

# Urea Amendment Alleviated Morphological and Physiological Damages and Yield Loss of Winter Wheat Subjected to Cold Stress at Jointing Stage

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## Research Article

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# Abstract

Cold stress in spring is an abiotic stress limiting the growth and productivity of winter wheat. A controlled pot experiment was done to explore the possibility of applying urea to alleviate the low temperature ( $3^{\circ}\text{C}/-4^{\circ}\text{C}$ , day/night) damage to wheat cultivar Yangmai 16 at jointing stage. Urea at different rates was applied at the 5th day after cold stress. Cold stress decreased grain yield and plant height. Compared with the unstressed control, the content of soluble sugar, proline, zeatin riboside (ZR), and abscisic acid (ABA) in the leaves was increased under longer cold stress. These above parameters in the stressed treatments without urea amendment were higher than those with urea amendment on the 10th day. The change in  $\text{GA}_3$  concentration was opposite to the concentrations of ABA and ZR. The decline in the concentrations of osmotic adjustment substances, balanced hormone concentrations, and increased grain number per ear and ear number were the main reasons for the increased grain yield after urea application. Remedial effects was enhanced with the higher urea remedial level under the same cold stress duration. Our study suggested that suitable urea remedial rates are recommended based on the freezing index in wheat to alleviate the impacts of low temperature on wheat production at jointing stage.

## 1. Introduction

Global warming is not only increasing average global temperature but also dramatically increasing the frequency and strength of severe cold days. The increasing occurrence of severe cold weather has become a restricting factor limiting crop production in many parts of the world (Neilson et al. 2010; FAO 2020). As one of the most important staple crops with wide global distribution, winter wheat is prone to severe cold injury during the growth stages, including germination, jointing, booting, and flowering (Fuller et al. 2007; Frederiks et al. 2015; Li et al. 2016; Ji et al. 2017; Flores et al. 2021).

China is one of the largest winter wheat producers in the world. However, low temperature stress has become a general issue for winter wheat growth during winter and early spring seasons, especially in the Huanghuai region and the middle and lower reaches of the Yangtze River. These areas include Henan, Shandong, Hebei, Anhui, and Jiangsu, which are the major wheat production provinces in China. Over the past decades, severe losses in wheat production have occurred due to low temperature. In Henan Province and Shandong Province, severe frost occurs frequently, with an occurrence frequency exceeding 30%. Alone in Henan Province, the occurrence frequency of frost stress on winter wheat has reached more than 50% in a 50-year period from 1964 to 2014 (Zhu et al. 2018). The wheat area suffering from frost injury was about 26.7 million ha in Henan Province in the 2004–2005 wheat season and about annual mean 0.22–0.45 million ha in Anhui Province over the 1986–2017 wheat seasons (Luo et al. 2011; Chen et al. 2020).

Low temperature in spring often has negative effects on wheat grain yield formation. In Henan province, wheat yield decreased by 19.9% when low temperature happened at jointing stage and reduced by 8.9% when low temperature happened at booting stage (Gao et al. 2015). It was reported that 3.1–56.4% yield reduction was recorded for two wheat cultivars treated with low-temperature at jointing stage (Ji et al. 2017). Therefore, freezing injury occurred in spring significantly limits the growth and yield of winter wheat.

The wheat plants under low temperature may undergo a series of morphological and physiological changes, which consequently inhibit plant growth and yield development. Limin and Fowler (2000) reported that low temperature markedly decreased carbohydrate production and accumulation. Low temperature stress also induced the overproduction of reactive oxygen species (ROS) such as hydrogen peroxide ( $\text{H}_2\text{O}_2$ ), superoxide radical, and singlet oxygen as well as hydroxyl radical (Alscher et al. 1997; Dai et al. 2009; Yu et al. 2020). Improved activity of guaiacol peroxidase and catalase can remove excess  $\text{H}_2\text{O}_2$  (Horvath et al. 2007; Yang et al. 2013; Valitovaa et al. 2019). Low temperature stress resulted in changes in proline content, sugar composition and accumulation, as well as hormonal levels and balance. Li et al. (2015) observed that proline and sugar were associated with typical stress responses and the overproduction of proline and sugar enhanced cold tolerance (Patton et al. 2007; Dörffling et al. 2009; Majláth et al. 2012).

Low temperature also caused the changes in endogenous hormones in plants (Penfield 2008). It has been reported that auxin participates in stress responses and functions by up-regulating or down-regulating a group of primary responsive genes to some extent (Majláth et al. 2012; Kalapos et al. 2016). Changes in the levels of hormones in wheat, such as cytokinins (CKs), indole-3-acetic acid (IAA), and nitric oxide, illustrate that wheat plants attempt to maintain growth during cold stress (Majláth et al. 2012; Yu. et al. 2020). Different strategies have been developed to combat the damages caused by low temperature stresses in wheat production (Li et al. 2013; Hussain et al. 2018). Nitrogen application is one of feasible remedial practices. But there is little information on the application of nitrogen on winter wheat after low temperature stress. We hypothesized that nitrogen amendment might be a feasible way to alleviate and combat the inhibitive effects of low temperature stress on winter wheat at jointing stage.

Therefore, the objectives of this study were to (1) illustrate the effects of low temperature at jointing stage on the yield and morphological attributes in winter wheat, (2) investigate the physiological responses of urea amendment following low-temperature stress on winter wheat growth recovery and yield loss alleviation, and (3) determine suitable nitrogen rates for alleviating low temperature stress at jointing stage.

## 2. Materials And Methods

A controlled pot experiment was done on Experimental Farm of Yangzhou University (119.42° E, 32.39° N), Jiangsu Province, China, in the two winter wheat growing seasons of 2012–2013 and 2013–2014. Yangmai 16, a widely extended winter wheat cultivar in the middle and lower reaches of the Yangtze River, was used.

### 2.1 Preparation of wheat plants

The wheat seedlings were developed in pots and transported to a phytotron for low temperature treatment. The pots were 0.28 m in diameter at the top, 0.22 m in diameter at the bottom, and 0.30 m in height. Each pot was filled with 13.0 kg sandy loam from a wheat field nearby. Soil containing 100.01 mg kg<sup>-1</sup> available N, 50.11 mg kg<sup>-1</sup> available P, and 149.08 mg kg<sup>-1</sup> available K in 2012 and 98.71 mg kg<sup>-1</sup> available N, 44.62 mg kg<sup>-1</sup> available P, and 156.83 mg kg<sup>-1</sup> available K in 2013.

For each pot, 12 seeds were sown at a seeding depth of 2.0 cm on October 31, 2012 and November 2, 2013, respectively. During the wheat growth period, 1.75 g N, 1.05 g P<sub>2</sub>O<sub>5</sub>, and 1.05 g K<sub>2</sub>O were applied for each pot, half of which was applied before sowing and half at jointing stage. At the 4th leaf stage, all the pots were thinned to 6 plants per pot. During the growth period, the pots were irrigated regularly to maintain a good moisture level and avoid water deficit. Other management practices were followed in accordance with local recommendations. At jointing stage, the plants were ready for low temperature treatment and urea amendment.

### 2.2 Low temperature treatment

A temperature-controlled phytotron was used in this study. It was set at 3°C/-4°C (day/night), relative humidity 70% (± 1%), 12 h photoperiod between 6:00 and 18:00, and photosynthetic photon flux density 900.0 μmol m<sup>-2</sup> s<sup>-1</sup>.

In the 2012–2013 wheat growing season, the plants were treated for 24 h and 48 h (referred to as T24 and T48 hereinafter). In the 2013–2014 wheat growing season, the plants were treated for 24 h, 48 h, and 72 h (referred to as T24, T48, and T72 hereinafter).

### 2.3 Urea amendment

Urea (N 46%) was applied on the fifth day after the low temperature treatment was finished. The amount of urea per pot was calculated based on the soil weight of the tillage layer in a nearby field where the soil in the pots were collected. The weight of tillage layer soil of the field is about 2.25 million kg hm<sup>-2</sup>. In the 2012–2013 wheat growing season, urea was

applied at 0, 0.9, and 1.3 g pot<sup>-1</sup>, which was approximately equivalent to 0, 150, and 225 kg N ha<sup>-1</sup> (referred to as N0, N150, and N225 hereinafter). In the 2013–2014 wheat growing season, urea was applied at 0, 0.6, 0.9, and 1.3 g pot<sup>-1</sup>, which was approximately equivalent to 0, 105, 150, and 225 kg N ha<sup>-1</sup> (referred to as N0, N105, N150, and N225 hereinafter). To promote the nitrogen uptake of wheat plants, urea was dissolved with 500 mL water and applied onto the soil surface of the pots.

In this study, a control growing in natural environment (without low temperature stress and urea amendment) was used. The study was arranged in a single factorial design with 8 replicates (each pot as a replicate). Thus, there were 7 and 13 treatments, respectively, in the 2012–2013 and 2013–2014 wheat growing seasons.

## **2.4 Observations and measurements**

### **2.4.1 Grade and proportion of freezing injury investigation**

The cold symptoms appeared on the fifth day after low-temperature treatment. We investigated the degree and proportion of freezing injury depending on the freezing injury standards used in wheat variety regional experiments in China and divided the freezing damage into five grades (Miao et al. 2007). Grade 1 was the lowest and indicated no freezing injury. Etiolation of one-third of the leaf tip corresponded to grade 2. One-third to one-half of the leaf tip being injured by freezing was grade 3. Withered and shed leaves were grade 4. Death of the main stems and tillers of wheat corresponded to grade 5.

### **2.4.2 Osmotic adjustment substances**

The second fully-expanded leaves from the top of the plant on the 10th day after urea amendment and from the cold-damaged wheat plants at jointing stage were used to determine the osmotic adjustment and hormonal content.

Proline content was measured according to Zhang and Qu (2003). Wheat leaves were extracted with 3.0% sulfosalicylic acid, incubated in a boiling water bath for 15 min, and then separated with filter paper. The reaction mixture containing 2 mL extract, 3.0 mL ninhydrin solution, and 2.0 mL acetic acid was incubated for 40 min at 100°C. The proline content was determined by monitoring the absorbance at 520 nm.

Soluble sugar content was measured according to Zhang and Qu (2003). Leaves were extracted with 80% ethanol, placed in a conical flask for 1 h, and then filtered with filter paper. The assay medium containing 0.5 mL extract, 4 mL anthrone (100 mL 72% H<sub>2</sub>SO<sub>4</sub> + 200 mg anthrone), and 1.5 mL distilled water was incubated for 15 min at 100°C, and then the absorbance was assessed at 620 nm.

### **2.4.3 Hormone contents**

The content of abscisic acid (ABA), zeatin riboside (ZR), and gibberellic acid (GA<sub>3</sub>) was measured by two-dimensional high-performance liquid chromatography (HPLC) following the method of Dobrev et al. (2005). Leaves were extracted and purified by immunoaffinity chromatography and quantified according to Pěnčík et al. (2009). The supernatant was separated using an Acquity ultra-high performance (UP) LC system (Waters 2695, U.S.A.) equipped with a Symmetry C<sub>18</sub> column (5 μm, 4.6 mm × 250 mm; Waters, U.S.A.). Methanol, 0.6% acetic acid, and acetonitrile (10: 9: 1 in volume) was used as a mobile phase, and its flow rate was 1.0 mL min<sup>-1</sup>. Pure ABA, GA<sub>3</sub>, and ZR were used as the standards.

### **2.4.4 Freezing injury grade proportion, freezing injury index, yield restoration effect, and nitrogen partial factor productivity**

The freezing injury grade proportion, freezing injury index, yield restoration effect, and nitrogen partial factor productivity were calculated as follows:

The freezing injury grade proportion (%) = the number tillers at each freezing injury grade/total tillers × 100

Freezing injury index =  $(\sum \text{freezing injury grade} (\geq \text{grade } 2) \times \text{proportion of freezing injury at the corresponding injury grade})/5$

Yield restoration effect (%) =  $(\text{yield with remedial fertilizer} - \text{yield without remedial fertilizer}) / \text{yield without remedial fertilizer} \times 100$

Nitrogen partial factor productivity ( $\text{g g}^{-1}$ ) = yield / total nitrogen amount

## 2.6.5 Yield assessment

At maturity, wheat plants from 8 pots in each treatment were harvested, and the grain yield per pot was assessed. The grain yield was adjusted to 13% moisture content.

## 2.7 Statistical analysis

The study was arranged in a single factorial design, with 7 and 13 treatments and 8 replicates (each pot as a replicate), respectively, in the 2012–2013 and 2013–2014 wheat growing seasons. For statistical analysis, analysis of variance (ANOVA) was performed ( $P < 0.05$ ) using DPS 7.05 Statistical Software (DPS, Zhejiang, China) to assess the effects of the treatments.

Unless specifically stated, the following result section focused on the 2013 to 2014 wheat growing season due to the fact that there were more low temperature treatments and urea rates in this growing season.

## 3. Results

### 3.1 Freezing injury grade after low-temperature stress at jointing stage

The freezing injury grade, proportion, and index in the wheat plants increased with longer low-temperature duration in the both growing seasons. As shown in Table 1, 55% of wheat plants were injured at different degrees when treated at 3°C/–4°C for 24 h (T24). The fifth level grade (main stems and tillers dead) was 2.5% under T24 and increased by 11.3% under T48 in the 2012–2013 wheat growing season. The average freezing injury index was 0.35, 0.50, and 0.66 under T24, T48, and T72, respectively, in the two wheat growing seasons, indicating that Yangmai 16 was sensitive to low temperatures at jointing stage.

Table 1  
Degree and proportion of freezing injury of wheat plants under low temperature stress at jointing stage

Year	Time (h)	Grade of freezing injury and proportion (%)					Freezing injury index
		1	2	3	4	5	
2012-2013	24 (T24)	45.34b	14.64a	21.68b	15.89b	2.45b	0.34
	48 (T48)	27.54c	10.42b	28.65a	22.06a	11.33a	0.50
	Control	100.00a	0.00c	0.00c	0.00c	0.00c	—
2013-2014	24 (T24)	42.83b	13.12a	26.53b	13.85c	3.67c	0.36
	48 (T48)	25.27c	11.46b	32.23a	21.65b	9.39b	0.51
	72 (T72)	12.06d	8.66c	24.75b	34.25a	20.28a	0.66
	Control	100.00a	0.00d	0.00c	0.00d	0.00d	—

\* Different small letters in the same column meant significant difference among treatments for the same year at the 0.05 level.

## 3.2 Effect of fertilizer amendment on plant morphology

Plant height ranged from 67.60 cm to 63.38 cm after low-temperature stress of 72 h at jointing stage, while the plant height of the control treatment was 87.27 cm (Fig. 1). The plant height was decreased by 22.5–27.4%. Plant height increased to 83.46 cm under T24N225 treatment, increased by 23.5% as compared with the cold treatment without the urea amendment. Plant height was 75.24 cm under T72N225 treatment, but dropped by 13.8% as compared with the plant in the natural environment. Remedial fertilizer after cold stress at jointing stage mainly promoted the length of the two upper internodes, which recovered the ear length and plant height to some extent. The stronger the cold stress was, the more significant the retrieval effects of remedial urea on plant height were.

## 3.3 Effect of fertilizer amendment after cold stress on yield formation

Grain yield exhibited a decreasing trend with increased low temperature duration (Table 2). Compared with the control, a 54.1–65.3% decrease in yield was observed under low temperature stress at jointing stage. The ear number was decreased from 34.3–38.1% with longer low-temperature stress in the 2013–2014 wheat growing season. Fertilizer amendment improved the ear number with increased urea application after cold stress of the same cold duration. The grain number per spike was significantly reduced with increased cold duration stress. The grain number per spike was increased by 9.3% under T24N225, compared with the treatment T24N0, while it was lower than that in the natural environment even though more remedial fertilizer had been applied. The 1000-grain weight was decreased with increased cold duration. In the same stress treatment, the yield restoration was the highest under the treatment with 225 kg ha<sup>-1</sup> urea amendment. The yield was restored by 33.9% under T24N225, 44.8% under T48N225, and 58.6% under T72N225 in the 2013–2014 wheat growing season. Their relative effects of yield restoration were also the highest among the different urea amendment levels.

Table 2  
Effects of remedial fertilizer after low temperature stress on wheat yield formation

Year	Time (h)	Freezing injury index	Remedial urea rate (kg ha <sup>-1</sup> )	Ear per pot	Grain number per spike	1000-grain weight (g)	Actual yield per pot (g)	Yield restoration effect (± %)	Partial factor productivity (g g <sup>-1</sup> )		
2012–2013	24(T24)	0.34	0 (N0)	24.00d*	36.52c	42.57ab	37.01e	-	21.15		
			150 (N150)	28.5c	42.28b	44.54a	52.43b	41.66	24.23		
			225 (N225)	29.67bc	42.50b	44.83a	54.78b	48.01	23.11		
	48(T48)	0.5	0 (N0)	22.00e	33.73f	38.51c	28.31f	-	16.18		
			150 (N150)	29.5bc	36.98c	41.77ac	43.25d	52.77	19.99		
			225 (N225)	30.67b	37.69c	42.57ab	48.92c	72.8	20.64		
			Control	–	32.50a	52.63a	44.17a	73.17a	-	41.81	
2013–2014	24(T24)	0.36	0 (N0)	23.00de	37.65bf	44.26ac	36.69ef	-	20.97		
			105 (N105)	25.67cd	39.93bd	45.85ab	45.19bc	23.17	22.16		
			150 (N150)	26.00bd	40.07bd	46.35ab	46.87bc	27.75	21.66		
			225 (N225)	26.33bd	41.15b	46.95a	49.13b	33.91	20.72		
	48(T48)	0.51	0 (N0)	23.00de	36.75dg	40.69cd	32.31fg	-	18.46		
			105 (N105)	26.33bd	39.00be	42.11bd	41.86ce	29.56	20.52		
			150 (N150)	26.33bd	39.93bd	42.9ad	44.99bc	39.24	20.79		
			225 (N225)	27.5bc	40.83bc	42.74ad	46.79bc	44.82	19.74		
			72(T72)	0.66	0 (N0)	21.67e	33.58g	38.71d	27.68g	-	15.82
					105 (N105)	28.00bc	34.23fg	39.31d	35.99ef	30.02	17.65
150 (N150)	28.33bc	35.46eg			40.51cd	38.47df	38.98	17.78			
225 (N225)	29.5b	36.86cg			41.93bd	43.89bd	58.56	18.51			

\* Values followed by the same letter in the same column for the same year are not significantly different ( $P < 0.05$ ).

Table3 Correlation coefficients between the parameters on the 10th day after applying remedial fertilizer and plant height as well as yield

Year	Time	Freezing injury index	Remedial urea rate	Ear per pot	Grain number per spike	1000-grain weight	Actual yield per pot (g)	Yield restoration effect ( $\pm$ %)	Partial factor productivity
	(h)		(kg ha <sup>-1</sup> )			(g)			(g g <sup>-1</sup> )
	Control	—		35.00a	52.28a	44.5ac	79.86a	-	45.63
* Values followed by the same letter in the same column for the same year are not significantly different ( $P < 0.05$ ).									
Table3 Correlation coefficients between the parameters on the 10th day after applying remedial fertilizer and plant height as well as yield									

At jointing stage, urea amendment mainly improved the grain yield by increasing the grain number per ear, with a path coefficient of 0.5258\*\* ( $P < 0.01$ ), followed by ear number with a path coefficient of 0.4580\*\* ( $P < 0.01$ ). With increased urea application under the same treatment duration, the growth of wheat plants was restored better and the loss of grain yield lessened.

### 3.4 Effect of remedial fertilizer on osmotic adjustment substance

**3.4.1 Soluble sugar** Soluble sugar in the leaves on the 10th day after urea amendment to the cold-damaged plants under T24, T48, and T72 was lower than that of the cold-damaged plants without fertilizer amendment treatment (Fig. 2A). Compared with the control, the treatment T24 with urea amendment had a higher soluble sugar concentration in the leaf on the 10th day after urea amendment. The differences in soluble sugars of the leaves between the treatments and control significantly narrowed after applying three urea levels under T24. The soluble sugar concentration was decreased from 127.36–71.74% on the 10th day after application of 150 kg ha<sup>-1</sup> and 225 kg ha<sup>-1</sup> urea under T24 compared with the control treatment (Fig. 2B). The soluble sugar concentration of the leaves on the 10th day under T72N225 treatment was lower than that of the treatment T72N0, with a decrease of 34.2%, implying that the cold-damaged wheat plants recovered growth gradually after applying remedial fertilizer. We observed that remedial fertilization after low-temperature stress had more positive effects on reducing the concentration of soluble sugar in wheat plants. The recovering speed of severe cold stress treatment (T72) was slower than that of mild cold stress (T24).

**3.4.2 Proline** Compared with the treatment T24N0, the proline concentration in the second fully-expanded leaves on the 10th day after urea amendment was decreased from 5% to 16% with the increased remedial urea rate under T24 (Fig. 2C, D). Under T48N225, the proline concentration was lower than that of T48N0, with a decrease by 13.0% on the 10th day. The changes in proline concentration under T72 exhibited the same trend as under T24 with remedial fertilizer at three levels. All the treatments under T72 with nitrogen amendment had the highest proline concentration among all cold stress treatments, indicating that the recovery effect of nitrogen amendment on T72 was worse than those of T24 and T48. Similar to soluble sugar, the differences in the proline concentration in the leaves between treatments with urea amendment and the control narrowed on the 10th day after remedial urea application under three low-temperature duration levels.

### 3.5 Effect of fertilizer amendment on hormone concentrations in the second expanded leaf from the top

**3.5.1 ABA** Compared with the control, the ABA concentration in the second upper leaves on the 10th day of the treatment without fertilizer application gradually increased with the duration of treatment time after low-temperature stress (Fig. 3A). Within the same duration, the concentration of ABA was decreased gradually with the increased rate of fertilization. Under T48 and T72, compared with the control treatment, the ABA concentration under 225 kg ha<sup>-1</sup> urea amendment was significantly increased by 87.6% and 120.2% on the 10th day.

**3.5.2 ZR** Compared with the treatment without urea amendment, the ZR concentration decreased following urea amendment (Fig. 3B). Under T24N225, the ZR concentration was significantly lower by 16.9% than that of T24N0 on the 10th day. The ZR concentration increased gradually with the increased treatment duration. Under the same low-temperature duration, the ZR concentration dropped with increased fertilizer amount.

**3.5.3 GA<sub>3</sub>** Compared with the control, the GA<sub>3</sub> concentration decreased following low temperature stress (Fig. 3C). Under the treatment of T24, T48, and T72, the GA<sub>3</sub> concentrations with 225 kg ha<sup>-1</sup> urea amendment was 54.6%, 63.5%, and 69.8% lower than that of the control treatment on the 10th day. The GA<sub>3</sub> concentration increased after urea application. The effect of increased urea application after low-temperature stress on recovering GA<sub>3</sub> concentration was more obvious.

## 3.6 Relationship between mechanism index and yield

Proline, soluble sugar concentrations, ABA, and ZR concentrations in the 2nd full expanded leaf from the top on the 10th day after remedial urea application were negatively correlated, while the GA<sub>3</sub> concentration was positively correlated with the length of the upper internode IV, internode V, plant height, and yield (Table 3). The higher proline, ABA, and ZR concentrations in the leaves under cold stress affected plant growth, resulting in shorter upper internode lengths, which influenced wheat yield formation.

## 4. Discussion

### 4.1 Response of osmotic adjustments to remedial fertilizer after low temperature stress at jointing stage

Carbohydrates play a crucial role in freezing tolerance (Livingston et al. 2006). Simple sugars, such as trehalose, sucrose, and raffinose, are especially correlated with enhanced cold tolerance (Kaplan et al. 2006; Hassan et al. 2021). In this study, significant increases in soluble sugar concentration in wheat under treatment without fertilizer amendment were observed, especially under the treatment T72N0 (Fig. 2A), indicating that wheat plants adapted to cold stress by increased soluble sugar concentration. Salicylic Acid-treated wheat enhanced total soluble sugar contents under low-temperature conditions (Wang et al. 2020). In our study, the concentration of soluble sugars were reduced from 13.2–20.6%, from 14.7–24%, and from 7–13.2% under T24, T48, and T72 with increased remedial fertilizer rate. These findings imply that remedial fertilizer after low temperature stress at jointing stage is an efficient way to increase sucrose inversion.

Another essential protective substance in the plant response to abiotic stress is proline. The level of proline was correlated with wheat low-temperature tolerance (Dörffling et al. 2009). In the present study, proline accumulation was enhanced on the 10th day after low-temperature stress without remedial fertilizer compared with the control and increased dramatically by 88.6% under T24N0, 107% under T48N0, and 130% under T72N0 (Fig. 2C, 2D). Proline may act as an osmolyte and function as a compatible ROS scavenger, protecting the plant from such oxidative stress. Its accumulation might balance the cell redox status and buffer the cytosolic pH (Majláth et al. 2012). High concentrations of proline can increase tissue turgor, advance osmotic regulation, and enhance plant resistance to low temperature (Li et al. 2017). In this study, the proline concentration was reduced significantly with increased remedial fertilizer at the same low-temperature duration, which demonstrated that remedial urea might contribute to the balance of proline metabolism and the growth recovery of the wheat plants. In summary, the lower soluble sugar and proline concentrations in wheat were likely to result in osmoregulatory recovery after remedial urea application, which ultimately contributed to faster recovery and growth of wheat plants.

### 4.2 Hormone changes of remedial fertilizer after low temperature stress at jointing stage

Plant endogenous hormones, such as ABA, GA, and CK<sub>s</sub>, function as signaling molecules and participate in cold resistance (Sun et al. 2009; Vaseva et al. 2009; Wang et al. 2018). Low temperature-induced ABA accumulation in winter wheat at booting stage altered the activity of enzymes related to sucrose metabolism, which led to sucrose synthesis and accumulation in the young ears, thus causing yield losses (Zhang et al. 2019). In this study, we also noticed that soluble sugar and ABA concentrations were increased in abundance after low-temperature stress from 24 h to 72 h (Fig. 2A, 3A). Lower GA<sub>3</sub> content and higher ABA content in the seeds after chilling injury was previously found to inhibit seed germination, while treating the seeds with GA<sub>3</sub> could mitigate the low-temperature injury to seed germination (Hou 2003). In this study, the GA<sub>3</sub> concentration changed conversely to the concentration of ABA not only after low temperature (Fig. 3C), but also after the remedial urea amendment. In addition, remedial urea significantly maintained lower levels of endogenous ABA and ZR and higher GA<sub>3</sub> levels, which enhanced the wheat growth rate. These results are well in accordance with the observations of Yang et al. (2013), who reported that low-temperature conditioning alleviated injury in kiwifruit by promoting higher ABA/GA<sub>3</sub> ratios.

In the present study, the levels of ABA and ZR in the treatment without remedial fertilizer gradually increased with the longer low-temperature duration at jointing stage, which are typical responses of wheat to low-temperature stress (Fig. 3A, 3B). These results suggested that wheat changed the balance of these hormones to adapt to low-temperature stress. Remedial urea was conducive to alleviating the low-temperature damage to wheat and improving wheat growth via the reduced ABA as well as ZR and increased GA<sub>3</sub> concentrations following low-temperature stress at jointing stage.

### **4.3 Contribution of remedial fertilizer after freezing injury to plant morphology and grain yield**

The warming of global mean winter temperatures promotes wheat growth in the previous winter, enhancing cold vulnerability in the next spring (Li et al. 2016). Low temperature can remarkably reduce the rates of wheat growth and development, resulting in changes in morphological characters, such as leaf area (Valluru et al. 2012), ear number, and grain number (Thakur et al. 2010; Li et al. 2017). Limin and Fowler (2000) reported that the cell size of wheat decreased and the young and new leaves became shorter and narrower following cold acclimation. In the present study, the degree of cold injury to the wheat plants and the freezing injury index were all increased from 0.3 to 0.5 with increased low temperature duration at jointing stage, resulting in lower plant height, especially the first and second upper internode length and ear length. Our results are consistent with the report of Li et al. (2017), who found that chilling stress treatment significantly decreased plant height and leaf area in hexaploid wheat. These results indicated that low temperatures affected wheat development mainly by reducing the growth rate, resulting in reduced plant height.

Several cold hardening experiments suggested that endogenous hormones, such as IAA and CKs, may actively participate in the control of plant growth under low-temperature stress (Majláth et al. 2012). Our findings confirmed that there was a strong negative correlation between plant height as well as the upper two internode length and related physiological parameters (Table 3), including proline, soluble sugar, ABA, and ZR concentrations, while GA<sub>3</sub> concentration was positively correlated with these. These data also confirmed that internode elongation was inhibited by higher ABA and ZR levels and lower GA<sub>3</sub> level under low temperature. In contrast, the balance among these hormones improved with remedial urea application after low-temperature stress, resulting in greater plant height (Fig. 1). Remedial fertilizer after cold stress at jointing stage mainly promoted the length of the two upper internodes and ear length, ultimately increasing the plant height.

Low temperature not only inhibits wheat growth and development but also results in the loss of grain yield. In Kansas, an additional day of freezing temperature in the spring was associated with a 3.3% yield reduction over the 1985–2013 period (Tack et al. 2015). In certain regions of Australian wheat belt, a high risk of  $\geq 10\%$  yield losses result from post-head-emergence frosts (Zheng et al. 2015). The wheat yield was reduced by 42.5–59.8% under low temperature ( $-2^{\circ}\text{C}/-8^{\circ}\text{C}$ ,

day/night) from 24 h to 72 h at tillering stage in Jiangsu province in China (Li et al. 2017). In the present study, the wheat yield was reduced by 49.42–65.11% under low temperature (3°C/–4°C, day/night) from 24 h to 72 h at jointing stage. Internode extension and dry matter accumulation were restricted and the spikelet was killed, and thus the grain yield was also decreased due to low temperature during stem elongation (Whaley et al. 2004). Our findings also demonstrated that the reduced number of fertile tillers and grain number per spike were primarily associated with the loss of yield under cold stress at jointing stage.

The cold resistance of wheat was correlated with the levels of expression of *TaEXPB7-B* in the tillering nodes, which was up-regulated by both low-temperature and ABA (Feng et al. 2019). As a result, young and new tillers were generated rapidly after cold stress at jointing stage (Liu et al. 2019). We also observed this phenomenon in our study. Treatment with remedial urea after low temperature at jointing stage generated more new tillers and enhanced grain yield. Of course, the yield under treatment with remedial urea could not wholly restore and reach the level in natural conditions. The principal component analysis results indicated that grain yield was positively correlated with GA<sub>3</sub>. The GA<sub>3</sub> concentration was higher 23.68%–52.65%, 16.67–41.4%, 25.6–79.8% with the increased rate of remedial urea than those with T24N0, T48N0 and T72N0, which could benefit from promoting wheat growth, inducing new generated tillers, and internode extension. With increased fertilizer application rate under the same treatment duration, the height of the wheat plants was better restored and the loss of grain yield was lessened.

These findings indicated that remedial fertilizer could increase the growth recovery of the cold-damaged wheat plants and reduce the loss of wheat yield. At jointing stage, considering the recovery effect and nitrogen partial factor productivity, 105 kg ha<sup>-1</sup> urea is recommended for nitrogen amendment when wheat plants are damaged slightly and the freezing injury index is about 0.3. When the freezing injury index is about 0.5, 150 kg ha<sup>-1</sup> urea is suggested. When the freezing injury index is about 0.7, 225 kg ha<sup>-1</sup> urea is recommended for recovering wheat growth after severe cold damage. Our findings can offer useful and practical approaches for alleviating the impacts of low temperatures in spring on wheat production. Of course, we need to think about the comprehensive income, including the remedial urea costs, the price of wheat, the negative effect of remedial urea on weak-gluten wheat quality, because the grain protein content will be improved with the remedial urea application after jointing stage.

## 5. Conclusion

Low-temperature damage at jointing stage significantly limited wheat yield and changed the concentrations of osmotic adjustment substances and related hormones, as well as inhibited plant growth, resulting in shorter internode length and plant height and fewer ears and grains per spike. Remedial urea to cold injured wheat reduced the concentrations of osmotic adjustment substances (proline and soluble sugar) and improved the balance of ABA, ZR, and GA<sub>3</sub> in the second fully-expanded leaves. Therefore, injured wheat plants amended with urea application recovered growth more rapidly than those without urea application, resulting in significant increases in the length of the upper two internodes, plant height, new reproductive tiller number, ear number, and wheat grain yield. These advantageous changes became more pronounced with a higher rate of urea application in the same cold duration treatment. Considering the recovery effects and nitrogen partial factor productivity, suitable urea remedial rates were suggested depending on the freezing cold index after cold damage at the wheat jointing stage. These findings provide effective and practical knowledge and approaches to alleviating the impacts of low temperature at jointing stage on wheat production.

## Declarations

**Conflicts of Interest:** The authors declare no conflicts of interest.

**Author Contributions:** Conceptualization, C.L. and X.Z.; formal analysis, J.Y. and M.Z.; investigation, J.Y.; Methodology, J.Y. and J.D.; project administration, W.G.; resources, J.D. and X.Z.; writing-original draft, C.L.; writing-review and editing, G.Z.;

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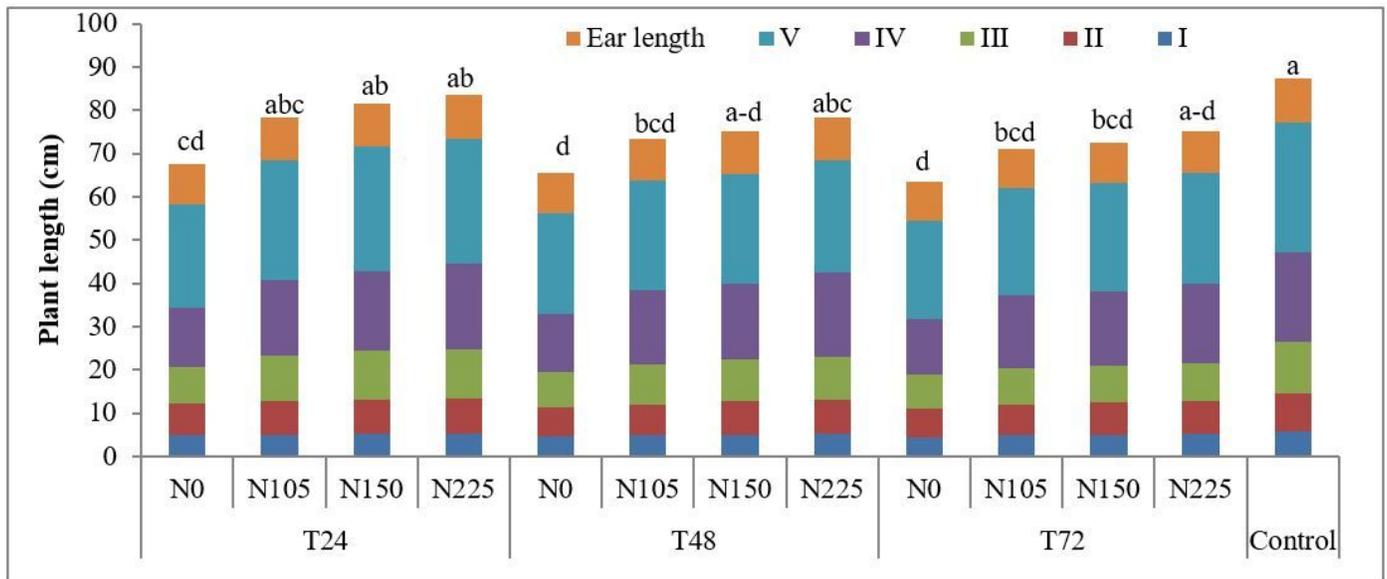
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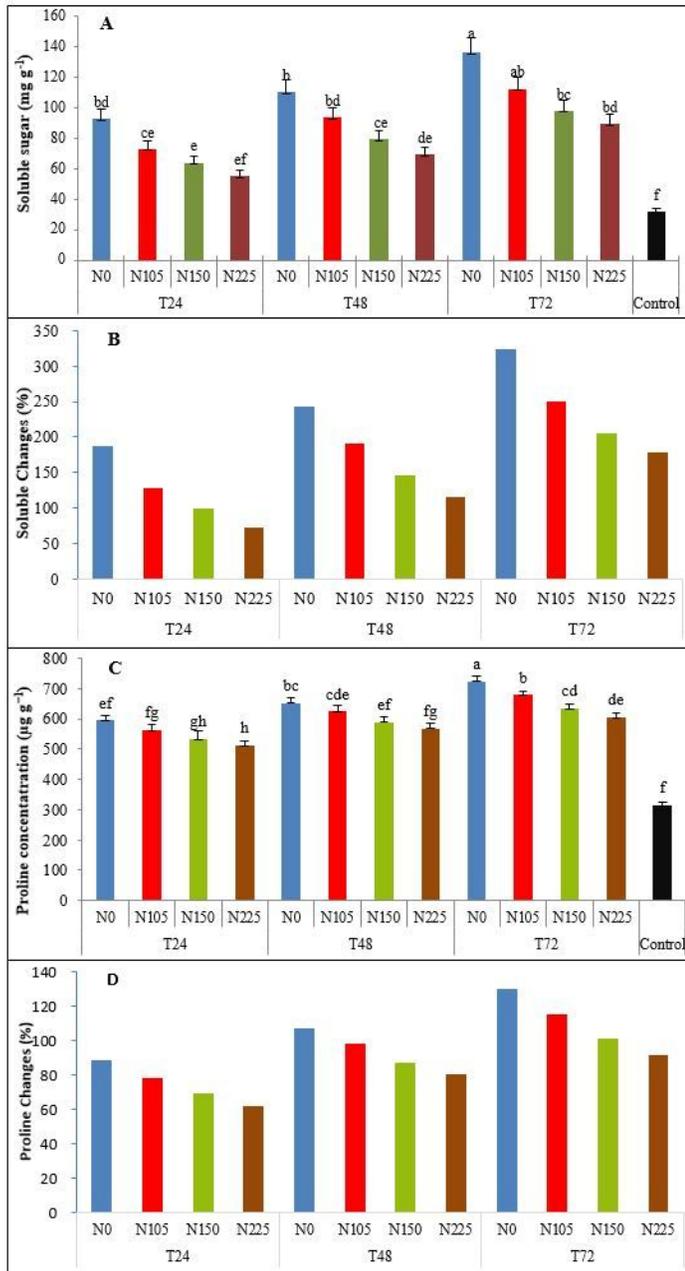
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## Figures



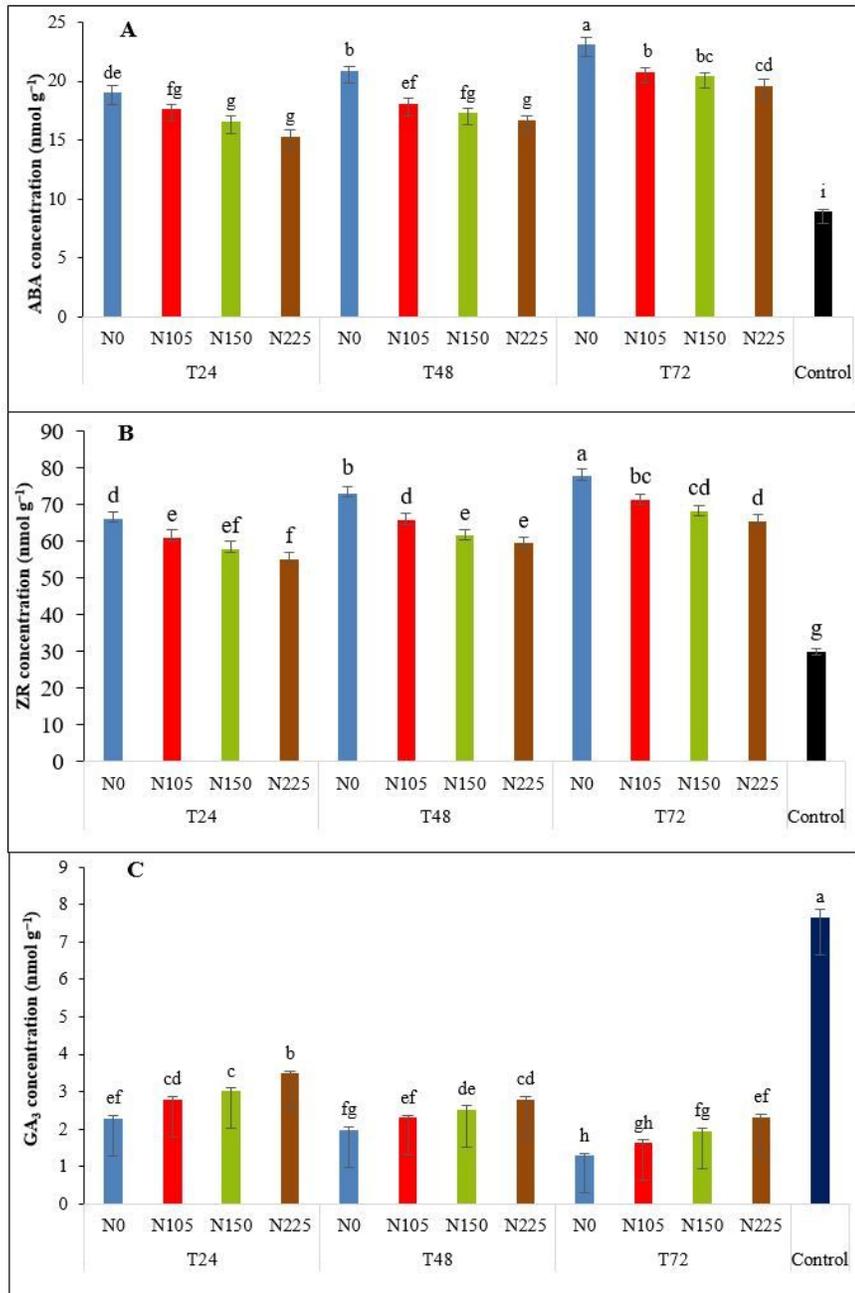
**Figure 1**

Changes in the length of internode, ear and plant after urea amendment to cold-damaged wheat at jointing stage (2013–2014 wheat growing season)



**Figure 2**

Changes in soluble sugar and proline concentrations in the second leaves from the top on the 10<sup>th</sup> day after urea amendment to cold-damaged wheat at jointing stage (2013–2014 wheat growing season)



**Figure 3**

Changes in ABA (A), ZR (B) and GA<sub>3</sub> (C) contents in the second leaf from the top after urea amendment to cold-damaged wheat plants at jointing stage in the 2013–2014 growing season.

ABA, abscisic acid; GA<sub>3</sub>, gibberellic acid; ZR, zeatin riboside.