

Blood Levels of Brain-Derived Neurotrophic Factor (BDNF) in People with Multiple Sclerosis (MS): A Systematic Review and Meta-Analysis

Parnian Shobeiri

Tehran University of Medical Sciences (TUMS), Children's Medical Center Hospital

Helia Ashourizadeh

Shahid Beheshti University of Medical Sciences

Boshra Akbarzadeh Pasha

Tehran University of Medical Sciences

Maryam Haghshomar

Tehran University of Medical Sciences (TUMS), Children's Medical Center Hospital

Tahmineh Jouzdani

Tehran University of Medical Sciences

Antônio L. Teixeira

The University of Texas Health Science Center at Houston

Nima Rezaei (rezaei_nima@yahoo.com)

Universal Scientific Education and Research Network (USERN)

Research Article

Keywords: brain-derived neurotrophic factor, multiple sclerosis, neurotrophins, BDNF, serum

Posted Date: May 31st, 2022

DOI: https://doi.org/10.21203/rs.3.rs-1680103/v1

License:
(i) This work is licensed under a Creative Commons Attribution 4.0 International License. Read Full License

Abstract

Background: Multiple sclerosis is an autoimmune demyelinating disease marked by the involvement of multiple pathophysiological pathways, including BDNF. BDNF (brain-derived neurotrophic factor) is one of the main neurotrophic factors in the adult brain. The amount of BDNF in the blood can be utilized as a surrogate for the central expression of this marker. Given contradicting reports, we set out to answer the question, "How do blood levels of BDNF differ in people with multiple sclerosis (PwMS) compared to controls?"

Methods: We performed a thorough search in MEDLINE, EMBASE, Web of Science, and the Cochrane Library databases, resulting in 13 eligible investigations. Eleven studies compared BDNF in serum of PwMS versus healthy controls (HC), and two studies provided BDNF levels in the plasma of PwMs. R version 4.0.4 was used for meta-analysis and visualizations. Mean difference (MD) was used for the measurement of effect size.

Results: The final analysis included thirteen studies with 689 patients with MS and 583 controls. The preliminary results indicated that MS patients had statistically significant lower levels of BDNF than controls: SMD -5.1992 (95% CI [-8.4488; -1.9496], *p-value* < 0.0001. Additionally, subgroup analysis revealed a statistically significant difference in serum and plasma levels (*p-value*=0.01). Performing univariate meta-regression, disease duration and the proportion of males had, respectively, a significant negative and positive correlation with BDNF levels.

Conclusion: Circulating levels of BDNF are decreased in MS. Future studies should investigate the role of BDNF as a biomarker of disease severity and/or progression for a personalized approach to MS.

1. Background

Multiple sclerosis (MS) is a central nervous system (CNS) inflammatory and demyelinating disease that primarily affects people between the ages of 20 and 40. It is a chronic condition marked by inflammation, demyelination, and axonal loss, and it often results in considerable degrees of disability (1–3). Axonal damage is a crucial factor in determining the severity of long-term neurological (motor, sensory and cognitive) impairments in people with multiple sclerosis (PwMS). Cognitive impairment is well recognized as a key characteristic of MS, affecting up to 70% of patients and having a significant impact on daily activities (4). Information processing speed, attention, working and episodic memory, executive skills, and visuospatial abilities are the most typically disrupted cognitive domains in adult patients. Language and general intelligence are relatively stable unaffected (4, 5). Cognitive impairment in MS has been associated with a variety of risk variables (6), with the function of brain-derived neurotrophic factor (BDNF) polymorphisms garnering increasing attention among genetic factors. Remarkably, BDNF has been associated with not only cognitive, but also motor/sensory performance in MS.

BDNF is a member of the neurotrophin family, including nerve growth factor (NGF) and neurotrophins 3 and 4 (NT3 and NT4). BDNF is secreted by microglial and astroglial cells, in autocrine loops, and across long distances through neural circuits (7–11). In both healthy people and patients with neurological diseases, BDFN is crucial for the development and survival of neurons and oligodendroglia (12). Synaptic plasticity, synaptic growth, dendritic branching, and excitatory/inhibitory neurotransmitter profile modification are all

assisted by BDNF (13). Following a variety of clinical assaults, human immune cells are also capable of generating BDNF, a molecule that protects neurons and axons from damage (14).

According to a substantial body of neuropathological and experimental data, BDNF may have essential roles in MS-related neuroinflammation, neuroprotection and repair (15). However, the results regarding peripheral levels of BDNF are conflicting. The current systematic review and meta-analysis aim to integrate the existing evidence on the blood levels of BDNF in PwMS.

2. Material And Methods

Study protocol, including objectives of the review, PICO Question, inclusion and exclusion criteria – for participants, types of studies, intervention/ exposure and outcome, search strategy, data collection, and methods for data synthesis/analysis, was developed according to The Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA 2020) guideline (16).

2.1. Search Strategy

The preliminary qualifying criteria comprised original publications assessing BDNF concentrations in serum and plasma in PwMS patients, with no language, publication date, or publication status restrictions (e.g., online first or published). Accordingly, a search for relevant articles was conducted using the keywords BDNF, Brain-Derived Neurotrophic Factor, Multiple Sclerosis, and MS. We did not consider CSF, serum, or plasma in our search terms, postulating that studies measuring BDNF in other source(s) could guide unpublished data on BDNF concentrations in CSF or serum. The search was carried out in MEDLINE, EMBASE, Web of Science, and Cochrane Library, and it was updated on September 11th, 2021. We also reviewed reference listings and contacted the corresponding authors of all the articles included in the current systematic review and meta-analysis to find additional investigations.

2.2. Inclusion and Exclusion Criteria

Included studies all met the following criteria: (1) original articles studying human subjects, (2) enrolling individuals with MS, (3) diagnosis of MS established by a neurologist according to international criteria, (4) measuring serum, plasma, or CSF levels of BDNF in MS patients, (5) use of immunoassay or enzyme-linked immunosorbent assay (ELISA) to measure BDNF levels, and (7) reporting sufficient data including the total number of participants and mean and standard deviation (SD) of BDNF measures in both cases and controls.

Exclusion criteria were as follows: (1) review articles, books, book chapters, (2) studying on animal subjects, (3) pediatric MS, (4) studies evaluating tissue expression of BDNF, (5) in-vitro studies or studies on cell cultures, (6) and studies on genetic polymorphisms of BDNF but not levels of BDNF.

2.3. Data Collection and Data Items

Two investigators (H.A and B.A.P) reviewed eligible studies independently. Five studies were selected to produce pilot data extraction sheets and define descriptive factors of study groups to be included in the final data extraction sheets. Finally, the following information was retrieved from each included article: author, publication date, case and control definitions, male to female ratio, age, MS type (RRMS, PPMS, SPMS),

disease severity, and sample size in each group. Any discrepancies in data extraction were handled by discussion or referring the case to a third investigator (P.S).

2.4. Quality Assessment and Risk of Bias of Individual Studies and Across Studies

The quality of included studies was assessed using the standardized critical appraisal instruments from the Joanna Briggs Institute (JBI, http://joannabriggs.org/research/critical-appraisal-tools.html). This is a critical evaluation checklist to see whether a study's design, conduct, and analysis were influenced by bias. Reviewers rated each study's study design based on clarity of inclusion criteria; setting; exposure; standard criteria or adequate matching for individual groups; identification and strategies to overcome confounding factors; validity and reliability of outcome measurement; and adequacy statistical analyses for each. Total positive ratings for each investigation are calculated by scoring each question as Yes (+), Unclear/Not Mentioned (-) or No (-). Studies having a total of fewer than five favorable evaluations were omitted from the synthesis because of their low quality. Two reviewers (H.A and B.A.P) independently examined all papers for methodological validity, and any disagreements were addressed through discussion or with the assistance of a third reviewer (P.S).

2.5. Statistical Analysis

The standardized mean difference (SMD) was used for the measurement of effect. Fixed effects and random effects were interchangeably used as the analysis model. If the values reported in the manuscript were given as a median and interquartile range (IQR) or median and range, and we were not able to retrieve the mean \pm standard deviation (SD) from the authors, we used statistical methods suggested by Luo et al. (17) and Wan et al. (18) to convert these values. Heterogeneity was determined using Q statistic tests and the I² index. According to the Cochrane guidelines, the I² < 40% would mean that the inconsistency across studies is not important. In this case, we planned to use the fixed effects model. If the I² estimates fluctuated more than 40%, we intended to use the random effects procedure as the analysis model. To further assess the causes of heterogeneity, we conducted a sensitivity analysis to identify influential cases for meta-analyses with significant heterogeneity, including ten or more studies. Each time we omitted one study and recalculated the effect size (Leave-One-Out Analyses).

The degree of funnel plot asymmetry and Egger's test (19) detect publication bias. In fact, funnel plots are commonly employed to identify publication bias visually. On the other hand, the Egger's test is an objective metric that allows users to corroborate visual clues offered by funnel plots. When there was evidence of publication bias, we adjusted the effect sizes using the trim-and-fill method (20).

All computations and visualizations were carried out using R version 4.0.4 (R Core Team [2020]. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria). We used following packages: "meta" (version 4.17-0), "metafor" (version 2.4-0), "dmetar" (version 0.0–9), and "tidyverse" (version 1.3.0). All plots were designed using R. A p-value of < 0.05 was considered statistically significant.

3. Results

3.1. Study Selection

The study selection details are depicted in Fig. 1. The database search returned a total of 2,622 entries. Based on the title/abstract, a preliminary screening eliminated 1,875 papers. The complete text of the remaining 214 publications was retrieved, and each item was carefully reviewed and analyzed against the inclusion criteria using the abstract and, where required, the full text. Twenty-six papers were chosen for comprehensive analysis, and their respective authors were contacted and asked to provide study group characteristics and BDNF levels. Additionally, we contacted the corresponding authors of six studies examining BDNF levels, anticipating they may contain unpublished data on serum/plasma BDNF levels. We omitted 13 articles due to the reasons depicted in Fig. 1. Finally, a comprehensive evaluation of BDNF concentrations in MS included 13 studies comprising 689 patients with multiple sclerosis (MS) and 583 healthy controls (HC).

3.2. Study Characteristics

As summarized in Table 1, thirteen studies published from 2003 to 2021 provided original data on BDNF levels in MS patients and HC (21-33). Ten studies compared BDNF levels in the serum of multiple sclerosis (MS) patients (n = 515) to those in healthy controls (HC) (n = 437) (22, 23, 25-30, 32, 33), while three studies provided additional information on plasma BDNF levels in MS patients (n = 174) and HCs (n = 146) (21, 24, 31). The mean age varied from 31.2 ± 7.99 to 54.07 ± 8.46 years among MS patients and from 29.00 ± 7.80 to 50.71 ± 12.08 years among HC. The EDSS score of the included participants ranged from 1.26 ± 0.73 to 3.22 ± 1.59 , reported by all but two studies (23, 31). The type of MS people included in each study is depicted in Table 1. All but three studies (21, 31, 33), contained information on the disease duration ranging from 3 to 31 years. Moreover, all but one (21) of the included studies assessed the BDNF levels using enzyme-linked immunosorbent assay (ELISA) as the analytical procedure.

3.3. Quality Assessment

Table 2 summarizes the results of the quality assessment of the included studies based on the JBI critical appraisal tool.

3.4. Serum and Plasma BDNF Levels in Multiple Sclerosis

Meta-analysis of ten between-group comparisons revealed that serum BDNF levels are significantly (*p-value* < 0.0001) decreased in MS patients (n = 338) in relation to HC (n = 443) with a standardized mean difference (SMD) of -0.9680 pg/mL (95% CI [-1.6092; -0.3268], I² = 93.1%, Fig. 2A). Regarding the plasma BDNF levels, meta-analysis of three between-group comparisons showed that the pooled SMD was – 5.1992 (95% CI [-8.4488; -1.9496], *p-value* < 0.0001, I² = 98.7%, Fig. 2A) for BDNF levels assessed in plasma. Overall, subgroup comparisons revealed a substantial difference in BDNF concentrations across individuals' serum and plasma samples (*p-value* = 0.0123). Table 3 summarizes the results of the subgroup meta-analysis.

Table 3 Results of meta-analysis and meta-regression of BDNF levels in patients with Multiple Sclerosis.

Comparison	No. of comparisons	No. of subjects		Meta-ana	alysis		Heterogeneity			
		MS	HC	Effect size	95% CI	p- value	 ² %	Q	p- value	
Serum vs. Plasma	13	689	583	-1.5738	-2.2637; -0.8839	0.0123	95.9%	289.34	< 0.0001	
Moderator	No. of comparisons	No. o subje	-	Meta-reg	ression		R ² Analog (proportion of variance explained)			
		MS	HC	Slope	95% Cl	p- value				
Sample Size	13	689	583	0.0049	-0.0080; 0.0177	0.4591	0.00%			
Age (mean, years)	13	689	583	0.1092	-0.0210; 0.2394	0.1001	0.00%			
EDSS score	11	630	530	-0.8327	-1.8139; 0.1484	0.0962	0.00%			
Disease duration	9	508	434	-0.3895	-0.5619; -0.2170	< 0.0001	9.34%			
Sex (male, %)	13	689	583	0.0740	0.0009; 0.1471	0.0472	0.00%			

3.5. Publication Bias

The funnel plot was asymmetric (Fig. 3A). Moreover, Egger's tests revealed significant evidence of publication bias among the included studies (Egger's test, *p-value* = 0.04).

3.6. Trim and Fill analysis (metatrim)

The *p*-value for Egger's regression test was 0.04, confirming the existence of publication bias. We conducted a trim and fill study to examine the effect of publication bias, as we thought that the funnel plot asymmetry was only due to publication bias, which may not be relevant to this type of data. Utilizing the 'trimfill' function, one unpublished investigation was discovered according to the trim and fill analysis. Considering this study in calculating the SMD of BDNF levels yielded an estimated SMD of -0.9247 (95% CI [-1.7514; -0.0980], $I^2 = 97.1\%$, Fig. 3B), when adjusted for the publication bias.

3.7. Sensitivity Analysis and Outliers' Identification

Each study's effect on the overall estimate was assessed through systematic omission of studies and comparison of the pooled estimate from the remaining 12 studies. MS patients had lower peripheral BDNF levels than controls, implying that removing any single study would have little effect on the overall results (Fig. 4).

Using the 'find.outliers' command in R software, eight studies (21, 26–31, 33) were identified as outliers; accordingly, we repeated the meta-analysis of five remained studies and obtained the following results: SMD – 0.6619 (95% CI [-0.9391; -0.3848], *p-value* < 0.0001, Fig. 2B), which indicates statistically significant lower BDNF levels in MS participants compared to controls.

3.8. Meta-regression Analysis

We utilized meta-regression analysis to determine the determinants of study heterogeneity and the effect of modifiers. Univariable meta-regression models showed no relationship between the sample size, mean age, and EDSS score and BDNF levels. Interestingly, disease duration and the proportion of males had a significant negative and positive association to BDNF levels, respectively. Additionally, disease duration could explain 9.34% of the existing heterogeneity. (Table 3, Fig. 5)

4. Discussion

BDNF is involved in the pathogenesis of a broad spectrum of diseases, specifically neurologic and psychiatric conditions. BDNF and its high-affinity receptor (TrkB) are widely expressed in the CNS and peripheral tissues, including liver, muscle cells (smooth and skeletal) (34), white blood cells (lymphocytes and monocytes) (35), endothelial cells (36), reproductive tract (37) and adipose tissue (38). BDNF is a key factor in neuronal growth, development, differentiation, synapse formation, and synaptic plasticity, ultimately in 'optimal' brain health (39-41). There is also evidence of the role of BDNF in bioenergy homeostasis, thermoregulation, and energy expenditure (42, 43).

Numerous studies have investigated peripheral levels of BDNF in neurologic diseases. In a systematic review and meta-analysis of studies involving patients with epilepsy, BDNF level was not significantly different from healthy controls. However, partial epilepsy patients had lower levels of BDNF than other types of epilepsy in the subgroup analysis (44). A recent systematic review and meta-analysis showed a significant decrease in plasma BDNF levels in PD patients compared to healthy controls (45). Nevertheless, increased plasma levels of BDNF in PD have also been reported and hypothesized to represent a CNS protective response to the neuronal loss resulting from PD pathogenesis (46, 47). In stroke, serum BDNF levels correlated with stroke severity in the acute phase (48), and later with rehabilitation treatment (49, 50). Altogether, these studies suggest the involvement of BDNF in the pathophysiology of CNS pathological conditions and the potential role of this molecule as a biomarker of disease severity and/or staging.

To investigate peripheral/blood levels of BDNF in PwMS, we did a systematic review of literature and metaanalysis of clinical studies investigating BDNF in PwMS compared with healthy controls. Our results demonstrated a statistically significant difference in serum and/or plasma BDNF concentration in PwMS and healthy non-MS individuals.

MS has complex pathophysiology involving inflammation-mediated demyelination and axonal loss. An immune-mediated inflammation against myelin sheath seems to be the main pathologic event leading to MS spectrum diseases (51). A growing body of evidence also implicates a role for BDNF in MS pathogenesis (52).

There is increased BDNF expression in active inflammatory MS lesions (53, 54), and increased peripheral levels of BDNF during the acute inflammatory MS relapses (24, 55, 56). Moreover, BDNF production by peripheral blood mononuclear cells was significantly increased during relapse and in the recovery phase compared with values detected in the stable phase of the disease (57). These findings suggest a neuroprotective role for BDNF that would promote axonal/neuronal regeneration, especially during MS relapses. The results are conflicting in the remission phase, i.e., outside the MS relapses, which we specifically investigated in our study. In some studies, the BDNF blood level was lower in patients than in healthy controls (22, 33, 58–63), whereas in other studies, no significant difference was observed (57, 64, 65). Studies investigating the correlation between BDNF and disease severity, as assessed by EDSS, also have conflicting results. (24, 56, 58)

Our results demonstrated a significant decreased serum and/or plasma BDNF concentration in PwMS compared with healthy non-MS individuals. This finding might reflect neuronal damage with subsequent decrease in BDNF production and/or BDNF consumption through the neuro-regenerative process (60, 63).

BDNF is produced by several neuronal cells in the CNS, including glial cells such as astrocytes and microglia (66) and non-neuronal non-glial cells in the peripheral tissues (36, 37, 67, 68). BDNF can also be transported through the blood-brain barrier in a bidirectional manner between the peripheral circulation and CNS (69). In the blood, around 90% of circulating BDNF is stored in the platelets (70), while in the CNS, BDNF is highly expressed by the hippocampus, amygdala, and cortex neurons (71, 72). As long as BDNF produced in the periphery can be transported through the blood-brain barrier, increased peripheral levels of BDNF could result in an increased entry of this neurotrophin into the CNS, contributing to remyelination and axonal regeneration (69).

Several factors can influence BDNF levels. Physical exercise, for instance, has been investigated as a relevant factor stimulating BDNF synthesis which is implicated in exercise-induced neurogenesis in MS and other neurologic disorders (73–76). Increased BDNF synthesis results in enhanced cognitive function, motor symptoms, and alleviation of psychiatric symptoms (77–79). Physical exercise in MS patients reduces episodes of relapses and disease progression, reduces fatigue, and improves the quality of life(80–83). Based on systematic literature reviews and meta-analyses, regardless of type and duration, physical exercise can increase BDNF circulating levels in MS and PD but not significantly in mild cognitive impairment (84, 85). Exercise-induced increase in BDNF levels was observed in 4 out of 8 studies within PwMS (86). Medication is another factor that can influence BDNF. Glatiramer acetate (GA) is a widely used medication among MS patients. One of the proposed mechanisms of action of GA is the downregulation of proinflammatory cytokines associated with increased production of BDNF (87). GA-treated MS patients showed a progressive increase in BDNF levels from baseline to month three (57). Other drugs which alter BDNF levels include antidepressants (88), Selegiline (89), Metformin (90), Risperidone, and Clozapine (91, 92).

High heterogeneity between studies is a valid concern in interpreting the meta-analyses results. Due to the high heterogeneity of included studies, we performed a meta-regression analysis to further investigate the source of methodologically and/or clinically diversity. Our meta-regression showed that the study heterogeneity could be explained by disease duration. Another explanation for high heterogeneity among included studies is the measurement bias. A diverse range of technical issues (e.g., sampling, ELISA kits) has

complicated BDNF measurement in human samples, resulting in poor reliability of available tests. This detection bias was well studied in a study investigating six commercially available ELISA kits for BDNF level measurement, in which only two of six assays showed reliable results (93). In addition, the limited number of studies with small sample sizes could further increase the heterogeneity of the analysis. Lack of enough data regarding the MS type of included patients and/or diverse MS types included in the studies could result in clinical diversity and bias. Although none of the patients in the included studies was at relapse or acute inflammatory state, there is still the possibility of bias in misclassifying the patients as remission or stable phase. This could falsely increase the BDNF level in the patient group. Disease duration and severity were among the data not comprehensively addressed in some studies, which could further increase the risk of bias. Other comorbid conditions, such as depression, can affect the BDNF level(94, 95). As discussed earlier, numerous drugs alter the BDNF level and expression in both MS patients and the control individuals. In most studies, the use and dosage of these medications were not properly controlled. Information about physical exercise is also missing.

5. Conclusion

Results of this systematic review of literature and meta-analysis showed that circulating BDNF level in MS patients is statistically decreased compared to healthy non-MS individuals. Future studies should investigate the role of BDNF as a biomarker of disease severity and/or progression for a personalized approach in MS.

Declarations

Ethics approval and consent to participate: Not applicable

Consent for publication: Not applicable

Availability of data and materials: Not applicable

Competing interests: The authors declare that they have no competing interests.

Funding: Not applicable

Authors' contributions: PS: Writing - original draft/ Conceptualization/ Formal analysis/ Visualization, **HA:** Writing - original draft/ Data curation, **BAP, MH, TJ**: Data curation, **ALT:** Writing - review & editing, **NR:** Supervision/ Writing - review & editing. All authors read and approved the final manuscript.

Acknowledgments: Not applicable

References

- 1. Karussis D. The diagnosis of multiple sclerosis and the various related demyelinating syndromes: a critical review. Journal of autoimmunity. 2014;48:134–42.
- 2. Gold R, Montalban X. Multiple sclerosis: more pieces of the immunological puzzle. The Lancet Neurology. 2012;11(1):9–10.

- 3. Naegele M, Martin R. The good and the bad of neuroinflammation in multiple sclerosis. Handbook of clinical neurology. 2014;122:59–87.
- 4. Sumowski JF, Benedict R, Enzinger C, Filippi M, Geurts JJ, Hamalainen P, et al. Cognition in multiple sclerosis: State of the field and priorities for the future. Neurology. 2018;90(6):278–88.
- 5. Benedict RH, Amato MP, DeLuca J, Geurts JJ. Cognitive impairment in multiple sclerosis: clinical management, MRI, and therapeutic avenues. The Lancet Neurology. 2020;19(10):860–71.
- 6. Amato MP, Prestipino E, Bellinvia A. Identifying risk factors for cognitive issues in multiple sclerosis. Expert review of neurotherapeutics. 2019;19(4):333–47.
- 7. Leibrock J, Lottspeich F, Hohn A, Hofer M, Hengerer B, Masiakowski P, et al. Molecular cloning and expression of brain-derived neurotrophic factor. Nature. 1989;341(6238):149–52.
- 8. Hartmann M, Heumann R, Lessmann V. Synaptic secretion of BDNF after high-frequency stimulation of glutamatergic synapses. The EMBO journal. 2001;20(21):5887–97.
- 9. Murer M, Yan Q, Raisman-Vozari R. Brain-derived neurotrophic factor in the control human brain, and in Alzheimer's disease and Parkinson's disease. Progress in neurobiology. 2001;63(1):71–124.
- Kerschensteiner M, Gallmeier E, Behrens L, Leal VV, Misgeld T, Klinkert WE, et al. Activated human T cells, B cells, and monocytes produce brain-derived neurotrophic factor in vitro and in inflammatory brain lesions: a neuroprotective role of inflammation? The Journal of experimental medicine. 1999;189(5):865–70.
- 11. Lindvall O, Kokaia Z, Bengzon J, Elme E, Kokaia M. Neurotrophins and brain insults. Trends in neurosciences. 1994;17(11):490–6.
- 12. Hu Y, Russek SJ. BDNF and the diseased nervous system: a delicate balance between adaptive and pathological processes of gene regulation. Journal of neurochemistry. 2008;105(1):1–17.
- 13. Giacobbo BL, Doorduin J, Klein HC, Dierckx RA, Bromberg E, de Vries EF. Brain-derived neurotrophic factor in brain disorders: focus on neuroinflammation. Molecular neurobiology. 2019;56(5):3295–312.
- Stadelmann C, Kerschensteiner M, Misgeld T, BruÈck W, Hohlfeld R, Lassmann H. BDNF and gp145trkB in multiple sclerosis brain lesions: neuroprotective interactions between immune and neuronal cells? Brain. 2002;125(1):75–85.
- 15. Nociti V. What is the role of Brain derived neurotrophic factor in Multiple Sclerosis neuroinflammation? Neuroimmunology and Neuroinflammation. 2020;7(3):291–9.
- 16. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. BMJ. 2021;372:n71.
- 17. Luo D, Wan X, Liu J, Tong T. Optimally estimating the sample mean from the sample size, median, mid-range, and/or mid-quartile range. Statistical methods in medical research. 2018;27(6):1785–805.
- 18. Wan X, Wang W, Liu J, Tong T. Estimating the sample mean and standard deviation from the sample size, median, range and/or interquartile range. BMC medical research methodology. 2014;14(1):1–13.
- 19. Egger M, Smith GD, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. Bmj. 1997;315(7109):629–34.
- 20. Duval S, Tweedie R. Trim and fill: a simple funnel-plot–based method of testing and adjusting for publication bias in meta-analysis. Biometrics. 2000;56(2):455–63.

- 21. Al Temaimi R, AbuBaker J, Al-Khairi I, Alroughani R. Remyelination modulators in multiple sclerosis patients. Experimental and Molecular Pathology. 2017;103(3):237–41.
- 22. Comini-Frota ER, Rodrigues DH, Miranda EC, Brum DG, Kaimen-Maciel DR, Donadi EA, et al. Serum levels of brain-derived neurotrophic factor correlate with the number of T2 MRI lesions in multiple sclerosis. Braz J Med Biol Res. 2012;45(1):68–71.
- 23. Devasahayam AJ, Kelly LP, Williams JB, Moore CS, Ploughman M. Fitness Shifts the Balance of BDNF and IL-6 from Inflammation to Repair among People with Progressive Multiple Sclerosis. Biomolecules. 2021;11(4).
- 24. Frota ER, Rodrigues DH, Donadi EA, Brum DG, Maciel DR, Teixeira AL. Increased plasma levels of brain derived neurotrophic factor (BDNF) after multiple sclerosis relapse. Neurosci Lett. 2009;460(2):130–2.
- 25. Gold SM, Schulz KH, Hartmann S, Mladek M, Lang UE, Hellweg R, et al. Basal serum levels and reactivity of nerve growth factor and brain-derived neurotrophic factor to standardized acute exercise in multiple sclerosis and controls. J Neuroimmunol. 2003;138(1–2):99–105.
- 26. Lalive PH, Kantengwa S, Benkhoucha M, Juillard C, Chofflon M. Interferon-β induces brain-derived neurotrophic factor in peripheral blood mononuclear cells of multiple sclerosis patients. Journal of Neuroimmunology. 2008;197(2):147–51.
- 27. Naegelin Y, Saeuberli K, Schaedelin S, Dingsdale H, Magon S, Baranzini S, et al. Levels of brain-derived neurotrophic factor in patients with multiple sclerosis. Ann Clin Transl Neurol. 2020;7(11):2251–61.
- 28. Oraby MI, El Masry HA, Abd El Shafy SS, Abdul Galil EM. Serum level of brain-derived neurotrophic factor in patients with relapsing–remitting multiple sclerosis: a potential biomarker for disease activity. Egyptian Journal of Neurology, Psychiatry and Neurosurgery. 2021;57(1).
- 29. Ozkul C, Guclu-Gunduz A, Irkec C, Fidan I, Aydin Y, Ozkan T, et al. Effect of combined exercise training on serum brain-derived neurotrophic factor, suppressors of cytokine signaling 1 and 3 in patients with multiple sclerosis. J Neuroimmunol. 2018;316:121–9.
- 30. Philippova ES, Bazhenov IV, Ziryanov AV, Bazarny VV. Impact of intradetrusor botulinum toxin A injections on serum and urinary concentrations of nerve growth factor and brain-derived neurotrophic factor in patients with multiple sclerosis and neurogenic detrusor overactivity. Neurourol Urodyn. 2021;40(1):95– 101.
- 31. Shajarian M, Alsahebfosoul F, Etemadifar M. The Effect of IFN-β Treatment on Plasma Levels of BDNF and IL-6 in Relapsing-Remitting Multiple Sclerosis Patients. Neuroimmunomodulation. 2021;28(3):150–7.
- 32. Tongiorgi E, Sartori A, Baj G, Bratina A, Di Cola F, Zorzon M, et al. Altered serum content of brain-derived neurotrophic factor isoforms in multiple sclerosis. J Neurol Sci. 2012;320(1–2):161–5.
- 33. Wens I, Keytsman C, Deckx N, Cools N, Dalgas U, Eijnde BO. Brain derived neurotrophic factor in multiple sclerosis: effect of 24 weeks endurance and resistance training. Eur J Neurol. 2016;23(6):1028–35.
- 34. Yarrow JF, White LJ, McCoy SC, Borst SE. Training augments resistance exercise induced elevation of circulating brain derived neurotrophic factor (BDNF). Neurosci Lett. 2010;479(2):161–5.
- 35. Kerschensteiner M, Gallmeier E, Behrens L, Leal VV, Misgeld T, Klinkert WE, et al. Activated human T cells, B cells, and monocytes produce brain-derived neurotrophic factor in vitro and in inflammatory brain lesions: a neuroprotective role of inflammation? J Exp Med. 1999;189(5):865–70.

- 36. Nakahashi T, Fujimura H, Altar CA, Li J, Kambayashi J-i, Tandon NN, et al. Vascular endothelial cells synthesize and secrete brain-derived neurotrophic factor. FEBS Letters. 2000;470(2):113–7.
- 37. Chow R, Wessels JM, Foster WG. Brain-derived neurotrophic factor (BDNF) expression and function in the mammalian reproductive Tract. Hum Reprod Update. 2020;26(4):545–64.
- 38. Zierold S, Buschmann K, Gachkar S, Bochenek ML, Velmeden D, Hobohm L, et al. Brain-Derived Neurotrophic Factor Expression and Signaling in Different Perivascular Adipose Tissue Depots of Patients With Coronary Artery Disease. J Am Heart Assoc. 2021;10(6):e018322.
- Bath KG, Lee FS. Neurotrophic factor control of adult SVZ neurogenesis. Dev Neurobiol. 2010;70(5):339–49.
- 40. Lee J, Duan W, Mattson MP. Evidence that brain-derived neurotrophic factor is required for basal neurogenesis and mediates, in part, the enhancement of neurogenesis by dietary restriction in the hippocampus of adult mice. J Neurochem. 2002;82(6):1367–75.
- 41. Tapia-Arancibia L, Aliaga E, Silhol M, Arancibia S. New insights into brain BDNF function in normal aging and Alzheimer disease. Brain Res Rev. 2008;59(1):201–20.
- 42. Krisztina Marosi MPM. BDNF Mediates Adaptive Brain and Body Responses to Energetic Challenges. Trends Endocrinol Metab. 2015;25(2):10.
- 43. Maria Carmela Di Rosa SZ, Miriam Wissam Saab, Marianna Flora Tomasello. The Pleiotropic Potential of BDNF beyond Neurons: Implication for a Healthy Mind in a Healthy Body. Life (Basel). 2021;11(11).
- 44. Nowroozi A, Salehi MA, Mohammadi S. Brain-derived neurotrophic factor in patients with epilepsy: A systematic review and meta-analysis. Epilepsy Res. 2021;178:106794.
- 45. Rahmani F, Saghazadeh A, Rahmani M, Teixeira AL, Rezaei N, Aghamollaii V, et al. Plasma levels of brainderived neurotrophic factor in patients with Parkinson disease: A systematic review and meta-analysis. Brain Res. 2019;1704:127–36.
- 46. Lima Giacobbo B, Doorduin J, Klein HC, Dierckx R, Bromberg E, de Vries EFJ. Brain-Derived Neurotrophic Factor in Brain Disorders: Focus on Neuroinflammation. Mol Neurobiol. 2019;56(5):3295–312.
- 47. Scalzo P, Kummer A, Bretas TL, Cardoso F, Teixeira AL. Serum levels of brain-derived neurotrophic factor correlate with motor impairment in Parkinson's disease. J Neurol. 2010;257(4):540–5.
- 48. Eleni Karantali DK, Vasileios Papavasileiou, Angeliki Prevezianou, Symela Chatzikonstantinou, Foivos Petridis, Jack McKenna, Alina-Costina Luca, Constantin Trus, Alin Ciobica, Ioannis Mavroudis. Serum BDNF Levels in Acute Stroke: A Systematic Review and Meta-Analysis. Medicina (Kaunas). 2021;57(3).
- 49. Koroleva ES, Tolmachev IV, Alifirova VM, Boiko AS, Levchuk LA, Loonen AJM, et al. Serum BDNF's Role as a Biomarker for Motor Training in the Context of AR-Based Rehabilitation after Ischemic Stroke. Brain Sci. 2020;10(9).
- 50. Mourão AM, Vicente LCC, Abreu MNS, Sant'Anna RV, Vieira ELM, de Souza LC, et al. Plasma levels of brain-derived neurotrophic factor are associated with prognosis in the acute phase of ischemic stroke. Journal of Stroke and Cerebrovascular Diseases. 2019;28(3):735–40.
- 51. Hemmer B, Archelos JJ, Hartung HP. New concepts in the immunopathogenesis of multiple sclerosis. Nat Rev Neurosci. 2002;3(4):291–301.

- 52. Hohlfeld R, Kerschensteiner M, Stadelmann C, Lassmann H, Wekerle H. The neuroprotective effect of inflammation: implications for the therapy of multiple sclerosis. Journal of Neuroimmunology. 2000;107(2):161–6.
- 53. Stadelmann C, Kerschensteiner M, Misgeld T, Brück W, Hohlfeld R, Lassmann H. BDNF and gp145trkB in multiple sclerosis brain lesions: neuroprotective interactions between immune and neuronal cells? Brain. 2002;125(Pt 1):75–85.
- 54. Azoulay D, Urshansky N, Karni A. Low and dysregulated BDNF secretion from immune cells of MS patients is related to reduced neuroprotection. J Neuroimmunol. 2008;195(1–2):186–93.
- 55. Weinstock-Guttman B, Zivadinov R, Tamaño-Blanco M, Abdelrahman N, Badgett D, Durfee J, et al. Immune cell BDNF secretion is associated with white matter volume in multiple sclerosis. J Neuroimmunol. 2007;188(1–2):167–74.
- 56. Sarchielli P, Greco L, Stipa A, Floridi A, Gallai V. Brain-derived neurotrophic factor in patients with multiple sclerosis. Journal of Neuroimmunology. 2002;132(1–2):180–8.
- 57. Sarchielli P, Zaffaroni M, Floridi A, Greco L, Candeliere A, Mattioni A, et al. Production of brain-derived neurotrophic factor by mononuclear cells of patients with multiple sclerosis treated with glatiramer acetate, interferon-beta 1a, and high doses of immunoglobulins. Mult Scler. 2007;13(3):313–31.
- 58. Yvonne Naegelin KS, Sabine Schaedelin, Hayley Dingsdale, Stefano Magon, Sergio Baranzini, Michael Amann, Katrin Parmar, Charidimos Tsagkas, Pasquale Calabrese, Iris Katharina Penner, Ludwig Kappos, Yves-Alain Barde. Levels of brain-derived neurotrophic factor in patients with multiple sclerosis. Annals of Clinical and Translational Neurology 2020;7(11):11.
- 59. Islas-Hernandez A, Aguilar-Talamantes HS, Bertado-Cortes B, Mejia-delCastillo GJ, Carrera-Pineda R, Cuevas-Garcia CF, et al. BDNF and Tau as biomarkers of severity in multiple sclerosis. Biomark Med. 2018;12(7):717–26.
- 60. Castellano V, White LJ. Serum brain-derived neurotrophic factor response to aerobic exercise in multiple sclerosis. J Neurol Sci. 2008;269(1–2):85–91.
- 61. Al-Temaimi R, AbuBaker J, Al-Khairi I, Alroughani R. Remyelination modulators in multiple sclerosis patients. Exp Mol Pathol. 2017;103(3):237–41.
- 62. Kalinowska-Łyszczarz A, Pawlak MA, Wyciszkiewicz A, Osztynowicz K, Kozubski W, Michalak S. Immunecell BDNF expression in treatment-naïve relapsing-remitting multiple sclerosis patients and following one year of immunomodulation therapy. Neurol Neurochir Pol. 2018;52(4):483–9.
- 63. Azoulay D, Vachapova V, Shihman B, Miler A, Karni A. Lower brain-derived neurotrophic factor in serum of relapsing remitting MS: reversal by glatiramer acetate. J Neuroimmunol. 2005;167(1–2):215–8.
- 64. Damasceno A, Damasceno BP, Cendes F, Damasceno A, Moraes AS, Farias A, et al. Serum BDNF levels are not reliable correlates of neurodegeneration in MS patients. Mult Scler Relat Disord. 2015;4(1):65–6.
- 65. Oraby MI, El Masry HA, Abd El Shafy SS, Abdul Galil EM. Serum level of brain-derived neurotrophic factor in patients with relapsing-remitting multiple sclerosis: a potential biomarker for disease activity. The Egyptian Journal of Neurology, Psychiatry and Neurosurgery. 2021;57(1).
- 66. Quesseveur G, David DJ, Gaillard MC, Pla P, Wu MV, Nguyen HT, et al. BDNF overexpression in mouse hippocampal astrocytes promotes local neurogenesis and elicits anxiolytic-like activities. Transl

Psychiatry. 2013;3:e253.

- 67. Katanuma Y, Numakawa T, Adachi N, Yamamoto N, Ooshima Y, Odaka H, et al. Phencyclidine rapidly decreases neuronal mRNA of brain-derived neurotrophic factor. Synapse. 2014;68(6):257–65.
- 68. Benros ME, Nielsen PR, Nordentoft M, Eaton WW, Dalton SO, Mortensen PB. Autoimmune diseases and severe infections as risk factors for schizophrenia: a 30-year population-based register study. Am J Psychiatry. 2011;168(12):1303–10.
- 69. Pan W, Banks WA, Fasold MB, Bluth J, Kastin AJ. Transport of brain-derived neurotrophic factor across the blood-brain barrier. Neuropharmacology. 1998;37(12):1553–61.
- 70. Hironobu Fujimura CAA, Ruoyan Chen, Takashi Nakamura, Takeshi Nakahashi, Jun-ichi Kambayashi, Bing Sun, Narendra N Tandon. Brain-derived neurotrophic factor is stored in human platelets and released by agonist stimulation. Thrombosis and Haemostasis. 2002;87(4):7.
- 71. Hofer M, Pagliusi SR, Hohn A, Leibrock J, Barde YA. Regional distribution of brain-derived neurotrophic factor mRNA in the adult mouse brain. The EMBO Journal. 1990;9(8):2459–64.
- 72. Timmusk T, Palm K, Metsis M, Reintam T, Paalme V, Saarma M, et al. Multiple promoters direct tissuespecific expression of the rat BDNF gene. Neuron. 1993;10(3):475–89.
- 73. Cotman C. Exercise: a behavioral intervention to enhance brain health and plasticity. Trends in Neurosciences. 2002;25(6):295–301.
- 74. Dalgas U, Langeskov-Christensen M, Stenager E, Riemenschneider M, Hvid LG. Exercise as Medicine in Multiple Sclerosis-Time for a Paradigm Shift: Preventive, Symptomatic, and Disease-Modifying Aspects and Perspectives. Curr Neurol Neurosci Rep. 2019;19(11):88.
- Velikonja O, Curic K, Ozura A, Jazbec SS. Influence of sports climbing and yoga on spasticity, cognitive function, mood and fatigue in patients with multiple sclerosis. Clin Neurol Neurosurg. 2010;112(7):597– 601.
- 76. Kalron A, Zeilig G. Efficacy of exercise intervention programs on cognition in people suffering from multiple sclerosis, stroke and Parkinson's disease: A systematic review and meta-analysis of current evidence. NeuroRehabilitation. 2015;37(2):273–89.
- 77. Szuhany KL, Bugatti M, Otto MW. A meta-analytic review of the effects of exercise on brain-derived neurotrophic factor. J Psychiatr Res. 2015;60:56–64.
- 78. Denham J, Marques FZ, O'Brien BJ, Charchar FJ. Exercise: putting action into our epigenome. Sports Med. 2014;44(2):189–209.
- 79. Devasahayam AJ, Downer MB, Ploughman M. The Effects of Aerobic Exercise on the Recovery of Walking Ability and Neuroplasticity in People with Multiple Sclerosis: A Systematic Review of Animal and Clinical Studies. Mult Scler Int. 2017;2017:4815958.
- 80. Pilutti LA, Platta ME, Motl RW, Latimer-Cheung AE. The safety of exercise training in multiple sclerosis: a systematic review. J Neurol Sci. 2014;343(1–2):3–7.
- 81. Wesnes K, Myhr KM, Riise T, Cortese M, Pugliatti M, Bostrom I, et al. Physical activity is associated with a decreased multiple sclerosis risk: The EnvIMS study. Mult Scler. 2018;24(2):150–7.
- 82. Motl RW, Gosney JL. Effect of exercise training on quality of life in multiple sclerosis: a meta-analysis. Mult Scler. 2008;14(1):129–35.

- 83. Nazanin Razazian MK, Hossein Moayedi, Alireza Daneshkhah, Shamarina Shohaimi, Masoud Mohammadi, Rostam Jalali,corresponding author, Nader Salari. The impact of physical exercise on the fatigue symptoms in patients with multiple sclerosis: a systematic review and meta-analysis. BMC Neurology 2020;20.
- 84. Ruiz-Gonzalez D, Hernandez-Martinez A, Valenzuela PL, Morales JS, Soriano-Maldonado A. Effects of physical exercise on plasma brain-derived neurotrophic factor in neurodegenerative disorders: A systematic review and meta-analysis of randomized controlled trials. Neurosci Biobehav Rev. 2021;128:394–405.
- 85. Shobeiri P, Karimi A, Momtazmanesh S, Teixeira AL, Teunissen CE, van Wegen EE, et al. Exercise-induced increase in blood-based brain-derived neurotrophic factor (BDNF) in people with multiple sclerosis: A systematic review and meta-analysis of exercise intervention trials. PloS one. 2022;17(3):e0264557.
- 86. Negaresh R, Motl RW, Zimmer P, Mokhtarzade M, Baker JS. Effects of exercise training on multiple sclerosis biomarkers of central nervous system and disease status: a systematic review of intervention studies. Eur J Neurol. 2019;26(5):711–21.
- 87. Blanco Y, Moral EA, Costa M, Gómez-Choco M, Torres-Peraza JF, Alonso-Magdalena L, et al. Effect of glatiramer acetate (Copaxone) on the immunophenotypic and cytokine profile and BDNF production in multiple sclerosis: a longitudinal study. Neurosci Lett. 2006;406(3):270–5.
- 88. Chanjuan Zhou JZ, Bin Zou, Liang Fang, Jianjun Chen, Xiao Deng, Lin Zhang, Xiang Zhao, Zehui Qu, Yang Lei, and Ting Lei. Meta-analyses of comparative efficacy of antidepressant medications on peripheral BDNF concentration in patients with depression. PLOS One. 2017;12(2).
- Feizipour S, Sobhani S, Mehrafza S, Gholami M, Motaghinejad M, Motevalian M, et al. Selegiline acts as neuroprotective agent against methamphetamine-prompted mood and cognitive related behavior and neurotoxicity in rats: Involvement of CREB/BDNF and Akt/GSK3 signal pathways. Iran J Basic Med Sci. 2020;23(5):606–15.
- 90. Keshavarzi S, Kermanshahi S, Karami L, Motaghinejad M, Motevalian M, Sadr S. Protective role of metformin against methamphetamine induced anxiety, depression, cognition impairment and neurodegeneration in rat: The role of CREB/BDNF and Akt/GSK3 signaling pathways. Neurotoxicology. 2019;72:74–84.
- 91. Chen CC, Huang TL. Effects of antipsychotics on the serum BDNF levels in schizophrenia. Psychiatry Res. 2011;189(3):327–30.
- 92. Xiu MH, Hui L, Dang YF, Hou TD, Zhang CX, Zheng YL, et al. Decreased serum BDNF levels in chronic institutionalized schizophrenia on long-term treatment with typical and atypical antipsychotics. Prog Neuropsychopharmacol Biol Psychiatry. 2009;33(8):1508–12.
- 93. Polacchini A, Metelli G, Francavilla R, Baj G, Florean M, Mascaretti LG, et al. A method for reproducible measurements of serum BDNF: comparison of the performance of six commercial assays. Sci Rep. 2015;5:17989.
- 94. Bocchio-Chiavetto L, Bagnardi V, Zanardini R, Molteni R, Nielsen MG, Placentino A, et al. Serum and plasma BDNF levels in major depression: a replication study and meta-analyses. World J Biol Psychiatry. 2010;11(6):763–73.

95. Kishi T, Yoshimura R, Ikuta T, Iwata N. Brain-Derived Neurotrophic Factor and Major Depressive Disorder: Evidence from Meta-Analyses. Front Psychiatry. 2017;8:308.

Tables

Tables 1 and 2 are available in the Supplementary Files section.

Figures

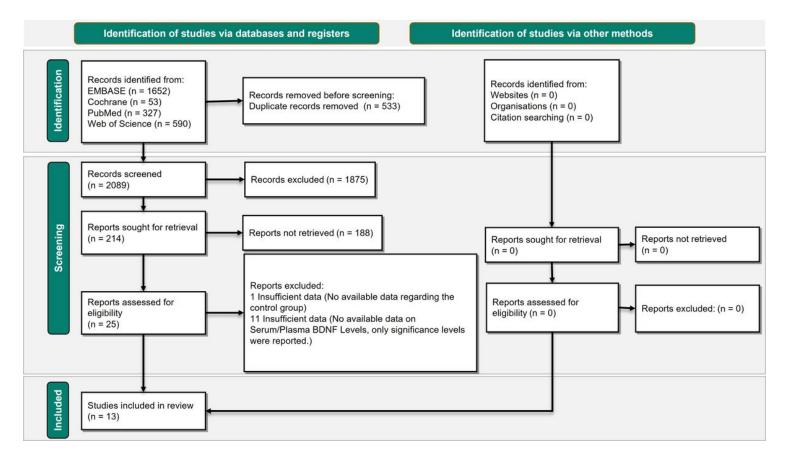


Figure 1

Flow diagram summarizing the selection of eligible studies based on the PRISMA guidelines.

Experiment						Control	Standardised Mean			
Author	N	Mean	SD	N	Mean	SD		SMD	95%-CI	Weight
Specimen = Serum										
Golda et al., 2003	25	4435.10	533.40	20	4712.20	491.80		-0.53	[-1.13; 0.07]	8.1%
Lalive et al., 2008	15	20400.00	6400.00	15	19500.00	7200.00		0.13	[-0.59; 0.85]	7.9%
Tongiorgi et al., 2012	20	24.45	5.89	20	29.50	6.47		-0.80	[-1.45; -0.15]	8.0%
Comini-Frota et al., 2012	28	1243.17	568.51	28	1840.39	903.60	in the second seco	-0.78	[-1.32; -0.24]	8.2%
Wens et al., 2016	22	11978.00	785.00	19	15200.00	1124.00		-3.30	[-4.27; -2.33]	7.4%
Ozkul et al., 2018	36	2233.21	3731.77	18	1520.43	1559.46		0.22	[-0.35; 0.79]	8.2%
Philippova et al., 2020	36	14264.00	2046.00	20	25407.00	1343.00	- 1	-6.00	[-7.28; -4.73]	6.7%
Naegelin et al., 2020	259	29410.00	7240.00	259	32690.00	8330.00		-0.42	[-0.59; -0.25]	8.6%
Oraby et al., 2021	60	384.26	410.42	30	265.47	81.73		0.35	[-0.09; 0.79]	8.4%
Devasahayam et al., 2021	14	56560.00	25120.00	8	57630.00	9480.00		-0.05	[-0.92; 0.82]	7.6%
Overall effect	515			437			•	-0.97	[-1.61; -0.33]	79.1%
Prediction interval									[-3.33; 1.39]	
Heterogeneity: / ² = 93% [89%	6; 96%	6], <i>p</i> < 0.01								
Specimen = Plasma										
Comini Frotaa et al., 2009	29	1241.54	564.43	24	1858.34	933.69		-0.81	[-1.37; -0.24]	8.2%
Al-Temaimi et al., 2017	100	2054.66	1644.32	77	3331.45	3842.47		-0.45	[-0.75; -0.15]	8.5%
Shajarian et al. 2021	45	374.79	3.56	45	437.03	4.09			[-18.53; -13.65]	4.2%
Overall effect	174			146				-5.20	[-8.45; -1.95]	20.9%
Prediction interval	2			2					[-46.37; 35.98]	
Heterogeneity: / ² = 99% [98%	6; >99	%], <i>p</i> < 0.01	1						-	
Overall effect	689			583			•	-1.57	[-2.26; -0.88]	100.0%
Prediction interval									[-4.32; 1.17]	
Heterogeneity: 12 = 96% [94%	6; 97%	[p < 0.01]								
Test for subgroup differences	$x_{1}^{2} =$	6.27, df = 1	(p = 0.01)				-15 -10 -5 0 5 10	15		

В)		E	xperimental	Control			Standardised Mean							
Study	Total	Mean		Total	Mean	SD		Di	ifferenc	е	SMD	9	95%-CI	Weight
Golda et al., 2003	25	4435.10	533.4000	20	4712.20	491.8000			H		-0.53	[-1.13;	0.07]	21.4%
Lalive et al., 2008	15	20400.00	6400.0000	15	19500.00	7200.0000					0.13	[-0.59;	0.85]	0.0%
Comini Frotaa et al., 2009	29	1241.54	564.4305	24	1858.34	933.6881			-		-0.81	[-1.37;	-0.24]	24.2%
Tongiorgi et al., 2012	20	24.45	5.8881	20	29.50	6.4705					-0.80	[-1.45;	-0.15]	18.4%
Comini-Frota et al., 2012	28	1243.17	568.5140	28	1840.39	903.5977		-	-		-0.78	[-1.32;	-0.24]	25.9%
Wens et al., 2016	22	11978.00	785.0000	19	15200.00	1124.0000					-3.30	[-4.27;	-2.33]	0.0%
Al-Temaimi et al., 2017	100	2054.66	1644.3200	77	3331.45	3842.4710					-0.45	[-0.75;	-0.15]	0.0%
Ozkul et al., 2018	36	2233.21	3731.7676	18	1520.43	1559.4575					0.22	[-0.35;	0.79]	0.0%
Philippova et al., 2020	36	14264.00	2046.0000	20	25407.00	1343.0000					-6.00	[-7.28;	-4.73]	0.0%
Naegelin et al., 2020	259	29410.00	7240.0000	259	32690.00	8330.0000					-0.42	[-0.59;	-0.25]	0.0%
Oraby et al., 2021	60	384.26	410.4238	30	265.47	81.7341					0.35	[-0.09;	0.79]	0.0%
Devasahayam et al., 2021	14	56560.00	25120.0000	8	57630.00	9480.0000			- 120		-0.05	[-0.92;	0.82]	10.2%
Shajarian et al. 2021	45	374.79	3.5638	45	437.03	4.0890					-16.09	[-18.53; -	13.65]	0.0%
Overall effect	689			583				-			-0.66	[-0.94;	-0.38]	100.0%
Prediction interval									-			[-1.11;	-0.21]	
Heterogeneity: $l^2 = 0\% [0\%; 79\%], p = 0.61$								1	1	1			-	
						ł	-2	-1	0	1	2			

Figure 2

A: Forest plot of subgroup meta-analysis of BDNF levels in MS patients compared to controls. **B:** Forest plot of meta-analysis of BDNF levels in MS patients removing outliers.

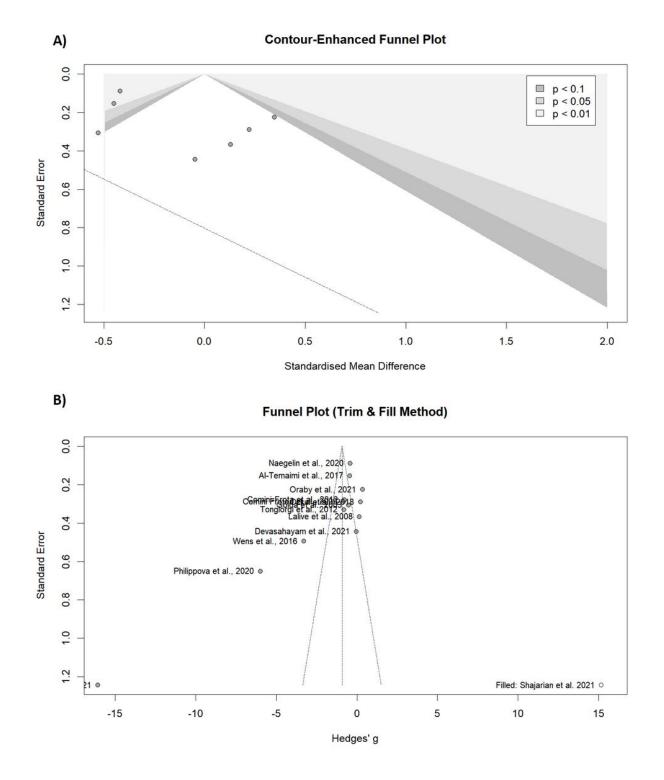


Figure 3

A: The counter-enhanced funnel plot showing evidence of publication bias, statistically supported by Egger's regression test. **B:** The funnel plot resulted from trim and fill analysis

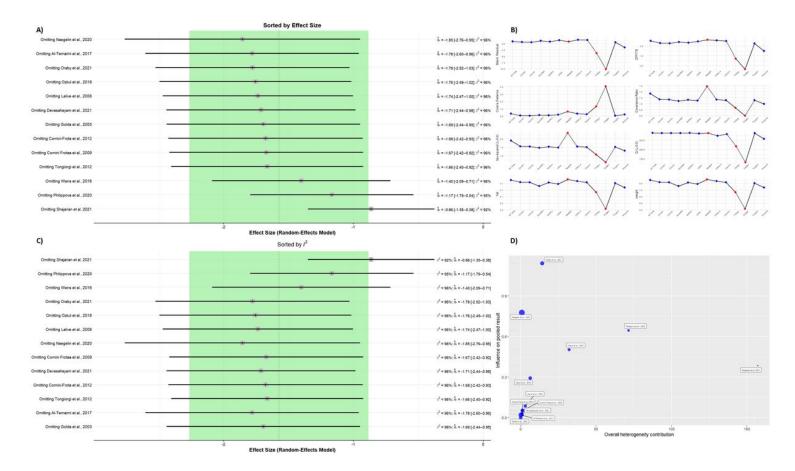
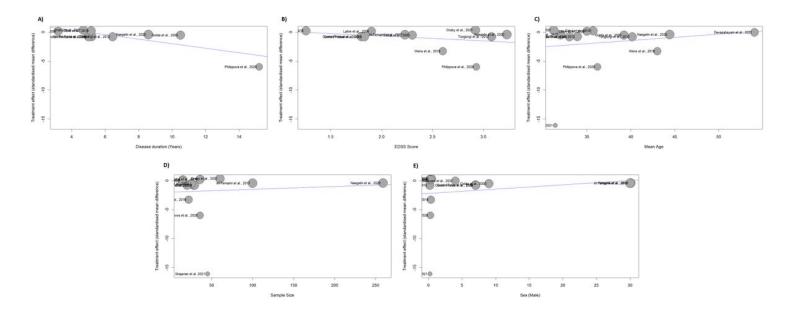


Figure 4

Results of Sensitivity analysis (leave-one-out analysis) of the meta-analysis **A**: Sorted by Effect Sizes; **B**: The influence analysis plot, showing different influence diagnostics including: Externally Standardized Residuals, DFFITS Value, Cook's Distance, Covariance Ratio, Leave-One-Out τ2 and Q Values, and Hat Value and Study Weight.; **C**: Sorted by I²; **D**: Baujat Plot.



Bubble plot of meta-regression **A:** Disease duration; **B:** EDSS Score; **C:** Mean Age; **D:** Sample Size; **E:** Sex (Male)

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- Table1.docx
- Table2.docx