

Impact of Ciprofloxacin With Autophagy on Acute Kidney Injury

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Abstract

Renal tubular injury caused by oxidative stress and inflammation results in acute kidney injury. Recent research reported that antibiotics may restore deteriorated renal tubules, but the underlying mechanism remains unclear. Therefore, we investigated the efficacy and mechanism of action of antibiotics against renal tubular injury. We screened ciprofloxacin, ceftizoxime, minocycline, and netilmicin and selected ciprofloxacin to examine further because of its low toxicity towards renal tubular cells. We evaluated the effect of ciprofloxacin on cell survival by analyzing apoptosis and autophagy. TUNEL assay results showed that the ciprofloxacin group had less apoptotic cells than the control group. The ratio of cleaved caspase 3 to caspase 3, the final effector in the apoptosis process, was decreased, but the ratio of Bax to Bcl-2 located upstream of caspase 3 was not decreased in the ciprofloxacin group. Therefore, apoptosis inhibition does not occur via Bax/Bcl-2. Conversely, the levels of phosphorylated Bcl-2, and Beclin-1, an autophagy marker, were increased, and that of caspase-3 was decreased in the ciprofloxacin group. This indicates that ciprofloxacin enhanced autophagy, increasing the amount of free Beclin-1 via phosphorylated Bcl-2, and inhibited caspase activity. Therefore, ciprofloxacin might increase renal cell viability through the autophagy activation in acute kidney injury.

Introduction

Studies on the homeostasis of mammalian cells have been actively conducted for a long time, and it is well known that autophagy, apoptosis, and necrosis are the most important mechanisms of programmed cell death in cells, and the fate of cells is decided by the coordinated action of this programmed cell death¹. Among them, apoptosis has been considered the important mechanism of programmed cell death in mammalian cells, but recent studies have reported that autophagy plays a dual role in both the survival and death mechanisms, according to the cell type and degree of stimulation². Cell homeostasis can be controlled by the interaction between autophagy and apoptosis at the molecular level³. In particular, autophagy and apoptosis play important roles in the homeostasis of kidney cells, such as podocytes and tubular epithelial cells, in hypoxia, nutrient deprivation, oxidative stress, and inflammation conditions^{4,5}. In other words, renal tubular injury caused by several injuries results in acute kidney injury (AKI), and the cells damaged by AKI recover or die based on the relationship between autophagy and apoptosis.

Recent research has reported that antibiotics may influence deteriorated kidney cells or tissues, although antibiotics have their own action mechanisms against bacteria, such as inhibition of replication and differentiation⁶. For example, tigecycline, which is a new broad-spectrum antibiotic drug that structurally and functionally resembles tetracycline, has been reported to have anti-cancer effects by activating the AMP-activated protein kinase/mammalian target of rapamycin (AMPK/mTOR) pathway, but not apoptosis, in human gastric cancer cells⁷. Minocycline, a protein synthesis inhibitor (30S inhibitor), and a representative tetracycline antibiotic, has been reported to be associated with apoptosis in acute kidney

injury and chronic inflammation⁸. In other words, antibiotics may restore renal tubules, but the underlying mechanism remains unclear.

Therefore, we aimed to investigate the effect of antibiotics on the survival of kidney cells, and the mechanism of action of antibiotics on cell viability associated with apoptosis and autophagy.

Results

Screening of the ideal concentration of antibiotics that assures MDCK cell viability in normoxia and hypoxia conditions

Ciprofloxacin, ceftizoxime, minocycline, and netilmicin were used for the screening of the concentration of antibiotics that assures the cell viability of Madin-Darby Canine Kidney (MDCK) cells after 48 hours of treatment using a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) assay. Under normoxia (21% O₂) conditions, ciprofloxacin showed a consistent increase in cell viability since the beginning of ciprofloxacin induction, compared with the control group, especially at a concentration higher than 100 µg/mL ($P < 0.001$). However, cell viability was mildly increased with the increase in the concentration (50 to 100 µg/mL) of ciprofloxacin, in comparison with the control group under hypoxia conditions (1% O₂) (Fig. 1A). This result indicates that ciprofloxacin can improve cell viability under both normoxia and hypoxia conditions. The maximal efficacies (cell viability of ciprofloxacin vs. control) of ciprofloxacin were 42.6% at 500 µg/mL under normoxia conditions, and 10.1% at 100 µg/mL under hypoxia conditions. Under normoxia conditions, ceftizoxime also showed an increase in cell viability after the introduction of ceftizoxime at a high concentration (500 to 1,000 µg/mL). However, there were no changes in cell viability with increasing concentrations of ceftizoxime under hypoxia conditions (Fig. 1B). Under normoxia and hypoxia conditions, minocycline (10 and 50 µg/mL) resulted in an increase in cell viability. However, cell viability rapidly decreased when the concentration of minocycline surpassed 50 µg/mL in normoxia and hypoxia conditions (Fig. 1C). Under normoxia and hypoxia conditions, the concentration of netilmicin was not correlated with cell viability (Fig. 1D).

Under normoxia conditions, ciprofloxacin showed the highest efficacy (30% – 45%) in comparison with other antibiotics. Minocycline had a 20% efficacy at a lower concentration, but the toxic effect increased as the concentration of minocycline was increased (Table 1). Based on these results, we chose ciprofloxacin to examine further, an antibiotic with a similar efficacy but a lower toxicity compared to other antibiotics.

Table 1
Efficacies of antibiotics on cell viability between normoxia and hypoxia

Spectrum	Name	Efficacy			
		Normoxia (21% O ₂)		Hypoxia (1% O ₂)	
		Con. (µg/ml)	Efficacy (%)	Con. (µg/ml)	Efficacy (%)
Quinolones	Ciprofloxacin	50	10.00 ± 2.36	50	9.27 ± 2.26**
		100	33.49 ± 1.22***	100	10.07 ± 2.91**
		150	38.87 ± 3.99***		
		200	42.62 ± 1.71***		
Cephalosporins	Ceftizoxime	500	21.77 ± 2.19***	No effect	
		1000	22.61 ± 2.93***		
Tetracyclines	Minocycline	10	15.98 ± 2.48***	50	4.42 ± 0.3*
		50	12.13 ± 1.91**		
Aminoglycosides	Netilmicin	No effect		No effect	
* Based on 48 hours					
Efficacies of 4 different antibiotics on improving cell viability under normoxic (21% O ₂) and hypoxic (1% O ₂) conditions at 48 hours of incubation with each antibiotic are presented. "Efficacy (%)" indicates a percentage for cell viability of group by treated antibiotics compared to the control group.					
The markers such as *, ** and, *** represented $P < 0.05$, $P < 0.01$, and $P < 0.001$, respectively.					

Effect of ciprofloxacin on apoptosis under normoxia conditions

We first investigated the effect of terminal deoxynucleotidyl transferase-mediated d-UTP nick end labeling (TUNEL) assay to determine whether ciprofloxacin improved cell viability by inhibiting apoptosis. We carried out the experiments using a 100 µg/mL concentration under normoxia conditions because ciprofloxacin improved cell viability under both normoxia and hypoxia conditions at this concentration, but its efficacy was more prominent under normoxia conditions. In the TUNEL assay, there were no apoptotic bodies at a concentration of 100 µg/mL in comparison with the control group (Fig. 2A). The ratio of apoptotic cells to total cells was significantly lower in the ciprofloxacin group than in the control group (7.17 ± 1.25 vs. 20.41 ± 1.04 , $P < 0.001$; Fig. 2B). Therefore, the efficacy of ciprofloxacin on the viability of MDCK cells was significantly higher in comparison with the control group under normoxia conditions by inhibiting apoptosis.

The efficacy of ciprofloxacin on cell viability under stressful conditions via the interaction between autophagy and apoptosis

To elucidate underlying mechanisms further, we investigated apoptosis and autophagy markers. First, the upstream production of caspase 3, which causes apoptosis, was investigated. Cleaved caspase-3 to caspase-3, which is the final effector caspase in apoptosis, was lower in the ciprofloxacin group than in the control group after 36 h at the beginning of treatment (Fig. 3A). Second, the ratio of Bcl-2-associated X protein (Bax) to B-cell lymphoma 2 (Bcl-2) was investigated located upstream of caspase 3. The ratio of Bax to Bcl-2 was higher in the ciprofloxacin group than in the control group 6 h after the beginning of the treatment (Fig. 3B). Third, the ratio of pBcl2 (phosphorylated Bcl-2, which is an inactive form of Bcl2) to β -actin was higher in the ciprofloxacin group than in the control group 12 hours after the beginning of treatment (Fig. 3C). The results of caspase-3 experiments favor anti-apoptosis whereas Bcl-2 experiments favor apoptosis. To accommodate apparent contradictory results, we assumed that ciprofloxacin may also affect autophagy. The ratio of Beclin-1 (BECN1), an autophagy marker, to β -actin increased with time and was higher in the ciprofloxacin group at all times, indicating that ciprofloxacin increases autophagy (Fig. 3D). Finally, this indicates that ciprofloxacin enhanced autophagy, increasing the amount of free Beclin-1 via phosphorylated Bcl-2, and inhibited caspase activity (Fig. 4).

Discussion

In our study, we showed that ciprofloxacin increased the viability of MDCK cells by inhibiting apoptosis through an autophagy mechanism. Although all cells eventually undergo apoptosis, it was noteworthy that ciprofloxacin could increase cell viability in the process of apoptotic cell death.

When energy is lacking, the organism tries to save cells by recycling dead cells through autophagy activation. At this time, it is not yet clear whether autophagy is being promoted or whether apoptosis is being suppressed. Apoptosis refers to programmed cell death, which can be observed through DNA breakage¹. In our study, the TUNEL assay demonstrated a status of apoptosis. Recently, several antibiotics have been reported to control cell viability. Therefore, for the first time, we screened using an MTT assay whether cell viability increased when antibiotics representing each class were administered. We examined four antibiotics: ciprofloxacin, a quinolone, ceftizoxime, a cephalosporin, minocycline, a tetracycline, and netilmicin, an aminoglycoside. Ceftizoxime was effective only at high concentrations under normoxia conditions, but not under hypoxia conditions. Netilmicin was not effective under normoxia or hypoxia conditions. Minocycline was effective at lower concentrations but deteriorated cell viability at higher concentrations under both normoxia and hypoxia conditions. Therefore, we used the most effective and safe antibiotic, ciprofloxacin, for further studies.

The first experiment was performed to investigate whether ciprofloxacin improved cell viability by inhibiting apoptosis using the TUNEL assay. The results showed that apoptosis was reduced qualitatively and quantitatively when ciprofloxacin was added under the normoxia conditions. Therefore, ciprofloxacin saves cells by inhibiting apoptosis.

The second experiment was performed to investigate the mechanism of increased ciprofloxacin-induced cell viability. In previous studies, apoptosis has been reported to be mainly responsible for programmed cell death⁹, but autophagy has also been reported to play important roles². In addition, although apoptosis and autophagy have been considered as different concepts, many studies have shown that the homeostasis of cells is maintained through the interaction between these two molecular mechanisms^{10,11}. Firstly, we investigated apoptosis mechanism with the relationship among caspase-3, Bax, and Bcl-2. In general, when Bax/Bcl-2 decreases and caspase-3 decreases, apoptosis is suppressed. Our study showed reduction of the caspase-3 level in the ciprofloxacin-treated group, and this result means that the inhibition of apoptosis occurs inhibiting caspase-3 activity by ciprofloxacin. However, conversely, ciprofloxacin-treated cells showed an increase in Bax/Bcl-2 level in this study. This may not be explained by the fact that the whole process is caused by apoptosis because this means that an increased Bax/Bcl-2 level caused more apoptosis, which leads to conflicting results (Fig. 2 and Fig. 3B). Therefore, we investigated the influence of ciprofloxacin on autophagy in an energy-insufficient environment. Since Beclin-1 is inhibited by Bcl-2, when Bcl-2 is phosphorylated and inactivated, Beclin-1 and Bcl-2 are separated from each other and an increased level of free Beclin-1 becomes operational, resulting in an increase in pBcl-2. This means that the inhibition of autophagy induced by Bcl-2 is reduced, which can lead to a better progression of autophagy. In other words, an increase in the amount of free Beclin-1 promotes autophagy¹². First, the level of Beclin-1 increases steeply. Second, when the level of unbound pBcl-2 is increased, the level of free Beclin-1 is increased due to a decrease in the binding to Beclin-1. Third, as Bax increases, more Bcl-2 and more bonds can be obtained, and the amount of free Beclin-1 can be increased. Finally, caspase-3 cleaves and deactivates Beclin-1, and the decreasing level of caspase-3 increases the level of free Beclin-1. Finally, in our study, ciprofloxacin enhanced autophagy, increasing the level of free Beclin-1, and inhibited caspase activity and reduced cell death due to the energy produced during autophagy as summarized in Fig. 4. To survive in the absence of energy, cells die to generate energy sources for the living cells. In other words, when the energy source is insufficient, apoptosis is promoted upstream, but at the same time, autophagy is promoted to decompose the unnecessary proteins in the cell and use them as energy sources. Accordingly, the activity of caspase-3 is inhibited, thereby suppressing apoptosis.

The strength of this study is that it has shown a positive effect on cell viability using antibiotics, and this is the first study to apply the concept of autophagy and apoptosis. In previous studies, minocycline had potent anti-apoptotic and anti-inflammatory effects and protected renal function in a rat model of ischemia-reperfusion injury^{8,13,14}. Because minocycline showed toxic effects at high concentrations in the MDCK cell line in our study, we could explain the effect of ciprofloxacin, which is safer than other antibiotics for the action of autophagy, on cell viability. Recently, these concepts of autophagy have been applied in several clinical fields, such as acute kidney injury¹⁵, chronic kidney disease (obstructive nephropathy, immunoglobulin A nephropathy, autoimmune kidney diseases)¹⁶, diabetic nephropathy¹⁷, autosomal dominant polycystic kidney disease¹⁸, cystinosis¹⁹, and chronic cyclosporine A renal toxicity²⁰. Our findings may have a positive impact on acute kidney injury and kidney diseases associated with chronic inflammation.

However, our study has some limitations. First, the order of apoptosis and the autophagy process is still unclear. Autophagy is known to precede apoptosis²¹. However, our study did not show the preceding autophagy process, although the apoptosis and autophagy markers were checked at 0, 6, 12, 24, 36, and 48 h. Second, in our study, research on other markers of autophagy and apoptosis was insufficient, and further studies are needed. Third, a slightly stronger stimulus leads directly to apoptosis without the action of autophagy. Although nutrient depletion is the most powerful factor in inducing autophagy, the combined metabolic stress of hypoxia and nutrient depletion damages organs, proteins, and DNA, which ultimately leads to apoptosis. Recent studies have shown that autophagy mitigates metabolic stress to protect cells against extreme cell damage⁴. Autophagy is not only necessary as a tool to generate energy in a nutrient-free environment, but it also plays an important role in regulating protein and organ function and maintaining homeostasis. It is also important in situations of metabolic stress, that is, when energy is limited and intracellular damage is accelerated. However, it is important to recognize that autophagy cannot inhibit apoptosis completely in conditions in which apoptosis is induced. In other words, when cells are exposed to stresses, autophagy takes place temporally, but it cannot completely prevent the progression to apoptosis. Finally, extreme environments with no nutrient supply and no oxygen are unlikely to reveal the mechanisms by which antibiotics affect cell viability and are most likely to be biased by apoptosis processes. In the future, it will be necessary to study the difference between normoxia and hypoxia conditions in a nutrient-free environment.

In conclusion, ciprofloxacin can help to restore cell viability against renal tubular damage under nutrient-free conditions associated with autophagy. Therefore, ciprofloxacin might be helpful in increasing renal cell viability through the activation of autophagy under acute kidney injury. Further studies are needed to confirm the mechanism by which ciprofloxacin affects cell viability.

Methods

1. Materials

We used the MDCK cell line, which is a tubular epithelial kidney cell line, which was purchased from Korean Cell Line Bank (Seoul, South Korea)^{22,23}. We used four representative antibiotics from each spectrum, ciprofloxacin (CJ Healthcare Corp., Seoul, South Korea), ceftizoxime (DONG-A ST, Seoul, South Korea), minocycline (Sigma-Aldrich Co., St. Louis, MO, USA), and netilmicin (KUHNIL Pharm. Seoul, South Korea), to screen for effective antibiotics that improve MDCK cell viability after 48 hours of antibiotic treatment using a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) assay. Ciprofloxacin is one of the quinolones which block DNA replication via inhibition of DNA gyrase. Besides its antibiotic activity, some studies have reported the immunomodulatory effects of ciprofloxacin in rodent models and human clinical trials^{24,25}. Ceftizoxime is a cell wall inhibitor and a member of the third-generation cephalosporin available for parenteral administration. It binds to and inactivates penicillin-binding proteins of the inner membrane of the bacterial cell wall. This leads to weakness of the bacterial cell wall. Minocycline is a protein synthesis inhibitor (30S inhibitor) and a representative

tetracycline antibiotic that has been reported to have immunomodulatory and anti-inflammatory effects. In particular, minocycline inhibits apoptosis in certain neurodegenerative disorders and has potent anti-apoptotic and anti-inflammatory effects, as observed in a rat model of ischemia-reperfusion injury¹³. Minocycline improved streptozotocin-induced kidney damage by protecting against long-term hyperglycemia-induced kidney cell apoptosis¹⁴. Netilmicin is a protein synthesis inhibitor (30S inhibitor) and one of the members of the aminoglycoside family. It is used to kill a wide range of bacteria. Its action resembles that of gentamicin, but it is less nephrotoxic.

Primary antibodies for Bcl-2, caspase-3, cleaved caspase-3 (Cell Signaling Technology, Beverly, MS, USA), Bax (Santa Cruz Biotechnology, Santa Cruz, CA, USA), phosphorylated Bcl-2 (pBcl-2), and phosphorylated Bax (pBax) (Thermo Fisher Scientific, Waltham, MA, USA) were used as apoptosis markers. Beclin-1 primary antibody (OriGene Technology, Rockville, MD, USA) was used as an autophagy marker. A primary antibody against β -actin (Santa Cruz Biotechnology, Santa Cruz, CA, USA) was used as the control. The secondary antibody used was horseradish peroxidase-conjugated secondary antibody (Enzo, Farmingdale, NY, USA).

2. Methods

2.1 Cell viability measurement

We divided the operating conditions into normoxia and hypoxia conditions. The glucose media were not supplemented. We evaluated the effect of antibiotics on cell viability according to the type and concentration of the four antibiotics: ciprofloxacin, ceftizoxime, minocycline, and netilmicin. MDCK cells were plated (at a density of 7×10^4 cells/well) in a 48-well plate and grown in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with penicillin (100 units/mL), streptomycin (100 μ g/mL), and fetal bovine serum (FBS, 10%) (Gibco BRL, Gaithersburg, MD, USA) at 37°C for 48 h under normoxia conditions (5% CO₂ balanced with air) in a humidified chamber (Forma Scientific Inc., Marietta, OH, USA). After the media were replaced by DMEM without glucose or serum, ciprofloxacin was added, and the cells were cultured under normoxia and hypoxia conditions. Four antibiotics were added to each well at different concentrations. Ciprofloxacin (0, 50, 100, 150, and 200 μ g/mL), ceftizoxime (0, 10, 100, 200, 500, and 1,000 μ g/mL), minocycline (0, 10, 50, 100, 200, and 500 μ g/mL), and netilmicin (0, 1, 10, 100, 500, and 1,000 μ g/mL) were used. The effects of the antibiotics on cell viability were evaluated using an MTT assay. An MTT assay, which is a tetrazolium-based colorimetric assay, was used to assess cell viability because it allows an easy and rapid reading of many samples²⁶. MTT (5 mg/mL) was added to each well. Because MTT solutions are sensitive to light, the exposure to light was minimized. The plates were placed in a 37°C and 5% CO₂ incubator for 2 h. Dimethyl sulfoxide (DMSO) was dispensed after removing the MTT solution. The plates were shaken for 10–15 min while being protected from light. The absorbance was measured using a plate reader at a wavelength of 570 nm.

2.2 TUNEL assay

MDCK cells were plated (at a density of 1×10^5 cells/well) on a Lab-Tek® 4 well chamber slide (Thermo Fisher Scientific, Waltham, MA, USA) and grown in DMEM culture medium, as described above. Twenty-four hours after plating, a TUNEL assay was performed to detect the nicks generated from DNA strand breaks¹⁰. The aim of the assay was to identify indicators of damage due to apoptosis²⁶. We used ciprofloxacin (100 µg/mL) and ApopTag® Peroxidase *In Situ* Apoptosis Detection Kit (Merck Millipore, Burlington, MA, USA). We fixed cells in 1% paraformaldehyde in phosphate-buffered saline (PBS), pH 7.4 and washed them twice with PBS for 5 min. We performed post-processing using precooled ethanol: acetic acid (2:1) at -20°C in a Coplin jar for 5 min, and drained it. We quenched the cells in 3.0% hydrogen peroxide in PBS at room temperature for 5 min. We rinsed the specimen twice with PBS for 5 min each time in a Coplin jar. An equilibration buffer (75 µL/5 cm²) was immediately added to the sample. We immediately pipetted the working strength terminal deoxynucleotidyl transferase (TdT) enzyme solution (55 µL/5 cm²) onto the section. The cells were incubated for 1 h at 37°C in a humidified chamber. The specimen was placed in a Coplin jar containing the working strength stop/wash buffer and stirred for 15 s and then incubated at room temperature for 10 min. We applied anti-digoxigenin peroxidase conjugate to the slide at room temperature (approximately 65 µL/5 cm² of the surface covered). The cells were incubated in a humidified chamber for 30 min at room temperature. We applied sufficient peroxidase substrate to completely cover the specimen (75 µL/5 cm²). We stained the cells for 3 to 6 minutes at room temperature. To determine the optimal staining time, we monitored the color change of cells under the microscope. We counterstained the cells in a Coplin jar for 10 min at room temperature using 0.5% methyl green. The specimens were dehydrated by moving the slides through three xylene jars while incubating for 2 min in each Coplin jar. We removed the slides from the Coplin jar and mounted them under a glass coverslip in a mounting medium.

2.3 Western blot analysis

Cell extracts were prepared by incubating the cells in cell lysis buffer [1X radioimmunoprecipitation assay buffer (RIPA) Buffer: 20 mM Tris-HCl (pH 7.5), 150 mM NaCl, 1 mM ethylenediaminetetraacetic acid disodium salt dehydrate (Na₂EDTA), 1 mM ethylene glycol-bis(β-aminoethyl ether)-N,N,N',N'-tetraacetic acid (EGTA), 1% nonyl phenoxypolyethoxyethanol (NP)-40, 1% sodium deoxycholate, 2.5 mM sodium pyrophosphate, 1 mM β-glycerophosphate, 1 mM Na₃VO₄, and 1 µg/mL leupeptin]. A total of 70 µg of caspase 3, cleaved caspase 3, and BECN1 proteins were separated using electrophoresis by using a 12% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to a polyvinylidene fluoride (PVDF) membrane. After the membranes were blocked with buffer [TBS-T buffer: 20 mM Tris (pH 7.5), 150 mM NaCl, and 0.1% Tween 20] containing 5% skim milk, the membranes were incubated with the primary antibodies. After washing, the membranes were incubated with the secondary antibody. Then, the membranes were washed with TBS-T buffer and developed using an enhanced chemiluminescence (ECL) Western blot assay detection reagent (GE Healthcare, Little Chalfont, UK). The intensity of each band was quantified using a ChemiDoc™ XRS + System with Image Lab™ Software (Bio-Rad, Hercules, CA, USA). A Western blotting assay was performed more than 3 times to investigate the

apoptosis and autophagy markers in order to assess the reproducibility, and thus, the reliability of the results.

2.4 Statistical analysis

Experiments were repeated at least three times to obtain consistent results. One-way analysis of variance (ANOVA) was performed. Shapiro-Wilk statistics were used to verify the normal distribution, and Levene statistics were used to analyze the homogeneity of variance. Tukey's test was performed for *post hoc* multiple comparisons. Student's *t*-test was performed and the data were expressed as the mean \pm standard deviation. In the non-normal distribution, the Kruskal-Wallis test was performed. The Statistical Package for the Social Science (SPSS) version 18.0 (SPSS Inc., Chicago, IL, USA) was used. The results were statistically significant at a P value lower than 0.05.

Declarations

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Authors' Contributions

Research idea and study design: J.W.L, S.H.L; data acquisition: W.Y.P, Y.K, J.H.P, K.J; data analysis/interpretation: W.Y.P, K.S.A; statistical analysis: W.Y.P; supervision: J.W.L, K.S.A, S.H. Each author contributed important intellectual content during manuscript drafting or revision and accepts accountability for the overall work by ensuring that questions pertaining to the accuracy or integrity of any portion of the work are appropriately investigated and resolved. J.W.L and K.S.A take responsibility that this study has been reported honestly, accurately, and transparently; that no important aspects of the study have been omitted; and that any discrepancies from the study as planned have been explained.

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Data Availability

All data generated or analyzed during this study are included in this published article and its Supplementary Information files. The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

Competing Interests Statement: The author(s) declare no competing interests.

References

1. Fuchs, Y. & Steller, H. Programmed cell death in animal development and disease. *Cell* **147**, 742-758, doi:10.1016/j.cell.2011.10.033 (2011).
2. Eisenberg-Lerner, A., Bialik, S., Simon, H. U. & Kimchi, A. Life and death partners: apoptosis, autophagy and the cross-talk between them. *Cell death and differentiation* **16**, 966-975, doi:10.1038/cdd.2009.33 (2009).
3. Gump, J. M. & Thorburn, A. Autophagy and apoptosis: what is the connection? *Trends in cell biology* **21**, 387-392, doi:10.1016/j.tcb.2011.03.007 (2011).
4. Havasi, A. & Dong, Z. Autophagy and Tubular Cell Death in the Kidney. *Seminars in nephrology* **36**, 174-188, doi:10.1016/j.semnephrol.2016.03.005 (2016).
5. Han, S. J. & Lee, H. T. Mechanisms and therapeutic targets of ischemic acute kidney injury. *Kidney research and clinical practice* **38**, 427-440, doi:10.23876/j.krcp.19.062 (2019).
6. Bbosa, G. S., Mwebaza, N., Odda, J., Kyegombe, D. B. & Ntale, M. J. H. Antibiotics/antibacterial drug use, their marketing and promotion during the post-antibiotic golden age and their role in emergence of bacterial resistance. **2014** (2014).
7. Tang, C. *et al.* Antibiotic drug tigecycline inhibited cell proliferation and induced autophagy in gastric cancer cells. *Biochemical and biophysical research communications* **446**, 105-112, doi:10.1016/j.bbrc.2014.02.043 (2014).
8. Sutton, T. A. *et al.* Minocycline reduces renal microvascular leakage in a rat model of ischemic renal injury. *American journal of physiology. Renal physiology* **288**, F91-97, doi:10.1152/ajprenal.00051.2004 (2005).
9. Galluzzi, L. *et al.* Molecular definitions of cell death subroutines: recommendations of the Nomenclature Committee on Cell Death 2012. *Cell death and differentiation* **19**, 107-120, doi:10.1038/cdd.2011.96 (2012).
10. Kim, J. H. *et al.* Nrf2-Heme oxygenase-1 modulates autophagy and inhibits apoptosis triggered by elevated glucose levels in renal tubule cells. *Kidney research and clinical practice*, doi:10.23876/j.krcp.18.0152 (2019).
11. Rubinstein, A. D. & Kimchi, A. Life in the balance - a mechanistic view of the crosstalk between autophagy and apoptosis. *Journal of cell science* **125**, 5259-5268, doi:10.1242/jcs.115865 (2012).
12. Gordy, C. & He, Y. W. The crosstalk between autophagy and apoptosis: where does this lead? *Protein & cell* **3**, 17-27, doi:10.1007/s13238-011-1127-x (2012).
13. Wu, Z. *et al.* Minocycline is effective in intracerebral hemorrhage by inhibition of apoptosis and autophagy. *Journal of the neurological sciences* **371**, 88-95, doi:10.1016/j.jns.2016.10.025 (2016).
14. Yuan, H. *et al.* Minocycline Attenuates Kidney Injury in a Rat Model of Streptozotocin-Induced Diabetic Nephropathy. *Biological & pharmaceutical bulletin* **39**, 1231-1237, doi:10.1248/bpb.b15-00594 (2016).
15. Kaushal, G. P. & Shah, S. V. Autophagy in acute kidney injury. *Kidney international* **89**, 779-791, doi:10.1016/j.kint.2015.11.021 (2016).

16. Liu, N., Shi, Y. & Zhuang, S. Autophagy in Chronic Kidney Diseases. *Kidney diseases (Basel, Switzerland)* **2**, 37-45, doi:10.1159/000444841 (2016).
17. Kume, S. & Koya, D. Autophagy: A Novel Therapeutic Target for Diabetic Nephropathy. *Diabetes & metabolism journal* **39**, 451-460, doi:10.4093/dmj.2015.39.6.451 (2015).
18. Ravichandran, K. & Edelstein, C. L. Polycystic kidney disease: a case of suppressed autophagy? *Seminars in nephrology* **34**, 27-33, doi:10.1016/j.semnephrol.2013.11.005 (2014).
19. Gahl, W. A., Thoene, J. G. & Schneider, J. A. Cystinosis. *The New England journal of medicine* **347**, 111-121, doi:10.1056/NEJMra020552 (2002).
20. Kim, H. S., Choi, S. I., Jeung, E. B. & Yoo, Y. M. Cyclosporine A induces apoptotic and autophagic cell death in rat pituitary GH3 cells. *PloS one* **9**, e108981, doi:10.1371/journal.pone.0108981 (2014).
21. Marino, G., Niso-Santano, M., Baehrecke, E. H. & Kroemer, G. Self-consumption: the interplay of autophagy and apoptosis. *Nature reviews. Molecular cell biology* **15**, 81-94, doi:10.1038/nrm3735 (2014).
22. Arthur, J. M. The MDCK cell line is made up of populations of cells with diverse resistive and transport properties. *Tissue & cell* **32**, 446-450, doi:10.1054/tice.2000.0135 (2000).
23. Dukes, J. D., Whitley, P. & Chalmers, A. D. The MDCK variety pack: choosing the right strain. *BMC cell biology* **12**, 43, doi:10.1186/1471-2121-12-43 (2011).
24. Fukumoto, R. *et al.* Ciprofloxacin modulates cytokine/chemokine profile in serum, improves bone marrow repopulation, and limits apoptosis and autophagy in ileum after whole body ionizing irradiation combined with skin-wound trauma. *PloS one* **8**, e58389, doi:10.1371/journal.pone.0058389 (2013).
25. Park, C. H. *et al.* Identification, biological activity, and mechanism of the anti-ischemic quinolone analog. *Bioorganic & medicinal chemistry* **15**, 6517-6526, doi:10.1016/j.bmc.2007.07.009 (2007).
26. van Meerloo, J., Kaspers, G. J. & Cloos, J. Cell sensitivity assays: the MTT assay. *Methods in molecular biology (Clifton, N.J.)* **731**, 237-245, doi:10.1007/978-1-61779-080-5_20 (2011).

Figures

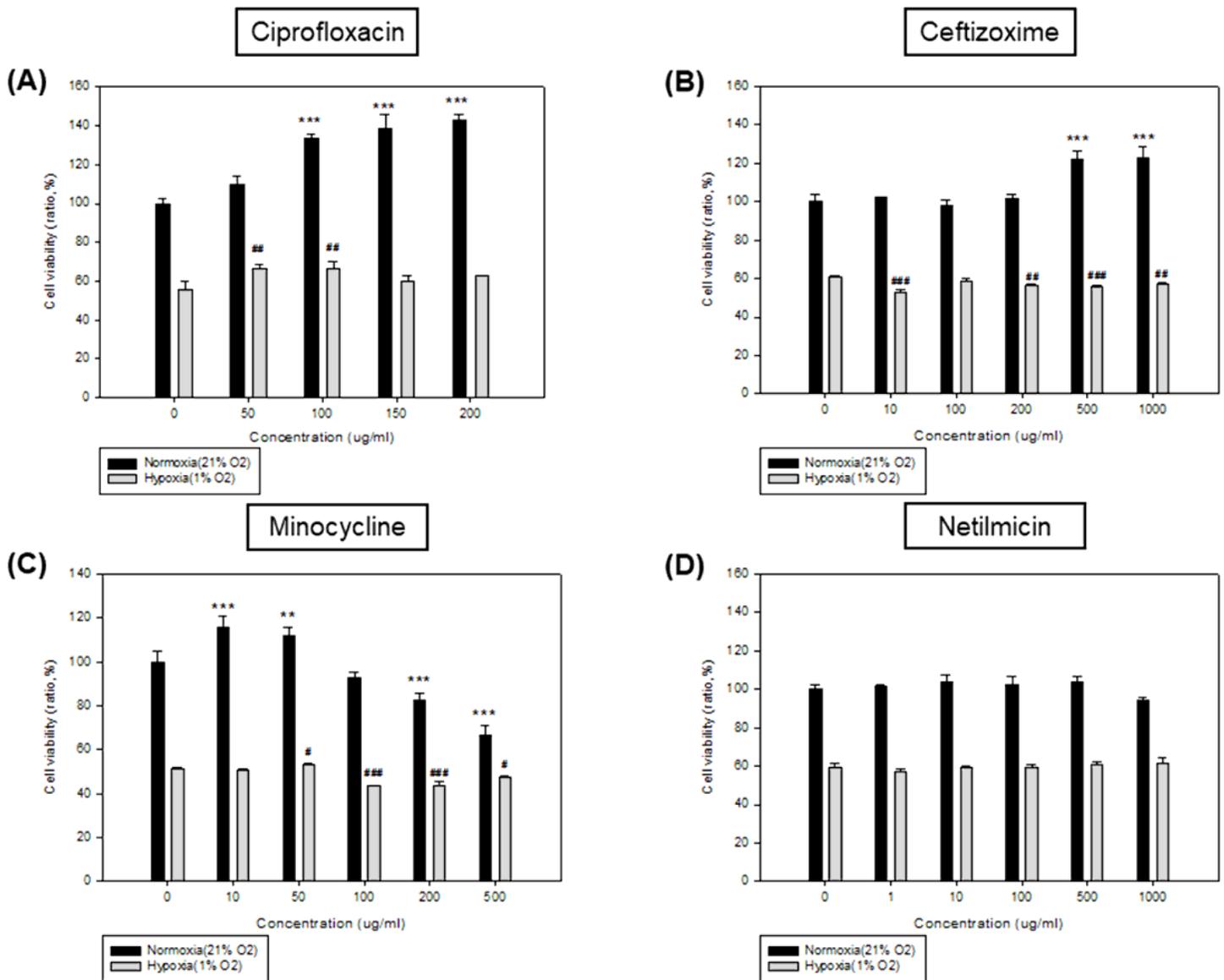


Figure 1

The relationship between the concentration of (A) ciprofloxacin, (B) ceftizoxime, (C) minocycline, and (D) netilmicin and cell viability. MDCK cells were plated in DMEM with glucose (1 g/L) and FBS (10%) at a density of 7×10^4 per well in 48-well plates under normoxia condition (21% O₂). After 48 hours of incubation, the medium was exchanged with DMEM without glucose or FBS. After antibiotics were added at appropriate concentrations, respectively, the cells were incubated further under normoxia condition (21% O₂) and hypoxia condition (1% O₂). Concentrations of antibiotics added were as follows; (A) Ciprofloxacin (0, 50, 100, 150, and 200 µg/mL); (B) Ceftizoxime (0, 10, 100, 200, 500, and 1000 µg/mL); (C) Minocycline (0, 10, 50, 100, 200, and 500 µg/mL); (D) Netilmicin (0, 1, 10, 100, 500, and 1000 µg/mL). Cell viability was assessed through MTT assay at 48 hours of antibiotics treatment. The experiment at ciprofloxacin concentration was performed more than 3 times. (Normoxia vs *** P < 0.001; Hypoxia vs ## P < 0.01).

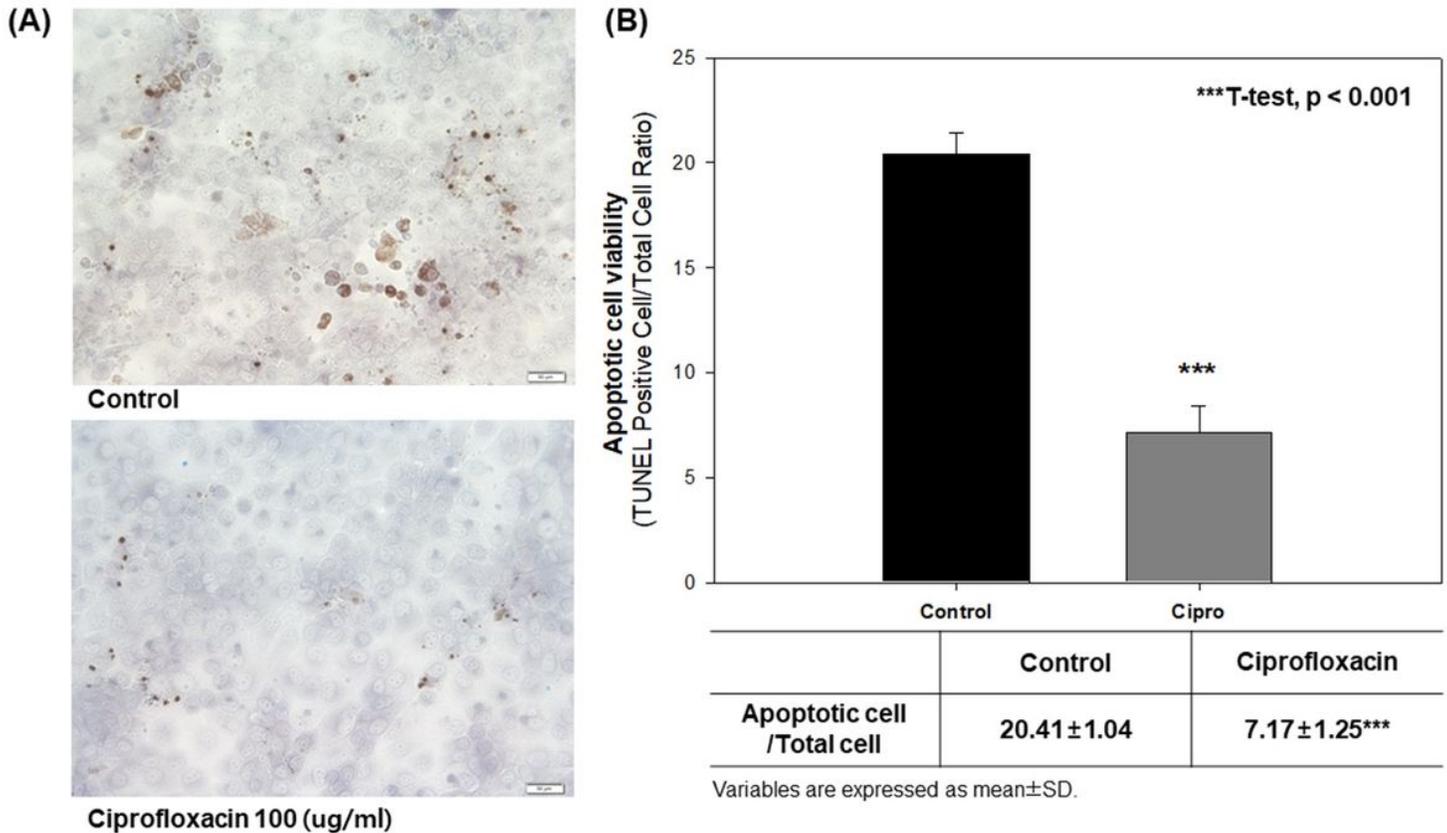


Figure 2

(A) The qualitative and (B) quantitative comparison of the difference of apoptotic bodies between in the control group and ciprofloxacin group. ApopTag® Peroxidase In Situ Apoptosis Detection Kit was used. A concentration of 100 µg/mL of ciprofloxacin was used in comparison with control group. The examination was done under normoxia condition (21% O₂). In the TUNEL assay, apoptotic cells from nicks of DNA strand breaks by labeling DNA strand breaks with fluorochromes were identified and quantified by fluorescence microscopy. The ratio of TUNEL positive cell to total cells was calculated for apoptotic cell viability. All data were performed more than 3 times in order to assess reproducibility and reliability of the results. Student's t-test was used statistically. ***T-test, $P < 0.001$ vs. control

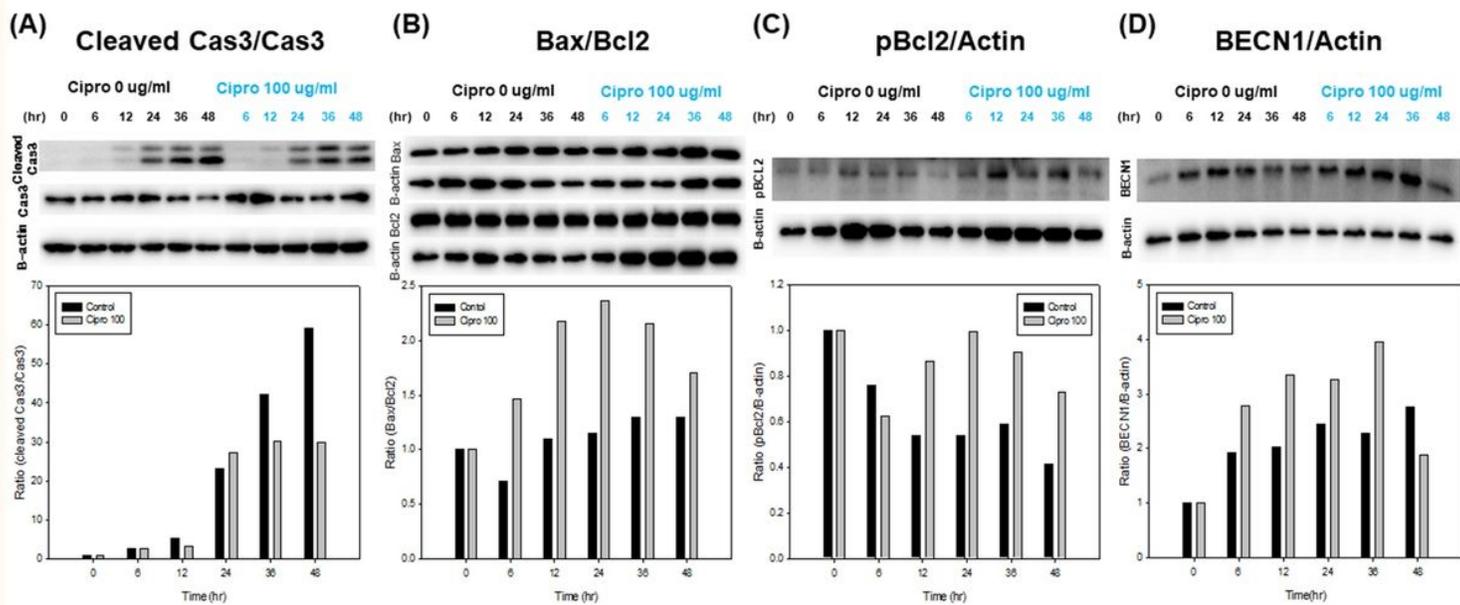


Figure 3

The comparison of (A) cleaved caspase 3/caspase 3, (B) Bax/Bcl2, (C) pBcl2/ β -actin, and (D) BECN1/ β -actin between in the control group and ciprofloxacin group. A concentration of 100 μ g/mL of ciprofloxacin was used in comparison with control group. β -actin was used for the control. The examination was done under normoxia condition (21% O₂). Proteins were detected by western blot analysis. Cell viability was checked at 0, 6, 12, 24, and 48 hours after treatments. All data were performed more than 3 times in order to assess reproducibility and reliability of the results.

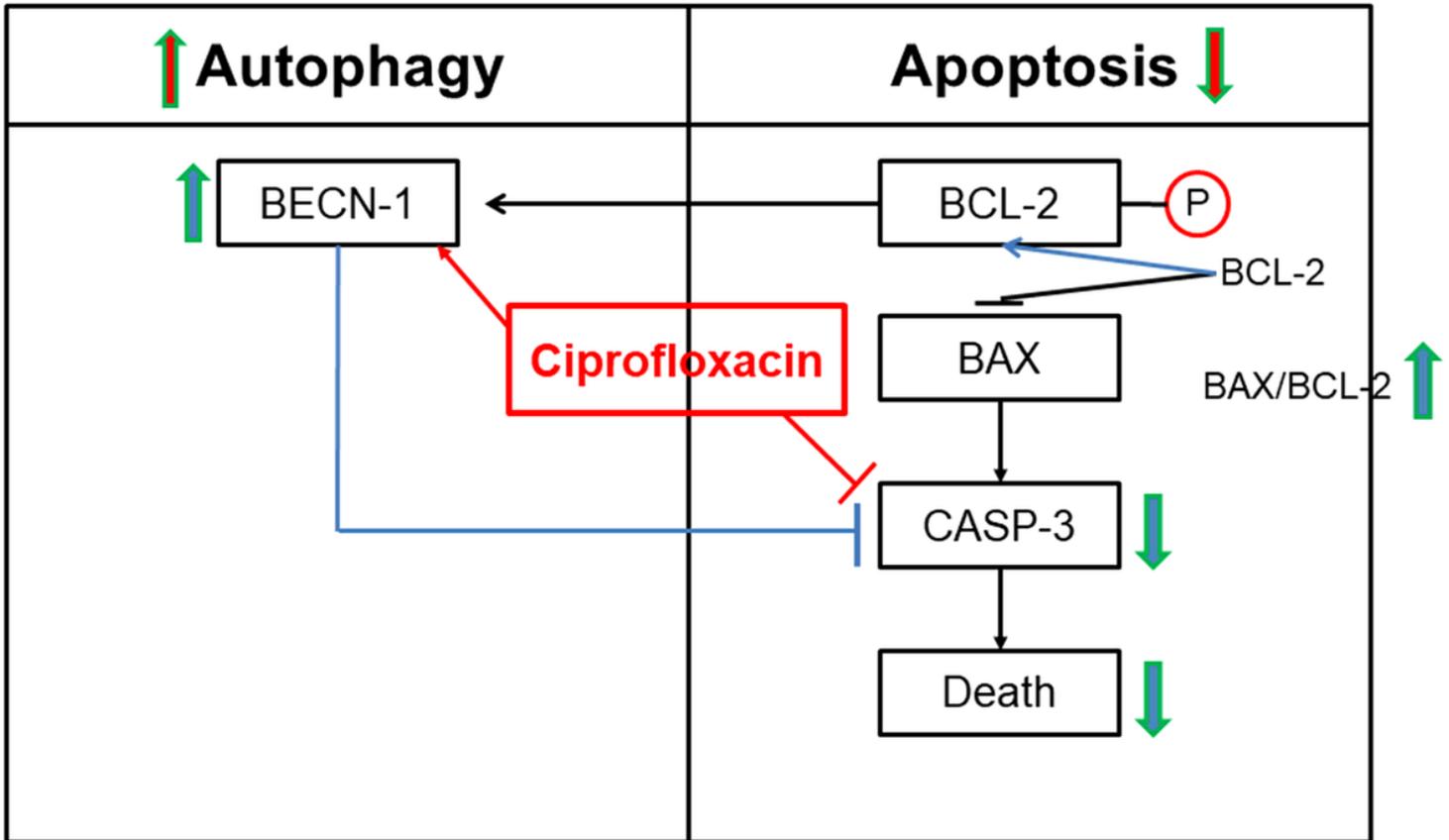


Figure 4

Overview of the molecular mechanisms of underlying the autophagy-apoptosis crosstalk by ciprofloxacin. This figure shows the relationship between autophagy and apoptosis for the effect of ciprofloxacin. The ratio of cleaved caspase 3 to caspase 3 (cleaved caspase 3/caspase 3) was decreased, but the ratio of Bax to Bcl-2 (Bas/Bxl-2) upstream of caspase 3 was not decreased in the ciprofloxacin group. Conversely, the levels of phosphorylated Bcl-2 and Beclin-1 were increased, and that of caspase-3 was decreased in the ciprofloxacin group.