

Immunogenicity evaluation after BNT162b2 booster vaccination in healthcare workers

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Abstract

Waning of the immune response upon vaccination in SARS-CoV-2 infection is an important subject of evaluation in this pandemic, mostly in healthcare workers (HCW) that are constantly in contact with infected samples and patients. Therefore, our study aimed to establish the specific humoral response of specific IgG and IgA antibodies upon vaccination, during the second year of pandemic and evaluating the booster shot with the same vaccine type. A group of 103 HCW with documented exposure to the virus were monitored for specific IgG and IgA levels prior to vaccination, after the first vaccination round, during the following 8 months and after the booster shot with the same vaccine type. After 8 months post-vaccination the humoral response in both IgG and IgA decreased, 2.4 times for IgG, and 2.7 times for IgA. Although the antibodies levels significantly decreased, no documented infection was registered in the group. After the booster shot, the entire group, displayed IgG increased levels, immediately after booster followed by the increase in specific IgA. IgG levels post-second round of vaccination are statistically higher compared to the first round, while IgA is restored at the same levels. Within the vaccination or booster routine for a multiple waves' pandemic that is generating new virus variants, populational immunity remains an important issue for future implementation of prevention/ control measures.

1. Introduction

Entering the third year of COVID-19 pandemic and registering in Romania already over 2,6 million cases ¹ with over 62,000 deaths ² the vaccination route of the population reached just a 36% percentage of the total population. As COVID-19 will enter its endemic phase, prevention and control raise severe challenges. In the third year of COVID-19 we still have no specific treatment, therefore promoting vaccinations and developing herd immunity are the only effective and economic measures to control the current pandemic ³.

Extended studies that focus on the antibody levels triggered by infection and/or by vaccination have reported the existence of an entire panel of specific immunoglobulines ⁴. Moreover, recent studies show that cross immunity against coronaviruses can be elicited by vaccination ⁵ but still we have to focus on the relevance of the booster vaccination.

Within the total population, healthcare workers (HCW) represent the highly exposed populational segment. Therefore, monitoring HCW characteristics and response to vaccination represents a good overall example of vaccine efficacy. Moreover evaluating vaccine efficacy against SARS-CoV-2 variants is seminal to sustain proper information to the large population and to guide public health in this pandemic ⁶. A recent finding suggests that the mRNA vaccine booster, associates with a good protection against Omicron and Delta variants when comparing the effect to unvaccinated or to the two doses vaccination ⁷. In an Italian cohort comprising almost one year of follow-up and over 33 million tested subjects important issues emerged. When epidemic phase registered Delta variant circulation vaccine effectiveness decreased from 82% to 33% at 7 months after the second dose. Moreover the study showed that high risk individuals aged ≥ 80 years after 7 months seemed not to be protected after the second

dose of vaccine. Therefore, the authors sustain a booster vaccination even earlier than six months after the primary vaccination cycle⁸. The Israeli reports done on immunity waning and booster recommendation are numerous. Thus, in August 2021, in Israeli HCW, the surge of SARS-CoV-2 infections, mostly by Delta variant, appeared in 21.4% individuals that received only the two dose regimen while the rate in the HCW group that have received a booster was only 0.7%. Therefore, in this group, **a booster vaccination** indicates substantial protection by a third vaccine dose⁹ while previous studies in the same country have shown that at 3 months most HCWs still had measurable antibodies¹⁰. Nevertheless in the same country at 5 months, a third dose of the BNT162b2 mRNA vaccine is effective in protecting subjects against severe COVID-19, compared with the subjects receiving only two doses¹¹. Additionally, half a year after first vaccination with the BNT162b2 vaccine second dose, the humoral response was found substantially decreased, more specifically in men, over 65 years of age or older, and among immunosuppressed subjects¹².

When examining total and neutralizing antibodies raised in HCW against SARS-CoV-2 Spike protein, from Washington-1 (WA-1), Beta, Delta and Omicron variants of concern it was shown that mRNA booster eliminates the immune escape phenomena observed with the Omicron variant after two-dose vaccination¹³. Another study has shown that although neutralizing antibodies raised by two-dose vaccination decreased 5 months after the second vaccination, specific T and B lymphocytes were still detectable, and upon 3rd dose induced a quick recall response. An interesting finding of the study showed that although HCWs with low antibodies response to two doses proved good specific immune memory, that was quickly recalled by the third dose¹⁴. In over 3,000 HCW subjects **from an Italian hospital, infection after vaccination occurred in 0.5% subjects** mostly asymptomatic with no predominance of a specific viral variant¹⁵. Somewhat similar results were obtained in a Turkish HCW cohort where 4.5% of vaccinated personnel were infected with SARS-CoV-2¹⁶ and the booster dose of CoronaVac was advised¹⁷.

Combination of vaccination has shown that combining mRNA-mRNA or vector-mRNA types induces high neutralization titers against SARS-CoV-2¹⁸. Another combination study done in Spanish HCW has reported results for the combination of one dose of ChAdOx1-S-nCoV-19 followed by a second dose of the Pfizer BNT162b2 vaccine as a booster. The heterologous vaccinated subjects proved a stronger neutralizing activity no matter of the SARS-CoV-2 variant. The enhanced neutralizing potential is due to the appearance of switched and activated memory B cells¹⁹. A study published almost concomitantly with the later one, has shown that T cell activation markers increase after vaccination. Plasma from previously infected subjects or 3 dose vaccinated subjects had a better neutralization capacity compared to the plasma harvested from non-infected individuals receiving two vaccine doses²⁰.

In CoronaVac vaccination it was shown that after 6 months post-vaccination almost all HCW subjects has proved a decreased antibody persistence²¹. AZD1222 (ChAdOx1) vaccination study has shown also an important decline in antibody levels in HCW, months after vaccination²². In a Cororean HCW BNT162b2 vaccinated cohort it was shown that after six months, serum antibody levels significantly declined²³. In Finland, mRNA vaccine displayed only 53% from the initial IgG level after 6 months, but antibody waning

was not observed against COVID-19 hospitalization²⁴. In a HCW Polish cohort it was reported that there are higher levels of specific antibodies 6 months after vaccination in subjects experiencing the disease after the first round of vaccination, the finding supporting once more the use of a booster dose, especially for non-infected subjects²⁵.

In Indonesian HCW specific IgG persisted 3 months post-vaccination with an inactivated SARS-CoV-2 vaccine. The authors point out that there is an increased decline of the specific antibodies in subjects without prior SARS-CoV-2 infection, finding that sustains the need for an additional booster dose²⁶.

IgA is an antibody that sustains the humoral mucosal immunity especially in viral respiratory infections, and that there are few studies that evaluate the circulatory form of the antibody in COVID-19^{27,28}. We have previously shown that post-vaccination, specific serum IgA is triggered in similar levels with IgG and having the same antibody dynamics²⁹, while other studies have reported saliva IgA in low levels upon vaccination³⁰. At 6 months, post-vaccination specific IgA serum levels showed a significant descending trend³¹. In a Dutch cohort vaccination with several vaccine types (mRNA-1273, BNT162b2, Ad26.CoV2-S or ChAdOx1-S) was studied and the authors point out that specific T cell responses were detectable one year post-vaccination while the humoral responses retained up to four months³².

Immune response waning upon vaccination in COVID-19 is an important issue in the current pandemics, mostly in HCW. Therefore, our study aimed to establish the specific humoral response of antibodies IgG and IgA, upon specific vaccination, during the second year of pandemic and evaluating the booster shot with the same vaccine type and dose.

2. Materials And Methods

Subjects. A total of 103 subjects, HCW in contact with SARS-CoV-2-infected samples and patients, constituted the test group followed-up between May, 2020 to October, 2021. The characteristics of the enrolled subjects, such as age and sex are presented in Table 1 along with their associated co-morbidities.

The group of 103 subjects were vaccinated in January, 2021 and they were followed-up before and after vaccination for measurement of the levels of serum IgG and IgA, during the 8 months of surveillance, prior to the 3rd booster received in October 2021 and after 3 weeks post-booster. Monthly RT-PCR tests during the 8 months follow-up yielded negative results for all subjects.

Vaccination. All the subjects received the Pfizer-BioNTech vaccine according to the supplier instructions, namely they received their first vaccine shot on the January 6, 2021 and the second dose on January, 27 and the results of a sample of the tested group were prior published by us focusing on the humoral response triggered by the first vaccination protocol²⁹. Subjects were followed the entire 2021 year and in October 2021 they have received the booster shot with the same Pfizer-BioNTech vaccine.

Dynamics of sampling. All the subjects were tested for the presence of IgA and IgG-specific antibodies recognizing the S1 domain of the SARS-CoV-2 Spike protein. All subjects were tested 1 day prior to Pfizer-BioNTech vaccination, after 2 weeks post-second second dose, 1 day before the 3rd booster and three weeks post-booster or for some of the subjects testing was done weekly as presented hereafter.

Blood sampling. Peripheral blood samples from subjects were collected by venipuncture during the morning hours in blood clot activator tubes (Vacutest Kima). Blood collection was carried out at the Colentina Clinical Hospital. Serum samples, separated by centrifugation (1,500 x g, 10 min at room temperature) within 4 h of blood collection, were used for ELISA. Serum samples were stored at -80⁰C for concomitant testing.

ELISA. Anti-SARS-CoV-2 ELISA (IgG and IgA) was used to determine the serum levels of specific IgG and IgA (EUROIMMUN Medizinische Labordiagnostika AG). The used protocol was as per the manufacturer's instructions. Details of the standard ELISA test were prior presented by us ²⁹. Results were calculated as indicated, namely the Ratio between the *Extinction of the patient sample* and the *Extinction of the calibrator*. The manufacturer recommends the following cut-off values: Ratio <0.8; Borderline Ratio ≥ 0.8 to <1.1; Positive Ratio ≥1.1.

The results are presented as index, as recommended by the IgG / IgA kit supplier. Data are presented as the mean ± standard deviation (SD). Comparison between groups, data analysis was performed using One-way ANOVA or Mann-Whitney tests using GraphPad Prism 9.3 (GraphPad Software, Inc.). All the tests and methods were performed in accordance with the relevant guidelines and regulations.

3. Results

Demographics characteristics. The tested group consists of mainly females having a mean age of 40.26 years with various comorbidities as presented in Table 1.

Table 1. Characteristics of the tested subjects

Parameter	Infected subjects until January 2021 (%)	Non-infected subjects until January 2021 (%)	Infected subjects during January – October 2021 (%)
Subjects (n)			
Female (90)	23	77	0
Male (13)	29	71	0
Average age of total, years	37.81	41.00	40.26
Average age of women, years	39.14	41.48	40.95
Average age of men, years	28.50	36.40	34.14
Major comorbidities (%)	Overweight		23
	Non-obesity overweight		13
	Obesity		10
	Cardiovascular disease		9
	Arterial hypertension		6
	Diabetes		4
	Non-immune thyroidian disease		4
	Hypothyroidity		4
	Autoimmune thyroidity		4
	Allergies		3
	Chronic venpus insufficiency		3
Various other comorbidities		Under 2%	

As previously reported by us, the gender differences did not statistically influence the level of antibody response upon vaccination, therefore the presented results comprise the entire group regardless of the gender.

Dynamics of IgG and IgA antibodies. The group received the Pfizer-BioNTech vaccination scheme in January 2021. Regardless of the infection status prior to vaccination, the entire group presented a high IgG and IgA levels post first round of vaccination (Figure 1).

After 8 month post first round of vaccination, the group had statistically decreased values for both antibodies (Figure 1). More specifically, for IgG the mean concentration decreased in 8 month 2.4 times,

while for IgA decreased 2.7 times ($p < 0.001$). To be mentioned that in the 8 month time period (January-October 2021) no documented infection with SARS-CoV-2 was registered.

The entire group was subjected to booster vaccination in October 2021 and post second round of vaccination the immunoglobulins serum concentrations (Figure 2) show that IgG increases immediately after booster 2.7 times, while IgA increased after the booster 2.5 times ($p < 0.001$).

To evaluate the level of humoral induction after booster vaccination we have compared, yet again the entire group with the values obtained after the first vaccination (Figure 3). Results shown that the IgG response after booster vaccination is statistically higher compared to the one obtained by the first vaccination ($p < 0.001$). In contrast, the IgA response after booster is almost identical to the values obtained after the first vaccination.

Some of the enrolled subjects were tested in a more detailed dynamics to evaluate the time frame in which the humoral response appears after the booster vaccination. Thus in a case where after 8 months post-first vaccination there are no detectable circulating antibodies, the booster induced a rapid (after the first week post-booster) a high value for both IgG and IgA, the levels continued to raise after two, respectively three weeks after booster. The concentrations of serum IgG and IgA were continuing to increase one month after booster (Figure 4). Both registered values were in this case higher than the ones registered post first-vaccination proving a proper immunological memory. However, 6 months after booster (174 days), the subject developed a mildly symptomatic form of COVID-19 with symptomatology associated with Omicron variant infection (sore throat, rinorhea, cough and harsh voice for 2 days, no fever, no headache, no other symptoms; oxygen saturation 96-100) having family members tested negative by rapid antigen tests.

Respiratory infections prior to booster vaccination. Out of the entire study group, during the 8 months follow up after first vaccination **none** of the subjects have contracted the SARS-CoV-2 infection pre-3rd vaccination booster. The assertion refers to the lack of any symptomatology related to the respiratory infection and to the fact that the in routine check-ups using RT-PCR testing no positive results were documented in this time frame.

Adverse effects upon vaccination. The presence of any adverse effects for the 3rd booster were registered for each subject. The adverse effects registered were compared to the ones registered in the first scheme of vaccination and in the entire group the adverse effects reported by us for the same group after the first round of vaccination²⁹ were less intense and far more reduced in number. Similar to the first round of vaccination, booster induced milder injection site pain in over 75% of the subjects.

Specific antibodies level upon 3rd vaccination. In the presented study, as the group displayed a decrease in both IgG and IgA increment of specific antibodies, the booster shot re-established, and for IgG even increased the specific humoral response in **all** of subjects. An interesting finding was that after booster vaccination the newly achieved level of IgA was statistically identical to the one achieved after the first

scheme of vaccination, while specific IgG surpassed the prior achieved antibody levels, sustaining the existence of a robust cellular memory.

COVID-19 after booster. 9 subjects from our study group (8.73%) developed COVID-19 after booster. One of the subjects developed the infection with Delta variant four months after booster (116 days) displaying a mild form of disease. Other 8 subjects were infected with Omicron variant in the time frame post-booster 4.5 - 6 months (127-174 days, medium 147 days = 4.90 months) displaying very mild forms of disease (minor symptoms for 1-3 days).

4. Discussion

Testing the humoral response in COVID-19 and further in vaccination,³³ is important to correctly evaluate the immune response to the natural and/or artificial immunization³⁴. Within the tested methods, the ELISA-related methodologies remain the most reliable and sensitive ones to evaluate the appearance of specific antibodies³⁵.

Adverse reactions to booster vaccination are reported as mainly pain at the injection site,³⁶ similar findings with our study. The adverse reactions panel reported in HCW receiving the first round of vaccination with BNT162b2 vaccine were significantly more frequent among HCWs with prior infection compared to infection-naïve individuals, and probably this process was due to the pre-existing cellular immunity. For the second round of vaccination the total adverse reactions were milder³⁷ thus this finding can reduce the overall negative attitude towards vaccines and vaccination.

A study performed on HCW in Greece has shown that the immune response after BNT162b2 vaccination depends on sex and age³⁸. We did not find statistically differences between the antibody response in correlation with gender, age, or the registered co-morbidities, therefore additional studies can clarify these dependencies.

The strategy to follow a 3rd vaccination shows that priority should be given to high-risk groups, elderly and immunodeficiency patients. Numerous studies have shown that heterologous boosters inflict a higher immune responses in comparison to homologous vaccination³⁹. Therefore, the COV-ADAPT study has presented the results obtained in HCW receiving various vaccination protocols. Homologous ChAdOx1 nCoV-19, homologous BNT162b2 or heterologous ChAdOx1nCoV-19/BNT162b2 vaccinations protocols have induced different Spike protein-directed humoral and cellular immune responses⁴⁰. An Israeli study has shown that BNT162b2, homologous booster dose was associated with a lower rate infection rate⁴¹. Our results show even after the first round of vaccination a reduced infection rate in our group and a low infection rate in the post-booster time frame. Although the post-booster infection was documented in almost 9% of the subjects, their symptoms were mild and the recovery was quick with no sequelae.

In Thailand, HCW receiving a third dose of AZD1222 were proved to trigger higher levels of specific IgG and IgA in comparison to the subjects receiving just two-dose vaccines. Moreover higher neutralizing potency against the wild type and variants of concern were found in the group receiving the 3rd dose of vaccine⁴². We have obtained higher levels of IgG in the entire group after booster compared to the levels obtained after the first round of vaccination, while the IgA levels were statistically similar, our study confirming thus an earlier report⁴². Moreover, our results are in accordance to the study performed in Germany in HCW subjects. Thus, in the study it was shown that SARS-CoV-2-specific IgM and IgA decrease rapidly over time, whereas IgG decreases more slowly. Prior infected subjects induced after booster vaccination higher IgG levels and to a lesser degree IgA levels⁴³. The link between the total antibodies and their neutralizing capacity is a question that still needs answers. A recent study, 2022, has shown that neutralizing titers are significantly higher post-boost compared to the titers obtained post two-dose series, as high as 15-fold increase in the neutralization capacity against Omicron variant. The mRNA booster dose induces an increase in both quantity and quality of the generated antibodies compared to the two-dose regimen¹³.

Re-infection after natural or artificial immunization after the booster shows that around 9% of our group showed documented respiratory infection, results that are in accordance with prior studies⁴⁴ pointing out that genetically distinct new variants can avoid established immune memory.

Study limitations

Comprehension of immune memory against SARS-CoV-2 viruses and their variants is still unknown. In general, studies show a 4, 6, 8 months waning of specific antibodies. Although this antibody waning appears, tests on immune memory cells could perfectly complete the immune pattern of this respiratory infection. Our study performed on 103 HCW subjects may be considered as small, but the subjects were and still are thoroughly documented during these 2 years of pandemics. There are similar studies performed on small well documented groups. A similar study performed on 90 HCW subjects has shown that the median IgGs titers are decreasing monthly in both previously infected individuals and naive subjects. Seven months after vaccination, it was shown a dramatically decrease of the humoral response in all subjects⁴⁵. Another study performed on 63 HCWs in Spain has shown that two months post-vaccination, antibody levels were decreased in naïve HCWs in comparison to previously infected HCWs. The authors report that ten months post-infection, the immune system has an immunological memory capable of producing a rapid and powerful secondary antibody response⁴⁶. In several cases that were weekly investigated post-boost we have shown that after vaccination, IgG level quickly increases, followed by a weekly increase of the IgA levels; this dynamic proving the clear existence of an immunological memory established by the first round of vaccination. The lack of correlation between the antibody response and the gender of the subjects can be explained by the fact that our group consisted of mainly females. We can not rule out that a more sex ratio group could have provided some correlations regarding gender differences in the post-vaccination humoral response.

5. Conclusion

The immune system has complex regulatory mechanisms in which the generation of immune memory is seminal in both natural infection and in the vaccination flow, but we still have to gather knowledge regarding antibody and cellular persistence.

Our results support the vaccination campaigns in healthcare workers receiving a booster dose of vaccine eight months after the primary vaccination cycle. The administration of a third dose of mRNA vaccine as a booster addresses the potential waning of immunity over time and by-passes the inefficacy against future viral variants. Though, more information and clinical studies are required to verify the safety of heterologous vaccination strategies and the evaluation of the necessity of a third dose of the vaccine. Although, our data show that there is a diminishing of the immune protection after 5 months after booster, the findings are opening the discussions for the need of an additional dose.

Within the vaccination or booster routine for a pandemic that is still on-going with its multiple waves and new variants, populational immunity remains an important issue for future implementation of prevention measures and control of this viral infection.

Declarations

CRedit authorship contribution statement

SZ: Conceptualization, Investigation, Formal analysis, Writing - original draft; CV: Investigation, Formal analysis, Writing - original draft. OD: Methodology, Writing - review & editing, Supervision; CC: Conceptualization, Investigation, Formal analysis, Writing - original draft, supervision; MN: Conceptualization, Investigation, Methodology, Formal analysis, Writing - original draft, supervision; All authors reviewed the manuscript.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Data Availability Statement: The dataset presented in this study is available from the corresponding authors upon reasonable request.

References

1. Romania COVID - Coronavirus Statistics - Worldometer (worldometers.info)
2. Popp, C. *et al.* Pandemics of our times: AH1N1 influenza versus COVID-19-features of fatal cases in Romania. *Virchows Archiv.* 479(1): S1-S2; (2021).
3. Neagu, M. The bumpy road to achieve herd immunity in COVID-19. *J Immunoassay Immunochem*; **41**(6): 928–945. DOI: 10.1080/15321819.2020.1833919 (2020).
4. Batra, M. *et al.* Role of IgG against N-protein of SARS-CoV2 in COVID19 clinical outcomes. *Sci Rep***11**, 3455 (2021). <https://doi.org/10.1038/s41598-021-83108-0>.
5. Ruggiero, A. *et al.* SARS-CoV-2 vaccination elicits unconventional IgM specific responses in naïve and previously COVID-19-infected individuals. *eBioMedicine***77**: 103888. <https://doi.org/10.1016/j.ebiom.2022.103888> (2022).
6. Neagu, M. *et al.* Back to basics in COVID-19: antigens and antibodies – Completing the puzzle. *J Cell Mol Med.* **25** (10): 4523-4533. <https://doi.org/10.1111/jcmm.16462> (2021).
7. Accorsi EK, Britton A, Fleming-Dutra KE, *et al.* Association Between 3 Doses of mRNA COVID-19 Vaccine and Symptomatic Infection Caused by the SARS-CoV-2 Omicron and Delta Variants *JAMA.* **327**(7):639-651. doi:10.1001/jama.2022.0470 (2022).
8. Fabiani, M. *et al.* Effectiveness of mRNA vaccines and waning of protection against SARS-CoV-2 infection and severe covid-19 during predominant circulation of the delta variant in Italy: retrospective cohort study. *BMJ***376**:e069052.doi: 10.1136/bmj-2021-069052 (2022).
9. Oster, Y. *et al.* The effect of a third BNT162b2 vaccine on breakthrough infections in healthcare workers: a cohort analysis. *Clin Microbiol Infect.* **S1198-743X**(22)00043-X.doi: 10.1016/j.cmi.2022.01.019 (2022).
10. Shachor-Meyouhas, Y. *et al.* Immunogenicity trends 1 and 3 months after second BNT162B2 vaccination among healthcare workers in Israel. *Clin Microbiol Infect.* **S1198-743X**(21)00660-1. doi: 10.1016/j.cmi.2021.11.014 (2021).
11. Barda, N. *et al.* Effectiveness of a third dose of the BNT162b2 mRNA COVID-19 vaccine for preventing severe outcomes in Israel: an observational study. *Lancet.* ;398(10316):2093-2100. doi: 10.1016/S0140-6736(21)02249-2 (2021).
12. Levin, E.G. *et al.* Waning Immune Humoral Response to BNT162b2 Covid-19 Vaccine over 6 Months. *N Engl J Med.* **385**(24):e84.doi: 10.1056/NEJMoa2114583 (2021).
13. Debes, A.K. *et al.* Comparison of total and neutralizing SARS-CoV-2 spike antibodies against omicron and other variants in paired samples after two or three doses of mRNA vaccine. *medRxiv.***2022.01.26.22269819**. doi: 10.1101/2022.01.26.22269819 (2022).
14. Liu, Y. *et al.* Robust induction of B cell and T cell responses by a third dose of inactivated SARS-CoV-2 vaccine. *Cell Discov.* **8**(1):10. doi: 10.1038/s41421-022-00373-7 (2022).
15. Lombardi, A. *et al.* Clinical characteristics of healthcare workers with SARS-CoV-2 infection after vaccination with BNT162b2 vaccine. *BMC Infect Dis.* **22**(1):97.doi: 10.1186/s12879-022-07083-1 (2022).

16. Çağlayan, D. *et al.* An analysis of antibody response following the second dose of CoronaVac and humoral response after booster dose with BNT162b2 or CoronaVac among healthcare workers in Turkey. *J Med Virol.* **94**(5):2212-2221. doi: 10.1002/jmv.27620 (2022).
17. Yigit, M. *et al.* Should a third booster dose be scheduled after two doses of CoronaVac? A single-center experience. *J Med Virol.* **94**(1):287-290. doi: 10.1002/jmv.27318 (2022).
18. Sølund, C. *et al.* Analysis of Neutralization Titers against SARS-CoV-2 in Health-Care Workers Vaccinated with Prime-Boost mRNA-mRNA or Vector-mRNA COVID-19 Vaccines. *Vaccines* (Basel). **10**(1):75. doi: 10.3390/vaccines10010075 (2022).
19. Pozzetto, B. *et al.* Immunogenicity and efficacy of heterologous ChAdOx1-BNT162b2 vaccination. *Nature.* **600**(7890):701-706. doi: 10.1038/s41586-021-04120-y (2021).
20. Lucas, C. *et al.* Impact of circulating SARS-CoV-2 variants on mRNA vaccine-induced immunity *Nature.* **600**(7889):523-529. doi: 10.1038/s41586-021-04085-y (2021).
21. Fonseca, M.H.G., de Souza, T.F.G., de Carvalho Araújo, F.M., de Andrade, L.O.M. Dynamics of antibody response to CoronaVac vaccine. *J Med Virol.* **94**(5):2139-2148. doi: 10.1002/jmv.27604 (2022).
22. Mishra, S.K. *et al.* Waning of Anti-spike Antibodies in AZD1222 (ChAdOx1) Vaccinated Healthcare Providers: A Prospective Longitudinal Study. *Cureus.* **13**(11):e19879. doi: 10.7759/cureus.19879 (2021).
23. Lim, S.H. *et al.* Serum Antibody Response Comparison and Adverse Reaction Analysis in Healthcare Workers Vaccinated with the BNT162b2 or ChAdOx1 COVID-19 Vaccine. *Vaccines* (Basel). **9**(12):1379. doi: 10.3390/vaccines9121379 (2021).
24. Poukka, E. *et al.* Cohort study of Covid-19 vaccine effectiveness among healthcare workers in Finland, December 2020 - October 2021. *Vaccine* **40**(5):701-705. doi: 10.1016/j.vaccine.2021.12.032 (2022).
25. Flisiak, R. *et al.* Effect of COVID-19 on Anti-S Antibody Response in Healthcare Workers Six Months Post-Vaccination. *Vaccines* (Basel) **9**(11):1325. doi: 10.3390/vaccines9111325 (2021).
26. Cucunawangsih, C., Wijaya, R.S., Lugito, N.P.H., Suriapranata, I. Antibody response to the inactivated SARS-CoV-2 vaccine among healthcare workers, Indonesia. *Int J Infect Dis.* **113**:15-17. doi: 10.1016/j.ijid.2021.09.078 (2021).
27. Russell, M.W., Moldoveanu, Z., Ogra, P.L., Mestecky, J. Mucosal Immunity in COVID-19: A Neglected but Critical Aspect of SARS-CoV-2 Infection. *Front Immunol.* **11**:611337, doi: 10.3389/fimmu.2020.611337 (2020).
28. Isho, B., *et al.* Persistence of serum and saliva antibody responses to SARS-CoV-2 spike antigens in COVID-19 patients. *Sci Immunol* **5**(52):eabe5511. doi: 10.1126/sciimmunol.abe5511 (2020).
29. Zurac, S. *et al.* COVID-19 vaccination and IgG and IgA antibody dynamics in healthcare workers. *Mol Med Rep.* **24**(2):578. doi: 10.3892/mmr.2021.12217 (2021).
30. Azzi, L. *et al.* Mucosal immune response in BNT162b2 COVID-19 vaccine recipients. *EBioMedicine.* **75**:103788. doi: 10.1016/j.ebiom.2021.103788 (2022).

31. Chivu-Economescu, M. *et al.* Kinetics and persistence of cellular and humoral immune responses to SARS-CoV-2 vaccine in healthcare workers with or without prior COVID-19. *J Cell Mol Med.* **26**(4):1293-1305. doi: 10.1111/jcmm.17186 (2022).
32. Mak, W.A. *et al.* SARS-CoV-2 antibody and T cell responses one year after COVID-19 and the booster effect of vaccination: A prospective cohort study. *J Infect.* **84**(2):171-178. doi: 10.1016/j.jinf.2021.12.003 (2022).
33. Constantin, C., Pisani, A., Bardi, G., Neagu, M. Nano-carriers of COVID-19 vaccines: the main pillars of efficacy. *Nanomedicine (Lond)* **16**(26):2377-2387. doi: 10.2217/nnm-2021-0250 (2021).
34. Neagu, M., Constantin, C., Surcel, M. Testing Antigens, Antibodies, and Immune Cells in COVID-19 as a Public Health Topic-Experience and Outlines. *Int J Environ Res Public Health.* **18**(24):13173. doi: 10.3390/ijerph182413173 (2021).
35. Carta, M. *et al.* Comparison of Anti-SARS-CoV-2 S1 Receptor-Binding Domain Antibody Immunoassays in Health Care Workers Before and After the BNT162b2 mRNA Vaccine. *Am J Clin Pathol.*;157(2):212-218. doi: 10.1093/ajcp/aqab107 (2022).
36. Hause, A.M. *et al.* Safety Monitoring of COVID-19 Vaccine Booster Doses Among Persons Aged 12-17 Years - United States, December 9, 2021-February 20, 2022. *MMWR Morb Mortal Wkly Rep.* **71**(9):347-351. doi: 10.15585/mmwr.mm7109e2 (2022).
37. Vizcarra, P. *et al.* BNT162b2 mRNA COVID-19 vaccine Reactogenicity: The key role of immunity. *Vaccine.* **39**(51):7367-7374. doi: 10.1016/j.vaccine.2021.10.074 (2021).
38. Tsatsakis A, *et al.* Immune response (IgG) following full inoculation with BNT162b2 COVID-19 mRNA among healthcare professionals. *Int J Mol Med.* **48**(5):200. doi: 10.3892/ijmm.2021.5033 (2021).
39. Meng, H., Mao, J., Ye, Q. Booster vaccination strategy: Necessity, immunization objectives, immunization strategy, and safety. *J Med Virol.* Jan 13. doi: 10.1002/jmv.27590 (2022).
40. Hollstein, M.M. *et al.* Interdependencies of cellular and humoral immune responses in heterologous and homologous SARS-CoV-2 vaccination. *Allergy.* 2022 Feb 6. doi: 10.1111/all.15247.
41. Spitzer, A. *et al.* Association of a Third Dose of BNT162b2 Vaccine With Incidence of SARS-CoV-2 Infection Among Health Care Workers in Israel. *JAMA.***327**(4):341-349. doi: 10.1001/jama.2021.23641 (2022).
42. Yorsaeng, R. *et al.* Immunogenicity of a third dose viral-vectored COVID-19 vaccine after receiving two-dose inactivated vaccines in healthy adults. *Vaccine.* **40**(3):524-530 doi: 10.1016/j.vaccine.2021.11.083 (2022).
43. Glück, V. *et al.* Immunity after COVID-19 and vaccination: follow-up study over 1 year among medical personnel. *Infection.* **50**(2):439-446. doi: 10.1007/s15010-021-01703-9 (2021).
44. Olariu, T.R., Ursoniu, S., Marincu, I., Lupu, MA. Dynamics of Antibody Response to BNT162b2 mRNA COVID-19 Vaccine: A 7-Month Follow-Up Study. *Medicina (Kaunas)* **57**(12):1330. doi: 10.3390/medicina57121330 (2021).

45. Rahman, S., *et al.* COVID-19 reinfections among naturally infected and vaccinated individuals. *Sci Rep* **12**, 1438 <https://doi.org/10.1038/s41598-022-05325-5> (2022).
46. Ontañón, J. *et al.* Influence of past infection with SARS-CoV-2 on the response to the BNT162b2 mRNA vaccine in health care workers: Kinetics and durability of the humoral immune response. *EBioMedicine*. **73**:103656.doi: 10.1016/j.ebiom.2021.103656 (2021).

Figures

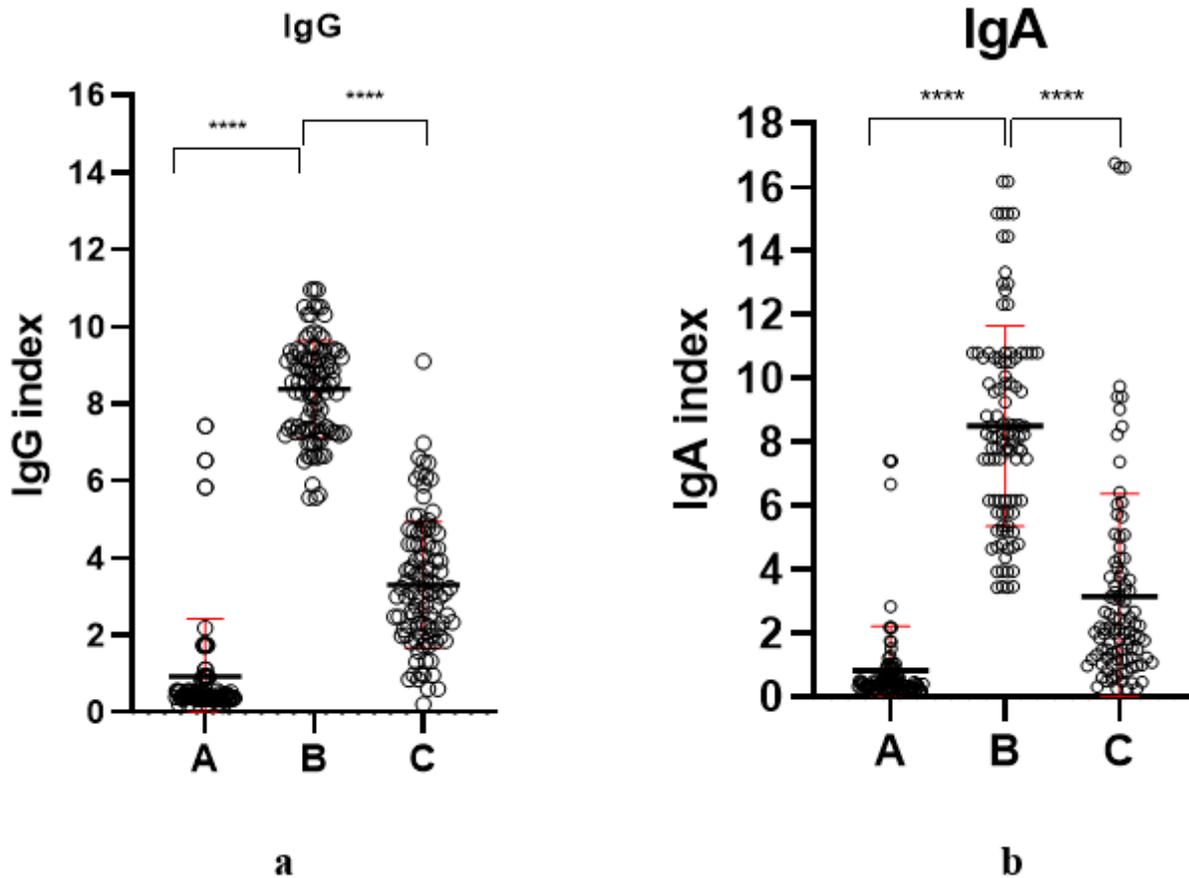


Figure 1

Ig indexes dynamics for the entire group regardless of their prior infection or not, before vaccination (A), after completion of the vaccination scheme (B) and 8 months after vaccination (C). **a.** IgG index; **b.** IgA index (red line mean \pm SD).

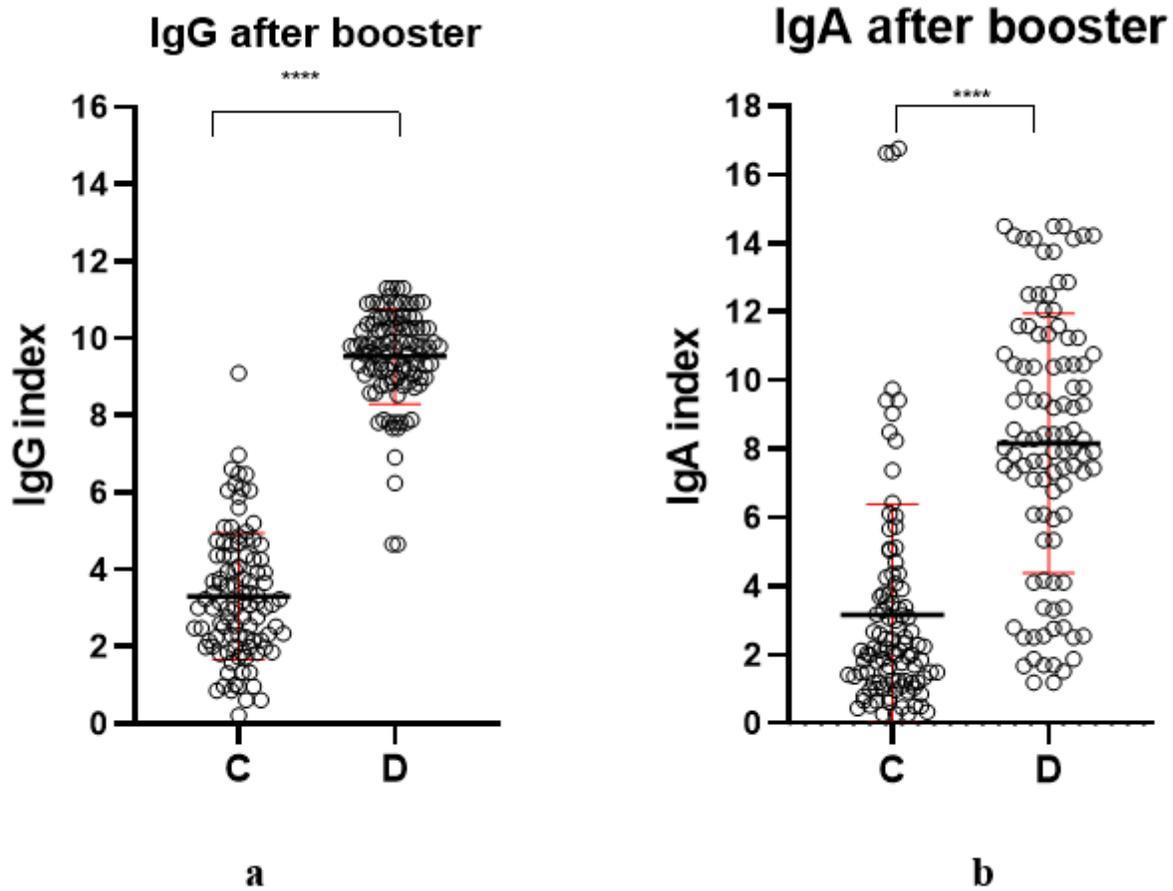


Figure 2

Ig indexes dynamics for the entire group regardless of their prior infection or not, 8 months after vaccination (C) and after booster vaccination (D); **a.** IgG index; **b.** IgA index (red line mean \pm SD).

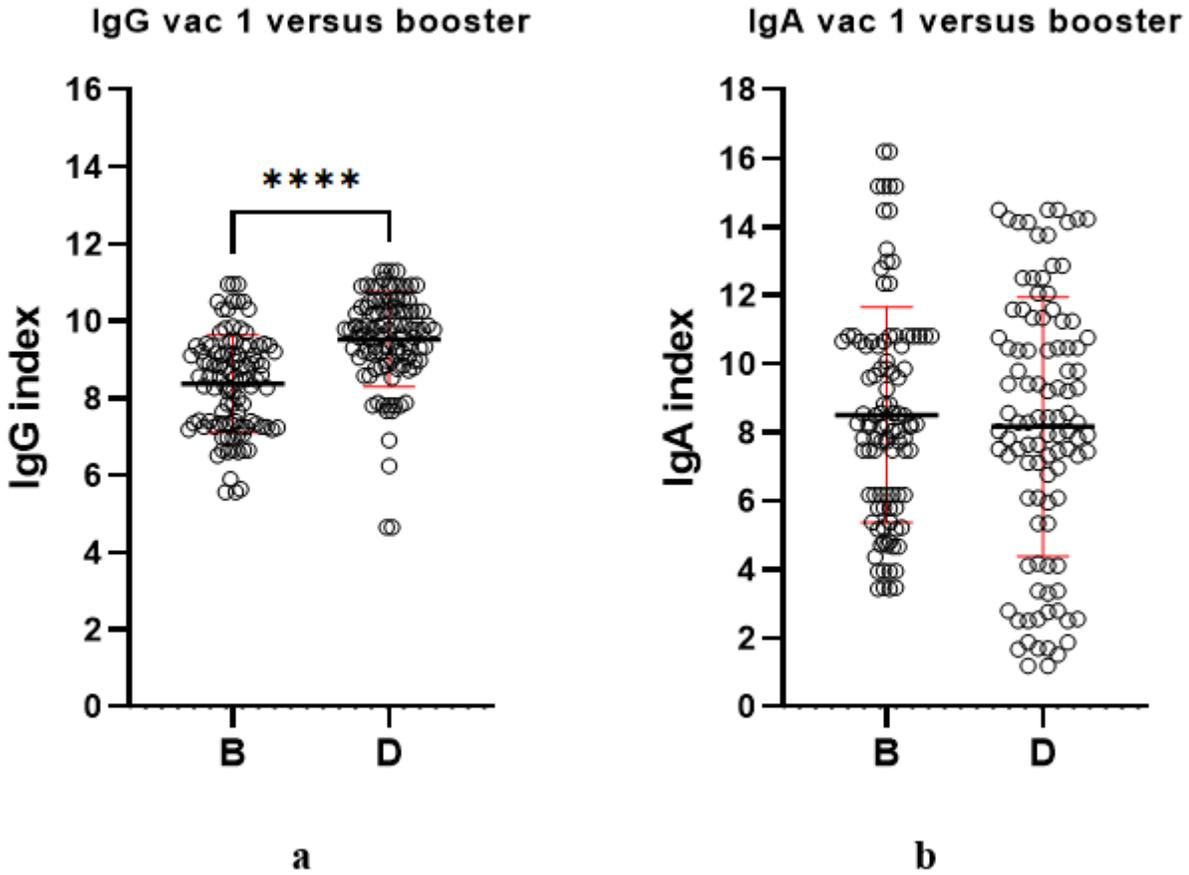


Figure 3

Ig indexes dynamics for the entire group regardless of their prior infection or not, first vaccination (B) compared to booster vaccination (D); **a**. IgG index; **b**. IgA index (red line mean \pm SD).

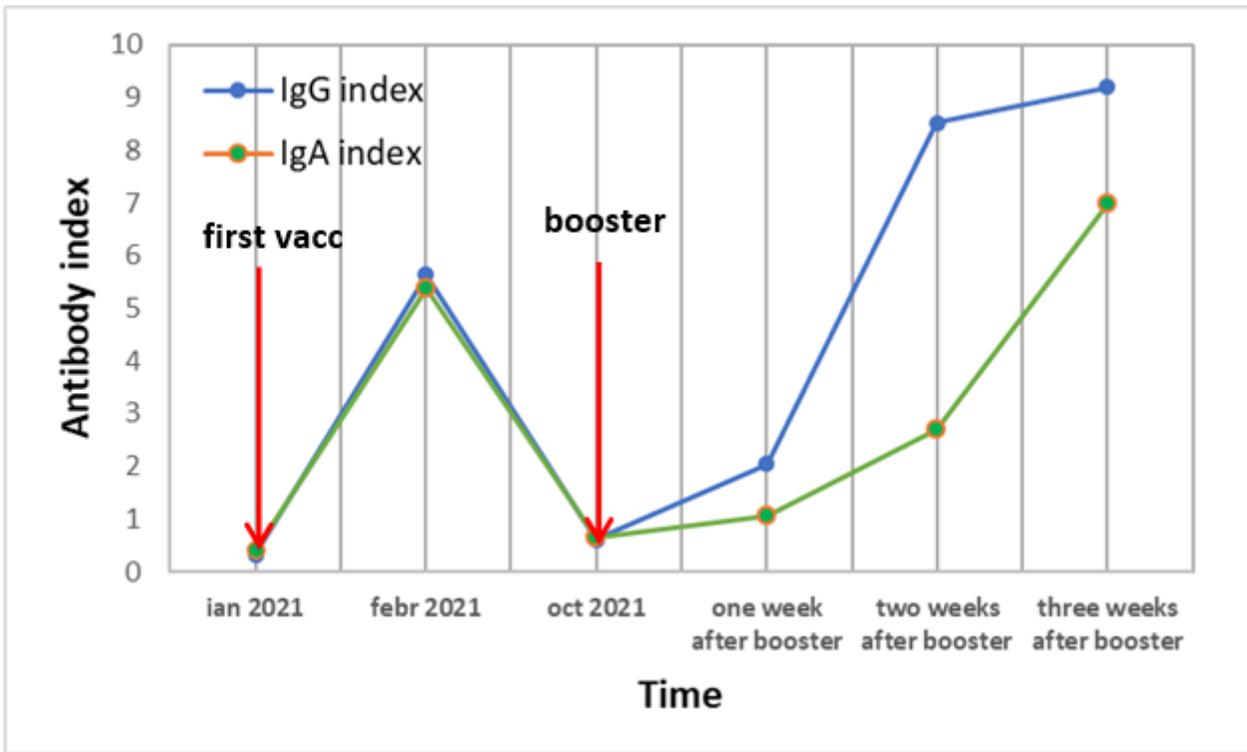


Figure 4

Individual values of IgG and IgA indexes after the first vaccination and booster in a non-infected individual