

# Gene mining of immune microenvironment in hepatocellular carcinoma

Zhi-Wei Xu (✉ [XuEmail840701040@qq.com](mailto:XuEmail840701040@qq.com))

Guangxi Medical University

---

## Research Article

**Keywords:** Hepatocellular carcinoma, Immune microenvironment, Bioinformatic analysis

**Posted Date:** May 2nd, 2022

**DOI:** <https://doi.org/10.21203/rs.3.rs-1593173/v1>

**License:**   This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

---

# Abstract

Hepatocellular carcinoma (HCC) is a common malignant tumor worldwide with a poor prognosis. Recent studies have shown that the occurrence, development and prognosis of liver cancer are closely related to tumor microenvironment (TME) and tumor immune infiltration. Therefore, important information on various diseases can be obtained from public databases such as The Cancer Gene Atlas (TCGA), and ideas or schemes that may be effective for the treatment of various diseases can be screened and analyzed by screening various conditions. In this study, 424 cases of liver hepatocellular carcinoma (LIHC) in the TCGA database and CIBERSORT algorithm were used to calculate the proportion of tumor-invasive immune cells. Combined with the clinical data from TCGA database, it was concluded that Tregs were correlated with the development and prognosis of hepatocellular carcinoma. Cox regression analysis was used to screen differentially expressed genes, and survival analysis was performed according to the screened differentially expressed genes to see whether there was a significant association with the prognosis of hepatocellular carcinoma. Then GO and KEGG analysis of differentially expressed genes were carried out to explore the possibility of differentially expressed genes becoming potential therapeutic targets of hepatocellular carcinoma. Finally, I identified the gene CENPO, which is associated with immune cells and improve the prognosis of hepatocellular carcinoma. CENPO may be a potential biological therapeutic target for hepatocellular treatment.

## 1. Introduction

The incidence and mortality of liver cancer are increasing year by year in the world, and liver cancer has become the second highest cause of cancer mortality in the world<sup>1</sup>. At present, hepatocellular carcinoma accounts for 90% of liver cancer. Nonalcoholic steatohepatitis is gradually becoming the most common risk factor leading to hepatocellular carcinoma, and nonalcoholic steatohepatitis has a specific and unique pathogenesis, so I need to find new targeted therapy or immunotherapy to improve the prognosis of liver cancer and hepatocellular carcinoma<sup>2</sup>.

TME is the microenvironment where the tumor is located. It not only contains tumor cells, but also a variety of cells, such as epithelial cells, fibroblasts, vascular endothelial cells, and a large number of immune cells with different functions<sup>3</sup>. Many studies have shown that the change of tumor microenvironment can normalize tumor cells, re culture stromal cells and play an anti-tumor role<sup>4</sup>. At the same time, innate immune cells and adaptive immune cells can interact with tumor cells, change the progress of tumor cells and improve the occurrence, development and prognosis of tumor<sup>5</sup>, for example, studies have shown that Kupffer cells can enhance the response of immune T cells, regulate the growth of tumor cells together with macrophages, and inhibit the development of tumor<sup>6</sup>. In recent years, it has been reported that the prognosis of hepatocellular carcinoma is related to immune cells, and the types and expression of immune cells in hepatocellular carcinoma caused by different etiologies are different<sup>7</sup>. The staging of liver cancer is closely related to its prognosis, which needs scientific staging guidance and treatment plan<sup>8</sup>. Due to the limited treatment options for advanced hepatocellular carcinoma, many

patients are difficult to adapt to the treatment methods of hepatectomy and liver transplantation. Drug immunotherapy is particularly important<sup>9</sup>. Immunotherapy for hepatocellular carcinoma has gradually attracted clinical attention and application<sup>10</sup>.

Therefore, this study attempts to use the LIHC data of TCGA database to explore and study whether there is correlation between immune cells in different stages of hepatocellular carcinoma and whether there are differences in immune cells in different stages of hepatocellular carcinoma, and find out the possible target genes.

## 2. Results

### 2.1 Data and immune cell expression estimation

In this study, I calculated the gene expression RNA sequence data (n = 424) of LIHC data in TCGA database in UCSC Xena with CIBERSORT algorithm, the number of repetitions was 1000, and then extracted the data with high significance, that is, P value < .05. In this way, I roughly estimated the approximate expression data and proportion of 22 immune cells in each patient(Fig. 1A). Then I matched and grouped them by the stage according to the clinical data(n = 469). Finally, it was divided into stage I, stage II and stage III.

### 2.2 Further analysis of immune cells

On this basis, I conducted enrichment Heatmap analysis(Fig. 1B) and correlation analysis(Fig. 1C) on immune cells, and compared them according to the stage of patients, especially stage I and stage III(Fig. 2C). I found that there are several immune cells with high differences: T cells follicular helper, T cells regulatory (Tregs), T cells gamma delta, Macrophages M2, Dendritic cells resting. Therefore, I analyzed the survival of patients based on the expression of significantly different immune cells(Fig. 3). T cells follicular helper(Fig. 3A) and T cells regulatory (Tregs) (Fig. 3B) have obvious differences in different stages of hepatocellular carcinoma. Then, I extracted the expression of these significantly different immune cells in stage I, stage II and stage III, and analyzed their data to obtain these results(Fig. 4). It can be seen from the results that there are significant differences in the prognosis between Tregs and hepatocellular carcinoma stage, and the decrease of Tregs content from stage I to stage III is significant(Fig. 4B), and the impact on survival and prognosis is also significantly different. According to the above situation, I believe that Tregs has a more obvious effect on hepatocellular carcinoma. High content of Tregs is harmful to the survival and prognosis of hepatocellular carcinoma, and low content of Tregs is beneficial to the survival and prognosis.

### 2.3 Screening and determination of differential genes

Therefore, I began to look for differential genes, that is, differential genes that can regulate Tregs, and differential genes have a significant impact on the survival and prognosis of patients with hepatocellular carcinoma. The screening of differential genes is based on the critical value set by log<sub>2</sub> fold change (log<sub>2</sub>

FC) > 1 and Padj < .05. After screening differential genes, I used the survival data in TCGA database (n = 463) for Cox regression analysis, and established a Cox regression analysis model to screen the top gene, in which p-value < .05 and the 95% confidence interval of gene risk ratio does not include 1. Because the high expression of Tregs has a bad effect on the survival and prognosis of hepatocellular carcinoma, the genes I selected are divided into two types. One is the gene of risk factor. When such gene is highly expressed, the expression of Tregs should be increased; When the expression of these genes is low, the expression of Tregs should be reduced, that is, these genes should be positively correlated with Tregs. The other is genes with protective factors. When such genes are highly expressed, Tregs should be lowly expressed; When the expression of such genes is low, the expression of Tregs should be increased, that is, such genes should be negatively correlated with Tregs. So I found the gene CENPO.

## 2.4 Analysis of differential genes

According to Cox regression analysis, gene CENPO is a gene with risk factors, so I drew the forest map according to the results of Cox regression analysis (Fig. 5A). According to the above inference, CENPO should be negatively correlated with the expression of Tregs (Fig. 5B). In addition, I need to know whether CENPO have significant differences in the survival and prognosis of hepatocellular carcinoma. I analyzed their survival data and drew their survival curve (Fig. 5C). In addition to the above studies, the expression content of CENPO in stage is also different, and there is an upward trend from stage I to stage III (Fig. 5D). Therefore, I calculated and plotted the correlation between CENPO and Tregs, whether the expression of the gene is related to the expression of 22 immune cells, and especially the content of Tregs (Fig. 5E). The results show that there was a significant difference between CENPO and the content of Tregs, which was negatively correlated with Tregs, and there was also a significant difference in the survival and prognosis of hepatocellular carcinoma. I used two GEO datasets gse25097 and gse36376 to verify whether CENPO is significantly different from adjacent hepatocellular carcinoma (Fig. 6).

## 2.5 GO and KEGG Enrichment Analysis

In order to better understand the expression of these genes in hepatocellular carcinoma, I conducted GO and KEGG enrichment analysis to analyze the aggregation of CENPO for metabolic pathways and pathways of hepatocellular carcinoma. According to GO analysis, CENPO is mainly enriched in organelle fission, nuclear division and response to xenobiotic stimulus in molecular function, in apical part of cell, synaptic membrane and apical plasma membrane in cellular component, and in channel activity and passive transporter activity in biological process (Fig. 7A). According to KEGG analysis, the pathway enrichment of CENPO on hepatocellular carcinoma is mainly in neuroactive ligand-receptor interaction (Fig. 7B).

## 2.6 Gene Set Enrichment Analysis (GSEA)

In order to explore the enrichment of up-regulated and down-regulated pathways of this gene, I performed GSEA, and I extracted the first five and immune related pathways. The results show that CENPO down regulates these pathways (Fig. 7C, 7D).

## 2.7 PPI Network

Firstly, I use CIBOPORTAL database(<http://www.cbioportal.org/>)<sup>11</sup> to analyze the co-expression genes of CENPO and screen out the genes with more than 0.7 correlation with CENPO. Then I use the string tool to establish the PPI network of these genes to analyze the direct relationship between CENPO and other genes, select the genes known to interact with CENPO and make the PPI network (Fig. 7E). The redder the color, the higher the degree.

## 3. Discussion

In our study, our main purpose is to find the difference of immune cell infiltration in different stages of hepatocellular carcinoma and screen out the differential genes that may regulate highly differential immune cells, so as to improve the survival and prognosis of patients with hepatocellular carcinoma. TME has a very key and important impact on the occurrence and development of tumor, survival and prognosis of patients. Therefore, I need to explore the potential therapeutic targets of tumor in TME, change tumor microenvironment and immune infiltration, and inhibit tumor development<sup>4,12</sup>. A large number of studies have shown that immune microenvironment plays an important role in tumors. Many tumor immune mechanisms have been explored and many immune targets have been found as treatment schemes<sup>13</sup>. I analyzed the RNA SEQ data of LIHC data in TCGA database. The results show that immune cells in TME had an important impact on the survival and prognosis of hepatocellular carcinoma. In recent years, targeted therapy has gradually been applied to the clinical treatment of hepatocellular carcinoma. In recent years, targeted therapy has gradually been applied to the clinical treatment of hepatocellular carcinoma. The application of regorafenib and sorafenib indicates the full opening of the second-line treatment of hepatocellular carcinoma, and more effective new drugs and the most influential immune checkpoint inhibitors(ICIs) have emerged, which have made a great contribution to prolonging the survival of patients with hepatocellular carcinoma.<sup>14</sup> Regulatory T cells (Tregs) is a subgroup of CD4 + T cells, which plays an indispensable role in immune tolerance<sup>15</sup>. Tregs also play a very important role in the tumor microenvironment. The immunosuppressive activity of Tregs is one of the mechanisms to promote tumor development<sup>16</sup>. Tregs in tumor microenvironment not only reduce the efficiency of anti-tumor immune response, but also support the immune escape mechanism of tumor cells and promote the proliferation and development of tumor, so Tregs has become one of the targets of immunotherapy<sup>17</sup>. There is a complex relationship between Tregs and ICIS. The depletion and high content of Tregs make the survival rate of patients low and the prognosis poor. ICIS can induce effector T cells to produce antitumor effect. However, Tregs mediated ICIS is easy to obtain drug resistance, which makes us need to further improve the treatment scheme, reduce or even overcome the drug resistance mechanism, and find new immunosuppressive points<sup>17,18</sup>.

Therefore, I screened the gene centromere protein o (CENPO), also known as mcm21r, icen-36. However, the function of CENPO is not clear<sup>19</sup>. According to the introduction of National Center for Biotechnology Information (NCBI, <https://www.ncbi.nlm.nih.gov/>), this gene is necessary in mitosis and encodes

alternative splicing transcriptional variants of many protein subtypes. It is reported that CENPO can promote the proliferation of gastric cancer and has a negative impact on the prognosis of gastric cancer<sup>20</sup>. Studies have also shown that upregulation of CENPO may lead to bladder cancer(BC) and may depend on P53 to regulate the development, proliferation and apoptosis of colorectal cancer<sup>21,22</sup>. Studies have suggested that the high expression of CENPO has an adverse effect on the survival of breast cancer and leads to poor prognosis. It is also considered that CENPO is an independent factor affecting the distant recurrence survival rate (DRFS) of breast cancer patients<sup>23</sup>. The role of this gene CENPO is gradually being developed and explored by researchers, and seems to be related to mental diseases<sup>24-26</sup>. In this study, I found that CENPO is also closely related to the prognosis of hepatocellular carcinoma. CENPO has a positive correlation with Tregs in the tumor microenvironment of hepatocellular carcinoma, and high expression of CENPO increases the content of Tregs, which can be consumed to make the tumor produce anti immunity or immune escape, and promote the development of hepatocellular carcinoma. Moreover, the survival rate of hepatocellular carcinoma with high expression of CENPO is low and the prognosis is poor.

The data in this study only come from TCGA database and GEO database, and the data analysis only exists at the theoretical level, which cannot be verified in animal model or cell experiment. I hope that the verification can be completed in the future.

## 4. Conclusion

In conclusion, this is a preliminary study on the survival and prognosis of hepatocellular carcinoma based on TCGA database. In this study, I searched and analyzed the effect of gene CENPO on hepatocellular carcinoma, which may be a potential target for the treatment of hepatocellular carcinoma. I hope to have the opportunity to use this base in the future, because the research direction can experimentally verify the impact on the prognosis of hepatocellular carcinoma.

## 5. Materials And Methods

### 5.1 Data Collection and Processing

The TCGA database is a very large genomics database with multiple cancer types which has genomics data of various cancers and genomics data of normal samples matched with corresponding cancers (<https://portal.gdc.cancer.gov/>)<sup>27</sup>. In this study, gene expression RNA-seq data(n = 424), clinical information(n = 469) and survival data(n = 463) were gained from TCGA database with level 3, which was downloaded through the University of California Santa Cruz Xena (UCSC Xena) (<http://xena.ucsc.edu/>)<sup>28</sup>. CIBERSORT algorithm<sup>29</sup> was used to calculate the transcriptome RNA sequence data, and the number of repetitions was 1000. I extracted highly significant data with  $P < .05$ , matched them with clinical data, grouped them according to the situation of stage, and then drew the landscape histogram of immune cells in different stages through the program package ggplot2 (version 3.3.5) of R software(version 4.1.2).

## 5.2 Screening of differential genes

I divided RNA SEQ data into tumor data and normal data, and calculated and screened them through `deseq2` package<sup>30</sup> (version 1.34.0) of R software. The screening conditions are  $\log_2$  fold change ( $\log_2FC$ )  $> 1$  and  $p_{adj} < .05$ . In this way, the screened differential genes can be considered to be highly significant and differential. Subsequently, I tested whether the genes I screened are different through two data (GSE25097, GSE36376) in The Gene Expression Omnibus (GEO, <http://www.ncbi.nlm.nih.gov/geo/>)<sup>31</sup> database.

## 5.3 Survival Analysis

I use log rank test to calculate the survival curve of the screened differential genes related to TME. The platform is R software, the program package is `survival` (version 3.2–13), and the set threshold is  $p < .05$ , so I can think that the differential genes have differences in the survival and prognosis of hepatocellular carcinoma. Survival data ( $n = 363$ ) is the survival data after screening and removing the paracancerous data. The pictures of Kaplan Meier survival analysis are also produced by R software.

## 5.4 Cox regression analysis

I established a univariate Cox regression model to analyze the survival data, calculated the risk ratio of the screened differential genes, and further screened the top gene on this basis. The threshold of the top gene was set as  $p_{cutoff} < .05$ , and the 95% confidence interval of the risk ratio was one side of 1, that is, between 0–1 or greater than 1. After selecting the top gene, I can make the forest map of gene risk ratio according to the calculated data. The platform is R and the program package is `forestplot` (version 2.0.1).

## 5.5 Heatmaps

The correlation heat map and clustering heat map of immune cells are also drawn by R software, and the program packages are `corrplot` (version 0.92) and `heatmap` (version 1.0.12) respectively. The correlation heat map of immune cells is calculated by Spearman correlation analysis algorithm.

## 5.6 GO and KEGG Enrichment Analysis

I will conduct GO, KEGG enrichment analysis on the finally screened genes. The platform is R and the program package is `org.Hs.Eg.db` (version 3.14.0), `clusterProfiler`<sup>32</sup> (version 4.2.2), `enrichplot` (version 1.14.2) and `ggplot2`. GO analysis reveals the main functions of genes in biological processes, cell components and molecular functions. KEGG analysis mainly analyzes the enrichment pathway of genes. I divided the expression of CENPO into high expression and low expression. After calculation,  $P$  value  $< .05$  was used as the screening threshold.

## 5.7 Gene Set Enrichment Analysis(GSEA)

GSEA uses a predefined gene set to sort the genes according to the degree of differential expression, and then tests whether the gene set is enriched at the top or bottom of the sorting table. I will conduct GSEA on the finally screened genes. The platform is R and the program package is org.Hs.Eg.db, clusterProfiler and enrichplot. I use C5 gene sets v7.0 collections. I calculated as the target gene set, with p-value < .05 and q-value < .05 as significant.

### 5.8 Violin Plot and Box Plot

The comparison of 22 immune cells shown in the violin diagram is calculated by Wilcoxon rank sum algorithm. The comparison of gene expression is divided into low expression and high expression based on the median of gene expression. It is also calculated by Wilcoxon rank sum algorithm. The above drawing and calculation are carried out on the R software platform, and the program package is vioplot (version 0.3.7). Box plot is also calculated and drawn by Wilcoxon rank sum algorithm on R software.

### 5.9 PPI Network Construction

The PPI network of the screened genes was obtained through the search tool for the retrieval of interacting genes (STRING, <https://string-db.org/>)<sup>33</sup> and imported into Cytoscape software<sup>34</sup> (version 3.9.1) for reconstruction.

### 5.10 Scatter diagram

The scatter diagram is drawn on the platform of GraphPad Prism (version 8.0.2), and the statistical method is nonparametric test.

## Declarations

### Data Availability

The datasets generated and analyzed during the current study are available in the TCGA database (<https://portal.gdc.cancer.gov/>) or UCSC Xena (<http://xena.ucsc.edu/>) and GEO database (<http://www.ncbi.nlm.nih.gov/geo/>).

### Conflicts of Interest

The author declare that there are no conflicts of interest regarding the publication of this article.

### Authors' Contributions

The author has read and approved the manuscript for publication.

### Ethics and consent to participate

Not Applicable



## Funding

Not Applicable

## References

1. Cao W, Chen HD, Yu YW, Li N, Chen WQ. Changing profiles of cancer burden worldwide and in China: a secondary analysis of the global cancer statistics 2020. *Chin Med J (Engl)*. Mar 17 2021;134(7):783–791. doi:10.1097/CM9.0000000000001474
2. Llovet JM, Kelley RK, Villanueva A, et al. Hepatocellular carcinoma. *Nat Rev Dis Primers*. Jan 21 2021;7(1):6. doi:10.1038/s41572-020-00240-3
3. Bolouri H. Network dynamics in the tumor microenvironment. *Semin Cancer Biol*. Feb 2015;30:52–9. doi:10.1016/j.semcancer.2014.02.007
4. Quail DF, Joyce JA. Microenvironmental regulation of tumor progression and metastasis. *Nature medicine*. Nov 2013;19(11):1423–37. doi:10.1038/nm.3394
5. Hinshaw DC, Shevde LA. The Tumor Microenvironment Innately Modulates Cancer Progression. *Cancer Res*. Sep 15 2019;79(18):4557–4566. doi:10.1158/0008-5472.CAN-18-3962
6. Sheng J, Zhang J, Wang L, et al. Topological analysis of hepatocellular carcinoma tumour microenvironment based on imaging mass cytometry reveals cellular neighbourhood regulated reversely by macrophages with different ontogeny. *Gut*. Jul 12 2021;doi:10.1136/gutjnl-2021-324339
7. Inada Y, Mizukoshi E, Seike T, et al. Characteristics of Immune Response to Tumor-Associated Antigens and Immune Cell Profile in Patients With Hepatocellular Carcinoma. *Hepatology*. Feb 2019;69(2):653–665. doi:10.1002/hep.30212
8. Forner A, Diaz-Gonzalez A, Llicioni A, Vilana R. Prognosis prediction and staging. *Best Pract Res Clin Gastroenterol*. Oct 2014;28(5):855–65. doi:10.1016/j.bpg.2014.08.002
9. Johnston MP, Khakoo SI. Immunotherapy for hepatocellular carcinoma: Current and future. *World J Gastroenterol*. Jun 28 2019;25(24):2977–2989. doi:10.3748/wjg.v25.i24.2977
10. Llovet JM, Castet F, Heikenwalder M, et al. Immunotherapies for hepatocellular carcinoma. *Nat Rev Clin Oncol*. Mar 2022;19(3):151–172. doi:10.1038/s41571-021-00573-2
11. Cerami E, Gao J, Dogrusoz U, et al. The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. *Cancer Discov*. May 2012;2(5):401–4. doi:10.1158/2159-8290.CD-12-0095
12. Bejarano L, Jordao MJC, Joyce JA. Therapeutic Targeting of the Tumor Microenvironment. *Cancer Discov*. Apr 2021;11(4):933–959. doi:10.1158/2159-8290.CD-20-1808
13. Hao X, Sun G, Zhang Y, et al. Targeting Immune Cells in the Tumor Microenvironment of HCC: New Opportunities and Challenges. *Front Cell Dev Biol*. 2021;9:775462. doi:10.3389/fcell.2021.775462
14. Huang A, Yang XR, Chung WY, Dennison AR, Zhou J. Targeted therapy for hepatocellular carcinoma. *Signal Transduct Target Ther*. Aug 11 2020;5(1):146. doi:10.1038/s41392-020-00264-x

15. Sakaguchi S, Yamaguchi T, Nomura T, Ono M. Regulatory T cells and immune tolerance. *Cell*. May 30 2008;133(5):775 – 87. doi:10.1016/j.cell.2008.05.009
16. Churov A, Zhulai G. Targeting adenosine and regulatory T cells in cancer immunotherapy. *Hum Immunol*. Apr 2021;82(4):270–278. doi:10.1016/j.humimm.2020.12.005
17. Sasidharan Nair V, Elkord E. Immune checkpoint inhibitors in cancer therapy: a focus on T-regulatory cells. *Immunology and cell biology*. Jan 2018;96(1):21–33. doi:10.1111/imcb.1003
18. Saleh R, Elkord E. Treg-mediated acquired resistance to immune checkpoint inhibitors. *Cancer Lett*. Aug 10 2019;457:168–179. doi:10.1016/j.canlet.2019.05.003
19. Izuta H, Ikeno M, Suzuki N, et al. Comprehensive analysis of the ICEN (Interphase Centromere Complex) components enriched in the CENP-A chromatin of human cells. *Genes Cells*. Jun 2006;11(6):673–84. doi:10.1111/j.1365-2443.2006.00969.x
20. Cao Y, Xiong J, Li Z, et al. CENPO expression regulates gastric cancer cell proliferation and is associated with poor patient prognosis. *Mol Med Rep*. Oct 2019;20(4):3661–3670. doi:10.3892/mmr.2019.10624
21. Liu Y, Xiong S, Liu S, et al. Analysis of Gene Expression in Bladder Cancer: Possible Involvement of Mitosis and Complement and Coagulation Cascades Signaling Pathway. *J Comput Biol*. Jun 2020;27(6):987–998. doi:10.1089/cmb.2019.0237
22. Liu Z, Chen C, Yan M, Zeng X, Zhang Y, Lai D. CENPO regulated proliferation and apoptosis of colorectal cancer in a p53-dependent manner. *Discov Oncol*. Feb 3 2022;13(1):8. doi:10.1007/s12672-022-00469-2
23. Zhang S, Xie Y, Tian T, et al. High expression levels of centromere protein A plus upregulation of the phosphatidylinositol 3-kinase/Akt/mammalian target of rapamycin signaling pathway affect chemotherapy response and prognosis in patients with breast cancer. *Oncol Lett*. May 2021;21(5):410. doi:10.3892/ol.2021.12671
24. Trampush JW, Yang ML, Yu J, et al. GWAS meta-analysis reveals novel loci and genetic correlates for general cognitive function: a report from the COGENT consortium. *Mol Psychiatry*. Mar 2017;22(3):336–345. doi:10.1038/mp.2016.244
25. Hewitt CA, Ling KH, Merson TD, et al. Gene network disruptions and neurogenesis defects in the adult Ts1Cje mouse model of Down syndrome. *PLoS One*. Jul 16 2010;5(7):e11561. doi:10.1371/journal.pone.0011561
26. Herve M, Bergon A, Le Guisquet AM, et al. Translational Identification of Transcriptional Signatures of Major Depression and Antidepressant Response. *Front Mol Neurosci*. 2017;10:248. doi:10.3389/fnmol.2017.00248
27. Tomczak K, Czerwinska P, Wiznerowicz M. The Cancer Genome Atlas (TCGA): an immeasurable source of knowledge. *Contemp Oncol (Pozn)*. 2015;19(1A):A68-77. doi:10.5114/wo.2014.47136
28. Goldman MJ, Craft B, Hastie M, et al. Visualizing and interpreting cancer genomics data via the Xena platform. *Nature biotechnology*. Jun 2020;38(6):675–678. doi:10.1038/s41587-020-0546-8

29. Newman AM, Liu CL, Green MR, et al. Robust enumeration of cell subsets from tissue expression profiles. *Nat Methods*. May 2015;12(5):453–7. doi:10.1038/nmeth.3337
30. Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome biology*. 2014;15(12):550. doi:10.1186/s13059-014-0550-8
31. Barrett T, Wilhite SE, Ledoux P, et al. NCBI GEO: archive for functional genomics data sets–update. *Nucleic acids research*. Jan 2013;41(Database issue):D991-5. doi:10.1093/nar/gks1193
32. Yu G, Wang LG, Han Y, He QY. clusterProfiler: an R package for comparing biological themes among gene clusters. *Omics: a journal of integrative biology*. May 2012;16(5):284–7. doi:10.1089/omi.2011.0118
33. Szklarczyk D, Gable AL, Lyon D, et al. STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. *Nucleic acids research*. Jan 8 2019;47(D1):D607-D613. doi:10.1093/nar/gky1131
34. Doncheva NT, Morris JH, Gorodkin J, Jensen LJ. Cytoscape StringApp: Network Analysis and Visualization of Proteomics Data. *J Proteome Res*. Feb 1 2019;18(2):623–632. doi:10.1021/acs.jproteome.8b00702

## Figures

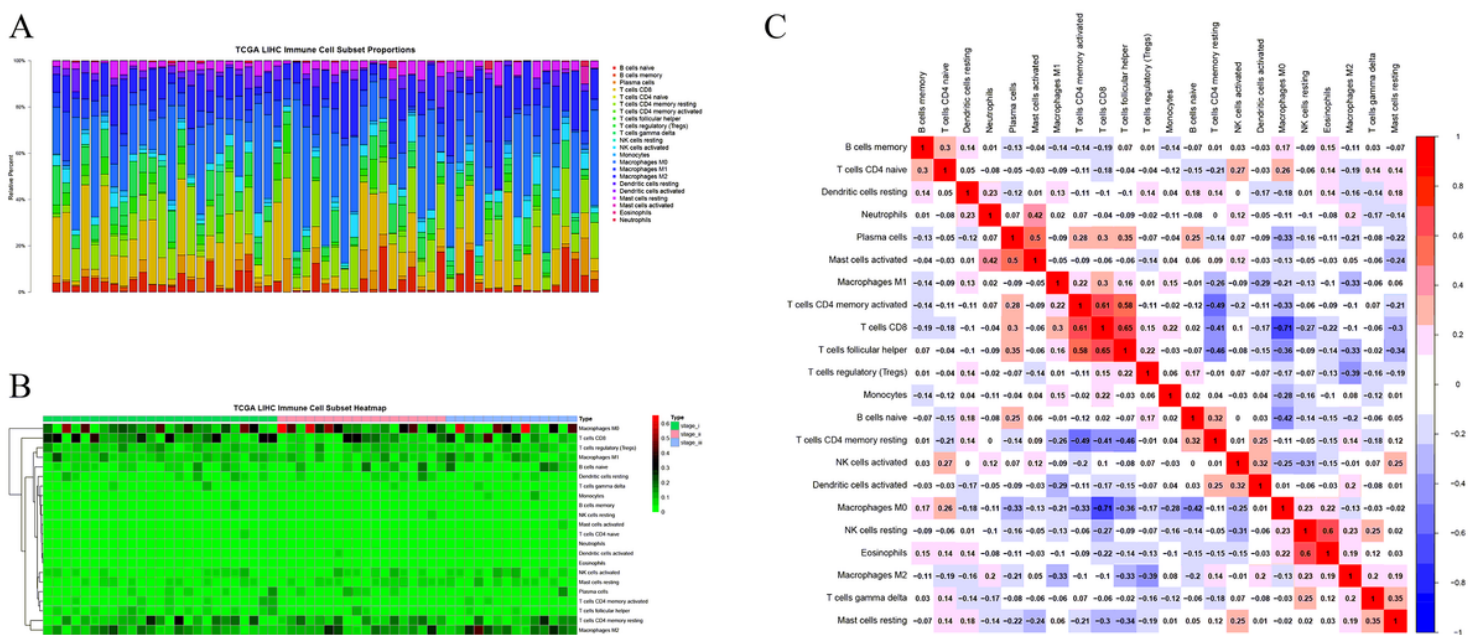
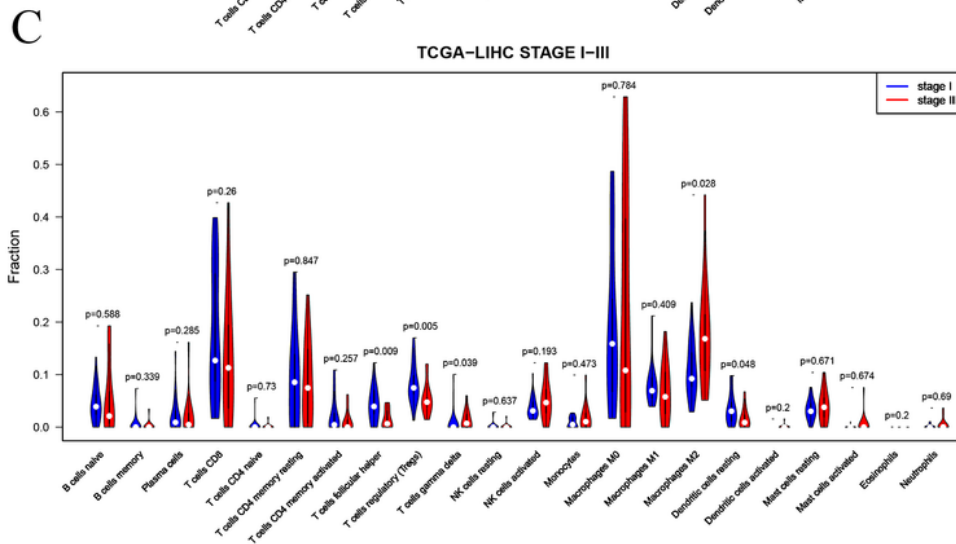
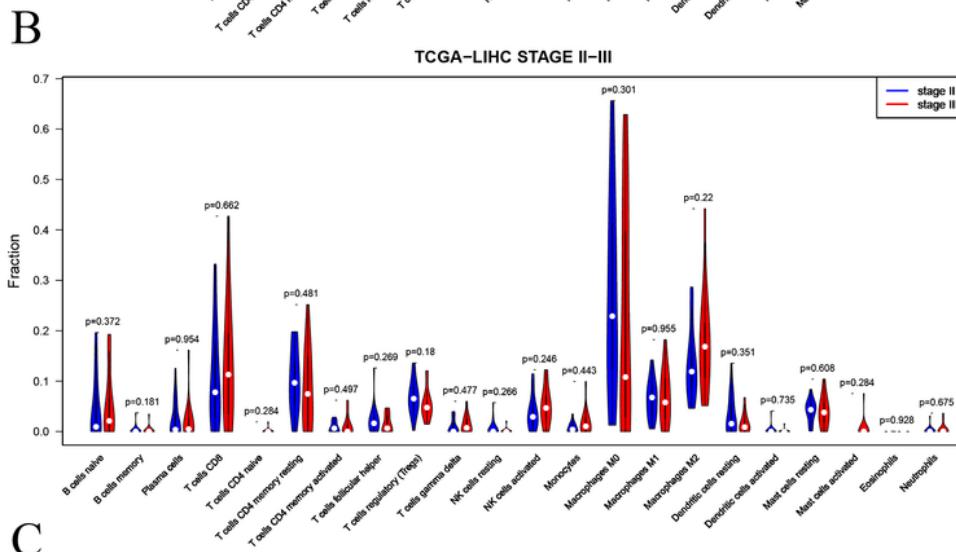
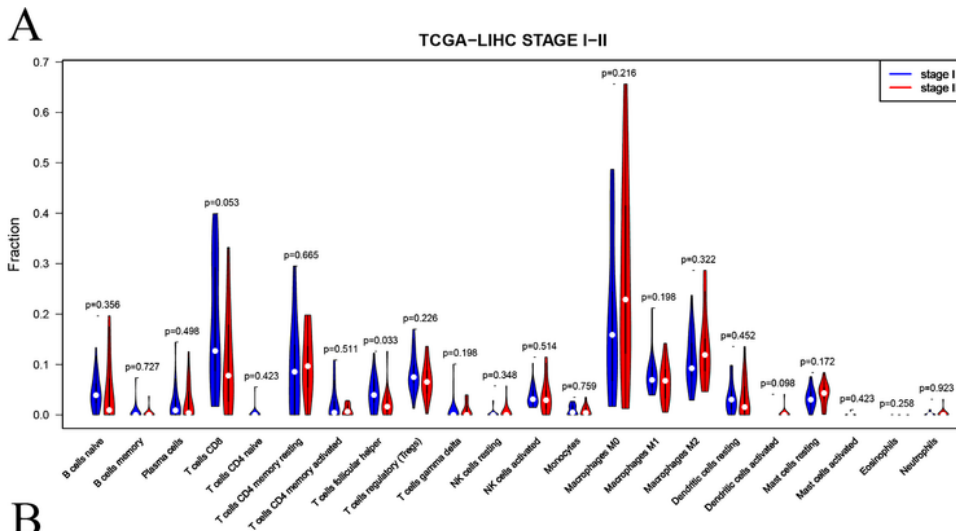


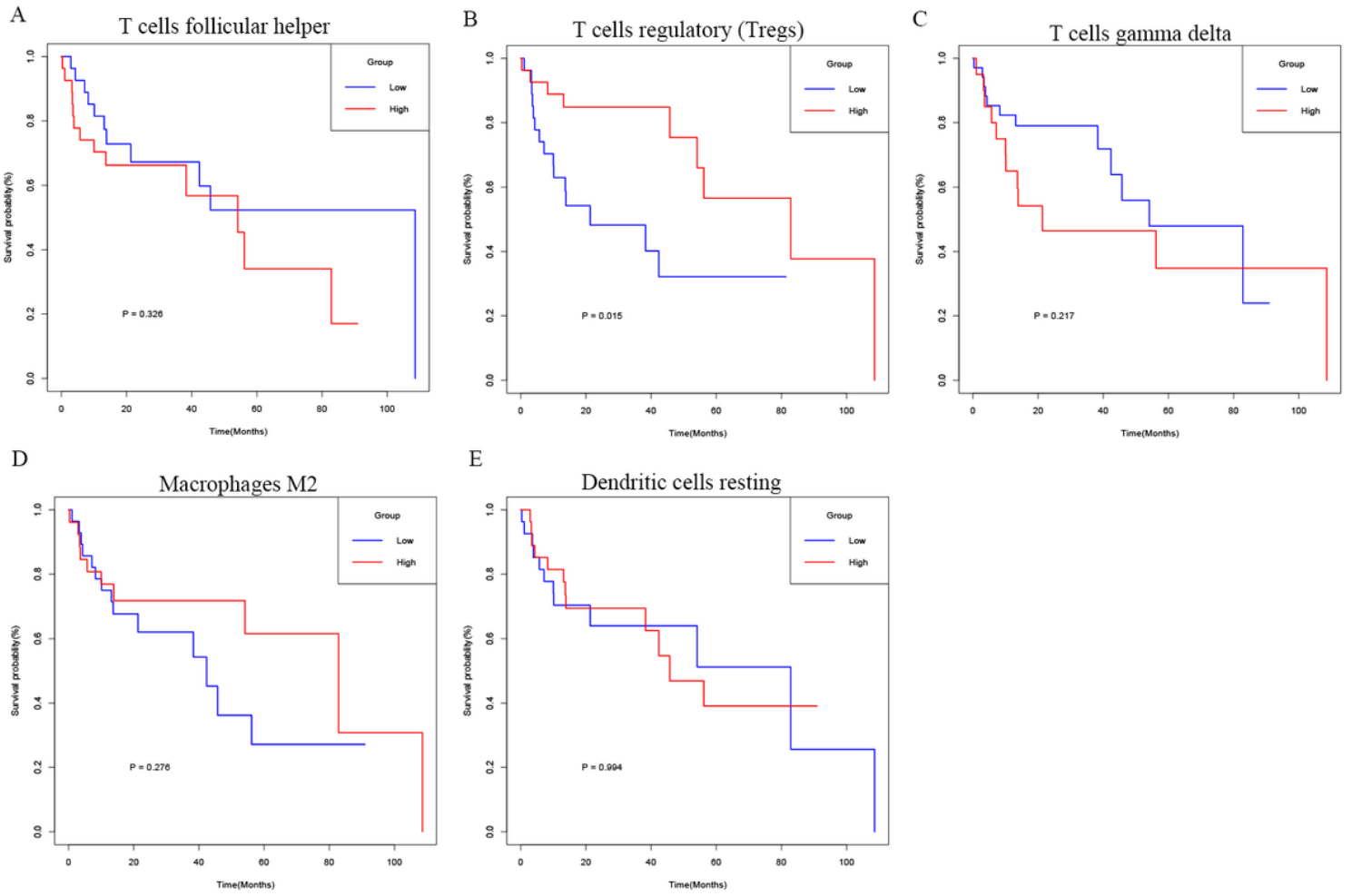
Figure 1

Legend not included with this version.



**Figure 2**

Legend not included with this version.



**Figure 3**

Legend not included with this version.

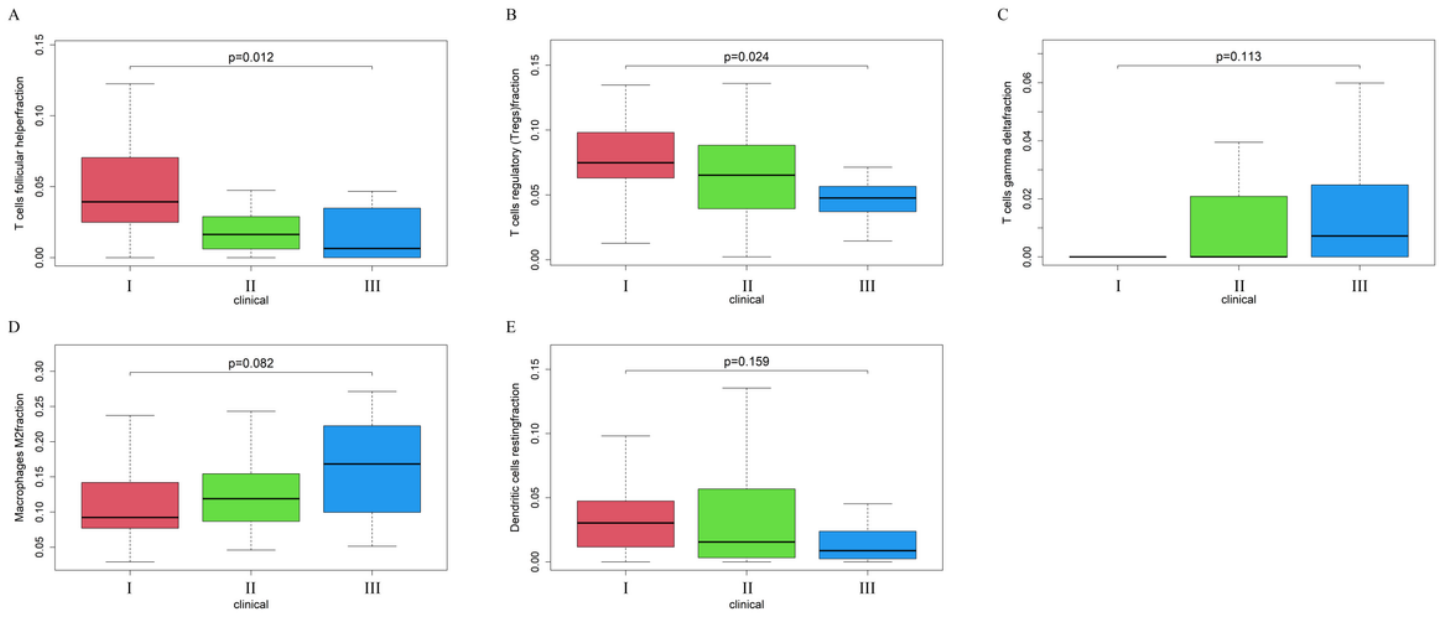
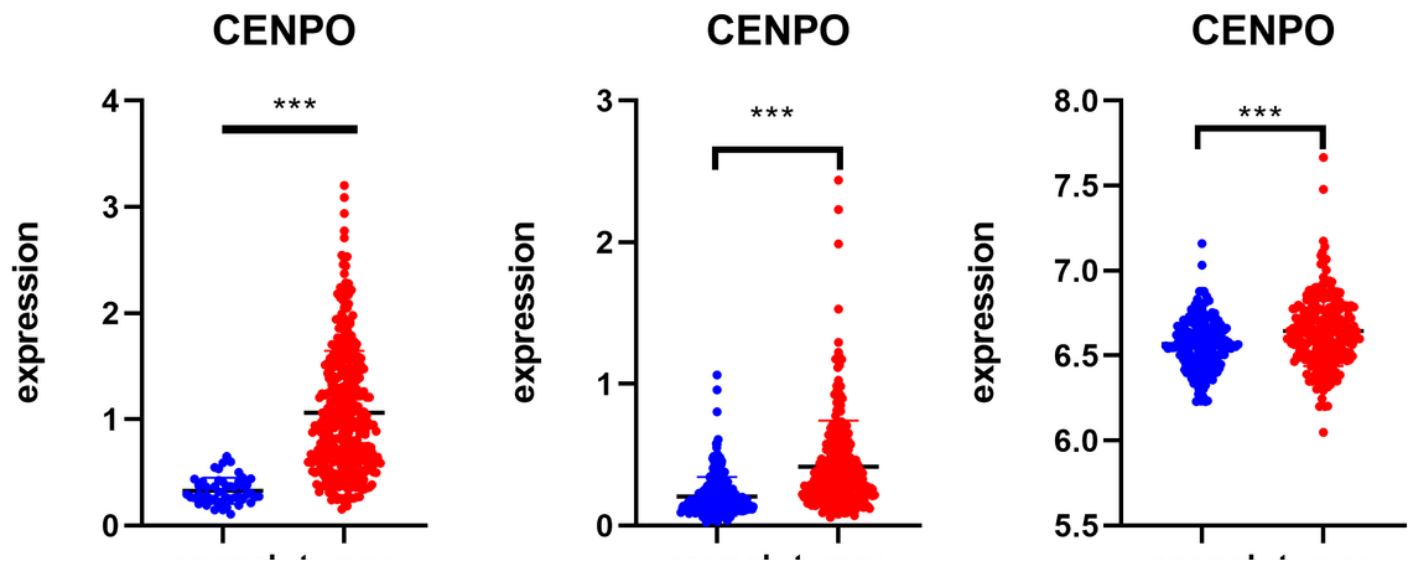


Figure 4

Legend not included with this version.

Figure 5

Legend not included with this version.



## **Figure 6**

Legend not included with this version.

## **Figure 7**

Legend not included with this version.