

Evaluation of GBLUP and Bayes-Alphabet Based on Different Marker Density For Genomic Prediction in Alpine Merino Sheep

Shaohua Zhu

Chinese Academy of Agricultural Sciences Lanzhou Institute of Husbandry and Pharmaceutical Sciences

Tingting Guo

Chinese Academy of Agricultural Sciences Lanzhou Institute of Husbandry and Pharmaceutical Sciences

Chao Yuan

Chinese Academy of Agricultural Sciences Lanzhou Institute of Husbandry and Pharmaceutical Sciences

Jianbin Liu

Chinese Academy of Agricultural Sciences Lanzhou Institute of Husbandry and Pharmaceutical Sciences

Jianye Li

Chinese Academy of Agricultural Sciences Lanzhou Institute of Husbandry and Pharmaceutical Sciences

Mei Han

Chinese Academy of Agricultural Sciences Lanzhou Institute of Husbandry and Pharmaceutical Sciences

Hongchang Zhao

Chinese Academy of Agricultural Sciences Lanzhou Institute of Husbandry and Pharmaceutical Sciences

Yi Wu

Chinese Academy of Agricultural Sciences Lanzhou Institute of Husbandry and Pharmaceutical Sciences

Weibo Sun

Chinese Academy of Agricultural Sciences Lanzhou Institute of Husbandry and Pharmaceutical Sciences

Xijun Wang

Gansu provincial Sheep Breeding Technology Extension Station

Tianxiang Wang

Gansu provincial Sheep Breeding Technology Extension Station

Jigang Liu

Gansu Provincial Sheep Breeding Technology Extension Station

Christian Keambou Tiambo

Centre for Tropical Livestock Genetics and Health, International Livestock Research Institute

Yaojing Yue

Chinese Academy of Agricultural Sciences Lanzhou Institute of Husbandry and Pharmaceutical Sciences

Bohui Yang (✉ yangbh2004@163.com)

Chinese Academy of Agricultural Sciences Lanzhou Institute of Husbandry and Pharmaceutical Sciences

Methodology article

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1 **Evaluation of GBLUP and Bayes-Alphabet Based on Different Marker Density for Genomic**
2 **Prediction in Alpine Merino Sheep**

3 Shaohua Zhu^{1,2}, Tingting Guo^{1,2}, Chao Yuan^{1,2}, Jianbin Liu^{1,2}, Jianye Li^{1,2}, Mei Han^{1,2}, Hongchang
4 Zhao^{1,2}, Yi Wu^{1,2}, Weibo Sun^{1,2}, Xijun Wang³, Tianxiang Wang³, Jigang Liu³, Christian Keambou
5 Tiambo⁴, Yaojing Yue^{2*} and Bohui Yang^{1*}

6 ¹.*Animal Science Department, Lanzhou Institute of Husbandry and Pharmaceutical Sciences,*

7 *Chinese Academy of Agricultural Sciences, Lanzhou, 730050, China.*

8 ².*Sheep Breeding Engineering Technology Center, Chinese Academy of Agricultural Sciences,*

9 *Lanzhou, 730050, China.*

10 ³.*Gansu Provincial Sheep Breeding Technology Extension Station, Sunan, 734400, China.*

11 ⁴. *Centre for Tropical Livestock Genetics and Health (CTLGH), International Livestock Research*
12 *Institute, Nairobi, 00100, Kenya.*

13 *** Correspondence:**

14 Yaojing Yue: Tel: +86-0931-2115273; Fax: +86-0931-2115272; E-mail: yueyaojing@126.com

15 Bohui Yang: Tel: +86-0931-2115272; Fax: +86-0931-2115272; E-mail: yangbh2004@163.com

16 **Email addresses:**

17 Shaohua Zhu: zhu87932890@126.com

18 Tingting Guo: guotingting@caas.cn

19 Chao Yuan: yuanchao@caas.cn

20 Jianbin Liu: liujianbin@caas.cn

21 Jianye Li: lijianye218@163.com

22 Mei Han: 82101186189@caas.cn

23 Hongchang Zhao: 18837101296@163.com

24 Yi Wu: 1593050417@qq.com

25 Weibo Sun: swb887246@126.com

26 Xijun Wang: 1091606575@qq.com

27 Tianxiang Wang: zhangyesyz@163.com

28 Jigang Liu: hcywtx@sina.com

29 Christian Keambou Tiambo: C.tiambo@cgiar.org

30 **Abstract**

31 **Background**

32 The marker density, the heritability level of trait and the statistical models adopted are critical to the
33 accuracy of genomic prediction (GP) or genomic selection (GS). The studies on the impact of the
34 above factors on accuracy of GP are usually focused on the comparison and discussion of simulated
35 datasets. If the potential of GS is to be fully utilized to optimize the effect of breeding and selection,
36 it is essential to incorporate these factors into real data for understanding their impact on GP
37 accuracy, more clearly and intuitively. Herein, we studied the genomic prediction of six wool traits
38 of sheep by two different models, including genomic best linear unbiased prediction (GBLUP), and
39 Bayes-Alphabet. We adopted 5-fold cross-validation to perform the accuracy evaluation based on
40 the genotyping data of Alpine Merino sheep (n=821).

41 **Results**

42 The GP accuracy of the six traits was found to be between 0.28 and 0.60, as demonstrated by the
43 cross-validation results. We showed that the accuracy of GP could be improved by increasing the
44 marker density, which is closely related to the model adopted and the heritability level of the trait.
45 Moreover, based on two different marker densities, it was derived that the prediction effect of
46 GBLUP model for traits with low heritability was better (GBLUP has the highest accuracy of 28.57%
47 higher than Bayes-Alphabet); while with the increase of heritability level, the advantage of Bayes-
48 Alphabet would be more obvious, therefore, different models of GP are appropriate in different traits.

49 **Conclusion**

50 This is the first study of optimization of GP has been applied to the domesticated Alpine Merino
51 sheep populations. The main aim was to study the influence and interaction of different models and
52 marker densities on GP accuracy. These findings indicated the significance of applying appropriate

53 models for GP which would assist in further exploring the optimization of GP.

54 **Keywords:** Genomic prediction; Alpine Merino sheep; Wool traits; GBLUP; Bayes-Alphabet;
55 Marker density

56 **Background**

57 The advancement in the field of quantitative genetics and molecular biology has improved the
58 selection and breeding methods of domestic animals [1]. Meuwissen et al. 2001 proposed a more
59 advantageous selection method, known as genomic selection (GS) or genomic prediction (GP) [2].

60 This method combines the genome-wide single nucleotide polymorphism (SNP) with phenotypic
61 data and implicates them for genetic evaluation [3-5]. It was first applied to the dairy cows [6] and
62 is now widely used in other model animals such as beef cattle [7], pigs [8], goats [9], and sheep [10],
63 aquatic animals like Atlantic salmon[11], rainbow trout[12], and plants [13, 14], such as wheat [15]
64 and alfalfa [16]. GS has made a substantial contribution to the modern breeding process, as
65 compared to traditional methods; the main advantages of this method include improved estimation
66 accuracy of breeding value (BVs) [17, 18], increased genetic progress, and reduced breeding costs
67 [19, 20]. With the successive publish of various livestock genome sequences and the continuous
68 upgrade of commercial SNP microarrays, different types and densities of microarrays have been
69 adopted in the GP of different livestock [21]. Accuracy and cost are generally the most critical
70 factors in GP, compared to low-density SNP microarrays, the high-density SNP microarrays could
71 accommodate more SNP sites that may lead to higher coverage of the genotype data [22]. However,
72 the cost of the high-density microarray was comparatively higher. In contrast, although the low-
73 density SNP microarrays has fewer SNP sites, it is more applicable in population breeding with a
74 huge dataset due to its lower cost. Both the methods have their own pros and cons and therefore, it
75 is difficult to conclude which density microarray is best suitable for GP.

76 For the first time, Meuwissen et al. 2001 proposed a GS based on Bayes method, which includes
77 BayesA and BayesB [2]. Based upon this approach, several other methods were also derived such
78 as BayesC π method [23], Bayesian least absolute shrinkage and selection operator (Bayesian
79 LASSO) method [24]. Subsequently, in 2013, Gianola summarized these methods as the Bayes-
80 Alphabet method [25]. In fact, the assumptions and strategies adopted by these methods are different.
81 The BayesA assumes that all SNPs have genetic effects and the variance of marker effects should
82 obey the t-distribution, whereas BayesB assumes that only a small proportion of SNPs have an effect.
83 Furthermore, the BayesC π is similar to BayesB, and estimates the proportion of sites with no effect
84 of π in the model. The Bayesian LASSO method assumes that all markers have effects, and the
85 variance of marker effects obeys the double exponential distribution also known as Laplace
86 distribution [25]. VanRaden et al. 2008, proposed another calculation method for GP and named it
87 as genomic best linear unbiased prediction (GBLUP). It calculates the relationship matrix of
88 individuals via genome-wide genotype information instead of traditional pedigree information.
89 Herein, the matrix denoted as G is applied to replace the A matrix in BLUP, to estimate the BVs
90 according to the BLUP method [26]. Another novel approach known as single-step GBLUP
91 (SSGBLUP or HBLUP) has been developed based on GBLUP [27]. This method integrates the
92 phenotype, pedigree and genomic information into a model, and combines the traditional kinship
93 matrix A with the genome relationship matrix G according to different weights to construct a new
94 relationship matrix H , then simultaneously estimate the genetic effects of all individuals (including
95 individuals with and without genotypes). Although there are various GP methods available, no
96 method could be suitable for all traits. Therefore, in this study, two methods based on Bayes and
97 GBLUP models were adopted to study the prediction accuracy of real data for different wool traits,

98 aiming to screen ideal GP models.

99 As an important domestic animal, sheep is one of the earliest domestic animals reared by humans
100 [28] and provides diverse resources such as mutton, wool, skin, and milk. Merino and Merino-
101 derived sheep breeds are distributed globally [29]. As the object of the current study, the Alpine
102 Merino sheep has Australian Merino and Tibetan sheep lineage. Thanks to their adaptation in high-
103 altitude hypoxia and excellent wool quality, they quickly adapted to the freezing Qinghai-Tibet
104 Plateau, living in high altitude and cold conditions for generations [30]. The length and strength of
105 the staple and fiber diameter are closely related to the wool quality and are the important economic
106 traits of fine-wool sheep. Therefore, adopting genome analysis to explore wool traits is crucial for
107 the selection and development of this population. However, the application of GP for this population
108 has just started obtain their genomic information through SNP microarray, and combined with
109 phenotypic datasets closely related to wool traits, then, adopted different methods of BVs estimation
110 and compare the results, in particular, genome analysis could be performed from two aspects include
111 genetic effects of markers and methods of GP. This could make a great contribution to the
112 application of GP and GS in Alpine Merino sheep population.

113 In the current study, two different densities of SNPs including low (50K) and high (630K) were
114 applied to estimate the genetic variance components of the Alpine Merino sheep datasets. Further,
115 based upon the SNP genotypes data, different models were adopted for GP and cross-validated to
116 compare the accuracy of different GP methods. The main purpose of this study is to investigate the
117 impact of different densities of SNP genotypes and different GP methods on the accuracy and
118 optimization methods of GP in Alpine Merino sheep populations.

119 **Results**

120 **Statistics and processing of phenotypic data**

121 A total of 6 wool traits were collected and the descriptive statistics of individual wool phenotype
122 data was presented in Table 1, including the abbreviation of each trait, the corresponding standard
123 error (S.E), the average value (represented by mean \pm S.D), and the number of individuals that were
124 effectively recorded (Numbers). For the wool traits, the standard deviation (S.D) ranged from 2.11
125 (FD) to 13.16 (SL), and the standard error (SE) ranged from 0.07 (FD) to 0.46 (SL).

126 **The polygenic heritability and the GP accuracy**

127 The phenotypic variance and the additive variance of the 6 wool traits based on L- and H-datasets
128 were estimated to calculate polygenic heritability (h^2). For L-datasets, heritability ranged from 0.37
129 (FER) to 0.70 (SL); and for H-datasets, heritability ranged from 0.29 (FER) to 0.68 (SL). The
130 estimated results of heritability (expressed as the proportion of additive variance in phenotypic
131 variance) shown in Table 3, states that SL was the highest and the FER was the lowest irrespective
132 of the L- or H-datasets. Moreover, the heritability estimated by L-datasets was slightly higher than
133 that of H-datasets for these 6 wool traits.

134 The GP accuracy was calculated using 5 methods based on two marker density datasets (Table 4).
135 For L-datasets, the GP accuracy of SL was the highest (0.59 for Bayesian LASSO model); and the
136 GP accuracy of FER was the lowest (0.28 for BayesA model). Correspondingly, for H-datasets, the
137 trait with the highest GP accuracy was also SL (0.58 for BayesA, BayesB, and Bayesian LASSO
138 model), and the trait with the lowest GP accuracy was FER (0.31 for BayesA model).

139 **Discussion**

140 **Genomic information and individual relationship matrix**

141 The analyses involved in this study are all based on genomic information obtained from genotyping

142 through microarrays, GP has replaced the traditional phenotype and pedigree information with the
143 dense markers, providing a new method to estimate genetic variance, which improves the accuracy
144 of prediction and selection [31]. Genomic information is not only suitable for a population with
145 pedigree information, but can also be applied to populations without pedigree information or
146 incorrect, incomplete and even missing genealogical records [32, 33]. In the GBLUP model, the
147 traditional individual relationship matrix A constructed by pedigree was replaced by the genome
148 matrix G , which represents the relationship between individuals more accurately, as it is based on
149 a dense genome-wide marker. More importantly, this may capture the genetic connections from
150 unknown common ancestors, because it represents confirmed gene sharing, and has advantages over
151 presumed or conceptualized ancestral sharing [4]. In GBLUP model, it was assumed that each SNP
152 has an effect, and the cumulative effect of SNPs obey a normal distribution [34], the assumption
153 might only be applicable to certain specific groups or traits. According to the hypothesis of Habier
154 et al., for some traits, only a few markers have a larger effect, while most markers have little or no
155 effect [23, 35]. Therefore, GBLUP may not be suitable for such trait, in other words, the GP accuracy
156 of GBLUP will be lower than other models, like the FD trait in current study, the GP accuracy (0.56
157 based on L-datasets) of the Bayesian LASSO model was higher than that (0.52 based on L-datasets)
158 of the GBLUP model. From the above results, GBLUP may not be applicable to FD traits and its
159 predictive ability may not achieve satisfactory results. Hence, it is necessary to adopt different GP
160 models. In the Bayes-Alphabet method, models such as BayesB and BayesC π assume that most of
161 the SNPs in the genome are located in regions without quantitative trait locus (QTL) and have no
162 effect [24]. while a small number of other SNPs existed in linkage disequilibrium (LD) together
163 with QTL, and accounts for most of the effect [34, 36]. According to reports, different Bayes-

164 Alphabet methods put forward a variety of prior hypotheses on the distribution of SNP effects (Table
165 2) [34]. In the current study, in addition to the GBLUP method, 4 typical Bayes-Alphabet methods
166 (BayesA, BayesB, BayesC π and Bayesian LASSO) were also used to compare the GP accuracy of
167 the 6 wool traits.

168 In most cases, GP suffers limitations while adopting the high-density or low-density SNP genomic
169 information, i.e., the number of marker effects that need to be estimated is often greater than the
170 number of individuals to be recorded. In this study, both the L-and the H-datasets showed that the
171 number (35,379 and 460,656) of markers was much larger than the number (821) of individuals.
172 Although many advanced statistical methods [37, 38] have been proposed to overcome this
173 challenge, the true distribution of QTL and SNP effects were unclear for many quantitative traits
174 [34]. Moreover, in contrast to L-datasets, the H-datasets microarrays contain more genomic
175 information, but it also involves more complex matrices and larger computation, which will
176 undoubtedly increase the cost of time and economy [36].

177 **Phenotypic statistics and estimation of heritability**

178 In the current study, the collected phenotypic statistics of wool traits were compared with the results
179 in previous reports: Moghaddar et al. collected 3000-8000 phenotypic records of various wool traits
180 from different breeds of sheep in 2014, including the Poll Dorset, White Suffolk and Border
181 Leicester. In their report, the statistical mean values of FD and FD_CV were 19.93 ± 5.39 and
182 19.26 ± 2.86 (mean \pm S.D) respectively. The statistical mean of SS and SL was 33.82 ± 9.82 ,
183 80.93 ± 13.06 , respectively [39]. In addition, according to the study by Hamadani (2019) et al. on
184 Rambouillet sheep [40], where they collected and recorded the wool traits of 4,108 samples from
185 1998 to 2007, the statistical mean value of FD and SL was 21.26 ± 0.03 (mean \pm S.E), 56.1 ± 0.05

186 respectively. The above comparison showed that the phenotypic statistics of the current study were
187 consistent with the earlier studies. It could be suggested that although the number of phenotypes
188 collected in this study was not as large as, the statistical values of phenotype measurement were still
189 reliable.

190 The additive and residual variance, and the heritability of the 6 wool traits of the Alpine Merino
191 sheep population were estimated. Daetwyler (2010) and Moghaddar (2014) et al. conducted the
192 genetic parameter estimation and GP studies on the sheep of multiple breeds including Merino,
193 Border Leicester, and White Suffolk. The results showed that the weighted average heritability of
194 SS and SL was in the range from 0.37 to 0.55 and 0.56 to 0.67, respectively. The weighted average
195 heritability of FD and FD_CV was between 0.62-0.75 and 0.47-0.57, respectively [39, 41]. In
196 addition, Safari (2005) and Fogarty (1995) et al. collected and summarized the genetic parameters
197 of 9 wool traits [42, 43]. Their results showed that the weighted average heritability of SS, SL,
198 CFWR, FD, FD_CV were 0.34, 0.46-0.48, 0.34-0.51 0.51-0.59 and 0.52, respectively. In the current
199 study, except for the slightly lower estimated value of FD (0.42-0.47), the other four wool traits
200 (Table 3) were close to the results reported in the previous literature. Especially, the SS (0.33-0.46)
201 was very close to them. The comparison with the previous literature suggested that the heritability
202 results estimated from the Alpine Merino dataset in the current study were reliable.

203 **GP results and accuracy of prediction**

204 If breeding scientists are to effectively apply genomic selection in their breeding programs, they
205 need to have a full understanding of the factors that affect the accuracy of the dataset predictions.
206 For effective application of GS and GP on sheep breeding programs, there should be a thorough
207 understanding of the factors affecting the accuracy of the dataset predictions. We collected 821

208 samples from the breeding program to investigate the influence and interaction of marker density
209 and GP on the accuracy of prediction. Previous studies suggested that the density of markers has an
210 essential impact on the accuracy of GP [44, 45]. Solberg and his collaborators (2008) adopted the
211 simulated data to analyze the correlation between accuracy and marker density, their results showed
212 that increasing the density of SNPs from 1 to 8 per centimorgan (cM) could improve the accuracy
213 of GP by 25 %[46]. but this did not mean that the accuracy could always improve with the increase
214 of marker density, in other words, there is a limit to this improvement. Heffner et al. (2011)
215 conducted a study using a wheat dataset and showed that with the increased density from 192 to
216 1,158 markers, the accuracy of GP could be improved by 10 %. However, when the marker density
217 increased from 192 to 384,it caused only a small increase in accuracy [47]. Most of the 10 %
218 improvement mentioned above occurred in the interval from 192 to 384 markers, and the increase
219 of the remaining markers did not significantly affect the accuracy. These results indicate that marker
220 density has a positive effect on the accuracy of GP, while the response of accuracy to density will
221 eventually stabilize [48].

222 Herein, we adopted the genome datasets based on the level of 50K and 630K microarray,
223 respectively. Table 1 shows that with the marker density increases, the improved accuracy of GP for
224 most traits, especially in FBS and FER, model Bayesian LASSO and BayesA increased by 12 and
225 11 %, respectively, while in other traits the accuracy was not significantly improved, such as CFWR
226 and FD_CV, the accuracy of GBLUP and BayesB increased only by 1 %; FBS and FER benefited
227 more from the increase in marker density than other traits, which could be explained by the fact that
228 quantitative genetic characteristics require more markers to accurately estimate their many small
229 effects of QTL[49]. Interestingly, there are exceptions in this study, for some traits, the accuracy

230 may even decrease: in FD trait, the accuracy of BayesA and Bayesian LASSO models were reduced
231 by 3 % and 5 %, respectively. Two reasons that may explain why increasing number of markers on
232 each chromosome led to a decrease in GP accuracy. Firstly, the number of markers in the microarray
233 is much larger than the number of samples, which may be due to excessively high density of markers
234 leading to the model overfitting [50]. Secondly, the increases in the number of markers will lead to
235 the addition of more unknown variables (marker effects) and a lack of accurate estimation. The
236 study from Fatemeh Alanoshahr et al. also showed that with the number of SNPs increased from
237 2000 to 3000, both BayesA and GBLUP model indicated a decrease in the accuracy of GP [51]. Our
238 results suggest that increasing the density of markers could indeed improve the GP accuracy, but it
239 is closely related to the trait itself. For traits with low heritability levels (FER and FBS), a small part
240 of the phenotypic variation was explained by additive effects[52], and the increase of marker density
241 may improve the accuracy of GP more obviously; correspondingly, for those traits with high
242 heritability levels (CFWR and FD), increasing the marker density has little benefit on the GP
243 accuracy, sometimes even has a negative impact on accuracy.

244 Among the 6 wool traits studied here, SL and FD_CV had the highest heritability ($h^2=0.53$ and
245 $h^2=0.58$, respectively), and their corresponding accuracy of GP was also the highest, which ranged
246 from 0.53 to 0.60 and 0.45 to 0.55, respectively. While for two traits with the lowest heritability,
247 FBS ($h^2=0.33$) and FER ($h^2=0.28$), the accuracy was 0.29 to 0.38 and 0.28 to 0.36, respectively,
248 which was lower than SL and FD_CV. For those traits with lower heritability, the correlation
249 between phenotypic value and genetic value will be lower, the effect value of markers distributed
250 across the genome may be estimated with lower accuracy [23], it suggested that higher heritability
251 has a positive effect on the accuracy of GP. Bolormaa et al. (2013) also reported that the prediction

252 of the trait with the highest heritability was more accurate [53], and also several studies have shown
253 that the accuracy of GP increases with the improved heritability[54, 55], the results of the current
254 study agreed with them. In addition, we found that for traits with low heritability, GBLUP had a
255 better prediction effect, whether it is adopting L- or H-datasets, but with the increase of heritability,
256 the advantage of GBLUP is not obvious. From Table 4, it could be observed that for the trait SL
257 with high heritability, the estimation accuracy of BayesB (0.58-0.60) and Bayesion LASSO (0.58-
258 0.59) models performed better, this may indicate that for some traits with high heritability, BayesB
259 and Bayesion LASSO assumes more reasonable distribution in marker effect, which leads to higher
260 prediction accuracy. Similar results were obtained in the study of Honarvar and his coworkers, based
261 on the simulation data of three different levels of heritability, they compared the accuracy of the
262 RRBLUP and bayesion-LASSO models, and the results showed that the GP accuracy of the
263 bayesion-LASSO model is higher than that of the RRBLUP model for these traits, but the former
264 has a more obvious advantage in traits with high heritability [56], and it should be noted that GBLUP
265 was equivalent to RRBLUP. In addition, the accuracy of GP was also related to the size and structure
266 of the reference group [57, 58]. We will collect and organize a larger dataset in future and try to take
267 the above factors into consideration in subsequent studies for better conclusive results.

268 **Conclusions**

269 To summarize, this study was based on two different densities of microarray genotyping data, using
270 Bayes-Alphabet (including BayesA, BayesB, BayesC π , Bayesion LASSO) and GBLUP model to
271 perform the GP. The heritability of 6 wool traits of Alpine Merino sheep was estimated, and the
272 accuracy of the BVs prediction of these traits under different conditions was evaluated through five-
273 fold cross-validation. To the best of our knowledge, this was the first study of optimization of GP

274 has been applied to the domesticated Alpine Merino sheep populations. We have observed that for
275 traits with low heritability (SS and FER), increasing the density of markers could improves the GP
276 accuracy, but it has little impact on traits with high heritability (SL), and even decreases the accuracy
277 (FD). The accuracy of the GBLUP model is generally higher than that of the Bayes-Alphabet model
278 for SS and FER, while with the improvement of heritability, the advantage of GBLUP is no longer
279 obvious. Therefore, from this study, we conclude that GBLUP is more suitable for traits with lower
280 heritability (FER and FBS), and Bayes-alphabet, especially BayesB and Bayesian LASSO, have
281 better GP effects for traits with high heritability (FD and SL), different GP models are applicable to
282 different traits.

283 **Methods**

284 **Animal resources and phenotypic data**

285 The original phenotypic dataset was obtained from the Sheep Breeding Technology Extension
286 Station of Gansu Province. These datasets consisted of 11,500 individuals based on 7 different herds
287 with information such as region (herd), gender, and date of birth. The individuals in the current
288 study included 821 Alpine Merino sheep (563 ewes and 258 rams) from HuangCheng pasture in
289 Gansu Province, China, all born between year, 2014 to 2018. This pasture was under the jurisdiction
290 of the Gansu Sheep Breeding Technology Extension Station which has a rigorously standardized
291 system of breeding and management, to ensure that all the individuals have unified feeding and
292 management conditions. The average age of each individual with phenotypic data was about 14
293 months. The wool traits involved in the current study were staple length (SL), clean fleece weight
294 rate (CFWR), average fiber diameter (FD), coefficient of variation of average fiber diameter
295 (FD_CV), staple strength (SS) and fleece extension rate (FER). The wool from individuals was

296 collected and evaluated according to the Agricultural Industry Standards of the People's Republic
297 of China (NO. NY/T 1236-2006). Wool samples (~150-200 grams) collected from the abdomen of
298 each individual, were weighed and stored in ziplock bags (Xingdeli Packaging Material Company
299 Ltd., Shenzhen, China). Within one week, the samples were sent to the National Animal and Rural
300 Ministry of Animal and Fur Quality Supervision and Inspection Center (Lanzhou, China) for
301 weighing, screening and quality identification of wool. Blood samples (~5 mL) were also collected
302 from each sheep from the jugular vein and immediately transferred to the vacutainer blood
303 collection tube (Yuli Medical Equipment Company Ltd., Jiangsu Province, China). Blood samples
304 were stored at -20°C for further genotyping [59]. The statistics used to estimate variance
305 components and GP of each wool trait are presented in Table 1.

306 **Genotypic data and quality control**

307 The customized Affymetrix HD 630K microarray was employed as the datasets for the genotype of
308 high-density SNP genotypes (H-datasets) for the Alpine Merino sheep. The genotyping platform for
309 analysis was based on the array plate processing workflow of GeneTitan system (Santa Clara,
310 California, USA) from Thermo Fisher (Affymetrix). The sites in the Illumina Ovine SNP 50K
311 microarray were screened out from the Affymetrix HD 630K microarray and used as the datasets of
312 low-density SNP genotypes (L-datasets). The H-and L-datasets were pre-processed using PLINK
313 v1.9b4 software prior to the statistical analysis and variance component estimation [60]. The SNPs
314 were eliminated with call rate (geno) below 95 %, minor allele frequency (MAF) below 0.01, which
315 seriously deviated from the Hardy Weinberg Equilibrium with a P-value below 10^{-6} . Here, the X, Y
316 chromosomes and mitochondrial chromosomes were excluded from the analysis. In addition, Beagle
317 software (version number; 12Jul19.0df) was used to impute the missing SNPs [61]. After quality

318 control and impute, a total of 821 individuals with 460,656 autosomal SNPs were retained for H-
319 datasets, and a total of 821 individuals with 35,379 autosomal SNPs for L-datasets.

320 **Statistical methods for GP**

321 We explored the application of SNP datasets of different densities in genome evaluation and further
322 compared the accuracy of GP adopting 5 different models, including Bayes-Alphabet (BayesA,
323 BayesB, BayesC π , Bayesian LASSO) and GBLUP. Six wool traits from 821 samples were used to
324 first, estimate the variance of each component, including the additive and residual variance; second,
325 five different models were adopted to perform GP, and its accuracy was compared via 5-fold cross-
326 validation, and all these models were evaluated in SNP datasets of H-and L-datasets. Replicate
327 measurements were not available for the individuals so that the effects of permanent environmental
328 were not modeled. The samples involved were from different herds and genders. These factors
329 altered the phenotype in a fixed pattern, and hence the system environmental effects were added to
330 the framework.

331 The statistical methods of Bayes-Alphabet involved can be written as:

$$332 \quad \mathbf{y} = \mathbf{Xb} + \sum_j^n \mathbf{Z}_{ij}\alpha_j + e \quad (1)$$

333 Here, \mathbf{y} represents the corrected phenotypic value of individuals, \mathbf{Xb} refers to a fixed term, and
334 \mathbf{b} contains a vector of 3 effects, including herds, genders, and mean of population. \mathbf{Z}_{ij} represents
335 the genotype of individual i at site j , and α_j represents the effect value of site j , and therefore
336 $\sum_j^n \mathbf{Z}_{ij}\alpha_j$ refers to the BV corresponding to individual i , e to the vector of residual effects.
337 According to the method from Meuwissen et al. and Habier et al [2, 23], adopted the R package
338 "BGLR" (<https://github.com/gdlc/BGLR-R>) to estimate the effect of markers [62]. The hypothetical
339 distribution of all markers' effects in different Bayes methods and the formula of effect distribution

340 are shown in Table 2.

341 The methods of GBLUP involved in the current study confirms to a linear model.

$$342 \quad \mathbf{y} = \mathbf{Xb} + \mathbf{Zu} + e \quad (2)$$

343 In Bayes-Alphabet model, in equation 2, \mathbf{y} , b , e and \mathbf{X} represent the same parameters as those
344 defined in equation 1, u is the vector of individuals breeding value, \mathbf{Z} is the design matrix
345 corresponding to the breeding value. The covariance matrix of additive effects is represented by
346 $Var(u) = G\sigma_a^2$, where G is the matrix of relationships between individuals obtained from genomic
347 information, calculated according to the approach of VanRaden [26] (equation 3) and also
348 implemented through the R package “BGLR” (<https://github.com/gdlc/BGLR-R>) [62].

$$349 \quad \mathbf{G} = \frac{\mathbf{W}_a \mathbf{W}_a^T}{2 \sum_{f=1}^m p_f (1-p_f)} \quad (3)$$

350 where \mathbf{W}_a represented the matrix of additive genetic effect markers, with dimension of the number
351 of individuals (n) by the number of loci (m), and p_f is the minor allele frequency (MAF) value of
352 locus f .

353 **Accuracy of GP by K-fold cross-validation**

354 Five-fold cross-validation was performed to compare the accuracy of different methods of GP.
355 During K-fold cross validation, the population should be divided randomly [34]. The datasets
356 consisting of 821 individuals were divided into five approximately equally-sized subgroups (each
357 subgroup contained around 165 individuals). For 5-fold cross-validation, four subgroups which
358 retain the phenotype and genotype, were regarded as training population (reference population) to
359 estimate the parameters. The remaining subgroup i.e., candidate population was used to verify the
360 samples, and correspondingly, the phenotype of this group of samples was set as missing (Not
361 applicable, NA).

362 According to the above mentioned five models, the cross-validation was performed based on two
363 types of genotypic data (H- and L-datasets), with different densities and the BVs of the validation
364 group (candidate population) were predicted. In addition, the above cross-validation was performed
365 in triplicates in order to ensure the randomness of individuals in the validation group. Finally, the
366 GP accuracy values were calculated for each validation, averaged and then recorded as the final
367 accuracy.

368 **Abbreviations**

369 GP: genomic prediction; GS: genomic selection; SNP: Single nucleotide polymorphism; GBLUP:
370 genomic best linear unbiased prediction; BV: breeding value; Bayesian LASSO: Bayesian least
371 absolute shrinkage and selection operator; SSGBLUP: single-step GBLUP; S.E: standard error; S.D:
372 standard deviation; CFWR: clean fleece weight rate; SS: staple strength; FER: Fleece extension rate;
373 FD: mean fiber diameter; FD_CV: Coefficient of variation of FD; SL: staple length; h^2 : polygenic
374 heritability; QTL: quantitative trait loci; LD: linkage disequilibrium; MAF: minor allele frequency

375 **Declarations**

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380 **Authors' contribution**

381 BY and YY conceived and designed the experiments and explained the data. SZ analyzed the content
382 of the data with the help of TG, CY, JL, MH, HZ and Christian. YW, WS, XW, TW and JL provided
383 assistance with sample and data collection. SZ drafted the manuscript with the help of BY and YY.

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390 **Availability of data and materials**

391 All analysis results data generated during this study are included in this manuscript. Requests for
392 the raw data should be made to the corresponding authors.

393 **Ethics approval and consent to participate**

394 All animal work carried out in the current study was performed per the guidelines for the care and
395 use of laboratory animals promulgated by the State Council of the People's Republic of China. The
396 study was approved (License Number: 2019-008) by the Animal Management and Ethics
397 Committee of Lanzhou, Institute of Animal Husbandry and Veterinary Sciences, Chinese Academy
398 of Agricultural Sciences.

399 **Consent to publish**

400 Not applicable

401 **Competing interests**

402 The authors declare that they have no competing interests.

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566

567

568 **Figure Legends**

569 **Figure 1.** Comparison of GP accuracy based on different density genotype datasets. The six traits
570 were clean fleece weight rate (CFWR); staple strength (SS); fleece extension rate (FER); mean
571 fiber diameter (FD); Coefficient of variation of FD (FD_CV); staple length (SL).

572 **Figure 2.** Based on genotype datasets of different densities, the GP accuracy of 5 models in
573 different heritability level. On the left is the result for the H-datasets, and on the right is the result
574 for the L-datasets. The six traits were clean fleece weight rate (CFWR); staple strength (SS);
575 fleece extension rate (FER); mean fiber diameter (FD); Coefficient of variation of FD (FD_CV);
576 staple length (SL). The five models were: BayesA (BA); BayesB (BB); BayesC π (BC); Bayesion
577 LASSO (BL); and GBLUP (GB).

578 **Tables**

579 **Table 1.** Descriptive statistics of phenotypic values of traits. ¹ S.E, standard error; ² S.D, standard
580 deviation.

581 **Table 2.** Different GS methods and effects distribution.

582 **Table 3.** Estimates of additive and residual components of variance obtained adopting 'BGLR' for
583 different datasets. ^a CFWR: clean fleece weight rate; SS: staple strength; FER: fleece extension
584 rate; FD: mean fiber diameter; FD_CV: Coefficient of variation of FD; SL: staple length; ^b
585 Polygenic heritability, the proportion of the additive effect variance to the total phenotypic
586 variance.

587 **Table 4.** Comparison of prediction accuracies of 6 traits based on 2 datasets via 5 models. ^a
588 Abbreviations of traits explained in Table 3; ^b S.E are in parenthesis; ^c BA: BayesA; BB; BayesB;
589 BC: BayesC π ; BL: Bayesion LASSO; GB: genomic best linear unbiased prediction, GBLUP.

Figures

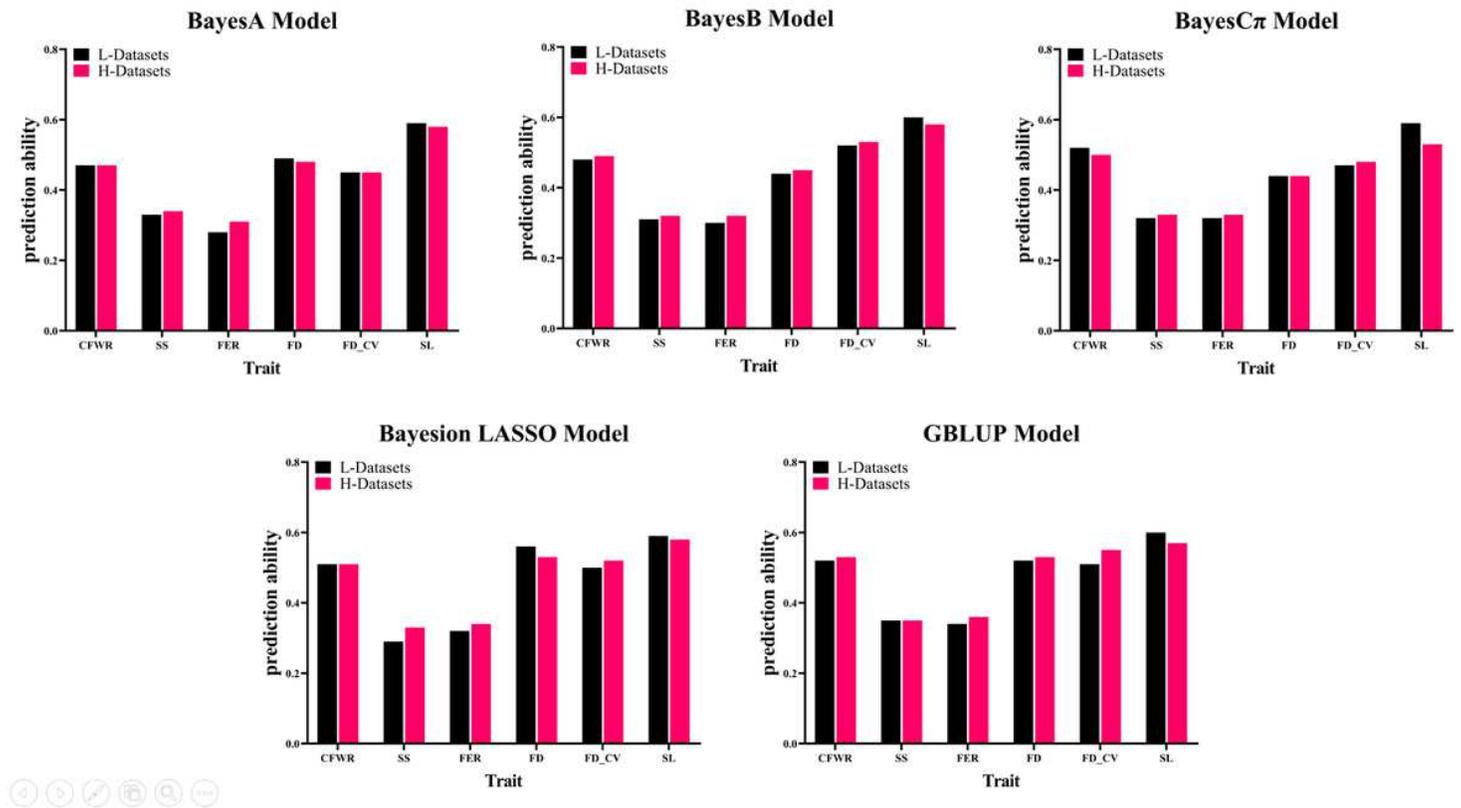


Figure 1

Comparison of GP accuracy based on different density genotype datasets. The six traits were clean fleece weight rate (CFWR); staple strength (SS); fleece extension rate (FER); mean fiber diameter (FD); Coefficient of variation of FD (FD_CV); staple length (SL).

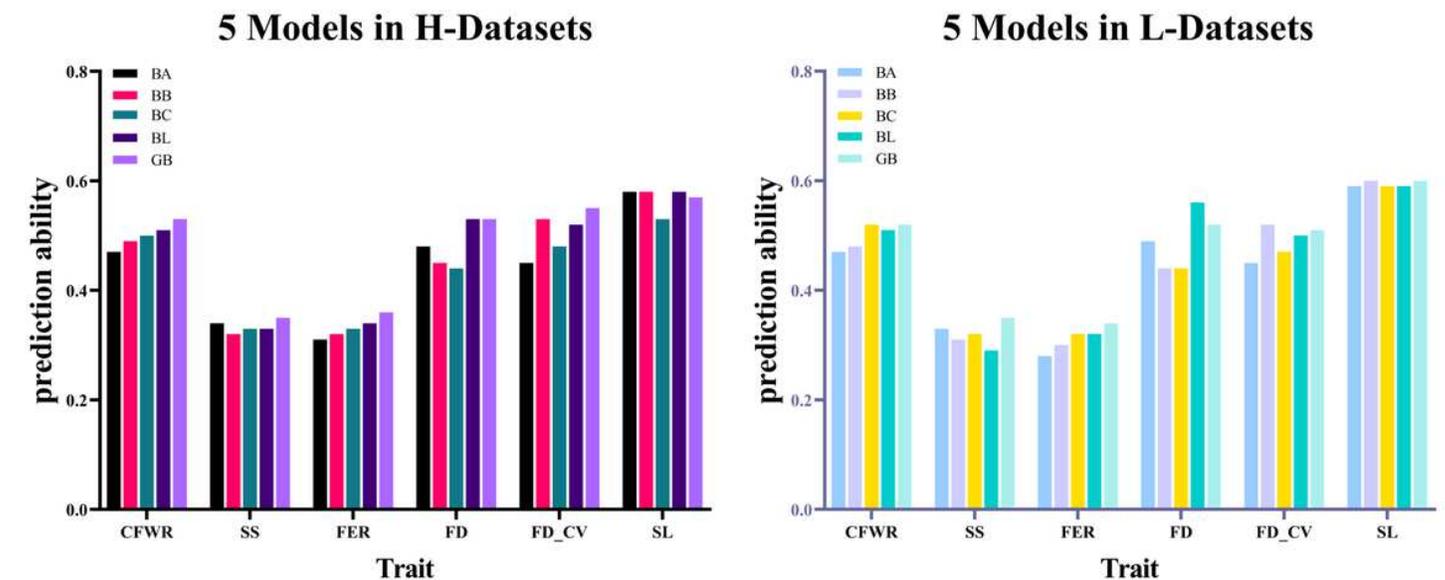


Figure 2

Based on genotype datasets of different densities, the GP accuracy of 5 models in different heritability level. On the left is the result for the H-datasets, and on the right is the result for the L-datasets. The six traits were clean fleece weight rate (CFWR); staple strength (SS); fleece extension rate (FER); mean fiber diameter (FD); Coefficient of variation of FD (FD_CV); staple length (SL). The five models were: BayesA (BA); BayesB (BB); BayesC π (BC); Bayesian LASSO (BL); and GBLUP (GB).

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