

Identification of QTLs and Candidate Loci Associated with Drought-Related Traits of the K/Z RIL Rice Population

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Abstract

Background: Rice is the main staple food for the global population and drought is one of the limited factor in rice production. In this research, progeny of a cross between an adapted U.S. rice cultivars with a *tropical japonica* and an *indica* rice genotype, were screened for drought resistant (DR) traits to identify DR loci, that would be useful for breeding U.S. rice cultivars for a water saving agricultural system.

Results: A recombinat inbred line (RIL) population, generated from selfed progeny of the cross between the drought resistant *tropical japonica* U.S. cultivar Kaybonnet and an *indica* drought sensitive cultivar ZHE733, was chosen for quantitative trait locus (QTLs) analysis of drought-resistance related traits. The DR traits were quantified by measuring different parameters of morphological traits, grain yield components and root architectural traits. K/Z RIL population of 198 lines were screened in the field at Fayetteville (AR), by giving controlled drought stress (DS) and well-watered (WW) treatment at the reproductive stage, consequently for three years and the effects of DS were quantify by measuring morphological traits and grain yield components. The effect of abscisic acid (ABA) sensitivity screen on parents and 198 lines at the V3 stage in culture media was quantified by measuring root architectural traits. QTL analysis was performed with a set of 4133 single nucleotide polymorphism (SNP) markers by using QTL IciMapping software version 4.2.53. A total of 41 QTLs and 184 candidate genes within the DR-QTL regions were identified for drought related traits. The potential candidate genes were validated by RT-qPCR of parental lines. The results of candidate DR genes revealed that the gene expression of 15 candidate DR genes with known annotations, and two candidate DR genes with unknown annotations within the DR-QTL regions were up-regulated in the drought resistant parent (Kaybonnet) compared to the drought sensitive parent (ZHE733) under DS conditions.

Conclusions: In this study, we detected 41 QTLs and 184 candidate genes within the DR-QTL regions, and most of the candidate genes were up-regulated in Kaybonnet as the drought resistant parent. The findings of this research provide important information to develop drought-resistant rice varieties with greater productivity under DS conditions.

Background

Rice (*Oryza sativa*) is one of the nutritional and commercially productive cereal crops providing the principal food for approximately 2.5 billion people world-wide, and a model species for monocot and cereal plants with a compact diploid genome size of around 500 Mb, with 12 chromosomes (Edwards and Batley, 2010). The primary rice-producing countries are China, India, Indonesia, Bangladesh, Vietnam, Thailand, Myanmar, Philippines, Brazil, Japan, U.S., Pakistan, and the Republic of Korea (FAOSTAT, 2018). Furthermore, U.S. is the third largest exporter of rice with Arkansas as the biggest rice producer of long and medium grain varieties (Quick Stats, 2016).

Based on several statistical estimates and surveys, the next 20 years have been predicted to be important for the global rice production to be increased by 30% to meet the food demands from the

increasing world population and economic development (Khush, 2001). Currently, the rate of global population growth exceeds the rate of the increase in food production. In addition, drought has become the most crucial constraint in rice production due to global climate change, and the competition with urban and industrial users of the limited water available (Tuong and Bouman, 2003; Farooq et al., 2009). Around 50% of the total rice area globally is affected by drought. Around 130 million ha of rice field in Asia are annually affected by drought and predicted to become more frequent in many areas in the future (Rahimi et al., 2013). When compared to the current drought levels, future drought could reduce rice production even more (Zhao et al., 2017). DS mainly affects physiological, morphological, and molecular-level factors of the rice crop plants (Ito et al., 1999). All of the rice growth stages; seedling, vegetative, and reproductive are affected by drought, with the reproductive stage being the most sensitive to DS conditions (Bunnag and Pongthai, 2013; Hsiao, 1982; O'Toole, 1982). Moreover, DS conditions at the reproductive stage cause a significant reduction in the grain yield components such as spikelet per panicle number, panicle length, primary panicle branch number, filled grain per panicle number, and hundred grain weight, all leading to a decrease in grain yield per plant (Sadeghi and Danesh, 2011).

A previous study reported that DS conditions at the reproductive stage might reduce the grain yield up to 77% (Ito et al., 1999). On the other hand, the production of 1 kg of rice needs 3,000-5,000 liters of water, which is 3 times more than other cereal crops like maize and wheat (Bouman, 2002). Annually, the total rice yield loss due to DS conditions is around 18 million tons. However, the use of advanced genomics technologies with high-quality rice genome sequence information is very useful to develop drought-resistant rice genotypes that perform better under DS conditions. The identification of quantitative trait loci (DR-QTL) regulating grain yield under DS conditions, can be done by employing genome information of several drought-resistant rice genotypes such as Vandana, Nagina-22 (N22), Bengal, and Kaybonnet in a drought molecular-breeding program (Dixit et al., 2014; Venuprasad et al., 2009; Kumar et al., 2017). Development of a drought-resistant rice genotype can help to increase yield production and stability for ensuring food security.

QTLs are chromosomal segments that encode genes for quantitative traits, whose effects are determined by making quantitative measurements. These quantitative traits are controlled by one or more genes and affected by environmental variation, with phenotypes such as plant height, grain yield, abiotic and biotic stress. QTLs are defined and mapped using molecular markers. This mapping method is very affordable to plant research programs because of the developments in genomic technology and statistical analysis methods (Zhu et al., 2008; Dwiningsih et al., 2021a). Single nucleotide polymorphisms (SNPs) are widely used DNA markers to identify QTLs for important traits that can effectively speed up plant breeding. These SNPs are characterized as the most abundant variation in rice genomes that are very useful for high-resolution genotyping and to produce the highest resolution maps (McCouch et al., 2010). Additionally, the use of SNPs are more efficient and cost effective (Edwards and Batley, 2010; Dwiningsih et al., 2020a). SNPs become the most popular DNA markers in the 21st century due to the development of genotyping by sequencing (GBS) techniques (Thomson, 2014). QTL analysis has been used to obtain genomic information about many agronomic traits. Genomic information from QTL analysis is therefore

very useful for enhancing plant breeding programs through marker-assisted selection (MAS). In the past few decades, QTL mapping for agronomic and physiological traits under abiotic stress conditions, has resolved several problems. A number of QTLs associated with drought resistant (DR) traits have been identified in *Oryza sativa* (Mardani et al., 2013). Furthermore, DR-QTL mapping has been very useful to identify the genes and chromosomal segments associated with complex DR traits.

The development of drought-resistant rice genotypes has been slow because of the genetic complexity that controls grain yield traits under drought and also the high genotype-environment (GxE) interaction associated with these traits (Barnabás et al., 2008). However, several studies at IRRI have reported that development of mapping populations derived from a drought-resistant variety and a high-yielding variety has proven effective in combining drought resistance with high yield potential. These mapping populations have also shown transgressive segregants with higher yield compared to the parents under drought and normal conditions.

Many genes related to drought stress resistance in rice have been identified by using several methods such as Expressed Sequence Tag (EST) profiling, transcript profiling via massively parallel signature sequencing (MPSS), microarrays and quantitative real time PCR, and RNA gel blot analysis (Rabello, et al., 2008), and also comparative proteome analysis (Xiong et al., 2010). On the other hand, only few genes have been functionally validated for their drought resistance ability in rice (Sahoo et al., 2013). Important examples include the stress-responsive rice NAC genes like SNAC1, OsNAC6/SNAC2, and OsNAC5 which enhance drought resistance when over-expressed (Nakashima et al., 2014).

The plant hormone abscisic acid (ABA) plays an important role in adaptive responses to drought stress conditions, by regulating stomatal closure to limit water loss through transpiration (Schroeder et al., 2001 and Finkelstein et al., 2002). Endogenous ABA concentration increase under drought stress conditions, for helping plants adapt to the water deficit (Xiong et al., 2002). In addition, ABA controls the expression of many drought-responsive genes involved in a protective response (Shinozaki and Yamaguchi-Shinozaki, 2007). Several transcription factors involved in the regulation of ABA-responsive gene expression include ABFs/AREBs (Kim, 2006), CBF/DREB, MYB, NAC, and WRKY (Berri et al., 2009; Yamaguchi-Shinozaki and Shinozaki, 2005).

The objective of this research is identification of drought resistance QTLs and candidate genes in the K/Z RIL population under both drought and well-watered conditions mainly measured with morphological traits, grain yield components, and root architectural traits changes related to ABA response. The QTLs showing high logarithm of odd (LOD) and phenotypic variation explanation (PVE) were further validated in the parental lines by using gene expression analysis. Identification of QTLs from the two contrasting cultivars for drought-related traits will contribute to our understanding of the genetic control of rice productivity at the sensitive reproductive stage under drought stress conditions, and lead to accelerate the development of drought-resistant rice varieties with improved grain yield under drought stress conditions.

Methods

Plant materials

A recombinant inbred line (RIL) population derived from the varieties Kaybonnet (*Oryza sativa*, an upland *japonica* type) and ZHE733 (*Oryza sativa*, an *indica* type), K/Z RILs (USDA, Stuttgart, Arkansas, U.S.). A total of 198 RILs from an F₂ population was generated by single seed descent (SSD), after selfing for F₁₀ generations and was used to study various morphological traits, grain yield components under well-watered (WW) and drought stress (DS) conditions, and also the response of ABA on root architectural traits.

Drought stress treatment at the reproductive stage

Seed of the K/Z RIL population and two parents (Kaybonnet and ZHE733) were germinated and grown in the greenhouse and uniform plants were transplanted to the field separately in 6 batches (at 7-day intervals) based on their heading day data from USDA to synchronize the drought treatment at the reproductive stage. This RIL population was evaluated in the field at Fayetteville, AR, U.S. of the growing seasons (May-November) in 2016, 2017 and 2018 with average annual rainfall of 849.63 mm. The population was grown in a randomized complete block design with five replications and two treatments of well-watered (WW) and drought stress (DS) conditions. The DS treatment was given at the reproductive stage (R3). The DS conditions were maintained continuously up to -70 kPa (severe stress) using tensiometer and the WW plot was watered continuously. The effect of drought stress was quantified by measuring morphological traits and grain yield components such as heading day (HD), plant height (PH), productive tiller number (TN), flag leaf width (FLW), leaf rolling score (LR), spikelet per panicle number (SP), panicle length (PL), primary panicle branch number (PPB), filled grain per panicle number (FG), hundred grain weight (HGW), and biological yield per plant (BY) with five replications per line. The data under WW and DS conditions for morphological traits and grain yield components were analyzed by analysis of variance (ANOVA) and the Tukey's HSD was performed to compare the means of the two treatments (WW and DS) (Tukey's HSD, $P < 0.05$) using JMP version 12.0. Shapiro-Wilk test was used to test a normal distribution for each trait by using SAS 9.4.

Screening for ABA sensitivity

A total of 2 parental genotypes and 198 recombinant inbred lines seeds were sterilized with 70% ethanol for 60 second and 30% bleach solution for 45 minutes and rinsed four times with sterilized water. Sterilized seeds (S0 stage) were germinated in 2 ml tubes contained germination media (Chu's N-6 Basal Salts with Vitamins, Macronutrients, Micronutrients) until S3 stage in the growth chamber (temperature: 28/22°C day/night, light intensity: 600 $\mu\text{mol}/\text{m}^2/\text{s}$, relative humidity: 60%) for uniform seedling growth. The seedlings at S3 stage of Kaybonnet, ZHE733, and 198 lines were transplanted into ABA media at different concentration levels: 0 μM (control), 3 μM , and 5 μM , then grown in the growth chamber until V3 stage. The effect of ABA sensitivity was quantified by measuring the root architectural traits: maximum root length (RL), root to shoot ratio (RSR), total root number (TRN), number of roots with

a shallow angle (0-45°) (SRN), number of roots with a deep angle (45-90°) (DRN), and root fresh weight (RFW) with five replications per line/treatment.

Measurements of Root Anatomical Phenotypes

Root samples from V3 stage of the parental genotypes (Kaybonnet and ZHE733), 18 drought-resistant lines, and 18 drought-sensitive lines were measured for root anatomical phenotypes with five biological replications. About 10 cm section from middle portion of primary root was used for fixing. For fixation, root segments were submerged in FAA (Formaldehyde Alcohol Acetic Acid, 10%:50%:5% + 35% water) solution for at least 24 hours at 4°C followed by dehydration in a gradation of ethanol series of 70%, 80%, 85%, 95%, and then 100% for an hour each. Dehydrated samples were then treated with toluene two times for an hour each and then embedded in paraffin. The embedded samples were cut 80–250µm with Microtome (Accu-Cut SRM 200 Rotary Microtome, Sakura Finetek USA, Inc., California, USA), the root sections were placed in the microscope slides and dried in the forced air dryer at 37°C for overnight. The root sections were stained with 0.05% toluidine blue for 3 minute and de-stained with distilled water, again air dried for 2 minutes and finally embedded in xylene. Root sections images were captured by using digital camera (Optiphot 2 Nikon and a D1 digital camera Nikon) mounted on microscope with 40x magnification. Root anatomical traits: metaxylem, aerenchyma, and cortical cells properties were measured using *RootScan2* software (Burton et al., 2012).

Genotyping

Genomic DNA was extracted from 2 week old seedling of the 198 selected lines and parents (Kaybonnet and ZHE733) by using cetyl tri-methyl ammonium bromide (CTAB) method as described (Murray and Thompson, 1980). The GBS libraries and analysis was performed by University of Minnesota Genomics Center. The library was created using single-end library type with enzyme *Pst*I and *Msp*I. NextSeq generated 1x150-bp read with total of two millions base pairs (bp) reads per sample. The reads with mean quality scores above Q30 was chosen for analysis.

SNP identification

SNP identification was analyzed from the sequence reads, and were generated in a FASTQ file. De-multiplexed FASTQ files were generated using Illumina BCL2FASTQ software. FASTQ files with more than the targeted number of reads (2,000,000) were subsampled down to 2,000,000 number reads. The first 12 bases were removed from the beginning of each read in order to remove adapter sequences, using Trimmomatic to remove adapter sequences at the 3' ends of reads. The FASTQ files were aligned to the reference genome of Nipponbare, *Oryza sativa* spp. Japonica version MSU7 by using Burrows-Wheeler Alignment (BWA) software. The sequences that perfectly matched and aligned were processed for SNP calling. Freebayes was used to jointly call variants across all samples simultaneously. The raw Variant Call Format (VCF) file generated by Freebayes was filtered using VCF tools to remove variants with minor allele frequency less than 1%, variants with genotype rates less than 95%, samples with genotype rates less than 50%, variants with 100% missing data, variants with monomorphic markers between parents,

and variants with more than 50% heterozygosity. The filtered data file with final set of SNPs in nucleotide-based hap map format was converted to an ABH-based format, where “A” represents donor allele, “B” represents recipient allele, and “H” represents heterozygous allele.

Linkage map construction and QTL mapping

The genotypic data for 198 lines of the K/Z RIL population with filtered SNP markers, were used for linkage map construction by using the linkage mapping function in the QTL IciMapping software version 4.2.53 (Meng et al., 2015) with a recombination frequency (r) set at 0.45. The Kosambi mapping function was used to convert recombination frequencies to map distance (cM) (Kosambi, 1943). Furthermore, the markers were ordered with a threshold logarithm of odd (LOD) set at 2.5. The morphological traits and grain yield components used to conduct QTL analysis were PH, SP, PL, PPB, FG, and BY; each trait under WW and DS conditions. In addition, data for HD, TN, FLW, and DLR were collected. The root architectural traits for ABA sensitivity screening, include RL, RSR, TRN, SRN, DRN, and RFW at different level of ABA concentrations: 0 μ M (control), 3 μ M, and 5 μ M that only RL used for QTL mapping. QTL analysis was done with 4133 SNP markers by using QTL IciMapping software version 4.2.53 with inclusive composite interval mapping (ICIM) function (Meng et al., 2015). QTLs explaining $\text{LOD} \geq 2.5$ were declared significant. QTL nomenclature used is based on the trait name, chromosome number, and their physical map position on the genome (Solis et al., 2018; McCouch, 2008). The left and the right markers flanking the QTLs were determined. Genotypic frequency was calculated according to the closest marker to the QTL peak.

Identification of candidate genes within the QTL regions

The candidate genes present within the QTL regions were identified based on the position of the SNP markers flanking the QTL regions, and the nearest predicted/annotated gene in the region, using the MSU rice *japonica* reference genome annotation release 7.0 as the reference. All the genes within 25 Kb of the identified QTLs position were classified into three major functional categories, including biological process, molecular function, and cellular component. The key functional genes regulating drought-related traits and ABA sensitivity were further analyzed by extracting RNA from the parental genotypes and used for analysis of their gene expression under DS conditions.

RT-qPCR validation of the key functional genes identified within the QTL regions regulating drought-related traits and ABA sensitivity

The leaf samples for RNA extraction and quantification of gene expression were collected from the two parental genotypes. RNA was extracted using the RNeasy® Plant Mini Kit (Qiagen Inc, Hilden, Germany) having a 260/280 ratio of 1.8–2.1, or 260/230 ratio ≥ 2.0 . One microgram RNA sample was reverse transcribed using GoScript® Reverse Transcription System (Promega, Madison, Wisconsin, USA). The RT-qPCR reaction samples were prepared in a total volume of 20 μ L, containing 10 μ L of SYBR green master mix, 1 μ L of cDNA template, 8 μ L of ddH₂O, and 1 μ L of each primer. RT-qPCR was performed with a BIO-RAD CFX-96 instrument (Bio-Rad Laboratories, Inc., Hercules, California, U.S.). The relative difference

in expression for each sample in individual experiments was determined by normalizing the threshold cycle (Ct) value for each gene against the Ct value of Ubiquitin and calculated relative to the respective control samples as a calibrator using the equation $2^{-\Delta\Delta Ct}$. The average of three biological replicates and three technical replicates for each sample was used to obtain each expression value (Ramegowda et al., 2014; Bevilacqua et al., 2015; De Freitas et al., 2016). The standard error was used to separate means for significant effects.

Analysis of genetic diversity in the K/Z RIL population

The raw fastq reads from 200 genotypes comprising 2 parental genotypes and 198 lines of the K/Z RIL population were mapped to the reference rice genome cv. Nipponbare (IRGSP 1.0) using the Burrows-Wheeler Aligner (BWA) (Li and Durbin 2009). The bam files were used to call SNPs using GATK (Van der Auwera and O'Connor 2020). Approximately 6.5 million high quality SNPs were retained (less than 2% missing rate and more than 5% minor allele frequency) and annotated. Of these, the SNPs present in the up- and down-stream region of the 26 drought loci were selected and analyzed further. Principal component analysis (PCA) was conducted using '-pca' flag in PLINK v.1.9 (Chang et al 2015) and visualized in R (R Core Team 2020).

Results And Discussion

The K/Z RIL population was developed at the USDA Dale Bumpers National Rice Research Center, Stuttgart, Arkansas, USA by crossing diverse parental genotypes from different subspecies, Kaybonnet (*tropical japonica*) and ZHE733 (*indica*) by SSD method to create segregating progenies with high genetic variability for selection desirable genes. Root growth and development vary among the rice genotypes studied here. Greub (2015) identified that the root length of Kaybonnet showed the longest root compared to diverse rice genotypes, such as Bengal, Sipirasikkam (GSOR 310428), *O. glaberrima*, IR64, Nagina-22 (N22), and Vandana. Root length is probably the most important architectural trait for drought avoidance, by enabling the roots to reach deeper water levels in the ground. Additionally, Greub (2015) also determined that Kaybonnet had higher metaxylem number than Bengal, *O. glaberrima*, IR64, Nagina-22 (N22), Vandana, and parental line ZHE733. The number and size of the metaxylem vessels are associated with their conductivity of water. Among these lines, Bengal and Nagina-22 (N22) are examples of drought-resistant rice genotypes, while IR64 and Nippobare as the reference for drought-sensitive genotypes.

The differences of the root anatomy and root morphology between Kaybonnet (drought resistant) and ZHE733 (drought sensitive) displayed in figure 1 and supplementary table 2. Kaybonnet exhibited longer root than ZHE733, and also Kaybonnet has two metaxylem number while ZHE733 only has one metaxylem in the stele area. Additionally, total aerenchyma area in Kaybonnet (0.34 mm²) is wider than ZHE733 (0.29 mm²). According to Zhu et al. (2010) aerenchyma improves drought resistance by decreasing root metabolic costs and greater water acquisition from dehydrated soil. Thus, Kaybonnet exhibited more drought resistance compared to ZHE733.

Based on the screening of grain yield under DS conditions at reproductive stage (R3), among the two parental genotypes, Kaybonnet is drought resistant while ZHE733 displays a drought sensitive phenotype. Under DS conditions, the number of filled grains per panicle in Kaybonnet reduced 20%, while ZHE733 reduced 50% (Fig. 2). The progeny of a cross between drought resistant and sensitive genotypes are useful to study the inheritance of drought resistance, and identification of important QTLs for variation in grain yield under DS conditions (Kumar et al., 2018; Islam et al., 2012). A total of 198 K/Z RILs was studied for filled grains per panicle number, and out of which, 13.13% were found to be highly drought resistant lines, 11.11% moderately drought resistant lines, and 75.75% drought sensitive lines, suggesting there are multiple factors involved in inheritance of drought resistant and sensitive phenotypes in the population.

Variation in morphological traits of RILs under drought stress conditions

Rice is more sensitive to DS conditions compared to the other cereal crops such as wheat, rye, and barley (Huang et al., 2014). The two parental lines are diverse in the morphological traits such as plant height (PH), tiller number (TN), heading day (HD), flag leaf width (FLW), and leaf rolling score (DLR). Kaybonnet (derived from variety Katty and Newbonnet) compared to ZHE733 is the donor parent, with high stature, low TN, late HD, wider FLW, and low DLR while ZHE733, the recurrent parent, has short stature, higher TN, early HD, narrow FLW, and high DLR under WW conditions. In addition to the two parental lines having variation for morphological traits when subjected to DS conditions, Kaybonnet showed overall superior performance to ZHE733 because Kaybonnet is drought resistance while ZHE733 is sensitive to DS conditions, thus exhibiting less resistance. Under DS conditions, PH of Kaybonnet reduced to 14.29%, while ZHE733 reduced to 39.22%. Meanwhile, TN of Kaybonnet reduced to 20%, and ZHE733 reduced to 14.3%. These results are in agreement with previous studies showing that water deficit conditions have a negative effect on plant development and growth due to a loss of turgor (Hsiao et al., 1970; Specht et al., 2001; Farooq et al., 2011; Todaka et al., 2015; Kumar et al., 2019). The reduction of the plant height and productive tiller number under DS conditions are associated with the reduction of the cell cycle processes, cell expansion and elongation (Mantovani and Iglesias, 2008).

In the RIL population, morphological traits such as PH, TN, HD, FLW, and DLR showed normal frequency distribution (Figs. 3 and 4) signifying quantitative inheritance, and thus the morphological traits were suitable for QTL analysis (Fang et al., 2019). Within the RIL population, there was a wide range of morphological traits that showed variation across WW and DS conditions for PH and TN, while HD, FLW, and DLR measured in DS conditions exhibited transgressive segregation pattern.

Within the RIL population, all the morphological traits under DS showed a significant reduction compared to WW conditions (Supplementary Table 1). Based on the PH and TN characteristics in WW conditions, the RIL population is more skewed toward parent ZHE733, plant height greater than 50 cm and tiller number greater than 8. The average PH in the RIL population under DS conditions showed 50% reduction, which is more than either of the parents, while the average of TN reduced to 13.90% which is less than either of the parents. Based on the PH reduction in the DS conditions compared to WW conditions,

54.04% of lines in the RIL population showed more than 50% reduction suggesting more than 50% of the lines are sensitive to DS conditions. Meanwhile, based on the TN reduction, 71.72% of lines in the RIL population showed drought resistant with less than 30% reduction suggesting that majority of the lines have reduced the tiller number and have characters of ZHE733. Among the RIL population, 28 lines showed less reduction ($\leq 30\%$) for both plant height and tiller number traits and these lines could be potential genetic resource to develop drought resistant varieties.

Genetic variation for grain yield components under reproductive stage drought stress

The effect of DS conditions on rice are different at every growth stage. Rice is very sensitive to DS conditions, especially at the reproductive stage, and grain yield is dramatically reduced even under slight DS conditions (Kamoshita et al., 2008; Palanog et al., 2014; Dwiningsih, 2020b). The effective way to reduce grain yield loss under DS conditions is to develop drought-resistant rice varieties, that is however very challenging due to the complexity of the drought resistant trait associated with grain yield components. The identification of drought QTLs associated with the grain yield components under water deficit conditions, and their utilization in molecular breeding, is an alternative method of increasing breeding effectiveness. Moreover, identified QTLs can be incorporated in a marker assisted breeding (MAB) strategy (Venuprasad et al., 2011).

The mean values of the grain yield components showed a significant difference in performance of both parents and population under DS conditions compared to WW, including biological yield (BY), spikelet per panicle number (SP), filled grain per panicle number (FG), panicle length (PL), and primary panicle branch number (PPB) (Table 1). ZHE733 showed a greater reduction in overall grain yield components compared to Kaybonnet. Under DS conditions, ZHE733 showed significant reduction ($P < 0.05$) in BY, SP, FG, PL, and PPB; 51.49%, 38.89%, 62.96%, 16.13%, and 12.5%, respectively. In addition, Kaybonnet showed lower reduction compared to ZHE733 in BY, SP, FG, and PL; 4.73%, 28.85%, 24.24% and 11.06%, respectively. The trait PPB in Kaybonnet showed greater reduction 31.58% compared to ZHE733.

In the RIL population, grain yield components showed a continuous frequency distribution under both WW and DS conditions, indicating polygenic control of the traits as shown in Fig. 5A (BY), Fig. 5B (SP), Fig. 5C (FG), Fig. 5D (PL), and Fig. 5E (PPB). Transgressive segregation was observed for the grain yield components under WW and DS conditions. Moreover, there were significant differences ($P < 0.05$) among the RILs for all the grain yield components under both WW and DS conditions. Under DS conditions, the average of BY, SP, FG, PL, and PPB in the RIL population reduced 12.5%, 38.75%, 47.62%, 15.82%, and 16.92, respectively compared to WW conditions. Based on the BY, SP, PL, and PPB reduction in the DS conditions compared to WW conditions, most of the lines in the RIL population is skewed toward drought resistant phenotypes with reduction less than 30%. Meanwhile, based on the FG, most of the lines belong to drought sensitive with reduction more than 50%. There are 12 lines that showed less reduction ($\leq 30\%$) for all the grain yield components and these RIL lines could be a potential genetic resource to develop drought resistant rice varieties.

Table 1 The average and range values of grain yield components of K/Z RIL population under WW and DS conditions

Traits	Treatments	Kaybonnet	ZHE733	K/Z RIL Population	
				Average	Range
Biological yield	WW	14.80	13.40	20	4 - 49
	DS	14.10	6.50	17.50	2 - 47
Spikelet per panicle number	WW	104	90	133.35	45 - 328
	DS	74	55	81.68	26.8 - 188.4
Filled grain per panicle number	WW	66	43.2	43.07	2 - 176
	DS	50	16	22.56	0 - 97
Panicle length (cm)	WW	21.7	18.6	21.24	11.90 - 33.30
	DS	19.3	15.6	17.88	11.64 - 26.62
Primary panicle branch number	WW	19	8	11.17	3 - 22
	DS	13	4	9.28	5.4 - 15.6

Variation in root architectural traits under ABA conditions

The parents, Kaybonnet and ZHE733 had contrasting responses under ABA conditions, where Kaybonnet as drought-resistant parent showed more sensitivity to ABA compared to ZHE733 as the drought sensitive parent. Both parents experienced a reduction in root length (RL), total root number (TRN), shallow root number (SRN), deep root number (DRN), and root fresh weight (RFW) under ABA conditions compared to control conditions. Kaybonnet showed a greater reduction in RL, TRN, SRN, DRN, and RFW compared to ZHE733 (Supplementary Table 2). Under ABA conditions (3 μ M), Kaybonnet showed high reduction in RL, TRN, SRN, DRN, and RFW; 61.94%, 62.12%, 100%, 66.22%, and 50%, respectively. However, ZHE733 showed less reduction under ABA conditions (3 μ M) in RL, TRN, SRN, DRN, and RFW; 17.59%, 5.43%, 4.68%, 5.79%, and 6.45%, respectively. Furthermore, under ABA conditions (5 μ M), ZHE733 also displayed less reduction in RL, TRN, SRN, DRN, and RFW compared to Kaybonnet.

The K/Z RIL population showed variation in their root architectural traits in response to ABA (0 μ M, 3 μ M, and 5 μ M), such as reduction in RL (Fig. 6A), increase in RSR (Fig. 6B), reduction in TRN (Fig. 6C), decrease of SRN (Fig. 6D), shortening in DRN (Fig. 6E), and lessening RFW (Fig. 6F). Furthermore, the distribution of root architectural traits studied under ABA conditions showed a near normal distribution. The range of RL, RSR, TRN, SRN, DRN, and RFW of the K/Z RIL population under ABA conditions also exhibited a wide variation (Supplementary Table 2). Additionally, the average of root architectural traits of the RILs lie between the parents in ABA conditions. The average of RL, TRN, SRN, DRN, and RFW in the RIL population under ABA conditions 3 μ M also showed reduction; 18.69%, 16.46%, 33.78%, 9.26%, and

2.94%, respectively. In addition, under ABA conditions 5 μ M; the average of RL, TRN, SRN, DRN, and RFW in the RIL population also showed reduction; 21.16%, 21.07%, 36.51%, 12.61%, and 23.53%. Although both parents and the RIL population exhibited a reduction in RL, TRN, SRN, DRN, and RFW under ABA conditions, the reduction was greater in Kaybonnet. Root to shoot ratio (RSR) increased 38% and 16% for Kaybonnet under ABA conditions 3 μ M and 5 μ M, while for RIL population under ABA conditions 3 μ M and 5 μ M increased 22.92% and 22.69%. Previous studies have demonstrated that ABA sensitivity is associated with drought stress resistance through its effect on the stomatal movement (Lim et al., 2015; Duan et al., 2008; Todorov et al., 1998). Lim et al. (2015) also indicated that drought-sensitive rice plants grown in media with 2 or 5 μ M ABA had significantly longer roots and shoots compared to the plants in control media. These data suggested that the drought-sensitive rice plants were insensitive to ABA and exhibited the ABA-dependent pathway in response to drought stress.

Correlation of morphological traits and grain yield components under WW and DS conditions with root architectural traits under ABA conditions

Correlation analysis increases an understanding of the overall contribution of various rice plant traits to each other (Gibert et al., 2016). A Pearson's correlation coefficient analysis was carried out on morphological traits and grain yield components under WW and DS conditions, and also on root architectural traits under ABA conditions, to analyze the correlations among them. Significant correlations were observed among all of the traits studied. Furthermore, FG-DS as the major trait among the grain yield components under DS conditions showed significant positive correlations with most of the morphological traits, including HD, TN-WW, PH-WW, PH-DS, FLW, and DLR. FG-DS also has positive correlations with other grain yield components, such as PL-WW, PL-DS, PPB-WW, SP-WW, SP-DS, and FG-WW (Supplementary Table 5). Additionally, FG-DS exhibited significant positive correlations with most of the root architectural traits under ABA conditions such as RL-ABA0, RL-ABA3, RL-ABA5, RSR-ABA0, RSR-ABA3, RSR-ABA5, TRN-ABA0, TRN-ABA3, TRN-ABA5, DRN-ABA0, SRN-ABA3, SRN-ABA5, DRN-ABA3, DRN-ABA5, RFW-ABA0, RFW-ABA3, and RFW-ABA5 (Supplementary Table 6). However, significant negative correlations were also observed between FG-DS and the morphological trait like TN-DS, and also with the grain yield components such as BY-WW, BY-DS, and PPB-DS. Several root architectural traits also showed significant negative correlations with FG-DS, including SRN-ABA0.

In this study, a positive correlation found between FG-DS with most of the morphological traits, the other grain yield components, and the major root architectural traits under ABA conditions, indicate that the rice drought-resistant plants maintain their grain yield under DS conditions through development of cell elongation, maintenance of cellular membrane integrity, and regulation of osmotic stress tolerance via ABA-mediated cell signaling (Kanbar et al., 2009; Ramegowda et al., 2015; Ding et al., 2016; Basu et al., 2016; Catalos et al., 2017; Nada et al., 2018; Hassaoni et al., 2018; Li et al., 2019; Kim et al., 2020). Furthermore, the negative correlation between FG-DS with BY under WW and DS conditions indicate that there is more assimilate distribution into the grains compared to the other parts of the plant. Moreover, ABA sensitivity was found correlated with the drought resistance of rice plants (Lim et al., 2015; Duan et al., 2008; Todorov et al., 1998).

Based on the correlation analysis, three lines (100007, 100036, and 100135) of the RIL population showed drought resistant, based on the morphological traits (PH and TN) and grain yield components (BY, SP, FG, PL, and PPB) and ABA sensitivity response characteristics similar to Kaybonnet as drought resistant parent. These three lines showed less than 30% reduction under DS conditions for PH, TN, BY, SP, FG, PL, and PPB. Meanwhile, for root architectural traits (RL, TRN, SRN, DRN, and RFW), these three lines showed more than 50% reduction. Furthermore, based on the morphological traits and grain yield components under WW conditions, these three lines displayed short PH and high number in TN, BY, SP, PL, FG, and PPB which become a good genetic resource to develop drought resistant varieties. Under WW conditions, the range of PH, TN, BY, SP, PL, FG, and PPB were 42 – 51 cm, 5 – 9, 17 – 24 g, 104 – 130, 17.5 – 23.7 cm, 100 – 103, and 10 – 12, respectively.

Relationships of root anatomy and drought resistance

Many studies have been reported that various root anatomical phenes such as metaxylem, aerenchyma, and cortical cells properties influence plant performance and productivity under DS conditions in cereal crops including maize (Chimungu et al., 2014a, b; Chimungu et al., 2015a, b), rice and wheat (Kadam et al., 2015); and also in legume crops such as soybean (Prince et al., 2017), common bean (Peña-Valdivia et al., 2010), chickpea, groundnut, pigeonpea, and cowpea (Purushothaman et al., 2013). Under DS conditions, plants allocate more energy and carbon resources to root growth rather than to shoot growth, which can increase water acquisition (Lynch and Ho, 2005; Palta and Gregory, 1997; Sharp and Davies, 1979). Aerenchyma and cortical cells contribute to increase root growth for soil exploration in the deeper soil layer, consequently, greater water acquisition, and improved productivity by reducing the root metabolic costs leading more carbon resources allocation to improve root growth (Jaramillo et al., 2013; Lynch, 2013). Understanding root adaptive mechanisms is important to maintain higher productivity under DS conditions.

Metaxylem vessels play an important role in water and nutrients uptake from the soil, and subsequent transport within the plant cells (Steudle and Peterson, 1998). A study by Kadam et al. (2015), metaxylem vessel properties are important traits to increase water-use efficiency under DS conditions. Parental Kaybonnet and drought-resistant lines showed higher number of metaxylem vessels compared to ZHE733 and drought-sensitive lines (Fig. 7). Kaybonnet has two metaxylem vessels (Fig. 1) and in an average, drought-resistant lines have three metaxylem vessels while ZHE733 and drought-sensitive lines only have one metaxylem vessels. Increasing of the metaxylem number leading to increase root hydraulic conductivity that decrease the metabolic costs for exploring water in deeper soil layer allowing water uptake efficiency and higher yield under DS conditions (Prince et al., 2017). The increased root hydraulic conductivity also leads to increase in performance of shoot physiological processes under DS conditions. The increased number of metaxylem vessels, improves water uptake efficiency under DS conditions thereby causing increased stomatal conductance and internal capture of carbon dioxide and thus overall maintains photosynthesis activity under DS conditions. In contrast, drought-resistant lines displayed smaller metaxylem size compared to drought-sensitive lines. The average of the metaxylem size of the drought-sensitive lines are 0.0008 mm² while drought-resistant lines are only 0.0005 mm². According to

Richards and Passioura (1989), the decrease of metaxylem size allows greater yield under DS conditions, because decreased metaxylem size is correlated to greater hydraulic conductivity, allowing greater water acquisition.

Aerenchyma is the enlarged air space in the root cortex resulting from programmed cell death that affects root respiration (Evans, 2004). Aerenchyma showed a positive correlation with drought resistance in rice. Kaybonnet and drought-resistant lines also showed higher percentage of aerenchyma than ZHE733 and drought-sensitive lines. The average coverage of aerenchyma in drought-resistant lines is 60% of the total root cross section while drought-sensitive lines is only 40%. Higher aerenchyma is associated with decreased root respiration costs, leading to more carbon allocation and improved root growth, thereby increasing water acquisition from the deep soil layer of drying soil and higher grain yield under DS conditions (Zhu et al., 2010).

Another root anatomical feature, important for improved drought resistance is cortical cell. Cortical cell area is the total transverse root cortex minus aerenchyma area. There are average of eleven cortical cell file number in drought-sensitive lines that is higher compared to five in the drought-resistant lines. Cortical cell file numbers are positively linked to higher root respiration (Jaramillo et al., 2013; Lynch, 2013) therefore the drought-sensitive lines have higher respiration compared to drought-resistant lines, causing reduced root growth, lower water acquisition and decreased productivity under DS conditions. According to Rieger and Litvin (1999), reduced cortical cell file number also showed a positive correlation to a smaller radial path that affect root hydraulic conductivity, thereby increasing water acquisition under DS conditions.

High-density genetic linkage map with GBS markers

A genetic linkage map is an important tool to explore the plant genome, and to obtain information of allele introgression during plant breeding efforts (De Soursa et al., 2015). By using high-density genetic linkage map leads to narrow down the location of QTLs into a specific region and predict more accurate candidate genes for gene cloning, then validation with reverse genetics approaches (Hattori et al., 2009). Based on the GBS analysis, 28,598 SNP markers were obtained from 200 samples (2 parental lines and 198 RILs) with heterozygosity level of 1.3% and non-parental alleles at 0.4%. The filtering process of the SNP markers was done based on the missing data ($\leq 90\%$), minor allele frequency (MAF $< 1\%$), polymorphic markers between parents, recombinant frequency, and percentage of heterozygosity. Furthermore, 4133 filtered SNP markers were obtained, and were used in the development of the high-density genetic linkage map by using QTL IciMapping software version 4.2.53 with the Kosambi mapping function.

The number of SNP markers mapped to each chromosome varied from 182 SNP markers found on chromosome 12 to 562 SNP markers on chromosome 1, with an average of 344.42 SNP markers per chromosome. The total length of the genetic linkage map was 6063.12 cM (varied from 343.72 cM on chromosome 10 to 676.52 cM on chromosome 2), with the average of 505.26 cM per chromosome. A calculated average genetic distance between two SNP markers across the chromosome was 1.58 cM

(ranged from 0.92 cM on chromosome 6 to 2.89 cM on chromosome 12). The density of the genetic linkage map was 0.69 SNP markers per cM or an average of 1 SNP marker every 1.5 cM. This high-density genetic linkage map covered 373 Mb of the rice genome and can be used to identify QTLs with higher resolution and reliability for application in rice breeding under DS conditions. Moreover, the high-density genetic linkage map lead to identification and selection of candidate genes within the QTL regions that are involved in improving drought resistance in the rice plants, and towards developing drought-resistant rice varieties. Many previous studies have also used high-density linkage maps for QTL mapping (Bhattarai and Subudhi, 2018; Sabar et al., 2019; Barik et al., 2019; Melandri et al., 2020; Barik et al., 2020; Dwiningsih et al., 2021b).

QTL mapping of morphological and yield traits under reproductive stage drought stress conditions and root architectural traits under ABA conditions

The identification of QTLs for morphological traits, grain yield components, and root architectural traits is important to understand the genetic complexity of the drought-related traits. The genetic variation of drought-related traits is regulated by numerous genes that have a large effect on the traits (Baisakh et al., 2020). In this research, 41 QTLs were identified for morphological traits and grain yield components under WW and DS treatments, and also root architectural traits under ABA treatments (Supplementary Table 3, Fig. 8). The identified QTLs varied under WW, DS, and ABA treatments. Additionally, the number of QTLs varied for morphological traits, grain yield components, and root architectural traits. A total of 12 QTLs were identified for multiple morphological traits, grain yield components, and root architectural traits suggesting that these QTLs have pleiotropic effect. For this study, the LOD values more than 2.5 was selected in order to filter unassociated candidate regions. The maximum values of LOD was 28.8 and PVE was 13.809% (Supplementary Table 3).

Most of the QTL identified in this study were mapped to approximately the same locations as previous reports (Mu et al., 2003; Courtois et al., 2003; Lanceras et al., 2004; Bernier et al., 2007; Bernier et al., 2009; Venuprasad et al., 2009; Mishra et al., 2013; Yadav et al., 2013; Wang et al., 2014; Palanog et al., 2014; Saikumar et al., 2014; Prince et al., 2015). Chromosome 1 harbored the highest number of QTLs for PH (Monna et al., 2000; Lanceras et al., 2004; Zhou et al., 2016; Jiang-xu et al., 2016; Yadav et al., 2019a; Zeng et al., 2019; Xu et al., 2020). The most QTLs associated with grain yield were located on chromosome 5 and 6 (Solis et al., 2018; Yadav et al., 2019b; Baisakh et al., 2020; Takahashi et al., 2001; Bernier et al., 2007; Xu et al., 2020). Moreover, chromosome 10 had the highest number of QTLs for root architectural traits (Xu et al., 2001; Lou et al., 2015; Kitomi et al., 2015; Gimhani et al., 2018). Eventhough there were overlapping genomic regions with the previous studies, the results of this study predicted QTLs/genomic regions in close association with genes of interest.

Candidate genes underlying QTL regions

Identification of candidate genes within a QTL region is useful for marker-assisted pyramiding to develop drought-resistant rice varieties (Bhattarai et al., 2018), and is also important for developing transgenic rice with enhanced drought resistance (Varshney et al., 2011). In this research, we identified candidate genes

involved in many biological processes, molecular functions, cell components, and drought response (Supplementary Table 3). The QTL clusters contain candidate genes with large pleiotropic effects. The total number of 184 candidate genes with an average of 5 genes per QTL were identified in 41 QTLs for morphological traits, grain yield components under WW and DS conditions, and root architectural traits under ABA conditions (Supplementary Table 3). These candidate genes were distributed unevenly on different chromosome. There were 15 candidate genes with high LOD and PVE score that have been previously studied for drought stress. In total 11 novel candidate genes with high LOD and PVE score were discovered in this analysis with unknown annotations. Many known genes present within these QTL regions include genes with homology to APETALA2 (AP2)/ETHYLENE RESPONSE FACTOR (ERF) transcription factor, malate dehydrogenase protein, photosystem II oxygen evolving complex protein PsbQ family protein, WRKY transcription factor, MYB transcription factor, Zinc Finger (ZFN) protein, endoplasmic reticulum protein, DEAD-box RNA helicase, glycosyl transferase protein, Late Embryogenesis Abundant (LEA) protein, and no apical meristem protein (NAC).

A transcription factor family well-known for drought response like APETALA2 (AP2)/ETHYLENE RESPONSE FACTOR (ERF) (Lata and Prasad, 2011; Mizoi et al., 2012; Licausi et al., 2013; Phukan et al., 2017), has family members present in several of the QTL regions such as QTLs for PH-WW on chromosome 1 (LOC_Os01g66270); RL-ABA3, RL-ABA5 on chromosome 2 (LOC_Os02g54160); and FG-WW, FG-DS, RL-ABA3, and RL-ABA5 on chromosome 5 (LOC_Os05g49010). Furthermore, the QTL regions for PH-WW on chromosome 1 is adjacent to the semi-dwarfing gene *sd1* locus (38.3 Mb). A strong linkage has been found previously between *sd1* and QTL for drought-related traits (Vikram et al., 2015). The *sd1* locus is also associated with underground and above ground traits in rice, such as plant height and root architectural traits (Yadav et al., 1997). Reduction in plant height under DS conditions, is the adaptation of rice plants to DS. An important QTL for grain yield components under DS, FG located on chromosome 5, overlaps with 12 candidate genes. There is also an overlap between QTL for root architectural traits, RL under ABA conditions and FG under DS, suggesting that ABA is involved in the drought stress resistance mechanism. Under DS conditions, ethylene biosynthesis is increased and interacts with AP2/ERF, and finally a response to water deficit (Abiri et al., 2017; Nakano et al., 2006; Ma et al., 2014). Additionally, AP2/ERF responds to ABA in order to help activate ABA dependent and independent stress responsive genes. Transgenic rice with over-expression of an AP2/ERF showed an increase in drought resistance (Pan et al., 2012). An understanding of the AP2/ERF gene functions in the drought resistance mechanisms in rice, may provide valuable information to facilitate the improved adaptation of rice to DS conditions.

An important candidate gene, LOC_Os03g56280, known to regulate malate dehydrogenase in response to drought stress was found in the QTL regions for BY-DS, RL-ABA3, and RL-ABA5, on chromosome 3. Another candidate gene that is responsible for carbohydrate metabolism was detected on chromosome 9 (LOC_Os09g08120) in the QTL regions for BY-DS and RL-ABA5. Malate dehydrogenase is an enzyme that catalyzes the oxidation of malate to oxaloacetate by using NAD(H)/NADP(H) as a cofactor. Additionally, this enzyme can be expressed in different parts of the rice plants, such as root, leaf, panicle, and stem and was induced in the presence of water deficit (Nan et al., 2020). Transgenic plants over-expressing

malate dehydrogenase exhibited increasing drought-resistance compared to wild-type. Malate dehydrogenase was also identified as a drought responsive protein by Agrawal et al. (2016). Under DS conditions, drought-resistant genotypes accumulate a higher level of malate dehydrogenase, that protects membranes from damage by reactive oxygen species (ROS) (Guo et al., 2018). By elucidating the function of malate dehydrogenase, the drought response in rice can be better understood.

A gene encoding photosynthesis function (LOC_Os04g44190) is present in the QTL regions for BY-WW, RL-ABA3, and RL-ABA5 on chromosome 4. This gene is involved in the light reaction of photosystem II (PSII) and is known to control stomatal closure, and protect plants from dehydration (Sasi et al., 2018). Under DS conditions, the photosynthetic rate is decreased due to reduction in photosynthetic electron transport and carbon assimilation, resulting in reduction of grain yield. Moreover, PSII is a pigment-protein complex in thylakoid membranes that is responsible for oxygen evolution, water splitting, and plastoquinone reduction (Lu, 2016). LOC_Os04g44190 encoding a PsbQ family protein that belongs to the class of PSII extrinsic proteins, and under DS conditions this protein changes expression due to change in PSII efficiency (Sasi et al., 2018). Therefore, PsbQ protein plays an important role in drought stress resistance.

A WRKY transcription factor that is involved in drought stress response and plant development was detected on chromosome 5 (LOC_Os05g49210) of the QTL regions for FGC and FGD. Shen et al. (2012) reported that transgenic rice with over-expression of OsWRKY30 showed improved drought resistance. Likewise, silencing WRKY genes in transgenic rice demonstrated increased drought sensitivity. In addition, expression of the WRKY transcription factor induced ABA accumulation under DS conditions, leading to stomatal closure and reduction in water loss (Chen et al., 2010; Schroeder et al., 2001). Yan et al. (2015) also reported that the expression of a WRKY transcription factor was increased by ABA treatment.

Genes present on chromosome 5 include LOC_Os05g49240 in the QTL regions for SP-DS and PPB-DS; and also on chromosome 10 (LOC_Os10g41460) in the QTL regions for BY-DS and RL-ABA3, were identified MYB transcription factor, a known transcription factor in drought response (Tang et al., 2019; Baldoni et al., 2015; Li et al., 2015; Dai et al., 2007; Ma et al., 2009; Yang et al., 2012; Xiong et al., 2014; Quan et al., 2010). Transgenic rice with over-expression of OsMYB6 exhibited increased resistance to drought compared to wild-type and contained higher proline catalase (CAT) and superoxide dismutase (SOD) activity. Additionally, OsMYB6 transgenic rice plants also showed higher expression of abiotic stress-responsive genes under DS conditions (Tang et al., 2019). Katiyar et al. (2012) also reported that the expression of MYB genes is controlled by drought. Increasing drought resistance correlated with over-expression of MYB genes and ABA accumulation (Xiong et al., 2014).

A stress-responsive transcription factor, Zinc Finger (ZFN) protein (LOC_Os06g49080), is located on chromosome 6 in the QTL regions for FG-WW, BY-WW, RL-ABA3, and RL-ABA5. The ZFN protein was reported to improve drought resistance in plants, suggesting that the ZFN protein contribute to the higher yield under DS conditions via control of stomatal closure (Huang et al., 2009; Ciftci-Yilmaz et al., 2000; Mukhopadhyay et al., 2004; Sakamoto et al., 2004).

A gene involved in sugar metabolism, OsSAC1 was found on chromosome 7 (LOC_Os07g02520) in the QTL regions for SP-DS. OsSAC1 regulates sugar partitioning in the carbon metabolism of young leaves and developing leaf sheaths (Zhu et al., 2018). OsSAC1 encodes an endoplasmic reticulum protein that causes sugar accumulation in the rice leaves and can be used to produce energy and construct carbon skeletons.

The genomic region on chromosome 8 (LOC_Os08g06344) in the QTLs for FG-DS, RL-ABA3, and RL-ABA5 encodes a DEAD-box RNA helicase which was reported to improve drought resistance in rice (Nawaz and Kang, 2019; Vashisht et al., 2006; Macovi et al., 2012). Over-expression of OsRH58, a chloroplast DEAD-box RNA helicase, in transgenic rice showed improved drought resistance, displayed by better survival rate than the wild-type under DS conditions (Nawaz and Kang, 2019). Furthermore, gene expression of the OsRH58 was increased under drought.

A gene regulating late embryogenesis abundant (LEA) protein was detected on chromosome 12 (LOC_Os12g02700) underlying the QTL regions for BY-DS. LEA protein has a major role in drought resistance in plants (Xiao et al., 2007; Duan and Cai, 2012; Magwanga et al., 2018; Liang et al., 2019; Chen et al., 2019; Kamarudin et al., 2019). Under DS conditions, LEA genes showed higher expression in the drought resistant plants compared to drought sensitive. In support of LEA functions, Xiao et al. (2007) reported that transgenic rice with over-expression a LEA protein gene OsLEA3-1 exhibited higher grain yield compared to wild-type under DS conditions.

Within the QTL region for SP-DS on chromosome 12, LOC_Os12g29330, (OsNAC139) was identified. OsNAC139 is a member of the NAC transcription factor family that is known to control plant response to drought (Kikuchi et al., 2003; Nakashima et al., 2007; Takasaki et al., 2010; Shim et al., 2018) by producing no apical meristem (NAM)/NAC protein. Rice contains 151 NAC genes (Puranik et al., 2013), from which several studies have reported that over-expression of OsNAC genes leads to improved drought resistance (Nakashima et al., 2007; Hu et al., 2008; Yokotani et al., 2009; Zheng et al., 2009; Jeong et al., 2010).

Among all the candidate genes identified within the QTL regions, various transcriptomes correlated with drought stress resistance were detected. Drought resistance of the rice plants can be either associated with metabolic regulation or osmoregulation. In conclusion, this study has revealed a number of potential candidate genes for developing drought-resistant rice varieties.

RT-qPCR validation of the key functional genes identified within the QTL regions regulating drought-related traits and ABA sensitivity

Information about the differences in the expression of drought resistance genes between drought-resistant and sensitive genotypes and their relationship to morphological characteristics, grain yield components, and root architectural traits under DS conditions has been limited. However, the study of the differences in gene expression between resistance and sensitive genotypes could improve the efficiency and opportunities of developing drought resistant varieties.

The identified candidate genes within the QTL regions regulate morphological traits and grain yield components under WW and DS, and also root architectural traits under ABA treatments, were further analyzed to exemplify their roles in increasing drought stress resistance in rice by identifying their expression under different conditions. In this research, QTLs controlling morphological and yield traits under reproductive stage drought stress conditions were identified and compared to those identified under WW conditions. The QTLs under DS conditions were different from those identified under WW conditions. This might be due to the difference in expression of the genes for morphological and grain yield traits under DS and WW conditions. Plants respond and adapt to the drought stress through several processes, such as physiological, biochemical, and molecular processes that are regulated by transcriptional regulators. When rice plants are exposed to drought stress, certain genes are activated or repressed. Proteins, as the products of the activated genes, will protect the plants from the damage of drought stress (Dai et al., 2007). The known genes and transcription factor families that have been proven to regulate the plant response to drought stress are AP2/ERF, WRKY, MYB, NAC, NAP, and bZIP (Wu et al., 2017; Mao et al., 2017; Tang et al., 2017; Butt et al., 2017; Zhu et al., 2018; Sun et al., 2018). Understanding the regulation of gene expression in response to drought stress is important to develop drought-resistant rice varieties. About 26 out of 184 candidate genes obtained from the QTL regions were selected for identifying their expression between drought-resistant parent (Kaybonnet) and drought sensitive parent (ZHE733) under DS conditions. These 26 candidate drought resistance genes comprise of 15 candidate genes with known annotations to be responsive to drought stress, and 11 candidate loci comprising of genes from the traits with high LOD and PVE with unknown annotations (Supplementary Table 3).

Out of 15 annotated candidate genes, LOC_Os01g66270 (ERF/Ethylene Response Factor) and LOC_Os10g41460 (MYB protein) showed up-regulation in Kaybonnet relative to ZHE733 under DS conditions when compared to WW conditions (Fig. 9C). All other 13 candidate genes, LOC_Os02g54160 (APETALA2/ERF transcription factor), LOC_Os03g56280 (Malate dehydrogenase), LOC_Os04g44190 (Light reaction photosystem II), LOC_Os05g49010 (APETALA2/ERF transcription factor), LOC_Os05g49210 (WRKY transcription factor), LOC_Os05g49240 (MYB protein), LOC_Os06g49080 (Brassinosteroid/BR signaling), LOC_Os07g02520 (Regulation of sugar partitioning in carbon-demanding young leaves and developing leaf sheaths), LOC_Os08g06344 (DEAD-box RNA helicase), LOC_Os09g08120 (Carbohydrate metabolic process), LOC_Os11g30760 (Galactosyltransferase activity), LOC_Os12g02700 (Late embryogenesis abundant/LEA protein), and LOC_Os12g29330 (No apical meristem/NAM protein domain) even though showed down-regulation in Kaybonnet relative to ZHE733 under DS conditions when compared to WW conditions do not support the phenotypic traits associated with each loci. These genes are inherently up-regulated in WW (Fig. 9A) and DS conditions (Fig. 9B) compared to ZHE733, so even though the relative amount was down-regulated in DS compared to WW (Fig. 9C), the intrinsic gene expression values of that genes are higher in Kaybonnet compared to ZHE733 suggesting a cause for better DS phenotype of Kaybonnet compared to ZHE733.

In order to verify the results from QTL mapping for two candidate regions for the traits, the loci high LOD and PVE score were studied for gene-expression between Kaybonnet and ZHE733. One of the region for

the trait PHC has LOD score 24.42 and PVE 13.81%. The window region of 25 Kb upstream and downstream was selected where the polymorphic SNP was detected on chromosome 2. This region has 6 candidate genes, LOC_Os02g44590, LOC_Os02g44599, LOC_Os02g44610, LOC_Os02g44620, LOC_Os02g44630, and LOC_Os02g44642 (Supplementary Table 3) with unknown annotations. The second region selected for gene-expression study also had high LOD score 21.82 and PVE 8.49% and is associated with traits BYD; and the region spanning the polymorphic marker selected included 5 genes: LOC_Os10g07030, LOC_Os10g07040, LOC_Os10g07050, LOC_Os10g07060, and LOC_Os10g07080 with unknown annotations on chromosome 10.

Four candidate genes (LOC_Os02g44590, LOC_Os02g44610, LOC_Os10g07040, and LOC_Os10g07080) out of 11 candidate genes with unknown annotations showed up-regulated in Kaybonnet relative to ZHE733 under DS conditions when compared to WW conditions (Fig. 8C). Seven other candidate genes (LOC_Os02g44599, LOC_Os02g44620, LOC_Os02g44630, LOC_Os02g44642, LOC_Os10g07030, LOC_Os10g07050, and LOC_Os10g07060) even though showed down-regulation in Kaybonnet relative to ZHE733 under DS conditions when compared to WW conditions (Fig. 8C), but the relative gene expression under DS (Fig. 8B) and WW conditions (Fig. 8A) showed higher expression in Kaybonnet and ZHE733, suggesting that higher intrinsic values of candidate genes in Kaybonnet compared to ZHE733 are probably enough for the traits. LOC_Os10g07040 and LOC_Os10g07080 up-regulated in Kaybonnet relative to ZHE733 under DS conditions when compared to WW conditions (Fig. 8C) and also showed higher relative gene expression under DS (Fig. 8B) and WW conditions (Fig. 8A) in Kaybonnet.

Among the candidate drought resistance genes not annotated to drought stress function, LOC_Os10g07040 showed high up-regulation in Kaybonnet compared to ZHE733 under DS and WW conditions (Figs. 9A, 9B, 9C), correlated with chalcone synthase, according to the MSU rice reference genome annotation release 7.0 that is involved in the drought stress response in rice (Hu et al., 2017), Arabidopsis (Nakabayashi et al., 2013), and tobacco (Hu et al., 2019). The other candidate drought resistance gene that also showed high up-regulated in Kaybonnet compared to ZHE733 under DS and WW conditions (Figs. 9A, 9B, 9C) was LOC_Os10g07080 is related to myosin (Jiang et al., 2004) and transposon protein (Cho et al., 2017) that regulates cell growth and developmental processes in rice. These candidate genes could be functioning in a cumulative manner in order to show a measurable positive effect on improving drought resistance in rice and the effect of genes can further be exploited to develop drought resistant cultivar.

A large number of genes were up-regulated in Kaybonnet (drought resistant parent), indicating that the drought resistant cultivar had higher capability to modulate drought resistance genes when exposed to DS conditions, thereby enhancing its resistance level compared to drought sensitive parent (ZHE733). Modulation of a higher number of up-regulated expressed genes with different transcription factor gene families is a crucial characteristics of drought resistant genotypes. Similar results were also obtained by Hayano-Kanashiro et al. (2009) who showed that drought-resistant maize genotypes induced more genes compared to the sensitive genotypes under DS conditions. All 15 candidate drought resistance genes identified within QTL regions have been strongly associated with direct roles in drought stress

resistance. For example, transcription factors MYB, NAP, NAC, ZIP, and APETALA2/ERF are responsive to dehydration induced by water deficit conditions (Tang et al., 2019; Baldoni et al., 2015; Li et al., 2015; Dai et al., 2007; Ma et al., 2009; Yang et al., 2012; Xiong et al., 2014; Quan et al., 2010; Lata and Prasad, 2011; Mizoi et al., 2012; Licausi et al., 2013; Phukan et al., 2017; Nakashima et al., 2007; Hu et al., 2008; Yokotani et al., 2009; Zheng et al., 2009; Jeong et al., 2010). These results provide strong evidence for genes expressed under DS conditions being involved in various physiological, biochemical, and molecular processes within the rice, in order to reduce the effects of drought stress, thereby enhancing their ability to resist the drought stress and maintain their grain yield production under DS conditions. Therefore, the up-regulation of the drought genes in Kaybonnet compared to ZHE733 provide important information to characterize the function of candidate drought resistance genes and to understand the drought stress mechanisms in rice.

Candidate genes within QTL regions involved in regulatory response to drought include a large family of genes expressed under DS conditions. Proteins expressed by known and candidate drought resistance genes played important roles in (1) cellular protection, including structural adaptation and osmotic adjustment, and (2) drought responses by interaction with other proteins and transcription factors, such as MYB, NAP, NAC, bZIP, and APETALA2/ERF. Under DS conditions, in drought resistant genotype Kaybonnet, exogenous ABA significantly improved the expression of ABA biosynthetic genes suggesting Kaybonnet genotype must be maintaining the water potential and cellular activity of the cell by closing the stomata.

Based on the RT-qPCR results, it may also be suggested that there is a correlation between gene expression, transcriptional regulation, and resistance to drought across resistance and sensitive genotypes. Therefore, the up-regulation of the drought genes and novel candidate genes in Kaybonnet compared to ZHE733 provided an important information to characterize the function of candidate drought resistance genes. All in all, these results enhance our understanding of the role of candidate drought resistance genes in the regulation of drought stress response, and this research has also revealed a number of potential candidate drought resistance genes that could be used to develop rice cultivars with greater drought resistance.

Genetic diversity in 26 loci across K/Z RIL population

The 26 drought loci described earlier are distributed across all 12 chromosomes with two distinct clusters on chromosome 2 (six loci) and chromosome 10 (five loci). The up- and down-stream region of the 26 drought loci was used for dissecting the genetic diversity in- and surrounding these loci in the diversity panel. Depending upon the proximity to the nearest neighbor gene, the region of interest was defined between 400 bp to 22Kbp for upstream, and 800 bp to 18Kbp for the downstream region.

We identified a total of 7,475 SNPs across these 26 loci in the 200-genotype of K/Z RIL population. The diversity panel represents six distinct subpopulations (Ind, *indica*; Aus, *aus*; TeJ, *temperate japonica*; TrJ, *tropical japonica*; Adm, *admixture* and Aro, *aromatic*). Each individual SNP has multiple effects on individual loci depending upon the 'impact', 'functional class', 'type' and 'region' where it is present as

shown (Supplemental Table 7). The SNPs were predominantly found in the upstream/promoter and intergenic regions. Five SNPs resulted in a stop codon gained in three loci (one stop gained in LOC_Os09g08120.1 and two stops gained in LOC_Os10g07030.1 and LOC_Os10g07050.1 each). SNP annotation showed that although most variants are moderate/modifying in effect, there are seven high impact and 200 low impact SNP effects that are potential candidates for further testing and validation. Moreover, the two QTLs on chromosomes 2 and 10 containing six and five loci, respectively show a distinct pattern of variation as compared to the overall variation in 26 loci.

To determine how the variation present in these 26 loci differentiates the 200 genotypes, we performed a principal component analysis (PCA) on the vcf files containing the SNP datasets. We performed three separate runs of PCA for i) all 26 loci ii) six-loci cluster on chromosome 2 and iii) five-loci cluster on chromosome 10 containing 7,475 SNPs, 565 SNPs and 4,375 SNPs, respectively (Supplemental Figure 8A-C). In the PCA for 26 loci, the first principal component (PC1) explains 34% of the variation in the data, and clearly separates a subset of *indica* subpopulation from the rest of the genotypes (Supplemental Figure 8.A). This is in contrast to the PCA performed for 6.5 million SNPs representing the whole genome where the *indica* subpopulation segregates as a single group (Gill et al. unpublished data). SNPs on chromosome 10 cluster also show a similar pattern of separation of *indica* subpopulation into two distinct groups with PC1 explaining 49% of the variation in the data (Supplemental Figure 8.C). However, the SNPs on chromosome 2 cluster show an interesting pattern where PC1 clearly separates the *japonicas* (both temperate and tropical) from the *indica* and *aus* subpopulations, and explains 84% of the variation in the data in doing so (Supplemental Figure 8.B). This indicates the presence of novel variation in the six drought loci on chromosome 2 (LOC_Os02g44590, LOC_Os02g44599, LOC_Os02g44610, LOC_Os02g44620, LOC_Os02g44630, LOC_Os02g44642) that distinguish the *japonicas* from the *indica* subpopulations.

Conclusions

In the RIL population, all the morphological traits, grain yield components, and root architectural traits showed normal frequency distribution, revealing quantitative inheritance. Furthermore, a positive correlation between FG-DS with most of the morphological traits, the other grain yield components, and the major root architectural traits under ABA conditions indicate that the rice drought-resistant plants maintain their grain yield under DS conditions by developing cell elongation, maintaining cellular membrane integrity, and regulation of osmotic stress tolerance via ABA-mediated cell signaling. QTL analysis was performed with 4133 SNPs markers by using QTL IciMapping. A total of 41 QTLs and 184 candidate genes within the QTL regions were identified for drought-related traits. The RT-qPCR results revealed that high number of genes were up-regulated in Kaybonnet as the drought-resistant parent, including 15 candidate drought resistance genes within QTL regions with known annotations showed higher intrinsic values in Kaybonnet, and two candidate genes with unknown annotations. Candidate genes identified within the QTL regions contribute to drought resistant traits and an understanding of the regulation of gene expression in response to drought stress, which is important to develop drought-resistant rice varieties.

Abbreviations

ABA, abscisic acid; BY, biological yield; DRN, deep root number; DR, drought resistant; DS, drought stress; EST, expressed sequence tag; FG, filled grain per panicle number; FLW, flag leaf width; GBS, genotyping by sequencing; HD, heading day; LR, leaf rolling score; MAB, marker assisted breeding; MAS, marker-assisted selection; MPSS, massively parallel signature sequencing; PCA, principal component analysis; PL, panicle length; PH, plant height; PPB, primary panicle branch number; QTLs, quantitative trait locus; RIL, recombinant inbred line; RFW, root fresh weight; RL, root length; RSR, root to shoot ratio; SRN, shallow root number; SNP, single nucleotide polymorphism; SP, spikelet per panicle number; TN, tiller number; TRN, total root number; WW, well-watered.

Declarations

Ethical approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Availability of supporting data

The supporting data will be available upon request.

Competing interests

The authors declare that they have no competing interests. The authors declare that the research was conducted in the absence of any commercial or financial relationships.

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Authors' contributions

Conceptualization, A.P., J.T., and A.K.; data curation, Y.D., A.K., C.R., J.T., N.B., and A.P.; formal analysis, Y.D., J.T., C.G., and N.G.; funding acquisition, A.P., J.T., and J.A.; investigation, Y.D., A.K., J.T., C.G., C.R. and N.G.; methodology, Y.D., J.T., A.K., and A.P.; project administration, A.P., J.T., J.A., Y.D., and A.K.; resources, A.P., J.T. and A.K.; supervision, A.P.; validation, A.P. and J.T.; writing—original draft preparation, Y.D., J.T.,

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Figures

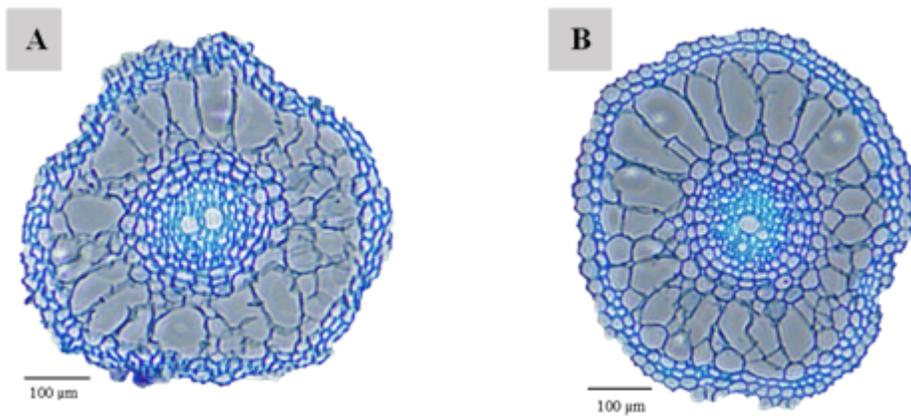


Figure 1

Root anatomy of Kaybonnet (A) and ZHE733 (B)

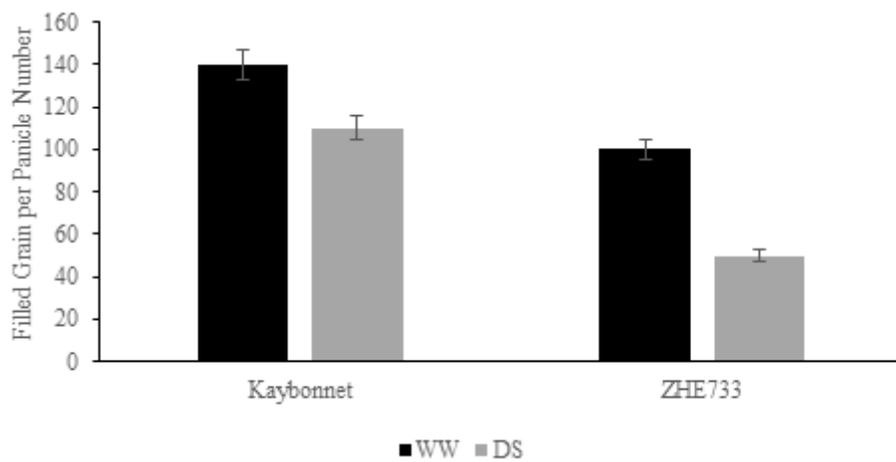


Figure 2

Number of filled grains per panicle in Kaybonnet and ZHE733. Kaybonnet maintained higher number of filled grains under DS than ZHE733

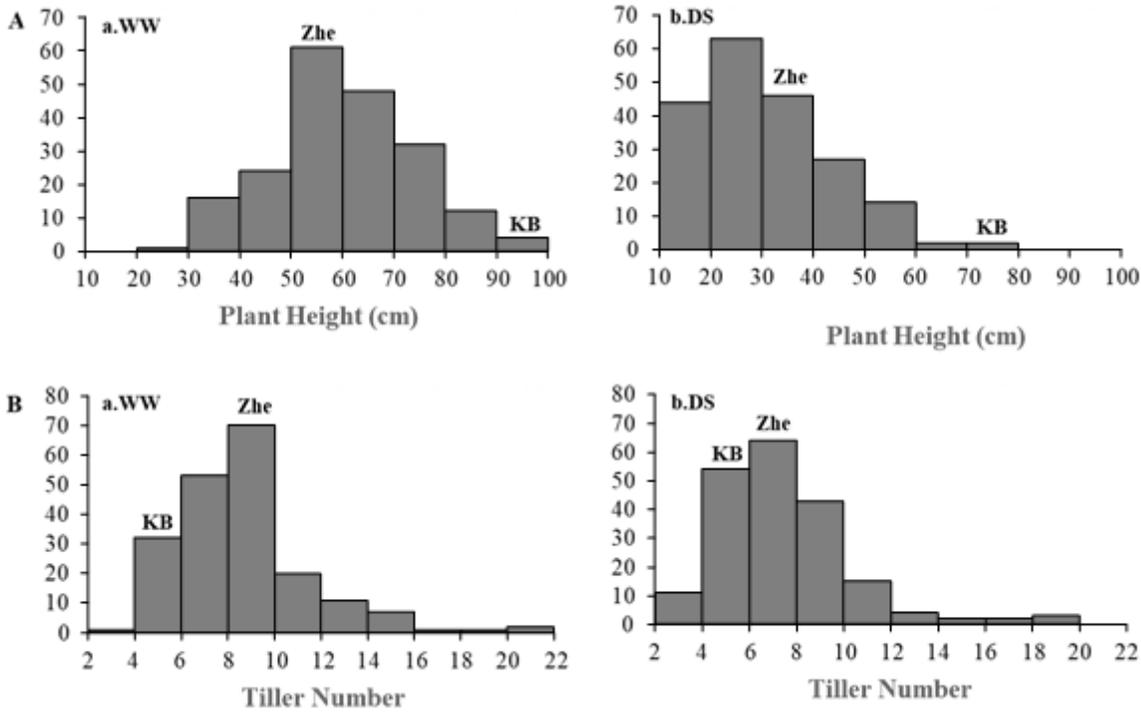


Figure 3

Frequency distribution of plant height (A) and productive tiller number (B) under WW (a) and DS (b) conditions

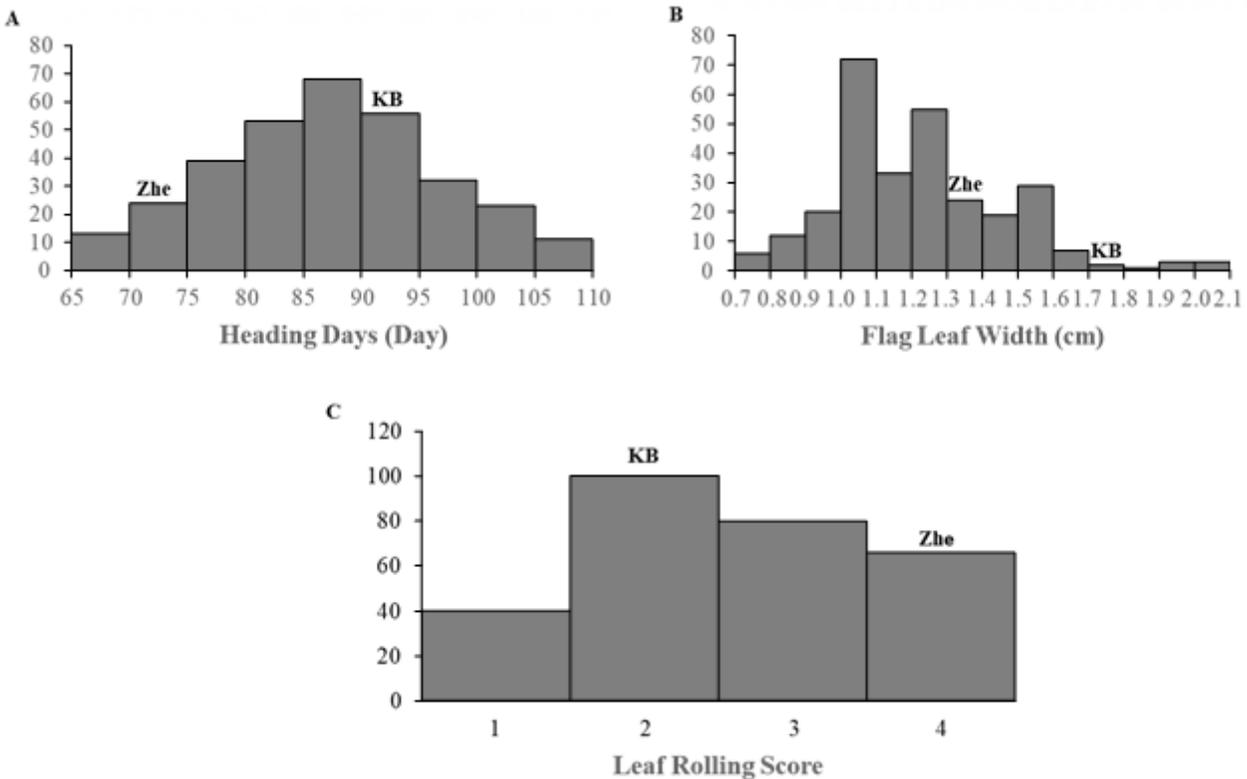


Figure 4

Frequency distribution of heading days (A), flag leaf width (B), and leaf rolling score (C)

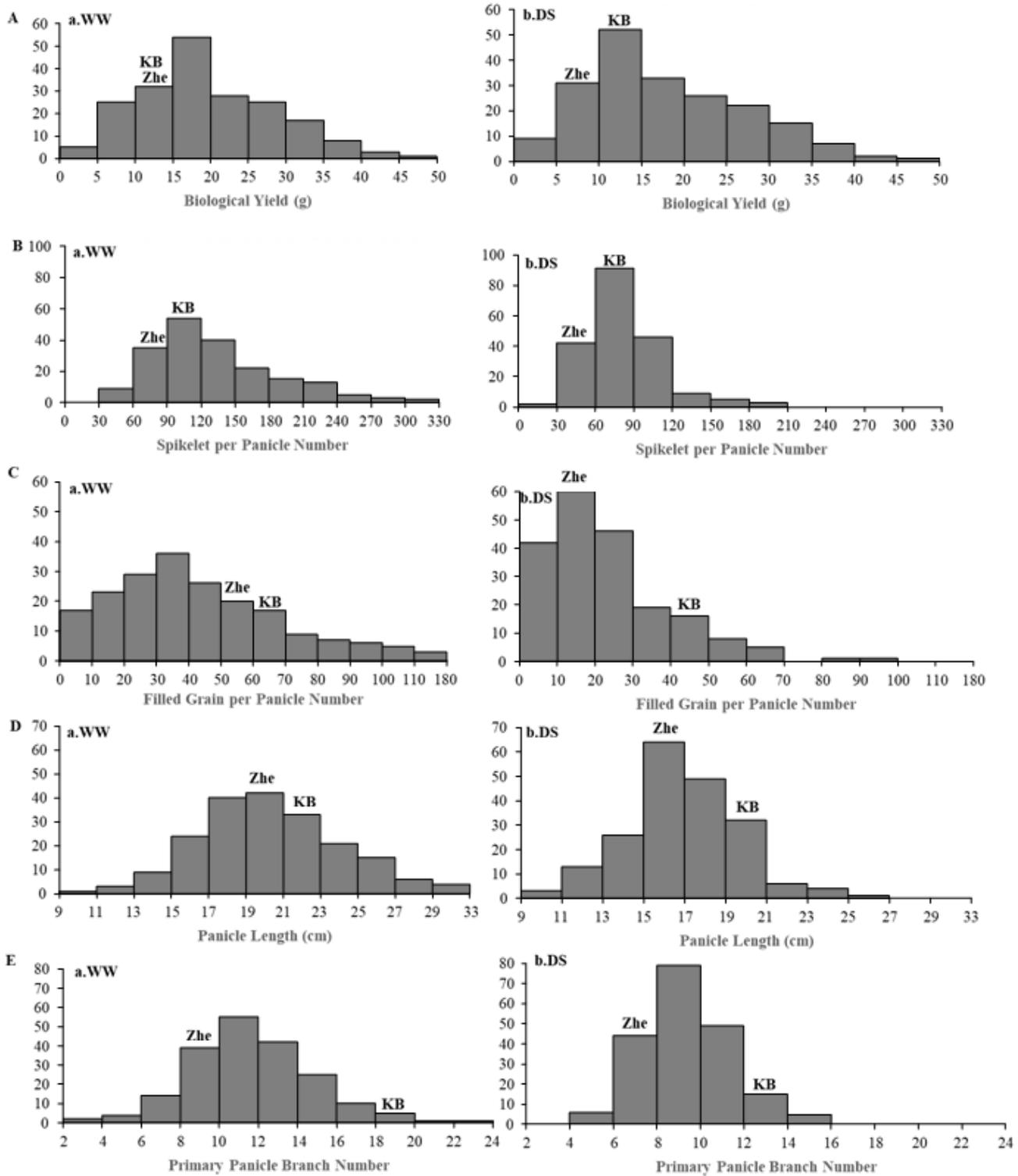


Figure 5

Frequency distribution of biological yield (A), spikelet per panicle number (B), filled grain per panicle number (C), panicle length (D), and primary panicle branch number (E) under WW (a) and DS (b) conditions

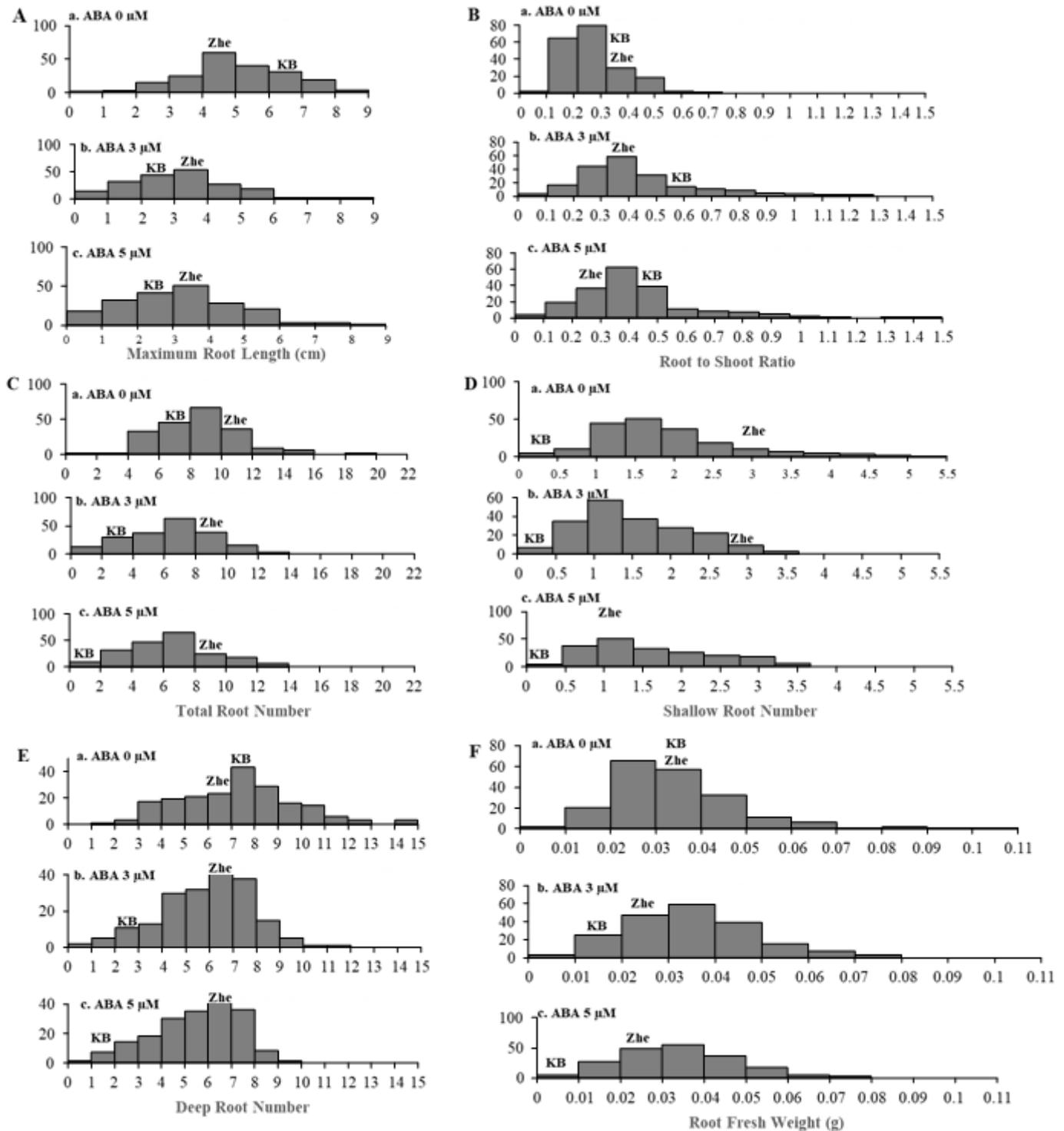


Figure 6

Frequency distribution of root length (A), root to shoot ratio (B), total root number (C), shallow root number (D), deep root number (E), and root fresh weight (F) under control (ABA 0 μM) (a), ABA 3 μM (b), and ABA 5 μM (c)

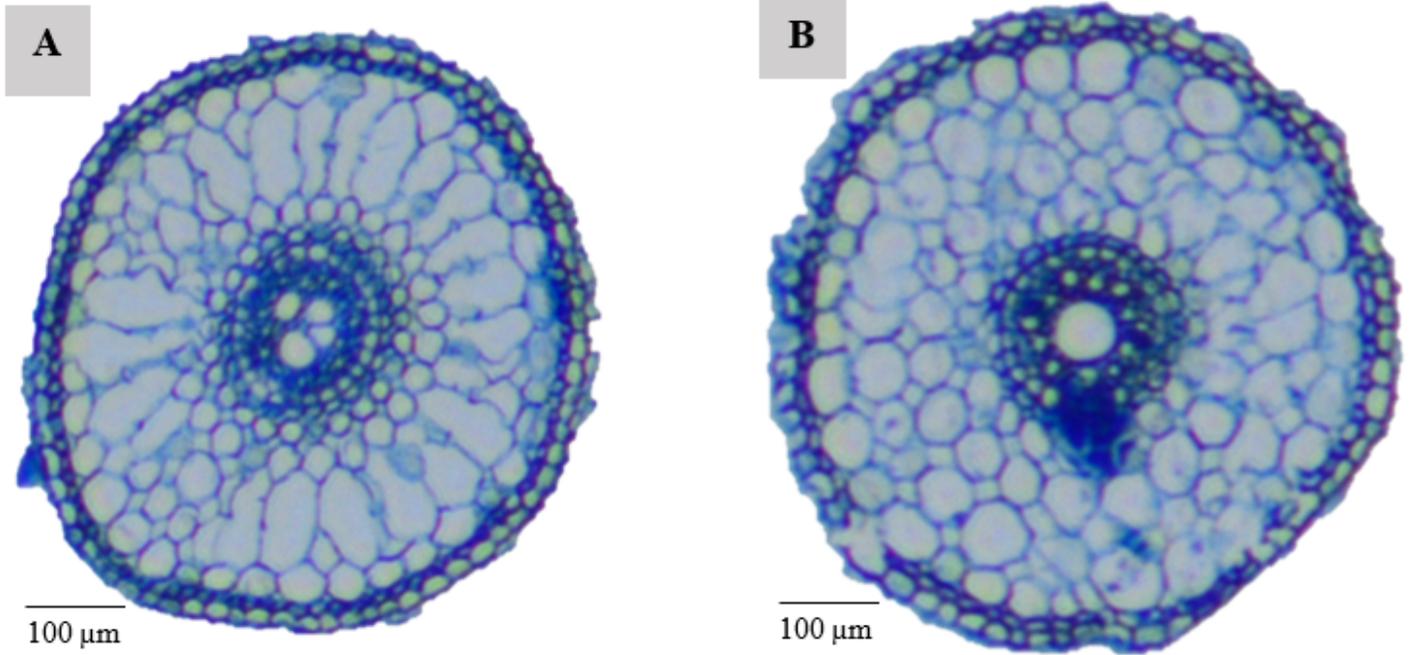


Figure 7

Root anatomy of 100162 as drought resistance line (A) and 100170 as drought sensitive line (B).

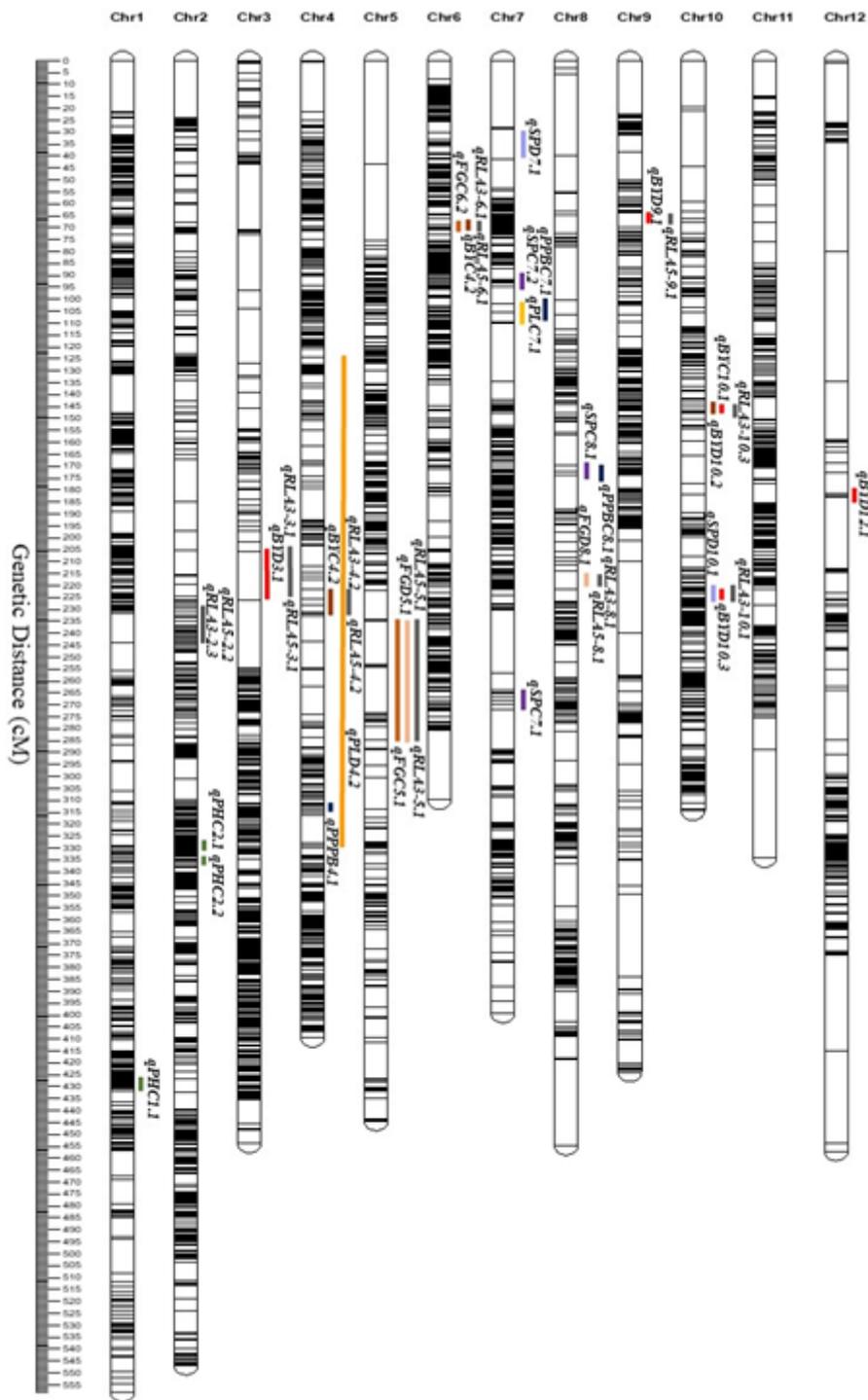


Figure 8

QTLs location of morphological and yield traits under DS and WW conditions and root architectural traits under ABA conditions on the 12 rice chromosomes. The genetic distance (cM) are shown on the left of the chromosome (chr)

Supplementary Files

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