

# Transient production of Receptor-Binding Domain of SARS-CoV-2 in *Nicotiana benthamiana* plants induces specific antibodies in immunized mice

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## Research Article

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# Abstract

The COVID-19 pandemic caused by the SARS-CoV-2 coronavirus has currently affected millions of people around the world. To combat the rapid spread of COVID-19 there is an urgent need to implement technological platforms for the production of vaccines, drugs and diagnostic systems by the scientific community and pharmaceutical companies. The SARS-CoV-2 virus enters the cells by the interaction between the receptor-binding domain (RBD) present in the viral surface spike protein and its human receptor ACE2. The RBD protein is therefore considered as the target for potential subunit-based vaccines. Here, we evaluate the use of *Nicotiana benthamiana* plants as the host to transiently-producing RBDr protein. The identity of the plant-produced RBDr was confirmed by immune assays and mass spectrometry. Immunogenicity was confirmed through the specific antibodies generated in all of the immunized mice compared to the PBS treated group. These findings support the use of plants as an antigen expression system for the rapid development of vaccine candidates.

## Introduction

The infectious disease COVID-19 produced by the SARS-CoV-2 coronavirus keeps the vast majority of countries on alert, due to the high mortality caused by this disease (1). Its rapid spread between continents has led the World Health Organization to declare it a pandemic, <https://www.who.int>. This has resulted not only in the development of therapies and quick affordable diagnostic systems, but also the rapid search for effective vaccines. Several vaccines and vaccine candidates are now being used and evaluated in clinical trials involving platforms based on nucleic acids, viral vectors and subunit vaccines (2).

SARS-CoV-2 is a virus enveloped with RNA genetic material, appearing like a crown, due to the presence of the S surface glycoprotein. This protein has two subunits known as S1 and S2. The amino acid sequence between the 319-591 residues of S1, corresponding to the receptor binding domain (RBD), has a central role in the viral infection. The RBD has eight cysteines and two N-glycosylation sites (N<sub>331</sub> and N<sub>343</sub>). The inclusion of glycans can play a relevant role in vivo, in the folding, dynamics, stability and accessibility of the solvents by the RBD (3). The receptor of this virus is the angiotensin converting enzyme 2 (hACE2), found on the external surface of the membrane of several types of human cells, forming the RBD-hACE2 complex. Consequently, the viral particle enters the host cell and produces the release of its genetic material in the cytoplasm of the infected cell (4). The above reasons have led to consider the RBD as a protein candidate for the development of a subunit vaccine.

There have been diverse biotechnological platforms for drug production, but most of the drugs and recombinant vaccines have been produced in mammal or microbial cells. These productive systems have large advantages, but also disadvantages such as their high production costs (5) leading to alternative production settings that may reduce costs significantly. Plants then arise as production systems for recombinant proteins, offering many advantages over conventional platforms, which include its simple cultivation, low cost, the uncomplicated increase in production scale, among others (6). It has also been shown that they can assemble simple molecules, such as antibody fragments, to produce more complex ones. The plants, as bioreactors, can also produce drugs in a stable or transitory manner, where the latter is a strong tool for rapidly obtaining heterologous proteins of pharmaceutical importance (7).

In this study, we describe the production of the RBD protein of SARS-CoV-2 in *N. benthamiana*, as a rapid and scalable alternative for making this antigen. The integrity and identity of the protein was confirmed by immunoblotting and mass spectrometry assays. Finally, we discuss the capacity of the RBD produced in *N. benthamiana* plants to generate a specific immune response.

## Materials And Methods

### Plant material and growing conditions

*N. benthamiana* seeds were grown on the TerraPlant<sup>R</sup> 2 substrate, in floating trays soaked with water under controlled conditions (25 °C with photoperiods of 16h light/8h darkness) to obtain plants of approximately 7 weeks of age. Every 15 days we sprinkled the foliar fertilizer Bayfolan® S- Bayer.

### **RBD recombinant variants as model antigens**

For western blot analysis his-RBD variant produced in *E. coli* (8) was used as positive control. Human ACE2 receptor (hACE2) and chimeric protein hFc-RBD-HRP were supplied by the Center of Molecular Immunology, Havana, Cuba. RBD produced in *Pichia pastoris* (9) was used as a positive control in the hACE2 inhibition assay.

### **Construction of the expression vector of the RBD**

The nucleotide sequence (amino acids 331-530 of the Spike protein from SARS-CoV-2) coding the RBD region that carried six histidine amino acids at the N-terminal end was extracted from the PET-28 plasmid (8). It was inserted in a plant expression vector, pCambiaHT, using the *Bam*HI restriction sites, and flanked by the 5' and 3' non-translatable (UTR) sequences of the Cowpea Mosaic Virus. The 35S promoter of the Cauliflower Mosaic Virus and the tNos nopaline synthase terminator were the signals used for starting and ending the transcription, respectively. The genetic design considered the fusion of the peptide signal of the sporamin from sweet potato (*Ipomoea batatas* L.), at the N-terminal end of the gene of interest. As a result, we obtained the binary vector pCambiahis-RBDapo (Fig. 1a), used in *N.benthamiana* transient transformation assays.

### **Transitory expression of the *N. benthamiana* leaves**

*Agrobacterium tumefaciens* strain GV3101 was individually transformed with the expression vector pCambiahis-RBDapo and pCambiaP19, by heat shock (Fig. 1a). The resulting strains were checked by PCR and cultivated in the YEB medium (lab-lemco 4g/L, saccharose 5g/L, lacto-pectone 5g/L, yeast extract 1g/L, MgSO<sub>4</sub> 2 mM, pH 7.2 ) supplemented with 50 mg/mL of rifampicin and kanamycin, while stirring at 200 rpm for 16h at 28 °C. We later transferred them to a YEB medium without antibiotics, under the same conditions where they reached the optical density (OD) of 0.6 to 0.7 at 600 nm. Cell were harvested by centrifugation at 3000 rpm for 30 min and each bacterial pellet was resuspended in the Murashige-Skoog liquid medium (Sigma, USA), with 30 mg/mL of acetosyringone (Sigma-Aldrich, USA) and then incubated at room temperature while slowly stirring in the dark for 4h.

The vacuum infiltration protocol was performed following the process shown in Fig. 1b, using a final solution, resulting from the mixture of the two bacterial solutions (1:1 proportion). At 7 weeks of growth, we took the plants from the floating tray and submerged them, with their roots upward, in the mixture of the *Agrobacterium*, within a vacuum chamber. The agro-infiltration procedure had 2 cycles consisting of 1 min of vacuum and 1 min of decompression each. Afterwards, we rinsed the treated plants with water to detach the excess bacteria, and they were incubated in water for 5 days at 23 °C with photoperiods of 16h of light and 8h of darkness in glass jars. Agro-infiltrated plants with pCambiaP19 were used as negative control (+P19).

### **RBDr expression and extraction**

The 5 day-post-infiltrated leaves (5dpi) were harvested and total soluble proteins were extracted (10). To determine the levels of expression of the RBDr we used an enzyme-linked immunosorbent assay (ELISA). Plates (Nunc MaxiSorp™) were coated with 10 µg/mL of the monoclonal antibody anti-RBD (AcM-RBD) produced at the CIGB, Sancti Spíritus, Cuba, and incubated for 16h at 4 °C. The plate was blocked with 5% skim milk in PBS-T, for 2h at 37 °C. In order to quantify the RBDr expression, (serial dilutions starting from 1:100) of *N. benthamiana* protein extracts containing RBDr were added to plate and kept it for 2h at 37 °C. As a second antibody was used monoclonal anti-poly-histidine, produced in mice and conjugated to the horseradish peroxidase (mAb-his-HRP, 1:2000, catalog A7058 Sigma-Aldrich), incubated for 1h at 37 °C. Intermediate washing were established between each step with PBS-T. In the quantification were used a standard curve of RBD produced in *Pichia pastoris*. As negatives controls were utilized crude extract +P19 and PBS. The reaction was detected after the

addition of 3,3',5,5'-tetramethylbenzidine and quantified using a microplate reader at 450 nm using a spectrophotometer (Thermo Scientific UV-Vis, USA).

### **RBD<sub>r</sub> antigen purification**

Before starting the purification, TSP was clarified by centrifugation at 9000 rpm for 20 min and the supernatant was filtrated with a membrane of 0.22µm (Sartorius Minisart, Germany). Recombinant His-tagged proteins were then purified by Immobilized Metal Affinity Chromatography (IMAC) using the procedure (9) with modifications described in the caption of Fig. 2. This fraction was concentrated through centricom 3 kDa (Millipore, USA). The protein concentration of the all fractions were determined by the Bradford method (11).

Samples from the fractions of the RBD<sub>r</sub> purification process were assayed in a 12% acrylamide SDS-PAGE stained with coomassie Blue G25 (Applichem, Germany). For Western blotting, the mAb-his-HRP was used to detect the his-tag of RBD<sub>r</sub>. his-RBD, a variant of RBD produced in *E.coli*, was used as a positive control. The substrate used for the colorimetric detection of the assay was diaminobenzidine (Sigma, USA).

### **Electrospray ionization mass spectrometry (ESI-MS) analysis**

The RBD<sub>r</sub> purified by IMAC was concentrated and separated using SDS-PAGE in polyacrylamide gel at 12.5% under reducing conditions. The bands of protein corresponding to RBD<sub>r</sub> were cut from the gel and divided into small cubes of approximately 1 mm<sup>3</sup>. The gel cubes were treated to obtain tryptic peptides. The peptide mixture was analyzed with a QToF-2 hybrid tandem mass spectrometer (Micromass, UK) according to (12)

### **Immunogenicity of the RBD<sub>r</sub> in mice**

According to the Institutional Committee for Animal Care and Use of the CIGB, Cuba, the regulations for the use of laboratory animals were followed to study the immunogenicity of RBD<sub>r</sub> in mice. Balb/c mice 6 weeks old were used in the experiment. They were maintained under controlled temperature and lighting, with feed and water *ad libitum*. Two experimental groups were designed, which consisted of nine animals per group for the subcutaneous injection. The first one was the group of interest in which we inoculated 5 µg RBD<sub>r</sub>. A second group identified as the negative control (-RBD) to which we administered the same volume of PBS. The inoculum was mixed with aluminum hydroxide in a 6:1 ratio (antigen/adjuvant). Three doses were assayed, corresponding to 0, 21 and 42 days, while mice bleeding were developed at 0, 21, 42 and 57 days in order to evaluate specific antibody production and their ability to inhibit the binding of RBD to hACE2.

### **Anti RBD<sub>r</sub> titers of immunized mice**

To evaluate the immune response against RBD<sub>r</sub>, the described protocol (9) was followed with the adaptations described below and only up to the plate reading step. Coating with 5 µg/mL of RBD<sub>r</sub> produced in *Pichia pastoris* and only the SS-1 antibody was used. In addition, the RBD<sub>r</sub> and -RBD sera used in the assay were diluted in PBS-T (dilutions from 1:25 to 1:10000).

### **RBD-hACE2 Inhibition Assay**

The capacity of the polyclonal antibodies generated by RBD<sub>r</sub> to inhibit the binding of the hACE2 receptor was evaluated through a competitive ELISA previously reported (9). Mice sera after the last immunization with RBD<sub>r</sub> (T57) were used for the analysis. As positive control was considered the sera of nine volunteers (90% inhibition) immunized with RBD of *P. pastoris* and mice sera immunized with PBS at T57 were used as a negative control. The percentage of inhibition was calculated according to the following formula:

$$\text{Inhibition \%} = (1 - (\text{OD}_{450\text{nm}} \text{ samples preincubated}) / (\text{OD}_{450\text{nm}} \text{ AB})) * 100$$

### **Statistics analysis**

The mean of the experimental values was represented and the standard deviation indicated as error bars. In the analysis of the data, we used the GraphPad Prism software version 8.0.2, with a Kruskal-Wallis test and Dunn's test.

## Results

### Transient transformation of *Nicotiana benthamiana*, expression and purification of RBDr

The strains of *A. tumefaciens* transformed with the genetic constructions represented in the figure 1a were used during the transitory co-transformation of *N. benthamiana* plants. Five days after vacuum infiltration (5dpi), no apparent damages were observed in the architecture of the plants (Fig. 1b). The antigen concentration in the total soluble protein extract was estimated by ELISA, quantifying an expression of 30 µg RBDr per g of leaves (Fig. 1c).

The designed RBDr included a polyhistidine peptide, so the purification included an immobilized metal chelate affinity chromatography step (Fig. 2a). The IMAC fractions were analyzed by SDS-PAGE and Western blot using mAb-his-HRP. In the eluates fractions (lanes 4 and 5) the identity of the recombinant protein was confirmed, as well as in the sample resulting from the final concentration (lane 8, several subpopulations with size between 25- 41 kDa approximately, presumably glycoforms thereof, coexisted). Additionally, bands above 41 kDa were observed in the SDS-PAGE, not detected in the Western blot. No recognition signals were detected in lanes 2 and 3 corresponding to unbound and washed fraction, either in lane 6 corresponding to the initial extract of agroinfiltrated plants only with pCambiaP19 vector (Fig. 2b).

### ESI-MS analysis of the RBDr protein

Mass spectrometry (MS) is the analytical tool of choice to confirm the amino acid sequence, to evaluate the integrity of the N- and C-terminal ends, and to detect posttranslational modifications in natural or recombinant proteins. We analyzed the RBDr through ESI-MS with the goal to confirm the amino acid sequence and to verify the correct assembling of disulfide bonds of RBDr. Sample processing, including an efficient proteolytic digestion, the recovery of the proteolytic peptides and a desalting step prior to ESI-MS analysis, plays a key role to obtain the highest sequence coverage. The ESI-MS spectrum through the RBDr protein digestion is shown in figure 3a. Some of the signals with stronger relative intensities, designed by the blue arrows, were chosen for sequencing. The sequence assignment based on the agreement between the expected and experimental m/z of tryptic peptides are summarized in Table 1. Two of the four-disulfide bonds present in the RBDr, C<sub>379</sub>-C<sub>432</sub> and C<sub>480</sub>-C<sub>488</sub>, were identified. This study confirmed coverage of the sequence of RBDr of 51.3 %, thereby indicating the identity of the antigen (Fig. 3b).

### Murine immunization and serum analysis

Animals subcutaneous administration was carried out with three doses (Fig. 4a) of protein containing 5 µg of RBDr mixed with aluminum hydroxide adjuvant. Through the duration of the experiment, mice showed no signs of disease or adverse effects following vaccination. Sera from mice immunized at times 0, 21, 42 and 57 days were evaluated by ELISA as described in materials and methods. Specific antibody titers were observed in all inoculated animals after the third dose compared to the control sera (-RBD). At 57 days, the highest antibody titers were detected in two of nine immunized mice. Antibody titers were statistically significant on T57 compared to earlier times (indicated with different letters) (Fig. 4b).

The inhibition of the binding of hFc-RBD-HRP to hACE2 mediated by sera from animals immunized with RBDr, suggests a probable decrease of the infective capacity of the virus, and it therefore offers information of the possible usefulness of a vaccine candidate against SARS-CoV-2. Fig. 5c represents inhibition of RBD binding to hACE2 in the presence of sera extracted at T57. Of all the sera tested, two of them showed an inhibition of approximately 90%, similar to the sera referred to as positive control. In the rest of the animals, the mean percentage of inhibition was between 40 and 50%. In contrast, at T0 the sera did not show any inhibitory capacity in the binding to hACE2, thus confirming that this inhibition is due to the anti-RBDr antibodies generated in the mice immunized with the antigen produced in *N. benthamiana*.

## Discussion

The propagation of the novel coronavirus SARS-CoV-2, the cause of COVID-19, has become the center of attention worldwide because of the levels of infection and mortality (13). This virus is genetically related to SARS-CoV, and it is characterized by a more rapid spread than the rest of the coronaviruses. It can infect cells of the lung, kidney, heart and intestine, among other tissues, producing the multiple organ dysfunction syndrome (MODS) (14). Among the treatment alternatives to survive and control the virus infection, SARS-CoV-2 specific antiviral drugs, redirected drugs and specific vaccines against the SARS-CoV-2 virus have been assayed (15).

In recent years, plants have been considered as a powerful expression system for the production of therapeutic proteins (16) that show similar, or even greater biological activity than the homologous proteins expressed in cultured mammalian cells (17). In this respect, we used in this study the transitory transformation of *N. benthamiana* plants due to the advantages it can offer in terms of speed, versatility and ease of agroinfiltration. Previous studies have demonstrated the production of SARS-CoV-2 related proteins in *N. benthamiana*. Recent studies estimated between 2-4  $\mu\text{g}$  RBD/g fresh weight using the same host as us.(18) Similarly, Rattanapisit obtained 8  $\mu\text{g}$  of such antigen/g fresh weight of leaf from the same plant (19). Also other studies confirm 10  $\mu\text{g}$  (3) and 25  $\mu\text{g}$  (20) of RBD in *N. benthamiana*. The RBDr expression level reported here is in the same order than those were previously informed, in spite of we assayed a platform involving a non-replicative methods by the construction of a vector that carried 5' and 3' UTR, of the Cowpea Mosaic virus, a successful design for the accumulation of other heterologous proteins (21). However, this does not imply that plant made vaccines against SARS-CoV-2 are not a possible alternative, taking into account the performance of clinical trials with vaccine candidates produced in *N. benthamiana* plants (22).

The preparation of the sample for the analysis of mass spectrometry plays a determinant role in the quality of the result. The confirmation of the main part of the amino acid sequence depends on the efficiency of the digestion and the recovery of the tryptic peptides (8). In our case, the digestion in gel under native conditions, made it possible to recover the necessary fragments to confirm 51.3% of the RBDr. Under our conditions, it was possible to identify the links of C<sub>379</sub>-C<sub>432</sub> and C<sub>480</sub>-C<sub>488</sub>. It was impossible to identify the first C<sub>336</sub>-C<sub>361</sub> because these Cys residues are included in the T2 and T6 fragments that could not be sequenced because of the presence of N-glycans, which make the peptides more hydrophilic, and therefore, difficult to recover by RP-HPLC. C<sub>391</sub>-C<sub>525</sub> was not identified because we were unable to sequence the C terminal end. Taking into account that the analysis was performed through a digestion in gel, we observed a greater coverage of sequences compared to other studies using fragments of the gel for the peptide analyses. In this sense, a study of the analysis of the protein extract of HeLa, optimizing six different gel digestion methods, revealed a sequencing coverage of less than 20% (23). In another case, was reached peptide sequencing of 44% by also using gel digestion (24). In contrast, when the variable region of an antibody was purified for its analysis by mass spectrometry, using digestion in solution, the coverage confirmed was of more than 80% of the sequence (25).

Both the humoral and cellular immune response developed with vaccines, have had great relevance in different viral disease protection (26). The doses used in the immunization have a determinant role in immunogenicity (27). In our study, the administration of 5  $\mu\text{g}$  of RBDr brought about the appearance of specific antibodies in all the animals, as of the third injection, compared to animals -RBD. In addition, approximately 22% of the animals immunized with RBD of plant origin developed IgG titers 1:10000 ( $\log_{10}$  4), very similar to the average the titers shown by Siri wattananon (20) in the first immunization but lower with respect to the second immunization evaluated by them with 10  $\mu\text{g}$  of RBD-Fc. It is presumed that the delay in the immunological response could be influenced by the dose used. In that sense, when used 10  $\mu\text{g}$  of RBD, produced in humans cell line, compared to the 5.0 and 2.5  $\mu\text{g}$  doses obtained higher antibodies titers (28). Also, the dose trials related to a vaccine candidate based on the mRNA of RBD in mice, showed that the highest dose (15  $\mu\text{g}$ ) produced the highest titers when compared to 2  $\mu\text{g}$  in the immunized mice (29). Another study showed that, mice injected with hemagglutinin (HA) from the Influenza virus, stimulated lower HA titers when using 3  $\mu\text{g}$  of the inoculum, compared to the highest dose vaccine, which induced higher HA titers (30). In addition, another factor that could influence the generation of

antibodies is the combination of antigens with potent adjuvants that increase the immune response regardless of the dose used (31). This is the case of the virus-like particle of protein S, produced in *N. benthamiana*, as a vaccine candidate that showed high anti-S antibody titers with one of the adjuvants employed (oily emulsion of DL- $\alpha$ -tocopherol and squalene), even when using low dosages of the antigen (22). Taking into account this report, RBD<sub>r</sub> could be combined with another adjuvant to increase the immunogenicity.

The capacity of viral RBD to bind itself to the hACE2 determines the success of the infectivity of SARS-CoV-2. In the protection against COVID-19 the formation of neutralizing antibodies that can inhibit the binding of the viral receptor to hACE2 is crucial (32). In this sense, some reports have shown a correlation between the percentage of inhibition of ACE2 binding, mediated by RBD antibodies, and the generation of neutralizing antibody responses (33). Although the inhibition percentage of the RBD<sub>r</sub> sera to reduce the protein binding to its receptor hACE2 average 40-50% compared with the mice treated with PBS, in this study was confirmed that those animals with the higher RBD-specific IgG titers had inhibition percentage near to 90% (coinciding with the highest IgG titers), indicating that the antigen herein obtained is functional. Similar results were obtained on evaluating a group of monoclonal antibodies with a high affinity against RBD, where they exceptionally demonstrated that five candidates showed more than 90% of inhibition, higher than the rest, with 50% of inhibition (34). Another study, evaluated the immunogenicity of RBD bound to Fc (2.5, 5 and 10  $\mu$ g) produced in human cell lines using Freud's adjuvant; with an immunization schedule similar to ours they obtained 66% of inhibition of the binding to ACE2 (28).

## Conclusions

In this study, we have confirmed the transient production of RBD<sub>r</sub> in *N. benthamiana*. By ESI-MS the identity of this protein and the correct assembling of two of their four disulfide bonds were demonstrated. While this paper does not describe dose studies necessary, the production of specific anti-RBD<sub>r</sub> antibody in mice immunized showed inhibition of binding of RBD to the hACE2 receptor, confirming the immunogenicity of the antigen produced in *N. benthamiana*, in spite of the low dosage assayed. This study suggests the transient expression *N. benthamiana* as a tool to assemble and present the correctly folded antigens to the immune system that could be used as vaccines candidates against SARS-CoV-2 virus which could be particularly useful when new variants of the virus have emerged.

## Declarations

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Y.C, A.L, A.H, A.E.G. design of the genetic construction, expression and purification of the protein. G.C. N-glycosylation analysis. I A. mass spectrometric analysis. Y.C, R.U.M. RBD<sub>r</sub> immunogenicity assays. Y.C. writing the manuscript. All authors contributed to data processing.

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**Conflict of Interest** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Availability of data and material** The data obtained during the investigation were included in the manuscript.

**Code availability** Not applicable

**Ethics approval** The experiments in animals were controlled by the Institutional Committee for Animal Use and Care of the CIGB, Cuba.

**Consent to participate** Informed consent was obtained from all individual participants included in the study.all individual participants included in the study.

**Consent for publication** The authors have given their consent to the disclosure of this study in the journal.

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## Tables

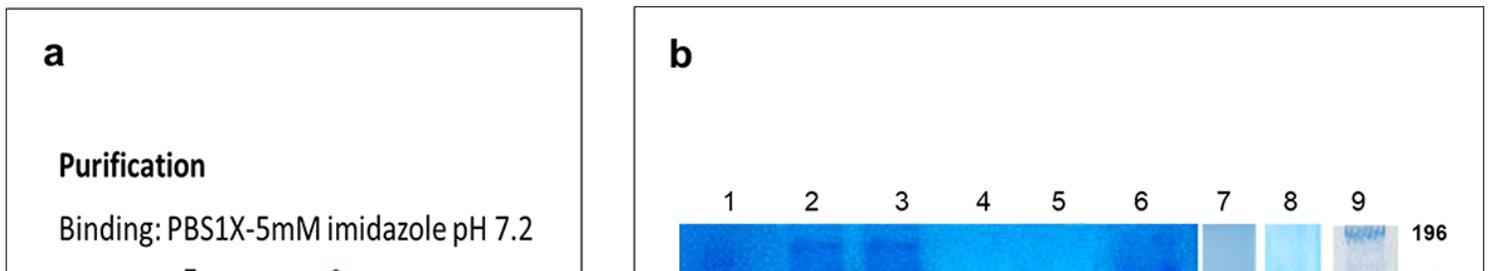
**Table 1** Representation of theoretical RBD peptides. Those marked with blue letters are the sequenced fragments that belonged to RBDr

Frg #	Res #	Sequence	Theor (Bo)	(M+H)	(M+2H)	(M+3H)	(M+4H)	(M+5H)
T1	1-17	MGSSHHHHHHSSGLVPR(G)	1898.88	1899.89	950.45	633.97	475.73	380.78
T2	18-39	(R)GSHMASNITNLCPFGEVFNATR(F)	4678.16	4679.16	2340.09	1560.39	1170.55	936.64
T6	51-71	(R)ISNCVADYSVLYNSASF						
<b>T3</b>	<b>40-48</b>	<b>(R)FASVYAWNR(K)</b>	<b>1112.54</b>	<b>1113.55</b>	<b>557.28</b>	<b>371.85</b>	<b>279.14</b>	<b>223.52</b>
<b>T4</b>	<b>49-49</b>	<b>(R)K(R)</b>	<b>146.11</b>	<b>147.11</b>	<b>74.06</b>	<b>49.71</b>	<b>37.53</b>	<b>30.23</b>
T5	50-50	(K)R(I)	174.11	175.12	88.06	59.05	44.54	35.83
<b>T7</b>	<b>72-79</b>	<b>(K)CYGVSPTK(L)</b>	<b>3059.40</b>	<b>3060.41</b>	<b>1530.71</b>	<b>1020.81</b>	<b>765.86</b>	<b>612.89</b>
<b>T12</b>	<b>118-137</b>	<b>(K)LPDDETGCVLAWNSNNLDSK(V)</b>						
<b>T8</b>	<b>80-96</b>	<b>(K)LNDLCFTNVYADSFVIR(G)</b>	<b>3966.04</b>	<b>3967.05</b>	<b>1984.03</b>	<b>1323.02</b>	<b>992.52</b>	<b>794.22</b>
T18	203-221	(R)VVVLSFELLHAPATVCGPK(K)						
T9	97-101	(R)GDEVR(Q)	574.27	575.28	288.14	192.43	144.58	115.86
<b>T10</b>	<b>102-110</b>	<b>(R)QIAPGQTGK(I)</b>	<b>898.49</b>	<b>899.50</b>	<b>450.25</b>	<b>300.50</b>	<b>225.63</b>	<b>180.71</b>
<b>T11</b>	<b>111-117</b>	<b>(K)IADYNYK(L)</b>	<b>885.42</b>	<b>886.43</b>	<b>443.42</b>	<b>296.15</b>	<b>222.36</b>	<b>178.09</b>
<b>T13</b>	<b>138-147</b>	<b>(K)VGGNYNYLYR(L)</b>	<b>1217.58</b>	<b>1218.59</b>	<b>609.80</b>	<b>406.87</b>	<b>305.40</b>	<b>244.52</b>
T14	148-150	(R)LFR(K)	434.26	435.27	218.14	145.76	109.57	87.86
T15	151-151	(R)K(S)	146.11	147.11	74.06	49.71	37.53	30.23
<b>T16</b>	<b>152-159</b>	<b>(K)SNLKPFER(D)</b>	<b>989.53</b>	<b>990.54</b>	<b>495.77</b>	<b>330.85</b>	<b>248.39</b>	<b>198.91</b>
<b>T17</b>	<b>160-202</b>	<b>(R)DISTEIYQAGSTPCNGVEGFNCY FPLQSYGFQPTNGVGYQP(YR)</b>	<b>4765.12</b>	<b>4766.12</b>	<b>2383.57</b>	<b>1589.38</b>	<b>1192.29</b>	<b>954.03</b>
T19	222-222	(K)K(-)	146.11	147.11	74.06	49.71	37.53	30.23

## Figures

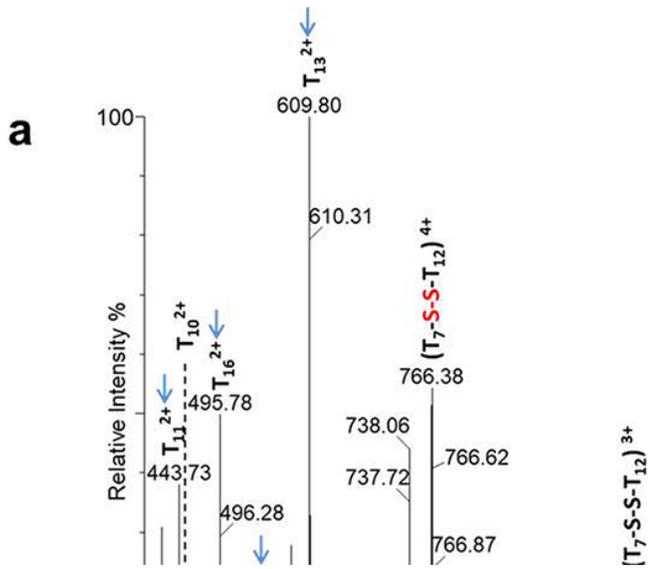
## Figure 1

Schematic representation of transitory transformation in plants of *N. benthamiana*. **a** Genetic construction of RBDr. 35S: promoter of the Cauliflower Mosaic Virus; 5' and 3' UTR: non-translational regions of the Cowpea Mosaic Virus; sp: sweet potato sporamin signal peptide; 6his: sequence coding for 6 histidines; tNos: nopaline synthase terminator; RBD: sequence of the receptor binding domain contained in the spike protein of the SARS-CoV-2 virus; P19: gene coding for RNA silencing suppressor p19 from Tomato Bushy Stunt Virus; RB and LB: right and left border, respectively, flanking the transcriptional unit. **b** Diagram of the vacuum agroinfiltration protocol. **c** RBDr expression levels



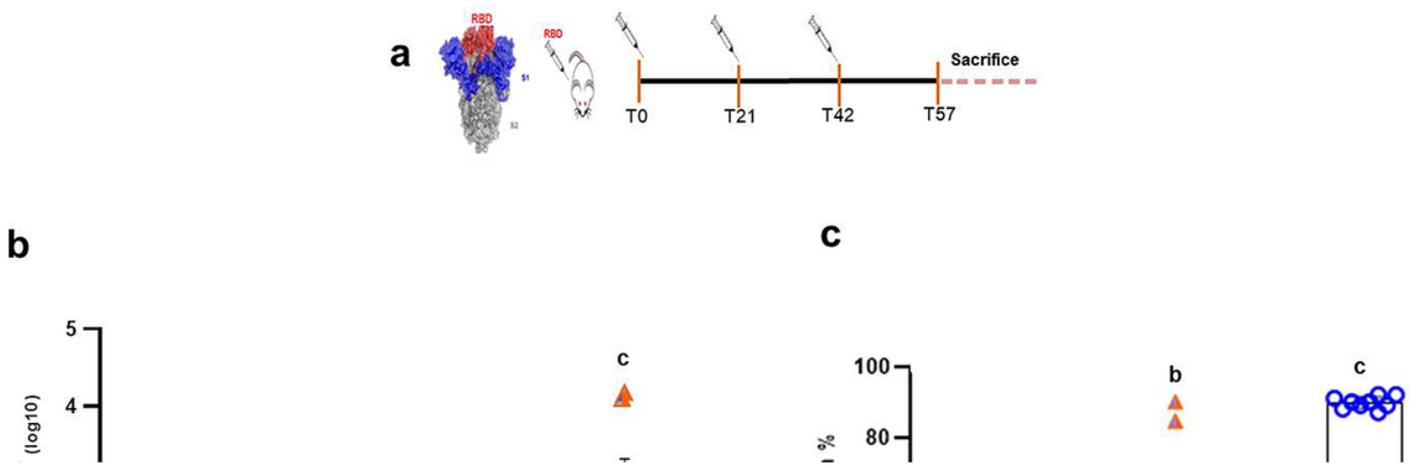
## Figure 2

Purification of RBDr from *N. benthamiana* leaves agroinfiltrated. **a** Schematic representation of recombinant antigen purification process. **b** Stained 12% SDS-PAGE loaded with 10 µg in line 1,2,3,6 ; 200 ng in 4,7 and 1 µg in 5,8. Bottom, Western blot using the anti-histidine monoclonal antibody conjugated with peroxidase of the fractions obtained during purification; 1: TSP extract containing the RBDr; 2: matrix unbound fraction; 3: washing of the process at 70mM of imidazole; 4 and 5: Elution of the RBDr at 500mM; 6: + P19 leaves extract; 7: RBD produced and purified from *E.coli*; 8: eluate of concentrated RBDr in centricon (3.0 MWCO, Sartorius); MW: Prestained SDS-PAGE standards, Broad Range, cat # 161-0318 (BIO-RAD, USA); simple arrow indicates the band corresponding to the RBDr protein and the double arrow is *E. coli* his-RBD



**Figure 3**

RBDr characterization through mass spectrometry. **a** ESI-MS spectrum through the digestion of the RBDr protein in polyacrylamide gel. The identified disulfide bonds between the cysteines ( $C_{379}-C_{432}$  and  $C_{480}-C_{488}$ ) are highlighted in red as -S-S-. The signs marked with blue arrows were the peptides that were analyzed with the MASCOT software as the criterion for the identity of the RBDr protein in the databases of the sequences. **b** Amino acid sequence of the RBDr protein. The residues highlighted in brown, correspond to the 4 amino acids contributed by the sub-cloning vector. The green colour highlights the six residues of the histidine in tandem for their purification by affinity chromatography using immobilized metal chelates. Those highlighted in blue are the residues of the spacer arm, and those in black correspond to the RBD theoretical sequence that includes the identified amino acids sequence in violet. Red letters are the cysteine residues. The red lines represent the disulfide bonds that link to cysteines,  $C_{336}-C_{361}$ ,  $C_{379}-C_{432}$ ,  $C_{480}-C_{488}$  and  $C_{391}-C_{525}$ . The asterisks represent the identified bonds



**Figure 4**

Titration of antibodies in mice immunized with RBDr produced in *N. benthamiana*. **a** Immunization schedule. From T0 to T42 we performed the administration of the recombinant antigen. The syringes (filled in red) throughout the entire period, show the time of the blood extraction of the inoculated animals. **b** Response in immunized mice in each trial period. The values represent the antibodies titers expressed as the base 10 logarithm. +RBDr: experimental group inoculated with the recombinant antigen produced in plants; -RBD: mice immunized with PBS; +RBDhCD: mice immunized with RBD from SARS-CoV-2 linked to the human CD immunomodulatory transiently produced in HEK293 mammalian cells. Graph show mean  $\pm$  standard deviation (SD) for  $n = 9$  mice/group. **c** Binding inhibition capacity of polyclonal antibodies RBDr to hACE2. +RBDr: sera of immunized mice with RBDr, C+ sera of persons immunized with RBD produced in *P. pastoris*, that showed high percentages of inhibition and taken as the positive control; -RBD: mice immunized with PBS. For the evaluation, we chose all sera at time T57. We observed two sera with high inhibition capacity, similar to those used as the positive control. Data are mean  $\pm$  SD. from  $n = 9$  repetitions/groups. The different letters represent significant differences for  $P < 0.0001$