

Identification of molecular markers associated with the progression and prognosis of lung adenocarcinoma: a bioinformatic study

Kexin Du (✉ 980204023@qq.com)

Beijing University of Chinese Medicine Affiliated Dongzhimen Hospital

Xioahan Wang

The Third Affiliated Hospital of Beijing University of Traditional Chinese Medicine

Research

Keywords: lung adenocarcinoma, pathway, sex, CCL14, GNRH2, CCR5, CCR2

Posted Date: December 21st, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-131100/v1>

License:  This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Abstract

Objective: For the identification of genes of prognostic significance related to tumor microenvironment (TME) in lung adenocarcinoma.

Methods and Materials: Transcriptome data and clinical data of lung adenocarcinoma originated from the Cancer Genome Atlas (TCGA) database. Immune scores and stromal scores were calculated by “Estimation of Stromal and Immune cells in Malignant Tumors using Expression data” algorithm. Based on these calculated scores, the samples were classified as high and low score groups. The average score of respective gene in different groups was calculated. A heat map was created to screen out genes exhibiting differential expressions. The interaction of up-regulated differentially expressed genes (DEGs) and down-regulated DEGs was harvested from a Venn diagram and then covered in the overlapping genes. The core genes influencing prognosis of lung adenocarcinoma were screened out by function enrichment analysis, protein-protein interaction network analysis and Kaplan-Meier (K-M) method on the overlapping genes.

Results: A total of 515 samples of lung adenocarcinoma were harvested from TCGA database. As revealed from the results, a high immune score was related to a high survival rate, while the matrix did not show significant relationships to survival rate. A total of 775 DEGs and 367 overlapping genes were harvested. The functions of these overlapping genes were tightly correlated with DEGs and immune response and were noticeably improved in cytokine-cytokine receptor interaction and chemokine signaling pathway. Eight genes, namely, CCR5, CCR2, CCL14, GNRH2, PKHD1L1, MS4A1, FCER2 and FDCSP, were correlated with prognosis of lung adenocarcinoma.

Conclusion: The genes and pathways affecting prognosis of lung adenocarcinoma were screened out, which offer ideas for subsequent study on lung adenocarcinoma.

1. Introduction

Cancer is affecting increasing people worldwide on a year to year basis; it has turned into a primary factor threatening human health. Besides, lung cancer heads the list of cancers in morbidity and mortality[1]. In accordance with the report of International Agency to study cancer (IARC), there are about 2.1 million lung cancer patients in the world. Lung cancer has two major types, namely, small-cell lung carcinoma (SCLC) (20%) and non-small-cell lung carcinoma (NSCLC) (80%)[2]. However, most patients with NSCLC were diagnosed in late stage, and the 5-year survival rate was less than 15%[3]. Though an increasing number of treatments have been used in clinic, drug resistance and intolerance still affect the therapeutic effect of lung cancer. Therefore, for the enhancement of the prognosis of lung cancer patients, it is necessary to understand the pathogenesis of tumor and explore more treatment strategies.

Tumor microenvironment(TME) is critical to tumor development. The inflammatory cells and immunomodulatory mediators in TME may participate in tumor progression[4-6]and may help tumor escape with the disease progression[7, 8]. For instance, tumor associated macrophages (TAMs) can

present different phenotypes according to the stimulation of TME (e.g., chemokines), promoting tumor progression or specific immune response [9-11]. Moreover, tumor associated fibroblasts (CAF) can promote immunosuppression by inducing myoid derived suppressor cell (MDSC) to generate reactive oxygen species (ROC) [12]. In terms of treatment, some studies proved that some drugs can exert high therapeutic effect by improving TME. For instance, dexamethasone Increases Cisplatin-Loaded Nanocarrier Delivery and Efficacy in Metastatic Breast Cancer by Normalizing the Tumor Microenvironment[13]. The vaccine targeting Legumain builds the novel paradigm since a reduction in the density of TAMs in the TME inhibits the release of factors potentiating tumor growth and angiogenesis. As a result, the TME is remodeled, and its immunosuppressive milieu decreases; thus, potentiates the DNA vaccine's ability to effectively suppress metastasis, vascularization, and proliferation of tumor cells [14]. Though we have some knowledge of the relationship between TME and tumor, the interaction between TME and lung adenocarcinoma has been rarely known.

The Cancer Genome Atlas (TCGA) database is a database that collects cancer patients' information. Thus far, it has collected more than 20000 patients' information, including 33 kinds of tumors. Big data has facilitated the development of all walks of life, and medicine is no exception. Yoshihara et al. developed an algorithm [10] that can select genes that affect tumor prognosis by calculating the immune score and stromal score of TME. This algorithm is applied in hepatocellular carcinoma and leukemia[15, 16], which it is proved to be feasible. In the present study, based on TCGA database and by the algorithm created by Yoshihara et al., the aim is to screen out genes affecting prognosis of lung adenocarcinoma and offer ideas for lung adenocarcinoma diagnosis and treatment.

2. Methods And Material

Transcriptome data and clinical data of lung adenocarcinoma originated from TCGA website(<https://portal.gdc.cancer.gov/>). The clinical data covered survival time, survival state, age, sex and stages. The Yoshihara's algorithm is adopted in the downloaded RNA expression data. The stromal, immune and estimate scores of cancerous tissue samples were calculated by R (3. 6. 2) estimate package, of which the survival score was the sum of stromal score and immune score.

The samples were split into high score group and low score group with the median of immune score and stromal score as the dividing line. Multivariate survival regression analysis was conducted with R package; meantime, survival time was adopted as an independent variable, and survival rate was adopted as a dependent variable ($p < 0.05$) to determine the relationships between stromal, immunity and survival.

To investigate the relationships between sex and immune, stromal score, R software was used to carry out Wilcoxon rank-sum test ($p < 0.05$) and a box plot was plotted.

The samples were split into high score group and low score group based on median of immune score and stromal score and Wilcoxon rank-sum test was adopted ($p < 0.05$). The average value of respective gene in different groups was calculated. Besides, the logarithm of "average value in high score group/average

value in low score group” was adopted, in which Log2FC was acquired. differentially expressed genes (DEGs) according to $\text{Log}_2\text{FC} > 1$ or $\text{Log}_2\text{FC} < -1$ and $\text{FDR} < 0.05$ were selected. Genes complying with $\text{Log}_2\text{FC} > 1$ and $\text{FDR} < 0.05$ were termed as up-regulated genes and genes complying with $\text{Log}_2\text{FC} < -1$ and $\text{FDR} < 0.05$ were down-regulated genes. Pheatmap in R package was used to plot the heat map which can observe the expression of genes in different groups.

The interactions of up-regulated genes in stromal group and immune group were collected, as well as those and of down-regulated genes. A Venn diagram was plotted with VennDiagram package in R package.

The overlapping genes were analyzed using clusterProfiler, org.Hs.eg.db, ggplot2, stringi, DOSE tool package, gene ontology (GO) and kyoto encyclopedia of genes and genomes (KEGG) in R language ($p < 0.05$, $q < 0.05$). The study screened out the top 10 interactions exhibiting the most highly enriched MF, CC and BP in GO and all 28 KEGG pathways. GO expressed the functions of genes in biological process, cellular component and molecular functions, while KEGG was used for delving into the enrichment ways of genes.

This study conducted PPI analysis on DEG genes on STRING (<https://string-db.org/>) data platform. minimum required interaction score = 0.9. After deleting free proteins, the top 30 most connected genes were taken as core genes with R, with which, a bar diagram was plotted.

K-M survival analysis on DEGs that affected prognosis of lung adenocarcinoma with survival package. $p < 0.05$ suggested a statistical significance. Genes correlated with prognosis were screened out.

3. Results

A total of 515 tissue samples from patients were harvested from TCGA database, including 535 cancer tissue samples and 59 normal sample. Clinical information of 522 patients was obtained, covering 242 male cases and 280 female cases (Table 1). The stromal score of 535 tumor samples was -1789.62–2097.96, with the median of 200.68. The immune score reached -942.51–3442.08, and the median was 1524.46. The estimate score reached -2357.12–4891.50, and the median was 1793.79.

Table 1 Clinical information

		Male(%)	Female(%)	Total(%)
	Number	242(46.4%)	280(53.6%)	522
Fustat	Alive	162(67.0%)	193(69.0%)	355(68%)
	Dead	80(33.0%)	87(31.0%)	167(32%)
Futime(year)	0-3	190(78.5%)	219(78.2%)	409(78.4%)
	3-5	22(9.1%)	33(11.8%)	55(10.5%)
	5-10	17(7.0%)	15(5.3%)	32(6.1%)
	≥10	4(1.7%)	3(1.1%)	7(1.4%)
	Unknown	9(3.7%)	10(3.6%)	19(3.6%)
Age	≤50	16(6.6%)	18(6.4%)	34(6.5%)
	50-75	182(75.2%)	210(75.0%)	392(75.1%)
	≥75	35(14.5%)	42(15.0%)	77(14.8%)
	Unknown	9(3.7%)	10(3.6%)	19(3.6%)
Stage	I	116(48.0%)	163(58.2%)	279(53.4%)
	II	68(28.1%)	56(20.0%)	124(23.8%)
	III	39(16.1%)	46(16.4%)	85(16.3%)
	IV	14(5.8%)	12(4.3%)	26(5.0%)
	Unknown	5(2.0%)	3(1.1%)	8(1.5%)

COX regression revealed that high immune score and estimate score were correlated with a high survival rate ($p=0.0207$ and $p=0.0257$, Fig. 1A and Fig. 1C). The effects in stromal score group was insignificant ($p=0.1030$, Fig. 1B).

From the box plot, the stromal score, immune score and estimate score of females were all higher than those of males ($p=0.1081$, $p=0.0360$ and $p=0.00362$, Fig.2A, Fig.2B and Fig. 2C).

Heat map which can observe the expression of genes in different groups was plotted based on stromal score and immune score (Fig. 3A and Fig. 3B). The pink represents the low score group, while the blue represents the high score group. Low expression is denoted in green, median expression is represented in black, while high expression is in red. According to $\text{Log}_2\text{FC}>1$ or $\text{Log}_2\text{FC}<-1$ and $\text{FDR}<0.05$, we found 683 up-regulated genes and 120 down-regulated genes in stromal score group. The immune score group included 611 up-regulated genes and 164 down-regulated genes. In Venn diagram, 300 up-regulated common genes (Fig. 4A) and 67 down-regulated common genes (Fig. 4B) were screened out.

GO enrichment analysis (Fig. 5) suggested a close relationships between DEGs and immune response. Immune response-activating cell surface receptor signaling pathway, regulation of lymphocyte proliferation, T cell activation and leukocyte proliferating process were highly enriched. In KEGG enrichment analysis, hematopoietic cell lineage,, viral protein interaction with cytokine and cytokine receptor chemokine signaling pathway and cytokine-cytokine receptor interaction were most highly up-regulated. The mentioned pathways displayed close relationships to immune reaction and virus.

Protein-protein interaction (PPI) network covered 152 nodes and 612 edges (Fig. 7A). The top 30 most connected genes included FPR2, C3AR1, MCHR1, CCR5, FPR1, CCL19, CCR2, CXCL10, ADORA3, AGTR2, CCL1, CCL13, CCL21, CCR4, CCR8, CNR2, CXCL11, CXCL13, CXCL6, CXCL9, FPR3, GPR183, NPW, P2RY12, P2RY13, IGLL5, CD3G, ITGAM, GPR84 as well as CD4 (Fig. 7B). These genes were critical to viral protein interaction with cytokine and cytokine receptor, cytokine-cytokine receptor interaction, chemokine signaling pathway, staphylococcus aureus infection and neuroactive ligand-receptor interaction.

In the study ascertaining the effect of single gene on prognosis of lung adenocarcinoma, 122 genes were significantly related to prognosis ($p < 0.05$), covering CCR5, CCR2, CCL14, GNRH2, PKHD1L1, MS4A1, FCER2 and FDCSP (Fig. 8). The prognosis was also correlated with neuroactive ligand-receptor interaction pathway.

4. Discussion

TME, composed of endothelial cells, inflammatory mediators, mesenchymal stem cells, immune cells and stromal cells, displays tight relationships to occurrence and development of a tumor[17, 18]. It has been demonstrated that the immune cells in TME of lung cancer are phase-dependent[19, 20], and the immune cells in different stages can act as potential biomarkers. Tumor cells were capable of modifying stromal cells in TME, while stromal cells can boost the growth of tumor cells[18, 21]. Stromal cells and immune cells are becoming a novel way for cancer treatment.

In this study, the estimate, stromal and immune scores of respective sample originating from TCGA database were calculated. According to the outcomes, high immune score displayed correlations with a high survival rate and longer survival time. The stromal score, immune score and estimate score of females were overall higher than those of males. The samples were split into high score group and low score group in accordance with immune score and stromal score. Genes exhibiting differential expressions were screened out. Interaction of genes was harvested and 300 up-regulated genes, 67 down-regulated genes were harvested. GO enrichment analysis, PPI network and KEGG enrichment analysis were performed on DEGs and as revealed from the results, there was a close relationships between DEGs and immune response. Cytokine-cytokine receptor interaction, chemokine signaling pathway and viral protein interaction with cytokine and cytokine receptor pathway were mostly highly enriched. FPR2, C3AR1 and MCHR1 were most connected in PPI network. As suggested from the investigation of the relationships between single gene and survival, CCR5, CCR2, CCL14, GNRH2, PKHD1L1, FCER2, MS4A1 and FDCSP exhibited a close relation with survival.

CCR5 and CCR2 are chemokine receptors, pertaining to G-protein-coupled receptors (GPCRs). Nearly 40% of drugs take effect via GPCRs[22, 23]. Multiple cells (e.g., B cells, natural killer cells and monocytes) can express CCR5. When chemokine ligand is combined with CCR5, they may activate immune reaction and induce migration[24, 25]. Innate $\alpha\beta$ T cells are capable of inducing CCR5-dependent immunogenic macrophage programming, activating and amplifying T cells, thereby inhibiting tumor growth[26]. CCR5 may help elevate the survival rate of tumor-infiltrating macrophages, and CCR2 can keep macrophages dominant in tumor infiltrating cells[27]. The chemotaxis experiment demonstrated proved that RUNX3 can facilitate CD8 T cell recruitment by up-regulating CCL3 and CCL20, in which CCR5 and CCR6 act as receptors [28]. In terms of treatment, anti-CCR5 therapy in colorectal cancer is capable of avoiding the use of tumor cells on immune cells[29, 30]. CCR5 also can facilitate tumor metastasis and migration[31-35]. PRDX6 boosts lung tumor development through its mediated and CCL5-related activation of the JAK2/STAT3 pathway[36]. CCR5 also can facilitate neovascularization[37, 38]. For CCR2, CCL2-CCR2 can impact the recruitment of inflammatory cells as well as differentiation of effector T cells[39, 40]. Moreover, it can also activate T cells and memory T cells[41]. Circulating fibrocytes make the preparation of the lung for cancer metastasis by adopting Ly-6C⁺ monocytes through CCL2[42]. In hepatocellular carcinoma, oncogene is able to induce aging hepatocytes to secrete cytokines and attract immune cells to clear away them, as an attempt to prevent the occurrence of cancer, which is termed "senescence surveillance". The process requires to be assisted by CCR2 myelocytes, so the reduction in CCR2 will facilitate the progression of[43]. In brief, CCR5 and CCR2 may affect tumor development by influencing the expression and migration of immune cells.

CCL14, also termed as chemokine 14, drives the cells to migrate to a specific site. Chemokine has been extensively employed; it also leads different subtypes of immune cells to TME[44, 45]. Moreover, chemokine is capable of directly affecting tumor cells and endothelial cells in TME, thereby regulating tumor growth, angiogenesis and tumor migration. It displays relationships to tumor development, so it marks a good prognosis[46, 47]. The role of CCL14 in tumor progression continues to be not clear. Some research proved that CCL14 inhibited hepatocellular carcinoma (HCC) in vitro through the inhibition of the progression of cell cycle and promoting cell apoptosis and inhibited HCC development in vivo through Wnt/ β -catenin signal pathway[48]. By suppressing CCL14, angiogenesis and metastasis of breast cancer can be inhibited [49]. A research highlighted that CCL14 acts as an individual element of epithelial ovarian cancer (EOC) prognosis, and upregulation of CCL14 is associated with a more conducive prognosis in EOC patients [50]. Thus, CCL14 may be a non-specific prognostic factor. In contrast, CCL14 may facilitate invasion, proliferation of bone marrow and macrophage polarization in multiple myeloma. It may also has a relation with chemotherapy resistance[51]. As indicated from the data on TCGA, CCL14 acts as a beneficial factor of prognosis, complying with the conclusions in breast cancer and hepatocellular carcinoma. More studies are required to clarify why CCL14 achieves different results in different cancers.

GNRH2 (gonadotropin releasing hormone 2) refers to a subtype of gonadotrophin releasing hormone, released by hypothalamus and facilitating secretion of luteinizing hormone[52-54]. GnRH2 may be the oldest form of GnRH[55] exhibiting a strong conservative property[56]and has remained almost

unchanged during the long-term evolution. GnRH2 is produced in kidneys, bone marrow, lymph glands, heart, lungs, etc.[57, 58]. GnRH2 has been proven capable of inhibiting proliferation of prostatic cancer[59, 60], ovarian cancer, breast cancer and endometrial cancer[61-64]; it also exerts better effects than GnRH1[65]. According to Ling Poon S et al., GnRH2 can inhibit ribosomal phosphoprotein of cancer cells and further inhibit protein translation and cell proliferation[66]. Extracellular vesicles may be its way to regulate tumor proliferation[67]. Moreover, GnRH2 is of implication in cell apoptosis. Several studies confirmed that GnRH2 facilitated apoptosis of human granulosa cells[68] and regulated autophagy[69]. In tumor invasion, GnRH2 can up-regulate the content of metal protease[70], which is crucial in different aspects of biological activities (e.g., cell proliferation, differentiation and vascularized cell migration)[71]. GnRH2 impacts cell apoptosis and cancer metastasis, as well as prognosis of cancer. Shiota M et al. reported that GnRH2 mutation would down-regulate the influences of androgen deprivation therapy in patients with metastatic prostatic cancer[72]. GnRH2 can impact tumor proliferation, metastasis, prognosis and others, and it plays a role in multiple tumors. Besides its stability, this gene is an ideal index.

In the present study, the genes and pathways tightly correlated with prognosis in TME of lung adenocarcinoma were explored with TCGA database and the algorithm created by Yoshihara et al.; they provide novel insights for subsequent studies. The mentioned genes and pathways were proven effective in other tumors, whereas their role in lung adenocarcinoma was not reported. Further experimental research is still required to determine the actual effects before their clinical application. The mentioned genes and pathways impact TME and TME while TME conversely impacts tumor proliferation and metastasis; thus, they interact with each other.

Some limitations remain in the present study. First, all the data are from TCGA database with an insufficient sample size; second, the mentioned results have not been experimentally verified. All these factors may cause outcome bias.

In brief, eight genes were tightly correlated with survival after an estimate analysis, and they may act as potential biological markers of lung adenocarcinoma

Declarations

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Ethics Statement

The data in this article are all from TCGA database, so there is no ethical approval and consent participation section.

Consent for Publication

Not applicable.

Availability of Data and Materials

The datasets generated during and analyses during the current study are available in the TCGA database.

Conflicts of Interest

The authors declare no conflict of interest.

Authors' Contributions

Kexin Du and Xiaohan Wang analyzed the data, and Kexin Du wrote the article.

Acknowledgement

We appreciated Dongyun Li for directing writing the manuscript.

References

- [1] B. F, F. J, S. I, S. RL, T. LA, J. A, Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries, *CA: a cancer journal for clinicians* 68(6) (2018) 394-424.
- [2] T. LA, B. F, S. RL, F. J, L.-T. J, J. A, Global cancer statistics, 2012, *CA: a cancer journal for clinicians* 65(2) (2015) 87-108.
- [3] T. LA, S. RL, J. A, Lung Cancer Statistics, *Advances in experimental medicine and biology* 893 (2016) 1-19.
- [4] B. A, P. A, M. E, R. T, N. DM, C. AR, A. A, Inflammatory angiogenesis and the tumor microenvironment as targets for cancer therapy and prevention, *Cancer treatment and research* 159 (2014) 401-26.
- [5] B. A, F. G, A. A, N. DM, A think tank of TINK/TANKs: tumor-infiltrating/tumor-associated natural killer cells in tumor progression and angiogenesis, *Journal of the National Cancer Institute* 106(8) (2014) dju200.
- [6] B. FR, C. M, H. T, The tumor microenvironment at a glance, *Journal of cell science* 125 (2012) 5591-6.
- [7] S. RD, O. LJ, S. MJ, Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion, *Science (New York, N.Y.)* 331(6024) (2011) 1565-70.
- [8] D. M, M. C, S. LM, C. AF, J. T, Expression of tumour-specific antigens underlies cancer immunoediting, *Nature* 482(7385) (2012) 405-9.

- [9] N. DM, D.L.B. A, V. N, M. L, A. A, Inflammation, inflammatory cells and angiogenesis: decisions and indecisions, *Cancer metastasis reviews* 27(1) (2008) 31-40.
- [10] C. LM, W. Z, Inflammation and cancer, *Nature* 420(6917) (2002) 860-7.
- [11] P. F, V. MA, B. D, N. A, M. D, S. A, D.S. C, N. L, D.P. M, A distinguishing gene signature shared by tumor-infiltrating Tie2-expressing monocytes, blood "resident" monocytes, and embryonic macrophages suggests common functions and developmental relationships, *Blood* 114(4) (2009) 901-14.
- [12] X. H, R. CP, H. J, Z. C, W. H, W. AA, A. R, G. P, S. MA, S. XS, C. PJ, C. M, M. JR, S. D, L. A, J. Y, M. LY, C. A, B. PE, Cancer-Associated Fibroblasts Promote Immunosuppression by Inducing ROS-Generating Monocytic MDSCs in Lung Squamous Cell Carcinoma, *Cancer immunology research* 8(4) (2020) 436-450.
- [13] M. JD, P. M, W. C, K. TT, M. MR, V. C, T. K, P. P, M. F, P. C, I. G, T. S, G. N, S. T, W. ME, M. VA, Q. S, N. J, L. RM, K. M, S. MD, S. T, K. K, C. H, Dexamethasone Increases Cisplatin-Loaded Nanocarrier Delivery and Efficacy in Metastatic Breast Cancer by Normalizing the Tumor Microenvironment, *ACS nano* 13(6) (2019) 6396-6408.
- [14] X. R, L. Y, N. AG, R. RA, Oral DNA vaccines target the tumor vasculature and microenvironment and suppress tumor growth and metastasis, *Immunological reviews* 222 (2008) 117-28.
- [15] D. Z, W. J, X. B, J. Z, W. G, Z. J, P. M, G. Y, W. Z, Mining TCGA Database for Tumor Microenvironment-Related Genes of Prognostic Value in Hepatocellular Carcinoma, *BioMed research international* 2019 (2019) 2408348.
- [16] N. J, W. Y, Q. F, L. X, Y. S, L. S, F. J, Z. Y, Screening the Cancer Genome Atlas Database for Genes of Prognostic Value in Acute Myeloid Leukemia, *Frontiers in oncology* 9 (2019) 1509.
- [17] H. D, W. RA, The hallmarks of cancer, *Cell* 100(1) (2000) 57-70.
- [18] H. D, C. LM, Accessories to the crime: functions of cells recruited to the tumor microenvironment, *Cancer cell* 21(3) (2012) 309-22.
- [19] K. J, B. SE, Y. GH, K. KH, H. ML, M. HE, H. JJ, L. SM, M. DK, M. MW, H. AM, Neutrophils dominate the immune cell composition in non-small cell lung cancer, *Nature communications* 8 (2017) 14381.
- [20] B. GA, T. A, P. SS, W. J, W. A, O. C, E. K, S. T, G. F, S. W, F. L, S. R, Immune and Inflammatory Cell Composition of Human Lung Cancer Stroma, *PloS one* 10(9) (2015) e0139073.
- [21] Q. DF, J. JA, Microenvironmental regulation of tumor progression and metastasis, *Nature medicine* 19(11) (2013) 1423-37.

- [22] O. JP, A.-L. B, H. AL, How many drug targets are there?, *Nature reviews. Drug discovery* 5(12) (2006) 993-6.
- [23] E. RM, R. T, New insights into GPCR function: implications for HTS, *Methods in molecular biology* (Clifton, N.J.) 552 (2009) 1-13.
- [24] V. A, S. A, B. V, M. B, The pros and cons of chemokines in tumor immunology, *Trends in immunology* 33(10) (2012) 496-504.
- [25] L. B, S. M, M. LJ, W. D, D. RW, Quantification of CD4, CCR5, and CXCR4 levels on lymphocyte subsets, dendritic cells, and differentially conditioned monocyte-derived macrophages, *Proceedings of the National Academy of Sciences of the United States of America* 96(9) (1999) 5215-20.
- [26] H. M, K. E, M. A, R. JAK, L. SM, L. KR, M. H, K. M, T. LE, O. A, L. J, S. RC, S. S, G. J, I. KMS, D. I, S. S, W. W, A. B, L. J, D. B, A. S, I. M, G. M, L. J, C. K, F. MS, W. B, W. J, S. S, A. DO, P. S, V. V, S. D, W. KK, C. LM, M. G, Innate $\alpha\beta$ T Cells Mediate Antitumor Immunity by Orchestrating Immunogenic Macrophage Programming, *Cancer discovery* 9(9) (2019) 1288-1305.
- [27] S. Y, U. S, K. M, S. T, T. JE, A. J, S. Y, K. M, K. K, M. N, M. K, Chemokine-mediated rapid turnover of myeloid-derived suppressor cells in tumor-bearing mice, *Blood* 111(12) (2008) 5457-66.
- [28] S. Q, S. J, Z. C, C. J, Z. L, W. X, Transcription factor RUNX3 promotes CD8 T cell recruitment by CCL3 and CCL20 in lung adenocarcinoma immune microenvironment, *Journal of cellular biochemistry* 121 (2020) 3208-3220.
- [29] Anti-CCR5 Therapy Circumvents Immune Cell Exploitation by Tumors, *Cancer discovery* 6(6) (2016) 571.
- [30] H. N, Z. I, B. A, K. C, K. F, S.-C. M, S. T, B. K, K. J, L. F, L. T, L.-M. C, U. A, K. M, W. J, S. M, B. MW, Z. L, H. T, B. A, K. C, L. S, S. C, G. N, F. CS, J. D, Tumoral Immune Cell Exploitation in Colorectal Cancer Metastases Can Be Targeted Effectively by Anti-CCR5 Therapy in Cancer Patients, *Cancer cell* 29(4) (2016) 587-601.
- [31] L. J, W. C, M. X, T. Y, W. C, F. Y, L. Y, High expression of CCR5 in melanoma enhances epithelial-mesenchymal transition and metastasis via TGF β 1, *The Journal of pathology* 247(4) (2019) 481-493.
- [32] K. G, F. C, Y. M, Y. KC, Reduced RhoA expression enhances breast cancer metastasis with a concomitant increase in CCR5 and CXCR4 chemokines signaling, *Scientific reports* 9(1) (2019) 16351.
- [33] v.D. HW, W. QP, B. DT, D. BK, O.C. BP, T. JP, S. JS, C-C chemokine receptor 5 on pulmonary fibrocytes facilitates migration and promotes metastasis via matrix metalloproteinase 9, *The American journal of pathology* 173(1) (2008) 253-64.
- [34] B. AC, P. N, T. RL, S. M, G. LA, G. ME, S. JR, F. RA, P. CA, Lung adenocarcinoma invasion in TGF β 2II-deficient cells is mediated by CCL5/RANTES, *Oncogene* 27(4) (2008) 557-64.

- [35] v.D. HW, O.C. W, B. WJ, A. RM, T. JP, S. JS, C-C chemokine receptor 5 on stromal cells promotes pulmonary metastasis, *Cancer research* 65(8) (2005) 3374-9.
- [36] Y. HM, P. KR, P. MH, K. DH, J. MR, K. JY, K. EC, Y. DY, H. SB, H. JT, PRDX6 promotes tumor development via the JAK2/STAT3 pathway in a urethane-induced lung tumor model, *Free radical biology & medicine* 80 (2015) 136-44.
- [37] W. Y, L. YY, M. K, B. T, M. N, CCL3-CCR5 axis regulates intratumoral accumulation of leukocytes and fibroblasts and promotes angiogenesis in murine lung metastasis process, *Journal of immunology* (Baltimore, Md. : 1950) 181(9) (2008) 6384-93.
- [38] S. MJ, G. C, A. VR, F. R, R. P, W. DE, D. CJ, N. I, E. B, W. M, R. K, S. A, M. V, P. N, M. AS, Cancer cell CCL5 mediates bone marrow independent angiogenesis in breast cancer, *Oncotarget* 7(51) (2016) 85437-85449.
- [39] R. BJ, R. H, R. R, N. RJ, G. RP, C. CL, R. BJ, High level monocyte chemoattractant protein-1 expression in transgenic mice increases their susceptibility to intracellular pathogens, *Journal of immunology* (Baltimore, Md. : 1950) 155(10) (1995) 4838-43.
- [40] C. SW, W. KS, R. JH, S. PS, L. P, K. SL, Role of monocyte chemoattractant protein-1 (MCP-1) in Th1 (mycobacterial) and Th2 (schistosomal) antigen-induced granuloma formation: relationship to local inflammation, Th cell expression, and IL-12 production, *Journal of immunology* (Baltimore, Md. : 1950) 157(10) (1996) 4602-8.
- [41] R. BJ, Chemokines, *Blood* 90(3) (1997) 909-28.
- [42] v.D. HW, P. DA, W. QP, M. EC, S. JS, Circulating fibrocytes prepare the lung for cancer metastasis by recruiting Ly-6C⁺ monocytes via CCL2, *Journal of immunology* (Baltimore, Md. : 1950) 190(9) (2013) 4861-7.
- [43] E. T, W. K, J. J, M. C, Y. T, K. S, M.-E. J, L. T, F. M, R. F, H. M, W. XW, Z. L, G. TF, Distinct Functions of Senescence-Associated Immune Responses in Liver Tumor Surveillance and Tumor Progression, *Cancer cell* 30(4) (2016) 533-547.
- [44] F. EF, K. EJ, B. EC, Integrating conflicting chemotactic signals. The role of memory in leukocyte navigation, *The Journal of cell biology* 147(3) (1999) 577-88.
- [45] F. EF, C. JJ, B. EC, Multistep navigation and the combinatorial control of leukocyte chemotaxis, *The Journal of cell biology* 139(5) (1997) 1349-60.
- [46] N. N, W. MS, Z. W, Chemokines in the cancer microenvironment and their relevance in cancer immunotherapy, *Nature reviews. Immunology* 17(9) (2017) 559-572.
- [47] B. F, Cancer and the chemokine network, *Nature reviews. Cancer* 4(7) (2004) 540-50.

- [48] Z. M, X. W, W. C, H. J, X. J, Z. Y, Z. Y, C. J, D. S, L. B, L. C, CCL14 serves as a novel prognostic factor and tumor suppressor of HCC by modulating cell cycle and promoting apoptosis, *Cell death & disease* 10(11) (2019) 796.
- [49] L. Q, S. L, G. B, Y. W, W. J, Z. D, H. X, Y. Z, S. Y, Binding of the JmjC demethylase JARID1B to LSD1/NuRD suppresses angiogenesis and metastasis in breast cancer cells by repressing chemokine CCL14, *Cancer research* 71(21) (2011) 6899-908.
- [50] C. Y, L. Y, H. L, H. H, C. X, X. Y, Z. Z, C. J, C-C motif chemokine 14 as a novel potential biomarker for predicting the prognosis of epithelial ovarian cancer, *Oncology letters* 19(4) (2020) 2875-2883.
- [51] L. Y, Z. Y, L. T, W. Q, Q. J, L. Y, Z. M, B. E, Y. M, R. F, Y. Q, C. Z, Chemokines CCL2, 3, 14 stimulate macrophage bone marrow homing, proliferation, and polarization in multiple myeloma, *Oncotarget* 6(27) (2015) 24218-29.
- [52] S. AV, A. A, B. CY, K. AJ, S. S, R. TW, Hypothalamic neurohormones regulating anterior pituitary function, *Recent progress in hormone research* 24 (1968) 497-588.
- [53] S. AV, A. A, K. AJ, M. H, B. Y, R. TW, N. RM, D. L, W. WF, Gonadotropin-releasing hormone: one polypeptide regulates secretion of luteinizing and follicle-stimulating hormones, *Science (New York, N.Y.)* 173(4001) (1971) 1036-8.
- [54] K. S, Evolution of the regulatory mechanisms for the hypothalamic-pituitary-gonadal axis in vertebrates-hypothesis from a comparative view, *General and comparative endocrinology* 284 (2019).
- [55] M. RP, L. ZL, P. AJ, F. CA, M. K, M. SR, Gonadotropin-releasing hormone receptors, *Endocrine reviews* 25(2) (2004) 235-75.
- [56] F. RD, W. RB, Gonadotropin-releasing hormone genes: phylogeny, structure, and functions, *Frontiers in neuroendocrinology* 20(3) (1999) 224-40.
- [57] N. JD, D. LW, S. JC, M. LC, K. JH, A regulator of G protein signaling, RGS3, inhibits gonadotropin-releasing hormone (GnRH)-stimulated luteinizing hormone (LH) secretion, *BMC cell biology* 2 (2001) 21.
- [58] M. R, L. S, C. D, P. A, M. S, T. B, O. T, M. M, L. G, S. R, F. B, S. G, K. R, T. E, K. A, A novel mammalian receptor for the evolutionarily conserved type II GnRH, *Proceedings of the National Academy of Sciences of the United States of America* 98(17) (2001) 9636-41.
- [59] D. S, S. J, K. MM, R. CN, L. HY, G. VJ, Expression of GnRH type II is regulated by the androgen receptor in prostate cancer, *Endocrine-related cancer* 14(3) (2007) 613-24.
- [60] P. S, H. JM, C. J, H. JI, S. JY, Apoptotic death of prostate cancer cells by a gonadotropin-releasing hormone-II antagonist, *PloS one* 9(6) (2014) e99723.

- [61] L. P, M. RM, M.M. M, M. M, The biology of gonadotropin hormone-releasing hormone: role in the control of tumor growth and progression in humans, *Frontiers in neuroendocrinology* 24(4) (2003) 279-95.
- [62] G. C, E. G, Role of gonadotropin-releasing hormone (GnRH) in ovarian cancer, *Reproductive biology and endocrinology : RB&E* 1 (2003) 65.
- [63] D. AT, C. RA, M. GA, F. JJ, L. CA, W. BR, LH-Independent Testosterone Secretion Is Mediated by the Interaction Between GNRH2 and Its Receptor Within Porcine Testes, *Biology of reproduction* 93(2) (2015) 45.
- [64] C. KC, A. N, L. PC, Expression and antiproliferative effect of a second form of gonadotropin-releasing hormone in normal and neoplastic ovarian surface epithelial cells, *The Journal of clinical endocrinology and metabolism* 86(10) (2001) 5075-8.
- [65] G. C, G. AR, M. RP, E. G, Expression of gonadotropin-releasing hormone II (GnRH-II) receptor in human endometrial and ovarian cancer cells and effects of GnRH-II on tumor cell proliferation, *The Journal of clinical endocrinology and metabolism* 87(3) (2002) 1427-30.
- [66] C. A, K. E, R. S, B.-A. N, O. E, K. Y, Two forms of gonadotropin-releasing hormone (GnRH) are expressed in human breast tissue and overexpressed in breast cancer: a putative mechanism for the antiproliferative effect of GnRH by down-regulation of acidic ribosomal phosphoproteins P1 and P2, *Cancer research* 62(4) (2002) 1036-44.
- [67] S. J, W. T, v.R. S, M. DH, G. L, S.-E. M, C. WT, C. BS, K. AM, B. XO, Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers, *Nature cell biology* 10(12) (2008) 1470-6.
- [68] H. IS, K. C, C. AP, L. PC, Gonadotropin-releasing hormone-I or -II interacts with IGF-I/Akt but not connexin 43 in human granulosa cell apoptosis, *The Journal of clinical endocrinology and metabolism* 97(2) (2012) 525-34.
- [69] K. DK, Y. JS, M. K, H. JI, K. K, S. D, A. Y, L. C, K. BC, K. HB, C. J, S. JY, A gonadotropin-releasing hormone-II antagonist induces autophagy of prostate cancer cells, *Cancer research* 69(3) (2009) 923-31.
- [70] E. N, G. AR, E. G, G. C, GnRH-II agonist [D-Lys6]GnRH-II inhibits the EGF-induced mitogenic signal transduction in human endometrial and ovarian cancer cells, *International journal of oncology* 29(5) (2006) 1223-9.
- [71] V. TH, W. Z, Matrix metalloproteinases: effectors of development and normal physiology, *Genes & development* 14(17) (2000) 2123-33.
- [72] S. M, F. N, T. A, K. E, D. T, I. J, T. K, Y. A, K. S, U. T, E. M, The Association of Polymorphisms in the Gene Encoding Gonadotropin-Releasing Hormone with Serum Testosterone Level during Androgen Deprivation

Figures

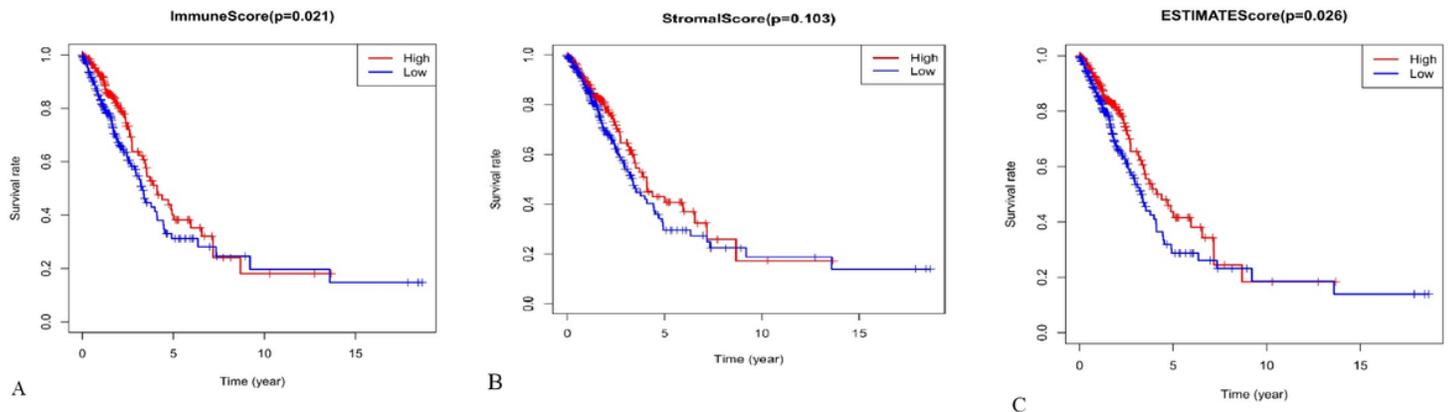


Figure 1

The relationships between stromal score, immune score, estimate score and survival. (A,C) High immune score and estimate score were correlated with a high survival rate (p=0.0207 and p=0.0257). (B) The effects in stromal score group was insignificant (p=0.1030).

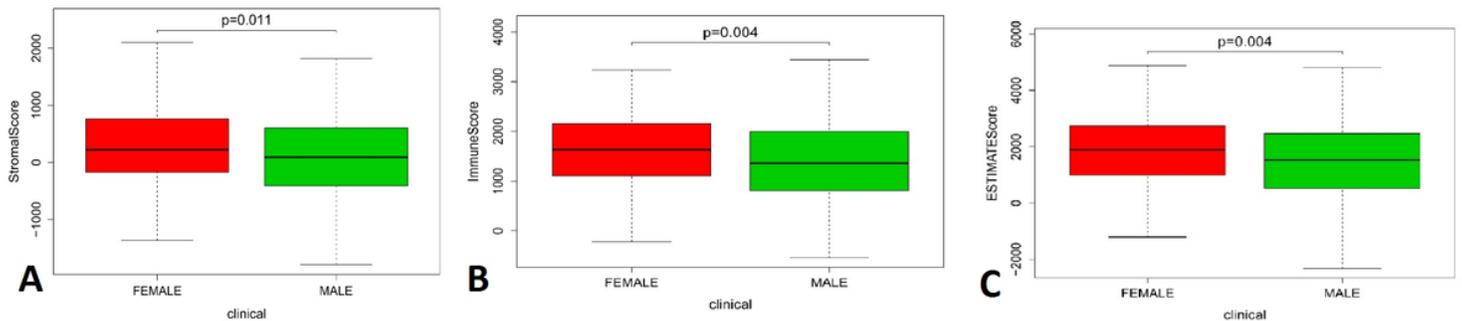


Figure 2

The relationship between sex and survival. (A-C) The stromal score, immune score and estimate score of females were all higher than those of males (p=0.1081, p=0.0360 and p=0.00362).

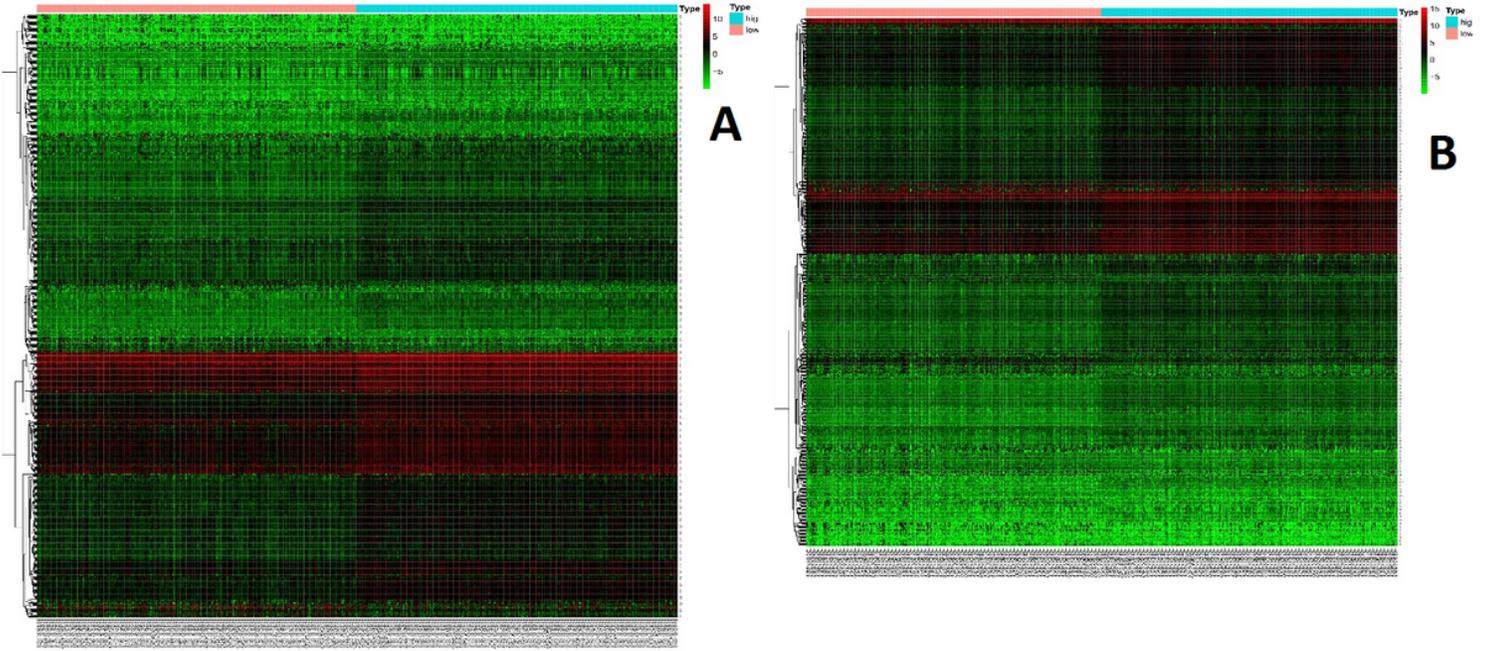


Figure 3

Heat map was plotted based on stromal score and immune score. (A,B) The pink represents the low score group, while the blue represents the high score group. Low expression is denoted in green, median expression is represented in black, while high expression is in red.

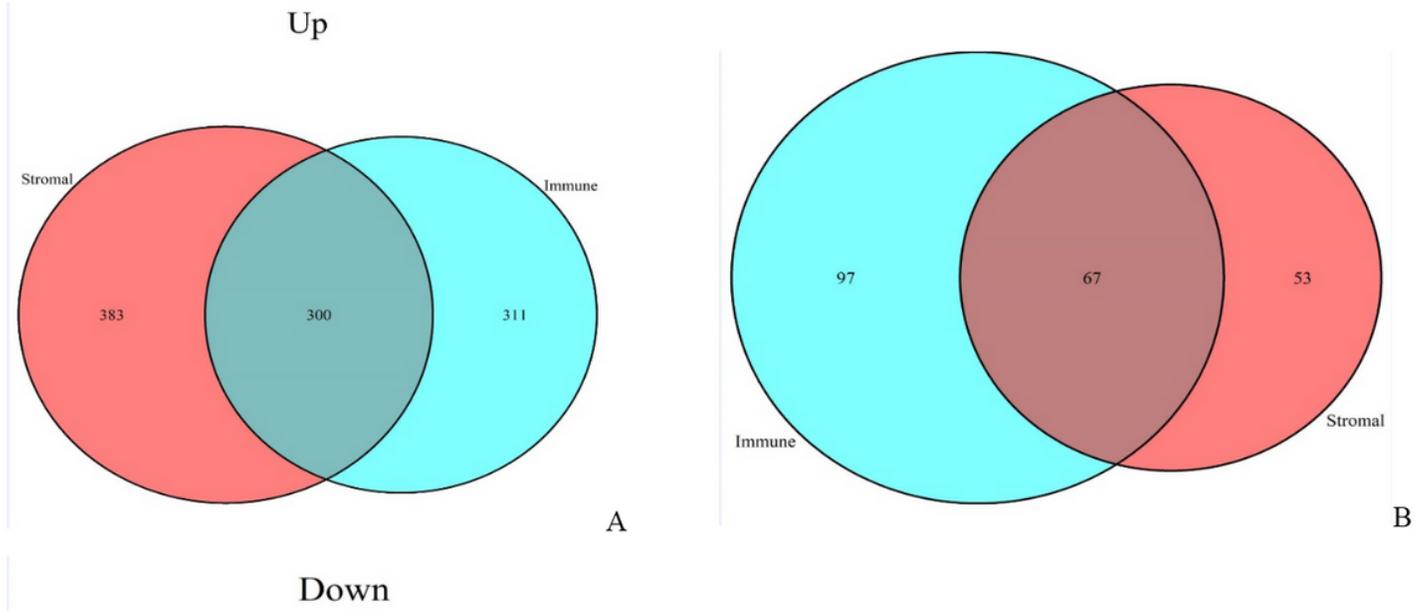


Figure 4

(A,B) 300 up-regulated common genes and 67 down-regulated common genes were screened out.

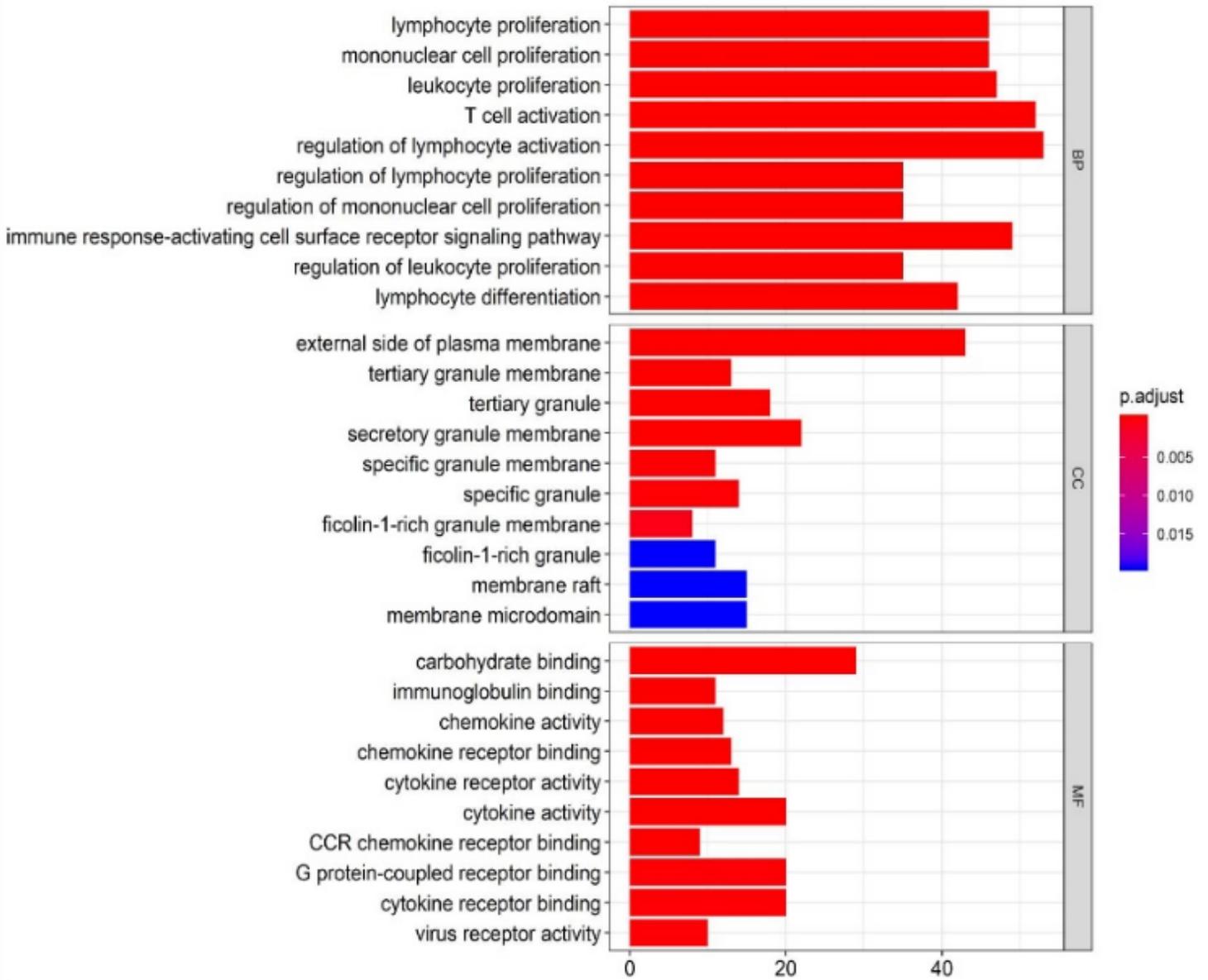


Figure 5

gene ontology (GO) enrichment analysis suggested a close relationships between differentially expressed genes (DEGs) and immune response. T cell activation, regulation of lymphocyte proliferation, immune response-activating cell surface receptor signaling pathway and leukocyte proliferation were highly enriched($p < 0.05$).

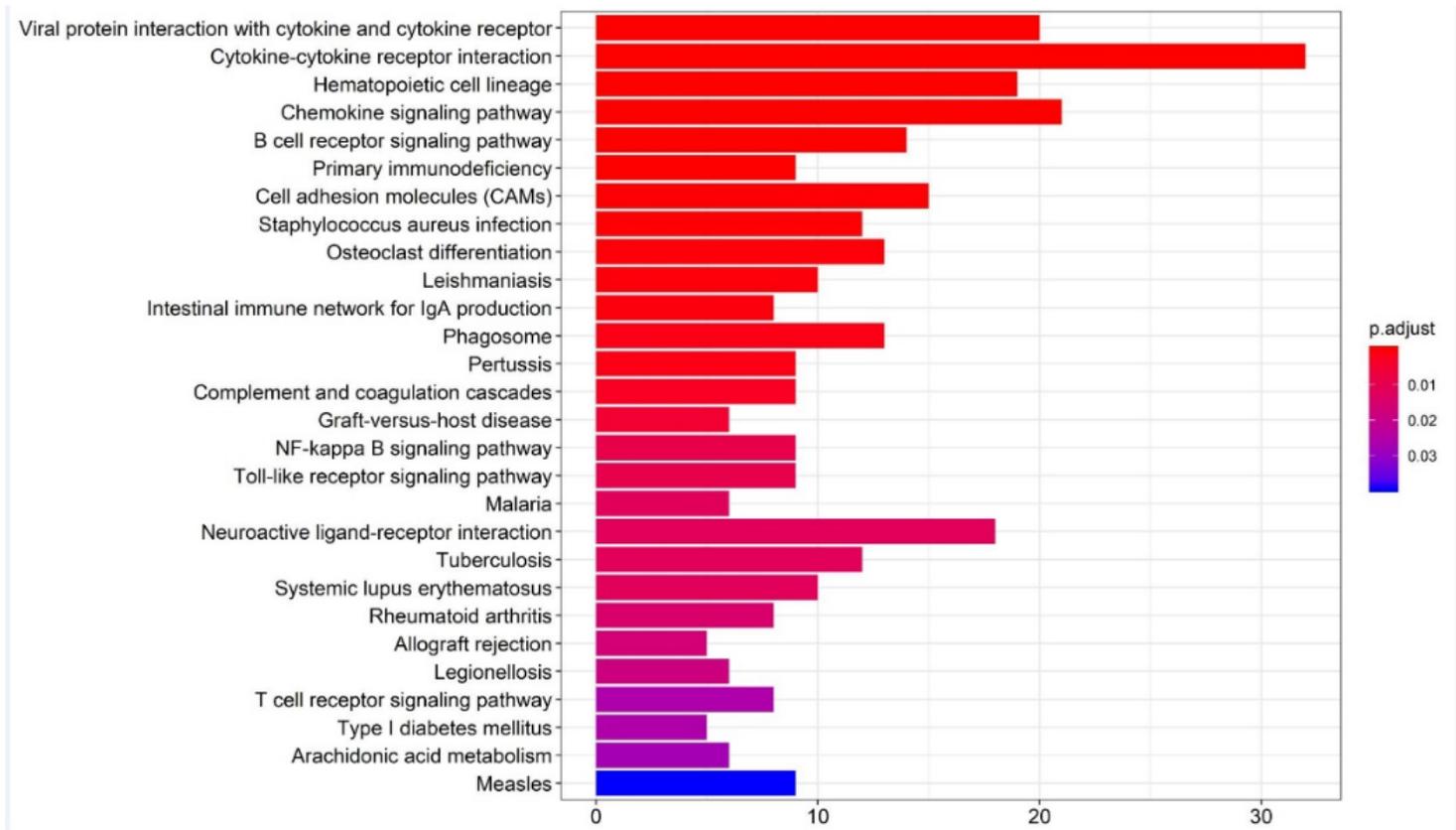


Figure 6

In kyoto encyclopedia of genes and genomes (KEGG) enrichment analysis, cytokine-cytokine receptor interaction, chemokine signaling pathway, viral protein interaction with cytokine and cytokine receptor and hematopoietic cell lineage were most highly up-regulated ($p < 0.05$).

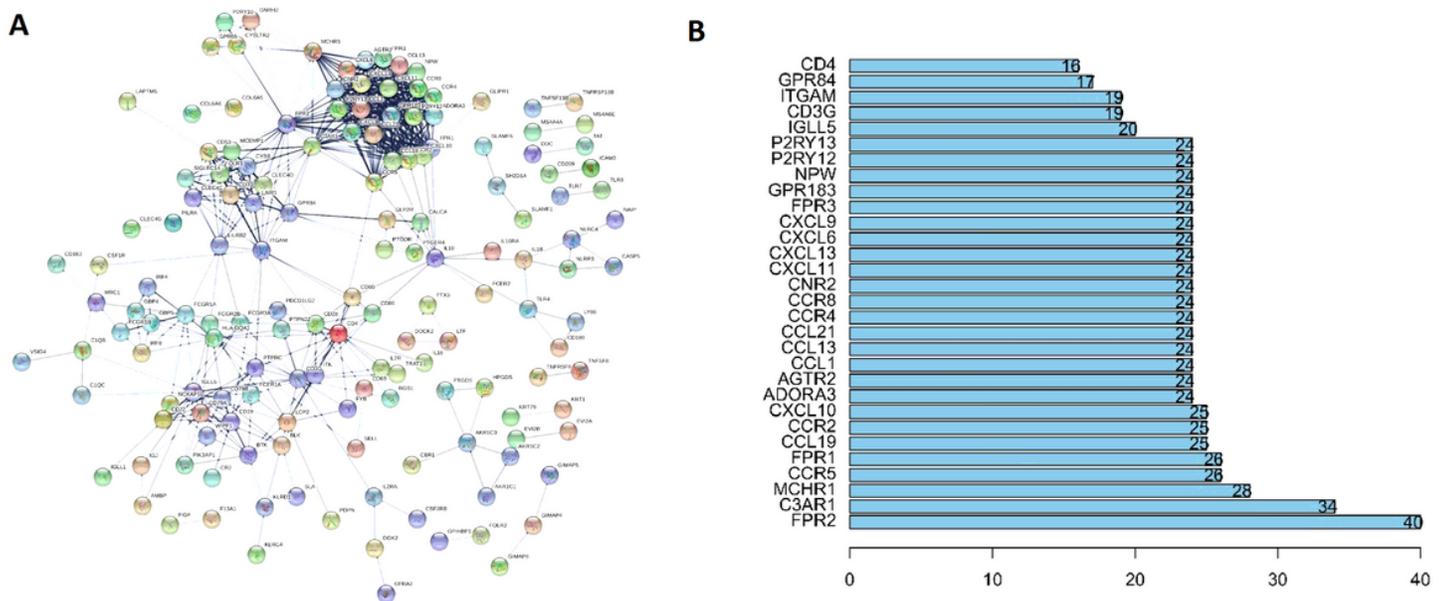


Figure 7

(A) Protein-protein interaction (PPI) network covered 152 nodes and 612 edges. (B) The top 30 most connected genes included FPR2, C3AR1, MCHR1, CCR5, FPR1, CCL19, CCR2, CXCL10, ADORA3, AGTR2, CCL1, CCL13, CCL21, CCR4, CCR8, CNR2, CXCL11, CXCL13, CXCL6, CXCL9, FPR3, GPR183, NPW, P2RY12, P2RY13, IGLL5, CD3G, ITGAM, GPR84 as well as CD4.

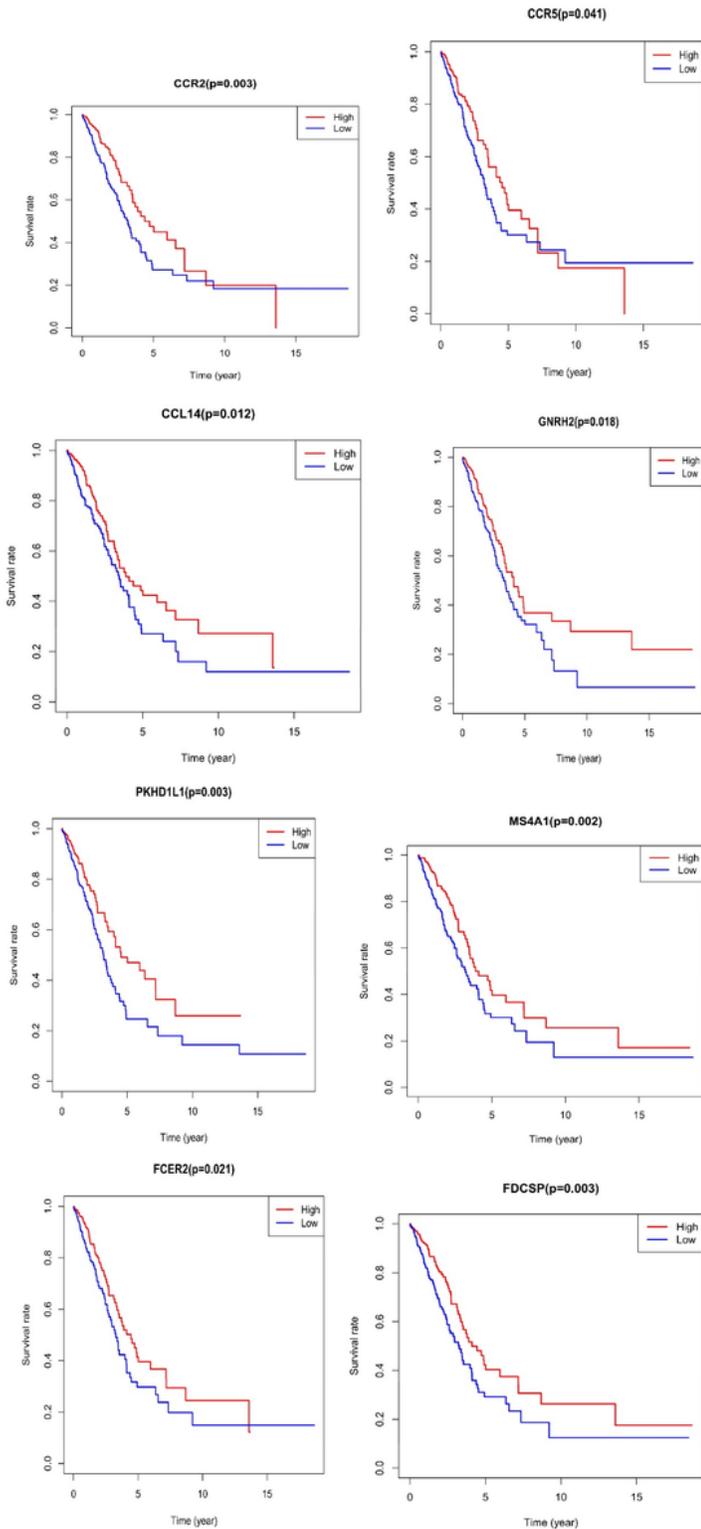


Figure 8

The relationships between CCR5, CCR2, CCL14, GNRH2, PKHD1L1, MS4A1, FCER2 ,FDCSP and survival($p < 0.05$).