Cell Sens Dimension was used to identify positive staining color (brown-yellow or tan), and calculate the percentage of positive staining cells in each field of vision. The positive imaging sample provided by reagent provider was used as the positive control, and PBS instead of primary antibody was used as the negative control. Similarly, Rabbit β-Catenin monoclonal antibody (Abcam, USA) and Rabbit MMP7 polyclonal antibody (OriGene, USA) were also used for the protein detection in clinical tissues.

2.5 Transwell migration and invasion experiment

The effect of S100A8/A9 on the migration ability of NPC cells was tested by transwell migration assay. CNE1, CNE2, 6-10B cells in logarithmic growth phase were cultured for 12 h for serum starvation, and then seeded into the upper chamber of transwell (1 × 10⁵ cells/well). RPMI 1640 medium containing 10% fetal bovine serum and 1 μg/ml S100A8/A9 were added in the lower chamber of transwell as the experimental group. Control group without S100A8/A9 protein was set for each cell line. After culture, five fields (200×, top, bottom, left, right and middle, respectively) on the down surface of the polycarbonate membrane were photographed using the inverted microscope (Olympus, Japan). The cells were counted and the average value was taken for statistics.

The effect of S100A8/A9 on the invasion ability of NPC cells was tested by transwell invasion assay. Matrigel was diluted with pre-cooled RPMI 1640 medium at the ratio of 1:6 and 100 μl mixture was evenly spread on the bottom of the upper chamber of transwell chamber to prepare the coating of basement membrane, subsequently the basement membrane was rehydrated using medium. The following steps are the same as the transwell migration experiment above. Similarly, 1 μg/ml S100A8/A9 was added to the lower chamber culture medium for the experimental group, while those without S100A8/A9 were set as the control groups.

2.6 Inhibition experiment of p38MAPK pathway and MEK pathway

Transwell assay was carried out after p38 MAPK pathway blockade using SB203580 to examine the effect of S100A8/A9 on NPC cell migration and invasion. SB203580 (CST, USA) dissolved in DMSO (Amresco, USA) was used to make 10 μM working concentration. Cells in logarithmic growth phase were cultured for 12 h for serum starvation, and then divided into two groups. Experimental group: cells pre-treated with SB203580 for 1 h and were cultured in RPMI 1640 medium containing 10% fetal bovine serum and 1 μg/ml S100A8/A9. Control group: cells pretreated with the same amount of DMSO as in the experimental group and cultured in RPMI 1640 medium containing 10% fetal bovine serum and 1 μg/ml S100A8/A9. Transwell migration and invasion experiments were carried out according to the experimental methods above. As for the MEK pathway blockade, the MEK1/2 selective inhibitor AZD6244 compound (CST, USA) was used, and the similar experiments were performed.

2.7 Statistics

All experimental data were statistically processed using SPSS 16.0 software. The area percentage of S100A8 and S100A9 positive staining cells was tested by normality and homogeneity of variance. They did not conform to the normal distribution, and the variances were not homogeneous. Therefore, the non-parametric statistical method was used. The comparisons between different groups were analyzed by mann-whitney U test. *P* < 0.05 was considered statistically significant. In the transwell migration and invasion experiment, these measurement data were shown as mean plus or minus standard deviation. Three independent experiments were repeated for each assay. These experimental results conform to the normal distribution. Student's t test was used for statistical analysis between different groups, and *P* < 0.05 was considered statistically significant.

3. Results

3.1 S100A8 and S100A9 proteins were frequently overexpressed in clinical NPC tissues.

To clarify the expression status of S100A8 and S100A9 proteins in NPC tissues, we performed the immunohistochemistry experiments to observe their expressions in 49 NPC cases and 20 chronic pharyngitis (CP) cases. Interestingly, we found a large number of brown-yellow staining signals for S100A8 proteins in the intercellular space and tumor cell cytoplasm of these NPC tissues including phase II, III and IV, while there was only a few brown-yellow staining in CP tissues (Figure. 1A and 1B). Similarly, a large number of brown-yellow staining signals for S100A9 proteins were also detected in the intercellular space and tumor cell cytoplasm of these NPC tissues including phase II, III and IV, while only a few brown-yellow staining for S100A9 was observed in CP tissues (Figure. 1C and 1D). The results indicated that abundant S100A8 and S100A9 proteins were expressed in NPC tissues including II, III and IV clinical stages, mainly distributed in the columnar epithelial interstitium and in the cytoplasm of NPC cells. In contrast, only a few S100A8 and S100A9 proteins existed in the CP tissues and most of them were concentrated in the columnar epithelial interstitium of tissues.

The statistically analyzed results showed that the positive staining area percentage (PSAP) of S100A8 and S100A9 in 49 cases of NPC tissues was 11.74 (8.08, 22.91) and 14.97 (10.55, 21.40), respectively, which were higher than those of 0.29 (0.07, 1.39) and 3.21 (1.98, 3.89) in 20 cases of CP tissues, with the significant differences (z-values - 6.34 and - 5.95, $P < 0.01$, respectively) (Figure. 1B and 1D, Table 1).

Table 1
Comparison of positive staining area percentage of S100A8 and S100A9 in NPC and CP tissues.

Group	N	S100A8 PSAP(%)	95% CI	Z	P	S100A9 PSAP(%)	95% CI	Z	P
NPC	49	11.74(8.08,22.91)	12.08-17.24	-6.34	0.000	14.97(10.55,21.40)	14.40-19.91	-5.95	0.000
CP	20	0.29(0.07,1.39)	0.30-1.23			3.21(1.98,3.89)	2.21-3.62		

3.2 S100A8 and S100A9 expression levels were closely related to NPC clinical stages.

In further stratified analysis, positive stained area percentages of S100A8 and S100A9 in II, III or IV stage NPC tissues, were both significantly higher than CP tissues ($P < 0.01$, respectively) (Figure. 1B and 1D, Table 2). In addition, the positive staining area percentages of S100A8 and S100A9 in advanced stage NPC (III stage or IV stage) were significantly higher than those in early stage NPC (II stage) ($P < 0.01$, respectively) (Table 2), while the III stage and IV stage were not statistical different (Figure. 1B and 1D, Table 2).

Table 2
Comparison of positive staining area percentage of S100A8 and S100A9 in NPC at different clinical stages and CP tissues.

G	N	S100A8 PSAP(%)	95% CI	Z	P	S100A9 PSAP(%)	95% CI	Z	P				
□	11	6.37(3.76,9.36)	4.46-10.49	△-4.27	^a -3.08	△0.000	^a 0.002	5.82(3.24,13.88)	5.50-12.26	△-3.51	^a -3.64	△0.000	^a 0.000
□	19	11.76(9.33,23.47)	11.60-19.93	△-5.34	^b -0.67	△0.000	^b 0.502	18.9(13.25,23.95)	16.01-23.57	△-5.34	^b -0.51	△0.000	^b 0.609
□	19	19.17(10.49,24.56)	13.25-22.19	△-5.11	^c -3.03	△0.000	^c 0.002	15.56(14.32,22.64)	14.13-24.51	△-5.34	^c -3.34	△0.000	^c 0.001
CP	20	0.29(0.07,1.39)	0.30-1.23					3.21(1.98,3.89)	2.21-3.62				

Importantly, we also explored the correlation between positive staining area percentage of S100A8 and S100A9 in NPC tissues and clinical features. Our data suggested that the expression levels of S100A8 and S100A9 in NPC tissues were not related to sex and age, tissue invasion and lymphatic node metastasis, but closely related to clinical stage ($P < 0.05$, respectively) (Table 3).

Table 3
Correlation between S100A8/S100A9 in NPC tissues and clinical features.

Characteristics	N	S100A8		B	S.E	Wald	P	OR	95%CI		
		High	Low						Lower	Upper	
Gender	Male	38	20	18	1.388	1.068	1.690	0.194	4.007	0.494	32.479
	Female	11	7	4							
Age	< 55	40	25	15	-2.259	1.196	3.568	0.059	0.104	0.010	1.089
	≥ 55	9	2	7							
TI	T1-T2	18	7	11	-1.700	1.317	1.665	0.197	0.183	0.014	2.416
	T3-T4	31	20	11							
LNM	N0-N1	15	4	11	-0.286	1.261	0.051	0.821	0.752	0.063	8.902
	N2-N3	34	23	11							
CS	I-II	11	1	10	4.724	2.096	5.078	0.024	112.576	1.850	6850.256
	III-IV	38	26	12							
Characteristics	N	S100A9		B	S.E	Wald	P	OR	95%CI		
		High	Low						Lower	Upper	
Gender	Male	38	22	16	-1.208	0.825	2.145	0.143	0.299	0.059	1.505
	Female	11	4	7							
Age	< 55	40	23	17	-0.044	0.982	0.002	0.965	0.957	0.140	6.557
	≥ 55	9	3	6							
TI	T1-T2	18	6	12	-0.249	0.955	0.068	0.794	0.780	0.120	5.062
	T3-T4	31	20	11							
LNM	N0-N1	15	4	11	-0.588	1.274	0.213	0.644	0.555	0.046	6.747
	N2-N3	34	22	12							
CS	I-II	11	1	10	3.855	1.900	4.117	0.042	47.234	1.140	1956.954
	III-IV	38	25	13							

3.3 S100A8/A9 stimulation promotes NPC cell proliferation, migration and invasion.

We previously found S100A8/A9 proteins were overexpressed in NPC cells and silencing of endogenous S100A8/A9 could significantly reduce NPC cell migration ability. [30] S100A8/A9 as a pair of secreted soluble inflammatory factors was also detected in the intercellular space besides the tumor cell cytoplasm in these NPC tissues as indicated above. To explore the effects of exogenous S100A8/A9 on NPC cell proliferation, we treated the NPC cells with S100A8/A9 at different concentrations to mimic S100A8/A9 infiltrated NPC microenvironment and detected cell proliferation by CCK-8 assay. The results suggested that all the three cell lines of CNE1 (high differentiation), CNE2 (low differentiation) and 6-10B (low tumorigenesis and metastasis) have already tended to increase their growths significantly in a dose dependent manner with S100A8/A9 treatment for 24 h, although there is still no overwhelming change in cell proliferation within the range of 0–5 µg/ml S100A8/A9 concentrations we tested (Supplementary Figure.S1).

In addition to the cancer cell proliferation, migration and invasion underlying metastatic dissemination is the key clinical problem in NPC. The present clinical investigation indicated NPC prognosis was remarkably associated with S100A8/A9 protein expression abundances. To further explore the effects of S100A8/A9 protein stimulation on NPC migration and invasion, we carried out the transwell migration and invasion experiments using these NPC cell culture models. Interestingly, the results indicated that as low as 1 µg/ml S100A8/A9 added to the lower chamber culture medium could already significantly driven the migration and invasion towards S100A8/A9 stimulation for all the three NPC cell lines including CNE1, CNE2 and 6-10B (Figure. 2A, 2B and 2C).

3.4 S100A8/A9 stimulation promotes migration and invasion of NPC cells via the p-38 MAPK pathway.

Mitogen-activated protein kinase (MAPK) pathway typically responds to extracellular stimulation and is involved into cancer metastasis. S100A8/A9 promote migration and invasion of gastric cancer through the p38 MAPK pathway. [31] To gain insight into the mechanism how S100A8/A9 regulates NPC migration and invasion, and explore whether the p-38 kinase is also involved into this process, a specific p-38 inhibitor SB203580 was used to block p38 MAPK pathway. The effect of S100A8/A9 on the migration and invasion ability of NPC cells was tested again by transwell experiments. Importantly, the results showed that p38 MAPK pathway inhibition after SB203580 pretreatment significantly reduced the migration and invasion abilities of these NPC cells even directly cultured in medium containing 1 µg/ml S100A8/A9 ($P < 0.05$, $P < 0.01$, respectively. Figure. 3A-3D). However, when we tested another MAPKs inhibitor AZD6244, which is a potent, selective MEK1/2 inhibitor as controls. The results indicated that AZD6244 treatment (MEK/ERK pathway inhibition) could not diminish the

migration and invasion abilities caused by S100A8/A9 stimulation in all the three NPC cell lines (Supplementary Figure.S2). These findings suggested that S100A8/S100A9 might promote the migration and invasion of NPC cells at least through p-38 MAPK pathway.

3.5 Tumor invasion and migration associated proteins β -catenin and MMP7 were elevated in clinical NPC tissues.

We previously discovered that S100A8 and S100A9 knockdown could significantly reduce the expressions of matrix metalloproteinases (MMPs) in NPC cancer cells[30]. In the present study, we further explored the downstream of signaling pathway and evaluated the β -catenin and MMP7 expression levels in clinical NPC tissues and chronic pharyngitis (CP) tissues, which are two important proteins in tumor cell invasion and migration. Here, the clinical tissues result indicated that abundant β -catenin and MMP7 proteins were expressed in NPC tissues including II, III and IV clinical stages. In contrast, only a few β -catenin and MMP7 proteins existed in the CP tissues (Fig. 4A and B). The statistically analyzed results showed that the positive staining area percentage (PSAP) of β -catenin and MMP7 in 42 cases of NPC tissues was 13.4(9.23,18.52) and 19.6(11.44,26.75), respectively, which were higher than those of 4.65(2.57,7.19) and 0.8(0.32,1.95) in 9 cases of CP tissues, with the significant differences (z-values - 3.83 and - 4.59, $P < 0.01$, respectively) (Table 4).

Table 4
Comparison of positive staining area percentage of β -Catenin and MMP7 in NPC and CP tissues.

Group	N	β -Catenin PSAP(%)	Z	P	MMP7 PSAP(%)	Z	P
NPC	42	13.4(9.23,18.52)	-3.83	0.000	19.6(11.44,26.75)	-4.59	0.000
CP	9	4.65(2.57,7.19)			0.8(0.32,1.95)		

Importantly, the positive stained area percentages of β -catenin and MMP7 in II, III or IV stage NPC tissues, were both significantly higher than CP tissues ($P < 0.01$, respectively) (Table 4). In further stratified analysis, the positive staining area percentages of β -catenin and MMP7 in advanced stage NPC (III stage or IV stage) were significantly higher than those in early stage NPC (II stage) ($P < 0.05$, $P < 0.01$, respectively), while the III stage and IV stage were not statistical different (Table 5).

Table 5
Comparison of positive staining area percentage of β -Catenin and MMP7 in NPC at different clinical stages and CP tissues.

G	N	β -Catenin PSAP (%)	Z	P	MMP7 PSAP (%)	Z	P
I	7	9.43 (6.84,9.61)	Δ -1.85	^a -2.36	Δ 0.064	^a 0.018	8.8 (7.54,10.46)
II	18	14.98 (8.64,18.52)	Δ -3.70	^b -0.45	Δ 0.000	^b 0.654	21.44 (14.42,26.86)
III	17	15.11 (10.59,25.91)	Δ -3.53	^c -2.89	Δ 0.000	^c 0.004	21.51 (16.76,30.52)
CP	9	4.65 (2.57,7.19)					0.8 (0.32,1.95)

4. Discussion

The expression of S100A8/S100A9 and their roles in NPC tissues is still not very clear up till the present moment. Over the past decade, Cheng, et al. identified several proteins including S100A8, S100A9 higher in NPC tissues than in normal nasopharyngeal epithelial tissues (NNET) by mass spectrometry (MS). [32] Li, et al. also independently discovered that the S100A9 protein in NPC was four times higher than that of NNET. [33] Other experiments including western blot were subsequently carried out to provide more evidences that S100A9 may be a potential biomarker in NPC tissue and the S100A9 level is markedly related to clinical typing of NPC. [34] However, it is still unclear that the association between these S100A8/A9 proteins and CP, a high risk stage as the early lesion before NPC, which might affect the specificity of these novel NPC biomarkers. Moreover, the molecular mechanism mediated by S100A8/A9 proteins in NPC migration and invasion is also currently not well understood.

In the past study, we discovered serum S100A8/A9 proteins roughly evaluated in NPC patients as the potential biomarkers, which are highly associated with NPC clinical stages. In the present study, we further detected the intracellular and extracellular S100A8/A9 proteins in NPC tissues and CP tissues. Similar results were observed in accordance with previous serum studies. These S100A8/A9 proteins in the intercellular space of NPC tissues and the cytoplasm of tumor cells were significantly higher than those of CP tissues. Meanwhile, we provide more solid evidences extracted from nearly fifty NPC patients that S100A8/A9 levels were closely related to the clinical stages of NPC, and the advanced stages were significantly higher than the early stages in these NPC tissues. The conclusion is supported by these above evidences that the expression level of S100A8/A9 in NPC tissues is dramatically elevated and closely related to the clinical stages.

To date, overexpressed S100A8/S100A9 proteins have already been observed in a variety of cancer types including breast cancer, [35, 36] prostate cancer, [37, 38] bladder cancer, [39, 40] and colon cancer. [41, 42] S100A8/S100A9 proteins play important roles in promoting cancer proliferation and enhancing their metastasis. Our previous study has shown that silencing of endogenous S100A8/A9 could obviously inhibit the migration of NPC cells. [30] It is well known that pathological stimulation of nasopharyngeal tissue, caused by bacterial/viral infection or inflammation is a risk factor of NPC. [43] The interactions between NPC cancer cells and stromal cells or immune cells including secrete cytokines act important roles in tumorigenesis. [44] Here, we use exogenous S100A8/A9 protein stimulation to mimic the S100A8/A9 infiltrated NPC microenvironment, where the secreted soluble inflammatory factors S100A8/A9 proteins are observed in the intercellular space and tumor cell cytoplasm of these NPC tissues. Interestingly, our results indicated that as low as 0–5 μ g/ml

concentration of S100A8/A9 proteins has already tended to promote NPC cell proliferation. In addition, the migration and invasion abilities were markedly enhanced by as low as 1 µg/ml S100A8/A9 proteins in a variety of NPC cell lines including the low-differentiated CNE2 as well as the high-differentiated CNE1 and even low metastatic 6-10B. Similarly, S100A8/A9 at a relatively low concentration (≤ 25 µg/ml) was reported to promote proliferation, migration and invasion of breast cancer cells. [45] S100A8/A9 proteins as low as 0.4-2 µg/ml were also discovered to promote migration and invasion abilities in one colorectal cancer study. [46] Taken together, it seems clear from present findings that S100A8/A9 protein stimulation could promote proliferation, migration and invasion of NPC cells at a low concentration level.

In addition, we also investigated whether the intracellular pathway is involved in S100A8/A9 stimulated NPC migration and invasion. Our findings indicated that this process might be involved into p38 MAPK pathway. When the p38MAPK pathway was inhibited, the migration and invasion abilities of NPC cells stimulated by S100A8/A9 were diminished. Consistent with the S100A8/A9 overexpression in NPC clinical tissues, the tumor invasion and migration associated proteins β -catenin and MMP7 were also elevated in these clinical NPC tissues. Therefore, here we hypothesized that overexpressed S100A8/A9 as the secreted soluble inflammatory factors in tumor microenvironment might enhance the activity/phosphorylation of p-38 MAPK pathway in cancer cells, which subsequently activated the transcriptional factors and elevated the tumor cell invasion and migration protein expression (e.g. MMP7), and finally promoted NPC cell proliferation, migration and invasion (Figure. 5). As is well known, p38 MAPK pathway is widely involved into cancer growth, development, proliferation, invasion, migration, metastasis, differentiation and other physiological processes. The abnormal or excessive activation of MAPK signaling pathway plays important roles in the malignant transformation and evolution of cells. Of note, similar mechanism was unraveled that activated p38 MAPK pathway under the stimulation of exogenous S100A8/A9 could enhance cell proliferation in breast cancer, [45] or promoted cell migration and invasion in gastric cancer. [31] In addition, clinical tumor metastasis and invasion/migration, is mainly dependent on the activities of protein family-matrix metalloproteinases (MMPs), which is involved into the process of Wnt/ β -Catenin and EMT signaling pathway [47].

5. Conclusion

Taken together, our study reveals that S100A8/A9 proteins are highly expressed in NPC tissues, markedly related to NPC clinical stages. Furthermore, S100A8/A9 overexpression in tumor microenvironment could promote NPC migration and invasion via the p38 MAPK signaling pathway and tumor cell invasion and migration protein overexpression (e.g. MMP7). The discovery of secreted soluble inflammatory factors S100A8/A9 as stimulators of NPC migration and invasion as well as better understanding of the S100A8/A9 actions in microenvironment could provide novel clues for NPC diagnosis and therapy.

Abbreviations

CP	chronic pharyngitis
iTRAQ	isobaric tags for relative and absolute quantitation
LC-MS/MS	liquid chromatography tandem mass spectrometry
MS	mass spectrometry
TME	tumor microenvironment
TI	tissue infiltration
LNM	lymph node metastasis
CS	clinical stage
IHC	immunohistochemistry
PSAP	positive staining area percentage
MAPK	mitogen activated protein kinase
NNET	normal nasopharyngeal epithelial tissues

Declarations

Author Contributions: All authors made a significant contribution to the work reported. In detail, conceptualization, X.Y. and Y.H.; methodology and data analysis, N.X., X.Y., Y.C., Q.H. and Z.K.; writing-original draft preparation, N.X., B.Z., X.Y., Y.C., Q.H. and Z.K.; writing-review and editing, N.X., B.Z., X.Y. and Y.H.; supervision, Y.H.; project administration, Y.H.; funding acquisition, Y.H.; All authors have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

Availability of data and materials: The datasets used and/or analyzed during the current study are available from the corresponding author on the reasonable request.

Ethics approval and consent to participate section:

The reporting studies involving human tissues in the present manuscript have included a statement on ethics approval and consent included the name of the ethics committee that approved the study and the committee's reference number.

Consent for publication section:

The reporting studies involving human tissues have included a statement on ethics approval and consent included the name of the ethics committee that approved the study and the committee's reference number in the present manuscript.

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Figures

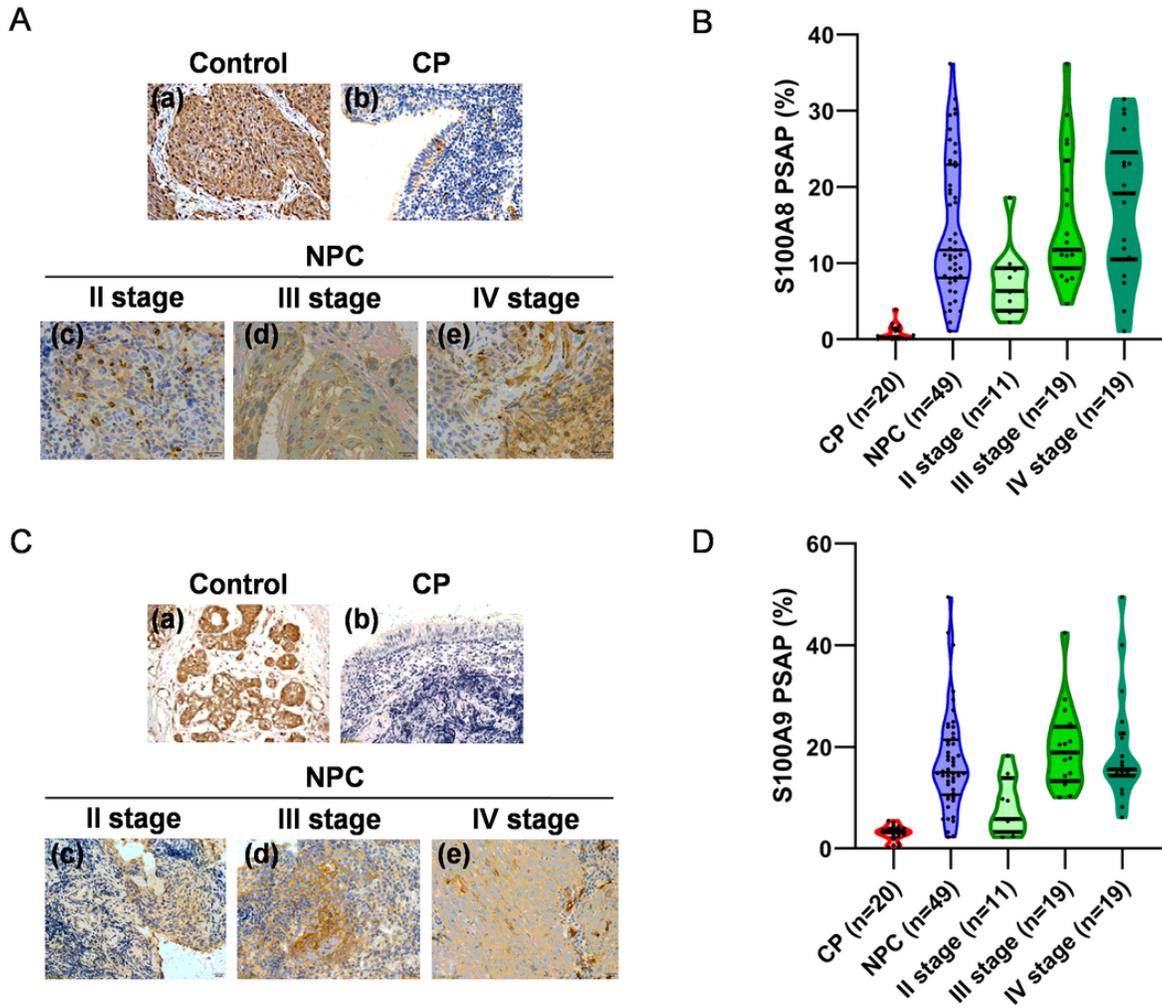


Figure 1

S100A8 and S100A9 expressions in NPC and CP tissues detected by immunohistochemistry method. A. Representative images of S100A8 expression (brown-yellow or tan staining) in positive control (a, human cervical carcinoma tissue provided by Abcam), CP tissue (b) or NPC tissues in stage II (c), III (d) and IV (e) (magnification 200×); B. Violin Plot of S100A8 positive staining area percentage (PSAP) for CP and NPC tissues including stage II, III, IV; C. Representative images of S100A9 expression (brown-yellow or tan staining) in positive control (a, human spleen tissue provided by Abcam), CP tissue (b) or NPC tissues in stage II (c), III (d) and IV (e) (magnification 200×); D. Violin Plot of S100A9 positive staining area percentage (PSAP) for CP and NPC tissues including stage II, III, IV. CP, chronic pharyngitis; NPC, nasopharyngeal carcinoma.

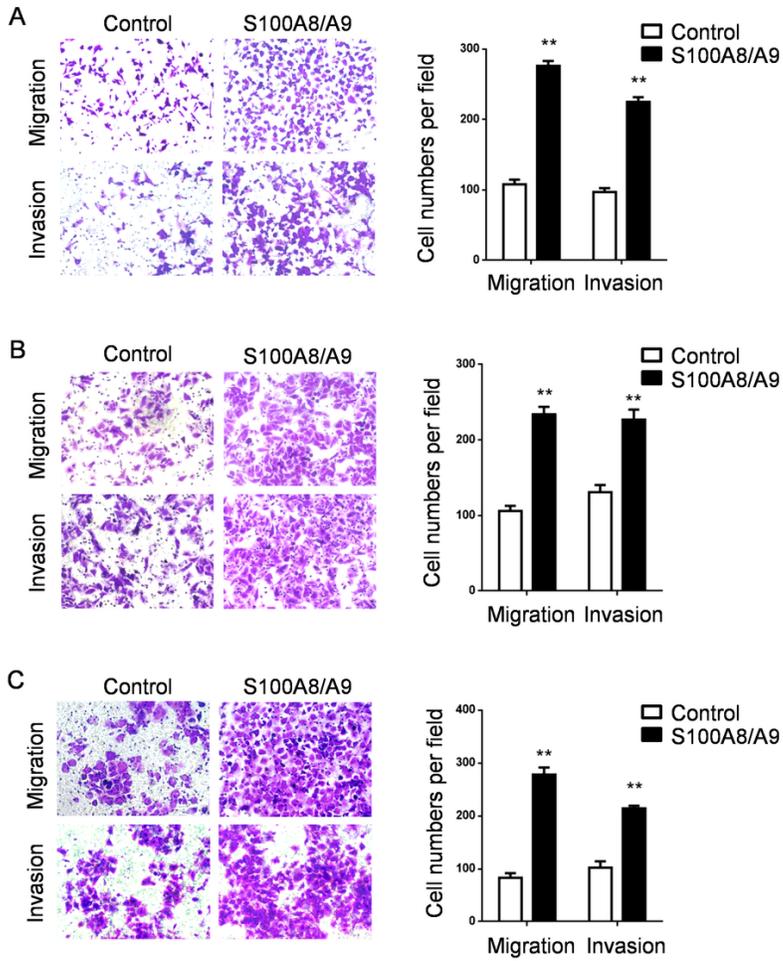


Figure 2

Effects of exogenous S100A8/A9 stimulation on the cell migration and invasion of NPC cells including CNE1, CNE2 and 6-10B. The lower chamber was filled culture medium with or without S100A8/A9 proteins (1 μ g/ml) to explore the migration and invasion abilities of NPC cells including CNE1 (A), CNE2 (B) and 6-10B (C). The left panels are representative images and right panels are quantifications. n=3, **P < 0.01.

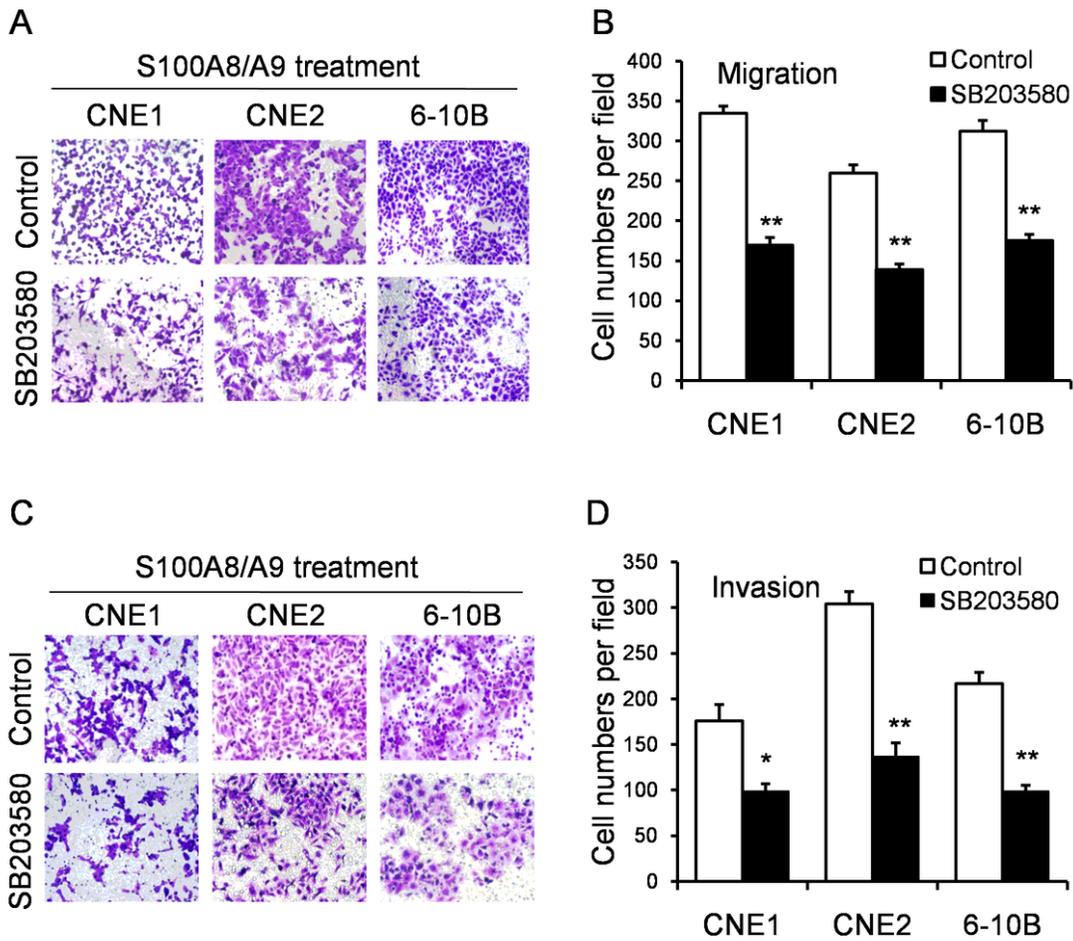


Figure 3

Effects of exogenous S100A8/A9 stimulation on p38-MAPK pathway in NPC cells. A. Representative images for the effects of S100A8/A9 proteins on migration of NPC cells after p38-MAPK inhibitor pretreatment (magnification 200 \times); B. Quantifications of S100A8/A9 protein effects on migration of NPC cells after p38-MAPK inhibitor pretreatment. $n=3$, ** $P < 0.01$; C. Representative images for the effects of S100A8/A9 proteins on invasion of NPC cells after p38-MAPK inhibitor pretreatment (magnification 200 \times); D. Quantifications of S100A8/A9 protein effects on invasion of NPC cells after p38-MAPK inhibitor pretreatment. $n=3$, * $P < 0.05$, ** $P < 0.01$.

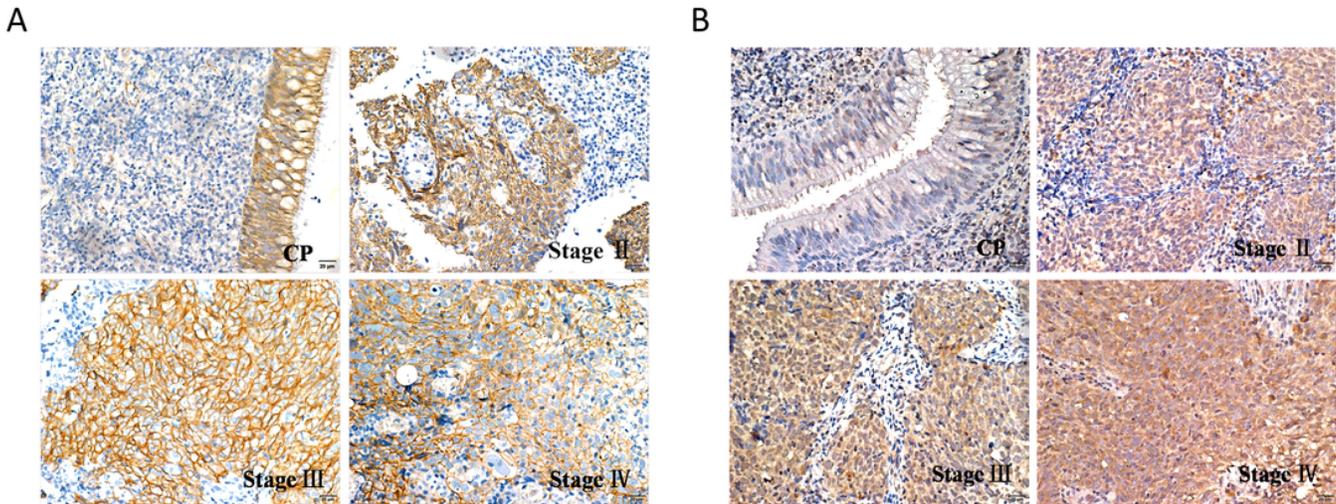


Figure 4
 β -catenin and MMP7 expressions in NPC and CP tissues detected by immunohistochemistry method. A. Representative images of β -catenin expression (brown-yellow or tan staining) in CP tissue or NPC tissues in stage II, III and IV (magnification 200 \times); B. Representative images of MMP7 expression (brown-yellow or tan staining) in CP tissue or NPC tissues in stage II, III and IV (magnification 200 \times); CP, chronic pharyngitis; NPC, nasopharyngeal carcinoma.

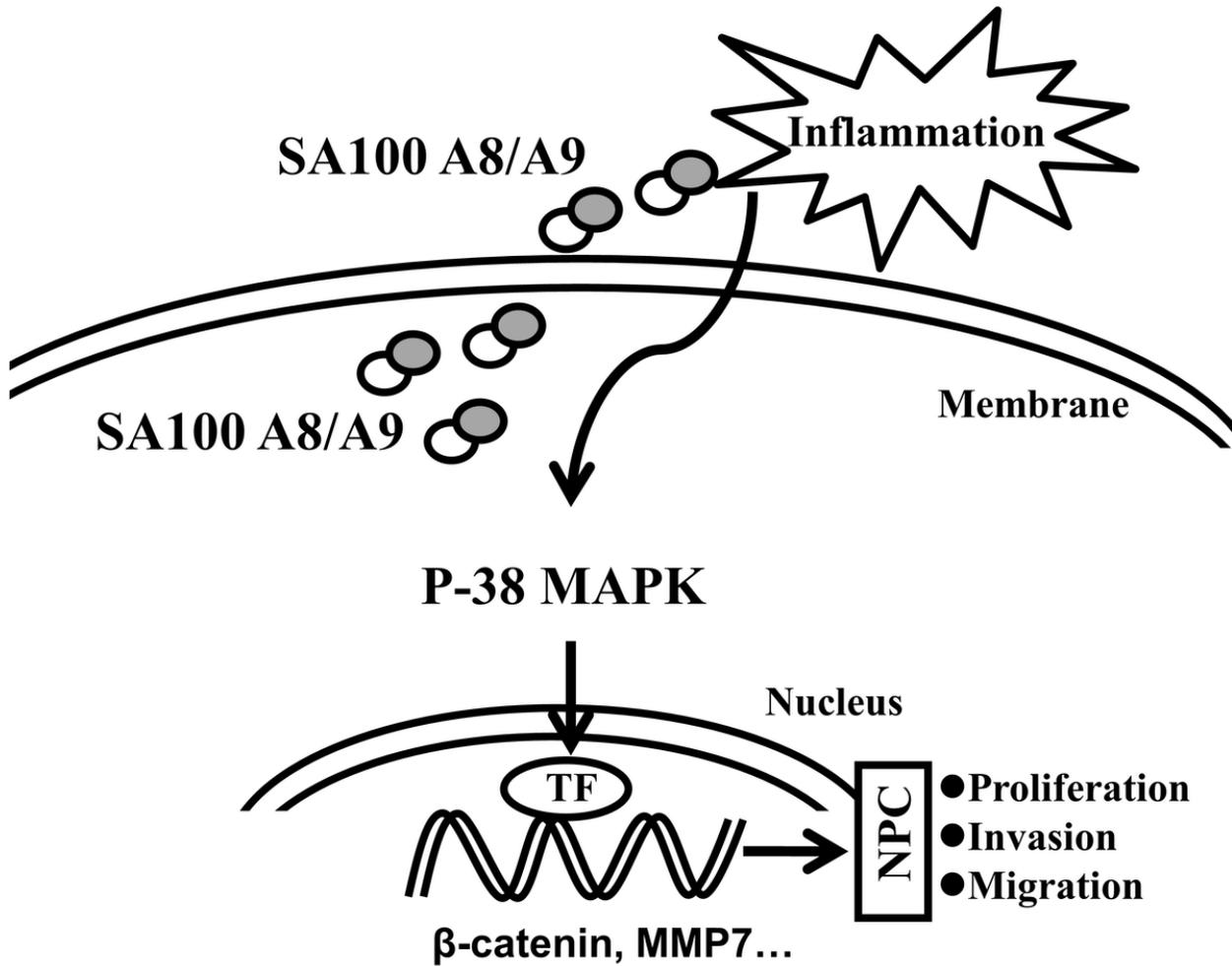


Figure 5

A hypothesis for the association between SA100A8/A9 proteins and NPC.

Supplementary Files

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