

# Analysis of the difference in characteristics of patients and IVF cycles and the parameters of in vitro embryo culture between normal individuals with high and low blastocyst formation rate

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## Research

**Keywords:** Useful blastocyst formation, normal patients, individual characteristics, basic LH, ovarian response, in vitro embryo culture

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# Abstract

**Background:** The main purpose of this study is to analyze the possible factors that were related with high useful blastocyst formation rate in normal patients.

**Methods:** This was a retrospective cohort study included 706 normal patients (excluding PCOS, DOR, endometriosis and other special diseases), according to the useful blastocyst in vitro formation rate (UBIVFR), the patients were divided into two groups respectively, group A (0-50%, including 50% and group B (50%-100%).

**Results:** There were no significant difference in patient ages between the two groups, compared with group A, the level of basic LH (bLH) was significantly increased. The clinical data showed that, compared with group A, the total Gn dose and the number of dominant follicles, oocytes retrieved, MII mature oocytes, normal fertilized oocytes rate, high-quality embryos rate and FET clinical pregnancy rate were significantly higher than group B.

**Conclusion:** Our study is to investigate the useful blastocyst formation rate in relation to bLH levels in IVF/ICSI cycles, showing that high bLH is associated with higher ovarian response and higher UBIVFR. AMH, bFSH and AFC which were defined as ovarian reserve and ovarian response are seen not associated with the useful blastocyst formation. **Key words:** Useful blastocyst formation; normal patients; individual characteristics; basic LH; ovarian response; in vitro embryo culture

## Background

ART is efficient technology (sometimes it is the only way) that allows infertile couples have a baby. Recent years the ART is developing at a high speed, expending the in vitro culture time from 3 days to 5/6 days is a big progress (1). Day 3 embryo was identified as cleavage embryo while Day 5/6 embryo was blastocyst stage embryo (a later developmental stage) (2). Not all the zygotes have the livebirth-potential, in fact, sometimes only a small number of zygotes can lead a livebirth. Therefore, how to choose the livebirth-potential embryo is very important. It is believed that blastocyst culture will contribute to embryo selection. This is because of some of embryos with limited developmental will be eliminated during expended in vitro culture. Numerous studies have reported that transfer of blastocyst resulted in increasing clinical pregnant and live birth rate and reduced abortion rate (3,4). Due to this, many reproductive centre have developed "all blastocyst culture strategy".

Couples produce more health embryos have more chance to have a baby. However, it is difficult to distinguish "good" embryos from total cleavage embryos that they may have the same morphology score (5). Blastocyst culture somewhat may improve this situation. In another word, more useful blastocyst obtained from a couple means more chance they have to have a baby. Therefore, the useful blastocyst in vitro formation capacity (UBIVFC) for any couples matters. We thought that the useful blastocyst formation rate (UBIVFR) based on a certain number of day 3 embryos for further culture can reflect

UBIVFC. Analysis of the difference in baseline characteristics of patients and IVF cycles between patient with high and low UBIVFC and to guide and improve the therapeutic strategy.

In the present study, from March 2017 to September 2018 in our center, women undergoing IVF/ICSI regimen for the treatment of infertility were recruited, excluding PCOS, DOR, endometriosis and other special diseases.

## Methods

### Patients

A retrospective cohort study was conducted at the Changzhou maternal and child health care hospital. From March 2017 to September 2018, women undergoing IVF regimen for the treatment of infertility were recruited. The study protocol was approved by the Ethics Committee of the Changzhou maternal and child health care hospital. It was conducted based on the Declaration of Helsinki for medical research. All participants provided informed consent after approving for infertility treatments and routine IVF procedures.

Since two D3 high-quality cleavage embryos were transplanted or frozen, more than two remaining embryos were used for expanding culture, only patients with a chance to reach blastocyst stage were included as defined by  $\geq 5$  normal fertilized oocytes. Polycystic ovary syndrome (PCOS), ovarian function decline (DOR), endometriosis and other diseases were excluded from the study population. The remaining patients were our subjects and defined as normal individuals.

### Blastocyst score

All our embryologists are trained and use this classification of Gardner and Schoolcraft (1999) to classify the blastocysts in daily clinical practice (7). Briefly, according to blastocysts degree of expansion and hatching status, blastocysts were given a numerical score from 1 to 6, as follows: 1, An early blastocyst whose blastocoel is less than half the volume of the embryo; 2, A blastocyst in which the blastocoel is half or more of the volume of the embryo; 3, The blastocoel is filled to the full embryo; 4, an expanded blastocyst with a blastocoel volume larger than that of the early embryo, with a thinning zona; 5, a hatching blastocyst with the trophectoderm starting to protrude through the zona; and 6, An incubated blastocyst in which the blastocyst has completely escaped from the zona. For blastocysts graded as 3–6 (full blastocysts onward), the development of the inner cell mass was assessed as follows: A, many cells, tightly packed; B, several cells, loosely grouped; or C, very few cells. The trophectoderm was assessed as follows: A, many cells forming a cohesive epithelium; B, few cells forming a loose epithelium; or C, very few large cells. According to the scoring rules, transferable blastocyst was analysed, not including 4CC, 5CC and 6CC. There were two embryologists with the six years' scoring experience to grade the blastocysts simultaneously in order to reduce the intra-variability and inter-variability.

According to the useful blastocyst formation rate, the patients were divided into two groups respectively, group A (0-50%, including 50%) and group B (50%-100%).

### **Serum hormone analysis**

Serum were collected on menstrual cycle day 3 and the trigger day respectively. Hormones including AMH, FSH, LH, E2 and progesterone levels were measured by chemiluminescence (Abbott Biologicals B.V., The Netherlands). The lower limits of sensitivity were as follows: FSH = 0.06 mIU/ml, LH = 0.09 mIU/ml, E2 = 10 pg/ml and P = 0.1 ng/ml.

### **Statistical Analyses**

All statistical analyses were performed with the use of Graphpad prism 5 (GraphPad Software Inc., San Diego, CA, USA). These data found to be normally distributed were presented as the mean  $\pm$  standard deviation (SD), and were tested to a Student's t-test. Those data that were not normally distributed were presented as the mean (interquartile range), and were analyzed using a Mann-Whitney U-test. Categorical variables were expressed as a frequency (percentage), and analyzed using a Chi-square test. A value of  $P < 0.05$  was considered to indicate statistical significance.

## **Results**

### **Patient characteristics**

No difference in age, and basal FSH, E2, P, T value was observed between the two groups. Similarly, type of infertility (n), patient diagnoses, ovulation promotion program, cycle number and methods of insemination were not significantly different between two groups. However, compared with the group A, the basal luteinizing hormone (bLH) in group B was significantly increased. (Table 1).

### **Follicle development, oocyte performance, and clinical outcomes**

The clinical data in two groups were described in the table 2. Compared with the group A, in group B the total Gn dose and the number of dominant follicles (diameter larger than 14 mm), oocytes retrieved, MII mature oocytes, normal fertilized oocytes rate, high-grade (grade I and grade II) embryos rate and FET

clinical pregnancy rate were significantly higher. The FSH levels were significantly lower ( $p < 0.05$ ) and E2 levels were significantly higher ( $p < 0.05$ ) on the HCG trigger day. There were no statistical differences in LH levels and P levels.

### **The subgroup analyses by age, BMI, bFSH and AFC between two groups**

Table 3 presents that the subgroup analyses by age, BMI, bFSH and AFC were conducted between two groups. In two groups, the proportion of women with advanced ages ( $> 37$  years), obese ( $BMI > 30 \text{ kg/m}^2$ ) and bFSH  $> 10 \text{ mIU/ml}$ , low AFC ( $< 5$ ) was very little respectively ( $< 5\%$ ), the women with low AMH ( $< 0.7 \text{ ng/ml}$ ) was zero. There was no statistical difference in the proportion of different subgroups of the same index.

## **Discussion**

Our study found that the basic LH levels of patients with high UBIVFR were significantly higher than the patients with low UBIVFR ( $5.31 \pm 3.19$  &  $5.68 \pm 2.83$ ) ( $p < 0.001$ ), suggested bLH levels may be associated with the blastocyst formation. The influence of LH on follicle maturation in vitro showed that follicle survival rate was significantly higher in medium supplemented with LH than without (8). Moreover, LH bioactivity had an effect on antral cavity formation (9). Some scholars have found that the threshold of LH is 1.2 in patients with hypogonadotropic hypogonadism (HH) [10]. If the threshold of LH is lower than this, the level of E2 will be low, which will affect the development of follicles. Other literature shows that women with LH less than 10 IU/L have higher pregnancy rate and lower abortion rate than those with basic LH greater than 10 IU/L [11, 12]. The basic LH of patients in our study was less than 10 ( $5.31 \pm 3.19$  &  $5.68 \pm 2.83$ ), it can be inferred that high basic LH in a certain range is better for the blastocyst formation and it may be specific reflective of the "LH window" [13] theory of follicular development.

The data of our study showed that the dosage of Gn in patients with UBIVFR is significantly more than in patients with low UBIVFR. Otherwise, the number of oocytes retrieved is significantly less than that in patients with low UBIVFR. Some study confirmed the "ovarian sensitivity index" (OSI, the ratio between the oocyte yield and the total dose of Gn defined ovarian response which is essential to the performance of assisted reproduction by IVF/ICSI) [14]. Previous study has confirmed an excellent correlation between the parameters of ovarian response and ISO [15]. Calculated that our results suggested ovarian response of patients with high UBIVFR better than the patients with low UBIVFR and it also confirmed that patients in our study are normal population, because the range of ISO with normal response is 1.697/IU-10.07/IU. Clinical outcome data show that the number of MII mature oocytes, normal fertilized oocytes, high-grade (grade I and grade II) embryos and FET clinical pregnancy rate in patients with high UBIVFR was significantly higher and suggested that the ovarian response may be associated with blastocyst formation.

In clinical practice, we often use AMH, bFSH and AFC as calculators of ovarian reserve and ovarian response. Previous study have indicated that elevated bFSH levels ( $> 10 \text{ IU/L}$ ) [16], low level of AMH ( $< 0.7 \text{ ng/ml}$ ) [17] and AFC ( $< 5$ ) [18] reduction correlated with poor response and poor pregnancy

outcomes, but there are no significant differences in three indexes in our study ( $p > 0.05$ ). On the one hand, due to the limited criteria for enrollment, the proportion of the patients with elevated bFSH levels, low level of AMH and AFC is lower and there is no difference, patients with AMH ( $< 0.7$  ng/ml) make up zero in two groups. On the other hand, we speculated that ovarian reserve markers serve as a proxy for follicle quantity but are considered poor predictors of ovarian response in normal patients. Some recent studies have confirmed this thought, the availability of records from 981 women 30–44 years of age without a history of infertility who trying to conceive for 3 months and 8241 patients 21–44 years of age undergoing fresh IVF cycles, do not support the use of bFSH, AMH and AFC to assess ovarian response, oocyte quality and embryo development [19][20]. The present results suggest that bFSH, AMH, and AFC is not associated with the ovarian response and the useful blastocyst formation of normal patients.

It proved that age is still remains the best predictor to oocyte quality and embryo development [21]. Aged oocytes exhibit increased chromosomal abnormalities and dysfunction of organelles, both of which factor affected oocyte quality [22][23]. But in our study there is no difference in the age of different UBIVFR, the reason is that it is possible the average age in the two groups was lower than 35 ( $30.1 \pm 3.62$  &  $29.8 \pm 3.47$ ). The results of age subgroup in the two groups showed that most of the patients were younger than 35 years, and only those older than 35 years accounted for 12.9% (308/354) and 9.4% (320/352) who may have good ovarian reserve and ovarian response. Several studies have confirmed that the reproduction of women decreases gradually but significantly starting approximately at 35 years and subsequently decreases more rapidly after 37 years [24][25]. Female age is independent affected the oocyte quality and blastocyst formation, our data show that age does not affect the formation of useful blastocysts, which may be the enrolled patients had the better ovarian responsiveness.

The limitation of the study is that it is retrospective and the sample is not enough big. Other clinical outcomes, such as live birth rate, fetal health, has not been observed in this study, expanded sample size is necessary to further verification.

## Conclusion

Our study is to investigate the useful blastocyst formation rate in relation to bLH levels in IVF/ICSI cycles, showing that high bLH is associated with higher ovarian response and higher rates of useful blastocyst. AMH, bFSH and AFC which were defined as ovarian reserve and ovarian response is seem not association with the useful blastocyst formation.

## Abbreviations

UBIVFC ☐ useful blastocyst in vitro formation capacity UBIVFR: useful blastocyst in vitro formation rate  
PCOS ☐ Polycystic ovary syndrome DOR: ovarian function decline

SD ☐ standard deviation BMI ☐ body mass index AFC ☐ the number of antral follicles

bLH ☐ the basal luteinizing hormone OSI ☐ ovarian sensitivity index

## **Declarations**

### **Acknowledgements**

None

### **Authors' contributions**

Chun-Mei Yu is the main contributor to study design, data collection, statistical plan, data interpretation, and manuscript draft/revision. Xiu-Liang Dai participated in designing the study, data collection and interpretation, and manuscript revision. Yu-Feng Wang participated in data collection and critically revised the manuscript. Li Chen assisted with gynecological expertise in study design, data interpretation, and manuscript revision. All authors have read and approved the final manuscript.

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### **Availability of data and materials**

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

### **Ethics approval and consent to participate**

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. The study protocol was approved by the Ethics Committee of the Changzhou maternal and child health care hospital(17020490718). Informed consent was obtained from individual participants included in the study.

## Consent for publication

Not applicable.

## Competing interests

The authors declare that they have no competing interest.

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## Tables

Table 1 Patient characteristics compared between two groups

characteristic	GroupA	GroupB	P-value
	354	352	
Ages(years)	30.1[4.0]	29.8[4.0]	0.46
BMI(kg/m2)	23.1[4.7]	22.8[4.2]	0.75
AMH	5.25[4.19]	5.12[2.98]	0.62
Basic FSH	6.45[1.94]	6.35[2.08]	0.89
AFC	11.28[6.00]	12.05[6.25]	0.10
Duration of infertility	3.19[2.0]	3.17[2.0]	0.88
Basic LH	5.31[3.20]	5.68[3.04]	0.0006
Basic E2	38.74[20.7]	34.51[21.7]	0.53
Basic P	7.21[20.5]	9.15[28.6]	0.16
Basic T	24.26[14.86]	23.81[15.57]	0.54
Partner Ages[years]	31.7[6.0]	31.5[6.0]	0.61
DFI	11.84[8.94]	11.79[8.66]	0.64
Partner BMI(kg/m2)	24.92[6.0]	24.98[5.0]	0.59
Diagnoses			
Endometriosis	12/345	18/345	0.35
Male factor	30/345	32/345	0.89
Ovulation disorders	43/345	37/345	0.55
Tubal factor	223/345	206/345	0.21
Unexplained	34/345	48/345	0.13
Infertility type			
Primary infertility	188/354	172/352	0.29
Secondary infertility	166/354	180/352	0.29
Ovulation promotion program			
GnRH-a program	254/354	272/352	0.10
Others	100/354	80/352	0.10

Table 2 The cycle characteristics of controlled ovarian stimulation in two regimens

characteristic	GroupA	GroupB	P-value
	354	352	
Gn dose (IU)	1883[1275]	1664[962]	0.001
Gn duration (d)	8.96[2.0]	8.68[1.0]	0.07
>14 mm follicles on hCG administration day (n)	13.87[6.25]	14.66[6.75]	0.02
Trigger day			
FSH(IU/L)	12.02[7.52]	10.84[5.42]	0.02
LH(IU/L)	2.13[1.30]	2.09[1.38]	0.80
E2 (ng/L)	4078[2446]	4450[2539]	0.02
P (ng/L)	0.90[0.49]	0.83[0.43]	0.07
Oocytes retrieved	12.20[4.25]	12.87[5.0]	0.008
MII oocytes (n)	11.56[5.0]	12.36[4.75]	0.004
Normal Fertilized	3308/4094 80.8%	3565/4349 82.0%	0.17
Oocytes rate (%)			
Top-quality embryosrate (%)	2200/3308 66.5%	2949/3565 82.7%	0.0001
FET clinical pregnancy rate	32/62 51.6%	45/61 73.7%	0.01

Table 3 The proportion of traditional ovarian reserve marker between two groups

characteristic	GroupA	GroupB	P-value
	354	352	
<b>Ages(years)</b>			
≤35	308/354 29.1±0.15	320/352 29.1±0.15	
35-37	31/354 35.7±0.77	25/352 35.7±0.79	0.15 3.78
≥37	15/354 39.0±1.13	7/352 40.0±2.88	
<b>body mass index(kg/m2)</b>			
≤18.5 0underweight	31/329 17.64±0.68	27/348 17.5±0.82	
18.5-24.9 0normal	228/329 21.43±1.71	247/348 21.36±1.64	0.86 0.76
25-29.9 0overweight	62/329 26.67±1.37	64/348 26.50±1.23	
≥30 0obese	8/329 32.29±1.16	10/348 33.05±2.69	
<b>Antimullerian hormone</b>			
≤0.7ng/ml	0	0	0.89 0.51
0.7-8.4ng/ml	212/244 4.06±2.0	224/260 4.09±1.62	
≥8.5ng/ml	32/224 12.66±3.55	36/260 11.51±2.75	
<b>Follicle stimulation hormone serum</b>			
≤10mIU/ml	334/343 6.28±1.40	333/344 6.21±1.60	0.82 0.99
≥10mIU/ml	9/343 11.96±2.96	11/344 11.15±1.32	
<b>Number of antral follicles</b>			
≤5 0a	6/220	3/243	
5-10 0a	96/220	101/243	0.43 1.70
≥10 0a	118/220	139/243	