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Re-recognizing Micro Locations of Nanoscale Zero-valent Iron in Biochar Using C-TEM technique

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Abstract: Biochar supported nanoscale zero-valent iron (NZVI/BC), prepared commonly by liquid reduction using sodium borohydride (NaBH_4), exhibits better reduction performance for contaminants than bulk NZVI. The better reducing ability was attributed to attachment of nanoscale zero-valent iron (NZVI) on biochar (BC) surface or into the interior pores of BC particles due to observations by scanning electron microscopy (SEM) and plan transmission electron microscopy (P-TEM) techniques in previous studies. In this study, cross-sectional TEM (C-TEM) technique was employed firstly to explore location of NZVI in NZVI/BC. It was observed that NZVI is isolated from BC particles, but not located on the surface or in the interior pores of BC particles. This observation was also supported by negligible adsorption and precipitation of $\text{Fe}^{2+}/\text{Fe}^{3+}$ and iron hydroxides on BC surface or into interior pores of BC particles respectively. Precipitation of Fe^{2+} and Fe^{3+} , rather than adsorption, is responsible for the removal of Fe^{2+} and Fe^{3+} by BC. Moreover, precipitates of iron hydroxides cannot be reduced to NZVI by NaBH_4 . In addition to SEM or P-TEM, therefore, C-TEM is a potential technique which should be employed to characterize the interior morphology of NZVI/BC for better understanding the reduction mechanisms and performance of contaminants by NZVI/BC.

Keywords: Nanoscale zero-valent iron, Biochar, Cross-sectional TEM, Interior morphology, Biochar supported nanoscale zero-valent iron, SEM

31 **TOC:**

1. Introduction

Nanoscale zero-valent iron (NZVI), as an environmental friendly material has been widely used in recent decades to reduce contaminants such as Cr(VI), As(V), nitrates, and chlorinated organics¹⁻⁶. Biochar (BC) supported NZVI (NZVI/BC), prepared commonly by liquid-phase reduction method using sodium borohydride (NaBH_4) as reducing agent, has better ability in reducing pollutants than NZVI⁷⁻⁹. The oxidation and agglomeration of NZVI can be effectively suppressed by BC, therefore, NZVI's surface energy and reduction activity can be maintained^{5,10-12}. In previous studies¹³⁻¹⁶, by scanning electron microscopy (SEM) and plan transmission electron microscopy (P-TEM) techniques, it was observed that NZVI particles were loaded on the surface or in the interior pores of BC particles, which is responsible for the improved reduction ability of NZVI¹⁷⁻¹⁹. Several possible ways were suggested to interpret the loading of NZVI on BC surface or in BC interior pores^{15,20-23} (i) $\text{Fe}^{2+}/\text{Fe}^{3+}$ ions are adsorbed on surface or into interior pores of BC and then be reduced to NZVI by NaBH_4 ; (ii) $\text{Fe}^{2+}/\text{Fe}^{3+}$ ions are hydrolyzed into precipitates of iron hydroxide, and then deposited on surface or into interior pores of BC particles for reduction to NZVI by NaBH_4 ; (iii) $\text{Fe}^{2+}/\text{Fe}^{3+}$ ions are reduced to NZVI by NaBH_4 in solutions and then deposited on BC surface or into the interior pores of BC particles.

SEM and P-TEM are widely used electron microscopy techniques with high resolution and high magnification to visualize the morphology of NZVI/BC²⁴⁻²⁶. However, by collecting secondary electrons reflected from sample surface, SEM technique can only visualize the sample top surface but not the interior structures^{27,28}.

For P-TEM technique, the accumulated signals of the sample from top to bottom including the interior structures can be obtained by collecting electrons through the sample²⁹. However, the accumulated signals of P-TEM are difficult to explore the interior structure of samples clearly because the interior structure signals are actually covered by the electron signals of top and bottom surfaces, i.e., there is a problem of superimposition for P-TEM technique^{30,31}. Therefore, both of SEM and P-TEM technique cannot explore the interior structure signals of NZVI/BC particle clearly for understanding the underlying improved reduction mechanisms of contaminants by NZVI/BC.

Cross-sectional TEM (C-TEM) technique, first reported in 1974³², is conducted through directly observing the inner ultra-thin slices (with thickness less than 100 nm) of samples using P-TEM technique. This technique can eliminate the problem of superimposition of P-TEM technique by avoiding the interfering of the sample surface signals, and thus, obtain the pure signals of interior structure of the sample³³⁻³⁵. Therefore, C-TEM technique has been applied widely in characterizing of solid materials such as bulk heterojunction films^{36,37}, biochar³⁸, and activated carbons³⁹ in previous studies. In this study, therefore, C-TEM technique was firstly employed to directly identify the interior structures of NZVI/BC, especially the micro locations of NZVI in NZVI/BC particles, for exploring the underlying reduction mechanisms of NZVI/BC for contaminants reduction.

2. Results and Discussion

The surface morphology of BC particles observed by SEM technique is relatively smooth in lamellar structure (Figure 1a), with abundant irregular pores (Figure 1b). However, in C-TEM images (Figure 1c and 1d), the slices of BC particles are in the sheet-like shape, showing abundant slit pores in uniform pore size. Bulk NZVI particles in SEM (Figure 2a) and C-TEM (Figure 2b) images are roughly spherical and aggregated significantly to a chain-like structure, which should be attributed to the intrinsic magnetic attraction between the NZVI particles^{4,5,12,40,41}. There are a lot of chain-like clusters and roughly spherical particles dispersed on BC surface in SEM images of NZVI/BC (Figure 3a and 3b). These chain-like clusters and roughly spherical particles on the surface of NZVI/BC are also observed in SEM images of bulk NZVI particles (Figure 2a) and in SEM images of NZVI/BC in previous studies^{11,18,40,42}, which should be attributed to NZVI particles. EDX spectrum of NZVI/BC shows that the main elemental composition of NZVI/BC is C, Fe and O (Figure 3c). The homogeneous distribution of C and Fe was also verified in SEM elemental mapping of NZVI/BC (Figure S2). NZVI particles formed in NZVI/BC are identified by the characteristic peak at 44.7° (Figure 3d) of XRD pattern of NZVI/BC. NZVI particles formed and dispersed on the surface of NZVI/BC were also observed in P-TEM images of NZVI/BC in previous studies^{19,22}. However, in C-TEM images of NZVI/BC (Figure 4a), NZVI particles are existed isolated from BC particles rather than attached on the surface of BC particles. The distinguished observations in C-TEM images of NZVI/BC from that in SEM and P-TEM images of NZVI/BC, could be attributed to the top surface

view of SEM technique^{27,29} and the superimposition problem of P-TEM technique^{30, 31}. Thus, both of SEM and P-TEM techniques cannot explore the interior structure signals of NZVI/BC particles. Moreover, NZVI particles are not observed in interior slit pores of BC particles (Figure 4b), i.e., NZVI particles cannot be embedded into the interior pores of BC particles. As reported in the previous literatures^{11,43,44}, the average particle size of NZVI on NZVI/BC is about 30~100 nm. In this study, the particle size of NZVI in NZVI/BC is in the range of 40-180 nm (Figure 3b). However, BC is basically composed of micropores and mesopores with average diameter less than 10 nm^{13,38,45}. The N₂ adsorption–desorption isotherm and pore size distribution of BC in this study are shown in Figure S3. The specific surface and average pore size of BC are 579.15 m²/g and 2.486 nm, respectively. Therefore, NZVI particles cannot enter into interior pores of BC particles, showing the clean and slit pores without any iron particles in C-TEM images of nZVI/BC (Figure 4). In addition, the surface morphology of NZVI/BC in SEM images (Figure 3a and 3b) and inner cross section structure of NZVI/BC in C-TEM images (Figure 4) as well as the XRD pattern (Figure 3d) of NZVI/BC are similar to that of NZVI+BC (Figure 3d and Figure 5), the sample mixed from NZVI and BC directly. Therefore, NZVI/BC is similar to NZVI+BC, without attachment of NZVI on BC surface or into BC interior pores, i.e., the better reduction ability of NZVI/BC for contaminants than NZVI cannot be interpreted by the attachment of NZVI on BC surface or into BC interior pores.

Adsorption of Fe²⁺ or Fe³⁺ ions on BC and then reduction of adsorbed Fe²⁺ or Fe³⁺ ions by NaBH₄ was proposed as a possible way to load NZVI on BC surface or in BC

interior pores ^{15,21,22}. However, we observed that the removal curves of Fe³⁺ in the presence of BC under various pH were overlapped with the precipitation curve of Fe³⁺ without BC (Figure 6a and 6b). Moreover, removal curves of Fe³⁺ were independent of the dosage of BC (20 and 200 mg) in Figure 6a or initial concentration of Fe³⁺ (50 and 150 mg/L) in Figure 6b. Therefore, the removal of Fe³⁺ ions in the presence of BC should be attributed primarily to precipitation by formation of Fe(OH)₃ rather than adsorption on BC⁴⁶. Although the removal curve of Fe²⁺ was not overlapped with the precipitation curve of Fe²⁺ without BC (Figure 6c), it was overlapped with the precipitation curve of Fe³⁺ without BC (Figure 6d), implying that the oxidation of Fe²⁺ to Fe³⁺ and then the precipitation of Fe³⁺ may be responsible for the removal of Fe²⁺ with BC. In order to check this hypothesis, four different points (A, B, C, D) were chosen to determine the mass of Fe²⁺ and Fe³⁺ in supernatants and solids, respectively (Figure 6c). Species distribution of Fe²⁺ and Fe³⁺ in supernatant and solids are listed in Table S1. Oxidation of Fe²⁺ to Fe³⁺ with or without BC were observed (Table S1). In the presence of BC and higher pH, more Fe²⁺ were oxidized to Fe³⁺ (Table S1). Therefore, the oxidation of Fe²⁺ to Fe³⁺ occurred in batch experiment for Fe²⁺ by BC. Characteristic peak of NZVI was not observed in XRD diffraction pattern of NZVI/BC*, a sample prepared by removing excessive free Fe²⁺ in Fe²⁺/BC system before adding NaBH₄ (Figure 3d), indicating almost no Fe²⁺ or Fe³⁺ adsorbed on BC. Moreover, the C-TEM diagrams of NZVI/BC* were similar to that of BC (Figures S4), indicating that no black NZVI particles observed in NZVI/BC, i.e., no Fe²⁺ or Fe³⁺ adsorption on BC

observed. Therefore, Fe^{2+} and Fe^{3+} cannot be fixed on BC surface or in interior pores through adsorption to form NZVI on BC by reduction using NaBH_4 .

Forming of $\text{Fe}(\text{OH})_2$ or $\text{Fe}(\text{OH})_3$ precipitates, deposited on BC and then reduced to NZVI by NaBH_4 was proposed as another possible way to load NZVI on BC surface or in BC interior pores^{20,22}. However, depositing of $\text{Fe}(\text{OH})_2$ or $\text{Fe}(\text{OH})_3$ precipitates on the surface or in the interior pores of BC particles were not observed in C-TEM images of NZVI/BC* (Figure S4), showing that the surface and pores of NZVI/BC* are clean and similar to that of BC (Figure S4). Moreover, NZVI can be obtained by reducing iron salt with NaBH_4 rather than hydroxide^{4,47}, i.e., $\text{Fe}(\text{OH})_2$ or $\text{Fe}(\text{OH})_3$ cannot be reduced to NZVI by NaBH_4 . For example, the suspensions of dark green $\text{Fe}(\text{OH})_2$ precipitates and reddish brown $\text{Fe}(\text{OH})_3$ precipitates (Figure 7a), prepared from Fe^{2+} and Fe^{3+} respectively via chemical precipitation, are not changed in color after dropping NaBH_4 (Figure 7b), i.e., No black ZVI particles formed after reduction treatment of $\text{Fe}(\text{OH})_2$ and $\text{Fe}(\text{OH})_3$ precipitates with NaBH_4 . The characteristic peak of NZVI was not observed in XRD diffraction patterns of the dried solid samples collected from NaBH_4 treated $\text{Fe}(\text{OH})_2$ and NaBH_4 treated $\text{Fe}(\text{OH})_3$ (Figure 3d), indicating NZVI cannot be obtained from $\text{Fe}(\text{OH})_2$ and $\text{Fe}(\text{OH})_3$ precipitates by reduction using NaBH_4 . Therefore, NZVI cannot be loaded on BC via depositing of $\text{Fe}(\text{OH})_2/\text{Fe}(\text{OH})_3$ precipitates on surface or into interior pores of BC particles as well as reduction of $\text{Fe}(\text{OH})_2/\text{Fe}(\text{OH})_3$ precipitates to NZVI by NaBH_4 .

Except for loading of NZVI on the surface or into the interior pores of BC, smaller size of NZVI in NZVI/BC was proposed to interpret the better reduction performance

of contaminants by NZVI/BC than bulk NZVI^{22,40,48}. Smaller-sized NZVI is more reactive, which can be attributed to its higher surface area-to-volume ratio and stronger quantum effects^{4,6,11}. In this study, bulk NZVI particles are aggregated into micron-size particles (Figure 2), which is consistent with the previous reports^{6,40,49}. However, the particle size of NZVI in NZVI/BC is in the range of 40-180 nm (Figure 3b and Figure 4), much smaller than that of bulk NZVI (Figure 2), implying that the agglomeration of NZVI particles is effectively alleviated by BC^{10,22}. The alleviation of NZVI agglomeration was also observed in sample NZVI+BC, showing that the particle size of NZVI in NZVI+BC is in the range of 50-220 nm (Figure 5). Therefore, the smaller size of NZVI in NZVI/BC than that of bulk NZVI could be possibly responsible for the better reduction performance of contaminants by NZVI/BC than bulk NZVI¹⁰⁻¹².

3. Conclusion and outlook

Employing C-TEM technique for liquid reduction method prepared NZVI/BC, it was observed in this study that NZVI particles are existed isolated from BC particles in NZVI/BC, rather than attached on surface or in interior pores of BC particles that was observed in previous study by SEM and P-TEM techniques. This observation was also supported by the negligible adsorption of $\text{Fe}^{2+}/\text{Fe}^{3+}$ and the negligible attachment of $\text{Fe}(\text{OH})_2/\text{Fe}(\text{OH})_3$ precipitates on BC surface or interior pore structure of BC particles, as well as the negligible reduction of $\text{Fe}(\text{OH})_2/\text{Fe}(\text{OH})_3$ to NZVI by NaBH_4 . Therefore, both of the widely used SEM and P-TEM techniques have difficulty in observing the inner structure of NZVI/BC. In addition to SEM and P-TEM, C-TEM is a useful

technique which should be employed to characterize the interior morphology of NZVI/BC and other porous materials supported NZVI for better exploring the underlying improved reduction mechanisms of contaminants by porous materials supported NZVI. The smaller size of NZVI particles in NZVI/BC than bulk NZVI rather than the attachment of NZVI particles on surface or into interior pores of BC particles in NZVI/BC, could be responsible for the better reduction performance of contaminants by NZVI/BC than bulk NZVI.

4. Materials and Methods

4.1. Chemicals

Ferrous sulfate heptahydrate ($\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$, 97%), sodium borohydride (NaBH_4 , 98%), and hydroxylamine hydrochloride ($\text{NH}_2\text{OH} \cdot \text{HCl}$, 99%) were supplied by Sinopharm Group Chemical Reagent, China. Ferric chloride hexahydrate ($\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$, 99%) was obtained from Aladdin Reagent Corporation.

4.2. Preparation of BC and NZVI materials

BC was prepared from wood chips (Ningbo, Zhejiang, China) by oxygen-limited pyrolysis method⁵⁰. Briefly, dried wood chips, grounded to pass through a 0.15 mm sieve, were packed in crucibles, covered with lids, and then, placed in a muffle furnace, pyrolyzed at 700°C for 6 h. The prepared BC was washed with 1 mol/L HCl ($m/v=0.1$ g/mL) by stirring for 24 h to remove impurities and then rinsed using deionized (DI) water to neutral pH. The obtained BC sample was dried overnight at 80°C in an oven for usage.

Bulk NZVI was synthesized using the liquid-phase reduction method^{11,12,51}. Briefly, under nitrogen atmosphere, 100 mL NaBH₄ solution (1.2 mol/L) was dropped (1-2 drops/s) into 100 mL FeSO₄ solution (0.44 mol/L) with stirring. The black NZVI particles were separated by centrifugation for 5 min at 3000 rpm. Then, NZVI particles were washed with deoxygenated DI water, followed by ethanol, and finally dried in a vacuum oven at 60°C for 12 h.

NZVI/BC was prepared according to the procedures reported in the previous literature⁵². Under nitrogen atmosphere, 100 mL of 0.44 mol/L FeSO₄ solution was mixed with 5.0 g BC for 3 h. Then, 100 mL of 1.2 mol/L NaBH₄ was added dropwise into the mixture. After reaction, NZVI/BC was collected by centrifugation at 3000 rpm for 5 min, washed with deoxygenated DI water and ethanol, and dried in a vacuum oven (60°C, 12 h).

NZVI/BC* was prepared as follows. 100 mL of 0.44 mol/L FeSO₄ solution was mixed with 5.0 g BC under nitrogen atmosphere for 24 h. Then the solid was collected by centrifugation at 3000 rpm for 5 min and followed by washing with deoxygenated DI water for three times to remove excess free Fe²⁺ that not adsorbed on BC. Under nitrogen atmosphere, 100 mL of 1.2 mol/L NaBH₄ was added dropwise into the suspension of BC which was pretreated with FeSO₄ solutions. Then, by centrifugation at 3000 rpm for 5 min, washed with deoxygenated DI water and ethanol, and dried in a vacuum oven at 60°C for 12 h, nZVI/BC* was collected.

NZVI+BC was prepared directly by mixing BC and NZVI particles. Briefly, 1.0 g BC and 0.5 g NZVI were mixed in 30 mL ethanol in a beaker and dispersed by

ultrasonic cell breaker (SCIENTZ-II D) for 10 min (20°C, 570W). Then, the mixed NZVI+BC sample was separated by centrifugation at 3000 rpm for 5 min and dried in a vacuum oven (60°C, 12 h).

4.3. Characterization of BC and NZVI materials

The inner structure information of BC and NZVI materials was observed by C-TEM technique directly^{38,39}. These samples were firstly embedded into embedding agent, i.e., spurr resin, which is a mixture of 10 g vinyl cyclohexene dioxide (ERL 4221), 8 g diglycidyl ether of polypropylene glycol (DER 736), 25 g nonenyl succinic anhydride (NSA) and 0.3 g 2-dimethylaminoethanol (DMAE), purchased from SPI-Chem Supplies. Then, spurr resin embedded samples were heated at 70°C for 12 h, and cut into ultra-thin slices with thickness less than 90 nm by ultramicrotome (Leica, EMUC7, Germany) equipped with diamond knife (Figure S1). The slices were finally collected on copper mesh grids for observation of transmission electron microscopy (TEM) (JEM1200, JEOL, Japan).

X-ray diffraction patterns of samples were obtained by an X-ray diffractometer (XRD, Philips, Netherlands) equipped with a CuK α radiation source and scanned at a speed of 2° per min. Surface morphologies of samples were examined by using a field emission SEM (FE-SEM) (SIRON, FEI, Netherlands) equipped with an energy dispersive spectroscopy (Oxford Inca EDS) at a voltage of 25.0 KV.

4.4. Batch experiments for Fe²⁺ and Fe³⁺ by BC

Fe²⁺ and Fe³⁺ solution was prepared from FeSO₄·7H₂O and FeCl₃·6H₂O respectively. Batch experiment of Fe²⁺ and Fe³⁺ was conducted in 22 mL screw cap vials

with a total solution volume of 20 mL sealed with Teflon-coated septa under ambient temperature (25°C). Briefly, 20 mg or 200 mg BC was added into vials having 18 mL background solution with various pH value, prepared from DI water with 0.01 mol/L HCl and NaOH. Then, 2 mL Fe²⁺ or Fe³⁺ solution (500 mg/L or 1500 mg/L), was added into the vials to give the initial Fe²⁺ or Fe³⁺ concentration of 50 mg/L or 150 mg/L, respectively. The vials were capped and transferred to a horizontal shaker (DHZ-D) at 150 rpm for 48 h. After centrifugation at 3000 rpm for 5 min, supernatant was taken from vials, and filtered with a 0.22 µm needle filter to remove the solid. The pH value of supernatant was measured using a glass pH electrode (Mettler Toledo). The concentration of Fe²⁺ iron in supernatant was determined by UV-spectrophotometer (UV-2450, Shimadzu, Japan) with 1,10-phenanthroline spectrophotometric method at a maximum wavelength of 510 nm⁵³. For measurement of Fe³⁺, Fe³⁺ in supernatant was reduced to Fe²⁺ firstly by NH₂OH·HCl⁵⁴, and then, determined by UV-spectrophotometer with 1,10-phenanthroline spectrophotometric method⁵³.

The removal efficiency of Fe²⁺ or Fe³⁺ was calculated by Equation 1:

$$R\% = (C_0 - C_e) / C_0 * 100\% \quad (1)$$

Where C₀ (mg/L) is the initial concentration of Fe²⁺ or Fe³⁺ in solution, C_e (mg/L) is the equilibrium concentration of Fe²⁺ or Fe³⁺ in solution.

4.5. Precipitation of Fe²⁺ and Fe³⁺ in aqueous

Precipitation of Fe²⁺ and Fe³⁺ was also conducted in 22 mL screw cap vials. 18 mL background water at various pH, prepared from DI water with 0.01 mol/L HCl and NaOH, and 2 mL Fe²⁺ or Fe³⁺ solution (500 mg/L) were mixed in the vials, i.e., the

initial Fe^{2+} or Fe^{3+} concentration in vials was 50 mg/L. The vials were capped and transferred to a horizontal shaker (DHZ-D) at 150 rpm for 48 h under ambient temperature (25°C). After centrifugation at 3000 rpm for 5 min, pH value and equilibrium concentration of Fe^{2+} or Fe^{3+} in supernatant were measured with glass pH electrode (Mettler Toledo) and UV-spectrophotometer (UV-2450, Shimadzu, Japan), respectively^{53,54}.

4.6. Reduction of $\text{Fe}(\text{OH})_2$ and $\text{Fe}(\text{OH})_3$ by NaBH_4

For reduction experiments of $\text{Fe}(\text{OH})_2$ or $\text{Fe}(\text{OH})_3$ precipitates by NaBH_4 , 10 mL 0.25 M Fe^{2+} or Fe^{3+} solution and 10 mL 1.0 M NaOH solution were mixed and then centrifuged at 3000 rpm for 5 min to collect $\text{Fe}(\text{OH})_2$ or $\text{Fe}(\text{OH})_3$ precipitates. The collected $\text{Fe}(\text{OH})_2$ or $\text{Fe}(\text{OH})_3$ precipitate was washed with deoxidizing DI water for three times to remove excessive Fe^{2+} or Fe^{3+} ions. Before reduction by NaBH_4 , another 20 mL deoxidizing DI water was added to redisperse $\text{Fe}(\text{OH})_2$ or $\text{Fe}(\text{OH})_3$ precipitates. Then, 20 mL 1.0 M NaBH_4 was added dropwise under nitrogen atmosphere to reduce the precipitates. After reduction reaction, solid product was collected by centrifugation at 3000 rpm for 5 min, washed with deoxidizing DI water and ethanol, and finally dried in a vacuum oven at 60°C for 12 h.

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Author contributions

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Competing interests

The authors declare no competing financial interest.

Supporting Information

Analysis methods of $\text{Fe}^{2+}/\text{Fe}^{3+}$ in supernatants and solids in the precipitation experiments (Text 1), species distribution of $\text{Fe}^{2+}/\text{Fe}^{3+}$ in supernatants and solids in the precipitation experiments (Table S1), schematic of ultra-thin slice of NZVI/BC preparation (Figure S1), SEM image (Figure S2a) and SEM elemental mapping (Figure S2b) of NZVI/BC, N_2 adsorption–desorption isotherm (Figure S3a), pore size distribution (Figure S3 b) of BC, and C-TEM images of NZVI/BC* (S4 a, c, e) and BC (S4 b, d, f) were presented.

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Caption of Tables and Figures

Figure 1. SEM images (a, b) and C-TEM images (c, d) of BC.

Figure 2. SEM image (a) and C-TEM image (b) of NZVI.

Figure 3. SEM images (a, b) and EDX (c) spectrum of NZVI/BC, as well as XRD patterns (d) of BC, NZVI/BC, NZVI/BC*, NZVI+BC, NZVI, NaBH₄ treated Fe(OH)₂ and NaBH₄ treated Fe(OH)₃.

Figure 4. C-TEM images (a, b) of NZVI/BC.

Figure 5. SEM (a, b) and C-TEM images (c, d) of NZVI+BC.

Figure 6. pH-dependent percent removal curves of Fe³⁺ or Fe²⁺ with or without BC.

Figure 7. Pictures of suspensions of Fe(OH)₂ and Fe(OH)₃ precipitates before (a) and after treated with NaBH₄ solution (b).

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Figures

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