

# Cellulose Based Cation-Exchange Fiber as Filtration Material for the Rapid Removal of Methylene Blue from Wastewater

**Xinyi Shao**

Shaanxi University of Science and Technology Xi'an Campus: Shaanxi University of Science and Technology

**Jian Wang** (✉ [zzwangjian@sust.edu.cn](mailto:zzwangjian@sust.edu.cn))

Shaanxi University of Science and Technology <https://orcid.org/0000-0003-1827-0239>

**Zetan Liu**

Shaanxi University of Science and Technology Xi'an Campus: Shaanxi University of Science and Technology

**Na Hu**

Shaanxi University of Science and Technology Xi'an Campus: Shaanxi University of Science and Technology

**Min Liu**

Shaanxi University of Science and Technology Xi'an Campus: Shaanxi University of Science and Technology

**Chao Duan**

Shaanxi University of Science and Technology Xi'an Campus: Shaanxi University of Science and Technology

**Ruimin Zhang**

Shaanxi University of Science and Technology Xi'an Campus: Shaanxi University of Science and Technology

**Cailin Quan**

Shaanxi University of Science and Technology Xi'an Campus: Shaanxi University of Science and Technology

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## Original Research

**Keywords:** cellulose, fibrous adsorbent, solid/liquid separation, bio-adsorbent, dye-containing wastewater

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# 1 Cellulose Based Cation-Exchange Fiber as Filtration Material for the 2 Rapid Removal of Methylene Blue from Wastewater

3 Xinyi Shao<sup>a</sup>, Jian Wang<sup>a, b, \*</sup>, Zetan Liu<sup>a</sup>, Na Hu<sup>a</sup>, Min Liu<sup>a</sup>, Chao Duan<sup>a</sup>, Ruimin Zhang<sup>a</sup>, Cailin Quan<sup>a</sup>

4 a: College of Bioresources Chemical and Materials Engineering, Shaanxi University of Science & Technology, Xi'an  
5 710021, China

6 b: National Demonstration Center for Experimental Light Chemistry Engineering Education, Shaanxi University of  
7 Science & Technology, Xi'an 710021, China

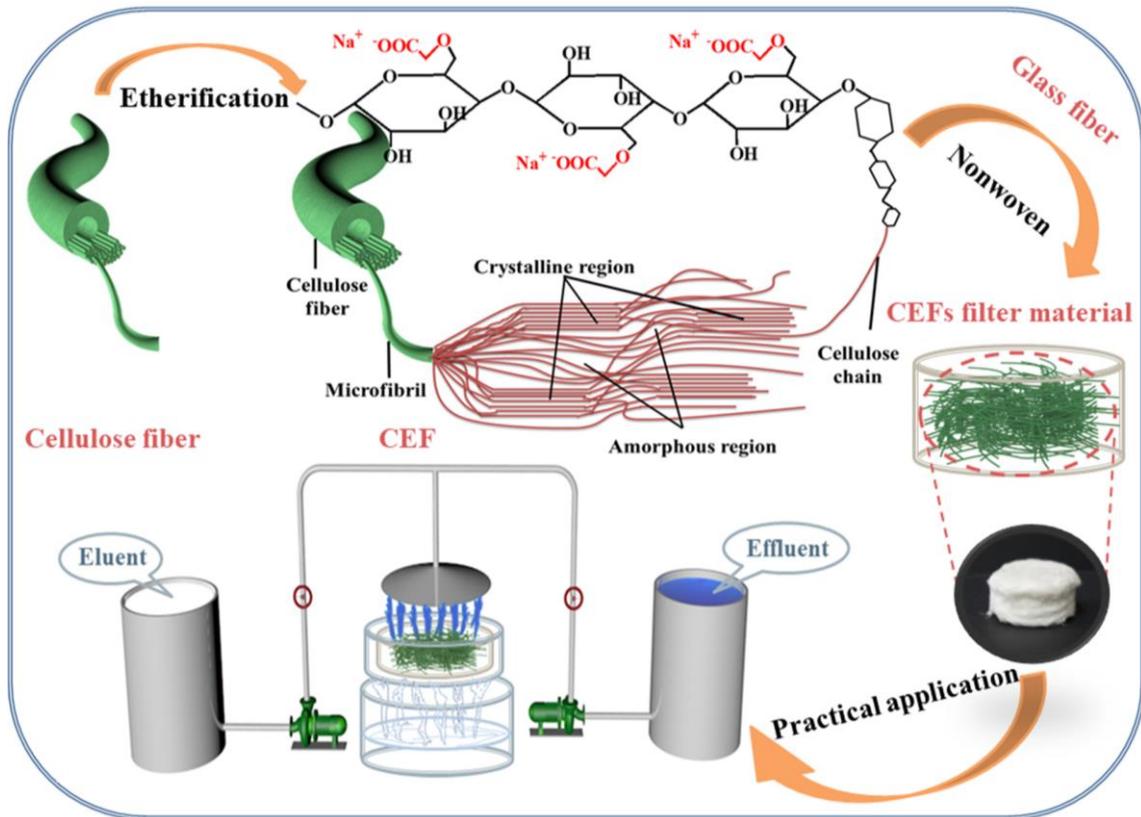
8 \*Corresponding author: Tel: +86-18229003848

9 E-mail: [zzwangjian@sust.edu.cn](mailto:zzwangjian@sust.edu.cn)

## 10 Abstract

11 The development of eco-friendly adsorbent with perfect solid/liquid separation performance is of  
12 great practical importance for the efficient purification of dye-containing wastewater. Herein, cellulose  
13 based cation-exchange fibrous bio-adsorbent (CEF) was successfully obtained via an industrialized  
14 cellulose etherification process. The fiber morphology can be maintained with the average degree  
15 substitution (DS) of CEF as about 0.19. The CEF showed high efficiency adsorption for methylene blue  
16 (MB): achieving 82% of equilibrium uptake ( $447.69 \text{ mg}\cdot\text{g}^{-1}$ ) within 5 min. Especially, the non-woven  
17 porous filter material, formed by CEFs and glass fibers, showed the praisable solid/liquid separation  
18 characteristics, fast water filtration rate ( $\approx 6.3 \text{ m}^3\cdot\text{m}^{-2}\cdot\text{h}^{-1}$ ) and high removal rate ( $\approx 99.2\%$ ) when it was  
19 used to adsorb MB (150 ppm) from wastewater. In addition, CEFs filter material can be regenerated via a  
20 simple method. This study demonstrates that green natural fiber materials are promising for the  
21 economical and efficient purification of dye-containing wastewater.

23 **Graphical abstract**



24

25 **Key words:** cellulose, fibrous adsorbent, solid/liquid separation, bio-adsorbent, dye-containing  
26 wastewater

27 **Introduction**

28 Water is the source of life, but unfortunately, water sources are threatened from contaminants and  
29 pollutants that are discharged into rivers due to industrialization (Jovan et al. 2019; Qi et al. 2019).  
30 Synthetic dyes, which are widely used in the textile, printing and plastic industries, are regarded as one of  
31 the most notorious pollutants on account of their high toxicity, high chroma and non-degradability (Huang  
32 et al. 2019). Thus, how to resolve dye pollution in wastewater has become an urgent issue. Various  
33 techniques have been applied for the wastewater treatment, including biologic treatment (Abiri et al., 2016;  
34 Baeta et al., 2015), photocatalytic degradation (Prabakaran and Pillay, 2019), chemical precipitation (Shi et al.,

35 [2015](#)), membrane separation (Neethu et al., [2018](#)) and adsorption (Jia et al., [2020](#)). Among these, adsorption  
36 has been regarded as one of the most effective and competitive method due to its low cost, easy  
37 separation and high efficiency. At present, many adsorbent materials have been investigated, including  
38 conventional materials such as activated carbons (Peláez-Cid et al. [2016](#)) and clay minerals (Li et al. [2018](#)),  
39 advanced materials such as metal-organic frameworks (MOFs) (Oveisi et al. [2017](#)), reduced graphene  
40 oxide/titanium dioxide nanotubes (rGO/TNT) (Nguyen and Juang, [2019](#)) and low-spin-state hematite (Shen  
41 et al. [2020](#)). Nevertheless, there is still need for further exploration on the adsorbents with sustainability,  
42 economy, nontoxicity and renewability.

43 Biomass raw materials are the most promising adsorbents to meet above requirements.  
44 Bio-adsorbents have been investigated to treat dye-containing wastewater, including forest and  
45 agricultural residues (Haque et al. [2020](#)), hemicellulose (Sun et al. [2015](#)), protein (Liu et al. [2016a](#)), lignin  
46 (Tang et al. [2014](#)), chitosan (Leon et al. [2018](#); Chen et al. [2020](#)), cellulose (Li et al. [2018](#)) etc. Cellulose,  
47 the most abundant biomass in nature, has received much attention in the development of these green  
48 bio-adsorbents (Chen et al. [2016](#)).

49 To this end, abundant specific functional groups in these structures are critical. For example, the  
50 cellulose modified with glycidyl methacrylate and sulfosalicylic acid leads to bio-adsorbent, with  
51 adsorption equilibrium for crystal violet of about  $135 \text{ mg}\cdot\text{g}^{-1}$  that is reached in 150 min (Zhou et al. [2014](#)).  
52 Amino-modified nanocellulose microgel achieved adsorption equilibrium of  $200 \text{ mg}\cdot\text{g}^{-1}$  for Congo red  
53 within 1 h (Xu et al. [2015](#)). Phosphate-modified cellulose powder reached the adsorption equilibrium of  
54  $46.95 \text{ mg}\cdot\text{g}^{-1}$  for rhodamine B (RhB) in 65 min (Silva et al. [2020](#)). Carboxymethyl cellulose-graphene  
55 oxide (CMC-rGA) aerogel showed a high absorption activity for RhB with  $186.33 \text{ mg}\cdot\text{g}^{-1}$  (Xiang et al.  
56 [2018](#)). The hydrogel from the mixtures of polyvinyl alcohol, CMC, rGA and bentonite reached the

57 adsorption equilibrium of 172.14 mg·g<sup>-1</sup> for MB within 200 min (Dai et al. 2017). The membrane  
58 fabricated using polyvinyl alcohol (PVA), CMC and ZSM-5 zeolite had an adsorption capacity of 7.83  
59 mg·g<sup>-1</sup> for MB with the contact time of 10 h (Sabarish et al. 2018).

60 However, most of cellulose based bio-adsorbents studied so far are in the form of aerogel, hydrogel,  
61 film or particle. Aerogel and hydrogel have a slow water filtration rate because the dye-containing  
62 wastewater needs to penetrate through multiple pore walls. The water filtration rate of membrane is fast,  
63 but its adsorption capacity is insufficient due to its small thickness. For particle, in order to have more  
64 adsorption sites, it is necessary to reduce the volume to increase the specific surface area, resulting in the  
65 difficulty of solid/liquid separation. The existing forms of cellulose based bio-adsorbents limit their  
66 practical application. Compared with them, fibrous bio-adsorbents have shown irreplaceable advantage  
67 owing to their perfect solid/liquid separation property, fast water filtration rate and superior wastewater  
68 treatment performance in terms of adsorption capacity and removal rate.

69 In this work, the cation-exchange fiber (CEF) based on cellulose were prepared via an  
70 industrialized cellulose etherification process using chloroacetic acid as the etherifying agent. The model  
71 cationic dye (MB) was employed to investigate the adsorption properties of CEF. Furthermore, a filter  
72 material expected to have perfect solid/liquid separation performance was formed by CEFs and glass  
73 fibers via non-woven way. The aim of this work is to provide a new strategy for using green and  
74 biodegradable natural fiber materials as promising bio-adsorbents for cost-effective and rapid purification  
75 of dye-containing wastewater.

## 76 **Experimental**

### 77 **Materials**

78 Softwood Kraft pulp fibers were obtained from the factory in Liao Cheng City, Shandong province,

79 China. Reagent grade sodium chloroacetate (MCA) (AR,  $\geq 98\%$ ), sodium hydroxide (AR,  $\geq 96\%$ ),  
80 hydrochloric acid (AR,  $\geq 36.0\%$ ), Methylene blue (Ind) and ethanol were received from Aladdin Reagent  
81 Co., Ltd (Shanghai, China). Glass fibers (length: 5 mm, width:  $26\mu\text{m}$ ) were obtained from Hengguang  
82 Co., Ltd (Shi Jia Zhuang, China). Deionized water was used throughout this work.

### 83 Synthesis of cellulose based cation-exchange fiber (CEF)

84 10 g softwood kraft pulp fibers, 15 g MCA and deionized water (20 mL) were added into beaker.  
85 The reaction was carried out at  $60^\circ\text{C}$  water bath for 2 h. Then the sample was immersed in the mixture  
86 solution (NaOH: 12.5 g, deionized water: 20 mL). The reaction was performed at  $80^\circ\text{C}$  for 8 h. The  
87 production was washed with ethanol to neutral, and finally air-dried.

### 88 Preparation of CEFs filter material

89 A porous filter material was formed by CEFs and glass fibers (weight ratio = 2:1) via non-woven  
90 way. The specific operation was that CEFs and glass fibers were fully dispersed in water, then filtered to  
91 remove excess water and dried in the oven at  $60^\circ\text{C}$ .

### 92 Characterization

93 Fourier Transform Infrared spectroscopy (FTIR, VERTEX 70, Bruker Optics Corporation; Germany)  
94 was carried out to analyze the functional group, the spectrum ranges from  $500$  to  $4000\text{ cm}^{-1}$ . The surface  
95 morphology of materials was observed by optical microscope (DMB5, Motic Co. Ltd., Xiamen, China).  
96 The energy dispersive spectrometer (EDS, VEGA 3 SBH) was used to analyze the elementary  
97 composition. The morphological properties of fibers were determined by the Fiber & Shive Analyzer  
98 (Morfi Compact FS-300, Techpap, France). Laser scanning confocal microscope (LSM, LSM800,  
99 Germany) was used to observe the cross-section of CEF. The adsorption capacity of CEF for MB was

100 measured by UV-vis spectrophotometer (UV-5000, Agilent, USA). The surface charges of fibers were  
101 measured by using a particle charge detector (PCD-03, Müttek, Germany). Zeta potential was determined  
102 by using a Zeta Nanosizer (SZTP06, Germany). The crystal structure of materials was analyzed by X-ray  
103 diffraction (XRD, D8 Advance, Bruker, Germany) at a scan rate of  $0.02^\circ \cdot \text{min}^{-1}$ . The crystallinity index  
104 (CrI) of the sample was calculated using the following equation (Liu et al. 2016b):

$$105 \quad CrI = (I_{002} - I_{am})/I_{002} \times 100 \quad (1)$$

106 Here  $I_{002}$  is the maximum diffraction intensity of crystalline from plane (002) at  $2\theta = 22.7^\circ$  for cellulose I,  
107 and  $2\theta = 21.7^\circ$  for cellulose II, while  $I_{am}$  is the intensity of amorphous cellulose at  $2\theta = 18^\circ$  for cellulose I,  
108 and  $2\theta = 16^\circ$  for cellulose II (Segal et al. 1959).

## 109 Carboxyl group content

110 The content of carboxyl group was determined by using standard method TAPPI T 237 with some  
111 appropriate modifications to suit the high carboxyl group content of CEF. Specifically, the amount of  
112 sodium bicarbonate-sodium chloride solution added was adjusted from 50 mL to 100 mL. The carboxyl  
113 content of the fiber was calculated according to the following modified equation:

$$114 \quad \text{Carboxyl content}(\text{mmol} \cdot 100\text{g}^{-1}) = \left[ B - \left( A + A \times \frac{c}{100} \right) \right] \times N \times \frac{400}{W} \quad (2)$$

115 where  $A$  (mL) is the volume of 0.010 N HCl consumed in titration of the pulp filtrate (25 mL);  $B$  (mL) is  
116 the volume of 0.010 N HCl consumed in titration of the sodium bicarbonate-sodium chloride solution (25  
117 mL);  $C$  (g) is the weight of water in pulp pad;  $N$  is the normality of HCl used in titration;  $W$  (g) is the  
118 weight of oven-dried test specimen;  $100$  (mL) is the amount of sodium bicarbonate-sodium chloride  
119 solution;  $400$  is derived as  $4 \times 100$ , where 4 is a factor to account for 25 mL aliquot taken for titration,  
120 and 100 is to express the result on 100 g of pulp.

121 Adsorption experiments

122 To study the adsorption capacity of CEF, a typical cationic dye MB was used. The maximum  
123 adsorption wavelength of MB was measured by UV-vis spectroscope. Subsequently, the  
124 absorbance-concentration (A-C) curve for MB was determined on the basis of the maximum absorption  
125 wavelengths of 664 nm, which was showed in Fig. 1. First of all, 20 mg CEF was added in 50 mL MB  
126 solution (initial concentration: 200 ppm) and stirred 30 min at room temperature. The pH value of MB  
127 was adjusted by 0.1 N HCl and NaOH. The residual concentration of the dye was detected using UV-vis  
128 spectrophotometer at 664 nm. The adsorption capacity of MB was calculated by

129 
$$q = \frac{(C_0 - C_t) \times V}{m} \quad (3)$$

130 Where  $q$  is the adsorption capacity of MB ( $\text{mg} \cdot \text{g}^{-1}$ );  $V$  is the solution volume (L);  $C_0$  is the MB initial  
131 concentration ( $\text{mg} \cdot \text{L}^{-1}$ );  $C_t$  is the MB concentration at time  $t$  ( $\text{mg} \cdot \text{L}^{-1}$ );  $m$  is the weight of adsorbent (g).

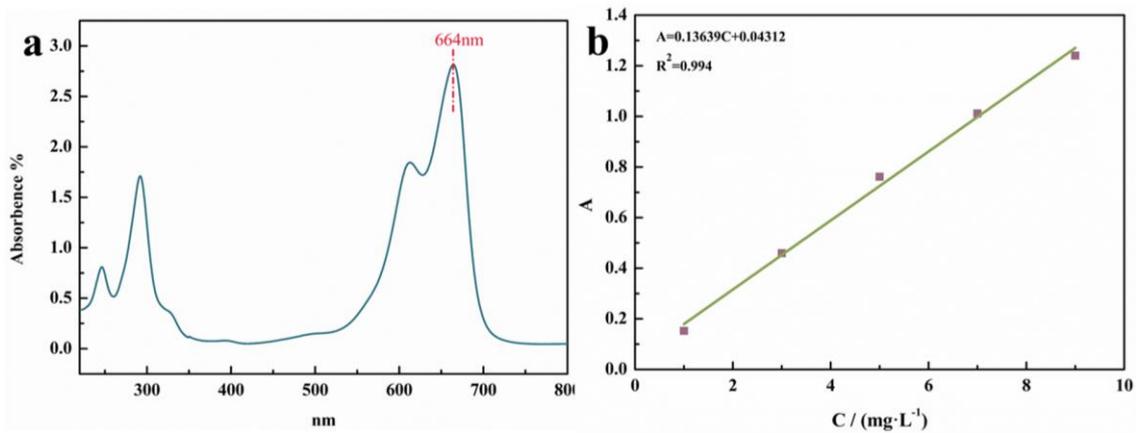


Fig. 1. (a) UV-Vis spectra of MB; (b) A-C curves of MB.

132 Adsorption kinetics

133 The adsorption kinetics could be calculated by pseudo-first-order (Eq. (4)) and pseudo-second-order  
134 models (Eq. (5)), shown as follows:

135 
$$\ln(q_e - q_t) = \ln q_e - K_1 t \quad (4)$$

$$\frac{t}{q_t} = \frac{1}{K_2 q_e^2} + \frac{1}{q_e} t \quad (5)$$

Where  $q_e$  and  $q_t$  are the amounts of dye adsorbed on the CEF at equilibrium and at time t, respectively;  $K_1$  and  $K_2$  are the rate constant of the pseudo-first-order and pseudo-second-order ( $\text{g}\cdot\text{mg}^{-1}\cdot\text{min}^{-1}$ ), respectively.

#### 140 *Adsorption isotherms*

141 The adsorption isotherm was detected by Langmuir (Eq. (6)) and Freundlich (Eq. (7)) models,  
142 respectively, the linear equations are presented below:

$$143 \frac{C_e}{q_e} = \frac{C_e}{q_m} + \frac{1}{q_m K_L} \quad (6)$$

$$144 \ln q_e = \frac{1}{n} \ln C_e + \ln K_F \quad (7)$$

145 Where  $q_e$  and  $q_m$  are the amounts of dye adsorbed on the CEF at equilibrium and the maximum saturated  
146 monolayer sorption capacity of adsorbent, respectively;  $K_L$  and  $K_F$  are the rate constant of the  
147 pseudo-first-order and pseudo-second-order, respectively.

#### 148 *Adsorption thermodynamics*

149 The thermodynamic factors could be obtained from temperature-dependent sorption isotherms. The  
150 values of  $\Delta H^0$  and  $\Delta S^0$  could be derived from the formula:

$$151 \Delta G^0 = -RT \ln K^0 \quad (8)$$

$$152 \ln K^0 = \frac{\Delta S^0}{R} - \frac{\Delta H^0}{RT} \quad (9)$$

153 Where  $R$  is ideal gas constant ( $8.314 \text{ J}\cdot\text{mol}^{-1}\cdot\text{K}^{-1}$ );  $T$  is the Kelvin temperature (K),  $\Delta G^0$  ( $\text{KJ}\cdot\text{mol}^{-1}$ ) is the  
154 standard Gibbs free energy change;  $K^0$  was the thermodynamic equilibrium constant;  $\Delta S^0$  ( $\text{KJ}\cdot\text{mol}^{-1}$ ) was  
155 the standard entropy change;  $\Delta H^0$  ( $\text{J}\cdot\text{mol}^{-1}\cdot\text{K}^{-1}$ ) is the standard enthalpy change.

#### 156 *Regeneration of CEFs filter material*

157 After the first adsorption, the used filter material was washed with HCl solution (0.1 N) at room

158 temperature to desorb the MB. Then it was washed with deionized water for several times until the pH  
159 was closed to natural, and dried for next test. This process was recycled four times. The regeneration  
160 efficiency (%RE) of adsorbent was calculated as follows:

$$161 \quad RE\% = \frac{q_r}{q_0} \times 100\% \quad (10)$$

162 Where  $q_0$  and  $q_r$  ( $\text{mg} \cdot \text{g}^{-1}$ ) are the adsorption capacities of the original adsorbent and regenerated adsorbent,  
163 respectively.

## 164 **Results and discussion**

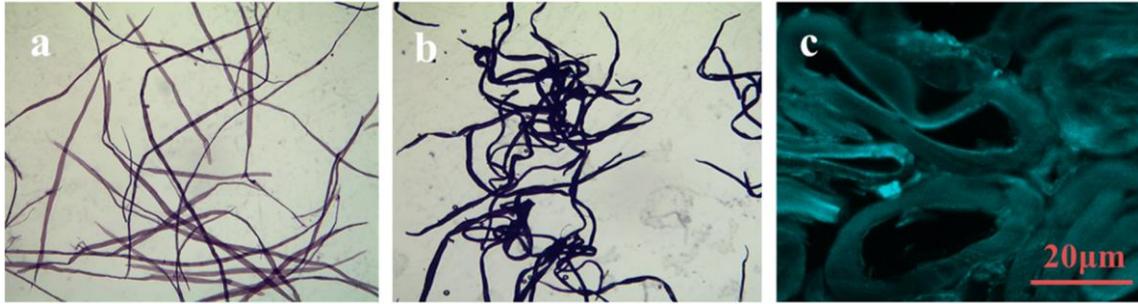
### 165 The morphological properties of fibers

166 As shown in Fig. 2a and b, CEF has the similar fibrous morphology as original cellulose fiber.  
167 Compared with cellulose fiber (Table 1), the average length of CEF was slightly decreased, which was  
168 due to the uneven reaction of fibers during etherification and the parts with violent reaction might be  
169 broken. In addition, the increasing of carboxyl content improves the hydrophilicity and water-swollen  
170 ability of the fiber, leading to a significant increase in the fiber width during the detection using water as  
171 medium. The cross-sectional morphology of CEF was observed by laser scanning confocal microscope  
172 (LSM). Fig. 2c clearly shows that CEF has a complete cell cavity, further confirming that the fibrous  
173 structure is still maintained after etherification.

174 **Table 1** Basic morphological characteristics of fibers before and after etherification.

Samples	Length/mm	Width/ $\mu\text{m}$
Cellulose fiber	$1.814 \pm 0.034$	$29.8 \pm 0.4$
CEF	$1.683 \pm 0.052$	$38.2 \pm 0.9$

175



**Fig. 2.** The optical micrographs of (a) cellulose fibers (×40) and (b) CEFs (×40). (c) The cross-sectional of CEFs (with fluorescent active labelling).

## 176 The carboxyl group content and zeta potential of fibers

177 It is well known that two phases coexist within the aggregation structure of cellulose is the reason  
178 why cellulosic biomass is insoluble in water. Table 2 showed that the total charges of fibers increased  
179 from 4.96 mmol/100g sample to 107.64 mmol/100g sample after carboxymethylation. According to Eq. (S1), the  
180 average degree of substitution (DS) of CEF is about 0.19, indicating that a small part of the hydroxyl  
181 groups on the cellulose molecular chains are substituted. It is speculated that CEF is still an aggregation  
182 structure with coexistence of two phases, which is the reason why CEF can maintain fibrous structure.  
183 The low DS makes the functionalized cellulose fiber still retains a large number of crystalline regions of  
184 original cellulose.

185 The introduction of carboxyl groups makes CEFs are strongly negatively charged on their surfaces,  
186 and the zeta potential is -38.3 mV, which is compared to -17.8 mV for the original cellulose fibers. The  
187 increased negative charges on CEFs surfaces lead to stronger electrostatic interactions with cationic  
188 substance.

189

190

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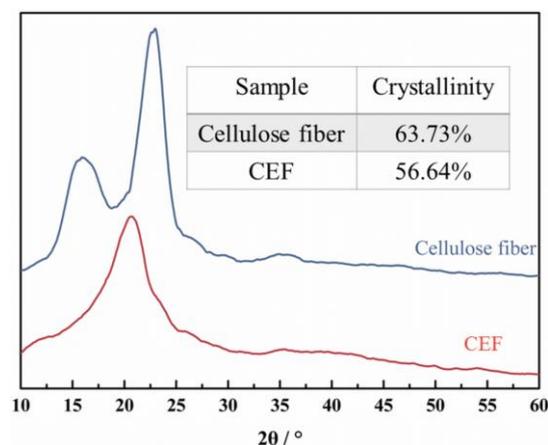
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**Table 2** The carboxyl group content and zeta potential value of fiber before and after etherification.

Samples	Carboxyl group content (the total charges)/ mmol /100g sample	Zeta potential value/mV
Cellulose fiber	4.96	-17.8
CEF	107.64	-38.3

## 193 XRD analysis

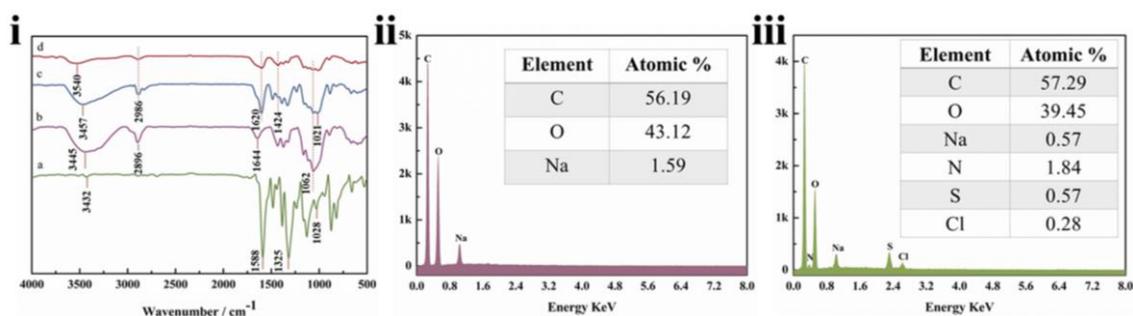
194 The destruction of Cellulose I was confirmed from X-ray diffraction (XRD) patterns (Fig. 3). The  
 195 peaks at  $2\theta = 15^\circ$  and  $22.5^\circ$  correspond to the crystal planes of (101) and (002) respectively, which are the  
 196 characteristic peaks of Cellulose I. After etherification, the presence of a new diffraction peak at  $2\theta = 20^\circ$ ,  
 197 suggesting that the transformation of Cellulose I to Cellulose II under the strong alkaline conditions  
 198 (Chen et al. 2018). Cellulose fiber has a crystallinity of about 63.73%, when it is etherified to CEF, the  
 199 crystallinity is reduced to about 56.64%. This result is in consistent with the above speculation, which is  
 200 that CEF still has a two-phase coexisting aggregation structure in the case of considerable carboxyl group  
 201 content (107.64 mmol/100g sample). Therefore, CEF can still maintain a stable fibrous structure when in  
 202 contact with water. Meanwhile, the reduction of crystalline phase enhances the accessibility of carboxyl  
 203 groups in the cell wall, facilitating the effective adsorption.

**Fig. 3.** XRD spectra of cellulose fiber and CEF.

204 Adsorption features of CEF

205 *FTIR analysis and EDS analysis*

206 The successful load of MB on CEF is confirmed by FTIR and EDS results. FTIR experimental  
207 results show that no covalent bonds are formed between CEF and MB, indicating that electrostatic  
208 interactions might be the main force. As shown in Fig. 4id, the characteristic signals of CEF at 1620 cm<sup>-1</sup>  
209 and 1424 cm<sup>-1</sup> correspond to the stretching and bending vibration of COO<sup>-</sup> group, respectively (Liu et al.  
210 2015). For the MB FTIR spectrum (Fig. 4ia), the peak at 1588 cm<sup>-1</sup> is attributed to C=N stretching on the  
211 benzene ring, the peaks at 1325 cm<sup>-1</sup> and 1028 cm<sup>-1</sup> are due to the C-N and C-S stretching of the benzene  
212 ring.<sup>3</sup> All these MB characteristics are present in Fig. 4ic (MB loaded CEF sample), confirming the  
213 successful adsorption of MB onto CEF. For the CEF sample, EDS results show no nitrogen, sulfur or  
214 chlorine are present (Fig. 4ii), while for the MB loaded CEF sample, these elements are evident and the  
215 sodium amount decreases (Fig. 4iii), indicating the ion exchange between CEF and MB.



**Fig. 4.** (i) FTIR spectra of (a) MB; (b) cellulose fiber; (c) MB loaded CEF; (d) CEF. EDS analyses of (ii) CEF and (iii) MB loaded CEF.

216 *Effect of pH value*

217 Generally, with varying levels of environmental pH, adsorbents exhibit different adsorption  
218 capabilities, because the pH influences the charge transfer at liquid-solid interface (Rethinasabapathy et al.  
219 2018). The same is true for CEF (Fig. 5), with the increasing of pH value, the adsorption capacity

220 gradually increases, and the rising trend begins to slow down at pH 7.0. This is because the adsorption of  
 221 CEF for MB mainly relies on the strong electrostatic interactions between carboxyl group and cationic  
 222 MB for ion exchange. The lower pH condition causes protonation of carboxyl group, leading to  
 223 electrostatic repulsion with MB. With the increasing of alkalinity, electrostatic interactions between  
 224 carboxyl group and cationic MB enhances, resulting in the improvement of adsorption capacity.

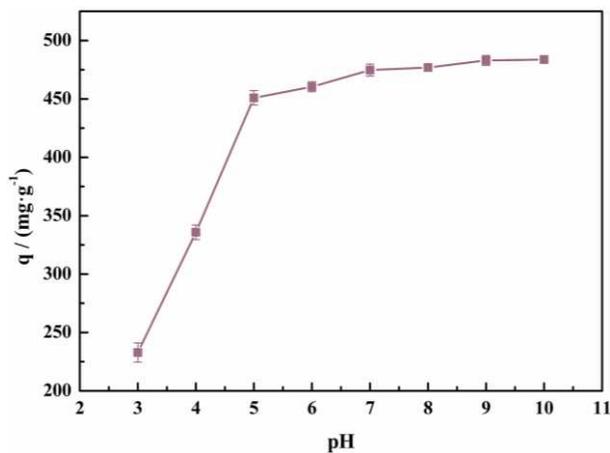


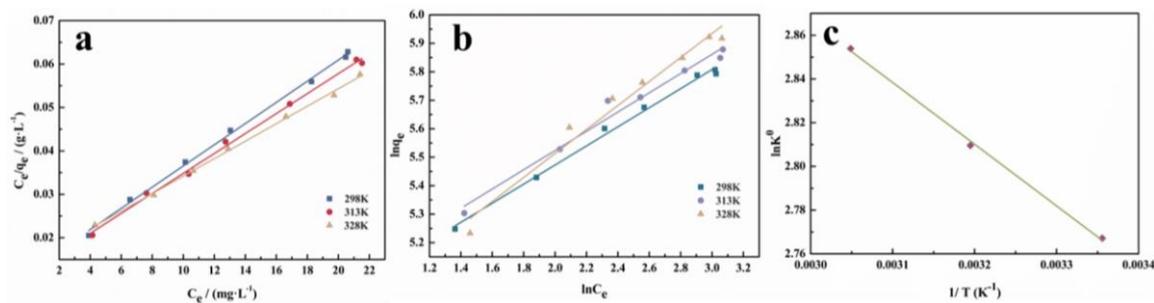
Fig. 5. The pH value effect on adsorption of CEF for MB.

### 225 *Adsorption isotherms and thermodynamics*

226 The adsorption isotherms of CEF for MB at three different temperatures (298, 313, 328 K) were  
 227 determined at pH 7.0, and the data were fitted by Langmuir model and Freundlich model. It obviously  
 228 observed that the results are a better match with monolayer adsorption (Langmuir model) (Fig. 6a and b  
 229 and Table 3). The higher the temperature, the higher the adsorption capacity. At 328 K, the saturation  
 230 capacity is 497.51 mg·g<sup>-1</sup>.

231 **Table 3** The isotherm parameters of the adsorption MB by CEF.

T(K)	Langmuir model			Freundlich model		
	q <sub>m</sub> (mg·g <sup>-1</sup> )	K <sub>L</sub> (L·mg <sup>-1</sup> )	R <sup>2</sup>	K <sub>F</sub> (L·mg <sup>-1</sup> )	n	R <sup>2</sup>
298	409.84	0.2001	0.999	121.79	2.9830	0.993
313	434.78	0.1944	0.998	127.46	2.9595	0.974
328	497.51	0.1428	0.997	106.80	2.3745	0.968



**Fig. 6.** (a) The Langmuir isotherm and (b) the Freundlich isotherm for the adsorption of MB. (c) The linear plots of thermodynamic for the adsorption of MB.

232 The thermodynamic data (Fig. 6c and Table 4) show a positive  $\Delta H^0$  and negative  $\Delta G^0$ , indicating that  
 233 the MB adsorption process is an endothermic and spontaneous process. This result further confirms that  
 234 high temperature is beneficial to the MB adsorption onto CEF.

235 **Table 4** Thermodynamic parameters at different temperatures.

T(K)	$\Delta G^0$ (KJ·mol <sup>-1</sup> )	$\Delta H^0$ (KJ·mol <sup>-1</sup> )	$\Delta S^0$ (J·mol <sup>-1</sup> ·K <sup>-1</sup> )
298	-6.86		
313	-7.31	2.35	30.87
328	-7.78		

### 236 *Adsorption kinetics*

237 The MB adsorption/removal is shown in Fig. 7a (200 ppm MB, 20 mg CEF, pH 7.0), the dye  
 238 removal reached a plateau value (447.69 mg·g<sup>-1</sup>) quickly. The theoretical adsorption capacity (i.e.,  
 239 electrostatic adsorption capacity) calculated from the carboxyl group content in CEF (107.64 mmol/100g  
 240 sample) is 344.29 mg·g<sup>-1</sup> (Eq. (S2)). Other factors, such as surface adsorption, van der Waals forces and/or  
 241 hydrogen bonding (Liu et al. 2015), also contribute to the MB adsorption onto CEF (Fig. 7e). Notably, the  
 242 adsorption capacity of CEF rapidly reached 113.13 mg·g<sup>-1</sup> ( $\approx$  25 % of its equilibrium uptake), 312.15  
 243 mg·g<sup>-1</sup> ( $\approx$  70 % of its equilibrium uptake) and 367.86 mg·g<sup>-1</sup> ( $\approx$  82 % of its equilibrium uptake) in a very

244 short period of just 1, 4 and 5 min, respectively.

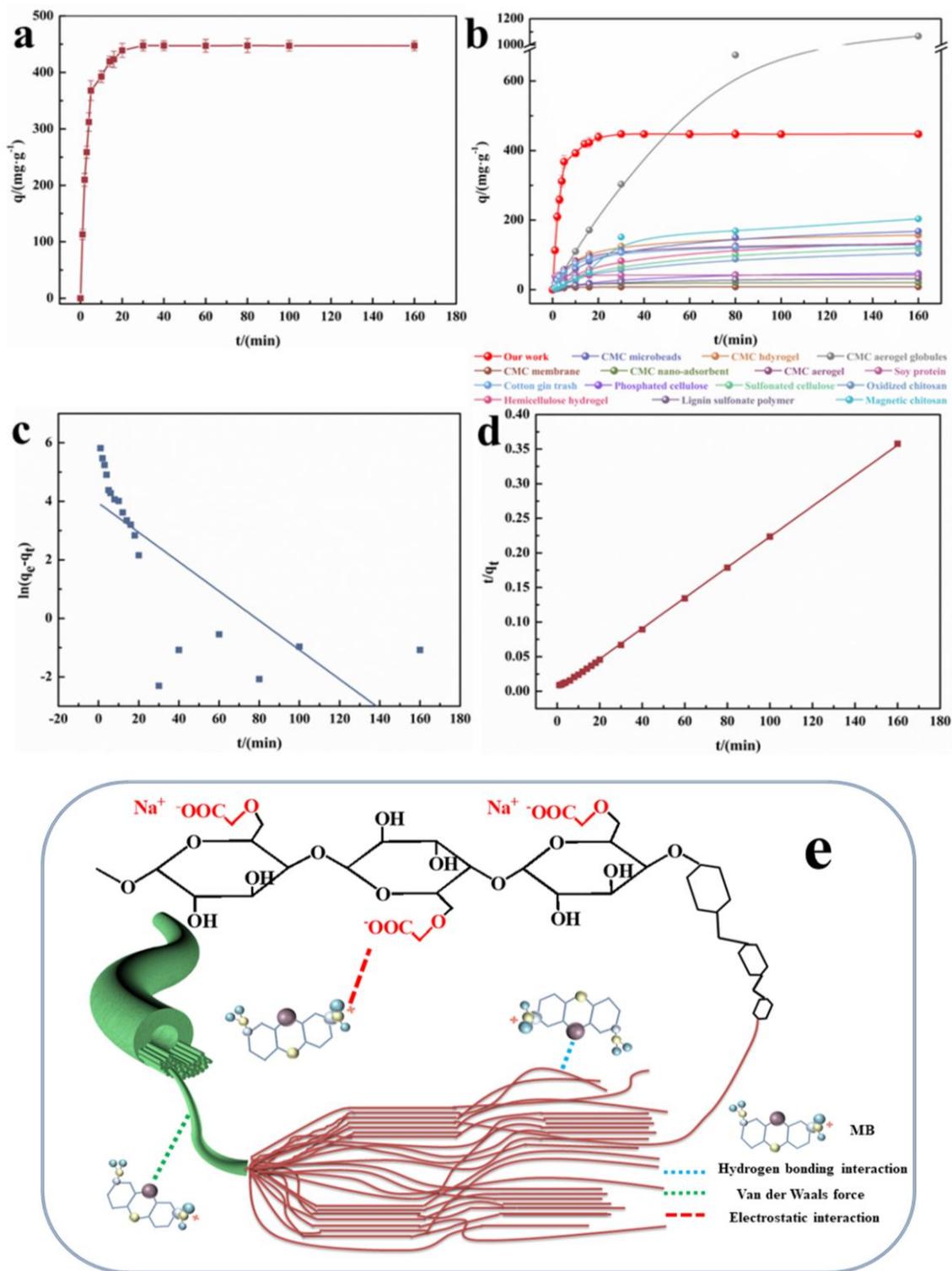


Fig. 7. (a) The adsorption capacity versus contact time for MB adsorption onto CEF. (b) Comparison of adsorption

capacity versus contact time of CEF, with other cationic dye bio-adsorbents, the model dyes used are listed in Table

S1. (c) The pseudo-first-order kinetic and (d) the pseudo-second-order kinetic plots of MB by CEF. (e) Schematic representation of the interactions between CEF and MB.

245 Shown in Fig. 7b are the comparisons of the adsorption capacity vs time plotting of the present study  
246 and those reported in the literature for cationic dye bio-adsorbents. One striking feature is that the  
247 as-prepared CEF sample has the fastest adsorption rate among all the samples, making it an excellent  
248 adsorbent for highly efficient MB removal from wastewater.

249 Two kinetic models, pseudo-first order kinetics and pseudo-second order kinetics were used to  
250 analyze the MB adsorption rates of CEF. The kinetic parameters were obtained by linear regression of the  
251 plots and summarized in Table 5. The theoretical equilibrium adsorption amount from pseudo-second  
252 order kinetics ( $452.49 \text{ mg}\cdot\text{g}^{-1}$ ) is approximate the actual adsorption amount ( $447.69 \text{ mg}\cdot\text{g}^{-1}$ ). Meanwhile,  
253 the value of  $R^2$  (0.9996) for pseudo-second order kinetics is much higher than that of pseudo-first order  
254 kinetics ( $R^2 = 0.5481$ ). The results indicate that the adsorption process of MB follows the pseudo-second  
255 order kinetics (Fig. 7c and d).

256 **Table 5** Kinetics parameters for adsorption MB of CEF.

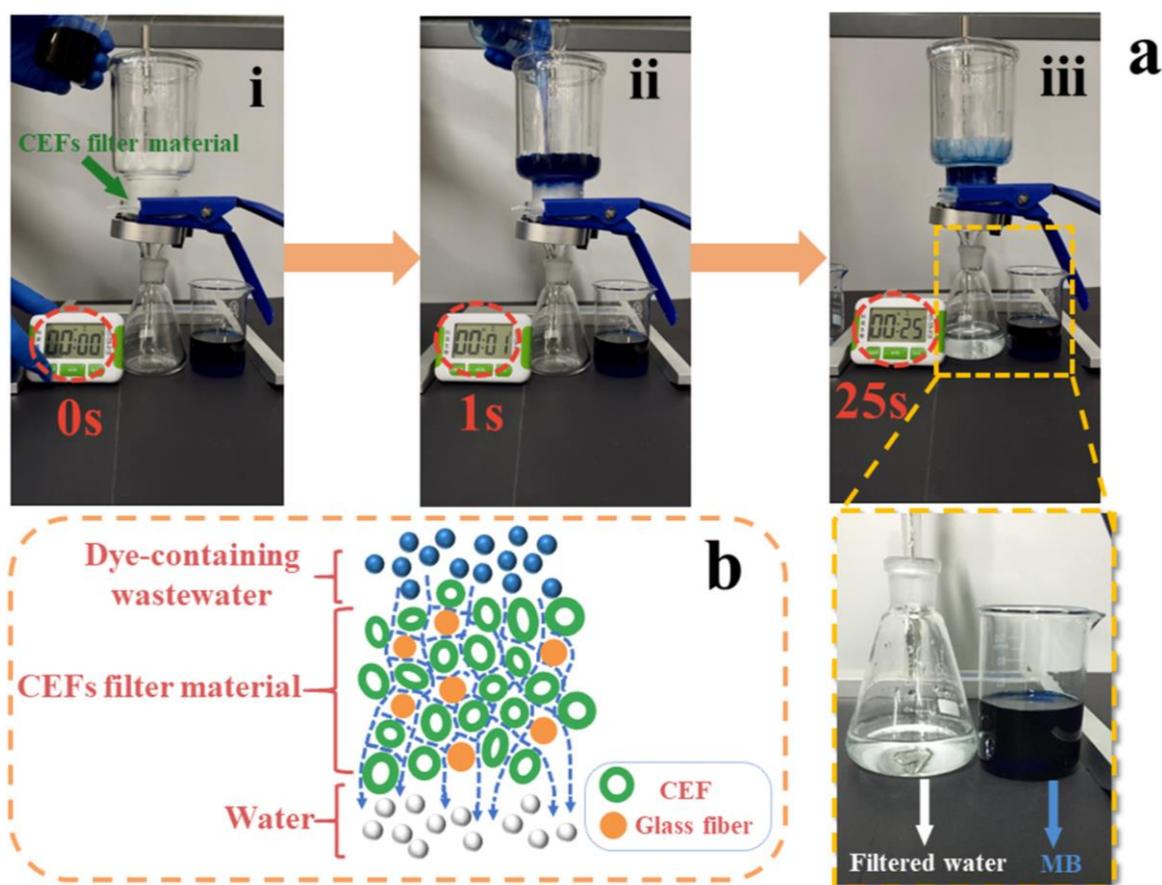
Pseudo-first order kinetics			Pseudo-second order kinetics		
$K_1(\text{g}\cdot\text{mg}^{-1}\cdot\text{min}^{-1})$	$q_e(\text{mg}\cdot\text{g}^{-1})$	$R^2$	$K_2(\text{g}\cdot\text{mg}^{-1}\cdot\text{min}^{-1})$	$q_e(\text{mg}\cdot\text{g}^{-1})$	$R^2$
0.05014	51.38	0.5481	0.00156	452.49	0.9996

### 257 *Construction of porous filter material*

258 A device with perfect solid/liquid separation performance was designed as shown in Fig. 8a and  
259 video. Here, a non-woven porous filter material is formed by CEFs and glass fibers, and the constructed  
260 multi-channel structure similar to polymer-based ion-exchange resins prepared by porogen (Vorotyntsev  
261 et al. 2018) effectively reducing the hindrance to fluid flow and increasing the water filtration rate (Fig.

262 8b).

263 The superior water filtration capability of adsorbent is critical, which means whether a sufficient  
264 amount of dye-containing wastewater can be treated to satisfy the practical need. In this study, water  
265 filtration capacity of CEFs filter material was evaluated at the MB concentration of 150 ppm and volume  
266 of 50 mL. As shown in the video and Fig. 8a i and ii, clean filtered water is immediately produced as the  
267 dye is poured. The absorbance of filtered water at 664nm, the characteristic absorption peak of MB, is  
268 0.21. The dye removal rate reached 99.2%. More importantly, CEFs filter material basically completed  
269 the purification process within 25 s (Fig. 8a iii). Therefore, the filtration rate of the CEFs filter material  
270 with a thickness of 17.5 mm is up to  $6.3 \text{ m}^3 \cdot \text{m}^{-2} \cdot \text{h}^{-1}$ .



**Fig. 8.** (a) A rapid removal of MB from the wastewater by CEFs filter material within 25 s. (b) Cross-sectional representation of the multi-channel CEFs non-woven materials to filter dye-containing wastewater.

271 The brilliant adsorption performance of CEFs filter material is inseparable from the fibrous  
272 structure, abundant carboxyl groups and low crystallinity of CEF. The perfect solid/liquid separation  
273 property, fast water filtration rate and excellent adsorption performance endow CEFs filter material the  
274 ability to efficiently purify dye-containing wastewater.

### 275 *Adsorption ability and reusability*

276 Adsorbent reuse/recycling is also of practical interest. The regeneration of dye-saturated CEFs filter  
277 material can be conducted by following a simple washing with 0.1 N HCl. As shown in Fig. 9, CEFs filter  
278 material still remains at about 90% of the original adsorption capacity after four cycles, indicating an  
279 excellent stability and reusability, which is a promising material for removal of dye from wastewater in  
280 practical application.

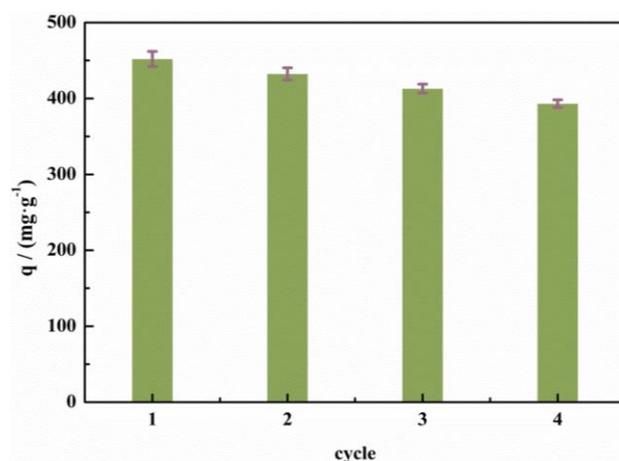


Fig. 9. Adsorption ability and reusability of CEFs filter material.

## 281 **Conclusion**

282 In summary, we synthesized a cation-exchange fibrous bio-adsorbent with desirable solid/liquid  
283 separation property using natural cellulose fibers as raw materials through a cellulose etherification  
284 process that controlling the average DS of 0.19, and prepared a filter material that similar to ion-exchange  
285 resins by non-woven way. CEF showed highly efficient adsorption of MB: achieving 82 % of its

286 equilibrium uptake in just 5 min. More importantly, the CEFs filter material with perfect solid/liquid  
287 separation performance has a fast water filtration rate ( $6.3 \text{ m}^3 \cdot \text{m}^{-2} \cdot \text{h}^{-1}$ ) and is easy to regenerate. The  
288 above advantages, together with the sustainability and biodegradability demonstrate the promise of using  
289 CEF as high-performance adsorbent for dye-containing wastewater treatment applications.

## 290 **Acknowledgments**

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## 292 **Conflict of interest**

293 The authors declared no potential conflicts of interest with respect to the research, authorship, and/or  
294 publication of this article.

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# Figures

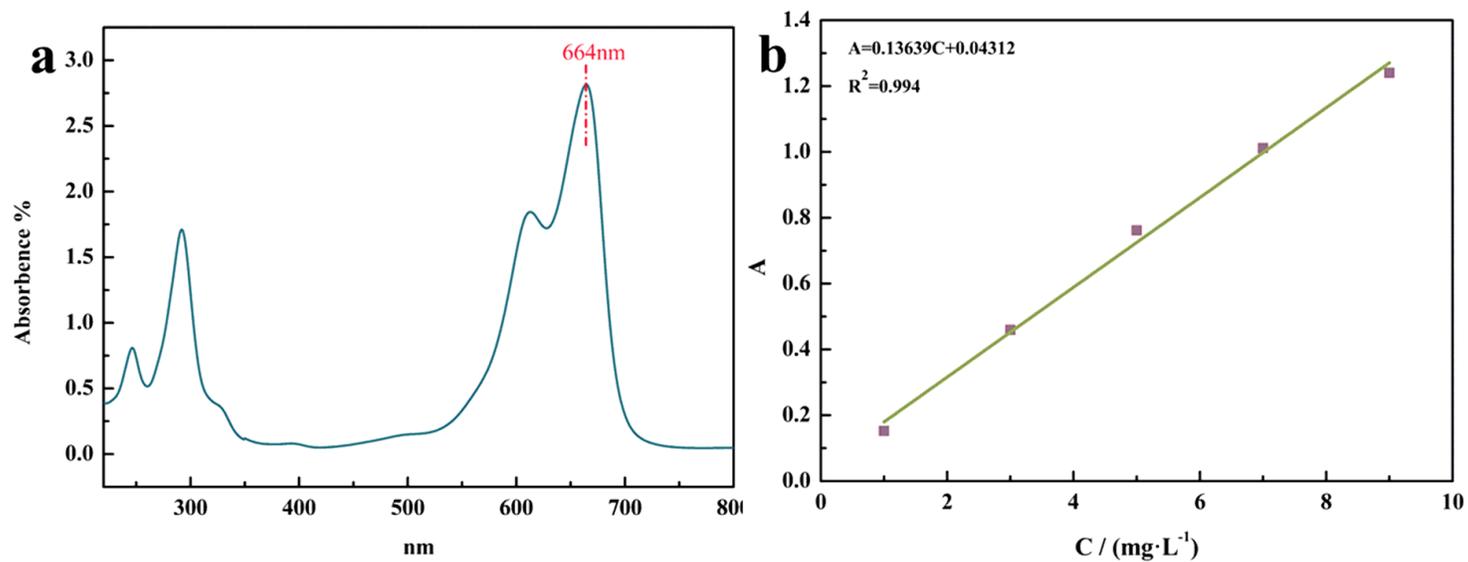


Figure 1

(a) UV-Vis spectra of MB; (b) A-C curves of MB.

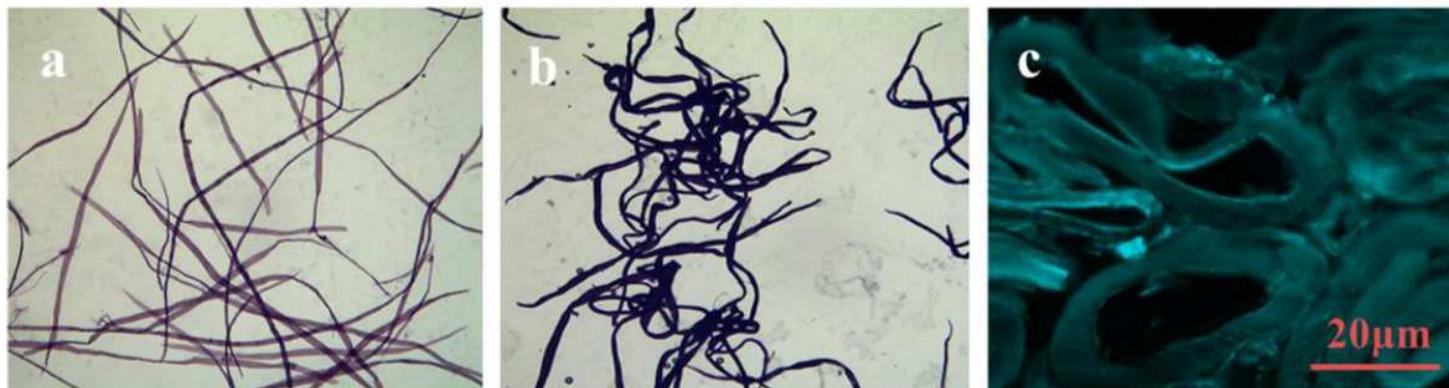


Figure 2

The optical micrographs of (a) cellulose fibers ( $\times 40$ ) and (b) CEFs ( $\times 40$ ). (c) The cross-sectional of CEFs (with fluorescent active labelling).

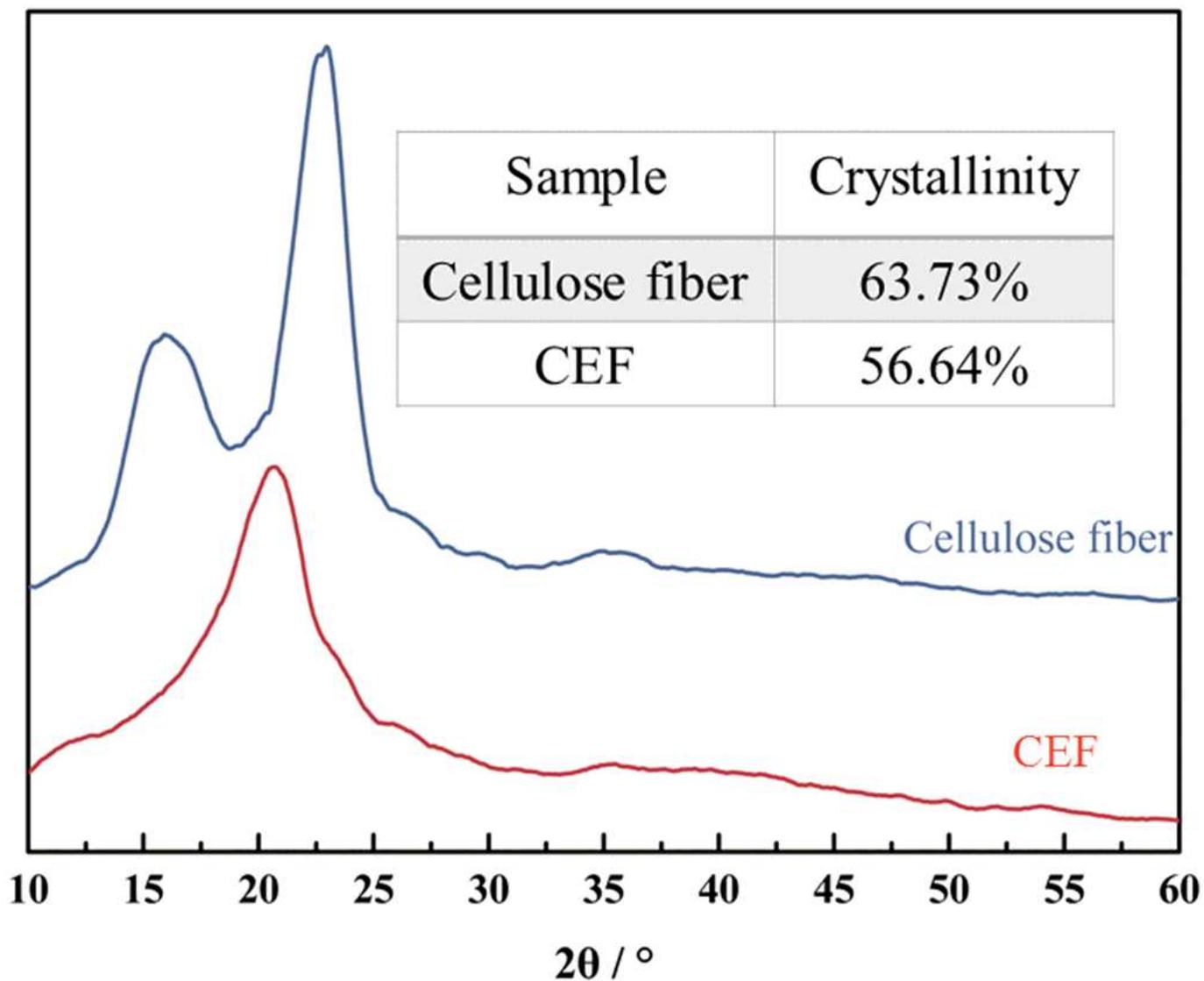
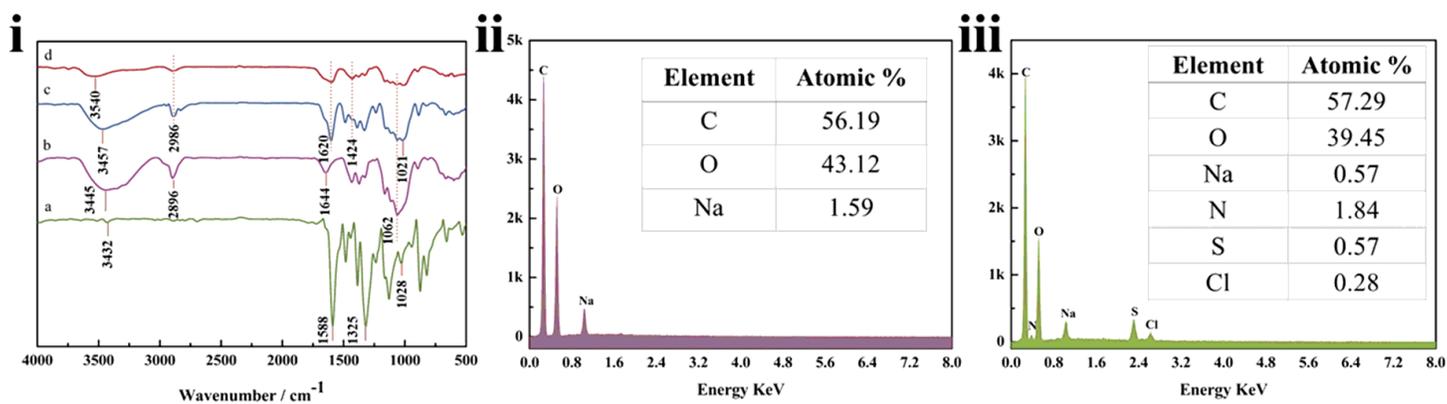


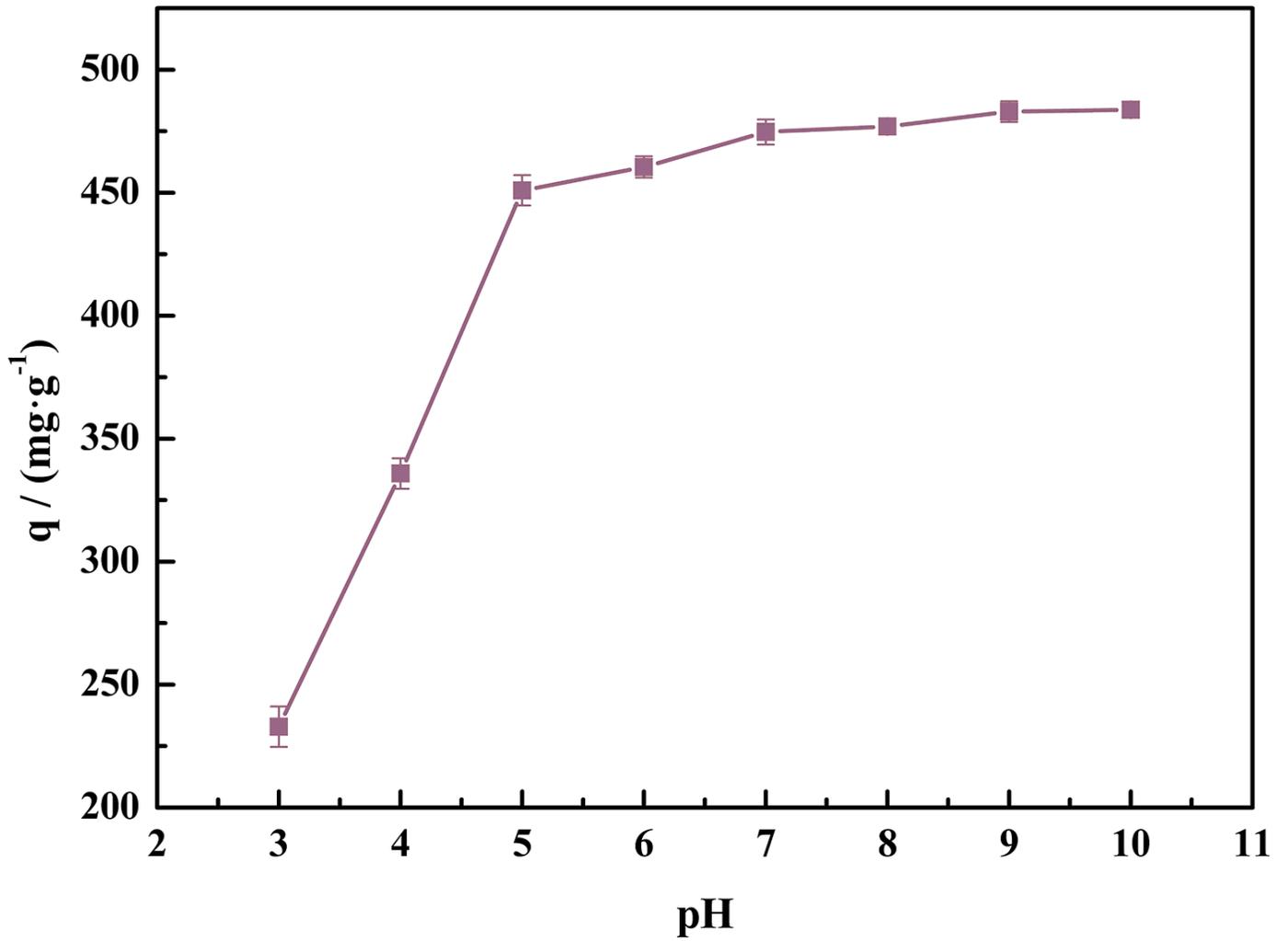
Figure 3

XRD spectra of cellulose fiber and CEF.



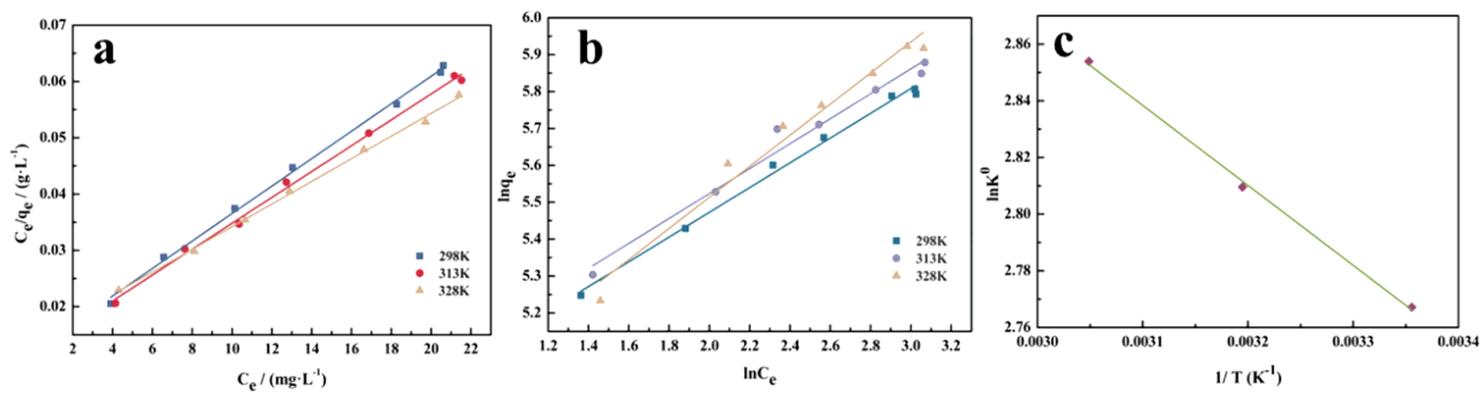
**Figure 4**

(i) FTIR spectra of (a) MB; (b) cellulose fiber; (c) MB loaded CEF; (d) CEF. EDS analyses of (ii) CEF and (iii) MB loaded CEF.



**Figure 5**

The pH value effect on adsorption of CEF for MB.



**Figure 6**

(a) The Langmuir isotherm and (b) the Freundlich isotherm for the adsorption of MB. (c) The linear plots of thermodynamic for the adsorption of MB.



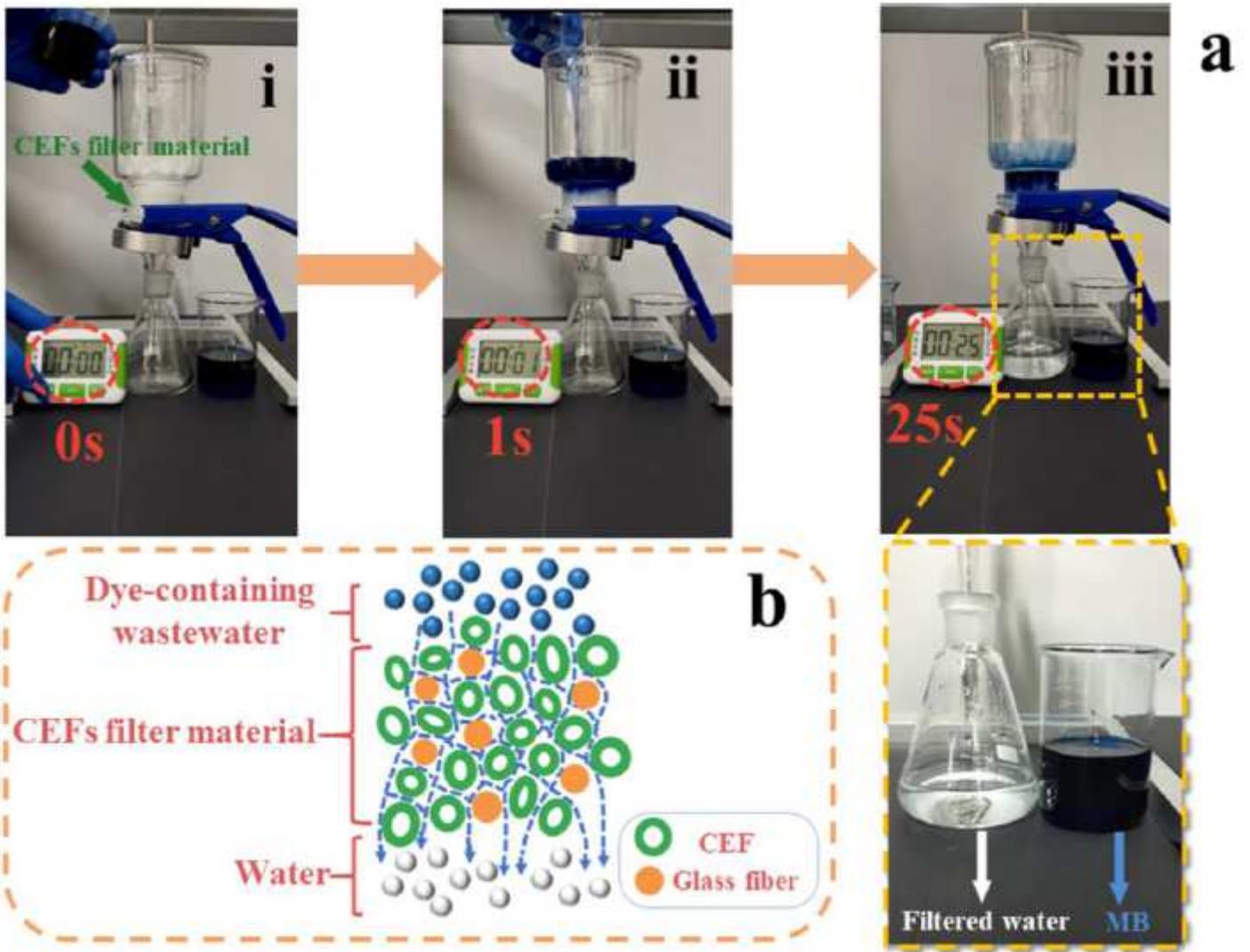


Figure 8

(a) A rapid removal of MB from the wastewater by CEFs filter material within 25 s. (b) Cross-sectional representation of the multi-channel CEFs non-woven materials to filter dye-containing wastewater.

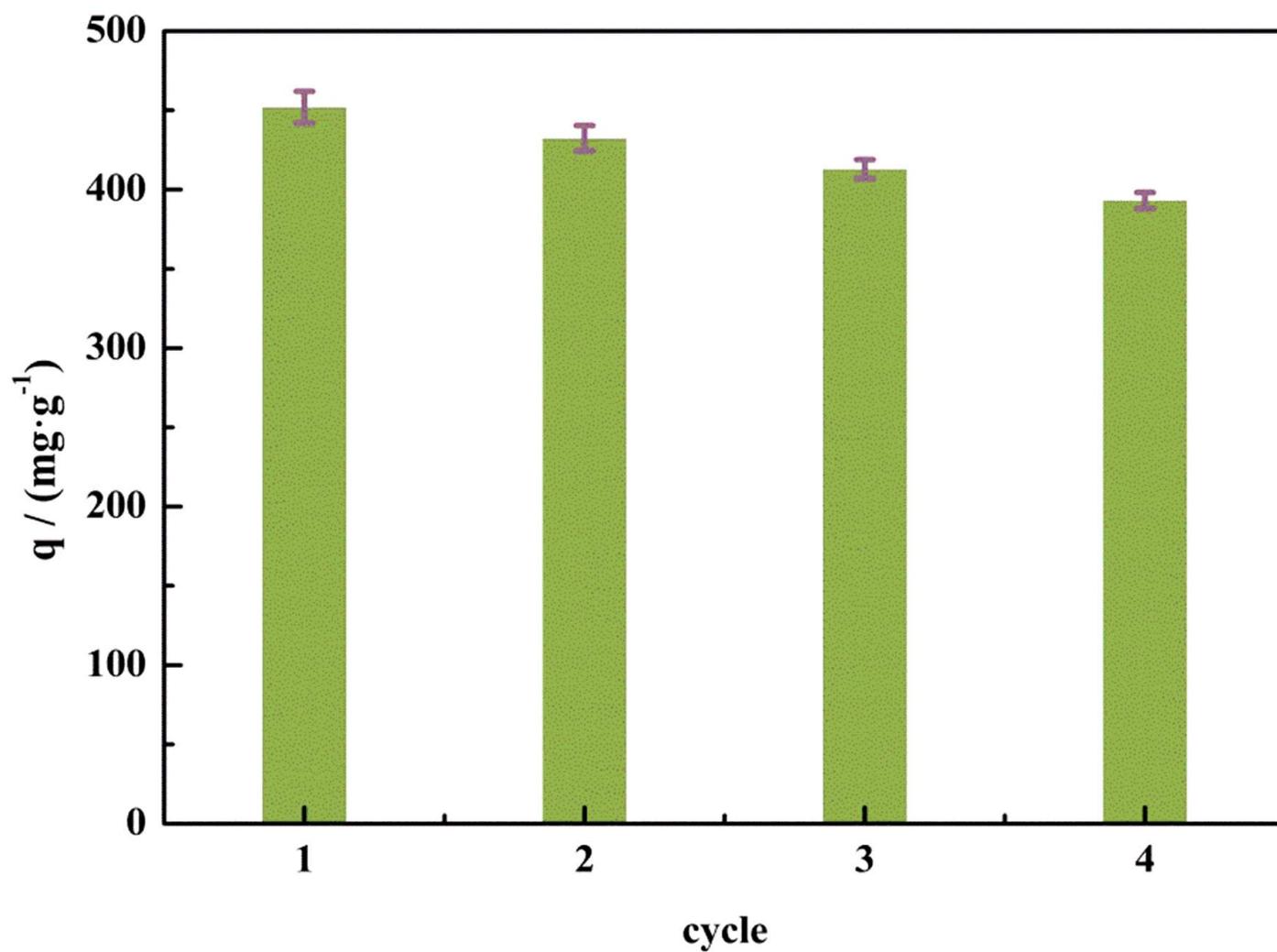


Figure 9

Adsorption ability and reusability of CEFs filter material.

## Supplementary Files

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