

The study of the performance of a biological fuel cell: A progress towards the improvement of low electrical bioenergy output by using an amplification system

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Abstract

A microbial fuel cell (MFC) has been conceived and constructed for the treatment of the sheep manure wastes and conversion them into clean sustainable renewable energy. The chemical oxygen demand (COD) removal of 10 days running MFC increased by 58.7% showing simultaneously an increase in the cell voltage. However, this MFC technology faces practical barriers since it produces low electric energy. A power management system including the MFC, operational amplifier, solar photovoltaic panel and a boost DC/DC converter, was elaborated in this respect. The low voltage output obtained from the MFC, was thus substantially increased by the amplifier prior to polarization by a photovoltaic module with solar radiation. The amplified voltage was thus sufficiently enough and in consequence, utilized to feed a light emitting diode. The low output voltage 0.5 V was simply harvested, successfully boosted up to approximately 2 V (i.e. 4 times higher) and effectively harnessed as a power supply. This novel application is very interesting to utilize the natural bioenergy contained in wastes to supply small electronic devices.

Highlights

- 1-Feasibility of power generation from animal waste using microbial fuel cell
- 2-Bioelectricity producing bacterial mixed consortia acquired from bio-anode polarization
- 3-Power generation using animal waste substrate with low cost electrodes
- 4-Mixed culture considered as cost-effective and environmentally sustainable process
- 5-Amplification and management of low voltage output by using common emitter amplifier polarized with a photovoltaic panel
- 6- MFC power amplification utilized to feed small electronic devices

Introduction

Microbial fuel cell (MFC) represents an eco-friendly approach to generating electricity while purifying [wastewater](#) concurrently, permitting [chemical oxygen demand](#) removal and electric power densities (Obileke et al. 2021). The system utilizes the metabolism power of bacteria for electricity generation, and application of [microorganisms](#) as available and inexpensive [biocatalysts](#) (Khajeh et al. 2020). The basic information and description of this novel technology are widely reviewed in the literature. However, scholars and researchers wish to know the problem of low power management and efforts deployed to increase it. Although there have been great advances in improving the [performance of the MFC by using bacterial natural metabolisms](#) (Adekunle et al. 2016), the low voltage output produced by such device seems to be the main drawback of this technology. For this reason, reliable system to make it beneficial on the industrial scale have been considered. There are thus two ways to undertake for increasing the low

power output of a MFC: the modification of electrodes and the use of amplification/management systems.

Recently, it has been proven that commercial application of MFCs can be increased through optimization of microorganism and invention of novel electrodes, which provide a promising option for cost effective bioelectricity generation (Choudhury et al. 2017). Really, due to low electricity generation performance and high cost of operation, the electrodes of a single-chamber air-cathode MFC have been modified with graphene and polymer polyaniline and then their effects on its performance have been assessed positively. Indeed, the modified electrode displayed higher catalytic activity toward oxygen reduction compared with unmodified electrodes (more than 6 times higher) (Wang et al. 2018). Furthermore, Fan *et al.* studied the effects of the polymer polypyrrole/iron(III) oxide composite modified anode on the electricity generation performance (i.e. the steady-state current density) and the sewage treatment capacity performance (i.e. higher rate of chemical oxygen demand removal) (Fan et al. 2021). Khajeh et al. have also made an investigation on the efficient improvement of a MFC performance by the modification of graphite cathode via electrophoretic deposition of copper/zinc oxide nanoparticles. The MFC performance was evaluated with and without visible light irradiation. A maximum voltage was achieved by the modified graphite electrode under the irradiation (Khajeh et al. 2020).

Although there have been a great deal of research towards the improvement of power outputs of MFCs, by modifying their electrodes and optimizing the electro active biofilms (biocatalysts), there still a drawback in using this technology in practical applications. For this reason, other researchers and scholar scientists have been focusing on the study the substantial increase of the output voltages of MFCs. Up to now, several MFCs were connected in series or in parallel to overcome the low voltage. Nevertheless, although a serially stacked MFCs unit could provide a higher voltage, it is ineffective due to voltage reversal, leading to a significant overall voltage decay (Kim et al. 2019). Moreover, much effort have been deployed to control the voltage reversal occurrence by connecting individual MFC units with a power point tracking system to charge a stacked polarized capacitor (Papaharalabos et al. 2017). This procedure allowed a maximum voltage which was fair enough to drive only low voltage electronic devices but not for real applications.

As a matter of fact, the power management unit (PMU) utilizing a combination of a DC/DC converter to boost the low MFC voltage to practical uses and a super capacitor to store electrical energy temporarily, has been proposed (Garita-Meza et al. 2018). As a result, a considerable boosted voltage was obtained from a wastewater single-chamber air-cathode MFC equipped with a low voltage booster (Koffi 2019). It was also proposed a new two-step "boost-and-multiply" system, in which the low output voltage was firstly boosted into an alternating current (AC) voltage by a transistor-based self-oscillating low voltage booster circuit. After, the boosted AC voltage was further multiplied and turned back into a direct current (DC) voltage by a multistage Single-Phase Cockroft-Walton voltage multiplier circuit. This newly designed low voltage booster multiplier was tested successfully and yield yielded higher voltage using a single-chamber air-cathode MFC treating domestic wastewater as a power source (Koffi et al. 2020). Specific converter topologies were also required to step-up in another manner the output voltage of a MFC using

another PMU for operation at low input voltage and at very low power in a completely autonomous way to capture energy from MFCs with the highest possible efficiency (Khaled et al. 2016). The power obtained was amply sufficient to supply a low-power temperature sensor.

To the best of our knowledge, there is no previous study on the amplification of the low voltage delivered by a MFC. In the present investigation, we have therefore fixed the goal to improve the performance of the MFC by amplifying its low output voltage in order to feed a Light Emitting Diode (LED). The amplification has been achieved with two electrical circuits by proposing the polarization of the operational with the conventional energy or that of solar radiation using photovoltaic panels. Furthermore, our PMU has been set up with a manner which quite different from that suggested recently by Koffi *et al*/ without utilizing any alternating current (AC) (Koffi 2019). A one-chamber MFC was thus inoculated and powered with leachate of sheep manure wastes, rich of EA bacterial source, that promises a sustainable MFC system. In the beginning, we studied the parameters useful for MFC conception, i.e. electron-generation, EA bacterial biofilm, power generation, and cell configuration. Then, we described all materials used and experimental procedure for energy harvesting, in particular the amplification of low energy generation. Finally, we presented the exploitation of the low bioenergy produced by the MFC, to feed for example the low electric input device LED. This has been achieved successfully by proposing the suitable amplification scheme.

Organic substrate degradation using bacterial metabolisms

As most organic substrates undergo combustion with the evolution of energy, the bio-catalyzed oxidation of organic substances carried out by means of microorganisms in the presence of O₂ at electrodes, permits the conversion of chemical energy into electric energy. In normal microbial catabolism, an organic substrate is oxidized initially without involvement of O₂, while its electrons are taken up by an enzyme-active site, which acts as a reduced intermediate.

Bioelectric power generation

In the MFC, the catalytic microorganisms oxidize the organic matter to produce electrical energy. They shuttle the electrons exogenously to the electrode surface without utilizing artificial mediators, are referred as exoelectrogens. The transfer of electrons, whether direct or indirect, takes place at the end of a cascade of oxidation and reduction reactions, considered as the keys to the catabolic processes necessary for the survival of the bacteria. These processes are the natural metabolisms of fermentation and cellular respiration. These latter are used by the bacteria to break down the organic matter (waste) and extract energy from it which will be stored in the form of ATP considered as a universal source of energy.

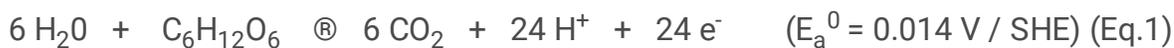
As illustrated in Fig. 1, the biodegradation of the organic substrate has been achieved by using the electrocatalytic ability of the bacteria. The electric energy can thus be generated following the two extracellular electron transfer pathways which are the trans Plasma Membrane Electron Transport (tPMET) and Electron Transport Chain (ETC) (Schaetzle et al. 2008). Both pathways allow the generation

of the electrons which go out through the bacterial cell. The electro catalytic oxidation of glucose can be performed thanks to NADH oxidation (Mardiana et al. 2015). Electrons are therefore produced and captured by the species under oxidized form. In anaerobic fermentation pathway, the recycling of NADH to NAD⁺ is important to keep the glycolysis process continuous (Nelson 2005)]. In all cases, the process results in the generation of electrons, protons and carbon dioxide. In this way, the substrate is biodegraded and the electric power is produced. Furthermore, the microbial growth is supported by the energy derived from the electron transfer process itself and results in a stable, long-term power production

The degradation of the organic matter (grape juice) by the microbial metabolism (fermentation), in the absence of oxygen, produces on one hand the energy which is stocked by the bacteria in the form of ATP, and on the other hand the electric energy recovered by the final electron acceptors (bio anode). This latter gives rise to an electric current. However, the bacteria continue to degrade by oxidation process the organic matter contained in the polluted environment by reducing its COD (chemical oxygen demand) without really using an external substrate (glucose). Microorganisms (bacteria, yeasts, etc.) can oxidize a wide variety of organic molecules (substrate), and thus produced electrons which are transformed into the energy useful for their growth and their metabolism.

Principle of voltage generation in MFC

The microbial generated electron can be harnessed by artificially introducing an adequate electrode (i.e an electron acceptor) into the MFC. This allows the stimulation of the development of the potential difference between the bacterial membrane and the anode, which contributes to the elaboration of the bio anode potential that helps in the electron delivery. The electrons that are drawn to the anode induce positive potential of the cathode. Thus, the potential difference between the positive cathode and the negative bio anode, is called the electromotive force or the cell voltage. This potential drives the generated electrons from the anode to the cathode through the external circuit across the load, that can be harvested as electricity (i.e. the electric current flows in the opposite way from the cathode to the anode). For example, the enzymatic-catalyzed oxidation of glucose (fuel) on the anode yields the electric potential versus standard hydrogen electrode (SHE) according to the reaction (Scott et al. 2012).



Whilst the reduction of the oxidant at the cathode proceeds according to the reaction



Experimental

Materials

Biocatalyst (sheep manure compost)

The biocatalyst of an MFC is generally the microorganisms, which adhere to the electrodes while performing the role of electro-catalyst. They come from a pure culture, mixed or a natural consortium. They are called anodophiles, exoelectrogens and electrochemically active bacteria (Lovely 2008).

As reviewed previously, the activity of bacterial growth in the sea sand/animal manure or sludge mixtures has been proved experimentally (El-Nahhal et al. 2020). In effect, it was thus clearly reported that *Bacillus* species in such wastes, can be isolated from all manure types. Nearly three bacterial species were characterized in the animal manure. They were grown under aerobic conditions and actively participate in the production of electricity. The microbial community was therefore responsible for the transfer of electrons to electrodes through electron transport chain during the oxidation reduction reactions that took place in the bio electrochemical fuel cell. It was moreover revealed the role of bacteria in harvesting electricity. After electricity generation, the manures had lower chemical oxygen demand due to consumption by bacterial community Alkhamis et al. 2021). In addition, owing to its potential bioenergy production by underground burial system heated with cascade-controlled solar water heated system, the sheep manure was used as an indicator of biomass potential contribution to power generation in less fossil energy countries (Zhang et al. 2012).

On this basis, the sheep manure was thus chosen by us in this study to test its potential electricity generation in a MFC for the production of energy and concomitant preservation of environment. So, a quantity of bulk sheep manure was collected directly from a sheep barn located in the town suburb farm. The manure samples were finely grinded and sieved to obtain very small particles in powder form. This latter was then mixed with potassium chloride solution (60 mM) to make the medium conducting for ionic motion of species obtained by oxidation of the organic matter. The mixture was after stirred for 24h and then filtered through a paper filter. The obtained manure sheep leachate was thus obtained and put inside a plastic cup under room temperature. Three forms of the waste manure were tested in the present study: solid (compost), sludge and leachate. The bio anode formed in the manure compost has shown efficient electro active properties with the grape juice (substrate) which readily oxidized in the presence of this biocatalyst. It has thus revealed a rich microbial flora with remarkable electro active ability, as it has been already reported in the literature (Parot et al. 2009).

Substrate (grape juice)

The fuel must be a biodegradable compound and not toxic to microorganisms. The first studies were focused on organic compounds of low molecular weight such as glucose (Chauduri et al. 2003). Later, complex fuels were tested such as cellulose (Schröder et al. 2005). Finally, studies have been developed on the possibility of using domestic or industrial waste (Liu et al. 2004). Complex industrial wastes are generally diluted to avoid undesirable planktonic metabolisms that may compete with the biofilm-catalyzed electricity generation and to decrease fouling on electrode and membrane surfaces as well (Cercado-Quezada et al. 2010).

As a matter of fact, in the present investigation, the grape juice was consequently tested, which generally represent most favorable environmental condition for microorganisms. Diverse metabolites such as

formate, succinate, lactate, acetate and propionate, are produced during glucose oxidation in MFC due to its fermentative nature but acetate has been identified as the dominant and most effectively utilized (Kim et al. 2011). As shown below, when it is oxidized in the presence of the appropriate catalyst (chemical or biological), the substrate glucose main constituent of the grape juice, releases electrons, protons and carbon dioxide.

Methods

Set Up of MFC

The one-compartment MFC reactor was used to generate electricity (Fig. 2). It was fully filled with leachate of sheep manure wastes. The anode and cathode electrodes made of carbon graphite (CG) were inserted inside the chamber, which was tightly closed with plastic paper to prevent penetration of oxygen and thus avoiding unwanted oxidation of present species. The two electrodes were then connected with an external of 1000 Ω , to allow the natural polarization and thus the growth of the EA biofilm. While using the sheep manure as the inoculum waste in the MFC, El-Nahhal *et al* described in details the mechanism of electricity generation (electrochemical potentials of both electrodes and biodegradation of organic matter), oxidation of inorganic molecules and electron transport chain reactions (El-Nahhal et al. 2020). The cell used in this study was operated in a batch mode with view to utilize it in future in the continuous mode for a real application. The generated voltage (V) was monitored using a digital voltmeter. The value of the electric current (I) was thus obtained by taking the ratio between the measured voltage and the discharged resistance (R). Moreover, in order to assess the performance of the MFC, the current density (mA/cm^2) was calculated by dividing the electric current by the projected area. The fluctuation in the current density response can be explained by the change in the bacterial population distribution more fermentative or electrogenic culture. Nevertheless, when the voltage of the MFC declined, 5 mL of commercial red grape juice of simple sugars concentration (i.e. 113 g/L) was added to the leachate to feed the microorganisms which resulted in the generation of electrons and an increase in the current density. The MFC started functioning since last summer and it is still working up to now as long as the electrodes are preserved from corrosion and correctly connected to the discharge resistance.

Chemical Oxygen Demand (COD)

COD is usually used as a measure of organic pollutants present in wastewater and it is expressed in (mg/L). The most common application of COD is in quantifying the amount and level of oxidizable pollutants found in the wastewater. It makes it possible to determine the concentration of organic matter in the wastewater through the quantity of oxygen necessary for their total chemical oxidation. A COD analysis was thus performed by us at the end of the MFC experiment to make in evidence the conversion of the polluted organic matter into bioenergy. It was measured according to the ISO 15705 standard (ISO 15705. 2002) based on the closed tube small scale method. The tubes were heated at 148° C for 2 hours. Then, the samples were left to cool down for 1 hour and diluted with distilled water. After that, both the blank (distilled water) and the experimental sample were analyzed with Mohr's salt solution (0.1 N) by titration

using the colored indicator (Ferroine). Then, the COD values (mg.L^{-1}) were calculated by using Eq.3 and subsequently, the corresponding COD removals were, therefore tabulated by using Eq. 4.

$$\text{COD} = (V_1 - V_2) \times 8000 \times \frac{N_{\text{MS}}}{V_3} \quad (\text{Eq.3})$$

$$\text{COD}_{\text{removal}} (\%) = 100 \times (\text{COD}_i - \text{COD}_f) / \text{COD}_i \quad (\text{Eq.4})$$

where

V_1 -volume (mL) of Mohr's salt solution (titrating solution) used for the blank test with distilled water.

V_2 -volume (mL) of Mohr's salt solution (titrating solution).

V_3 -volume (3 mL) of the sample (titrated solution)

N_{MS} - normality of Mohr's salt solution (0.1 N).

COD_i and COD_f are initial and final COD values respectively

Amplification of low voltage output

Following previous work carried out by Ridvan Umaz who studied the power management system of the low power obtained from MFCs (Umaz et al. 2020). The proposed system was comprised of a fronted converter, a super capacitor, a charge control circuit and a second converter. In the fronted converter, the maximum power extraction from a MFC can be achieved. In our present investigation, we thus carried on another procedure by harvesting the maximum output voltage by using amplifying system polarized by a solar photovoltaic panel. Although the MFC can produce renewable energy from wastes, the generated power is practically unusable. However, in order to extract usable power from it, we have used two amplifiers: operational amplifier and common emitter amplifier.

Operational Amplifier (OA)

An operational amplifier is a high gain differential electronic device. It is used to greatly amplify an electric potential difference present at its inputs. Initially, the amplifier was designed to perform mathematical operations in analog calculators: it easily implemented basic mathematical operations such as addition, subtraction, integration, derivation and others. Subsequently, it was used in other applications such as the control of electric motors, voltage regulation, current sources or even oscillators.

As depicted in Fig. 3a, we have a positive input signal which becomes negative at output. The gain (G) of the assembly is fixed by the value of the resistors. In effect, in the mesh 1, $V_{\text{in}} = R_1 I_1 + \varepsilon$. So for ε small enough, we get $V_{\text{in}} = R_1 I_1$. The same reasoning applies for the mesh 2, but this time with a negative sign due to the inversion between the input and the output in this amplification system. Indeed, $V_{\text{out}} = (-R_2 I_2)$. This results in the gain which is the ratio between the two voltages. For example, for $R_1 = 100 \Omega$

and $R_2 = 3000 \Omega$, the gain is equal to (-30). The negative sign highlights the amplification reversal system between the input and the output.

Common Emitter Amplifier (CEA)

As illustrated in Fig. 3b, a common emitter amplifier assembly is one of the three elementary configurations for amplifying a bipolar transistor. The other two are called common base or common collector. In this assembly, the emitter (indicated by an arrow) is connected to the common ground or to a reference voltage, while the base of the transistor is connected to the input and the collector to the output load. The analog circuit using a field effect transistor is called a common source amplifier circuit.

The common-emitter amplifier circuit consists of a C_e decoupling capacitor which is chosen to have a low enough impedance, when studied in small signals, to short-circuit the resistor R_e . The presence of C_e makes it possible to significantly increase the gain of the assembly, but on the other hand the circuit has lower input and output impedances. The emitter resistor R_e therefore makes it possible to create a feedback called degeneration of the emitter, which ensures good characteristics of stability and linearity of the circuit, in particular in response to temperature variations. In the case where C_e is absent, the impedance of R_e reduces the overall trans conductance $G_m = g_m$ of the circuit by a factor $(g_m \cdot R_e + 1)$. Thus the gain in tension is expressed by:

An electronic amplifier is used to amplify AC signals with a very large amplification gain. However, in the microbial fuel cell, it can be applied to increase the low voltages, which are variable over time, in order to use them to power small electronic devices such as LED. The active components used in electronic amplifiers, in particular the transistor, make it possible to control their output current as a function of an electrical quantity (current or voltage), the image of the signal to be amplified. The output current of these active components is taken directly from the amplifier power supply. It can be, however, replaced by a mixed association (series/ parallel) of a set of MFCs. Depending on how they are installed in the amplifier, the active components thus make it possible to increase the voltage and/or the current of the input electrical signal.

Direct current converter

As it is commonly well known, the voltage harvested from the MFC is relatively low (less than 0.5 V) and requires thus an electric management system to step-up the low voltage to acceptable voltage values (i.e. > 1.0 V). This management system contains in general, a charge pump or a direct current (DC) converter to boost the voltage and capacitors to store the energy (Donovan et al. 2011). The DC converter utilized in the present investigation is illustrated in Fig. 4.

Results

Evolution of MFC voltage in open circuit

The carbon graphite electrode was inoculated with leachate of sheep manure waste to build a bio anode. This latter has been set up as soon as the MFC has been started. At the beginning, most of bacteria were still present in planktonic state in the solution. Overtime, the bacteria start to colonize on the anode surface and form a biofilm. As shown in Fig. 2, the colony grows larger and therefore the biofilm becomes visible. With a pure strain such as *Escherichia coli*, the performance is much better (Mukhtar et al. 1914). However, the system is very expensive. Moreover, as the MFC is discharged to an electric load (resistance), the self-developed current contributes to the EA biofilm development and consequently the fuel cell is thus polarized naturally. Previous studies reported the formation of the EA biofilm, once the bio anode was polarized by applying a constant stimulating potential (Ojima et al. 2020).

Fig. 5 shows a plot of the voltage generation of the MFC versus time. The generated voltage expresses the potential difference between the CG bio anode and the stainless steel plate cathode both dipped in the same chamber containing the leachate of the sheep manure wastes. In the beginning, the observed voltage is very low because the potential difference between the bare CG anode and the cathode is narrow. Along with the biofilm growth, we observe an increase in voltage generation and stability of the MFC (Kosimaningrum. 2018). As a result, the potential difference between the bio anode and the cathode becomes wider, mainly due to the formation of EA biofilm on the anode surface. Biofilm attachment involves extracellular polymeric substances (EPS) that can facilitate electron transfer from the bacterial cell to the anode (Zhang. 2008). These substances surround the cell by acting as transient media in the extracellular electron transfer (EET) process. Xiao, *et al.* suggested the electron hopping to be the molecular mechanism of the (EET) through (EPS) (Zhang et al. 2014).

As soon as the EA biofilm was built up and agglomerated around the anode, it catalyzed the oxidation of the organic matter in the leachate inside the MFC chamber; thus, the voltage increased steadily and reached the value 650 mV. Then, it declined as the substrate was consumed around the bio anode. However, upon the addition of substrate (5 mL of grape juice), the voltage of the MFC rose up again drastically and attained its highest value i.e. 850 mV. Regardless the efficiency of the MFC, after a long working period, the voltage decreased naturally, because of the depletion of the substrate and the nutriment in the leachate, leading to lower activity of the bacterial consortium.

Polarization and power curves

As shown in Fig. 6, the polarization curve presents regions expressing three types of loss: 1) activation loss due to activation polarization; 2) ohmic loss due to polarization; and 3) concentration loss. At low current density, the potential drops drastically, due to energy loss for oxidation and reduction initiations at anode and cathode respectively, as well as energy loss during electron transfer from the bacterial cell membrane to electrode surface.

At intermediate current density, the ohmic loss originates mainly from the higher resistance of ions to travel inside the leachate (low ionic conductivity), and the electrical connection between the two electrodes. However, in order to reduce the ohmic loss, one should reduce the electrodes spacing and increasing the ionic conductivity of the electrolyte. Whilst at higher current density, the cell performance

falls sharply, owing to the mass transport limit of the reactants towards the electrode and subsequently, the products out of the electrode (Lin et al. 2015). The polarization curve was obtained when the cell was connected to an external variable electric resistance in the decreasing order from the highest resistance 10 M Ω down to lowest resistance 1 Ω), avoiding unwanted rapid discharge of the MFC. The voltage produced on the load, was measured with a multimeter of higher input resistance, in order to minimize any unwanted current flow in the voltmeter. The resulting current was therefore calculated by using Ohm's law. As shown in the figure, the MFC presents an open circuit voltage (V_{OC}) of 484 mV (i.e. $R=\infty$) and a short circuit current (I_{SCC}) of 2611 mA/cm² (i.e. $R=0$).

Besides, the power production is one of the key parameters to determine its performance. The output power was determined from the measured MFC's voltage and the resulting delivered current. It determines the maximum power point (MPP) that can be supplied by the MFC at a specific electric resistance (Logan. 2007). At this point, the load resistance can be deduced and should be placed permanently to yield the highest output. Adapting a MFC to the power of the respective load plays a crucial role in practical applications. Whenever possible, the power of the load should always be close to the MPP (Heliocentris GmbH. 2000). Table 1 summarizes the most important values obtained with the MFC after having worked for 140 days.

Applied potential to formation of EA biofilm on bio anode

The efficiency of the EA biofilm is characterized by the maximum current density of projected bio anode area and by the faradic yield, it achieves. In effect, an anodic EA biofilm performs well when it is capable of producing a high current density at the lowest possible potential. Thus, the choice of the potential to be imposed on the bio anode generally depends on two major objectives: the increase of the chance of obtaining an EA biofilm and the selection of the most effective one.

It has been described previously that in the same reactor, when microorganisms compete for the consumption of the organic substrate, the biofilm formed under negative polarization compared to the standard hydrogen electrode, made it possible to obtain better performance (Torres et al. 2009). In addition, it has also been shown that imposing a very negative potential for a long period allows the evolution of a pure culture of *G. Sulfurreducens* towards a more efficient strain with a density of current produced more than 5 times greater than that of the starting strain (Yi et al. 2009). The authors thus show that the negative potential acts as a selective pressure towards the evolution of a more efficient strain. On the other hand, it has been made in evidence that a low negative potential, despite a lower quantity of biomass produced, allowed maximum powers to be obtained much more quickly than higher potentials (Aelterman et al. 2008).

Thus, on this basis of these considerations, we have polarized the bio anode of our MFC with positive and negative potentials successively. The value of this polarizing potential was +/- 0.6 V which is quite close to the RedOx potential of CO₂/Glucose (i.e.- 0.43 V/HSE); Glucose being the main constituent of the grape juice used by us as substrate to feed the existing bacteria in the sheep manure inoculum. As shown

in Fig. 7, the evolution curve of the MFC discharged to the resistance 1000 Ω , decreases sharply with time during the period of 8 hours, reaching the lowest voltage with the MFC polarized positively. The voltage decreases from 400 mV down to less than 1 mV. In contrast, the voltage of the MFC polarized negatively, started with 300 mV and lowers as well but stabilizes readily to a constant value, which is greater (i.e. 155 mV). However, the voltage of the MFC without prior polarization, lies between.

During the positive polarization, as the current arrived to the bio anode, the electrons were pumped out from the EA biofilm and as a result, the voltage of the MFC dropped to lower values. In contrast, during the negative polarization, as the current left out the bio anode, the electrons were added massively to the EA biofilm, inducing an increase of the voltage. This result is in quite agreement with that reported by Ketep *et al.* while they studied the lowering of the applied potential during successive scratching/re-inoculation for the improvement of the performance of microbial anodes for microbial fuel cells. The microbial anodes were formed under polarization at -0.2 V/SCE on smooth graphite plate electrodes with paper mill effluents (Ketep et al. 2013). The effect of anode polarization on biofilm formation and electron transfer in *Shewanella oneidensis* /graphite felt microbial fuel cells, supports also the results obtained in the present study (Pinto et al. 2018).

COD removal

Amongst other applications, the MFC technology allows maximum COD removal and its conversion into bio-energy in a treatment plant. We have adopted this application in the present work, in order to establish the relationship between the production of the bioenergy and the depollution of the medium by assessing the COD removal. The MFC was continuously operated for 10 days, achieving a COD removal of 58.7% with a pH ranging between 7 and 8, similar to those observed in typical biological wastewater treatment processes. However, the COD removal would have been higher, if the MFC had been left working longer. Nevertheless, the overall efficiency observed for the COD removal confirms the effective wastewater treatment process using the MFC technology. Moreover, the increase in the COD removal is well correlated with an increase in the cell voltage, making in evidence the conversion of the biomass into the green eco-electric energy. These findings revealed the anaerobic development of the microorganisms, which were electrochemically active. The same behavior of the COD removal has been observed previously, by studying the effects of the resistance load, the geometry and the design of the MFC (Lee et al. 2016), as well as other effects such as electrodes modification and biofilm scratching. In general, at the end of the experiment, the color of the leachate of the sheep manure inside the MFC, became very much less darker, almost clear, which means that the maximum COD has been effectively removed, yielding sufficient bio-electrochemical energy conversion. It resulted in an electric current, which was produced after having grown an electro active biofilm that caused the reduction of the COD.

Discussion

Amplification of low voltage output

In order to extract usable power from it, we have used two amplifiers: operational amplifier and common emitter amplifier. The low output MFC voltage (ca. 0.5 V) was successfully boosted up to 1.9 V, which was the highest voltage that has been ever reported. Moreover, the boosted voltage was stably maintained for more than 2 hours. The energy harvesting efficiency increases as the polarization of the amplifier is utilized. These results clearly suggest that the proposed system is an efficient and self-starting energy harvester and storage for low-power generating MFCs. This finding is consistent with another renewable system that concerns the high voltage generation from wastewater by MFCs equipped with newly designed low voltage booster multiplier (Nguyen et al. 2019).

As shown in Fig. 8, the low voltage of the single-compartment MFC was exploited by amplification by using an amplifier polarized with the photovoltaic solar module of characteristics given in appendix 1 (Table 2), for the improvement of output voltage. The low voltage of the single-compartment MFC was exploited by amplification by undertaking five different ways. In the first case, as shown in Fig. 9, the voltage 0.5 V delivered by the MFC was amplified with the operational amplifier, but unfortunately it generated a low current of 160 μA which lasted just 20 s with a drastic drop in the input voltage to the end (a-a'). On the other hand, the series association of four MFCs made it possible to increase the voltage substantially to 1.64 V, but as before the current drawn was always very low (14 s) and the amplification time was shorter (b-b'). Whilst, the parallel association of the four MFCs made it possible to maintain the average voltage of 0.5 V overall, but the current was increased relatively to 650 μA (c-c'). The advantage of this amplification lies in its long operating time (65 min). Finally, the polarization voltage of the amplifier was biased with 3 V using a current generator in our case it can be replaced by that of microbial fuel cells associated in series and in parallel (d-d'). Thus, it was restarted by delivering a voltage a little higher (0.65 V) but by delivering the same current 650 μA . This polarization therefore results in an increase in the duration of amplification (120 min).

Furthermore, the common emitter amplifier was biased with a photovoltaic panel powered with solar radiation (Fig. 10). The first test was carried out at the end of the day where the LED was illuminated for 135 minutes. It went off when the sun set down completely. However, The LED could remain illuminated as long as there is lightening, in particular solar radiation in our case. So, from a renewable energy point of view, this method is advantageous because the polarization of the common-emitter amplifier has been replaced by solar radiation. During the night, one is obliged to polarize with a current generator in order to keep LED on.

Integration of DC/DC Boost Converter for effective exploitation

Moreover, in order to remedy to the deficiency encountered when using only the amplifier alone, we have included another potential electronic device to harvest the maximum energy delivered by the MFC. The energy harvested from the MFC required thus a power management assembly to step-up the low voltage to acceptable level. It contained thus the power supply (MFC), the amplifier and in addition a DC/DC converter to boost the voltage (Fig. 11).

As a matter of fact, we have therefore thought of the design of the precision amplifier, in particular for continuous signals. We therefore looked for appropriate circuits with very low drift, because the smallest detectable signals are limited by the signals error created by the amplifier. Really, the amplification of AC signals, noise and risk of oscillation can be avoided by just short-circuiting DC errors with capacitive couplings. However, this decoupling is no longer possible if utilizing amplified continuous signals. In this case, the continuous errors due to parasite voltages and currents and their drifts are very restrictive. An optimized boost-type step-up voltage converter (DC/DC) is therefore necessary to increase the voltage delivered by the MFC. So in order to meet the requirements for the real applications, the common emitter amplifier and the DC/DC converter boost should be used, so that the voltage and current of the MFC can be increased.

Fig. 12 illustrates the variation in voltage used and the result of manual conversion using the pulses due to the position of the switch (S) of the boost converter. When the switch is closed (on state), the current in the inductor increases, and as a result, energy of magnetic origin is stored. The diode is therefore blocked, and the load is disconnected. On the other hand, when the switch is open, the inductance is then in series with the generator and its voltage is added to that of the generator: called the booster effect. This therefore results in a transfer of cumulative energy in the inductance to the capacitance.

Performance of MFCs after amplification

After having been tested successfully in powering the LED, the performance of MFCs has again been tested by plotting the power density curves. In Fig. 13, we compare the power density curves of the single-compartment MFC and the parallel association, after amplification. Whilst, in Fig. 14, we compare the MFCs with parallel and series associations successively. The figure reveals the enormous difference between them. Indeed, the parallel association yields the maximum power density with respect to the series association. Although the parallel association is quite beneficial in powering the LED, the stored energy was exhausted and in consequence, the MFCs stack requires a longer time to recover. Whilst, the series association yields higher voltage, but unfortunately lower current output. Table 3 summarizes the values of current and power densities of MFCs associated in series and in parallel after having been used in the amplification.

Conclusion

In this present investigation, the energy harvested from the MFC was demonstrated from the biodegradation of the sheep manure solid waste by using the bacterial catalyst. The output voltage of the MFC was substantially increased upon the addition of the GJ substrate (i.e. from 650 mV to 850 mV) making in evidence the electro activity of the biofilm. Really, the bio anode was much more effective in current production when it was polarized negatively (from 300 down to 155 mV) than positively (from 400 mV down to less than 1 mV). This allowed the possibility of adapting the microorganisms, rendering them more electro active and thus producing higher current densities. The COD removal of 10 days running MFC increased by 58.7% showing simultaneously an increase in the cell voltage. Furthermore, a

power management unit including the MFC and an operational amplifier was utilized to power supply a light emitting diode (LED). The initial relative low output voltage 0.5 V was successfully boosted up to approximately 2 V (i.e. 4 times higher) and effectively harnessed as a power supply. The amplifier was tested satisfactorily with parallel association of MFCs. However, both single stack and series association of MFCs were not able to achieve the required current for functioning the LED. The common emitter amplifier biased with a photovoltaic panel powered with solar radiation, allowed to feed the device as long as it was illuminated. This application is very interesting and promising to utilize the green and natural energy in the treatment and recovery of wastes, to supply monitoring and measuring systems.

Abbreviations

Microbial Fuel Cell (MFC)

Direct Current (DC)

Light Emitting Diode (LED)

Adenosine TriPhosphate (ATP)

trans Plasma Membrane Electron Transport (tPMET)

Electron Transport Chain (ETC)

Nicotinamide Adenine Dinucleotide Hydrogen (NADH)

Carbon Graphite (CG)

Operational Amplifier (OA)

Common Emitter Amplifier (CEA)

Electro Active (EA)

Extracellular Polymeric Substances (EPS)

Extracellular Electron Transfer (EET)

Open Circuit Voltage (V_{OC})

Short Circuit Current (I_{SCC})

Maximum Power Point (MPP)

Alternating Current (AC)

Reduction/Oxidation (RedOx)

Saturated Calomel Electrode (SCE)

Grape Juice (GJ)

Chemical Oxygen Demand (COD)

Standard Hydrogen Electrode (SHE)

Declarations

Author Disclosure Statement

The authors must disclose any financial and personal relationships with other people or organizations that could inappropriately influence their work.

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References

1. Obileke KC, Onyeaka H, Meyer EL, Nwokolo N (2021) Microbial fuel cells, a renewable energy technology for bio-electricity generation: A mini-review. *Electrochem commun* 125:107003. <https://doi.org/10.1016/j.elecom.2021.107003>
2. Khajeh RT, Aber S, Nofouzi K (2020) Efficient improvement of microbial fuel cell performance by the modification of graphite cathode via electrophoretic deposition of CuO/ZnO. 240:122208. <https://doi.org/10.1016/j.matchemphys.2019.122208>. *Mater. Chem. Phys.*
3. Adekunle A, Garipey Y, Lyew D, Raghavan V (2016) Energy recovery from cassava peels in a single-chamber microbial fuel cell. *Energ Source Part A* 38(17):2495–2502. <https://doi.org/10.1080/15567036.2015.1086909>

4. Choudhury P, Uday USP, Bandyopadhyay TK, Ray RN, Bhunia B (2017) Performance improvement of microbial fuel cell (MFC) using suitable electrode and Bioengineered organisms: A review. *Bioeng* 8(5):471–487. DOI: 10.1080/21655979.2016.1267883
5. Wang Y, Wu J, Yang S, Li H, Li X (2018) Electrode Modification and Optimization in Air-Cathode Single-Chamber Microbial Fuel Cells. *Int J Environ Res Public Health* 15:1349. <https://doi.org/10.3390/ijerph15071349>
6. Fan L, Xi Y (2021) Effect of Polypyrrole-Fe₃O₄ Composite Modified Anode and Its Electrodeposition Time on the Performance of Microbial Fuel Cells. *Energies* 14:2461. <https://doi.org/10.3390/en14092461>
7. Kim T, Yeo J, Yang Y, Kang S, Paek Y, Kwon JK, Jang JK (2019) Boosting voltage without electrochemical degradation using energy-harvesting circuits and power management system-coupled multiple microbial fuel cells. *J Power Sources* 410–411:171–178. <https://doi.org/10.1016/j.jpowsour.2018.11.010>
8. Papaharalabos G, Stinchcombe A, Horsfield I, Melhuish C, Greenman J, Ieropoulos I (2017) Autonomous energy harvesting and prevention of cell reversal in MFC stacks. *J Electrochem Soc* 164:H3047–H3051. DOI: 10.1149/2.0081703jes
9. Garita-Meza MA, Ramírez-Balderas LA, Contreras-Bustos R, Chávez-Ramírez AU, Cercado B (2018) Blocking oscillator-based electronic circuit to harvest and boost the voltage produced by a compost-based microbial fuel cell stack. *Sustain Energy Technol. Assess.* 29:164–170. 10.1016/j.seta.2018.08.007. **DOI**
10. Koffi NJ (2019) Domestic wastewater treatment and electricity recovery by a PVDF-based air-cathode MFC coupled with a low voltage booster multiplier, PhD thesis, Hokkaido University. https://eprints.lib.hokudai.ac.jp/dspace/bitstream/2115/80053/1/NDah_Joel_KOFFI.pdf
11. Koffi NJ, Okabe S (2020) High voltage generation from wastewater by microbial fuel cells equipped with a newly designed low voltage booster multiplier (LVBM). *Sci Rep.* 10(1):18985. 10.1038/s41598-020-75916-7. **doi**
12. Khaled F, Ondel O, Allard B (2016) Microbial fuel cells as power supply of a low-power temperature sensor. *J Power Sources* 306:354–360. <https://doi.org/10.1016/j.jpowsour.2015.12.040>
13. Schaetzle O, Barriere F, Baronian K (2008) Bacteria and yeasts as catalysts in microbial fuel cells: electron transfer from micro-organisms to electrodes for green electricity. *Energy Environ Sci* 1:607–620. DOI: 10.1039/b810642h
14. Mardiana U, Innocent C, Jarrar H, Cretin M, Buchari, Gandasasmita S (2015) Electropolymerized Neutral Red as Redox Mediator for Yeast Fuel Cell. *Int J Electrochem Sci* 10:8886–8898
15. Nelson DL, Cox MM (2005) *Principles of biochemistry*, 4th edn. W.H. Freeman, New York
16. Scott K, Yu EH, Ghangrekar B, Erable B, Duteanu NM (2012) Biological and microbial fuel cells. *Compr Renew Energy* 4:277–300. doi: 10.1016/B978-0-08-087872-0.000412-1
17. Lovely DR (2008) The micro electric: conversion of organic matter to electricity. *Curr Opin Biotech* 19:564–571. DOI: 10.1016/j.copbio.2008.10.005

18. El-Nahhal YZ, Al-Agha MR, El-Nahhal IY, Aila E, El-Nahal NA, Alhalabi FI, RA (2020) Electricity generation from animal manure. *Biomass Bioenergy* 136:105531. <https://doi.org/10.1016/j.biombioe.2020.105531>
19. Alkhamis TM, Alzoubi AI, Alma'atah BM (2021) Biogas Production from Sheep Manure by a Simulated Underground Burial System Heated with Cascade-Controlled Solar Water Heated System, as an Indicator of Biomass Potential Contribution to Power Mix in Jordan. *Journal of Environmental Protection* 12:125–140. <https://www.scirp.org/journal/jep>
20. Zhang GZ, Hao Q, Jiao Y, Wang K, Lee DJ, Rena N (2012) Biocathode microbial fuel cell for efficient electricity recovery from dairy manure. *Biosens Bioelectron* 31:537–543. <https://doi.org/10.1016/j.bios.2011.11.036>
21. Parot S, Nercessian O, Delia ML, Achouak W, Bergel A (2009) Electrochemical checking of aerobic isolates from electrochemically active biofilm formed in compost. *J Appl Microbiol* 106:1350–1359. doi.org/10.1111/j.1365-2672.2008.04103.x
22. Chauduri SK, Lovely DR (2003) Electricity generation by direct oxidation of glucose in mediatorless microbial fuel cells. *Nat Biotechnol* 21(10):1229–1232. <https://doi.org/10.1038/nbt867>
23. Niessen J, Schröder U, Harnisch F, Scholz F (2005) Gaining electricity from in situ oxidation of hydrogen produced by fermentative cellulose degradation. *Lett Appl Microbiol* 41:286–290. DOI: 10.1111/j.1472-765X.2005.01742.x
24. Liu H, Ramnarayanan R, Logan BE (2004) Production of electricity during wastewater treatment using a single-chamber microbial fuel cell. *Environ Sci Technol* 38(7):2281–2285. doi.org/10.1021/es034923g
25. Cercado-Quezada B, Delia ML, Bergel A (2010) Testing various food-industry wastes for electricity production in microbial fuel cell. *Bioresour Technol* 101:2748–2754. <https://doi.org/10.1016/j.biortech.2009.11.076>
26. Kim IS, Kim KY, Chae KJ, Choi MJ, Ajayi FF, Jang AM, Kim CW (2011) Enhanced coulombic efficiency in glucose-fed microbial fuel cells by reducing metabolite electron losses using dual-anode electrodes. *Bioresour Technol* 102:4144–4149. DOI: 10.1016/j.biortech.2010.12.036
27. Water quality - (2002) Determination of chemical oxygen demand index (ST-COD) - Small scale closed tube method, ISO15705
28. Umaz RA (2020) Power Management System for Microbial Fuel Cells With 53.02% Peak End-to-End Efficiency. *IEEE Trans Circuits Syst II Express Briefs* 67(11):2592. doi: 10.1109/TCSII.2019.2951810
29. Donovan C, Dewan A, Peng H, Heo D, Beyenal H (2011) Power management system for a 2.5W remote sensor powered by a sediment microbial fuel cell. *J Power Sources* 196:1171–1177. <https://doi.org/10.1016/j.jpowsour.2015.12.040>
30. Mukhtar S, Capareda S, Smith EG (1914) Director, Texas Cooperative Extension, The Texas A&M University System.
31. Ojima Y, Kawaguchi T, Fukui S et al (2020) Promoted performance of microbial fuel cells using *Escherichia coli* cells with multiple-knockout of central metabolism genes. *Bioprocess Biosyst Eng*

- 43:323. <https://doi.org/10.1007/s00449-019-02229-z>
32. Kosimaningrum WE (2018) Modification of Carbon Felt for Construction of Air-Breathing Cathode and Its Application in Microbial Fuel Cell, PhD thesis, University of Montpellier, France
33. Zhang J (ed) (2008) (Ed.). PEM Fuel Cell Electrocatal. Catal. Layer Fundam. Appl. Springer, London, pp 1–79
34. Zhang L, Shen J, Wang L (2014) Stable operation of microbial fuel cells at low temperatures (5 – 10°C) with light exposure and its anodic microbial analysis. *Bioprocess Biosyst Eng* 37:819–827. <https://doi.org/10.1007/s00449-013-1054-8>
35. Lin CW, Wu CH, Huang WT, Tsai SL (2015) Evaluation of different cell-immobilization strategies for simultaneous distillery wastewater treatment and electricity generation in microbial fuel cells. *Fuel* 144:1–8. <https://doi:10.1016/j.fuel.2014.12.009>
36. Logan BE, Cells MF (2007) John Wiley and Sons, Inc
37. Hydro-Genius Professional experimentation models (2000) Heliocentris GmbH, Rudower Chaussee 29, 12489, Berlin, Germany
38. Torres CI, Krajmalnik-Brown R, Parameswaran P, Marcus AK, Wanger G, Gorby Y, Rittmann BE (2009) Selecting anode-respiring bacteria based on anode potential: phylogenetic, electrochemical, and microscopic characterization. *Environ Sci Technol* 43:9519–9524. <https://doi.org/10.1021/es902165y>
39. Yi H, Nevin KP, Kim BC, Franks AE, Klimes A, Tender LM, Lovley DR (2009) Selection of a variant of *Geobacter sulfurreducens* with enhanced capacity for current production in microbial fuel cells. *Biosens Bioelectron* 24:3498–3503. doi: 10.1016/j.bios.2009.05.004
40. Aelterman P, Freguia S, Keller J, Verstraete W, Rabaey K (2008) The anode potential regulates bacterial activity in microbial fuel cells. *Appl Microbiol Biotechnol* 78:409–418. doi: 10.1007/s00253-007-1327-8
41. Ketep F, Bergel A, Bertrand M, Achouak W, Fourest E (2013) Lowering the applied potential during successive scratching/re-inoculation improves the performance of microbial anodes for microbial fuel cells. *Bioresour Technol* 127:448–455. <https://doi.org/10.1016/j.biortech.2012.09.008>
42. Pinto D, Coradin T, Laberty-Robert C (2018) Effect of anode polarization on biofilm formation and electron transfer in *Shewanella oneidensis* /graphite felt microbial fuel cells. *Bioelectrochemistry*, 120:1-9. [ff10.1016/j.bioelechem.2017.10.008](https://doi.org/10.1016/j.bioelechem.2017.10.008)[ff. fahal-01651845](https://doi.org/10.1016/j.bioelechem.2017.10.008)
43. Lee Y-Y, Kim TG, Cho K-S (2016) Characterization of the COD removal, electricity generation, and bacterial communities in microbial fuel cells treating molasses wastewater. *Journal of Environmental Science and Health Part A* 1–8. doi:10.1080/10934529.2016.1199926
44. Nguyen CL, Tartakovsky B, Woodward L (2019) Harvesting Energy from Multiple Microbial Fuel Cells with a High-Conversion Efficiency. *Power Management System ACS Omega* 21:18978. [https://doi: 10.1021/acsomega.9b01854](https://doi.org/10.1021/acsomega.9b01854)

Tables

Table 1. The characteristics values of the MFC

Power density at MPP (mW/m ²)	Current density at MPP (mA/m ²)	Potential at MPP (mV)	Electric resistance at MPP (W)	Open Circuit Potential (mV)	Potential ratio (%)	Energy released during 140 days (mJ/g)
184.84	274.76	145	900	484	29.95	0.019

Table 2. Characteristics of photovoltaic solar module used for polarization of operational amplifier

Standard Irradiation (W/m ²)	1000
Temperature(°C)	25
(A)	0.36
(V)	14.4
(A)	0.33
(V)	12
Power Pm (W)	4
Resistance Rs (Ω)	0.2
Ns	48

Table 3. Values of current and power densities of MFCs associated in series and in parallel after having been used in amplification

Type of association	Interval of current density (mA/m ²)	Maximal power density (mW/m ²)	Current at maximum power density (mA/m ²)
Single MFC	0 - 2571	186	858
Parallel	0 - 160	21	80
Series	0 - 10	5.6	3.6

Figures

Figure 1

Biodegradation of organic substrate (glucose) by bacteria allowing electric energy generation using two distinct pathways: Electron Transport Chain (ETC) and trans Plasma Membrane Electron Transport (tPMET) (Inspired from Figs.2&3 of Ref. (Schaetzle et al. 2008))

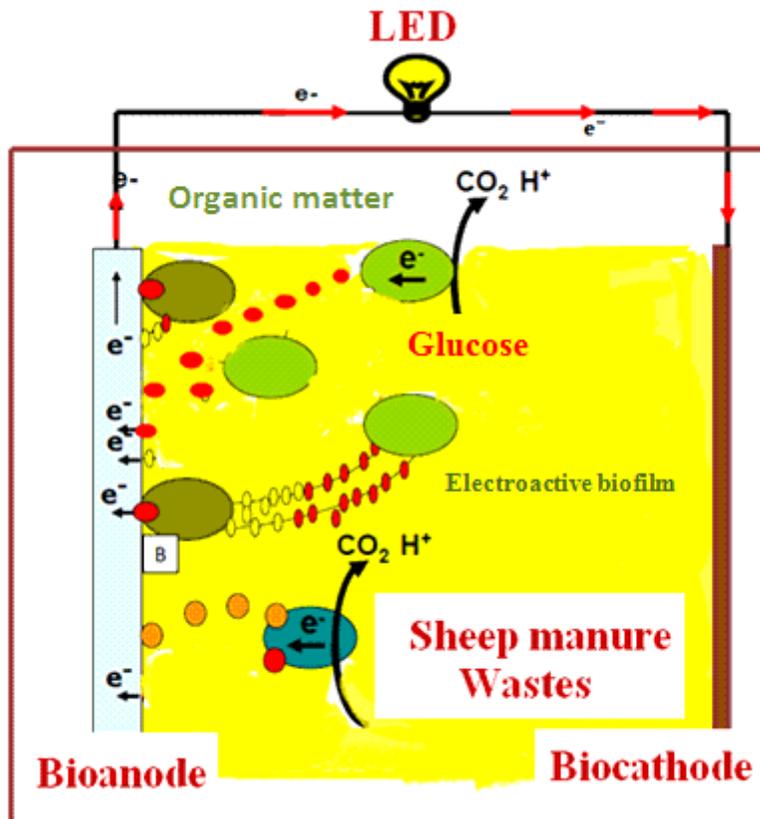


Figure 2

Electrochemical assembly of one-chamber MFC using sheep manure wastes

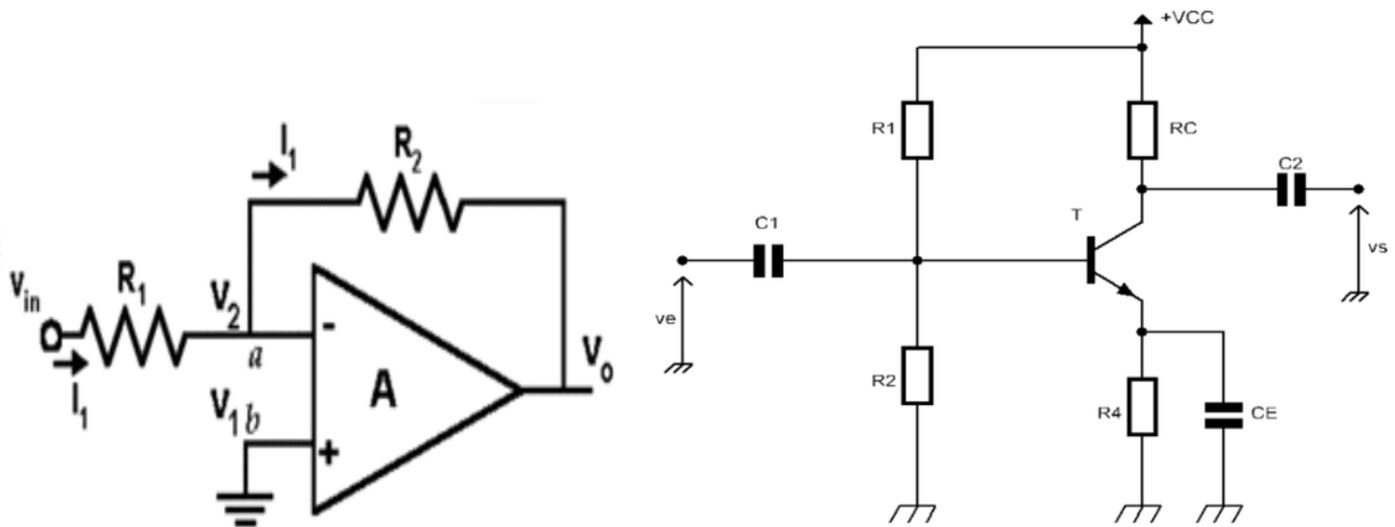


Figure 3

Synoptic diagram of (a): operational amplifier, (b): with common emitter amplifier

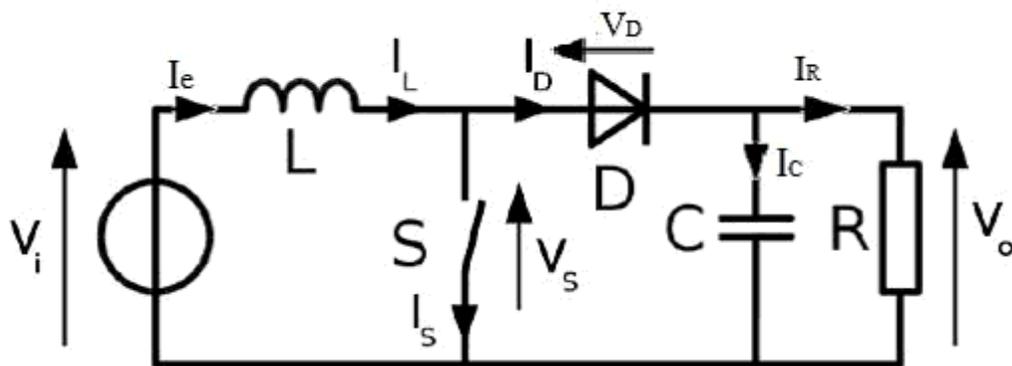


Figure 4

Equivalent circuit of the DC/DC boost converter.

Figure 5

Evolution of MFC open circuit voltage during 870 hours

Figure 6

Polarization and power curves of MFC

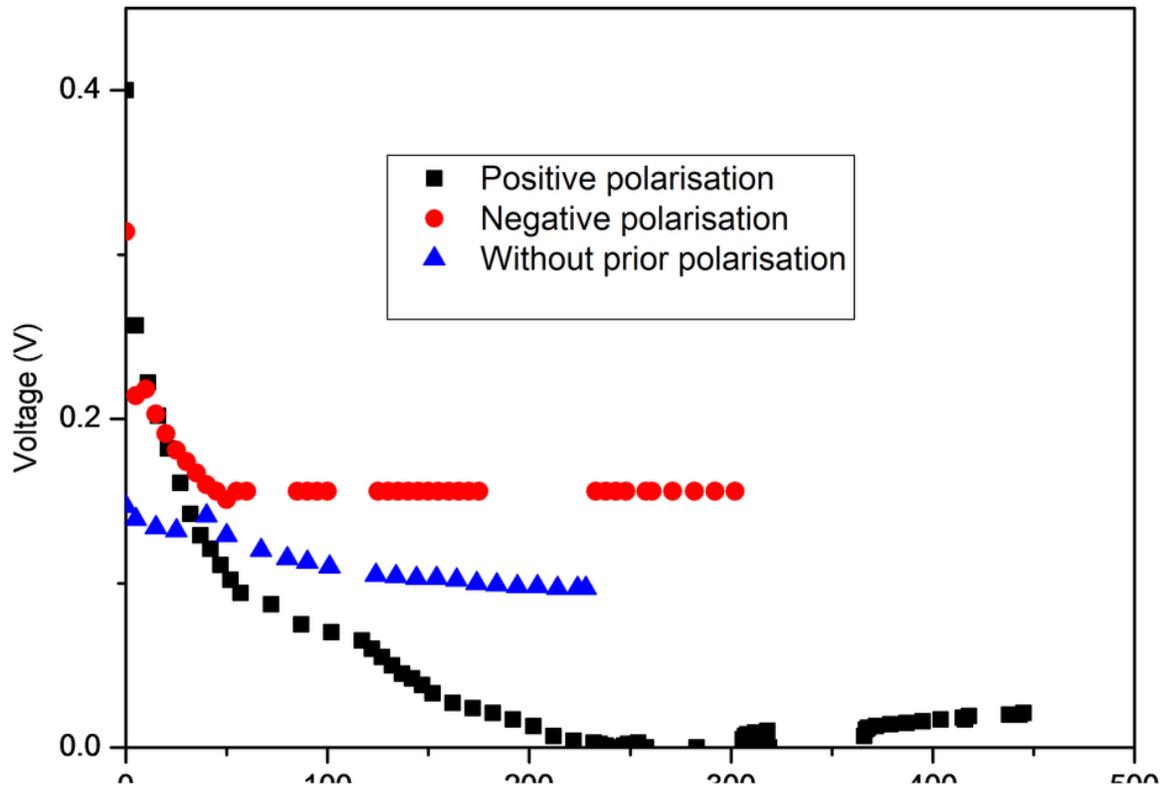


Figure 7

Evolution of MFC discharged to electric resistance 1000 Ω upon positive and negative polarizations

Figure 8

Amplifier polarized with photovoltaic solar module for improvement of output voltage.

Figure 9

Evolution of input and amplified voltages using different configurations: (a-a') single stack of MFC; (b-b') series association of MFCs; (c-c') parallel association of MFCs; (d-d') parallel association of MFCs with polarization voltage

Figure 10

Evolution of output amplified voltage with solar polarization voltage.

Figure 11

Experimental set up containing MFCs, amplifier and DC/DC converter.

Figure 12

Evolution of output amplified voltage using solar polarization and DC/DC boost converter.

Figure 13

Power density curves of MFC: Single-compartment and parallel association

Figure 14

Power density curves of MFC: Series association and parallel association