

Preprints are preliminary reports that have not undergone peer review. They should not be considered conclusive, used to inform clinical practice, or referenced by the media as validated information.

Flux Decline and Fouling Analysis in Reverse Osmosis of Watermelon Juice

Quoc Dat Lai (Iqdat@hcmut.edu.vn)

Ho Chi Minh City University of Technology (HCMUT)

Ngoc Thuc Trinh Doan Ho Chi Minh City University of Technology (HCMUT)

Hoang Dung Nguyen

Ho Chi Minh City University of Technology (HCMUT)

Research Article

Keywords: Reverse osmosis, watermelon juice, concentration, modelling, lycopene, antioxidant activity

Posted Date: March 30th, 2022

DOI: https://doi.org/10.21203/rs.3.rs-1487272/v1

License: (a) This work is licensed under a Creative Commons Attribution 4.0 International License. Read Full License

FLUX DECLINE AND FOULING ANALYSIS IN REVERSE OSMOSIS OF WATERMELON JUICE

2		Lai Quoc Dat ^{1,2*} , Doan Ngoc Thuc Trinh ^{1,2} , Nguyen Hoang Dung ^{1,2}
3		*Corresponding Authors' email: lqdat@hcmut.edu.vn
4	1)	Department of Food Technology, Faculty of Chemical Engineering, Ho Chi Minh City University of
5		Technology (HCMUT), 268 Ly Thuong Kiet Street, District 10, Ho Chi Minh City, Vietnam
6	2)	Vietnam National University Ho Chi Minh City, Linh Trung Ward, Thu Duc District, Ho Chi Minh City,
7		Vietnam
8		
9	A	bstract

10 In this paper, the concentration of watermelon juice by HR98PP membrane was investigated at 30 and 40 bar 11 of operating pressure. The semi-empirical model of permeate flux was determined. The predominant fouling mechanism in the concentration was found to be complete blocking by Hermia's model. Recovery yield and content 12 of total solid, lycopene and antioxidant capacity in concentrate was analyzed. Results indicated that, concentration of 13 14 watermelon juice by HR98PP at 40 bar exhibited the higher effectiveness than that at 30 bar. At 40 bar and 2.25 of 15 CF, contents of total sugar (TS), lycopene and antioxidant capacity in concentrate were 171.31 g/L, 83.9 mg/L and 16 124.74 mg TEAC/L, respectively. Recovery yields of TS, lycopene and antioxidant capacity in concentrate were 17 98.67. 90.03 and 82.39 %, respectively. Maximum CF value of concentration at 40 bar of operating pressure was 2.64. 18 The mathematical model was built for estimation of the change in the contents of lycopene and TS versus time. 19 Maximum theoretical content of lycopene and TS at 40 bar were estimated as 185 mg/L and 341 g/L, respectively. 20 The cleaning procedure with NaOH solution at pH10 could fully recovery permeate flux after the concentration. 21 Results imply that, concentration of watermelon juice by HR98PP was feasible. 22

- 23

Keywords: Reverse osmosis; watermelon juice; concentration; modelling; lycopene; antioxidant activity.

- 24
- 25

1. Introduction

27 Watermelon (Citrullus lanatus) is a fruit with high content of lycopene: 8.20 - 59.17 mg/100 g watermelon 28 flesh (Oberoi and Sogi 2017). Due to richness of lycopene, watermelon juice exhibits the high antioxidant capacity 29 (Neglo et al. 2021). Di Mascio et al. (1991) and Ribaya-Mercado et al. (1995) reported that lycopene exhibits the 30 quenching capacity of singlet oxygen in vitro with quenching constant being 2 and 10 times higher than that of β – 31 carotene and α – tocopherol. Lycopene-rich foods are considered to relate to a lower risk of some degeneration 32 diseases (Liang et al. 2019). Lycopene plays roles as a chemopreventive agent of digestive-tract cancers, lung cancer 33 and prostate cancer (Anlar and Bacanli 2020; Bano et al. 2020; Mirahmadi et al. 2020; Qi et al. 2021). However, 34 thermal processes cause the isomerization, consequently, reduction of biological activity of lycopene (Gupta et al. 35 2010; Murakami et al. 2018; Saini et al. 2019). Watermelon juice is also a source of citrulline with content being 1.1 36 -4.7 g/kg in flesh (Tarazona-Díaz et al. 2011). Citrulline is precursor for arginine for human (Bahadoran et al. 2020). 37 In addition, watermelon juice contains many nutritious constituents, such as: vitamins (A, B1, B2, B6, C, E) and 38 minerals (Maoto et al. 2019). In general, watermelon juice is an extremely valuable source of nutrition for human 39 health.

40 Recently, watermelon juice has been consumed in form of fresh juice, concentrate and instant powder. In concentrate and instant powder production juice, the concentration is one of the most important steps because it 41 42 determines quality and energy consumption. Recently, concentration of juice has been conducted by vacuum 43 evaporation. However, thermally sensitive constituents are destroyed by influence of high temperature. With lycopene, 44 thermal processes cause the isomerization and oxidation, consequently, reduction of biological activity (Colle et al. 45 2013; Murakami et al. 2018; Saini et al. 2019). In addition, sensory properties of juice, especially flavor, is significant 46 change under thermal processing (Pendyala et al. 2020). Consequently, it leads to decrease in quality of final products. 47 Reverse osmosis (RO) is membrane separation process with pressure driving force. The mechanisms of separation are based on sieving and diffusion effects (Wenten and Khoiruddin 2016). This process can be conducted 48 49 at ambient temperature, thus remains the thermally sensitive compounds (W. Barker 2011). Besides, RO process 50 consumes lower energy than the evaporation (Cassano et al. 2021). In addition, the RO system is also low capital cost, 51 easy to install, operate and maintain (Anis et al. 2019). Because of these advantages, RO process has been used to 52 concentrated a variety of juices, e.g. apple (Ahmad et al. 2020), orange (Destani et al. 2020), pomegranate (Bagci et 53 al. 2020), etc. Dos Santos Gomes et al., 2011 evaluated the concentration of watermelon juice by polyamide composite

RO membrane. The results showed that the process concentrated juice from 6.5 to 24 °Brix and increased lycopene content 3.1 times. However, this study did not clearly show the impact of operating condition and kinetic in process of concentrating watermelon by RO membrane. Meanwhile, these information help to easily optimize the operation process.

The objective of this research was to investigate the application of RO for concentration of watermelon juice. Influence of operating pressure on performance of process was studied. The recovery yield of total sugar (TS), lycopene, and antioxidant capacity in concentration by RO membrane was also determined. The determination of predominant fouling mechanisms and the mathematical models were also investigated to describe the watermelon juice concentration by RO membrane. We aim to assess the feasibility of RO process for concentration of watermelon ijuice.

64

2. Materials and methods

65 **2.1. Materials**

Watermelon juice: Watermelon (*Citrullus lanatus*) fruits were purchased from a local market in Ho Chi Minh
City (Vietnam). Their weight was 3 – 4 kg per fruit, with total red flesh. The fruits were clean by water. Then, it was
peeled, followed by recovering flesh. The flesh was crushed by a steel sieve with 1 mm of mesh to obtain the juice.
The juice was stored at 4 °C and using in 24 hours.

70 **2.3. Membrane**

Reverse osmosis membrane was HR98PP, a composite membrane made from polypropylene, manufactured by
Alfa Laval (Denmark). NaCl rejection by this membrane is higher than 96% at 2,000 ppm of NaCl, 15.5 bar of
operating pressure and 25 °C of operation temperature (information supplied by manufacturer). Prior to use, membrane
was cleaned as following procedure: Flush with clean water with approximately 5 times of the system hold up volume;
then, flush by full recirculation of NaOH solution (pH10) in 30 min without applied pressure and 15 L/min of feed
flowrate; finally, flush with clean water at 5 bar of applied pressure until pH of permeate and retentate reached 6.0 –
7.0.

78 **2.4. Membrane apparatus**

The Labstak M20, a plate and frame system manufactured by Alfa Laval (Denmark) was used to carry out the concentration of watermelon juice (**Fig. 1**). The unit consisted of 4 couples of membrane sheets with 0.144 m² of active area (0.018 m²/sheet). The pressure was supplied by Hydra – Cell pump, a piston pump manufactured by 82 Wanner Engineering Inc. (USA). The operating temperature was at ambient. The system was operated as concentration

83 mode with full recirculation of retentate (Fig. 1). Feed flow rate was 15 L/min. In this research, the operation pressure

was investigated at 30 and 40 bar to evaluate the effects of operating pressure on the RO filtration. All experiments of 84

85 concentration were in duplicate with difference in permeate flux being lower than 5%.

86 Concentration factor (CF) was expressed as the ratio of initial volume of feed (V_F , L) to the volume of the 87 retentate (V_R , L):

$$CF = \frac{V_F}{V_P} \tag{1}$$

The recovery yield of component i in retentate side (Y) was expressed as the following formula: 88

$$Y_i = \frac{C_{R,i} V_R}{C_{F,i} V_F} \tag{2}$$

Where, $C_{R,i}$ and $C_{F,i}$ were contents (g/L) of component *i* in retentate and feed, respectively. 89

90 After each run of RO, crossflow membrane system was cleaned as the following procedure:

91 - Flush with clean water with approximately 5 times of the system hold up volume.

92 - Recirculate clean water in 30 min at 5 bar of applied pressure and 15 L/min of feed flowrate.

93 - Recirculate NaOH solution (pH10) in 30 min without applied pressure and 15 L/min of feed flowrate.

94 - Flush with clean water at 5 bar of applied pressure until pH of permeate and retentate was 6.0 - 7.0.

95 2.5. Fouling mechanisms analysis

96 In this work, the fouling mechanism was determined using the models described by Hermia (Equation (3) and 97 **Table 1**). The equation was determined by nonlinear regression, the coefficient of determination (\mathbb{R}^2) and the root 98 mean square deviation (RMSE) was used to find out the mechanism for each assay evaluated.

$$\frac{d^2t}{dV^2} = k_n \left(\frac{dt}{dV}\right)^n \tag{3}$$

99 Where, n: blocking index, t: filtration time (h), V: accumulated permeate volume (L), k_n : resistance coefficient.

Besides that, fouling and cleaning effectiveness in NF were evaluated by fouling and recovery indices and 100 101 expressed as the following equations:

102 Fouling index:

$$FI = 100 \frac{PWP_{final}}{PWP_{initial}}$$
(4)

103 Where, $PWP_{initial}$ (L.m⁻²h⁻¹) and PWP_{final} (L.m⁻²h⁻¹) were the initial pure water permeability of membrane and 104 the pure water permeability when NF was completed.

105 *Recovery index:*

$$RI = 100 \frac{PWP_{cleaning}}{PWP_{initial}}$$
(5)

106 Where, $PWP_{initial}$ (L.m⁻²h⁻¹) and $PWP_{cleaning}$ (L.m⁻²h⁻¹) were the initial pure water permeability of membrane 107 and the pure water permeability when the cleaning was completed.

108 2.7. Mathematical model

In order to model the permeate flux in concentration of watermelon juice by RO process, the following equationwas applied:

$$J_v = k \cdot \ln(\frac{\alpha}{CF}) \tag{6}$$

111 Where, J_{ν} : permeate flux (L.m⁻².h⁻¹), *k*: representative of mass transfer in boundary layer on membrane surface 112 (L.m⁻².h⁻¹) and α : considered as maximum of CF value.

113 This semi – empirical model derived from osmotic pressure model (Nabetani et al. 1995). In osmotic pressure 114 model, permeate flux is limited by difference in osmotic pressure between two sides of membrane. When CF reaches 115 α value, the difference in osmotic pressure between two sides of membrane equals to transmembrane pressure, 116 consequently, permeate flux is zero. This model was successfully applied for modeling permeate flux in concentration 117 chicken extract (Nabetani et al. 2012) and nanofiltration for concentration of fish sauce (Lai and Nguyen 2021), coffee 118 extract (Pan et al. 2013).

119 The variation of retentate volume (V_R , L) during the concentration by membrane filtration:

$$\frac{dV_R}{dt} = -AJ_v \tag{7}$$

120 Where, A: the membrane area (m^2) , J_{ν} : permeate flux $(L/m^2.h)$

According to the mass balance, the following equation describes the relationship between individual solute variation in retentate flows:

$$\frac{d(V_R C_{R,i})}{dt} = -AJ_\nu C_{R,i}(1-R_i)$$
⁽⁸⁾

Where: R_i : solute retention (-); R_i , A, V_R and $C_{Ro,i}$ were determined from experimental data. The set of equations (equation (7) and equation (8)) were solve by the fourth order Runge – Kutta method in Matlab software (version R2018a). This mathematical model has been successfully applied in modeling in concentration waste fresh tea leaf
extract by RO (Lai et al. 2021), as well as purification and concentration rice protein by ultrafiltration (Doan and Lai
2021).

128 **2.6. Analytical methods**

Total sugar (TS) content of watermelon juice was determined by using sulfuric acid to form furan compounds,
then adding phenol as indicator. The absorbance at 490 nm of wavelength was measured (Nielsen 2010).

Lycopene: Total lycopene were analyzed by spectrometric method (Fish et al. 2002). 10 mL of n-hexane was delivered into a tube. Then, adding 5 mL of ethanol 95% (v/v) and 5 mL of BHT 0.05% (w/v) in acetone. The mixture was vortexed. A given weight (approximately 0.5 gram) of watermelon juice was added into the mixture and shaking at 180 rpm in 15 min. Then, 3 mL of distilled water was added and shaking at 180 rpm in 5 min. After shaking, the tube was remained in 5 min for phase separation. Three mL of upper layer (n-hexane) was taken to determine the absorbance at 403 nm of wavelength. All steps were conducted at 20 °C. Content of lycopene was calculated by the following equation:

$$Lycopene (mg/kg) = \frac{A_{503}x31.2}{m_{juice}(g)}$$
(9)

Antioxidant capacity of watermelon juice was determined as the scavenging capacity of 1,1-diphenyl-2picrylhydrazyl (DPPH) (Oms-Oliu et al. 2009). Watermelon juice was centrifuged at 6000g for 15 min at 4 °C. Then, 0.01 mL of the supernatant was added to 3.9 mL of 0.025 g/L of DPPH in methanol. Then, adding 0.090 mL of distilled water. The mixture was vortexed and kept in 30 mins in darkness. Then, the absorbance was determined at 515 nm of wavelength. Blank was methanol. AC was estimated as mg/L of Trolox equivalent antioxidant capacity (TEAC).

143

3. Results and discussion

144 **3.1. Permeate flux**

Permeate flux in concentration of watermelon juice by HR98PP was showed in **Fig. 2**. Permeate flux declined with increase in CF. Due to higher driving force, at same CF value, permeate flux at 40 bar was higher than that at 30 bar of operating pressure. At initial point, permeate flux at 40 bar was 19.74 L.m⁻².h⁻²; whereas, that at 30 bar was 13.54 L.m⁻².h⁻². The curves of permeate flux at 30 and 40 bar of operating pressure was nearly offset. It means that the difference in permeate flux decline between at 30 and 40 bar of operating pressure was insignificant during the concentration. It implies that, based on resistance series model, influence of operating pressure on fouling in concentration by HR98PP was also insignificant (W. Barker 2011). It also implies that, increase in total solid in retentate insignificantly influenced on concentration polarization due to the high turbulence of fluid on membrane surface. Thus, the decline of permeate flux with increase in CF can be explained by increase in osmotic pressure, which caused by increase in total solid in retentate.

Based on equation (6), the linear regression was applied for modeling of permeate flux. Result in **Fig. 2** and **Fig. 3** and the approximate 1.0 of R^2 indicates that model exhibited the good agreement with experimental data. The estimation of value in concentration of watermelon juice by HR98PP was stated in **Table 2**. The mass transfer (*k*) value at 30 bar was lower than that at 40 bar. Wijmans et al. (1984) proved that increase in operating pressure leads to increase in mass transfer in boundary layer on membrane surface. And result in **Table 2** also indicates that higher operating pressure lead to higher maximum value of CF due to higher transmembrane pressure.

161 The statistical analysis results are summarized in **Table 3**. The resistance coefficient (k_n) , coefficient of 162 determination (R^2) and the root mean square deviation (RMSE) were used to compare the numerical predictions and the experimental data, and to choose the best fit mechanism for fouling and permeate decline over time evaluation. 163 164 The results showed that blocking models were better correlated with experimental results than cake filtration models. 165 This was completely consistent with the explanation for flux variation as a function of concentration factor. At both survey pressure conditions, the complete blocking model had the highest R² value, and best fit to the experimental 166 167 data. On the other hand, the standard blocking mechanism had the highest resistance coefficient indicating that it was the most important fouling mechanism. For standard blocking the R² was about 0.986 and 0.946, respectively, at 30 168 169 and 40 bar that also showed the good agreement between the model and the experimental data. This is explained that 170 the watermelon juice is a complex solution containing solutes of varying sizes, the multiple pore blockings 171 mechanisms can occur simultaneously. This phenomenon reported in concentration of strawberry juice by Arend, 172 Rezzadori, et al. (2019). However, due to the molecular weight of components in the watermelon juice, as well as the 173 high operating pressure (30 - 40 bar), the foulants were less likely to enter the pore and reduce diameter. Therefore, 174 the standard pore blocking did not occur significantly. This was also observed in several other studies (Lamdande et 175 al. 2020; Lin 2017).

The resistance coefficient of the cake filtration mechanism was extremely low, indicating that there was no boundary layer formation in the concentration of watermelon juice. This reaffirmed that, as total solid concentration increased, concentration polarization had no significant effect on permeate flux. This suggested that the complete 179 blocking mechanism is dominant during concentration of watermelon juice. Both complete pore blocking and cake 180 filtration mechanism occur when the sizes of most solute molecules in watermelon juice were greater than the 181 membrane pore size. As results, particles are unable to enter into the pore and permeate through the membrane (Khan 182 et al. 2020). However, cake layer occurs in the presence of factors such as high concentration and binding of molecules, 183 binding to the membrane and they can deposit on the membrane surface. It has a great influence on the permeate flux. 184 While, in the case of the complete blocking mechanisms, a molecule never settles on another molecule that has 185 previously deposited on the membrane surface (Vela et al. 2008). It is easily eliminated by strong impacts such as high pressure or turbulent flow, and it has little effect on permeate flux. These prove that HR98PP membranes were 186 187 suitable for watermelon juice concentration. Furthermore, the resistance coefficient was greater at 40 bar of operating 188 pressure than at 30 bar. This suggested that the increased pressure was responsible for the increase in turbulent flow, 189 thereby reducing fouling phenomena. The complete blocking mechanism has also observed in many concentration 190 processes by RO, such as waste fresh tea leaf extract (Lai et al. 2021), skim milk (Arend, Castoldi, et al. 2019).

191

3.2. Recovery yields and contents of components in retentate

192 Content in retentate and recovery yield of TS in concentration of watermelon juice by HR98PP is showed in Fig. 193 4. Relationship between TS content and CF was linear. It means that, rejection of TS was insignificant change in 194 concentration of the juice. However, slope of TS content versus CF at 30 bar was slightly lower than at 40 bar. It 195 implies that rejection of TS at 30 bar was lower than that at 40 bar. Tsuru et al. (1991) proved that, increase in operating 196 pressure leads to increase in rejection in RO process. Comparing to at 40 bar, at 30 bar, the lower rejection of TS led 197 to the lower recovery yield. At 40 bar, recovery yield was 98.6% when reaching 2.25 of CF; whereas, at 30 bar, the 198 one was 90.7% when reaching 1.78 of CF. Ratio of C_R/C_F to CF at 40 bar was approximately 1.0. It means that, 199 rejection of TS at 40 bar was approximately 100%. Whereas, that ratio at 30 bar was higher than 0.92, implies that 200 rejection of TS at 30 bar was higher than 92%. The high rejection of TS also led to low concentration of TS in permeate 201 (Fig. 5). Result in Fig. 5 indicates that, content of TS in permeate was very lower than one in retentate. At 40 bar of 202 operating pressure and 2.25 of CF, TS content in permeate was 2.41 g/L. The one at 30 bar of operating pressure and 203 1.79 of CF was 1.93 g/L. At same CF value, TS content in permeate at 30 bar of operating pressure was higher than 204 that at 40 bar because that, rejection of TS at 30 bar was lower than that at 40 bar.

Estimation based on equation (6) indicates that, maximum of CF in retentate could reach 2.6 folds higher than that in feed. Dos Santos Gomes et al. (2011) reported that, when concentrating watermelon juice by RO membrane at 60 bar, CF and TS content can reach 4.4 and 30 °Brix, 3.2 folds in relation to feed. The CF and TS content in that work was higher than ones in our work. Nevertheless, the process carried out by Dos Santos Gomes et al. (2011) was conducted at higher operating pressure and recovery of TS was lower, in comparing with present work.

210 Content and recovery yield of lycopene in concentration of watermelon juice by HR98PP is showed in Fig. 6. 211 Rejection of lycopene in RO process insignificantly changed due to linear relationship between lycopene content and 212 CF. Result also indicates that, lycopene in permeate was not detected. At 2.25 of CF and 40 bar of operating pressure, recovery yield of lycopene was 90%. Whereas, at 1.79 of CF and 30 of operating pressure, the one was 86.8%. It 213 214 means that, there was the loss of lycopene by oxidation resulted in a reduction in recovery yield along with increase 215 in CF. Comparing at 40 bar, the recovery yield of lycopene at 30 bar was lower due to operating time at 30 bar was 216 longer. The loss of lycopene by oxidation and isomerization reactions was also observed in report by Dos Santos 217 Gomes et al. (2011). However, the loss of lycopene in membrane process was not much as comparing to concentration 218 by evaporation. The concentration of watermelon juice from 8 °Brix to 30 °Brix at 50 °C and 100 mg Hg of pressure 219 in rotary vacuum evaporator, 33.7% of lycopene was degraded (Jaju et al. 2017).

220 The processing properties of watermelon juice in concentration at 30 and 40 bar based on proposed mathematical 221 model are presented in Fig. 7. The theoretical results are given from mathematical models that were well consistent 222 with the experimental data. The content of lycopene and TS at 40 bar was higher than that at 30 bar due to the faster 223 concentration process caused by the higher permeate flux. It implies that, at 40 bar, the concentration of watermelon 224 juice was more effective compared to at 30 bar. In mathematical model, rejection was assumed to be constant and 225 estimated as mean of experimental rejection, and permeate flux varied linearly with CF, but experimental rejection 226 and permeate flux slightly changed. As a result, there is a discrepancy between calculating data and measured data. 227 However, they fluctuated slightly and asymptotically around the predicted value. The reasons for this were the fouling 228 phenomena that cough occur after a time of operation and increase in the viscosity of the retentate. Thus, the predictive 229 model can be used to predict the time operation for a target solutes concentration at the end of the concentration. From 230 the mathematical model, the maximum theoretical value of lycopene content at 30 and 40 bar were 172 and 185 mg/L, 231 respectively, increased by 4.5 and 3.5 folds, with operating time of 36.8 and 27.1 hours. Along with that, the maximum 232 of TS content was 245 and 341 g/L, respectively. It implies that, the concentration of watermelon juice was more 233 effective at 40 bar, due to shorter time and higher of molecules content. Compared with other studies on applying RO 234 for concentration, the maximum theoretical value of TS content obtained at 40 bar in watermelon juice concentration

235 by HR98PP membrane is significantly higher than soluble solids content obtained in concentration by RO membrane 236 of orange juice (30 °Brix at 60 bar, (Jesus et al. 2007), apple juice (28.1 °Brix at 60 bar, (Aguiar et al. 2012)), 237 chokeberry juice (24.9 °Brix at 55 bar, (Pozderović et al. 2016)), pomegranate juice (18 °Brix at 30 bar, (Bagci et al. 238 2019)). The maximum theoretical value of lycopene and TS content in watermelon juice obtained by concentration 239 with HR98PP membrane suggests that, the concentrate exhibited a good recovery particles which could be used in 240 another process without concentrated process more. However, more research is necessary to identify the optimal multi-241 stage RO system for a watermelon juice product with high concentration of bioactive compounds and soluble solids. 242 Summary of concentration of watermelon juice by HR98PP was stated in Table 4. Result in Table 4 indicated 243 that the antioxidant capacity of watermelon juice was lost 29.89 and 17.61% when concentration by RO at 30 bar, 244 1.79 of CF and 40 bar, 2.25 of CF, respectively. Result also indicated that, the fouling in RO process with HR98PP at 30 bar of operating pressure was more severe than that at 40 bar. FI values of RO process at 30 bar and 40 bar were 245 246 67.11 and 76.64%. However, the cleaning as proposed procedure was eliminated fouling. RI values of RO process 247 with HR98PP at 30 and 40 bar of operating pressure were higher than 95%. It means that, permeate flux was fully 248 recovered.

4. Conclusions

250 Results indicated that the concentration of watermelon juice by HR98PP at 40 bar of operating pressure was 251 more effective than that at 30 bar. The semi-empirical model has shown good performance when used to simulate 252 permeate flux. The complete blocking mechanism was observed to be the reason causing fouling phenomena and the 253 decline in permeate flux in concentration of watermelon juice by HR98PP. Theoretical maximum value of CF was 254 2.64. Recovery yields of TS, lycopene and antioxidant at 40 bar of operating pressure and 2.25 of CF were 98.67, 255 90.03 and 82.39, respectively. Result showed that, at 40 bar, the maximum contents of lycopene and TS increased up to 4.5 folds. TS and lycopene contents at that conditions of RO process were 171.31 g/L and 83.90 mg/L, respectively. 256 257 From mathematical model, the theoretical permeate flux, content of compounds in retentate over time were observed, 258 and they were in good agreement with the experimental data. The fouling in RO process were severe; however, 259 cleaning with NaOH solution at pH10 was fully eliminated fouling. It is feasible to utilize HR98PP membrane for 260 concentration of watermelon juice.

261 ACKNOWLEDGEMENT

- 262 We acknowledge the support of time and facilities from Ho Chi Minh City University of Technology (HCMUT),
- 263 VNU–HCM for this study.
- 264 CONFLICT OF INTEREST
- 265 None
- 266 ETHICAL APPROVAL
- 267 Ethics approval was not required for this research.

268 DATA AVAILABILITY STATEMENT

269 Data available on request from the authors.

270 AUTHOR'S CONTRIBUTION STATEMENT

- 271 Q.D.L and H.D.N designed and directed the project. Q.D.L and N.T.T.D. wrote the main manuscript text.
- 272 N.T.T.D. processed the experimental data, performed the analysis. All authors reviewed the manuscript.

273 **REFERENCES**

- Aguiar, I. B., Miranda, N. G. M., Gomes, F. S., Santos, M. C. S., de GC Freitas, D., Tonon, R. V, & Cabral, L. M. C.
- (2012). Physicochemical and sensory properties of apple juice concentrated by reverse osmosis and osmotic
 evaporation. *Innovative Food Science & Emerging Technologies*, *16*, 137–142.
- 277 Ahmad, S., Marson, G. V., Zeb, W., Rehman, W. U., Younas, M., Farrukh, S., & Rezakazemi, M. (2020). Mass
- transfer modelling of hollow fiber membrane contactor for apple juice concentration using osmotic membrane
 distillation. *Separation and Purification Technology*, 117209.
- Anis, S. F., Hashaikeh, R., & Hilal, N. (2019). Reverse osmosis pretreatment technologies and future trends: A
 comprehensive review. *Desalination*, 452, 159–195.
- Anlar, H. G., & Bacanli, M. (2020). Lycopene as an antioxidant in human health and diseases. In *Pathology* (pp. 247–
 254). Elsevier.
- Arend, G. D., Castoldi, S. M., Rezzadori, K., Soares, L. S., & Brião, V. B. (2019). Concentration of skim milk by
- reverse osmosis: characterization and flow decline modelling. *Brazilian Journal of Food Technology*, 22.
- Arend, G. D., Rezzadori, K., Soares, L. S., & Petrus, J. C. C. (2019). Performance of nanofiltration process during
 concentration of strawberry juice. *Journal of food science and technology*, *56*(4), 2312–2319.
- 288 Bagci, P. O., Akbas, M., Gulec, H. A., & Bagci, U. (2019). Coupling reverse osmosis and osmotic distillation for

- 289 clarified pomegranate juice concentration: Use of plasma modified reverse osmosis membranes for improved
- 290 performance. Innovative Food Science & Emerging Technologies, 52, 213–220.
- Bagci, P. O., Kahvecioglu, H., Gulec, H. A., & Bagci, U. (2020). Pomegranate juice concentration through the
 consecutive application of a plasma modified reverse osmosis membrane and a membrane contactor. *Food and Bioproducts Processing*, *124*, 233–243.
- Bahadoran, Z., Mirmiran, P., Kashfi, K., & Ghasemi, A. (2020). Endogenous flux of nitric oxide: Citrulline is preferred
 to Arginine. *Acta Physiologica*, e13572.
- Bano, S., Ahmed, F., Khan, F., Chaudhary, S. C., & Samim, M. (2020). Targeted delivery of thermoresponsive
 polymeric nanoparticle-encapsulated lycopene: in vitro anticancer activity and chemopreventive effect on
 murine skin inflammation and tumorigenesis. *RSC Advances*, *10*(28), 16637–16649.
- Cassano, A., Castro-Muñoz, R., Conidi, C., & Drioli, E. (2021). Recent Developments in Membrane Technologies
 for Concentration of Liquid Foods and Food Ingredients. In K. Knoerzer & K. B. T.-I. F. P. T.
- 301 Muthukumarappan (Eds.), *Innovative Food Processing Technologies: A comprehensive Review* (pp. 100–121).
- 302 Oxford: Elsevier. https://doi.org/https://doi.org/10.1016/B978-0-08-100596-5.23036-9
- Colle, I. J. P., Lemmens, L., Van Buggenhout, S., Van Loey, A. M., & Hendrickx, M. E. (2013). Modeling lycopene
 degradation and isomerization in the presence of lipids. *Food and Bioprocess Technology*, 6(4), 909–918.
- Destani, F., Naccarato, A., Tagarelli, A., & Cassano, A. (2020). Recovery of Aromatics from Orange Juice Evaporator
 Condensate Streams by Reverse Osmosis. *Membranes*, 10(5), 92.
- 307 Di Mascio, P., Murphy, M. E., & Sies, H. (1991). Antioxidant defense systems: the role of carotenoids, tocopherols,
 308 and thiols. *The American Journal of Clinical Nutrition*, 53(1), 194S-200S.
 309 https://doi.org/10.1093/ajcn/53.1.194S
- Doan, N. T. T., & Lai, Q. D. (2021). Ultrafiltration for recovery of rice protein : Fouling analysis and technical
 assessment. *Innovative Food Science and Emerging Technologies*, 70, 102692.
 https://doi.org/10.1016/j.ifset.2021.102692
- 313 Dos Santos Gomes, F., Albuquerque da Costa, P., Domingues de Campos, M. B., Couri, S., & Cabral, L. M. C. (2011).
- Concentration of watermelon juice by reverse osmosis process. *Desalination and Water Treatment*, 27(1–3),
 120–122.
- 316 Fish, W. W., Perkins-Veazie, P., & Collins, J. K. (2002). A quantitative assay for lycopene that utilizes reduced

volumes of organic solvents. Journal of food composition and analysis, 15(3), 309-317.

- Gupta, R., Balasubramaniam, V. M., Schwartz, S. J., & Francis, D. M. (2010). Storage stability of lycopene in tomato
 juice subjected to combined pressure- heat treatments. *Journal of agricultural and food chemistry*, 58(14),
 8305–8313.
- Jaju, N. S., Patel, S. S., Birwal, P., Deshmukh, G., Sukhdev, B. S., & Nema, P. K. (2017). Effect of Vacuum
 Evaporation Concentration on Lycopene Content and Rheological Properties of Watermelon Juice. *Int. J. Pure*
- 323 *App. Biosci*, 5(3), 1018–1024.
- Jesus, D. F., Leite, M. F., Silva, L. F. M., Modesta, R. D., Matta, V. M., & Cabral, L. M. C. (2007). Orange (Citrus sinensis) juice concentration by reverse osmosis. *Journal of Food Engineering*, *81*(2), 287–291.
- Khan, I. A., Lee, Y.-S., & Kim, J.-O. (2020). A comparison of variations in blocking mechanisms of membrane fouling models for estimating flux during water treatment. *Chemosphere*, 259, 127328.
- Lai, Q. D., Doan, N. T. T., & Nguyen, H. D. (2021). Technical assessment of reverse osmosis for concentration of fresh tea leaf extract. *Journal of Food Process Engineering*, *44*(7), e13725.
- Lai, Q. D., & Nguyen, H. D. (2021). Enhancement of fish sauce quality by application of nanofiltration. *LWT*,
 151(January), 112181. https://doi.org/10.1016/j.lwt.2021.112181
- 332 Lamdande, A. G., Mittal, R., & K.S.M.S., R. (2020). Flux evaluation based on fouling mechanism in acoustic field-
- assisted ultrafiltration for cold sterilization of tender coconut water. *Innovative Food Science and Emerging Technologies*, *61*, 102312. https://doi.org/10.1016/j.ifset.2020.102312
- Liang, X., Ma, C., Yan, X., Liu, X., & Liu, F. (2019). Advances in research on bioactivity, metabolism, stability and
 delivery systems of lycopene. *Trends in Food Science & Technology*, *93*, 185–196.
- Lin, Y. L. (2017). Effects of organic, biological and colloidal fouling on the removal of pharmaceuticals and personal
 care products by nanofiltration and reverse osmosis membranes. *Journal of Membrane Science*, *542*, 342–351.
- 339 https://doi.org/10.1016/j.memsci.2017.08.023
- Maoto, M. M., Beswa, D., & Jideani, A. I. O. (2019). Watermelon as a potential fruit snack. *International Journal of food properties*, 22(1), 355–370.
- Mirahmadi, M., Azimi-Hashemi, S., Saburi, E., Kamali, H., Pishbin, M., & Hadizadeh, F. (2020). Potential inhibitory
 effect of lycopene on prostate cancer. *Biomedicine & Pharmacotherapy*, *129*, 110459.
- 344 Murakami, K., Honda, M., Takemura, R., Fukaya, T., Kanda, H., & Goto, M. (2018). Effect of thermal treatment and

- 345 light irradiation on the stability of lycopene with high Z-isomers content. *Food chemistry*, 250, 253–258.
- Nabetani, H., Abbott, T. P., & Kleiman, R. (1995). Optimal Separation of Jojoba Protein Using Membrane Processes.
 Industrial and Engineering Chemistry Research, *34*(5), 1779–1788. https://doi.org/10.1021/ie00044a029
- Nabetani, H., Hagiwara, S., Yanai, N., Shiotani, S., Baljinnyam, J., & Nakajima, M. (2012). Purification and
 concentration of antioxidative dipeptides obtained from chicken extract and their application as functional food.
- 350 *Journal of Food and Drug Analysis*, 20(SUPPL.1), 179–183. https://doi.org/10.38212/2224-6614.2145
- Neglo, D., Tettey, C. O., Essuman, E. K., Kortei, N. K., Boakye, A. A., Hunkpe, G., et al. (2021). Comparative
 antioxidant and antimicrobial activities of the peels, rind, pulp and seeds of watermelon (Citrullus lanatus) fruit.
 Scientific African, *11*, e00582.
- Nielsen, S. S. (2010). Phenol-sulfuric acid method for total carbohydrates. In *Food analysis laboratory manual* (pp.
 47–53). Springer.
- Oberoi, D. P. S., & Sogi, D. S. (2017). Utilization of watermelon pulp for lycopene extraction by response surface
 methodology. *Food chemistry*, 232, 316–321.
- Oms-Oliu, G., Odriozola-Serrano, I., Soliva-Fortuny, R., & Martín-Belloso, O. (2009). Effects of high-intensity pulsed
 electric field processing conditions on lycopene, vitamin C and antioxidant capacity of watermelon juice. *Food chemistry*, *115*(4), 1312–1319.
- Pan, B., Yan, P., Zhu, L., & Li, X. (2013). Concentration of coffee extract using nanofiltration membranes.
 Desalination, *317*, 127–131.
- Pendyala, B., Patras, A., Ravi, R., Gopisetty, V. V. S., & Sasges, M. (2020). Evaluation of UV-C irradiation treatments
 on microbial safety, ascorbic acid, and volatile aromatics content of watermelon beverage. *Food and Bioprocess Technology*, *13*(1), 101–111.
- Pozderović, A., Popović, K., Pichler, A., & Jakobek, L. (2016). Influence of processing parameters on permeate flow
 and retention of aroma and phenolic compounds in chokeberry juice concentrated by reverse osmosis. *CyTA- Journal of Food*, *14*(3), 382–390.
- Qi, W. J., Sheng, W. S., Peng, C., Xiaodong, M., & Yao, T. Z. (2021). Investigating into anti-cancer potential of
 lycopene: Molecular targets. *Biomedicine & Pharmacotherapy*, *138*, 111546.
- Ribaya-Mercado, J. D., Garmyn, M., Gilchrest, B. A., & Russell, R. M. (1995). Skin lycopene is destroyed
 preferentially over β-carotene during ultraviolet irradiation in humans. *The Journal of nutrition*, *125*(7), 1854–

373 1859.

- Saini, R. K., A. Bekhit, A. E.-D., Roohinejad, S., Rengasamy, K. R. R., & Keum, Y.-S. (2019). Chemical stability of
 lycopene in processed products: A review of the effects of processing methods and modern preservation
 strategies. *Journal of agricultural and food chemistry*, 68(3), 712–726.
- Tarazona-Díaz, M. P., Viegas, J., Moldao-Martins, M., & Aguayo, E. (2011). Bioactive compounds from flesh and
- by-product of fresh-cut watermelon cultivars. *Journal of the Science of Food and Agriculture*, 91(5), 805–812.
- Tsuru, T., Nakao, S., & Kimura, S. (1991). Calculation of ion rejection by extended Nernst-Planck equation with
 charged reverse osmosis membranes for single and mixed electrolyte solutions. *JOURNAL OF CHEMICAL*
- 381 ENGINEERING OF JAPAN, 24(4), 511–517. https://doi.org/10.1252/jcej.24.511
- Vela, M. C. V., Blanco, S. Á., García, J. L., & Rodríguez, E. B. (2008). Analysis of membrane pore blocking models
- applied to the ultrafiltration of PEG. Separation and Purification Technology, 62(3), 489–498.
 https://doi.org/10.1016/j.seppur.2008.02.028
- W. Barker, R. (2011). *Membrane technology and applications*. (R. W.Barker, Ed.)*Membrane Technologies and Applications*. England: John Wiley & Sons, Ltd. https://doi.org/10.1201/b11416
- Wenten, I. G., & Khoiruddin. (2016). Reverse osmosis applications: Prospect and challenges. *Desalination*, 391, 112–
 125. https://doi.org/10.1016/j.desal.2015.12.011
- Wijmans, J. G., Nakao, S., & Smolders, C. A. (1984). Flux limitation in ultrafiltration: Osmotic pressure model and
 gel layer model. *Journal of Membrane Science*, 20(2), 115–124. https://doi.org/10.1016/S0376-7388(00)81327-

391

7

392

394	FIGURE LEGENDS
395	Fig. 1. Schema of flat and frame pilot LabstakM20 system. 1: feed tank, 2: pump, 3: pressure gauge, 4:
396	membranes, 5: permeate tank, 6: flowmeter
397	Fig. 2. Permeate flux against CF in concentration of watermelon juice by HR98PP (●: 40 bar, ▲: 30 bar; Solid
398	line: estimation, spot: experimental data)
399	Fig. 3. Permeate flux against Ln(CF) in concentration of watermelon juice by HR98PP (●: 40 bar, ▲: 30 bar;
400	Solid line: estimation, spot: experimental data).
401	Fig. 4. Content and recovery yield of TS in retentate in concentration of watermelon juice by HR98PP (•: 40
402	bar, ▲: 30 bar; filled spot: recovery yield, non-filled spot: content)
403	Fig. 5. Content of TS in permeate in concentration of watermelon juice by HR98PP (●: 40 bar, ▲: 30 bar)
404	Fig. 6. Content and recovery yield of lycopene in retentate in concentration of watermelon juice by HR98PP (•:
405	40 bar, ▲: 30 bar; filled spot: recovery yield, non-filled spot: content)
406	Fig. 7. Content of lycopene and TS in retentate vs. operating time in concentration of watermelon juice by
407	HR98PP. Plot: experiment (at 40 bar, \circ : lycopene, Δ : TS; and at 30 bar, \Box : lycopene, +: TS). Line: calculation (at 40
408	bar:: lycopene, $$: TS; at 30 bar,: TS)





Fig. 1. Schema of flat and frame pilot LabstakM20 system. 1: feed tank, 2: pump, 3: pressure gauge, 4:

membranes, 5: permeate tank, 6: flowmeter





Fig. 2. Permeate flux against CF in concentration of watermelon juice by HR98PP (●: 40 bar, ▲: 30 bar;

Solid line: estimation, spot: experimental data)



417

418 Fig. 3. Permeate flux against Ln(CF) in concentration of watermelon juice by HR98PP (●: 40 bar, ▲: 30
419 bar; Solid line: estimation, spot: experimental data).





421 Fig. 4. Content and recovery yield of TS in retentate in concentration of watermelon juice by HR98PP (●: 40
422 bar, ▲: 30 bar; filled spot: recovery yield, non-filled spot: content)



Fig. 5. Content of TS in permeate in concentration of watermelon juice by HR98PP (●: 40 bar, ▲: 30 bar)





426 Fig. 6. Content and recovery yield of lycopene in retentate in concentration of watermelon juice by HR98PP



(●: 40 bar, ▲: 30 bar; filled spot: recovery yield, non-filled spot: content)



429Fig. 7. Content of lycopene and TS in retentate vs. operating time in concentration of watermelon juice by430HR98PP. Plot: experiment (at 40 bar, \circ : lycopene, Δ : TS; and at 30 bar, \Box : lycopene, +: TS). Line: calculation (at43140 bar: ----: lycopene, ---: TS; at 30 bar, ----: lycopene, ---: TS)

432	TABLE LEGENDS
433	Table 1. Flux expressions of Hermia models
434	Table 2. Estimation of parameters of model of permeate flux in concentration of water melon juice
435	Table 3. Models fitting accuracy for concentration of watermelon juice by HR98PP
436	Table 4 . Summary of concentration of watermelon juice by HR98PP membrane.
437	

Fouling mechanism	n value	Derivative equation						
Complete pore blocking	2	$\ln(J^{-1}) = \ln(J_0^{-1)} + kt$						
Standard pore blocking	1.5	$J^{-0,5} = J_0^{-0,5} + kt$						
Intermediate pore blocking	1	$J^{-1} = J_0^{-1} + kt$						
Cake formation	0	$J^{-2} = J_0^{-2} + kt$						
J and J_0 (L/m ² h) are the permeate flux at t (h) of operating time and initial								

time, respectively.

Table 2. Estimation of parameters of model of permeate flux in concentration of water melon juice

Operating pressure (bar)	k (L.m ⁻² .h ⁻¹)	α	R ²	RMSE
30	17.302	2.13	0.992	0.2921
40	19.669	2.63	0.9898	0.5061

 Table 3. Models fitting accuracy for concentration of watermelon juice by HR98PP

Pressure		Cake filtration	Intermediate blocking	Standard blocking	Complete blocking
	k _n	0.00006	0.0023	0.2024	0.0385
30 bar	\mathbb{R}^2	0.8568	0.9557	0.986	0.9964
	RMSE	4.6947	1.4839	0.4437	0.1524
	kn	0.00003	0.0019	0.2740	0.0169
40 bar	R^2	0.7164	0.8806	0.9462	0.9856

 Table 4. Summary of concentration of watermelon juice by HR98PP membrane.

Operating	Fresh juice		Concentrate juice		Recovery yield							
pressure (bar)	TS ^a	Ly ^b	ACc	TSª	Ly ^b	ACc	TSª	Ly ^b	ACc	CF	FI	RI
30	70.12	48.46	70.81	114.53	76.54	88.86	90.32	86.90	70.11	1.79	67.11	95.72
40	76.41	41.05	67.83	171.31	83.90	125.74	98.67	90.03	82.39	2.25	76.64	96.12
a: g/L, b: mg/L, c: mg TEAC/L												