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1 **Nanocellulose/chitosan biohybrid aerogel bead adsorbent for removal of**
2 **diclofenac sodium from wastewater: preparation, characterization, and**
3 **adsorption mechanism**

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15 **Abstract:** In this work, a novel nanocellulose/chitosan biohybrid aerogel (NCBA)
16 bead adsorbent was successfully prepared from oxidized bamboo nanocellulose and
17 chitosan to remove diclofenac sodium (DS) from wastewater. Fourier transform
18 infrared spectroscopy, scanning electron microscopy, and X-ray photoelectron
19 spectroscopy were used to characterize the preparation of NCBA bead adsorbent. The
20 results demonstrated that the adsorption kinetics were well described by the
21 pseudo-second-order model, and the adsorption isotherms were well fitted by the Sips
22 isotherm model with the maximum adsorption capacity of 321.26 mg/g on DS, which

23 is much higher than most of the previously reported works. The absorption
24 mechanism was governed by electrostatic interaction and hydrogen bond interaction.
25 Moreover, the resulting bead remained a high removal efficiency after 5 regeneration
26 cycles. In summary, it had been proved that the aerogel bead was an efficient
27 bio-sorbent for removing DS from aqueous solutions.

28 **Keywords:** Adsorption, Adsorbent, Nanocellulose, Chitosan, Diclofenac sodium

29 **1. Introduction**

30 Diclofenac sodium has been a widely used nonsteroidal anti-inflammatory drug,
31 which is commonly applied for medical purposes, such as analgesic, anti-arthritic and
32 antirheumatic agent [1]. Owing to its broad applications, residual DS is found in
33 groundwater and drinking water around the world [2]. Although DS can be degraded
34 via natural and other auxiliary methods [3], residual DS in the environment is still a
35 potential toxic contaminant. It can cause adverse gastrointestinal reactions and acute
36 renal insufficiency [4]. Many efforts, including biodegradation [5], nanofiltration [6],
37 adsorption [7], and photocatalysis [8], have been made to remove the residual DS in
38 aqueous solution. Among them, adsorption is the most popular method due to its
39 low-cost, simple operation, and reusability [7-12]. Aerogels are defined as
40 three-dimensional networks of hydrophilic polymers, which also can be easily
41 separated from water [13]. They have received widespread attention for DS removal
42 because of their low density, high porosity, and high removal efficiency [14, 15].

43 Compared with synthetic polymer-based aerogels, the natural-based ones (such
44 as cellulose, chitosan, and sodium alginate) are particularly attractive because of

45 their biodegradability, reproducibility and green preparation process [16]. Biohybrid
46 aerogel prepared from TEMPO-oxidized cellulose and chitosan has a good
47 performance on the adsorption of pollutants from wastewater because it can combine
48 the advantages of cellulose and chitosan, including ample resources, renewability, and
49 functional groups [17]. Physical cross-linking is a widely used method to produce
50 biohybrid hydrogels *via* electrostatic attraction, hydrogen bonding, and hydrophobic
51 interactions [18]. After freeze-drying, the physically cross-linked biohybrid aerogels
52 will have a 3D porous network with abundant functional groups (-OH, -COOH, and
53 -NH₂) [19]. Typically, the physically cross-linked biohybrid aerogels have relatively
54 weak mechanical properties [20]. Therefore, the physically cross-linked aerogels need
55 to be modified to improve their structural stability.

56 Alum is one of the most widely used coagulants to enhance the physically
57 cross-linked aerogels in the wastewater treatment industry [21]. It will rapidly
58 produce a great number of cationic metal species (aluminum hydroxide) after
59 dissolving in water [22]. These cationic metal species can efficiently adsorb
60 negatively charged contaminants. Therefore, the introduction of alum can not only
61 enhance the stability of physically cross-linked aerogel but also improve the
62 adsorption efficiency. γ -glycidoxypropyl trimethoxysilane (GPTMS) is a promising
63 alkoxy silane coupling agent [23]. It will functionalize as a coupling agent to create
64 covalently bonded compounds between chitosan and cellulose [24]. Therefore, it will
65 enhance the graft strength of alum onto cellulose and chitosan. Meanwhile, it also
66 enhances the mechanical strength of physically cross-linked aerogels [25].

67 Biohybrid aerogels usually have tunable surface characteristics through surface
68 modification, which makes them more capable of absorb pollutants [26].
69 Polyethyleneimine (PEI), a hydrosoluble cationic polyelectrolyte, has been recognized
70 by its abundant amino groups [3]. Meanwhile, it has been extensively used to adsorb
71 dyes [27], heavy metal ions [28], and pharmaceutical residues [29]. Thus, PEI can be
72 used as an amino-functionalized agent to boost the adsorption performance of
73 biohybrid aerogel towards DS.

74 Herein, with the aim of efficiently removing DS from wastewater, a
75 nanocellulose/chitosan biohybrid aerogel (NCBA) bead adsorbent was prepared from
76 waste bamboo paper and chitosan. The NCBA bead was characterized by FTIR, XPS,
77 and SEM. In addition, these vital factors that influence the adsorption properties of
78 NCBA bead were investigated and the DS removal mechanisms were studied as well.

79 **2. Material and methods**

80 **2.1 Materials**

81 Waste bamboo paper was collected from the laboratory. Glacial acetic acid,
82 polyethyleneimine (PEI), γ -glycidyloxypropyl trimethoxysilane (GPTMS), chitosan
83 (CS), and diclofenac sodium (DS) were purchased from Aladdin Chemical Reagent
84 Co. Ltd. (Shanghai, China). All chemicals were used as received.

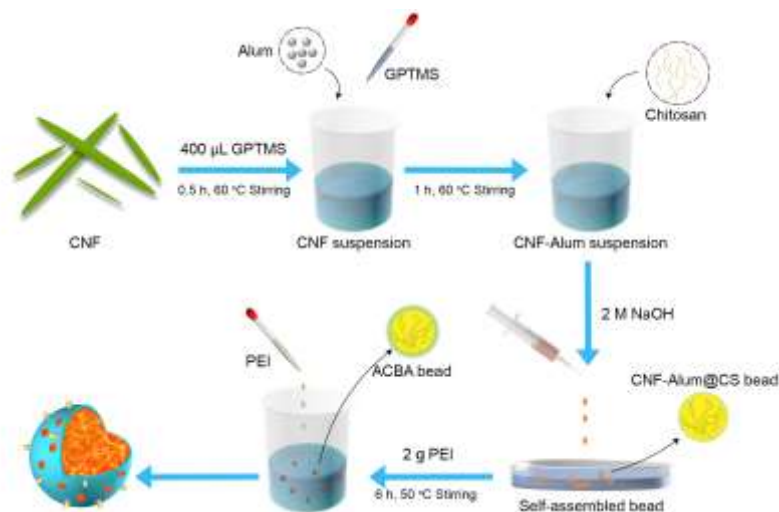
85 **2.2 Preparation of CNF-Alum**

86 The oxidized bamboo nanocellulose (CNF) was prepared according to our
87 previous work [17]. Oxidized CNF (0.5 g) was dissolved in 50 mL distilled water.
88 The suspension was then sonicated for 10 min. Then, GPTMS (400 μ L) was added to

89 the solution and subsequently, the solution was stirred at 60°C for 0.5 h. After the
90 reaction was completed, the solution was mixed with 0.25 g alum under stirring for 1
91 h to prepare CNF-Alum.

92 2.3 Preparation of NCBA bead

93 The CNF-Alum (50 mL) suspension was mixed with acetic acid (1 mL) under
94 constant stirring at 60 °C for 20 min. Then, the CNF-Alum suspension was mixed
95 with 1 g CS. The CNF-Alum@CS suspension was stirred rapidly at 50 °C until a
96 homogeneous mixture was obtained. The CNF-Alum@CS suspension was dropwise
97 added into a precipitation bath (1 M NaOH) to form CNF-Alum@CS bead. The
98 obtained bead was washed with deionized water. Then, it was mixed with deionized
99 water (200 mL) and PEI (2 g) at 50 °C for 6 h to produce an amino-functionalized
100 nanocellulose/chitosan biohybrid aerogel (NCBA) bead, and then the resulting bead
101 was freeze-dried for 48 h. The synthetic procedure of NCBA bead is shown in **Fig. 1**.



102

103

Fig. 1. Synthetic procedure of NCBA bead.

104

2.4 Characterization

105 The morphology of NCBA bead was carried out by scanning electron
106 microscopy (SEM, ZEISS Gemini 300). The chemical structure of NCBA bead was
107 characterized by Fourier-transform infrared spectroscopy (FTIR, Nicolet 670), and
108 X-ray photoelectron spectroscopy (XPS, Thermo Scientific K-Alpha). The point of
109 zero charge (pH_{PZC}) of NCBA bead was measured according to the pH drift method
110 [17].

111 2.5 Adsorption and desorption experiments

112 The adsorption experiments were performed by placing 50 mg NCBA bead in a
113 series of vials containing 50 mL DS solution with desired concentration. After
114 shaking the solution at 180 rpm at 25 °C for a predefined time. The residual DS
115 concentration was measured by UV-vis spectrometer at the DS maximum adsorption
116 wavelength of 276 nm. The effect of initial DS concentration on the adsorption was
117 investigated from 50 to 800 mg/L. The effect of contact time was carried out with
118 adsorption time from 0 to 240 min. All tests were conducted in triplicate. The
119 adsorption capacity (q_e , mg/g) and removal ratio (R , %) were calculated as follows:

$$120 \quad q_e = \frac{C_0 - C_e}{m} V \quad (1)$$

$$121 \quad R = \frac{C_0 - C_e}{C_0} \times 100 \quad (2)$$

122 where: q_e (mg/g), C_0 (mg/L), C_e (mg/L), m (mg), and V (mL) are adsorption capacity,
123 the initial and equilibrium concentrations, adsorbent dosage and solution volume,
124 respectively.

125 Desorption experiments were performed as follows. The initial DS concentration,
126 solution pH, and adsorbent dose were 100 mg/L, 5.2, and 1 g/L, respectively. After 24

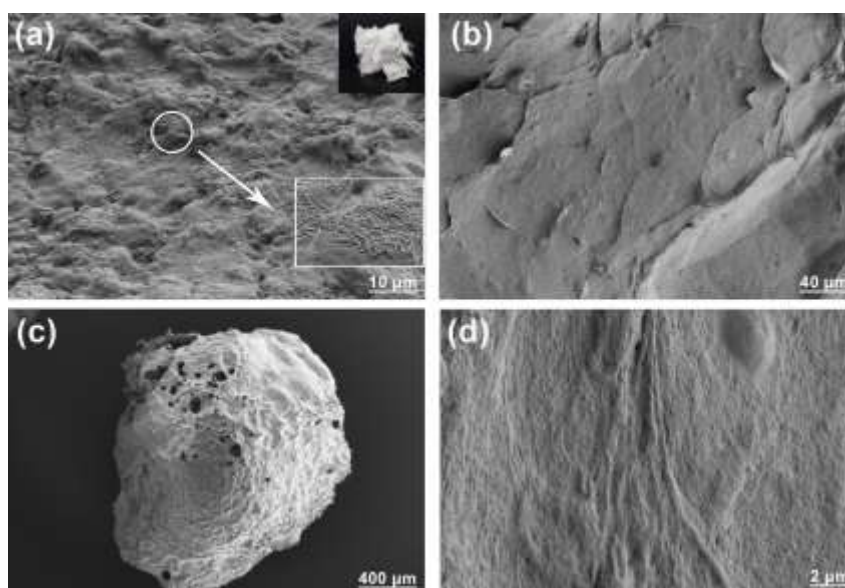
127 h adsorption, the residual concentration of DS was measured. The NCBA bead was
128 soaked in NaOH solution (0.5 mol/L) for 4 h, followed by vacuum-dried at 60 °C for
129 the next cycle. It was repeated 5 times to study its reusability.

130 3. Results and discussion

131 3.1 Characterization of NCBA bead

132 3.1.1 SEM analysis

133 As shown in **Fig. 2a**, the surface of CNF was covered by alum, which exhibited a
134 loose surface structure of CNF-Alum, suggesting the successful grating of alum onto
135 CNF [30]. Interestingly, a smoother surface of CNF-Alum@CS bead was formed
136 after introducing CS (**Fig. 2b**), which may be ascribed to the strong inter-hydrogen
137 and intra-hydrogen bond forces [31]. In addition, a rough bead surface with irregular
138 pores could be observed on NCBA bead after involving in PEI (**Fig. 2c and d**), which
139 was expected to have a large specific area. This structure was benefit to providing
140 more accessible adsorption sites for DS.



141

142

Fig. 2. SEM images of (a) CNF-Alum; (b) CNF-Alum@CS; (c-d) NCBA bead.

143 3.1.2 FTIR analysis

144 **Fig. 3** shows the FTIR spectra of CNF-Alum, CNF-Alum@CS, and NCBA bead.
145 It was observed that the CNF-Alum had characteristic peaks at 3440, 1720, 1610, and
146 897 cm^{-1} , which associated with the symmetry vibrations of -OH, C=O, C-OH, and
147 glucosidic ring, respectively. Additionally, the existence of three adsorption peaks at
148 1050, 634, and 475 cm^{-1} were assigned as asymmetric stretch vibrations from Si-O-Al
149 and Si-O-Si [32]. It indicated that the alum was successfully grafted on the oxidized
150 bamboo nanocellulose *via* cross-linking with GPTMS. In the spectrum of
151 CNF-Alum@CS, the characteristic peaks at 1650, 1520, and 1230 cm^{-1} were
152 attributed to the stretching vibration of C=C and C=O, which evidenced the
153 introduction of CS [33, 34]. As compared with CNF-Alum@CS, the band at 1600
154 cm^{-1} was weakened in NCBA bead, which may be ascribed to the Schiff base reaction
155 with grafting PEI [35]. It was noted that the N-C-N asymmetric stretching at 1453
156 cm^{-1} was observed in the NCBA bead. It was also confirmed that the PEI
157 (amino-functionalization) was successfully grafted onto the surface of
158 CNF-Alum@CS [36].

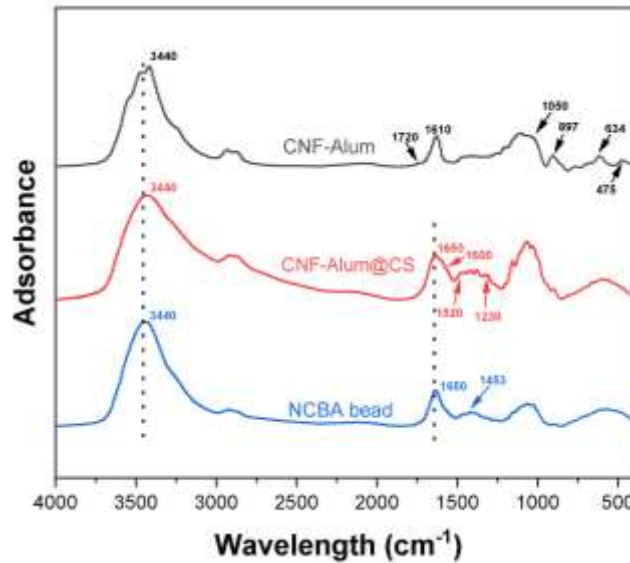
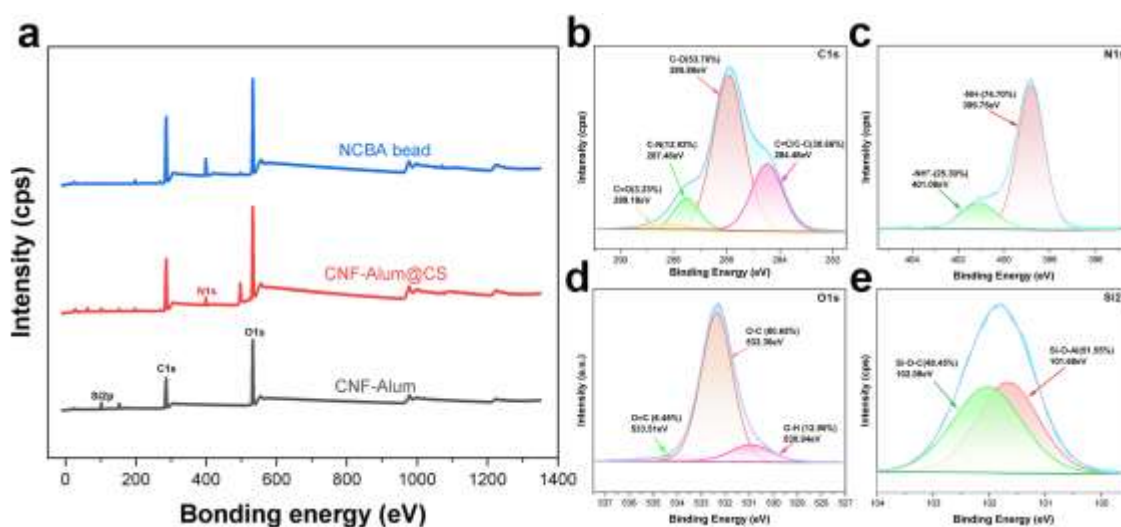


Fig. 3. FTIR spectra of CNF-Alum, CNF-Alum@CS, and NCBA bead.

3.1.3 XPS analysis

The XPS spectra of CNF-Alum, CNF-Alum@CS, and NCBA bead were recorded to investigate the element states on their surfaces. As shown in Fig. 4a, the peaks at 101.8, 286.08, 399.08, and 532.08 eV were related to Si2p, C1s, N1s and O1s electrons, respectively. As compared with CNF-Alum, the intensities of C1s and O1s were significantly increased in the NCBA bead. In addition, a new peak of N1s was observed in CNF-Alum@CS, while the peak of Si2p disappeared in CNF-Alum. These results suggested that CNF-Alum was covered by CS [37]. For NCBA bead, peaks at 284.88, 285.88, 287.48, and 288.18 eV in the high-resolution C1s spectrum (Fig. 4b) were attributed to C=C/C-C, C-O, C-N, and C=O, respectively [28]. This observation was in agreement with the FTIR results and confirmed the grafting of PEI onto CNF-Alum@CS. Two peaks ascribed to -NH₂ (399.75 eV) and -NH₃⁺ (401.08 eV) were observed in the high-resolution N1s spectrum of the NCBA bead (Fig. 4c), indicating the abundance of amino groups that resulted from the introduction of -NH₂

175 from CS and PEI [38]. In the O1s spectrum (**Fig. 4d**), three peaks at 530.94, 532.36,
 176 and 533.51 eV corresponded to O-H, O-C, and O=C, respectively [28]. It
 177 demonstrated the abundant carboxyl groups and hydroxyl groups of NCBA bead.
 178 These results further indicated that the physical cross-linking included electrostatic
 179 attraction ($-\text{COO}^-$ and $-\text{NH}_3^+$) and hydrogen bonding ($-\text{OH}$ groups) [39]. As shown in
 180 **Fig. 4e**, the peaks at 102.08 and 101.68 eV were associated with Si-O-C and Si-O-Al,
 181 respectively, proving that the alum was successfully grafted on oxidized bamboo
 182 nanocellulose *via* cross-linking with GPTMS. The peak of Si-O-Al was mainly due to
 183 the reaction between $\text{Si-OC}_2\text{H}_5$ or Si-OH and $-\text{OH}$ in alum when dissolving in water
 184 [40].



185
 186 **Fig. 4.** (a) High-resolution XPS spectra of CNF-Alum, CNF-Alum@CS, and NCBA bead;
 187 high-resolution XPS spectra of (b) C1s, (c) N1s, (d) O1s, and (e) Si2p of NCBA bead.

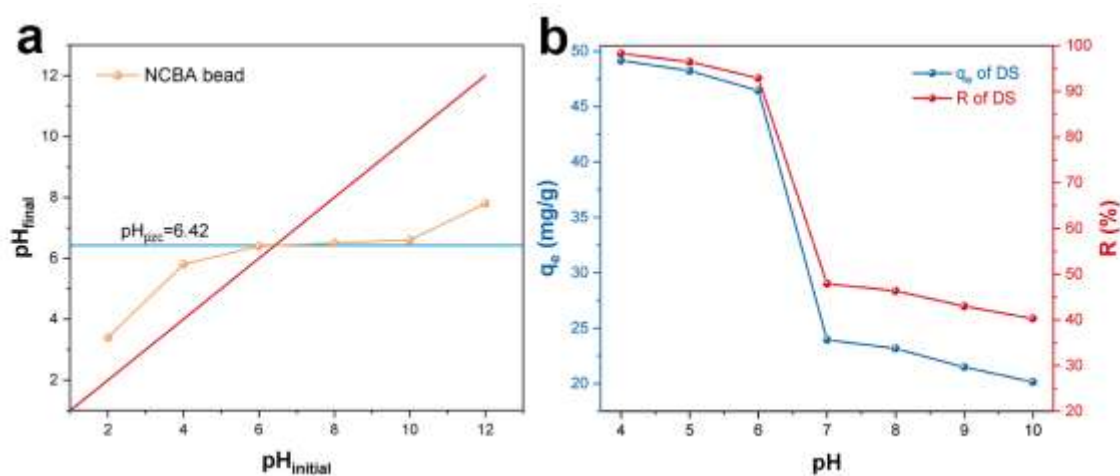
188 3.2 Adsorption and desorption of DS

189 3.2.1 Effect of initial pH

190 The solution pH is an essential parameter in water remediation by adsorption.
 191 The point of zero charge (pH_{PZC}) is an independent model to characterize the surface

192 charge of adsorbent in an aqueous solution. The pH_{PZC} of NCBA bead was 6.42, as
 193 shown in **Fig. 5a**. It indicated that the surface of NCBA bead was negatively charged
 194 at the solution $pH > 6.42$, while its surface was positively charged at the solution pH
 195 < 6.42 . Therefore, the electrostatic attraction between diclofenac anions and the
 196 NCBA bead is promoted at low pH solutions.

197 The adsorption capacity of DS was decreased from 49.17 to 20.15 mg/g and DS
 198 removal efficiency decreased from 98.31% to 50.08% with increasing solution pH
 199 from 4 to 10 (**Fig. 5b**). This was expected because the deprotonation of the amino
 200 ($-NH_2$) and carboxyl ($-COO^-$) groups resulted in a negatively charged NCBA bead
 201 surface that induced the electrostatic repulsion between the NCBA bead and
 202 diclofenac anions, thereby reducing the diclofenac uptake [7].



203

204

Fig. 5. (a) Zero point position; (b) The effect of pH on DS solution.

205 3.2.2 Adsorption kinetics

206 As shown in **Fig.6**, the adsorption capacity was very fast within the first 60 min.

207 Then, it slowed down until reaching the equilibrium at 240 min. The DS removal

208 efficiency increased from 22.65% to 70.30% by prolonging the contact time. It was

209 ascribed to the available adsorption sites in NCBA bead. They were gradually
210 occupied with prolonging contact time.

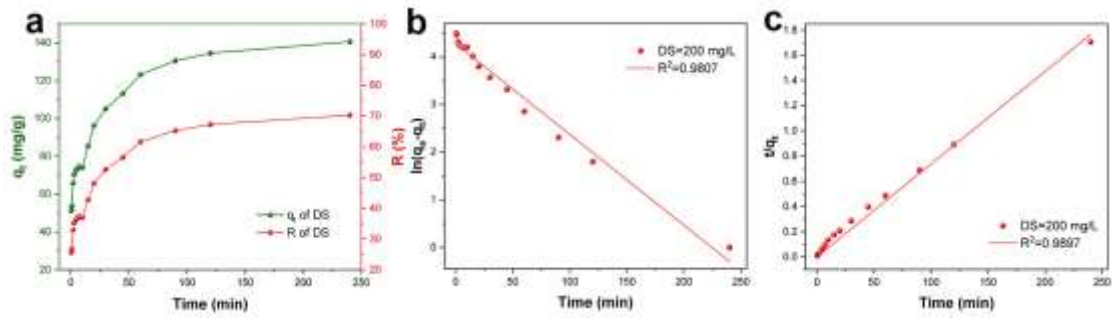
211 In order to study the adsorption process, the kinetic experimental data of DS
212 adsorption by NCBA bead were fitted using pseudo-first-order and
213 pseudo-second-order models [34, 41]. The corresponding equations were shown as
214 follows:

215 Pseudo-first-order equation: $\ln(q_e - q_t) = \ln q_e - k_1 t$ (3)

216 Pseudo-second-order equation: $\frac{t}{q_t} = \frac{1}{k_2 q_e^2} + \frac{t}{q_e}$ (4)

217 where q_e (mg/g) and q_t (mg/g) represent the adsorbate uptakes at equilibrium and time
218 t (min), respectively. k_1 (1/min) and k_2 (g/(mg, min)) are the pseudo-first-order and
219 pseudo-second-order rate constant, respectively.

220 **Fig.6** and **Table 1** show the fitting curves and parameters, respectively. The
221 correlation coefficient ($R^2=0.9897$) of pseudo-second-order kinetic model was higher
222 than that of pseudo-first-order kinetic model. Additionally, the q_e (135.82 mg/g)
223 calculated from pseudo-second-order kinetic model was in good agreement with the
224 experimental data (140.60 mg/g). These findings indicated that the adsorption process
225 of DS was well described by the pseudo-second-order kinetic model. This model
226 suggested that the adsorption process of DS was mainly controlled by chemical
227 adsorption involving electron exchange between DS and adsorbent [41].



228

229 **Fig. 6.** (a) Effect of contact time on DS uptake; (b) Linear plots of pseudo-first-order for DS uptake; (c)

230 Linear plots of pseudo-second-order model for DS adsorption.

231

Table 1 Kinetic parameters for the adsorption of DS by NCBA bead.

Isotherm models	Parameters	Adsorbate
		DS
Pseudo-first-order	q_e	71.8093
	k_1	0.0190
	R^2	0.9807
Pseudo-second-order	q_e	135.8229
	k_2	-0.0005
	R^2	0.9897

232

233 3.2.3 Adsorption isotherms

234 The concentration-dependent adsorption of DS by NCBA bead was studied with
 235 an initial concentration from 50 to 800 mg/L at 25 °C (pH= 5.0). As shown in **Fig. 7**,
 236 the adsorption capacity was increased with increasing initial DS concentration. It
 237 could be ascribed to the increase of mass driving force when the initial DS
 238 concentration increased [45].

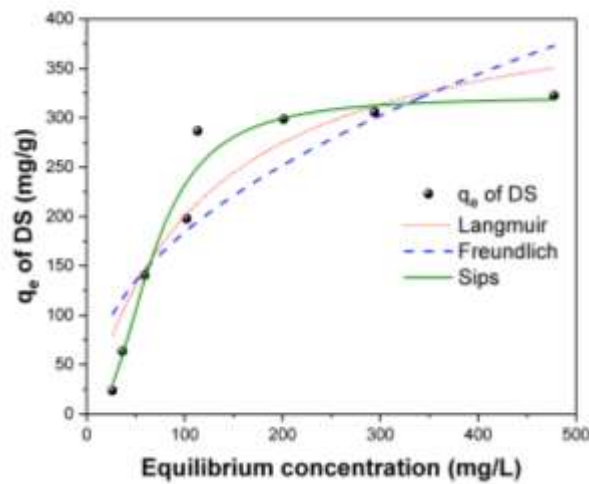
239 Adsorption isotherms are widely applied to describe the adsorption behavior.
 240 The equilibrium data were evaluated by the Langmuir, Freundlich, and Sips isotherm
 241 models [43, 44]. The corresponding equations were expressed as follows:

242 Langmuir equation:
$$\frac{C_e}{q_e} = \frac{1}{bq_m} + \frac{C_e}{q_m} \quad (5)$$

243 Freundlich equation: $\log q_e = \log K_F + \frac{1}{n} \log C_e$ (6)

244 Sips equation: $q_e = \frac{q_m (k_s C_e)^\gamma}{1 + (k_s C_e)^\gamma}$ (7)

245 where q_m is the maximum adsorption capacity, $1/n$ and $1/b$ are the Langmuir and
 246 Freundlich constants, respectively, and K_F is a constant representing the adsorption
 247 capacity. K_S is the Sips constant, and γ represents the surface inhomogeneity of the
 248 adsorbent.



249
 250 **Fig. 7.** Isotherm plots for the adsorption of DS by NCBA bead.

251 **Table 2** Isotherm parameters for the adsorption of DS by NCBA bead.

Isotherm models	Parameters	Adsorbate DS
Langmuir	k	0.0085
	q_m	437.0703
	R^2	0.8818
Freundlich	K_F	22.9980
	n	2.2140
Sips	R^2	0.9708
	q_m	321.2583
	K_S	0.0387
	R^2	0.9702

252
 253 The fitting results are shown in **Fig. 7**, and the fitting data are given in **Table 2**.
 254 The R^2 value (0.9702) calculated from the Sips isotherm model was much higher than

255 other models (**Table 2**), indicating that the Sips isotherm described well the
 256 adsorption of DS by NCBA bead. This model indicated that the adsorption process of
 257 DS was followed by a combined model: monomolecular (at high DS concentration)
 258 and diffuse (at low DS concentrations) [46]. The maximum adsorption capacity
 259 obtained from the Sips isotherm model was 321.26 mg/g for DS, which was higher
 260 than most of the previously reported works (**Table 3**).

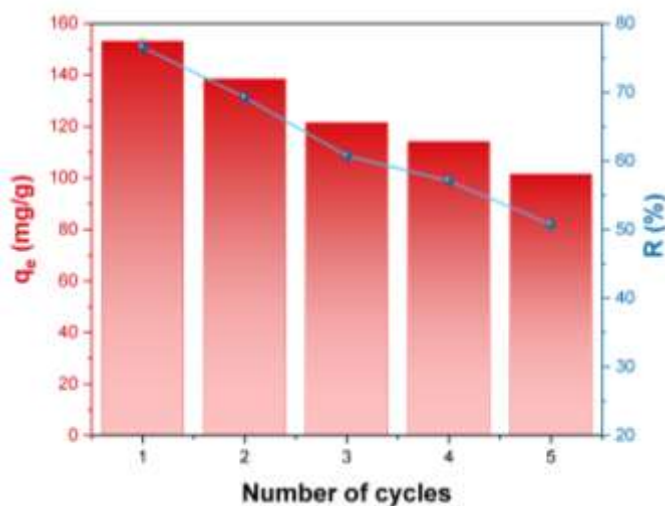
261 **Table 3** Comparison of the maximum DS adsorption capacity by various adsorbents.

Adsorbents	q_{max} (mg/g)	Adsorption time (min)	Ref.
Chitosan-based magnetic composite	196.00	720	[38]
CNC-ED@CS-ED	444.44	50	[29]
rGO	56.67	200	[2]
Zeolitic-CTAB	60.58	90	[47]
Fe ₃ O ₄ @SiO ₂ /SiHTCC	240.40	300	[45]
Carbon xerogels	182.50	2880	[48]
(PAA/PEI) ₂₅	32.42	120	[49]
NCBA bead	321.26	240	This work

262 3.2.4 Reusability of adsorbents

263 To evaluate the regeneration property of the NCBA bead, five
 264 adsorption-desorption cycle experiments were carried out using NaOH as eluent. The
 265 removal efficiency remained above 50% after five adsorption-desorption processes

266 (Fig. 8), indicating that it had excellent reusability. The adsorption capacity was
267 decreased from 153.82 to 103.74 mg/g after five cycles. It was probably attributed to
268 the loss weight of adsorbent during the recycling process [50].



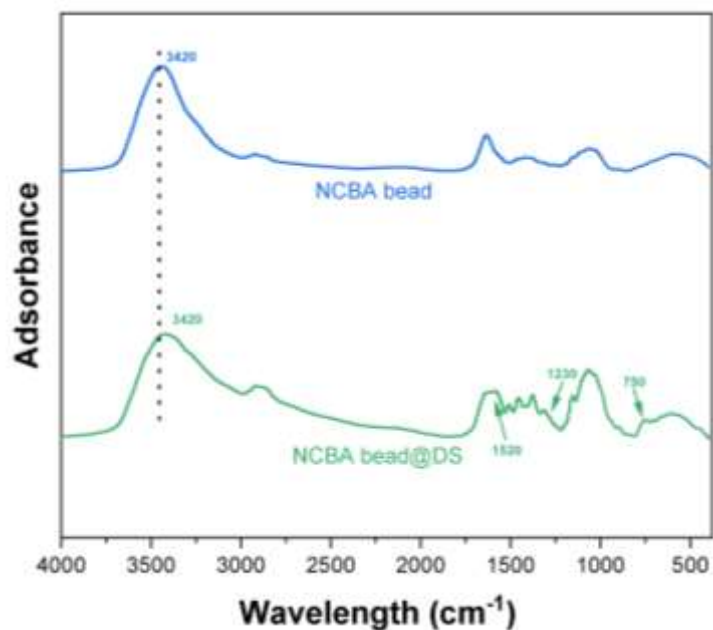
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270

Fig. 8. Five cycles of DS adsorption and desorption.

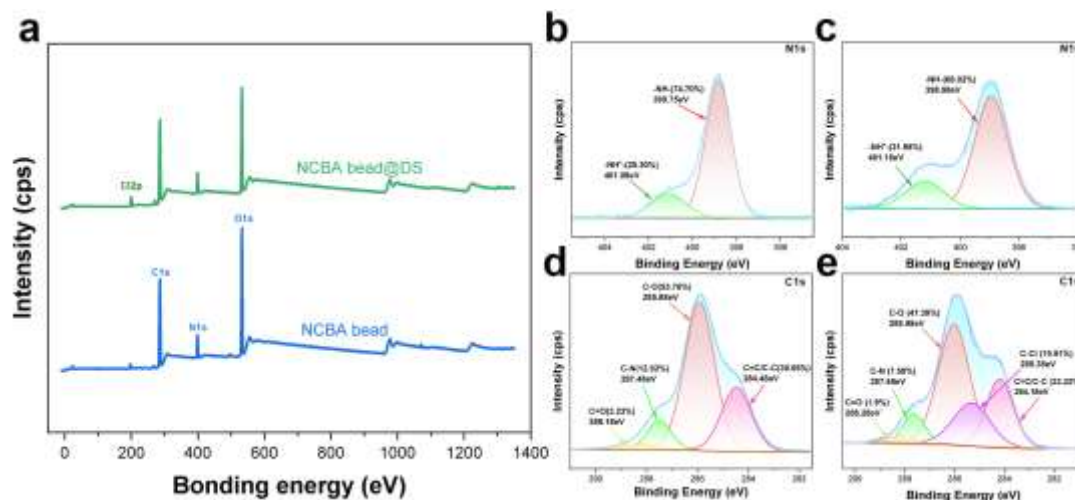
271 3.3 Adsorption mechanism study

272 FTIR spectra of NCBA bead before and after DS adsorption are shown in Fig. 9.
273 After adsorbing DS, the characteristic peak of hydroxyl groups was red-shifted to
274 3420 cm⁻¹ and its intensity was decreased, indicating that the hydrogen bonding was
275 involved in DS adsorption process. The bands at 1520, 1230, and 750 cm⁻¹ were
276 assigned to the stretching vibration of C=O and C-Cl in DS [51]. After adsorption,
277 these characteristic peaks appeared in NCBA bead@DS, suggesting successful DS
278 adsorption.



279
280 **Fig. 9.** The FTIR of NCBA bead before and after adsorption DS.

281 XPS results are presented in **Fig. 10**. After absorbing DS, the intensities of C1s
282 and O1s were slightly decreased in NCBA bead@DS, and both of them shifted to
283 lower binding energy. Besides, a new peak of Cl2p was found in NCBA bead@DS.
284 These results indicated successful DS adsorption [51]. As shown in **Fig. 10b**, the N1s
285 spectrum was divided into two peaks, corresponding to -NH₂ and protonated -NH₃⁺,
286 respectively. After DS adsorption, the -NH₃⁺ increased, while the -NH₂ decreased
287 (**Fig. 10c**). It was probably due to the electrostatic attraction between -NH₃⁺ and DS
288 [29]. **Fig. 10d** displayed that the C1s spectrum consisted of four components, which
289 were associated with C=O, C-N, C-O, and C=C/C-C, respectively. After DS
290 adsorption, the C=O and C-O peaks were shifted to lower binding energies, indicating
291 that the adsorption of DS was triumphant [51]. A new characteristic peak C-Cl could
292 be seen in C1s spectrum (**Fig. 10e**). The Cl element was the characteristic element in
293 DS, also confirming the successful DS adsorption [52].



294

295 **Fig. 10.** (a) High-resolution XPS spectra of NCBA bead before and after adsorption; (b) N1s and (d)

296 C1s before adsorption; (c) N1s and (e) C1s after adsorption.

297 The DS adsorption mechanism was proposed in **Fig. 11**. At solution pH=

298 4.20-6.42, the amino groups on the NCBA bead surface were protonated ($-\text{NH}_3^+$)

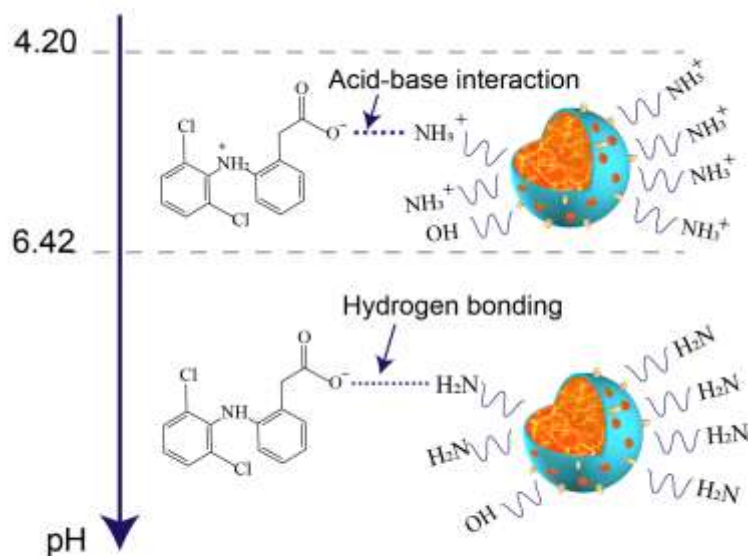
299 acquiring positive charges, which favored electrostatic attraction with the groups of

300 DS. In addition, the hydrogen bonding between $-\text{OH}$ groups (oxidized bamboo

301 nanocellulose, $\text{Al}(\text{OH})_3$, GPTMS, and CS) and deprotonated DS had a positive

302 contribution to the DS adsorption. At solution pH > 6.42, the removal mechanism was

303 the DS interaction with hydroxyl groups on the surface of the NCBA bead.



304

305 **Fig. 11.** Proposed schematic diagram of DS adsorption mechanism by NCBA bead.

306 **4. Conclusion**

307 In this study, a nanocellulose/chitosan biohybrid aerogel (NCBA) bead adsorbent
 308 was prepared for removing DS from aqueous solutions. The point of zero charge
 309 (pH_{PZC}) obtained from the pH drift method was 6.42. The adsorption of DS was fitted
 310 well by pseudo-second-order kinetic model, suggesting the involvement of chemical
 311 adsorption. The adsorption process of DS was well described by the Sips isotherm
 312 model. The maximum adsorption capacity for DS was evaluated as 321.26 mg/g
 313 according to the Sips isotherm model. Additionally, the adsorption mechanism was
 314 governed by electrostatic interaction and hydrogen bond interaction. This study
 315 indicated that the NCBA bead had a great potential application for DS wastewater
 316 purification.

317 **Declaration of Competing Interest**

318 The authors declare that they have no known competing financial interests or

319 personal relations that have appeared to influence the work reported in this paper.

320 **Acknowledgment**

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323 Wildlife Research Center, Mississippi State University.

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