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Wang Ruru

Anhui Agricultural University

Mingxing Gu

Anhui Agricultural University

Yanzhong Zhang

Anhui Agricultural University

Qinglin Zhong

Anhui Agricultural University

Linbo Chen

Yunnan Academy of Agricultural Sciences

Daxiang Li

Anhui Agricultural University

Zhongwen Xie (✉ zhongwenxie@ahau.edu.cn)

Anhui Agricultural University

Article

Keywords:

Posted Date: April 15th, 2022

DOI: <https://doi.org/10.21203/rs.3.rs-1529592/v1>

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Long Term Green Tea Intervention with Exercise Efficiently Ameliorate Hepatic Steatosis and Obesity Complication in High Fat Diet Induced Mice

1 Ruru Wang^{1†}; Mingxing Gu^{1†}; Yanzhong Zhang^{2†}; Qinglin Zhong¹; Llinbo Chen³;
2 Daxiang Li¹; Zhongwen Xie^{1*}

3 ¹State Key Laboratory of Tea Plant Biology and Utilization, School of Tea and Food
4 Sciences and Technology, Anhui Agricultural University, Hefei, China

5 ²Department of Sports Sciences, Anhui Agricultural University, Hefei, China

6 ³Tea Research Institute, Yunnan Academy of Agricultural Sciences, Menghai, China

7 † **These authors contribute equally.**

8 * **Correspondence:** Zhongwen. Xie, E-mail: zhongwenxie@ahau.edu.cn

9 **Abstract**

10 Dietary habits and exercise play important roles in wellbeing of human health.
11 Currently, how long of drinking tea with exercise could be efficiently ameliorating
12 hepatic steatosis and obesity complication still need to be investigated. Here, we
13 comparatively explored short and long term green tea consumption plus exercise improve
14 hepatic steatosis and obesity complication in high fat diet (HFD) mice. Our results
15 showed that Yunkang 10 green tea (YKGT) plus exercise exhibited synergistic
16 prevention effects on ameliorating hepatic steatosis and obesity complication at the 8-
17 week intervention. Moreover, 22-week intervention with YKGT or exercise improved all
18 symptoms of obesity complication, which indicated that long term intervention exhibited
19 profound preventive effects than short term. All treatments inhibited NF-κB activation
20 and pro-inflammatory cytokine expression at both 8- and 22-week intervention, indicated
21 that inflammation may be a cause event for developing hepatic steatosis. The key
22 molecules for regulating lipid and glucose metabolism SCD1 were obviously down-
23 regulated, and GLU2 and PPAR γ were significantly up-regulated by YKGT and exercise
24 in the liver of HFD mice. This study demonstrated that long term intervention with
25 YKGT and exercise effectively relieved hepatic steatosis and obesity complication by
26 ameliorating hepatic inflammation, reducing lipid synthesis and accelerating glucose
27 transport.

28 **1 Introduction**

29 Dietary habits and exercise play important roles in the well-being of human health.
30 Currently, how long drinking tea with exercise could efficiently ameliorate hepatic
31 steatosis and obesity complication still need to be investigated. Hepatic steatosis
32 associated with obesity is the most common cause of chronic liver disease and may
33 progress to nonalcoholic steatohepatitis and end-stage liver disease¹. The risk for
34 developing hepatic steatosis is closely related to diet and lifestyle. The rapid development
35 of the economy and technology has resulted in increasing food supply and declining
36 physical activity^{2, 3}. Over the past decades, sedentary lifestyles and over eating habits
37 are primary causes for the growing rate of obesity complications across the world⁴⁻⁶.
38 Although great progress has been made in the treatment of obese complications, some
39 potential side effects are inevitable^{6, 7}. At present, lifestyle improvement, diet
40 intervention, and exercise therapy are the basic strategies for the prevention of obesity
41 complications. Regular exercise training increases oxidative capacity, lipid metabolism,
42 reduces serum triglycerides, blood pressure^{8, 9}. However, what types of diet and exercise
43 would exhibit synergistic beneficial effects? How long intervention could perform
44 profound beneficial effects? And what is the underlying molecular mechanism? All these
45 questions need to be further investigated.

46 Tea is one of the most popular beverages because of its natural and healthy
47 properties. Green tea has a high amount of monomeric catechin, and (-)-epigallocatechin
48 gallate (EGCG) is the major component of catechin. Many reports have previously
49 demonstrated that catechins, especially EGCG, promote fat oxidation, lower blood total
50 cholesterol and triglycerides, and reduce body weight¹⁰. Yun Kang 10 (*Camellia sinensis*
51 var. *assamica* cv. Yun Kang 10) is a widely cultivated tea cultivar in Southwestern China.
52 The chemical profiling and volatile components of Yun Kang 10 have been reported¹¹.
53 In a previous paper, our group reported that YKGT has a higher amount of monomeric
54 catechin, and a combination of YKGT supplement with aerobic exercise synergistically
55 ameliorated existing metabolic syndrome¹². Previous studies also reported that a
56 combination of green tea and exercise facilitates sports performance and endurance
57 capacity, and effectively prevents obesity^{13, 14}. The goal of this study is to investigate
58 whether a combination YKGT supplement and physical exercise exhibits synergistically

59 preventive effects, and to compare how long intervention period (8 week vs. 22 week)
60 will show effectively preventive effects on hepatic steatosis and obese complication
61 induced by HFD in C57BL/6J Mice.

62 **2 Material and methods**

63 **2.1 Tea sample**

64 YKGT was produced by steaming the leaves collected from Yun Kang 10 tea trees
65 (*Camellia sinensis* var. *assamica* cv. Yun Kang 10) in Menghai County of Yunnan
66 province China following a standard protocol. YKGT was further crushed into powder by
67 BLENDER 800S (Warning, Corp., Torrington, CT, USA), and was added into HFD at
68 the concentration of 5% (w/w).

69 **2.2 Animal experiments and ethics approval**

70 The specific pathogen-free male C57BL/6J mice, age of five weeks, the weight of
71 18~21 g, were purchased from Vital River Laboratory Animal Technology Co., Ltd.
72 (Beijing, China). The animals were housed in cages at the Animal Facility Center of
73 Anhui Agricultural University, which was controlled with constant temperature ($22\pm 1^\circ\text{C}$)
74 and humidity ($50\pm 5\%$) with a 12:12 h light-dark cycle falls on 8:00 a.m. to 8:00 p.m. The
75 mice were provided with standard AIN93 food and water ad libitum. After 3-week of
76 acclimation, the mice were fed with low-fat-diet (LFD, TP23303, 11% of energy derived
77 from fat), high-fat-diet (HFD, TP23300, 60% of energy derived from fat), HFD with 5%
78 YKGT (YKGT), HFD with treadmill exercise (Ex), HFD with YKGT plus Ex
79 (YKGT+Ex), respectively. Each group has 12 mice. All diets were obtained from Trophic
80 Animal Feed High-tech Co., Ltd (Nantong, China). The composition of diets was listed
81 as Supplementary table S1. The exercise mice received treadmill running 6 days per week
82 during the experimental period. A detailed treadmill running schedule was provided in
83 Supplementary Table S2. Food intake and water drink were recorded daily. Body weight
84 was monitored with a weight scale weekly.

85 **2.3 Serum and tissue samples collection**

86 After 8 weeks of treatment, or at the end of 22 weeks of intervention, upon
87 overnight fasting, 6 mice from each group were anesthetized via injection with 4%

88 chloral hydrate (10 mL/kg, i.p.), and were then sacrificed. Peripheral blood was collected
89 from the ophthalmic vein. Serum was obtained by centrifugation at 3000 rpm/min for 5
90 min at 4°C, and then stored at -80°C. Liver and abdominal fat weights were measured on
91 a scale. A small piece of liver tissues was preserved in RNA stabilization solution
92 (Thermo Fisher Scientific, Baltics, USA) for gene expression analysis, and was fixed in
93 formaldehyde solution (Zhanyun, Jiangsu, China) for the histological experiment,
94 respectively. The rest of liver tissues were immediately liquid nitrogen frozen and stored
95 at -80°C for protein expression studies.

96 2.4 Serum biochemical parameter analysis

97 The serum LDL and TC were measured using micro test kits (Johnson medical
98 equipment, Shanghai, China). The enzymatic activity of ALT was analyzed using enzyme
99 kits (Jiancheng Biotechnology, Nanjing, China). Fasting blood glucose was measured
100 using Nova StatStrip XpresstM Glucose CR Meter (Nova Biomedical, Waltham, UK)
101 with Nova StatStrip XpresstM Glu-test Strips (Nova Biomedical, Waltham, UK).

102 2.5 Quantitative Real-Time PCR Assay

103 Real-time PCR was performed with SYBR Green Master Mix using Real-Time PCR
104 Detection System (CFX96 Touch, Bio-RAD, USA) following the previous protocol¹⁵.
105 Primer sequences used for this study are listed in supplementary table S3.

106 2.6 Western blot analysis

107 Western blot was performed following the method described previously¹⁶. The
108 primary antibodies included total-IKK β , phosphorylation-IKK α/β , total and
109 phosphorylation -I κ B α , total and phosphorylation P 65 (Cell Signaling Technology, MA,
110 USA), SCD1, GLUT2, PPAR γ and β -actin (Santa Cruz, CA, USA). Appropriate HRP
111 conjugated secondary antibodies were from Santa Cruz (CA, USA). The intensities of
112 protein expression were analyzed using Image J software. β -actin was used as an internal
113 control.

114 2.7 Hematoxylin-eosin staining

115 Hematoxylin-eosin (HE) staining was performed following the published protocol¹⁷.
116 All the images were obtained using microscope (LEICA DM500, USA). Hepatic adipose
117 infiltration cells were counted manually using Image J software.

118 2.8 Statistical analysis

119 The statistic results were presented as mean \pm S.E.M. Graph Pad Prism5 software
120 was used for statistical analysis. Multiple groups were compared by one-way or two-way
121 ANOVA with Tukey's test when appropriate. The student's t-test was conducted to
122 determine significant differences between specific two groups. $P < 0.05$ is considered a
123 statistically significant difference.

124 3 Results

125 3.1 YKGT and Ex ameliorated obese complication in HFD mice

126 An 8-week HFD feeding significantly increased C57BL/6J mice body weight,
127 abdominal fat weight to body weight ratio, serum glucose, TC, LDL, and activity of ALT
128 (**Figure 1 A-G**). Ex alone for 8-week just averted body weight and TC increase (**Figure 1**
129 **A, F**). YKGT alone for 8-week significantly prevented the increases of body weight,
130 abdominal fat weight to body weight ratio, glucose, and ALT activity. However,
131 intervention with YKGT+Ex for 8-week exhibited synergetic effects and prevented all
132 these indexes increases (**Figure 1 A-G**). HFD feeding or YKGT, Ex, YKGT+Ex
133 treatment for 8-week did not alter liver weight to body weight ratio (**Figure 1C**). While,
134 22-week intervention with YKGT, Ex, YKGT+Ex improved all indexes (**Figure 1A-G**).
135 Our data indicated that long-term intervention exhibited more profound preventive effects
136 than short-term. Liver tissue sections showed the same physiological structures (**Figure 2**
137 **A-E**) and existed little amount adipose infiltration cells (**Figure 2 F**) from all 8-week
138 intervention groups of mice. However, 22-week HFD feeding mice displayed aberrantly
139 fatty hepatocytes with high volumes of lipid droplets. Treatment with YKGT, Ex, or
140 YKGT plus Ex persevered the normal liver architecture with minimal deposition of fat
141 droplets in hepatocytes (**Figure 2G-L**).

142 3.2 YKGT and Ex ameliorated liver inflammation in HFD mice

143 The mRNA expression of pro-inflammatory cytokines IL-6, TNF- α , and MCP-1 was
144 significantly up-regulated in the liver of HFD group mice compared to that of LFD group
145 mice. However, intervention with YKGT, Ex, or YKGT plus Ex for both 8-week and 22-
146 week significantly prevented the increases of the cytokines genes expression in liver
147 tissue (**Figure 3 A-C**). The phosphorylation of IKK and I κ B α is a key process for

148 activating of NF- κ B pathway. The western blotting showed that the phosphorylation of
149 $\text{IKK}\alpha/\beta$, $\text{I}\kappa\text{B}\alpha$, and P65 protein were all dramatically increased in the HFD mice liver
150 compared to that of LFD mice. Intervention with YKGT, Ex, or YKGT+Ex for both 8-
151 week and 22-week dramatically prevented these phosphorylated protein increases
152 compared to HFD mice (**Figure 3 D-O**).

153 The data showed that the phenotype of fatty liver was not altered when mice were
154 fed HFD for 8-week (**Fig 2 A-F**). However, pro-inflammatory cytokines IL-6, TNF- α ,
155 and MCP1 were significantly up-regulated by 8-week HFD feeding (**Fig 3 A-C**). Feeding
156 HFD for 8-week, the mRNA and protein expression of SCD1 did not change in the liver
157 of HFD mice compared to that of LFD mice. However, in an 8-week intervention with
158 YKGT, Ex, or YKGT+Ex, the mRNA and protein expression of SCD1 was significantly
159 down-regulated (**Figure 4 A, B, D**). HFD feeding for 22-week significantly up-regulated
160 the SCD1 expressions in the liver of HFD group mice compared to LFD group mice
161 (**Figure 4 A, C, E**). However, the increase of SCD1 gene and protein expression was
162 significantly prevented by YKGT, Ex, or YKGT+Ex intervention (**Figure 4 A, C, E**).
163 Our data further found that either YKGT or Ex prevented both mRNA and protein
164 expression of SCD1 in the liver of HFD mice (**Fig 4 A-E**).

165 3.3 YKGT and Ex accelerated glucose metabolism in HFD mice

166 HFD feeding for 22-week significantly decreased GLUT2 gene and protein
167 expression in the liver of HFD mice. However, a supplement of YKGT, or Ex or YKGT
168 plus Ex, significantly increased GLUT2 gene and protein expression in the liver (**Figure**
169 **5 A, C, E**). In addition, the gene and protein expression of PPAR γ did not alter in the
170 liver of HFD group mice compared to that of LFD group mice. While, YKGT, Ex, or
171 YKGT plus Ex obviously increased PPAR γ gene and protein expression in the liver of
172 treated group mice (**Figure 5 B, D, F**). Our data further revealed that intervention with
173 YKGT, Ex, or YKGT plus Ex for 22-week significantly up-regulated GLUT2 gene and
174 protein expression in the liver of HFD mice (**Figure 5 A, C, E**), which may contribute to
175 ameliorates HFD induced hepatic steatosis. The results found that gene and protein
176 expression of PPAR γ was increased in the liver of HFD mice treated with YKGT, Ex, or
177 YKGT plus Ex for 22-week (**Figure 5 B, D, F**).

178 4 Discussion

179 Obesity complication includes obesity, dyslipidemia, diabetes, and fatty liver.
180 Inflammation initiates a vicious cycle between obesity and nonalcoholic fatty liver
181 disease¹⁸. Fatty liver disease is characterized by chronic hepatic inflammation.
182 Numerous inflammatory cytokines are involved in the process of developing into hepatic
183 steatosis. Martin et al. reported that short-term GT supplementation did not affect glucose
184 kinetics, however, GT was associated with attenuated insulinemia¹⁹. Recently, Bagheri
185 et al. found that the combination of GTE and exercise promotes a further decrease in
186 weight, body mass index, and body fat percentage than exercise alone in inactive
187 overweight women²⁰. Khoo et al. also reported that the combination of decaffeinated
188 green tea extract and voluntary exercise synergistically mitigated nonalcoholic fatty liver
189 disease in HFD mice²¹. The current data indicated that hepatic inflammation is a
190 preceding event for developing hepatic steatosis. YKGT supplement and treadmill
191 exercise together only exert synergetic beneficial effects for short-term intervention. The
192 long-term 22-week intervention exhibited profound preventive effect than that of the 8-
193 week treatment. The schematic diagram for this study is shown in **Figure 6**.

194 Indeed, Ma et al found that HFD elevated plasma IL-6, and blockade of IL-6
195 signaling ameliorated systemic insulin resistance and improves hepatic steatosis in HFD
196 fed mice²². The NF- κ B contains multiple proteins complex that controls cytokine
197 production. The phosphorylation of IKK and I κ B α is a key factor for activation of NF- κ B
198 pathway²³. Our results showed that YKGT, Ex or YKGT+Ex all decreased the
199 phosphorylation of IKK α/β and I κ B α , and thus inhibited the transcript of pro-
200 inflammatory cytokines at both short and long term intervention. Moreover, the
201 phosphorylated NF κ B p65 subunit were significantly down-regulated by YKGT, Ex or
202 YKGT+Ex at 8- and 22-week intervention. Li et al. reported that GTE decreased
203 phosphorylation of the NF κ B p65 subunit and alleviated nonalcoholic steatohepatitis
204 NASH in HFD induced mice²⁴. Our data further found that YKGT, exercise or YKGT
205 drinking plus exercise ameliorated hepatic steatosis via inhibiting NF κ B activation in
206 liver of HFD mice.

207 The accumulation of excess fat in the liver is a primary cause of hepatic steatosis.
208 SCD1 is a key regulator for de novo lipogenesis in the liver. SCD1 is also a central
209 regulator of fuel metabolism and catalyzes the synthesis of monounsaturated fatty acids

210 (MUFA). Li et al. reported that hepatic SCD1 plays a key role in the prevention of
211 steatohepatitis by partitioning excess lipid into MUFA²⁵. Zhou et al. found that aqueous
212 extract of post-fermented tea reversed the hepatic steatosis of hyperlipidemia rats by
213 down-regulating the hepatic SCD1 genes expression in HFD fed rats²⁶. Consistent with
214 previous reports, this study showed that an increase of inflammatory cytokines is a
215 preceding inducer in the development of hepatic steatosis, while YKGT and Ex inhibited
216 the expression of SCD-1 in the liver of HFD mice. Moreover, the liver is an important
217 organ for nutrient metabolism. GLUT2 is the predominant glucose transporter in
218 hepatocytes and PPAR γ is the master regulator of glucose metabolism which plays a
219 significant role in protecting the liver from inflammation, oxidation, fibrosis, fatty liver²⁷.
220 Few studies have investigated the exact signaling molecules that may explain the
221 mechanism following exercise training with or without longer-term GTE intake in human
222 and animal studies²⁸. The GLUT2 represents the main gate of glucose uptake by the liver.
223 Ahmed et al. reported that genetic depletion of Soat2 diminished hepatic steatosis via
224 genes regulating de novo lipogenesis and by GLUT2 in female mice²⁹. Green tea
225 polyphenol extract up-regulates the GLUT2 expression in the liver of rats fed a high
226 fructose diet³⁰. Importantly, we further extended our mechanistic exploration in this
227 study and found that short-term drinking tea and exercise might perform synergetic health
228 benefits, and long-term tea-drinking habit or regular exercise could produce the same
229 beneficial effects.

230 Overall, this study demonstrated that YKGT supplement and exercise effectively
231 relieved hepatic steatosis and obesity complication in HFD Mice by ameliorating hepatic
232 inflammation, reducing lipid synthesis, and accelerating glucose transport and
233 metabolism. Our results suggested that long-term green tea drinking and regular aerobic
234 exercise might be a good habit for preventing hepatic steatosis and obesity complication
235 for the human population.

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328

329 **Data Availability Statement**

330 The original contributions presented in the study are included in the article or
331 supplementary materials, further inquiries can be directed to the corresponding author.

332 **Author Contributions**

333 Ruru Wang, Mingxing Gu, and Yanzhong Zhang performed the experiments and drafted
334 the manuscript. Qinglin Zhong and Linbo Chen designed the experiments and performed
335 the statistical analysis. Daxiang Li, Zhongwen Xie designed the study, edited the
336 manuscript. All authors contributed to the article and approved the submitted version.

337 **Funding**

338 This work was supported by the key joint grant for regional innovation from the National
339 Natural Science Foundation of China to Z.X. [Grant Number U19A2034]; a key grant for
340 the University Synergy Innovation Program of Anhui Province to Z.X. [Grant Number
341 GXXT-2019-49]; an open grant from State Key Laboratory of Tea Plant Biology and
342 Utilization to Y. Z [Grant Number SKLTOF20170102]; and a grant from China
343 Agriculture Research System of MOF and MARA [CARS-19].

344 **Ethics Statement**

345 All the animal experimental procedures imposed in this study were by guidelines of
346 institutional animal care and use committee (IACUC) of Anhui Agricultural University
347 with ethical approval code AHAU 2016-002.

348 **Statement**

349 The study is reported in accordance with ARRIVE guidelines.

350 **Additional information**

351 **Conflict of interest**

352 The authors declare that they have no known competing financial interests or personal
353 relationships that could have appeared to influence the work reported in this paper.

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356 and institutional affiliations.

357

358 **Figure captions**

359 **Fig. 1.** YKGT and exercise improved symptoms of obesity complications in HFD
360 C57BL/6J mice. Body weight (A), abdominal fat weight to body weight ratio (B) and
361 liver weight to body weight ratio (C), glucose (D), LDL (E), TC (F), and ALT (G) were
362 measured at 8 and 22-week intervention, respectively. * P<0.05, ** P<0.01, ***P<0.001,
363 compared to LFD; and #P<0.05, ##P<0.01, ###P<0.001 compared to HFD. YKGT,
364 Yunkang 10 Green Tea; LDL, low-density lipoprotein; TC, total cholesterol; ALT,
365 alanine aminotransferase activity. (n=6, mean ± SEM).

366 **Fig. 2.** YKGT and exercise ameliorated fatty liver in HFD C57BL/6J mice. The HE
367 staining of liver sections of LFD groups (A, G), HFD groups (B, H), HFD supplement
368 YKGT groups (C, I), HFD with Ex groups (D, J), HFD with YKGT plus Ex groups (E,
369 K), and statistic results of hepatic adipose infiltration cell (F, L) were presented at 8 and
370 22-week intervention, respectively. ***P<0.001, compared to LFD; and #P<0.05,
371 ##P<0.01, compared to HFD (Note: 5 images per section and 3 sections per mice were
372 randomly selected for statistical analysis, n=3, mean ± SEM).

373 **Fig. 3.** YKGT and exercise inhibited inflammation and NFκB activation in the liver of
374 HFD C57BL/6J mice. The mRNA expression of *IL-6* (A), *TNFα* (B), *MCPI* (C) in the
375 liver tissues were quantified by Real Time PCR at 8 and 22 weeks of intervention,
376 respectively. The representing images of protein expression of total IKKβ and
377 phosphorylated IKKα/β (D, E), and statistical results of D (F) and E (G); total and
378 phosphorylated IκBα (H, I), and statistical results of H (J) and I (K); total and
379 phosphorylated P65 (L, M), and statistical results of L (N) and M (O) in the liver tissues
380 at 8 and 22 weeks intervention, respectively. *P<0.05, **P<0.01, ***P<0.001, compared
381 to LFD; and #P<0.05, ##P<0.01, ###P<0.001 compared to HF. NF-κB, nuclear factor
382 kappa light chain enhancer of activated B cells; IL-6, interleukin 6; TNFα, tumor necrosis
383 factor α; MCP-1, monocyte chemotactic protein-1 (n=3-6, mean ± SEM).

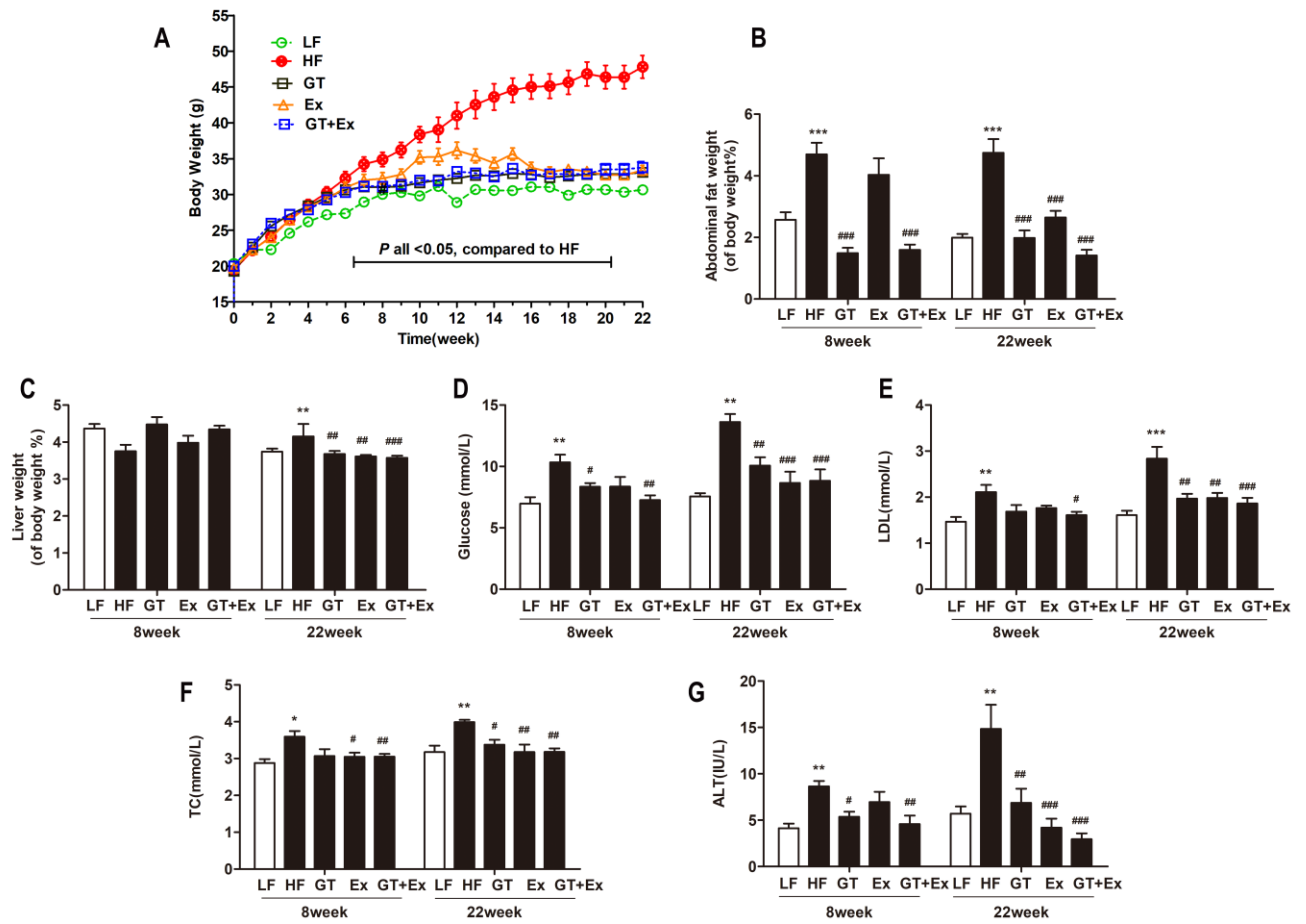
384 **Fig. 4.** YKGT and exercise inhibited SCD1 gene and protein expression in the liver of
385 HFD C57BL/6J mice. The mRNA expression of *SCD1* (A), representing images of
386 protein expression of SCD1 (B, C), and statistical results of B (D) and C (E) were
387 presented in the liver tissues at 8 and 22 weeks intervention, respectively. *P<0.05,

388 ***P<0.001, compared to LF; and #P<0.05, ##P<0.01, ###P<0.001 compared to HF.
389 SCD1, Stearoyl CoA desaturase 1 (n=3, mean ± SEM).

390 **Fig. 5.** YKGT and exercise increased GLU2 and PPAR γ gene and protein expression in
391 the liver of HFD C57BL/6 mice. The mRNA expression of *GLU2* mRNA (A) and *PPAR γ*
392 (B), representing images of protein expression of GLU2 (C) and PPAR γ (D), statistical
393 results of C (E) and D (F) were presented in the liver tissues at 22 weeks intervention,
394 respectively. *P<0.05, compared to LF; and #P<0.05, ##P<0.01, compared to HF. GLU 2,
395 glucose transporter 2; PPAR γ , peroxisome proliferator-activated receptor γ (n=3, mean ±
396 SEM).

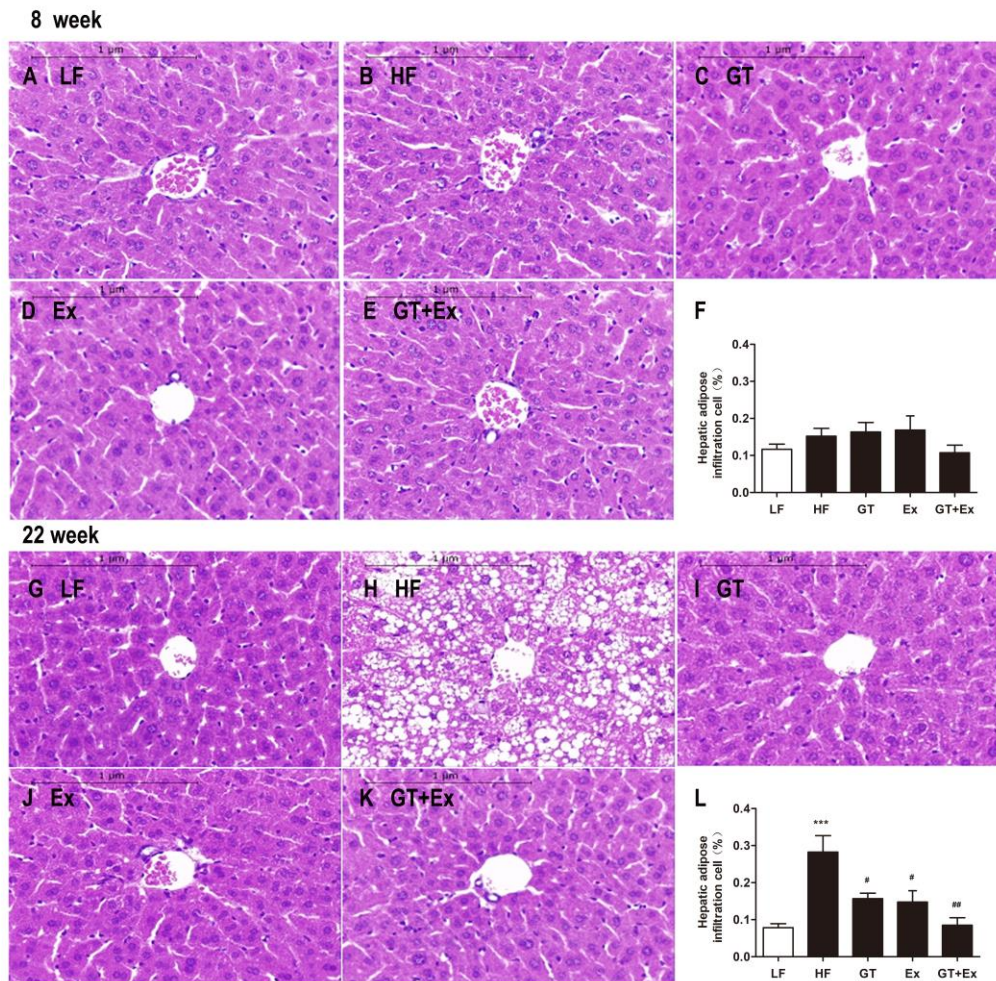
397 **Fig. 6.** The schematic diagram for this study.

398



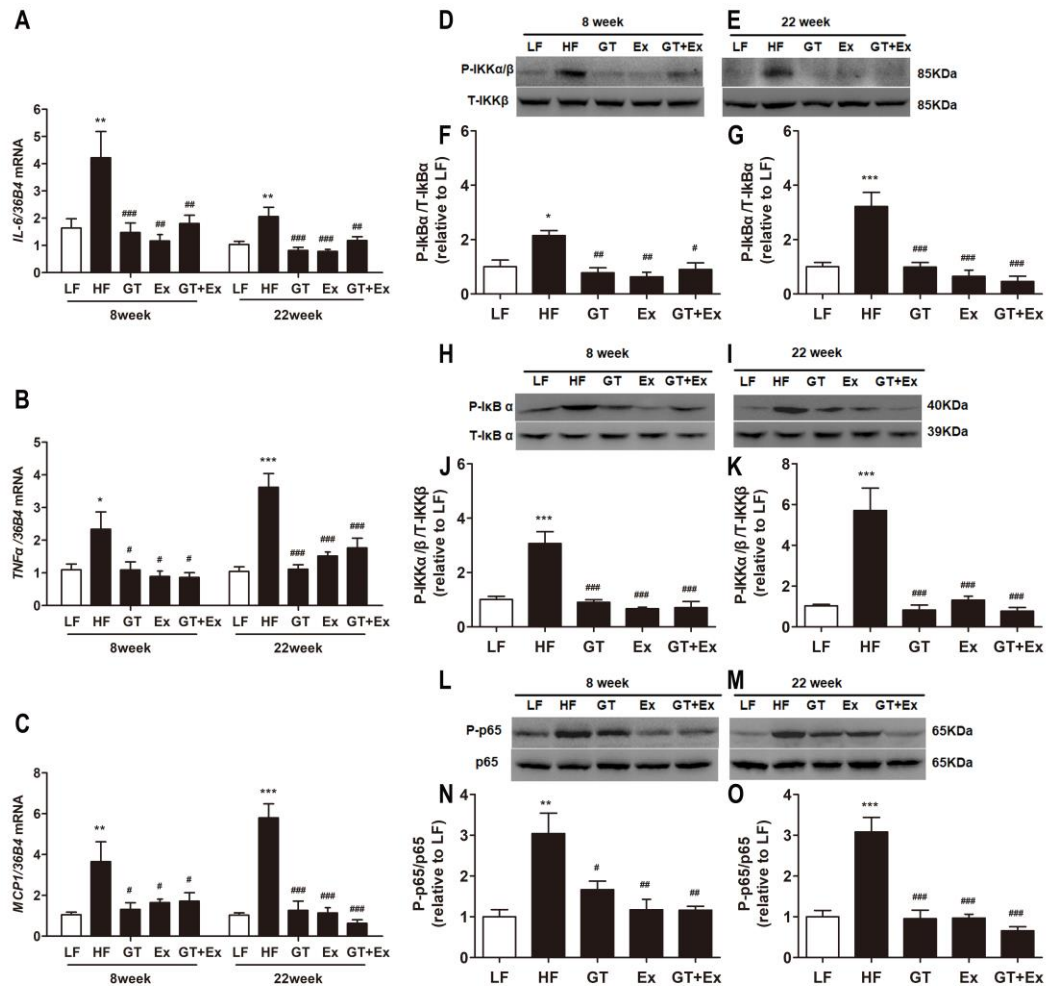
399

400 Fig. 1. YKGT and exercise improved symptoms of obesity complications in HFD
 401 C57BL/6J mice. Body weight (A), abdominal fat weight to body weight ratio (B) and
 402 liver weight to body weight ratio (C), glucose (D), LDL (E), TC (F), and ALT (G) were
 403 measured at 8 and 22-week intervention, respectively. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$,
 404 compared to LFD; and # $P < 0.05$, ## $P < 0.01$, ### $P < 0.001$ compared to HFD. YKGT,
 405 Yunkang 10 Green Tea; LDL, low-density lipoprotein; TC, total cholesterol; ALT,
 406 alanine aminotransferase activity. (n=6, mean \pm SEM).



407

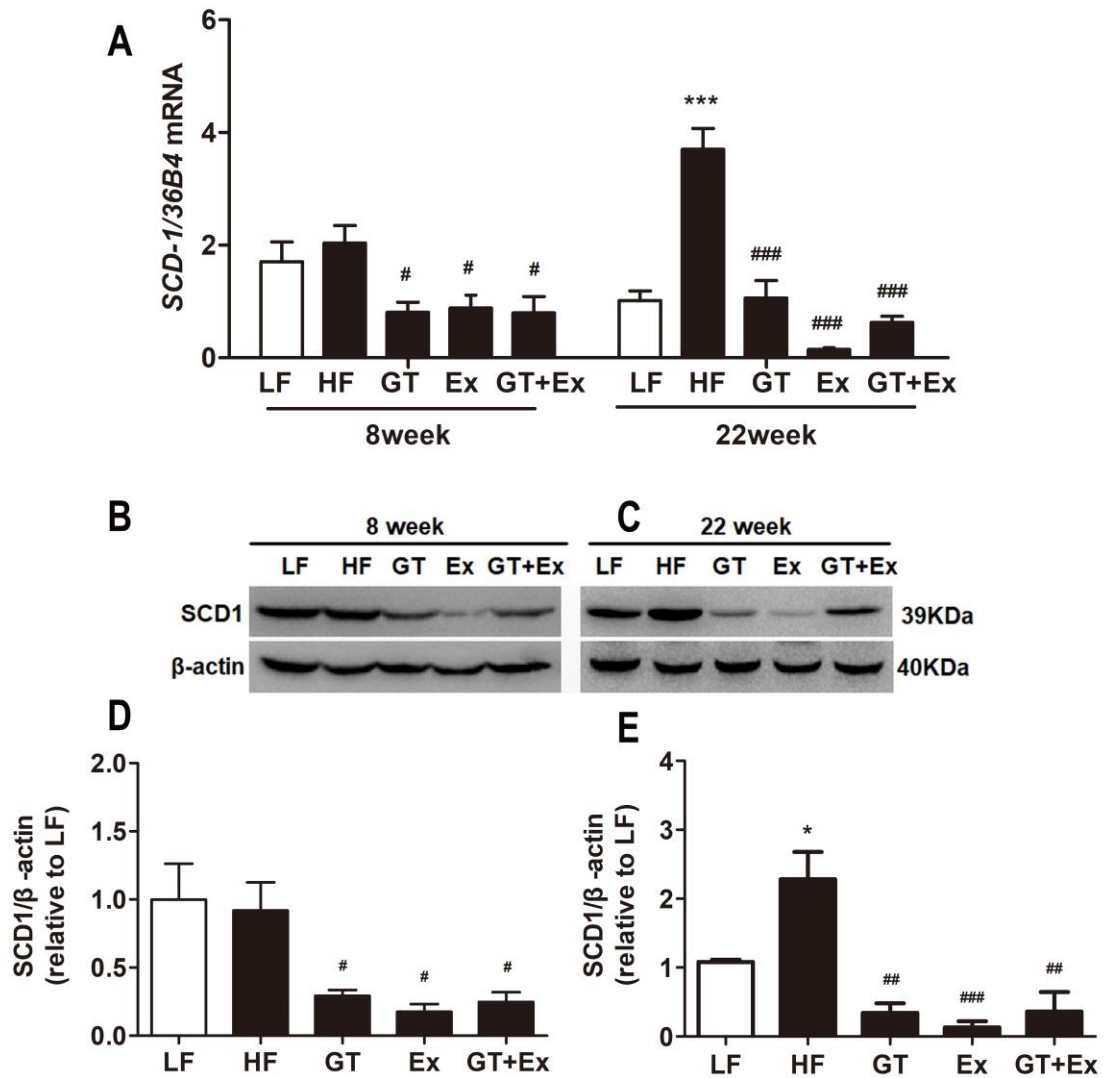
408 Fig. 2. YKGT and exercise ameliorated fatty liver in HFD C57BL/6J mice. The HE
 409 staining of liver sections of LFD groups (A, G), HFD groups (B, H), HFD supplement
 410 YKGT groups (C, I), HFD with Ex groups (D, J), HFD with YKGT plus Ex groups (E,
 411 K), and statistic results of hepatic adipose infiltration cell (F, L) were presented at 8 and
 412 22-week intervention, respectively. *** $P < 0.001$, compared to LFD; and # $P < 0.05$,
 413 ## $P < 0.01$, compared to HFD (Note: 5 images per section and 3 sections per mice were
 414 randomly selected for statistical analysis, $n = 3$, mean \pm SEM).



415

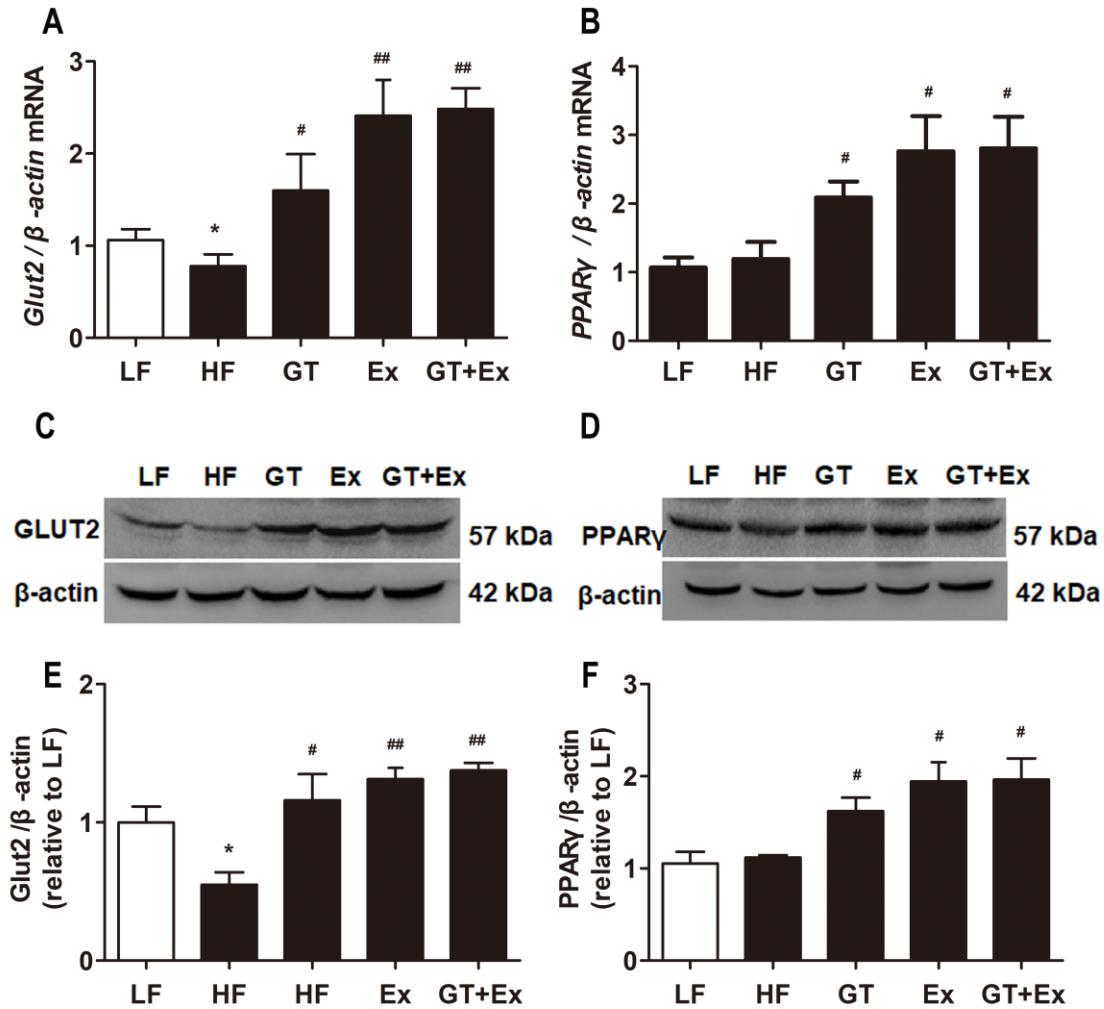
416 Fig. 3. YKGT and exercise inhibited inflammation and NFκB activation in the liver of
 417 HFD C57BL/6J mice. The mRNA expression of *IL-6* (A), *TNFα* (B), *MCP1* (C) in the
 418 liver tissues were quantified by Real Time PCR at 8 and 22 weeks of intervention,
 419 respectively. The representing images of protein expression of total IKKβ and
 420 phosphorylated IKKα/β (D, E), and statistical results of D (F) and E (G); total and
 421 phosphorylated IκBα (H, I), and statistical results of H (J) and I (K); total and
 422 phosphorylated P65 (L, M), and statistical results of L (N) and M (O) in the liver tissues
 423 at 8 and 22 weeks intervention, respectively. *P<0.05, **P<0.01, ***P<0.001, compared
 424 to LFD; and #P<0.05, ##P<0.01, ###P<0.001 compared to HF. NF-κB, nuclear factor
 425 kappa light chain enhancer of activated B cells; IL-6, interleukin 6; TNFα, tumor necrosis
 426 factor α; MCP-1, monocyte chemotactic protein-1 (n=3-6, mean ± SEM).

427



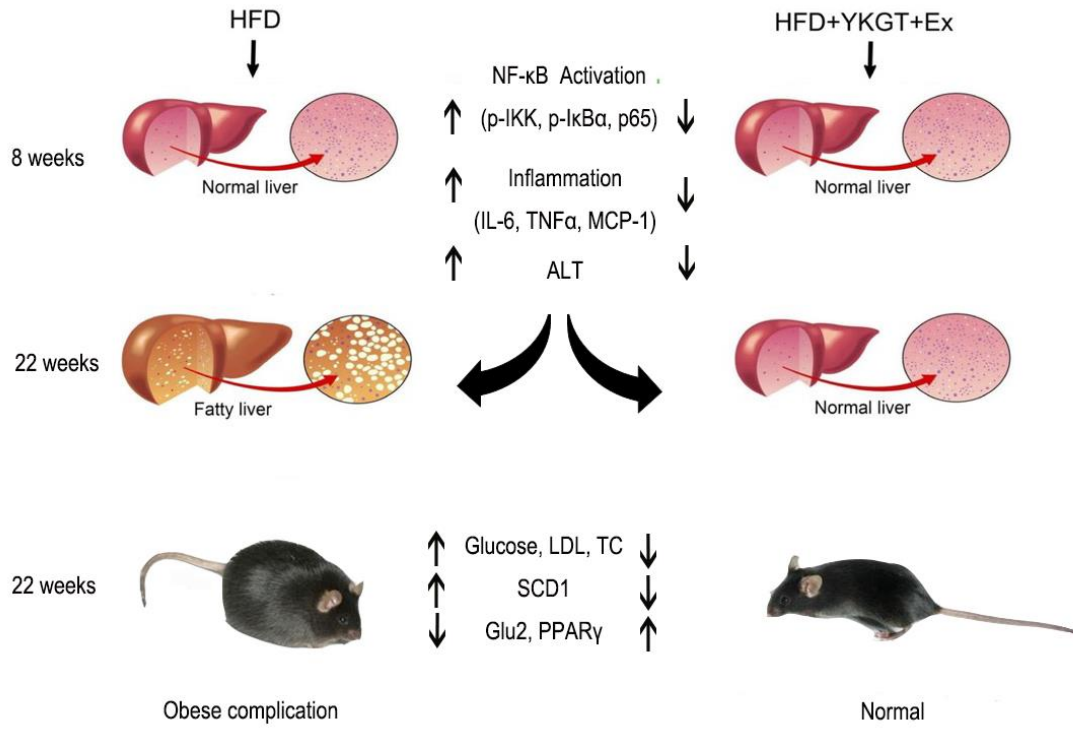
428

429 Fig. 4. YKGT and exercise inhibited SCD1 gene and protein expression in the liver of
 430 HFD C57BL/6J mice. The mRNA expression of *SCD1* (A), representing images of
 431 protein expression of SCD1 (B, C), and statistical results of B (D) and C (E) were
 432 presented in the liver tissues at 8 and 22 weeks intervention, respectively. * $P < 0.05$,
 433 *** $P < 0.001$, compared to LF; and # $P < 0.05$, ## $P < 0.01$, ### $P < 0.001$ compared to HF.
 434 SCD1, Stearoyl CoA desaturase 1 (n=3, mean \pm SEM).



435

436 Fig. 5. YKGT and exercise increased GLU2 and PPAR γ gene and protein expression in
 437 the liver of HFD C57BL/6 mice. The mRNA expression of *GLU2* mRNA (A) and *PPAR γ*
 438 (B), representing images of protein expression of GLUT2 (C) and PPAR γ (D), statistical
 439 results of C (E) and D (F) were presented in the liver tissues at 22 weeks intervention,
 440 respectively. * $P < 0.05$, compared to LF; and # $P < 0.05$, ## $P < 0.01$, compared to HF. GLUT 2,
 441 glucose transporter 2; PPAR γ , peroxisome proliferator-activated receptor γ (n=3, mean \pm
 442 SEM).



443

444 Fig. 6. The schematic diagram for this study.

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447

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