

# Production of Basil (*Ocimum Basilicum* L.) Under Different Soilless Cultures

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## Research Article

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# 1 **Production of basil (*Ocimum basilicum L.*) under different soilless cultures**

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10 **Abstract: Soilless cultivation systems that provide plant management in soilless**

11 **conditions in which the supply of water and of minerals is carried out by nutrient**

12 **solution, with or without a growing medium. The main aim of this paper was to**

13 **investigate the possibility of growing basil under three soilless systems (aeroponic,**

14 **hydroponic and peatmoss slab systems). A model was developed to predict the**

15 **nutrients consumption by basil plants. Shoot and root height, fresh and dry mass of**

16 **whole plant, nutrients uptake, and oil content were studied during the growth period**

17 **(after 4 and 7 weeks from transplanting). The results indicated that the shoot lengths**

18 **of basil plants were  $62.00 \pm 2.65$ ,  $57.83 \pm 7.42$  and  $48.77 \pm 2.89$  cm after 4 weeks and**

19  **$71.67 \pm 2.89$ ,  $65.67 \pm 1.15$  and  $62.33 \pm 2.31$ cm after 7 weeks from transplanting for**

20 **aeroponic, hydroponic and peatmoss slabs, respectively. The highest value of root**

21 **height of basil plants was  $37.67 \pm 6.66$  cm for aeroponic system. The dry mass of shoot**

22 **of basil plants ranged from  $28.48 \pm 0.91$  to  $44.77 \pm 0.97$  and  $72.98 \pm 0.83$  to  $117.93 \pm$**

23  **$1.40$  g plant<sup>-1</sup> after 4 and 7 weeks from transplanting, respectively. The highest values**

24 **of the N, P, K, Ca and Mg uptakes were  $262.50 \pm 6.84$ ,  $74.34 \pm 2.90$ ,  $195.13 \pm 4.09$ ,**

25  **$132.41 \pm 1.54$  and  $41.81 \pm 0.83$  mg plant<sup>-1</sup> and  $753.99 \pm 5.65$ ,  $224.88 \pm 3.05$ ,  $449.75 \pm$**

26  **$4.59$ ,  $529.12 \pm 6.63$  and  $112.44 \pm 1.67$  mg plant<sup>-1</sup> after 4 and 7 weeks from**

27 **transplanting, respectively. The basil oil content ranged from  $1.129 \pm 0.020$  to  $2.520 \pm$**

28  **$0.021$  and  $2.664 \pm 0.291$  to  $6.318 \pm 0.375$  g plant<sup>-1</sup> after 4 and 7 weeks from**

29 **transplanting, respectively at the same pervious order. The production costs of basil**

30 plant were 2.93, 5.27 and 6.24 EGP kg<sup>-1</sup> of plant. The model results were in a  
31 reasonable agreement with the experimental ones.

32 **Keywords:** Aeroponic, Hydroponic, Peatmoss Slab, Basil plant, Shoot, Root, Oil Content,  
33 model

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## 34 **Introduction**

35 There is an increasing interest recently for growing sweet basil (*Basilici herba*) in  
36 greenhouse soilless culture, which offer a suitable condition for maximization of  
37 production<sup>1,2</sup>. It is cultivated commonly in an open field with a variability in productivity and  
38 quality<sup>3</sup>.

39 Basil has high nutritional contents with low caloric values. It is used as a  
40 pharmacological raw material. Also, it contains vitamins A, B<sub>6</sub> and C as well as carotene  
41 besides calcium, potassium, phosphorus, magnesium, iron. Therefore, it needs a warm  
42 climate and high temperature and soil should be fecundity<sup>4,5</sup>.

43 The advantages of the soilless culture are the earliest growth and higher yield compared  
44 to traditional culture. Also, this system assures an equal supply of nutrient solution, so it can  
45 obtain a homogeneous crop. The mineral elements concentration and composition are well  
46 adjusted. Also, the buffer capacity of nutrient solution is low. pH and mineral composition of  
47 solution are easily changeable. Soilless culture decreases the time of adjusting solution<sup>6</sup>.

48 Soilless cultivation systems provide plant management under controlled water and  
49 minerals supply of the nutrient solution with or without medium. There are three systems of  
50 soilless cultivation namely, system with solid medium, in a liquid medium and aerated  
51 medium<sup>7,8</sup>.

52 Hydroponic system is a way plants without soil in water having a nutritional solution.  
53 The soil is used in traditional cultivation as a medium to add water and minerals in it, this soil  
54 is not needed in hydroponic because the minerals are added directly to water where the plants

55 grow. It is more efficient and controlled water can be reused after adjustment. It decreases the  
56 use of pesticides. It is used for many crops such as beets, radishes, carrots, potatoes, cereal  
57 crops, fruits, ornamentals and seasonal flowers can be grown on inert supporting medium  
58 instead of soil<sup>9,10,11</sup>.

59 **QI**<sup>12</sup> reported that the aeroponic system is a type of growing plants in air or mist  
60 environment without using any soil. In hydroponic, plant's roots are growing in water with  
61 nutrients. But for aeroponic, the nutrients are added through mist spray by sprinkles to plant's  
62 roots. The aeroponic system consist of a pump, nozzles, and growing chamber. There are a  
63 few types of aeroponic like low pressure type, high pressure type and commercial system.  
64 Basil is used as fresh and dried leaves a medicinal herb<sup>13,14</sup> for its diuretic and stimulating  
65 properties and also used in perfume compositions<sup>15</sup>. Basil is growing better in soilless  
66 systems than conventional systems and many studies have used basil as aquaponic or  
67 hydroponic crop<sup>16</sup>.

68 The most severe problem in the hydroponic system and soilless is the root rot which is  
69 due to the low oxygen level in the nutrient solution, therefore, proper aeration is required to  
70 overcome this problem. Aeroponic system is the proper solution to provide the plant with the  
71 required oxygen and nutrients. Besides, demand of organic production is increasing day after  
72 day. Therefore, this study aimed to improve the basil production under three soilless systems.

### 73 **Materials and methods**

74 The experiment was conducted at Agricultural and Bio-Systems Engineering  
75 Department, Faculty of Agriculture Moshtohor, Benha University, Egypt (latitude 30° 21` N  
76 and 31° 13` E). During the period of May to July, 2019 season.

### 77 **Culture systems description**

78 Fig. 1a and b show the experimental setup. It shows the system which consists of  
79 hydroponic system, aeroponic system, soilless substrate, solution system and pumps.

80 The hydroponic system (Deep Water Culture (DWC)) consists of three rectangular  
81 polyethylene tanks that used for basil plants culture. Dimensions of each tank are 80 cm long,  
82 40 cm wide and 30 cm high. The slope of hydroponic tanks was 2 % and stand 1 m high  
83 above the ground. The hydroponic tanks were covered with foam boards to support the  
84 plants. Each hydroponic tank provided with an air blower (Model NS 780 – Flow Rate 850 L  
85 h<sup>-1</sup> – Head 1.5 m – Power 15 W, China) to increase dissolved oxygen concentrations. The  
86 solution was circulated by a pump (Model First QB60 – Flow Rate 30 L min<sup>-1</sup> – Head 25 m –  
87 Power 0.5 hp, China) from the solution tank to the upper ends of the hydroponic tanks. Small  
88 tubes (16 mm) were used to provide tanks with solution in a closed system.

89 **Fig. 1a.**

90 **Fig. 1b.**

91 Aeroponic system consists of three rectangular polyethylene tanks that used for basil  
92 plants culture. Dimensions of each tank are 80 cm long, 40 cm wide and 50 cm high. The  
93 aeroponic tanks were established 1 m above the ground. Each aeroponic tank was divided  
94 into two parts, the lower part was made from polyethylene and the upper part was made from  
95 wood. The aeroponic tanks were covered with foam boards to support the plants. Each  
96 aeroponic tank was provided with two fog nozzles (8 L h<sup>-1</sup> discharge) located at the bottom of  
97 the tank sprayed nutrient solution into the tank in order to keep the roots wet. Small tubes (16  
98 mm) were used to provide aeroponic tank with solution in a closed system.

99 Soilless substrates consist are placed in three rows are 2 m long. Each row consists  
100 standard peat moss slabs (1.00 m x 0.20 m x 0.075 m). Basil plants were placed on row peat  
101 moss slabs with a drip irrigation system. There were three plants per slab giving a mean  
102 density of 9.0 plant m<sup>-2</sup>. Each plant was fed by a single drip.

103 The circular polyethylene tank of the nutrient solution system 500 liter capacity was  
104 used for collecting the drained solution by gravity from the ends of the three systems. The

105 nutrient solutions were prepared manually once per ten days by dissolving appropriate  
106 amounts of  $\text{Ca}(\text{NO}_3)_2$ ,  $\text{KNO}_3$ ,  $\text{K}_2\text{SO}_4$ ,  $\text{KH}_2\text{PO}_4$ ,  $\text{MgSO}_4$  and chelates for trace elements into  
107 preacidified groundwater. pH and Electrical Conductivity (EC) were further adjusted to  
108 6.5-7.0 and 1.4-1.8 dS  $\text{m}^{-1}$ , respectively, after salt addition. The minimum and maximum air  
109 ambient temperatures were 16 and 32 °C, respectively, and the minimum and maximum  
110 water temperatures were 15.8 and 27.5 °C, respectively. The Average relative humidity was  
111 65.4%.

## 112 **Basil Plants**

113 Basil seedlings were sown in the plastic cups (7 cm diameter and 7 cm height) filled with  
114 peat moss. The cups were irrigated daily using water with nutrient solution. Two weeks old  
115 basil seedlings were planted at 9.0 plant  $\text{m}^{-2}$  in the experimental tanks.

## 116 **Measurements**

117 Plant samples were taken during the vegetative and flowering stages (four and seven  
118 weeks after transplanting, respectively) for growth measurement and chemical analysis.  
119 Plant height, root length and the fresh and dry weight of leaves, stems and roots were  
120 determined. After measuring fresh mass, the plants were oven dried at 65 °C until constant  
121 weight was reached. Total content of macro elements was evaluated after being digested<sup>17</sup>.  
122 Nitrogen was determined by Kjeldahl digestion apparatus<sup>18</sup>. Potassium, Calcium and  
123 magnesium were determined by Photofatometer (Model Jenway PFP7 – Range 0 - 160 mmol  
124  $\text{L}^{-1}$ , USA) and phosphorus (P) was determined colorimetrically method<sup>19</sup>. The content of oil  
125 was determined in different organs: leaves, stems and inflorescences according to<sup>20</sup>.

126 Water samples were taken, at inlet and outlet of the culture units for measuring Nitrogen  
127 (N), Phosphorus (P), Potassium (K), Calcium (Ca) and Magnesium (Mg) were measured  
128 every week at 10 am during the experimental period.

## 129 **Total production cost**

130 The cost calculation based on the following parameters was also performed:

131 **Fixed costs (Fc)**

132 - Depreciation costs ( $D_c$ )

133 
$$D_c = \frac{P_d - S_r}{L_d} \quad (1)$$

134 Where:

135  $D_c$  is the depreciation cost, EGP year<sup>-1</sup>.

136  $P_d$  is the system price, EGP.

137  $S_r$  is the salvage rate (0.1 $P_d$ ) EGP.

138  $L_d$  is the system life, year.

139 - Interest costs ( $I_n$ ):

140 
$$I_n = \frac{P_d + S_r}{2} \times i_n \quad (2)$$

141 Where:

142  $I_n$  is the interest, EGP year<sup>-1</sup>.

143  $i_n$  is the interest as compounded annually, decimal. (12%)

144 Shelter, taxes and insurance costs ( $S_i$ ):

145 Shelter, taxes and insurance costs were assumed to be 3 % of the purchase price of the  
146 automatic feeder ( $P_m$ ).

147 Then:

148 
$$\text{Fixed cost} = D_c + I_n + 0.03 P_m / \text{hour of use per year} \quad (3)$$

149 **Variable (operating) costs (Vc):**

150 - Repair and maintenance costs ( $R_m$ ):

151 
$$R_m = 100 \% \text{ depreciation cost} / \text{hour of use per year} \quad (4)$$

152 - Energy costs (E):

153 
$$E = EC \times EP \quad (5)$$

154 Where:

155 E is the energy costs, EGP h<sup>-1</sup>.

156 EC is the electrical energy consumption, kWh.

157 EP is the energy price, 0.57 EGP kW<sup>-1</sup>.

158 Labor costs (L<sub>a</sub>):

$$159 \quad L_a = \text{Salary of one worker} \times \text{No. of workers} \quad (6)$$

160 Where:

161 L<sub>a</sub> is the Labor costs, EGP h<sup>-1</sup>.

162 Salary of one worker = 10 EGP h<sup>-1</sup>.

163 No. of workers = 1

164 Then:

$$165 \quad \text{Variable costs} = R_m + E + L_a \quad (7)$$

166 **Total costs (T<sub>c</sub>):**

$$167 \quad \text{Total costs} = \text{Fixed costs} + \text{Variable costs} \quad (8)$$

168 Table 1 shows the input parameters of calculate total production costs of basil plants  
169 grown in different soilless systems.

170 **Table 1.**

171 **Nutrients consumption rate**

172 The Nutrients consumption rate were calculated as the differences between the Nutrients  
173 at inlet and outlet of culture units by the following formula<sup>21</sup>:

$$174 \quad C_{Nc} = \frac{Nc_{in} - Nc_{out}}{\text{Number of plants}} \times Q \times 24 \quad (9)$$

175 Where:-

176 C<sub>Nc</sub> is the Nutrients consumption rate, mg day<sup>-1</sup> plant<sup>-1</sup>

177 N<sub>cin</sub> is the Nutrients at inlet of the hydroponic unit, mg L<sup>-1</sup>

178  $N_{c_{out}}$  is the Nutrients at outlet of the hydroponic unit,  $mg L^{-1}$

179  $Q$  is the discharge,  $L h^{-1}$

### 180 **Model development of nutrient consumption**

181 Model assumptions:

182 - N, P, K, Ca and Mg are the nutrients used in study.

183 - The plants are uniformly distributed in the solution, so they work as a  
184 uniform sink for water and minerals with space at any time.

185 - The root systems are uniformly dispersed in the solution with uniform root length  
186 density at any time.

187 - The whole root system uptake characteristics are uniform.

188 - Water losses by evaporation are negligible.

189 The simplest nutrient consumption models relate the nutrient consumption to the  
190 concentration gradient using some sort of proportionality factor such as root permeability or  
191 conductivity<sup>22,23</sup>. The nutrient consumption was determined by using the following equation:

$$192 \quad NC = a_{NC} \cdot \Delta C \quad (10)$$

193 Where:-

194  $NC$  is the nutrient consumption,  $mg plant^{-1} day^{-1}$

195  $\Delta C$  is the concentration gradient,  $mg plant^{-1} day^{-1}$

196  $a_{NC}$  is the proportionality factor, dimensionless

197 A similar model of nutrient consumption takes into consideration the differing effects  
198 caused by variations in root growth stage. Assuming that growth follows a first order  
199 differential equation and assuming that the root growth is exponential<sup>24</sup>, then Equation (3)

200 can be derived. This equation is presented in similar form to Equation (2) and use the  
 201 following equation:

$$202 \quad NC = \left( \frac{(C_{plant} - C_{plant0})}{A_r - A_{r0}} \right) \cdot \left( \frac{\ln\left(\frac{A_r}{A_{r0}}\right)}{t - t_0} \right) \cdot A_r \quad (11)$$

203 Where:

204  $C_{plant0}$  is the concentration of the nutrients in the plant at time  $t_0$ , mg plant<sup>-1</sup>

205  $A_r$  is the root surface area at time  $t$ , cm<sup>2</sup> plant<sup>-1</sup>

206  $A_{r0}$  is the root surface area at time  $t_0$ , cm<sup>2</sup> plant<sup>-1</sup>

207 Root surface area was calculated from root length and mean root radius using the  
 208 following equation:

$$209 \quad A_r = 2\pi r_0 L_r \quad (12)$$

210 The root length increment using the following equation<sup>25</sup>:

$$211 \quad \Delta L_r = \Delta DW_{root} v \quad (13)$$

212 Where:

213  $\Delta L_r$  is the root length increment, cm day<sup>-1</sup>

214  $\Delta DW_{root}$  is the daily amount of root dry mass increment, g day<sup>-1</sup>

215  $v$  is the ratio of root length and mass of roots, cm g<sup>-1</sup>

216 The daily amount of dry weight of roots is calculated from the following equation<sup>26</sup>:

$$217 \quad \Delta DW_{root} = \begin{cases} 5LAI & \text{for } LAI \leq 0.5 \\ 2.5 + 23.9(LAI - 0.5) & \text{for } LAI > 0.5 \end{cases} \quad (14)$$

218 Where:

219 LAI is the leaf area index,  $m^2 m^{-2}$

220 Leaf area index was changed in the same proportions as root length density to  
221 maintain a constant ratio between roots and shoots. The leaf area index is calculated from the  
222 following equation<sup>27</sup>:

$$223 \quad LAI = \frac{LAI_{\max}}{1 + K_2 e^{(-k_1 t)}} \quad (15)$$

224 Where:

225  $LAI_{\max}$  is the maximum leaf area index,  $m^2 m^{-2}$

226  $K_2$  and  $k_1$  are the coefficients of the growth functions

227 All computational procedures of the model were carried out using Excel spreadsheet.  
228 The computer program was devoted to mass balance for predicting the nutrients  
229 consumption. The differences between the predicted and measured values were evaluated  
230 using RMSE indicator (root means square error) which is calculated using the following  
231 equation:

$$232 \quad RMSE = \sqrt{\frac{\sum (Predicted - Measured)^2}{n}} \quad (16)$$

233 The parameters used in the model that were obtained from the literature are listed in  
234 Table 2. Fig. 2 shows flow chart of the model.

235 **Table 2.**

236 **Fig. 2.**

### 237 **Statistical analysis**

238 Four replicates of each treatment were allocated in a Randomize Complete Block Design  
239 (RCBD) in the system. Data were analyzed one-way ANOVA (analysis of variance) using  
240 statistical package for social sciences (spss v21). Means were separated using New Duncan

241 Multiple Range Test (DMRT). Data presented are mean  $\pm$  standard division (SD) of four  
242 replicates.

## 243 **Results and discussion**

### 244 **Shoot length**

245 Fig. 3 shows the shoot length of basil plants grown in different soilless systems  
246 (Aeroponic, hydroponic and peatmoss slabs) at the vegetative stage (4 weeks after  
247 transplanting) compared to the flowering stage (7 weeks after transplanting). The results  
248 indicate that the shoot in aeroponic was taller than those of hydroponic system and peatmoss  
249 slabs at the vegetative and flowering stages. It could be seen that the shoot length of basil  
250 plants were  $62.00 \pm 2.65$ ,  $57.83 \pm 7.42$  and  $48.77 \pm 2.89$  cm for Aeroponic, hydroponic and  
251 peatmoss slabs, respectively, after 4 weeks from transplanting, which they were  $71.67 \pm 2.89$ ,  
252  $65.67 \pm 1.15$  and  $62.33 \pm 2.31$  cm for aeroponic, hydroponic and peatmoss slabs, after 7  
253 weeks from transplanting at the same previous. These results agreed with those obtained by<sup>30</sup>  
254 whose found that the plants grown aeroponically were twice as high as those in hydroponics  
255 and 4 times taller than those grown in sand. These previous results may be due that the roots  
256 of aeroponics systems are hanged in mid-air inside containers or chambers at 100 % humidity  
257 and fed up a fine mist of nutrient solutions. This pervious system stimulates absorption of  
258 roots to much needed oxygen and nutrients, those increasing metabolism and rate of growth  
259 compared with soil<sup>31</sup>.

260 The statistical analysis showed that the differences between the obtained data of shoot  
261 length due to the effect of culture system (A) and plant age (B) were significant. The analysis  
262 showed also that the interaction between both AB was significant.

263 **Fig. 3.**

### 264 **Root length**

265 Fig. 4 shows the root length of basil plants grown in different soilless systems  
266 (Aeroponic, hydroponic and peatmoss slabs) at the vegetative stage (4 weeks after  
267 transplanting) compared to the flowering stage (7 weeks after transplanting). The results of  
268 measurements of root of the plants grown in aeroponic system were taller than those of  
269 hydroponic system and peatmoss slabs at the vegetative and flowering stages. It could be  
270 seen that the highest value of root length of basil plants was  $37.67 \pm 6.66$  cm for aeroponic  
271 system, while, the lowest value of root length of basil plants was  $27.67 \pm 0.58$  cm was found  
272 with peatmoss slabs. The root length for basil plants grown in aeroponic system were 1.68  
273 and 2.12 times taller than those grown in peatmoss slabs after 4 and 7 weeks from  
274 transplanting, respectively. These results agreed with those obtained by<sup>32</sup>. Also, many studies  
275 showed that the aeroponic system enhance the rates of plants growth by promoting the root  
276 aeration because of the root system is grown totally suspended at the air, giving the plant  
277 stem and roots systems access to 100% of the available oxygen at the air<sup>33</sup>. These results are  
278 in agreement with findings which were reported by<sup>34</sup> that they showed that plant root length,  
279 area, volume of aeroponic system were significantly exceeded the hydroponic and substrate  
280 systems.

281 The statistical analysis showed that the differences between the obtained data of root  
282 length due to the effect of culture system (A) and plant age (B) were significant. The analysis  
283 showed also that the interaction between both AB was significant.

#### 284 **Fig. 4.**

#### 285 **Fresh and dry mass of shoot**

286 Figs. 5a and b show the fresh and dry mass of shoot of basil plants grown in different  
287 soilless systems (Aeroponic, hydroponic and peatmoss slabs) at the vegetative stage (4 weeks  
288 after transplanting) compared to the flowering stage (7 weeks after transplanting). The results  
289 indicate that the fresh and dry of shoot grown in aeroponic system were better than those of

290 hydroponic system and peatmoss slabs at the vegetative and flowering stages. It could be  
291 seen that the fresh and dry mass of shoot of basil plants were  $140.00 \pm 13.76$ ,  $139.02 \pm 10.19$   
292 and  $102.06 \pm 35.54$  g plant<sup>-1</sup> and  $44.77 \pm 0.97$ ,  $32.36 \pm 0.68$  and  $28.48 \pm 0.91$  g plant<sup>-1</sup> for  
293 Aeroponic, hydroponic and peatmoss slabs, respectively, after 4 weeks from transplanting.  
294 Meanwhile, the results also indicate that the fresh and dry mass of shoot of basil plants were  
295  $438.61 \pm 42.61$ ,  $229.33 \pm 10.30$  and  $187.99 \pm 24.84$  g plant<sup>-1</sup> and  $117.93 \pm 1.40$ ,  $77.85 \pm 0.77$   
296 and  $72.98 \pm 0.83$  g plant<sup>-1</sup> for aeroponic, hydroponic and peatmoss slabs, respectively, after 7  
297 weeks from transplanting. We can explain those that the aeroponic system enhance the rates  
298 of plants growth by promoting the root aeration because of the root system is grown totally  
299 suspended at the air, giving the plant stem and roots systems access to 100% of the available  
300 oxygen at the air<sup>35</sup>.

301 The statistical analysis showed that the differences between the obtained data of fresh  
302 mass of shoot due to the effect of culture system (A) and plant age (B) were significant. The  
303 analysis showed also that the interaction between both AB was significant. Also, the  
304 statistical analysis showed that the differences between the obtained data of dry mass of  
305 shoot due to the effect of culture system (A) and plant age (B) were significant. The analysis  
306 showed also that the interaction between both AB was non-significant.

307 **Fig 5.**

### 308 **Fresh and dry mass of root**

309 Figs. 6a and b show the fresh and dry mass of root of basil plants grown in different  
310 soilless systems (Aeroponic, hydroponic and peatmoss slabs) at the vegetative stage (4 weeks  
311 after transplanting) compared to the flowering stage (7 weeks after transplanting). The results  
312 indicate that the fresh and dry of root grown in aeroponic system were better than those of  
313 hydroponic system and peatmoss slabs at the vegetative and flowering stages. It could be  
314 seen that the fresh and dry mass of root of basil plants were  $150.52 \pm 0.72$ ,  $128.15 \pm 2.32$  and

315  $49.17 \pm 4.52 \text{ g plant}^{-1}$  and  $39.11 \pm 2.14$ ,  $33.82 \pm 1.57$  and  $24.73 \pm 1.76 \text{ g plant}^{-1}$  for aeroponic,  
316 hydroponic and peatmoss slabs, respectively, after 4 weeks from transplanting. Meanwhile,  
317 the results also indicate that the fresh and dry mass of root of basil plants were  $452.02 \pm 8.94$ ,  
318  $337.97 \pm 12.20$  and  $324.94 \pm 5.48 \text{ g plant}^{-1}$  and  $114.22 \pm 5.05$ ,  $97.16 \pm 3.35$  and  $66.88 \pm 2.36$   
319  $\text{g plant}^{-1}$  for Aeroponic, hydroponic and peatmoss slabs, respectively, after 7 weeks from  
320 transplanting.

321 The statistical analysis showed that the differences between the obtained data of fresh  
322 mass of root due to the effect of culture system (A) and plant age (B) were significant. The  
323 analysis showed also that the interaction between both AB was significant. Also, the  
324 statistical analysis showed that the differences between the obtained data of dry mass of root  
325 due to the effect of culture system (A) and plant age (B) were significant. The analysis  
326 showed also that the interaction between both AB was non-significant.

327

328

### **Fig. 6.**

#### **Nutrients uptake**

330 Table 3 shows the nitrogen (N), phosphorus (P), potassium (K), calcium (Ca) and  
331 magnesium (Mg) uptake of basil plants grown in different soilless systems (Aeroponic,  
332 hydroponic and peatmoss slabs) at the vegetative stage (4 weeks after transplanting)  
333 compared to the flowering stage (7 weeks after transplanting). The results indicate that the  
334 uptake of nitrogen (N), phosphorus (P), potassium (K), calcium (Ca) and magnesium (Mg)  
335 by the basil plants were higher in aeroponic system compared those of hydroponic system  
336 and peatmoss slabs at the vegetative and flowering stages. It could be seen that the nitrogen  
337 (N) uptake of basil plants values were  $262.50 \pm 6.84$ ,  $145.01 \pm 4.91$  and  $185.58 \pm 4.22 \text{ mg}$

338 plant<sup>-1</sup> and 753.99 ± 5.65, 409.10 ± 5.28 and 387.50 ± 5.29 mg plant<sup>-1</sup> after 4 and 7 weeks  
339 from transplanting, respectively, for aeroponic, hydroponic and peatmoss slabs.

340 The results indicate that the phosphorus (P) uptake by basil plants values were 74.34 ±  
341 2.90, 48.34 ± 2.05 and 46.40 ± 3.28 mg plant<sup>-1</sup> and 224.88 ± 3.05, 131.86 ± 2.77 and 128.13  
342 ± 2.85 mg plant<sup>-1</sup> after 4 and 7 weeks from transplanting, respectively, for aeroponic,  
343 hydroponic and peatmoss slabs. The potassium (K) uptake by basil plants values were 195.13  
344 ± 4.09, 136.10 ± 5.51 and 135.06 ± 2.97 mg plant<sup>-1</sup> and 449.75 ± 4.59, 375.91 ± 4.34 and  
345 371.00 ± 3.97 mg plant<sup>-1</sup> after 4 and 7 weeks from transplanting, respectively, for aeroponic,  
346 hydroponic and peatmoss slabs. The calcium (Ca) uptake by basil plants values were 132.41  
347 ± 1.54, 92.86 ± 0.84 and 83.51 ± 1.32 mg plant<sup>-1</sup> and 529.12 ± 6.63, 371.91 ± 3.97 and 262.50  
348 ± 3.20 mg plant<sup>-1</sup> after 4 and 7 weeks from transplanting, respectively, for aeroponic,  
349 hydroponic and peatmoss slabs. The magnesium (Mg) uptake by basil plants values were  
350 41.81 ± 0.83, 30.53 ± 0.90 and 24.74 ± 0.58 mg plant<sup>-1</sup> and 112.44 ± 1.67, 84.53 ± 1.08 and  
351 71.88 ± 1.10 mg plant<sup>-1</sup> after 4 and 7 weeks from transplanting, respectively, for aeroponic,  
352 hydroponic and peatmoss slabs.

353 The highest values of the N, P, K, Ca and Mg uptakes were 262.50 ± 6.84, 74.34 ± 2.90,  
354 195.13 ± 4.09, 132.41 ± 1.54 and 41.81 ± 0.83 mg plant<sup>-1</sup> and 753.99 ± 5.65, 224.88 ± 3.05,  
355 449.75 ± 4.59, 529.12 ± 6.63 and 112.44 ± 1.67 mg plant<sup>-1</sup> after 4 and 7 weeks from  
356 transplanting, respectively, were found with aeroponic system. While, the lowest values of  
357 the N, P, K, Ca and Mg uptakes were 185.58 ± 4.22, 46.40 ± 3.28, 135.06 ± 2.97, 83.51 ±  
358 1.32 and 24.74 ± 0.58 mg plant<sup>-1</sup> and 387.50 ± 5.29, 128.125 ± 2.85, 371.00 ± 3.97, 262.50 ±  
359 3.20 and 71.88 ± 1.10 mg plant<sup>-1</sup> after 4 and 7 weeks from transplanting, respectively, were  
360 found with peatmoss slabs. These results agreed with those obtained by<sup>34</sup> they reported that  
361 the nutrients uptake of both aeroponic and hydroponic were higher than that in substrate  
362 cultivated.

363 The statistical analysis showed that the differences between the obtained data of  
364 nutrients uptake due to the effect of culture system (A) and plant age (B) were significant.  
365 The analysis showed also that the interaction between both AB was significant as shown in  
366 table 3.

### 367 **Table 3.**

#### 368 **Content of oil**

369 Fig. 7 shows the basil oil content in different soilless systems (aeroponic, hydroponic  
370 and peatmoss slabs) at the vegetative stage (4 weeks after transplanting) compared to the  
371 flowering stage (7 weeks after transplanting). The results indicate that the basil oil content  
372 higher in aeroponic system compared to those of hydroponic system and peatmoss slabs at  
373 the vegetative and flowering stages. It could be seen that the basil oil content values were  
374  $2.520 \pm 0.021$ ,  $1.722 \pm 0.026$  and  $1.129 \pm 0.020$  g plant<sup>-1</sup> for Aeroponic, hydroponic and  
375 peatmoss slabs, respectively, after 4 weeks from transplanting. Meanwhile, the results also  
376 indicate that the basil oil content were  $6.318 \pm 0.375$ ,  $4.359 \pm 0.404$  and  $2.664 \pm 0.291$  cm for  
377 aeroponic, hydroponic and peatmoss slabs, after 7 weeks from transplanting at the same  
378 previous. The statistical analysis showed that the differences between the obtained data of  
379 basil oil content due to the effect of culture system (A) and plant age (B) were significant.  
380 The analysis showed also that the interaction between both AB was significant.

### 381 **Fig. 7.**

#### 382 **Production costs**

383 Table 4 shows the total production costs of basil plants grown in different soilless  
384 systems (aeroponic, hydroponic and peatmoss slabs) at the end growing period. It could be  
385 seen that the results indicate that the production costs of basil plant were 2.93, 5.27 and 6.24  
386 EGP kg<sup>-1</sup> of plant. The total production costs of basil plants grown in hydroponic system  
387 were 1.8 times higher than those basil plants grown in aeroponic system, also the total

388 production costs of basil plants grown in peatmoss slabs were 2.1 times higher than those  
389 basil plants grown in aeroponic system. Besides it is considered as an organic product which  
390 is safe for the human health.

391 **Table 4.**

392 **Model results and validation:**

393 The model was validated using the experimental data. Figs. 8 and 9 show the predicted  
394 and the measured nitrogen (N), phosphorus (P), potassium (K), calcium (Ca) and magnesium  
395 (Mg) consumption of basil plants during the whole growth period. It could be seen that the N,  
396 P, K, Ca and Mg consumption by basil plants increased gradually until it reached the peak  
397 after 6 week and then decreased. The results indicate also that, the average daily N, P, K, Ca  
398 and Mg consumption by the model was in a reasonable agreement with those measured,  
399 where, the nitrogen ranged 2.657 to 13.763 mg plant<sup>-1</sup> day<sup>-1</sup> theoretically while it was from  
400 2.024 to 13.459 mg plant<sup>-1</sup> day<sup>-1</sup> experimentally during the whole period. The phosphorus  
401 ranged 0.417 to 3.593 mg plant<sup>-1</sup> day<sup>-1</sup> theoretically while it was from 0.292 to 3.739 mg  
402 plant<sup>-1</sup> day<sup>-1</sup> experimentally during the whole period. The potassium ranged 8.635 to 29.511  
403 mg plant<sup>-1</sup> day<sup>-1</sup> theoretically while it was from 5.963 to 28.318 mg plant<sup>-1</sup> day<sup>-1</sup>  
404 experimentally during the whole period. The calcium ranged 3.076 to 14.442 mg plant<sup>-1</sup> day<sup>-1</sup>  
405 theoretically while it was from 3.495 to 13.853 mg plant<sup>-1</sup> day<sup>-1</sup> experimentally during the  
406 whole period. The magnesium ranged 0.471 to 1.376 mg plant<sup>-1</sup> day<sup>-1</sup> theoretically while it  
407 was from 0.427 to 1.344 mg plant<sup>-1</sup> day<sup>-1</sup> experimentally during the whole period. RMSE of  
408 N, P, K, Ca and Mg consumption were 0.73, 0.21, 1.5, 0.21 and 0.11, respectively, which  
409 means the predicted values were close to the measured values.

410 **Fig. 8.**

411 **Fig. 9.**

412 The best fit for the relationship between the predicted and the measured values of  
413 nutrients consumption was in the following form:

$$414 \quad \text{NC}_P = a\text{NC}_M + b \quad (17)$$

415 Where:

416  $\text{NC}_P$  is the predicted nutrients consumption,  $\text{mg plant}^{-1} \text{ day}^{-1}$

417  $\text{NC}_M$  is the measured nutrients consumption,  $\text{mg plant}^{-1} \text{ day}^{-1}$

418 The constants of these equation and coefficient of determination are listed in Table 4.

419 **Table 4.**

## 420 **Conclusions**

421 The experiment was carried out to study was conducted to investigate the possibility of  
422 growing basil under three soilless systems (aeroponic, hydroponic and peatmoss slabs). To  
423 achieve that study the effect of different soilless systems and plant age on shoot and, root  
424 length, fresh and dry mass of shoot, fresh and dry mass of root, nutrients uptake, basil oil  
425 content and costs. A mathematical model for mass balance of the system was developed  
426 successively for predicted the nutrients consumption by basil plant. It is concluded that the  
427 highest values of shoot length, root length, fresh and dry mass of shoot, fresh and dry mass of  
428 root were obtained for the basil grown under aeroponic system. The highest values of the N,  
429 P, K, Ca and Mg uptakes were found with aeroponic system, while, the lowest values of the  
430 N, P, K, Ca and Mg uptakes were found with peatmoss slabs. The highest value of the basil  
431 oil content was found with aeroponic system. The production costs of basil plant were 2.93,  
432 5.27 and 6.24 EGP  $\text{kg}^{-1}$  of plant. Besides it is considered as an organic product which is safe  
433 for the human health. Further work should be carried out to study the effect of more  
434 parameters such the environmental parameters on the plant growth and yield. The model  
435 results were in a reasonable agreement with the experimental ones.

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532

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537

538 **Author Contributions**

539 El-Sayed Khater, Adel Bahnasawy, Wael Abass, Osama Morsy, Hossam El-Ghobashy,

540 Yousry Shaban and Mohsen Egela: Investigation, Resources, Writing—Original Draft

541 Preparation, Writing—Review and Editing.

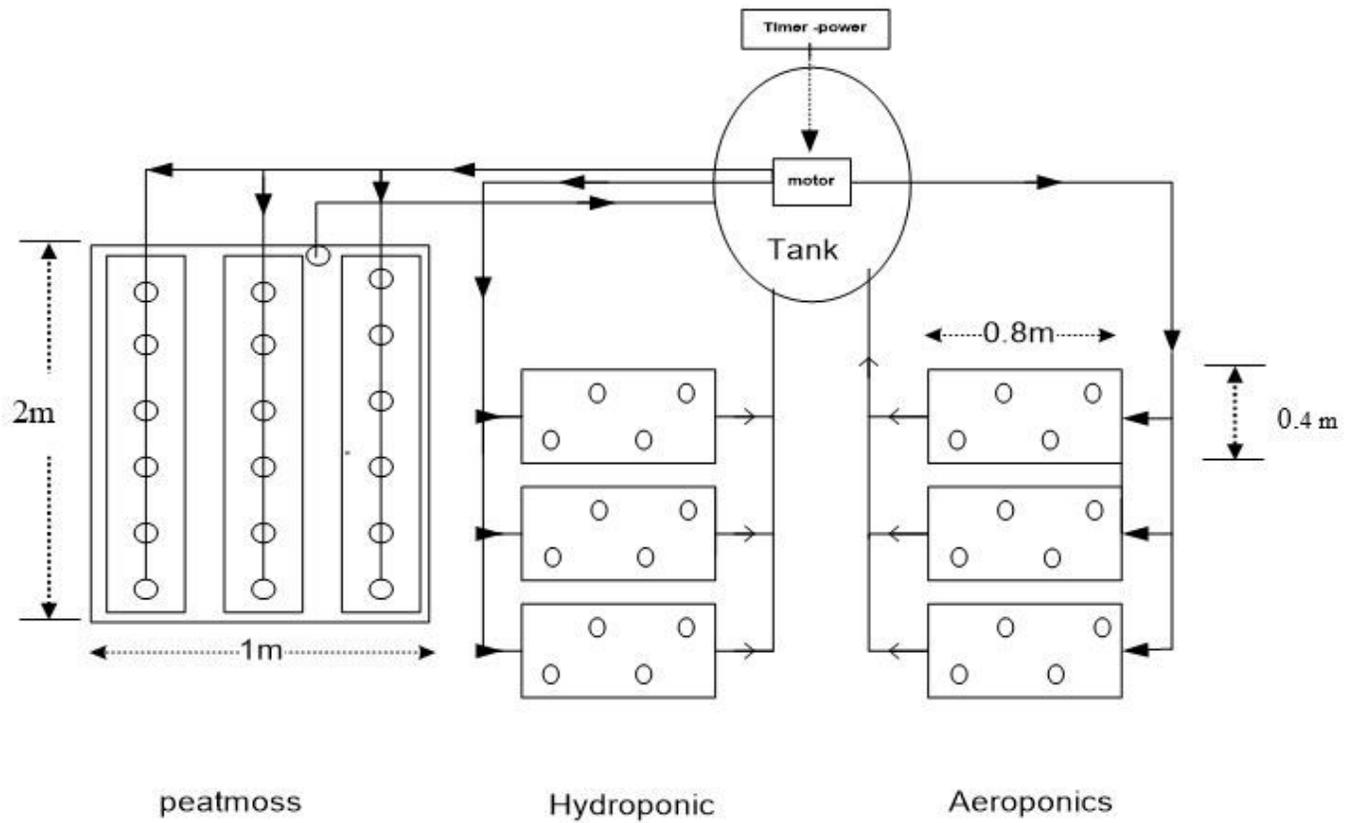
542

543 **Competing interests**

544 The authors declare no competing interests.

545

# Figures



**a.**



Peatmoss



Hydroponic

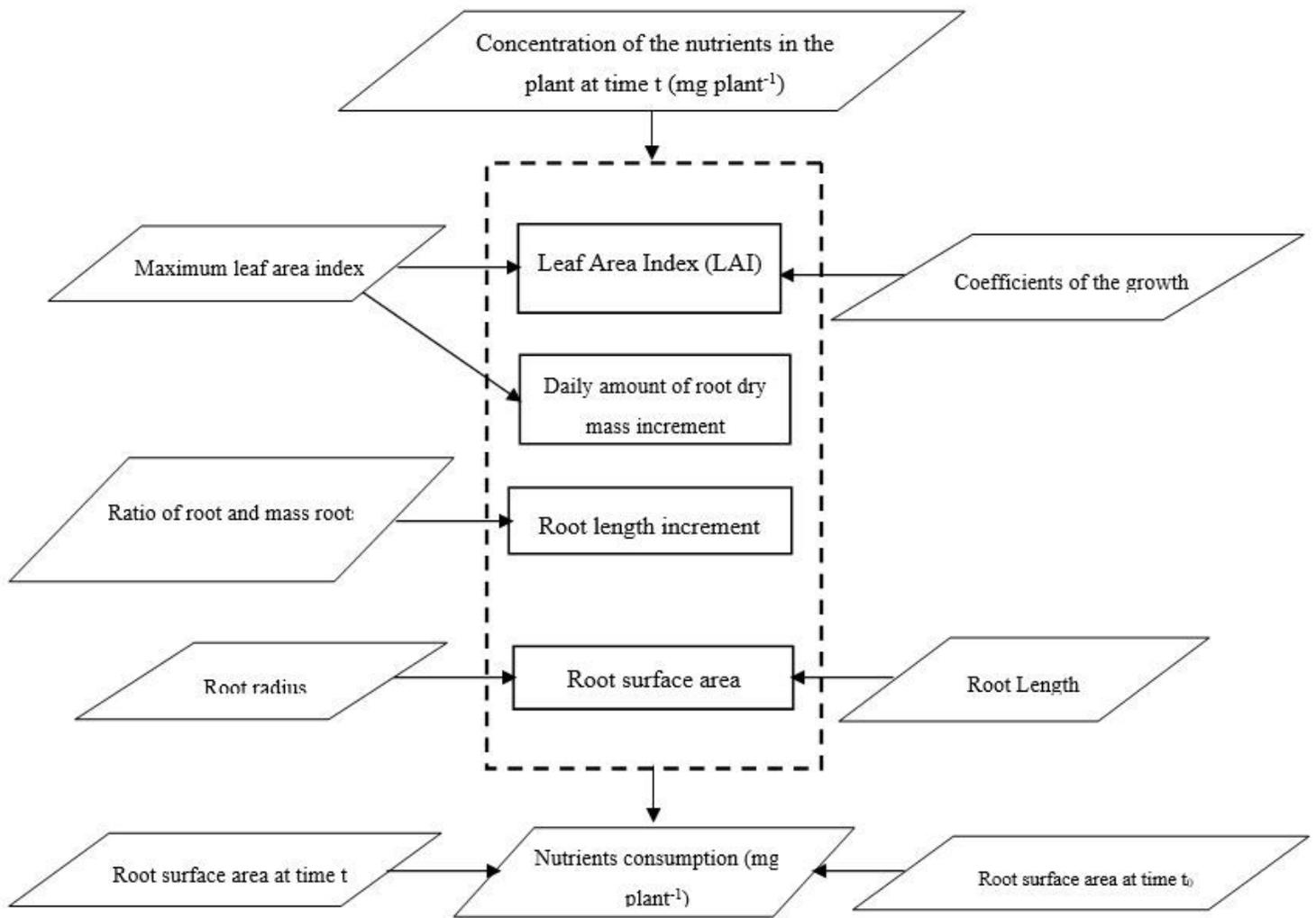


Aeroponic

**b.**

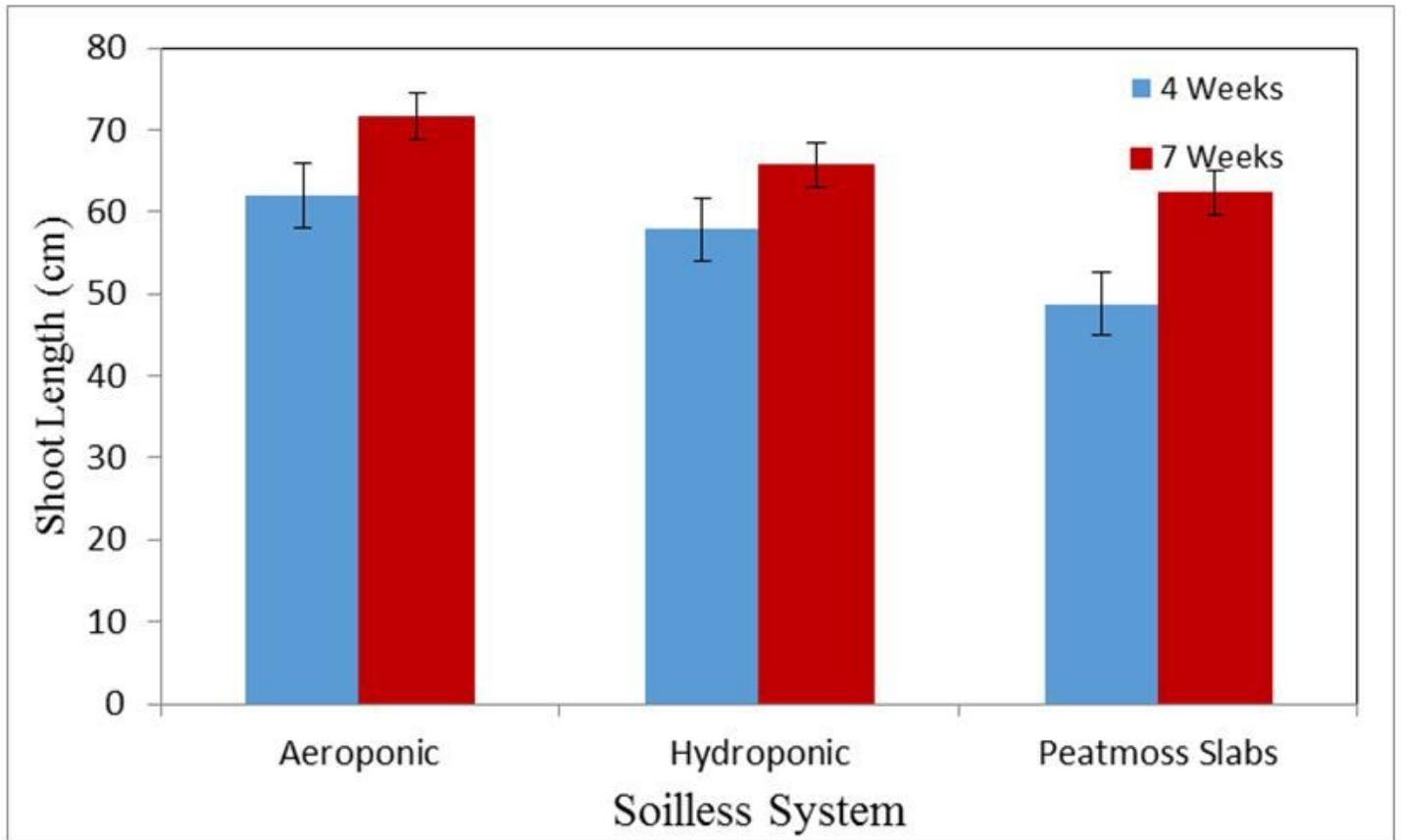
**Figure 1**

1a. The experimental setup 1b. Images of system



**Figure 2**

Flow chart of nutrients consumption rate.



**Figure 3**

The shoot length of basil plants grown in different soilless systems.

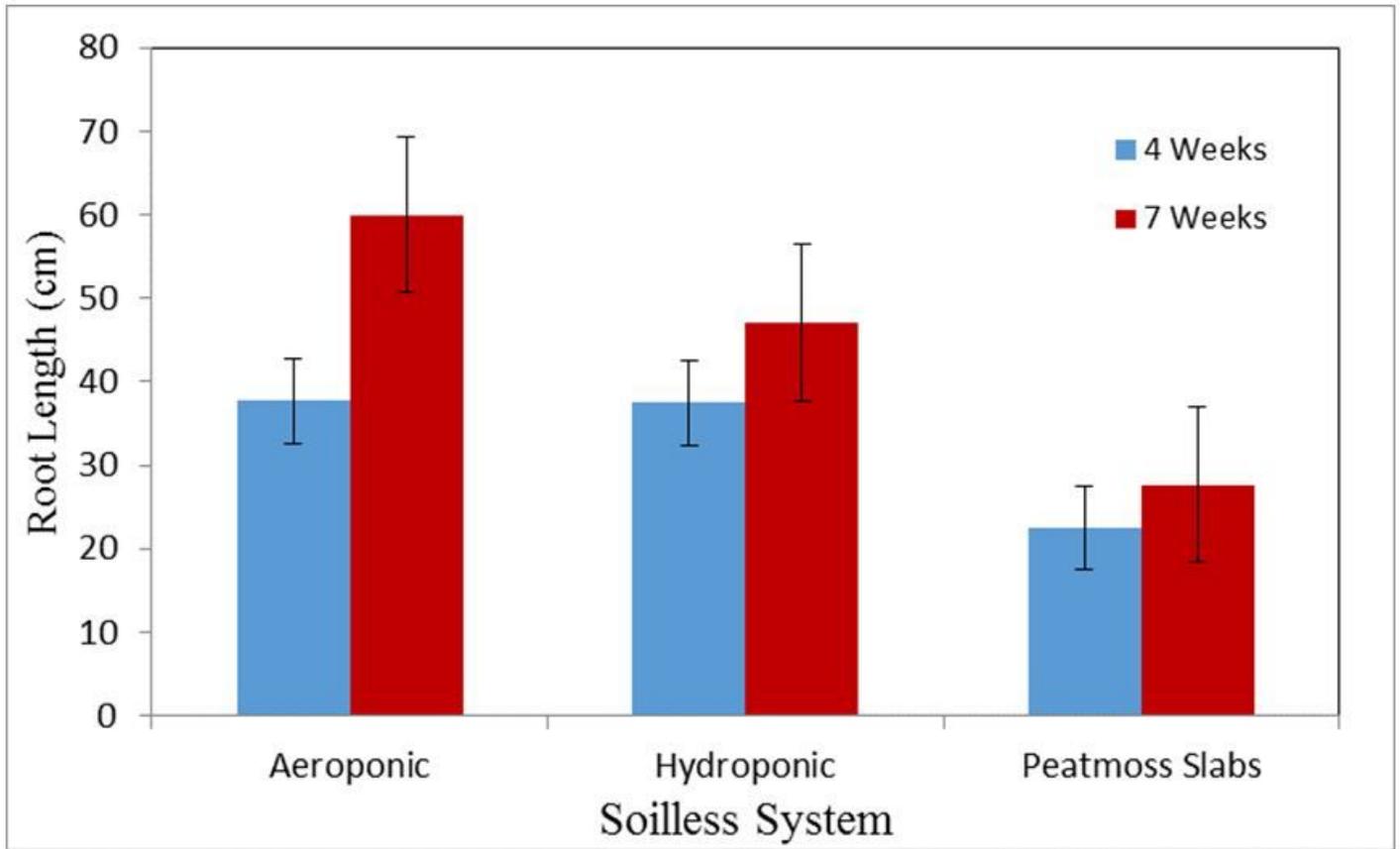


Figure 4

The root length of basil plants grown in different soilless systems.

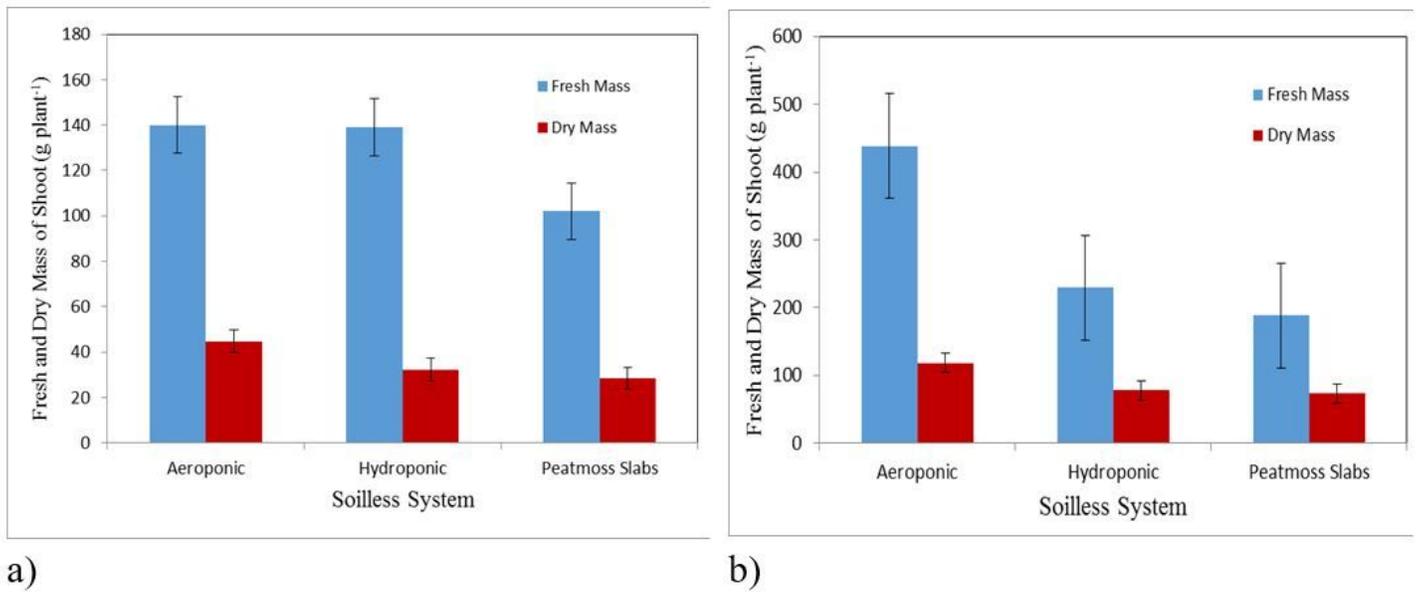
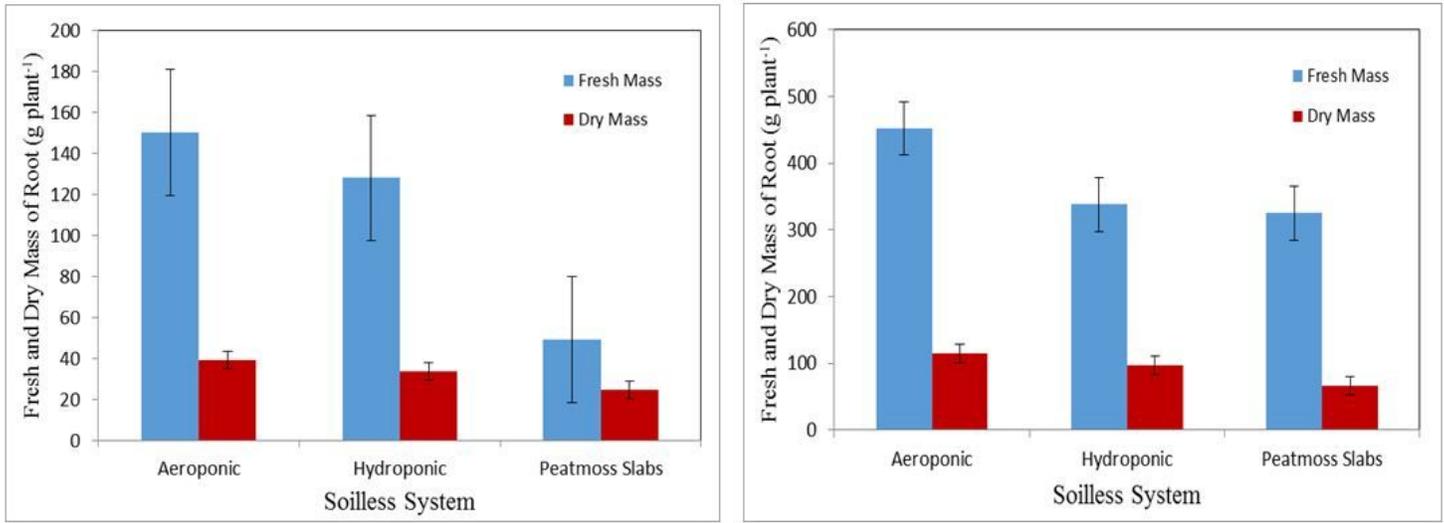


Figure 5

Fresh and dry mass of shoot of basil plants, a) At vegetative stage, and b) at flowering stage.



a)

b)

Figure 6

Fresh and dry mass of root of basil plants, a) at vegetative stage and b) at flowering stage.

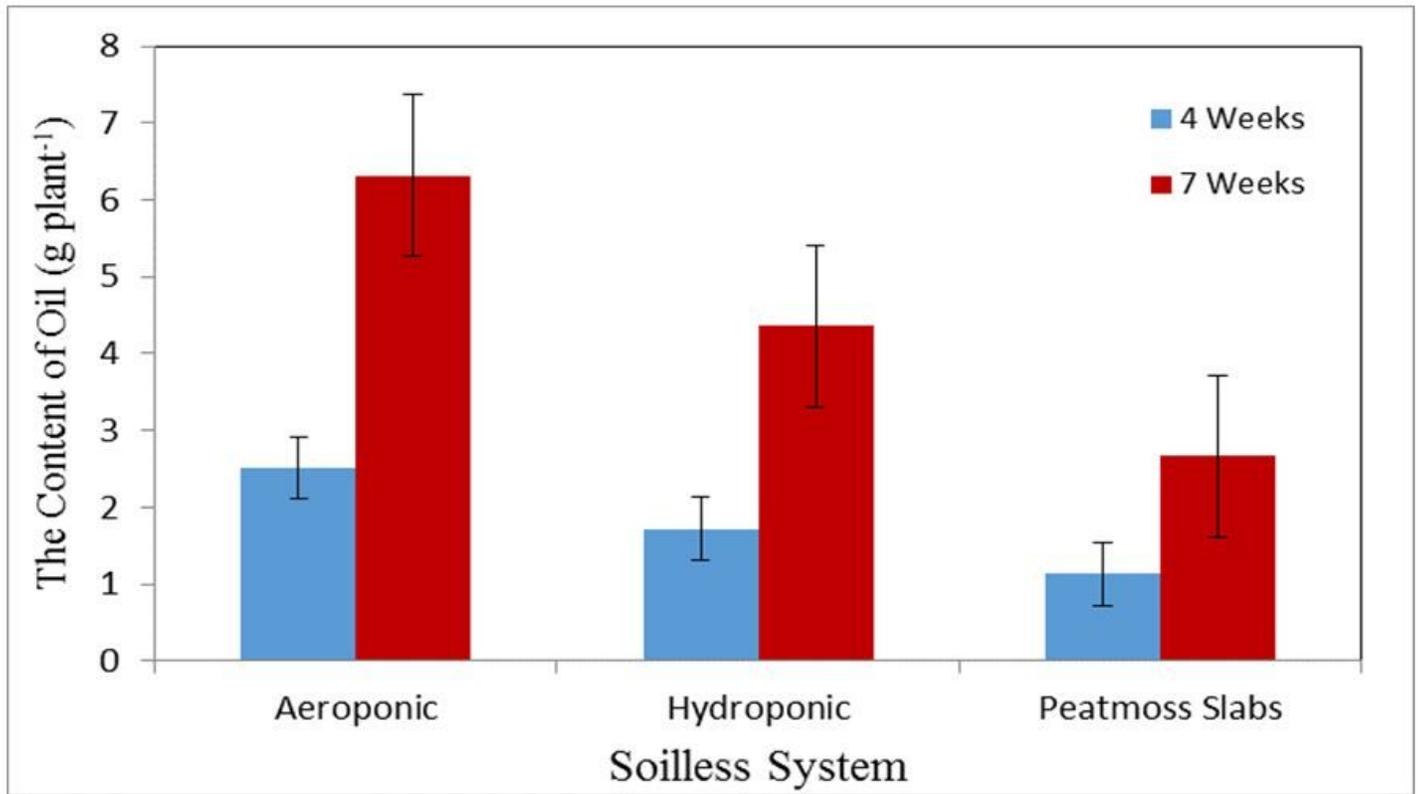
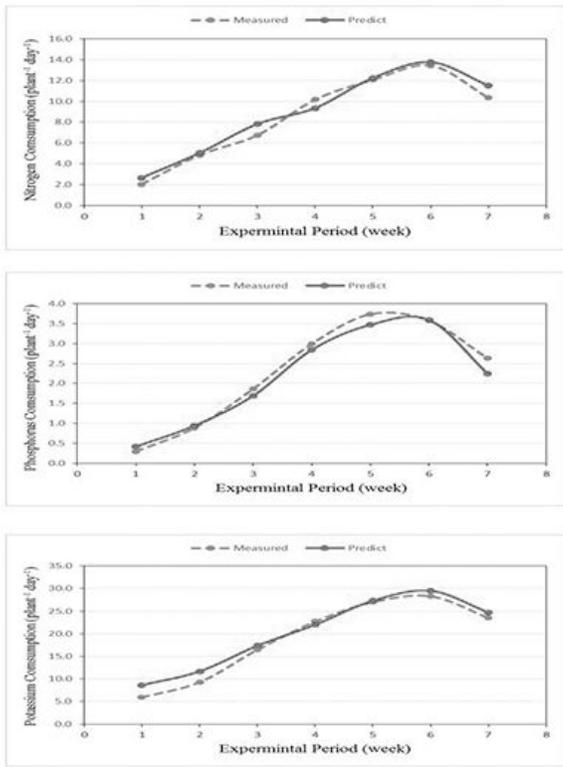


Figure 7

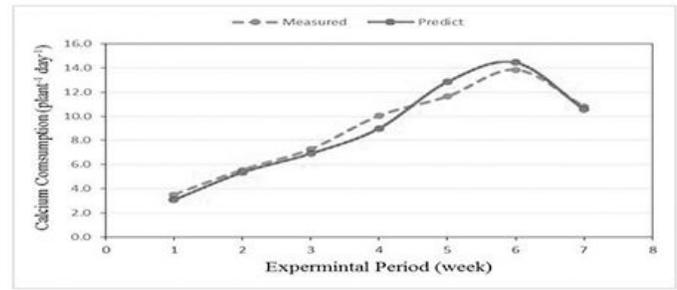
The basil oil content grown in different soilless systems.



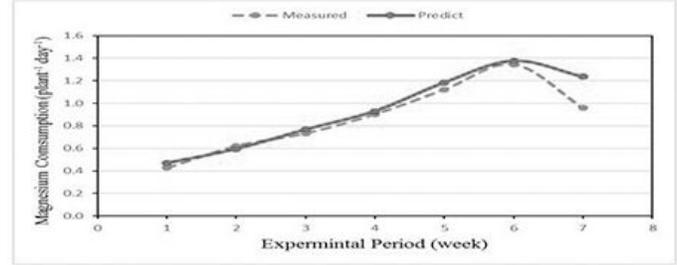
a

b

c



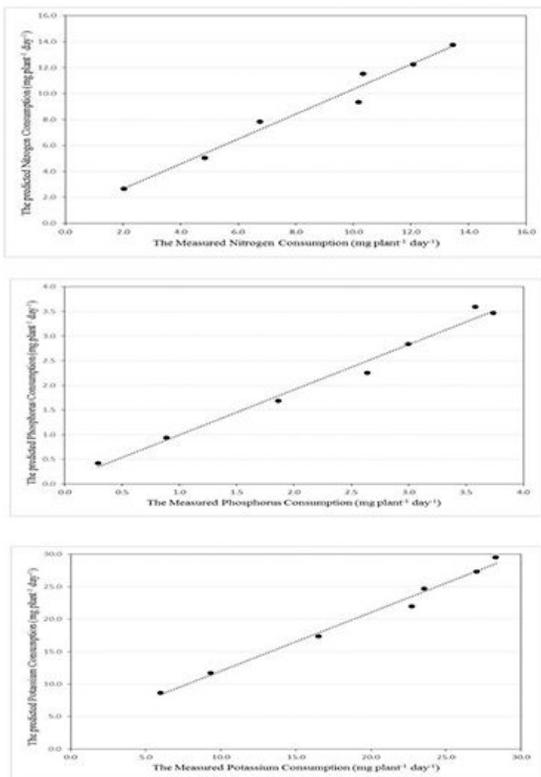
d



e

**Figure 8**

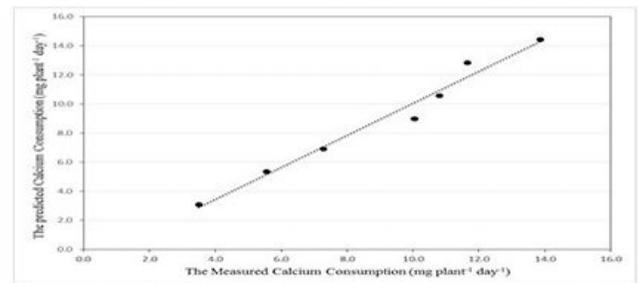
The predicted and the measured nutrients consumption by basil plants during the whole growth period. a: N b: P c: K d: Ca e: Mg



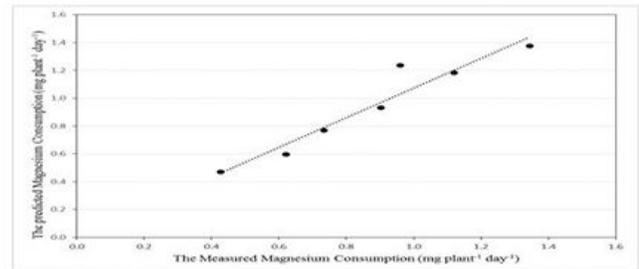
a

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d



e

**Figure 9**

The comparison between the predicted and the measured nutrients consumption by basil plants during the whole growth period. a: N b: P c: K d: Ca e: Mg