

Association Between Plasma Glycocalyx Component Levels and Poor Prognosis in Influenza Type A (H1N1)

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Research

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Abstract

Background: Influenza A virus infection causes a series of diseases, but the factors associated with disease severity are not fully understood. Disruption of the endothelial glycocalyx contributes to acute lung injury in sepsis, but has not been well studied in H1N1 influenza. We aim to determine whether the plasma glycocalyx components levels are predictive of disease severity in H1N1 influenza.

Methods: This prospective observational study included 53 patients with influenza A (H1N1), who were admitted to the Pulmonary and Critical Care Medicine and Intensive Care Unit, and 30 healthy controls between November 2017 and March 2019 in our hospital. Patients were grouped by severity and survival. We collected clinical data and blood samples at admission. Inflammatory factors (tumor necrosis factor- α , interleukin-6, interleukin-10) and endothelial glycocalyx components (syndecan-1, hyaluronan, heparan sulfate) were measured.

Results: The plasma levels of syndecan-1, hyaluronan, and heparan sulfate were significantly higher in patients with severe influenza A (H1N1) than in mild cases. At a cutoff point >81.0 ng/mL, syndecan-1 had a 92.1% sensitivity and 86.7% specificity for diagnosing severe H1N1. Syndecan-1 and hyaluronan were positively correlated with disease severity, which was indicated by the APACHE II and SOFA scores and lactate levels, and negatively correlated with albumin levels. Non-survivors had higher syndecan-1 levels than did survivors, and syndecan-1 was strongly predictive of 28-day mortality (area under the curve, 0.855).

Conclusions: An increased plasma syndecan-1 level was an independent risk factor for mortality and may be indicative of disease severity in patients with influenza A (H1N1).

Trial registration: CHINA, ChiCTR2000040921. Retrospectively registered

Background

Influenza A is an acute viral infection that causes mild upper respiratory tract infection and acute respiratory distress syndrome (ARDS). According to World Health Organization reports, influenza A causes global annual infection rates of 5% -10% in adults, 3–5 million cases of severe illness, and about 500,000 deaths [1, 2]. Early antiviral therapy with neuraminidase inhibitors can improve the outcome of influenza A [2]; however, a small number of patients experience a rapid progression to primary viral pneumonia or secondary bacterial infections, resulting in a significant number of deaths despite the use of antivirals [3]. Therefore, improving antivirals alone may not be sufficient to minimize morbidity and mortality, and the pathogenesis of influenza A needs to be further explored.

Severe influenza virus infections cause a dysregulated inflammatory response, leading to the release of proinflammatory cytokines in the lungs and blood, a condition often referred to as a “cytokine storm”. Inflammatory cytokines can lead to diffuse alveolar damage, and interstitial and airspace inflammation, which adversely affect outcomes in patients with severe influenza infections [4, 5]. An abnormal immune

response to influenza A can lead to endothelial damage, alteration of microvascular permeability, tissue edema, deregulation of coagulation, and even shock [6, 7].

Influenza A can directly and indirectly cause lung endothelial cell activation and injury, inducing microvascular leakage [8]. The surface of the vascular endothelium is coated with a “thick” glycocalyx, a dynamic and complex biochemical structure consisting of core proteins (syndecans and glypicans) and glycosaminoglycan chains (heparan sulfate [HS], hyaluronan [HA] etc.), which play a key role in limiting vascular permeability and regulating platelet and leukocyte adhesion [9]. Syndecan-1 (SDC-1) belongs to the family of syndecans, and is involved in leucocyte recruitment, the chemokine gradient, and extracellular matrix remodeling during inflammatory diseases [10]; it also plays a pivotal role in glycocalyx integrity and function. Glycocalyx dysfunction and component shedding have been described in many clinical pathophysiologic processes, including sepsis [11], ARDS [12], and coronavirus disease 2019 (COVID-19) [13]. Furthermore, elevated plasma SDC-1 and HA levels are associated with cumulative fluid volumes, degree of organ failure, and increased mortality in patients with sepsis [14, 15]. Lipopolysaccharide induced shedding of HS in the pulmonary endothelial glycocalyx uncovers endothelial cell surface adhesion molecules, thereby accelerating neutrophil adhesion and alveolar exudation [16]. Thus, the role of endothelial glycocalyx injury and activation in the pathogenesis of ARDS caused by sepsis is known, but the role of the endothelial glycocalyx in influenza A (H1N1) severity has not been elucidated. To our knowledge, no studies have compared plasma glycocalyx components in patients with mild and severe influenza A (H1N1).

We further hypothesized that the degree of endothelial glycocalyx degradation is associated with severity and mortality in patients with influenza A(H1N1). In this study, we aimed to determine whether the plasma levels of glycocalyx components are indicative of influenza A (H1N1) severity.

Methods

Study design and patient recruitment

This prospective observational study included patients admitted to the Department of Pulmonary and Critical Care Medicine (72 beds) and intensive care unit (48 beds) of Binzhou Medical University Hospital (Binzhou, Shandong, China), from November 2017 to March 2019. The inclusion criteria were the age of ≥ 18 years, and a positive real-time reverse transcription polymerase chain reaction (PCR) finding from an airway specimen. Viral specimens were collected from the patients via nasopharyngeal swab, whereas lower airway specimens were obtained via endotracheal tube and bronchoalveolar lavage fluid. Influenza A (H1N1) reverse transcription PCR testing was performed in accordance with the Centers for Disease Control and Prevention guidelines. A total of 30 age- and sex-matched healthy donors were recruited as controls. Patients who met any one of the following criteria were classified as severe: ARDS, shock, multiorgan failure, requiring ICU admission, or mechanical ventilation for medical reasons. This study was conducted in accordance with the amended Declaration of Helsinki, and the protocol was approved by the Ethics Committee of the Binzhou Medical University Hospital, and was registered in the Clinical

Trials Register (ChiCTR2000040921). Informed consent was obtained from patients (or their caregivers) and healthy controls before enrollment.

Data and blood sample collection

Demographic characteristics, including sex, age, comorbidity, and symptoms at onset of illness, were recorded on admission. The Sequential Organ Failure Assessment (SOFA) score, and the Acute Physiology, Age, Chronic Health Evaluation II (APACHE II) score were calculated to assess illness severity. Clinical outcomes were assessed according to the 28-day mortality rate.

Venous blood samples were collected from the patients and centrifuged at $4000 \times g$ for 10 min, and plasma was collected and stored at $-80\text{ }^{\circ}\text{C}$ until analysis. The lactate level, blood routine index, hepatic and renal function, and coagulation/fibrinolysis markers were analyzed in a standardized laboratory. Plasma levels of tumor necrosis factor- α (TNF- α) (950.090.096, Diaclone, France), interleukin-6 (IL-6) (950.030.096, Diaclone, France) and IL-10 (950.060.096, Diaclone, France), SDC-1 (950.640.096, Diaclone, France), HA (DHYAL0, R&D Systems, USA), and HS (MBS285287, MyBioSource, USA) were measured using an enzyme-linked immunosorbent assay kit according to the manufacturer's instructions.

Statistical analysis

All categorical variables are expressed as numbers and percentages. Between-group comparisons of frequencies were analyzed using the chi-square test. Summary statistics for normally distributed continuous variables are presented as mean \pm standard deviation, and non-normally distributed continuous variables are presented as median and interquartile range. Differences between groups were tested for significance using either the non-parametric Mann-Whitney U test or an unpaired Student's t-test for two groups and the Kruskal-Wallis analysis of variance for more than two groups. Correlations were analyzed by the Spearman rank correlation test. The efficacy of each parameter in predicting the diagnostic power of each marker is expressed as the area under curve (AUC) using the receiver operating characteristic (ROC) analysis. Odds ratios with 95% confidence intervals (CIs) were computed using a multivariate logistic regression model with 28 day mortality of H1N1 cases as the dependent variable. For all tests, a P-value of less than 0.05 was considered statistically significant. Statistical analyses were conducted using IBM SPSS 25.0.

Results

Patient characteristics

We initially screened 59 hospitalized patients with a high suspicion of influenza A infection (Fig. 1). We excluded one patient with avian influenza H7N9, and five patients with influenza B antigen (+). Thus, we enrolled 53 patients with a positive PCR test for influenza A (H1N1) and 30 healthy controls. The 30 healthy controls comprised 15 men and 15 women with a median age of 59 years. The 53 patients with influenza A (H1N1) comprised 20 men (37.73%) and 33 women (62.26%) with a median age of 57 years. Using the program for diagnosis and treatment of influenza A (H1N1), H1N1-infected patients were

divided into two different groups based on clinical severity: the mild group (n = 15) and the severe group (n = 38). There were 30 healthy controls, 15 of which males, and the median age was 59 years. Of these 58 patients with influenza A (H1N1), 23 were men (39.66%) and 35 were women (60.34%); the median age was 57 years. The patient characteristics are presented in Table 1.

Table 1
Characteristics and symptoms of patients infected with Influenza A (H1N1).

Characteristics	Control subjects (N = 30)	Total Influenza A (H1N1) (N = 53)	Mild Influenza A (H1N1) (N = 15)	Severe Influenza A (H1N1) (N = 38)	P Value
Age, years (Median, P25-P75)	59 (53–63)	57 (47, 69)	56 (47, 76)	59 (44, 69)	0.722
Sex (Male/Female)	15/15	20/33	7/8	13/25	0.399
Smoker (n, %)		20, 37.73%	10, 66.67%	10, 26.32%^a	0.006
Lactate (mmol/L)		1.5 (1.0, 2.7)	1.1 (0.9, 1.5)	2.1 (0.9, 3.1)^b	0.012
≥ 2 (n, %)		21, 39.62%	1, 6.67%	20, 52.63%^a	0.002
APACHE II score		11 (8, 15)	8 (5, 9)	14 (10,18)^b	< 0.001
SOFA score		5 (2, 7)	2 (1, 2)	6 (4, 8)^b	< 0.001
Comorbidity (n, %)					
Hypertension		15, 28.30%	2, 13.33%	13, 34.21%	0.129
Cardiovascular disease		6, 11.32%	2, 13.33%	4, 10.53%	0.771
Diabetes		8, 15.09%	1, 6.67%	7, 18.42%	0.282
Cerebrovascular disease		3, 5.66%	1, 6.67%	2, 5.26%	0.842
Chronic obstructive		3, 5.66%	1, 6.67%	1, 2.63%	0.487
Asthma		2, 3.77%	2, 13.33%	0, 0^a	0.022
Chronic hepatitis		2, 3.77%	0, 0	2, 5.26%	0.365
Malignancy		3, 5.66%	2, 13.33%	1, 2.63%	0.144
Without		9, 16.98%	1, 6.67%	8, 21.05%	0.209
Symptoms at onset of illness					
Highest temperature, °C		38.5 (38.0, 39.1)	38.6 (37.9, 39.1)	38.5 (38.0, 39.1)	0.993

Data are expressed as median (inter-quartile range), mean ± SD, or number (%). Abbreviations: APACHE, Acute Physiology, Age, Chronic Health Evaluation; SOFA, Sequential organ failure assessment; P/F, PaO₂/FiO₂. a, vs. Mild group, χ^2 tests; b, vs. Mild group, Mann - Whitney U test.

Characteristics	Control subjects (N = 30)	Total Influenza A (H1N1) (N = 53)	Mild Influenza A (H1N1) (N = 15)	Severe Influenza A (H1N1) (N = 38)	P Value
< 37.3 (n, %)		5, 9.43%	2, 13.33%	3, 7.89%	0.542
37.3–38 (n, %)		13, 24.53%	3, 20.00%	10, 26.31%	0.630
38.1–39 (n, %)		19, 35.85%	6, 40.00%	13, 34.21%	0.692
> 39 (n, %)		16, 30.19%	4, 26.67%	12, 31.58%	0.726
Cough (n, %)		42, 79.25%	11, 73.33%	31, 81.58%	0.505
Sputum production (n, %)		33, 62.26%	9, 60.00%	24, 41.38%	0.831
Oppression (n, %)		22, 41.51%	9, 60.00%	13, 34.21%	0.086
Chilly (n, %)		14, 26.41%	4, 26.67%	10, 26.32%	0.979
Fatigue (n, %)		8, 15.09%	2, 13.33%	6, 11.32%	0.822
Myalgia (n, %)		5, 9.43%	2, 13.33%	3, 5.17%	0.542
Headache (n, %)		3, 5.66%	2, 13.33%	1, 2.63%	0.129
Chest pain (n, %)		4, 7.55%	3, 20.00%	1, 2.63%^a	0.031
Pharyngalgia (n, %)		3, 6.66%	1, 6.67%	2, 3.45%	0.842
Anorexia (n, %)		5, 9.43%	0, 0	5, 8.62%	0.134
Vomiting (n, %)		1, 1.88%	0, 0	1, 1.72%	0.526
Dizziness (n, %)		3, 5.66%	1, 6.67%	2, 3.45%	0.842
P/F ratio		223 (112, 311)	328 (297, 366)	164 (76, 205)^b	< 0.001
During hospitalization stay					
Ventilatory support (n, %)					
Nasal catheters for oxygen			10, 66.67%	2, 5.26%	
Non invasive ventilation			0, 0	6, 15.79%	

Data are expressed as median (inter-quartile range), mean \pm SD, or number (%). Abbreviations: APACHE, Acute Physiology, Age, Chronic Health Evaluation; SOFA, Sequential organ failure assessment; P/F, PaO₂/FiO₂. a, vs. Mild group, χ^2 tests; b, vs. Mild group, Mann - Whitney U test.

Characteristics	Control subjects (N = 30)	Total Influenza A (H1N1) (N = 53)	Mild Influenza A (H1N1) (N = 15)	Severe Influenza A (H1N1) (N = 38)	P Value
High flow nasal cannula			0, 0	8, 21.05%	
Invasive mechanical ventilation			0, 0	22, 57.89%	
Steroid treatment		20, 37.74%	6, 40.00%	14, 36.84%	0.831
Hospital length of stay, days		11 (6, 18)	8 (6, 14)	14 (9, 21)^a	0.044
ICU length of stay, days			-	9 (4, 13)	
28-day mortality, (n, %)		16, 30.19%	-	16, 42.11%	
Data are expressed as median (inter-quartile range), mean \pm SD, or number (%). Abbreviations: APACHE, Acute Physiology, Age, Chronic Health Evaluation; SOFA, Sequential organ failure assessment; P/F, PaO ₂ /FiO ₂ . a, vs. Mild group, χ^2 tests; b, vs. Mild group, Mann - Whitney U test.					

The most common clinical features at illness onset included a fever of $> 38^{\circ}\text{C}$ (n = 35, 66.04%), cough (n = 42, 79.25%), sputum production (n = 33, 62.26%), oppression (n = 22, 41.51%) and chilly (n = 14, 26.41%). Less common symptoms were myalgia, chest pain, pharyngalgia, anorexia, etc. The most common comorbidities were hypertension (n = 15, 28.30%), cardiovascular disease (n = 6, 11.32%), and diabetes (n = 8, 15.09%); however, 9 patients presented without comorbidity. Two patients in the mild group presented with asthma; this comorbidity was not present among patients in the severe group. The P/F ratio tended to be lower in the severe group than in the mild group. Invasive mechanical ventilation was required in $> 50\%$ of the patients in the severe group. There was no statistically significant difference in the number of patients taking steroid treatment between the two groups. The length of hospital stay was significantly longer in the severe group than in the mild group. All participants were followed-up for 28 days, at which point there were 37 survivors and 16 non-survivors.

Laboratory records and inflammation markers

The median white blood cell count in both the mild and severe groups were within the normal range; however, the count was significantly higher in the severe group; 50% of the patients in the severe group had white blood cell counts of $> 10 \times 10^9/\text{L}$. The lymphocyte count and percentage were significantly lower in the severe group than in the mild group. The levels of alanine aminotransferase (ALT) and aspartate aminotransferase (AST) were significantly higher in the severe group than in the mild group. Concurrently, the proportion of patients with increased AST was significantly higher in the severe group than in the mild group (24/38 [63.16%] vs. 1/15 [6.67%], $P < 0.001$). The lactate dehydrogenase and procalcitonin levels were significantly higher in the severe group than in the mild group. However, the

albumin level was significantly lower in the severe group than in the mild group. The severe group also exhibited marked increases in TNF- α , IL-6, and IL-10 levels relative to the mild group. The laboratory findings and inflammation markers of patients with influenza A (H1N1) are presented in Table 2.

Table 2

Laboratory records and inflammation markers of patients infected with Influenza A (H1N1) on admission.

Characteristics	Control subjects (N = 30)	Mild Influenza A (H1N1) (N = 15)	Severe Influenza A (H1N1) (N = 38)	P Value
White blood cell count, ×10⁹/L	4–10	7.2 (5.5, 9.0)	10.3 (6.0, 16.1)^b	0.024
< 4		2, 13.33%	2, 5.26%	0.044
4–10		11, 73.33%	17, 44.74%	...
> 10		2, 13.33%	19, 50.00%	...
Lymphocyte count, ×10⁹/L	1–3	0.9 (0.6, 1.6)	0.6 (0.5, 0.9)^b	0.042
≤ 0.5		3, 20.00%	16, 42.11%^a	< 0.001
0.5- 1.0		7, 46.67%	15, 39.47%	...
≥ 1.0		5, 33.33%	7, 18.42%	...
Lymphocyte percentage	20–40	14.5 (8.6, 22.4)	7.2 (4.6, 12.4)^b	0.017
CD4 + T Lymphocytes (%)	30.1–44.4	41.4 ± 8.2	36.4 ± 11.8	0.244
CD8 + T Lymphocytes (%)	20.7–29.4	22.2 ± 5.3	23.2 ± 8.0	0.733
CD4/CD8	1.02–1.94	1.77 (1.40, 2.66)	1.71 (0.92, 2.41)	0.569
Platelet count, × 10 ⁹ /L	100–300	233.5 (173.0, 291.0)	154.0 (105.0, 255.0)	0.508
D-dimer (µg/mL)	0-0.5	0.32 (0.22, 0.68)	3.75 (1.50, 13.27)^b	< 0.001
Albumin, g/L	40–55	38.0 ± 4.8	27.6 ± 4.0^c	< 0.001
Alanine aminotransferase, U/L	9–50	18.5 (16.4, 39.9)	34.3 (20.5, 56.1)^b	< 0.001
≤ 35		11	21	0.226
>35		4	17	...
Aspartate aminotransferase, U/L	15–40	23.1 (17.8, 27.0)	47.8 (34.8, 82.5)^b	0.033
Data are expressed as median (inter-quartile range), mean ± SD, or number (%). Abbreviations: TNF, tumor necrosis factor; IL-6, interleukin-6; IL-10, interleukin-10. ^a , vs. Mild group, χ^2 tests; ^b , vs. Mild group, Mann - Whitney U test; ^c , unpaired Student t test.				

Characteristics	Control subjects (N = 30)	Mild Influenza A (H1N1) (N = 15)	Severe Influenza A (H1N1) (N = 38)	<i>P</i> Value
≤ 40		14	14 ^a	< 0.001
> 40		1	24	...
Creatinine, μmol/L	0-135	60.3 (43.6, 66.9)	59.8 (47.3, 91.3)	0.622
Lactate dehydrogenase, U/L	0-430.6	223.9 (193.9, 239.5)	635.7 (375.6, 864.4) ^b	< 0.001
Creatine kinase	25–200	65.9 (45.1, 130.5)	86.3 (42.8, 272.3)	0.365
Procalcitonin, μg/L	0-0.5	0.06 (0.04, 0.13)	0.68 (0.40, 4.80) ^b	0.001
Inflammation markers				
TNF-α (pg/ml)	3.7 (1.3, 5.6)	8.4 (6.7, 13.3)	13.3 (11.7, 23.2) ^b	0.021
IL-6 (pg/ml)	1.9 (0.5, 5.9)	3.4 (2.6, 5.5)	71.2 (16.6, 182.0) ^b	< 0.001
IL-10 (pg/ml)	5.9 (4.7, 6.5)	9.1 (6.6, 15.9)	22.4 (12.3, 52.6) ^b	0.004
Data are expressed as median (inter-quartile range), mean ± SD, or number (%). Abbreviations: TNF, tumor necrosis factor; IL-6, interleukin-6; IL-10, interleukin-10. ^a , vs. Mild group, χ ² tests; ^b , vs. Mild group, Mann - Whitney U test; ^c , unpaired Student t test.				

Glycocalyx component levels and biomarker accuracy

The plasma levels of SDC-1 (257.9 [142.1–658.8] vs. 58.6 [49.4–75.9] ng/ml), HA (334.4 [159.3–413.9] vs. 156.6 [116.1–230.5] ng/ml), and HS (305.1 [193.9–516.9] vs. 228.8 [198.9–312.4] ng/ml) were significantly higher in the severe group than in the mild group (Fig. 2). We conducted a ROC analysis to determine the utility of glycocalyx components as markers of severe disease. The diagnostic accuracy for the identification of patients progressing to severe influenza A (H1N1) was highest for SDC-1, with an AUC of 0.942 (95% CI: 0.881–1.000; *P* < 0.001; Fig. 2d). At a cutoff point of > 81.0 ng/mL, SDC-1 had a sensitivity of 92.1% and specificity of 86.7% for diagnosing severe influenza A (H1N1).

SDC-1 levels and disease severity

To identify the clinical relevance of endothelial glycocalyx components, we analyzed the correlation between the following variables: SDC-1, HA, and HS levels; SOFA score; APACHE II score; and various

biochemical parameters including lactate, albumin, inflammatory markers (TNF- α , IL-6, and IL-10), and global hemostatic markers (platelet count and D-dimer).

The SOFA and APACHE II scores indicate organ dysfunction and disease severity in critically ill patients independent of the underlying disease, and are therefore commonly used to predict mortality in ICU patients. Spearman's correlation analysis showed that the SDC-1 and HA levels were significantly correlated with the APACHE II ($r = 0.526, P < 0.001$; $r = 0.439, P = 0.001$) and SOFA scores ($r = 0.515, P < 0.001$; $r = 0.409, P = 0.002$) (Table 3). High lactate level is traditionally considered as a marker of tissue hypoxia, and can be considered as a warning signal for organ dysfunction [17]. Among the glycocalyx components, lactate levels were significantly correlated with SDC-1 ($r = 0.392, P = 0.004$) and HA ($r = 0.372, P = 0.006$; Table 3) levels. The platelet count and D-dimer concentration reflect coagulation function. SDC-1 and HA were negatively correlated with the platelet count and positively correlated with the D-dimer concentration. We also observed that SDC-1 and HA were positively correlated with IL-6 and IL-10 levels (Table 3).

Table 3
Correlations between glycocalyx components levels and various clinical parameters.

Characteristics	SDC-1		HA		HS	
	r	PValue	r	PValue	r	PValue
APACHE II score	0.526	< 0.001	0.439	0.001	0.047	0.739
SOFA score	0.515	< 0.001	0.409	0.002	0.063	0.652
Lactate	0.392	0.004	0.372	0.006	0.198	0.155
Albumin	-0.639	< 0.001	-0.399	0.003	0.03	0.831
Platelet	-0.313	0.017	-0.473	< 0.001	0.071	0.616
D-dimer	0.593	< 0.001	0.299	0.031	0.209	0.138
TNF- α (pg/ml)	0.276	0.046	0.245	0.077	-0.13	0.352
IL-6 (pg/ml)	0.63	< 0.001	0.549	< 0.001	0.20	0.15
IL-10 (pg/ml)	0.50	< 0.001	0.622	< 0.001	0.152	0.279

Abbreviations: APACHE, Acute Physiology, Age, Chronic Health Evaluation; SOFA, Sequential organ failure assessment; TNF, tumor necrosis factor; IL-6, interleukin-6; IL-10, interleukin-10; SDC-1, syndecan-1; HS, heparan sulfate; HA, hyaluronan. r, Spearman's correlation coefficient.

Predictors of 28-day mortality

All study participants were followed-up for 28 days and divided into the survivor and non-survivor groups (Table 4). The APACHE II and SOFA scores and lactate, D-dimer, IL-6, IL-10, SDC-1, and HA levels were

significantly higher in non-survivors than in survivors. The albumin level and lymphocyte percentage were significantly lower in non-survivors than in survivors. We constructed ROC curves to determine the sensitivity and specificity of the APACHE II and SOFA scores, laboratory records, inflammation markers, and endothelial glycocalyx markers, in order to predict 28-day mortality (Fig. 3). The AUC for the SDC-1 was 0.855 (95% CI, 0.75–0.96), which was higher than that for other indicators. At a cutoff point of > 173.9 ng/ml, SDC-1 provided a specificity of 81.3% and a sensitivity of 70.3% for predicting 28-day mortality.

Table 4

The APACHE II and SOFA scores, laboratory records, inflammation markers, and endothelial glycocalyx markers in the survivors and non-survivors (28-day death) on the admission.

Characteristics	Survivor (N = 37)	Non-survivor (N = 16)	P Value
APACHE II score	9 (7, 13)	15 (14, 18) ^b	< 0.001
SOFA score	3 (2, 6)	7 (5, 9) ^b	0.001
Lactate	1.2 (0.9, 1.6)	2.2 (1.5, 3.5) ^b	0.001
Lymphocyte count, ×10 ⁹ /L	0.9 (0.5, 1.4)	0.6 (0.3, 0.9) ^b	0.048
Lymphocyte percentage	11.5 (7.1, 18.6)	5.4 (4.2, 7.8) ^b	0.002
Platelet	210.5 (144.0, 272.0)	136.5 (85.5, 230.0) ^b	0.075
D-dimer (µg/mL)	1.52 (0.49, 3.72)	6.26 (2.27, 22.1) ^b	0.012
Albumin, g/L	32.2 ± 6.6	27.2 ± 4.2 ^c	0.007
Inflammation markers			
TNF-α (pg/ml)	11.6 (8.4, 21.6)	14.1 (10.0, 35.7) ^b	0.322
IL-6 (pg/ml)	15.4 (4.5, 96.7)	82.5 (15.8, 376.1) ^b	0.046
IL-10 (pg/ml)	11.27 (7.0, 23.1)	30.1 (14.1, 57.6) ^b	0.036
Endothelial glycocalyx markers			
SDC-1 (ng/ml)	117.7 (65.3, 258.5)	576.3 (204.7, 755.3) ^b	0.003
HA (ng/ml)	190.4 (108.4, 348.2)	400.5 (213.1, 431.3) ^b	0.02
HS (ng/ml)	233.8 (178.8, 331.7)	419.7 (201.4, 1446.6) ^b	0.034
Data are expressed as median (inter-quartile range), mean ± SD, or number (%). Abbreviations: APACHE, Acute Physiology, Age, Chronic Health Evaluation; SOFA, Sequential organ failure assessment; TNF, tumor necrosis factor; IL-6, interleukin-6; IL-10, interleukin-10; SDC-1, syndecan-1; HS, heparan sulfate; HA, hyaluronan. ^b , vs. Mild group, Mann - Whitney U test; ^c , unpaired Student t test.			

Independent risk factors for mortality

Univariate logistic regression showed that the variables associated with 28-day mortality in patients with influenza A (H1N1) were a reduced lymphocyte count and percentage, albumin level, elevated lactate, IL-6, IL-10, SDC-1, and HA. Multivariate logistic regression indicated that SDC-1 was an independent risk factor

for mortality; a small increase in plasma SDC-1 concentration of only 1 ng/ml was associated with a 0.4% increased risk of 28-day mortality, with an odds ratio of 1.004 (95% CI: 1.001–1.007). The multivariate logistic regression results are provided in Table 5.

Table 5

Multivariate logistic regression of risk factors for hospitalized patients with influenza A (H1N1).

Characteristics	Regression coefficient	odds ratio	95% confidence interval	P Value
SDC-1	0.004	1.004	1.001–1.007	0.032
Albumin	-0.074	0.929	0.804–1.072	0.312
HA	0.001	1.001	0.996–1.007	0.620
Abbreviations: SDC-1, syndecan-1; HA, hyaluronan.				

Discussion

In this study, we aimed to determine whether the plasma levels of glycocalyx components are indicative of influenza A (H1N1) severity. We found that SDC-1, HS, and HA levels were significantly higher in patients with severe influenza A (H1N1) than in patients with mild disease. In addition, the severity of endothelial glycocalyx shedding was closely associated with disease severity, and facilitated the identification of patients with severe influenza A (H1N1). Furthermore, we found that the level of SDC-1 was an independent risk factor for mortality among patients with influenza A (H1N1). To our knowledge, this study is the first to compare plasma glycocalyx components in patients with mild and severe influenza A (H1N1).

Pandemic 2009 influenza A (H1N1) viral infections continue to be a public health threat [18]. Influenza A (H1N1)-infected patients most often have a mild clinical course; only 6–31% of hospitalized patients with influenza A require treatment in the ICU [19–21]. However, this study observed a higher proportion (71.7%) of hospitalized patients with influenza A (H1N1) requiring treatment in the ICU. We considered that a large number of patients with mild influenza A (H1N1) received oseltamivir in the early stages of upper respiratory tract infection, thus reducing the positive rate of influenza A (H1N1) virus testing, and the inclusion of patients with mild influenza A (H1N1). In this study, the proportion of patients with a history of smoking and chest pain was significantly higher in the severe group than in the mild group. Furthermore, we found that approximately 21% of patients with severe H1N1 had no underlying disease. Several studies have found that both young subjects and adults may develop a severe clinical course of H1N1 infection without having any known risk factors [19, 22]. The underlying pathogenic mechanisms have not been fully elucidated. In the ICU, patients with influenza A (H1N1) often present with viral pneumonia, severe hypoxemic respiratory failure, and ARDS. Currently, treatment for patients with severe influenza A (H1N1) is limited to antiviral drugs and symptomatic treatment, and which mainly consists of the optimization of oxygen supply and transfer through ventilation.

The severity of influenza A has been attributed to a systemic and inflammatory process that damages not only the lungs, but also multiple organs, including the central nervous system and cardiovascular disease [23, 24]. In our study, we found that patients with severe influenza A (H1N1) had increased levels of ALT, AST, and D-dimer, indicating injuries to multiple organs, including the liver, and coagulation disorders. This systemic form of influenza A may be due to inflammation and vascular endothelial cell injury [25]. The cytokine storms caused by influenza have been associated with proinflammatory response disorder, which may lead to significant immunosuppression and poor prognosis [5, 26, 27]. The levels of pro-inflammatory cytokines are closely related to outcomes in patients with severe influenza infections [26–28]. This is consistent with our observation of significantly higher levels of inflammatory cytokines (IL-6 and IL-10) in non-survivors compared to survivors.

The levels of endothelial glycocalyx components, including SDC-1, HS, and HA, are useful biomarkers for sepsis [14, 15], ARDS [12, 16], and COVID-19 [13, 29]. Further, the APACHE II and SOFA scores may be used to assess the risk of death in critically ill patients, regardless of the primary disease. We found that plasma levels of SDC-1 and HA were significantly higher in patients with severe H1N1 than in patients with mild H1N1, and were positively correlated with the APACHE II and SOFA scores. Furthermore, at a cutoff point of > 173.9 ng/ml, SDC-1 showed a specificity of 81.3% and sensitivity of 70.3% for predicting 28-day mortality. Thus, SDC-1 is a simple and reliable predictor of severity and mortality among hospitalized patients with influenza A (H1N1).

The importance of platelets in the regulation of hemostasis and blood coagulation is well-known. Chappell et al. reported that protection of glycocalyx shedding reduces platelet adhesion in ischemia/reperfusion injury [30]. In addition, Fraser et al. reported that endothelial glycocalyx degradation in critically ill COVID-19 patients had implications for microvascular platelet aggregation [31]. We previously found that increasing levels of SDC-1 can be used as a biomarker for predicting DIC development with sepsis [10], and that non-anticoagulant heparin can improve coagulation by inhibiting the activity of heparinase and reducing the shedding of glycocalyx in sepsis rats [32]. In the present study, we found that the levels of SDC-1 and HA were negatively correlated with the platelet count in patients with influenza A (H1N1). It is well known that a sharp increase in D-dimer, a secondary fibrinolytic specific molecular marker, typically indicates the existence of a thrombus. Wang reported that abnormally increased D-dimer at the preliminary diagnosis is a general predictor of respiratory failure or even ARDS in patients with 2009 novel influenza A (H1N1) [33]. Our study found that the D-dimer level was significantly higher in non-survivors than in survivors, and was positively correlated with SDC-1 and HA levels.

Albumin has multiple biological activities including antioxidant effects, and maintains vessel wall integrity. Although albumin has a net negative charge, its amphoteric nature promotes tight binding to the glycocalyx, with the net effects of reducing hydraulic conductivity across the vascular barrier, resisting glycocalyx degradation (i.e., protecting against shedding), and contributing to the maintenance of vascular integrity and normal capillary permeability [34, 35]. In this study, we observed a significant negative correlation ($r = -0.639$) between the plasma albumin level and SDC-1 level. Although reduced albumin was not identified as an independent risk factor for disease development in the multivariate

logistic regression analysis, there was a strong correlation between the albumin level and glyocalyx function.

Our study has several limitations. First, this study was conducted at a single center and involved a relatively small number of patients with influenza A (H1N1); our findings require large-scale clinical validation in order to be generalized. Therefore, our data should be interpreted with caution. Second, the data presented here were based on single time-point measurements and were not consecutive. Although decreased SDC-1 was observed to be an independent risk factor for 28-day mortality in patients with influenza A (H1N1), the dynamic change was unknown, and the predictive value needs further evaluation. Despite the above limitations, we believe that our study has yielded important and novel findings regarding the prediction of mortality in hospitalized patients with influenza A (H1N1).

Conclusions

Plasma glyocalyx component levels are significantly higher in patients with severe influenza A (H1N1) than in mild cases. Specifically, SDC-1 and HA are closely correlated with APACHE II and SOFA scores, albumin, platelets, and D-dimer. Plasma SDC-1 levels of > 173.9 ng/ml were independently associated with 28-day mortality in critically ill patients with severe influenza A (H1N1). Thus, increased plasma SDC-1 may be a biomarker of disease severity and could be useful for predicting 28-day mortality in patients with influenza A (H1N1).

Abbreviations

ALT: alanine aminotransferase; APACHE II: Acute Physiology, Age, Chronic Health Evaluation II; AST: aspartate aminotransferase; ARDS: acute respiratory distress syndrome; AUC: area under curve; COVID-19: coronavirus disease 2019; CIs: confidence intervals; HA: hyaluronan; HS: heparan sulfate; IL-6: interleukin-6; IL-10: interleukin-10; ROC: receiver operating characteristic; RT-PCR: reverse transcription polymerase chain reaction; SDC-1: syndecan-1; SOFA: Sequential Organ Failure Assessment; TNF- α : tumor necrosis factor- α

Declarations

Ethics approval and consent to participate

The study was approved by the Human Research Ethics Committee of Binzhou Medical University Hospital and was conducted in accordance to the ethical standards set forth in the Declaration of Helsinki. The study was registered in the Clinical Trials Register (ChiCTR2000040921). All patients provided written informed consent on the day of admission.

Consent for publication

Not applicable.

Availability of data and material

The datasets used in this study are available from the corresponding author on reasonable request.

Competing interests

The authors declare that they have no competing interests.

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Authors' contribution:

XH, FL and HH T drafted the study and wrote the manuscript. HR H, QM S, DH and WW Z performed the statistical analyses and interpreted the data. GQ K, FY N, XH M and JL F contributed biomarker measurements and analyses. T W and XZ W supervised the study design, data interpretation, and manuscript preparation. All authors made significant intellectual contributions to the final manuscript and approve its submission.

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Figures

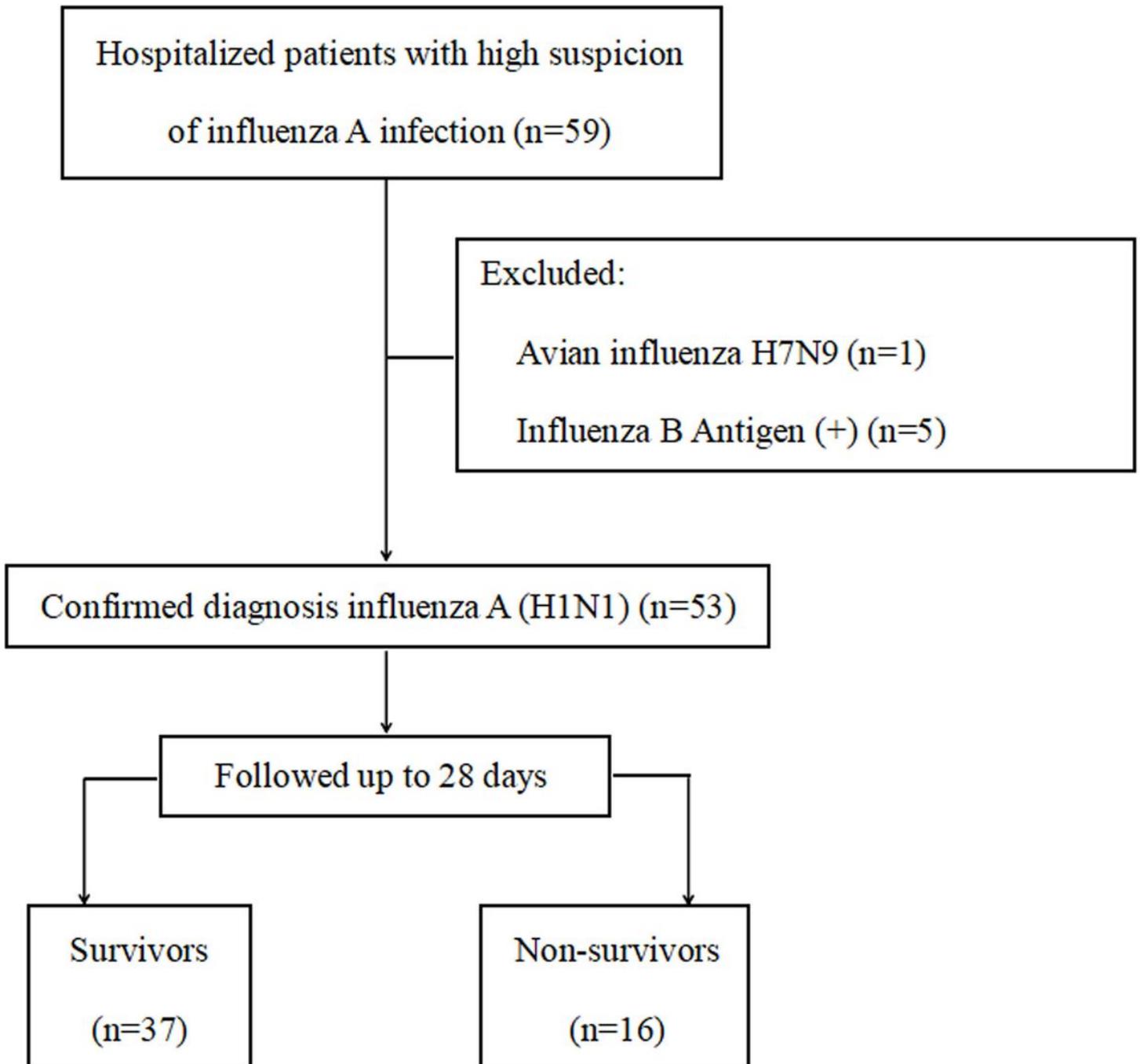


Figure 1

Flowchart of the study.

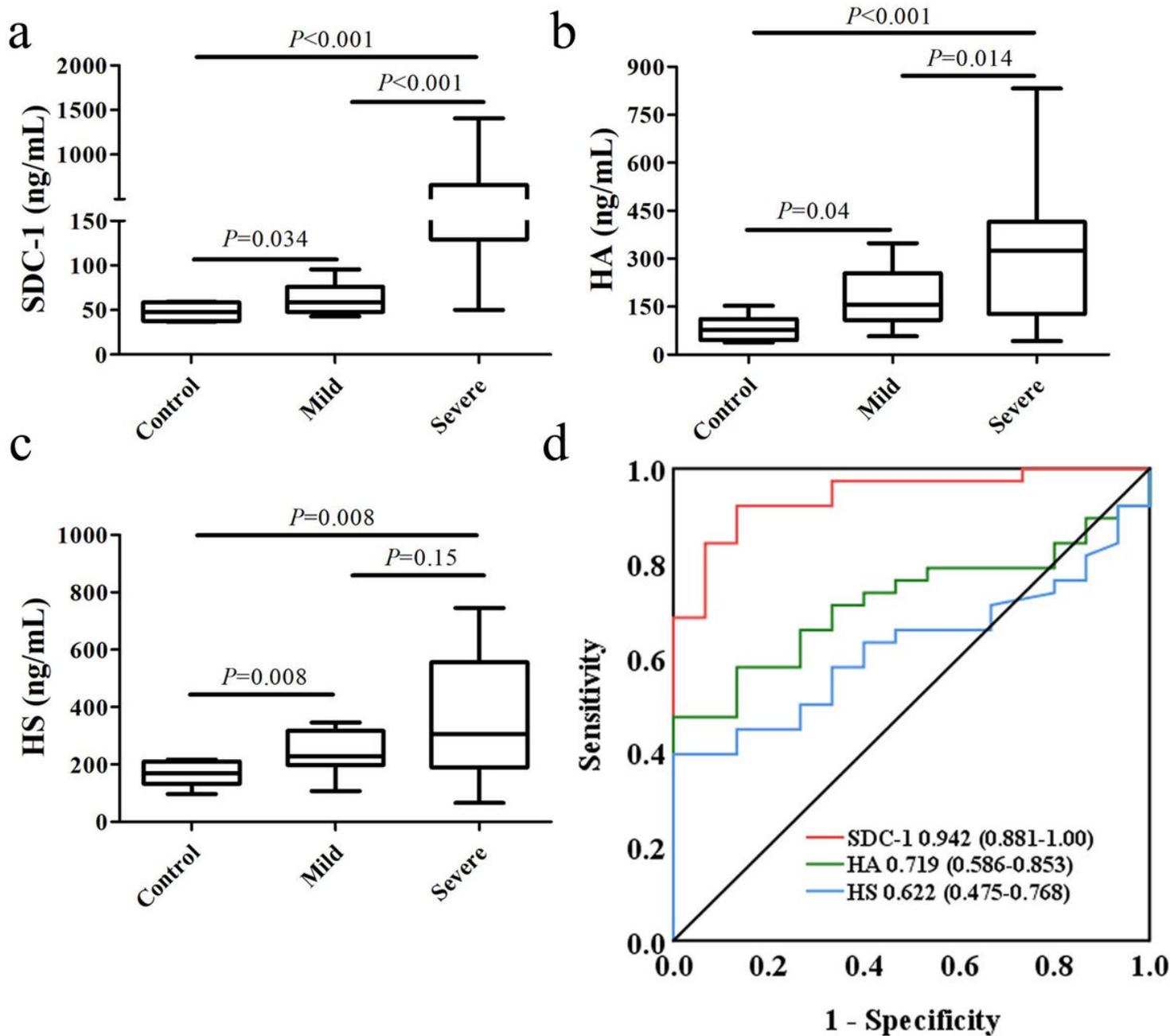


Figure 2

Plasma levels of glyocalyx components and ROC curves for the prediction of severe influenza A (H1N1). Plasma SDC-1 (a), HA (b), and HS (c) levels were significantly elevated in septic shock patients compared with those in sepsis patients. ROC curves (d) for the prediction of septic shock. AUC (95% confidence interval). The red rulers in figures A and B represent the median with range. The red rulers in figure C represent the mean with SD. Three-group comparisons of frequencies were analyzed by Kruskal-Wallis test. SDC-1, syndecan-1; HS, heparan sulfate; HA, hyaluronan.

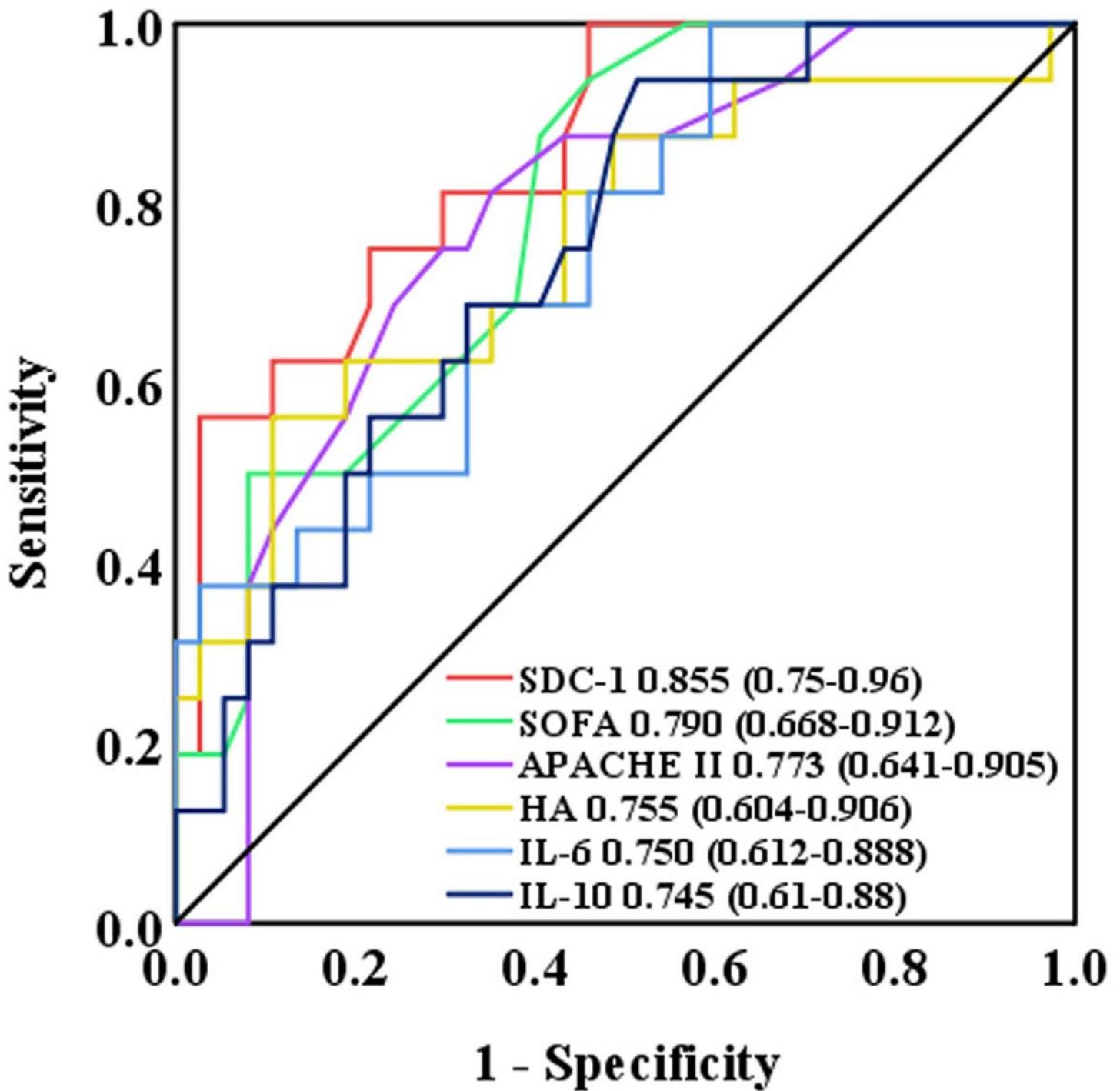


Figure 3

ROC curves for the prediction of 28-day mortality in patients with influenza A (H1N1).