Effect of Inhibition of GLUT1 Expression and Autophagy Modulation on the Growth and Migration of Laryngeal Carcinoma Stem Cells under Hypoxic and Low-Glucose Conditions

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Research

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Abstract

Background: Enhanced glucose uptake and autophagy are means by which cells adapt to stressful microenvironments. We investigated the roles of glucose transporter-1 (GLUT-1) and autophagy in laryngeal carcinoma stem cells under hypoxic and low-glucose conditions.

Methods: CD133⁺ Tu212 laryngeal carcinoma stem cells were purified by magnetic-activated cell sorting and subjected to hypoxic and/or low-glucose conditions. Proliferation was evaluated using a cell-counting kit and a clone-formation assay, and migration was evaluated through a Transwell assay. Autophagy was assessed via transmission electron microscopy. GLUT-1 and beclin-1 expression were silenced using an shRNA and autophagy was manipulated using rapamycin, 3-MA, or chloroquine. Gene expression levels were evaluated by quantitative reverse transcription-polymerase chain reaction and protein concentrations were assessed via Western blotting.

Results: Compared to CD133⁻ stem cells, CD133⁺ cells showed increased proliferation and migration, and reduced apoptosis, under hypoxic or low-glucose conditions. They also showed increased expression of GLUT-1 and autophagy markers. Finally, GLUT-1 knockdown or autophagy inhibition reduced their proliferation and migration.

Conclusions: Enhanced glucose uptake and autophagy maintain the functions of CD133⁺ laryngeal carcinoma stem cells under hypoxic and low-glucose conditions.

Background

Laryngeal cancer is a common malignant tumor of the head and neck. The 5-year survival rate of laryngeal cancer has not improved in the past 40 years despite the near-continuous development of diagnostic and treatment methods [1]. The lack of understanding of the mechanisms underlying the functions of laryngeal cancer cells has hampered the development of effective therapeutic strategies.

Compared to ordinary tumor cells, cancer stem cells (CSCs) are typically resistant to apoptosis and therapeutic agents [2,3]. CSCs are closely related to the occurrence, development, metastasis, recurrence, and chemoradiotherapy resistance of tumors [4]. Laryngeal CSCs reportedly express CD133 as a marker [5]. CD133⁺ CSCs have higher tumorigenic and invasive abilities than CD133⁻ cancer cells [5–7,10]. However, whether CD133⁺ CSCs have specific metabolic mechanisms is unclear.

Autophagy provides energy for cells from the degradation of proteins and organelles. It can restrain the growth of tumor cells. However, it is also an adaptive response to stressful tumor microenvironments, preventing apoptosis of tumor cells [11,12]. A basal level of autophagy may promote the survival of cancer cells [11,12]. However, prolonged and excessive activation of autophagy may induce self-degradation and death of tumor cells [13].

CD133⁺ CSCs show greater proliferation and a lower frequency of apoptosis compared to CD133⁻ cancer cells in multiple types of tumors, particularly under hypoxic or nutrient-deprived conditions [14,15]. Interestingly, autophagy induction reportedly promotes the conversion of non-stem pancreatic cancer cells into CD133⁺ stem-like cells under intermittent hypoxia [16]. The expression of glucose transporter-1 (GLUT-1) is reportedly associated with autophagy activation in CSCs under hypoxic or nutrient-deprived conditions [17–20]. Autophagy may also affect cellular glucose uptake [21–23]. Beclin-1, an autophagy marker, plays an important role in the initiation of autophagy [24]. It promotes the localization of other autophagy-related proteins to autophagosomes, thus promoting the formation and maturation of autophagosomes. High expression of beclin-1 is typically accompanied by enhanced autophagy, increased GLUT-1 expression, and increased glucose uptake in certain types of tumors, such as non-small-cell lung carcinoma and breast cancer [25–27]. However, in one study, beclin-1 and GLUT-1 expression were negatively correlated in 29 cases of head-and-neck squamous cell carcinoma [28], indicating that the association between autophagy and glucose metabolism may be cancer-type related. In this work, we investigated the regulation by GLUT-1 and autophagy of the functions of laryngeal carcinoma stem cells under hypoxic or low-glucose conditions, as well as the underlying mechanisms.

Materials And Methods

Cell culture and treatment

Tu212 cells were purchased from the Cell Research Institute of the Chinese Academy of Sciences (Shanghai, China). Tu212 cells were cultured in Roswell Park Memorial Institute-1640 medium (Gibco-BRL, Gaithersburg, MD), supplemented with 10% heat-inactivated fetal bovine serum (FBS; Hyclone, Logan, UT), 100 U/mL penicillin, and 100 g/mL streptomycin at 37°C in an atmosphere containing 5% CO₂.

Sorting and identification of Tu212 laryngeal carcinoma cells

Magnetic sorting. Briefly, 1×10^7 Tu212 cells were resuspended in phosphate-buffered saline (PBS). The resuspended cells were added to 100 μ L FcR Blocking Reagent and 100 μ L CD133 MicroBeads, mixed, and incubated at 4°C for 30 min. Next, 2 mL PBS was added, and the cells were centrifuged at $300 \times g$ for 10 min; the supernatant was discarded. Subsequently, the cell pellet was resuspended in 500 μ L PBS. The magnetic separation (MS) column was clipped to the magnetic separator and 500 μ L PBS was added to moisten the column. Next, the cell suspension was added to the MS sorting column. The MS separation column was washed three times with 500 μ L PBS to remove unbound cells. The column was removed from the magnetic separator, 1 mL PBS was added, and the cells were expelled from the column using a push rod. After centrifugation at $300 \times g$ for 10 min, the supernatant was discarded, and the cells were resuspended in 1 mL PBS and enumerated.

Determination of the purity of Tu212 CD133⁺ cells. Cells (2×10^5) were removed before and after separation, centrifuged, and the supernatant was discarded. Next, 80 μ L PBS and 10 μ L anti-human

CD133-PE were added. The sample was gently mixed using a micropipette and incubated at 4°C for 10 min. The cells were centrifuged at 1000 rpm for 5 min and the supernatant was discarded. Precooled PBS (1 mL) was added, and unbound excess antibody components were removed by two centrifugation and washing steps. After adding 4% paraformaldehyde and incubation at 4°C for 20 min, the supernatant was centrifuged. The cells were transferred to a flow tube and stored at 4°C protected from light. Flow cytometry was performed using the standard procedure (Beckman, Fullerton, CA).

Experimental groups

Relationship between the proliferation and migration of Tu212 CD133+ cells and the levels of GLUT-1 and autophagy under hypoxia and low-glucose conditions. Eight groups were used in this experiment: CD133⁺ (20% O₂, 25 mM Glu); CD133⁻ (20% O₂, 25 mM Glu); CD133⁺+hypoxia (1% O₂, 25 mM Glu); CD133⁻+hypoxia (1% O₂, 25 mM Glu); CD133⁺+low Glu (20% O₂, 2.5 mM Glu); CD133⁻+low Glu (20% O₂, 2.5 mM Glu); CD133⁺+hypoxia+low Glu (1% O₂, 2.5 mM Glu); and CD133⁻+hypoxia+low Glu (1% O₂, 2.5 mM Glu).

Cell growth, migration, and levels of GLUT-1 and autophagy-related proteins. Fourteen groups were used in this experiment: CD133⁺+hypoxia+low Glu (1% O₂, 2.5 mM Glu); CD133⁻+hypoxia+low Glu (1% O₂, 2.5 mM Glu); CD133⁺+hypoxia+low Glu (1% O₂, 2.5 mM Glu)+NC shRNA; CD133⁻+hypoxia+low Glu (1% O₂, 2.5 mM Glu)+NC shRNA); CD133⁺+hypoxia+low Glu (1% O₂, 2.5 mM Glu+GLUT-1 shRNA); CD133⁻+hypoxia+ low Glu (1% O₂, 2.5 mM Glu)+GLUT-1 shRNA); CD133⁺+hypoxia+low Glu (1% O₂, 2.5 mM Glu+beclin-1 shRNA); CD133⁻+hypoxia+low Glu (1% O₂, 2.5 mM Glu+beclin-1 shRNA); CD133⁺+hypoxia+low Glu (1% O₂, 2.5 mM Glu+3-MA); CD133⁻+hypoxia+low Glu (1% O₂, 2.5 mM Glu+3-MA); CD133⁺+hypoxia+low Glu (1% O₂, 2.5 mM Glu+CQ); CD133⁻+hypoxia+low Glu (1% O₂, 2.5 mM Glu+CQ); CD133⁺+hypoxia+low Glu (1% O₂, 2.5 mM Glu)+rapamycin); and CD133⁻+hypoxia+low Glu (1% O₂, 2.5 mM Glu)+rapamycin).

Clonogenic assay

The cell suspension was dispersed; the percentage of individual cells was greater than 95%. Next, cells were counted, and the cell density was adjusted to 250/mL by adding culture medium. The cell suspension was added to the wells of a six-well plate (2 mL per well), and the plate was gently shaken. The plate was placed in an incubator for 2 to 3 weeks, and the medium was replaced every 3 days. The culture was terminated when clones became visible. The medium was discarded, and the cells were gently washed twice with PBS, stained with 1% crystal violet at room temperature for 1 h, and photographed.

Cell-Counting Kit-8 assay

Cells were incubated at 37°C in an atmosphere containing 5% CO₂ for 48 h. Next, 20 µL Cell Counting Kit-8 (CCK-8) solution was added, and the cells were incubated in the dark for 1 h. The absorption at 450 nm

of the suspension was measured using a Spectra Plus Microplate Reader (Molecular Devices, Sunnyvale, CA).

Flow cytometry

Briefly, 10× Binding Buffer was diluted 1:10 with deionized water. Cells were collected by centrifugation for 5 min, exposed to reagents, digested, and resuspended in 500 µL Annexin V binding buffer. Next, 5 µL fluorescein isothiocyanate and 10 µL propidium iodide (Sigma Aldrich Co., St. Louis, MO) were added for 10 min in darkness at RT. Finally, the proportions of non-apoptotic and apoptotic cells were determined in triplicate by flow cytometry with ModFit LT software (Becton Dickinson, Mountain View, CA)

Transwell assay

Cells were digested with trypsin and the culture medium was discarded. Next, the cells were washed once or twice with PBS and resuspended in serum-free medium (containing 0.2% bovine serum albumin) to a density of 1×10^6 /mL. Cell suspension (200 µL) was added to the upper Transwell chamber and 600 µL FBS was added to the lower chamber. The cells were incubated in a 5% CO₂ atmosphere at 37°C for 24 h. The Transwell chamber was removed, and the culture medium was discarded. Then the chamber was washed twice with calcium-free PBS, fixed in formaldehyde for 30 min, air-dried, and the cells were stained with 0.1% crystal violet for 20 min. Finally, the upper layer of unigrated cells was gently removed using cotton swabs and washed three times with PBS.

Quantitative real-time polymerase chain reaction

The cells were collected, washed three times with precooled PBS, centrifuged at 1500 rpm for 3 min, and lysed on ice in the presence of TRIzol. Total RNA was extracted from the cells according to the manufacturer's instructions. Briefly, 1 µg RNA was reverse-transcribed using a First-Strand cDNA Synthesis Kit (K1622; Fermentas, Burlington, ON, Canada) and amplified by PCR using a SYBR Green qPCR Kit (Merck, Darmstadt, Germany). The PCR program was 37°C for 60 min, 85°C for 5 min, and 4°C for 5 min. The amplification products were stored at –20°C. The primers for GLUT-1, Beclin-1, Atg7, Atg5, and LC3 were designed and synthesized by Sangon Biotech (Table 1). The $2^{-\Delta\Delta C_t}$ method was used to calculate relative gene expression levels.

Western blotting

Total proteins were extracted from cells and tumor tissues in radioimmunoprecipitation assay buffer. The cells were collected, washed three times with precooled PBS, and centrifuged at 1500 rpm for 3 min. An appropriate volume of cell lysate was added, and the cells were left on ice for 30 min. The supernatant was centrifuged at 1200 rpm at 4°C for 30 min and stored at –80°C. After assaying the protein concentration, samples were added to 4× sodium dodecyl sulfate loading buffer, boiled for 5 to 10 min, and centrifuged at 12,000 × g for 1 min. Proteins (30 µg) were subjected to SDS-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to a polyvinylidene difluoride membrane (Millipore). Primary

antibodies against GLUT-1 (Abcam), beclin-1, LC3 (Proteintech), Atg7, Atg5, and β -actin (Abcam) were added and incubated at 4°C overnight; β -actin served as the internal control. After washing three times with Tris-buffered saline/Tween 20, the secondary antibodies were added for 1 h at room temperature. The signal was developed using an enhanced chemiluminescence assay kit (Beyotime Biotech) and analyzed semi-quantitatively using the ChemiDoc XRS+ System (Bio-Rad).

Transmission electron microscopy

Cells were collected, washed in PBS, fixed in 2.5% glutaraldehyde, post-fixed in 1% osmium tetroxide, and dehydrated in ethanol and acetone. After embedding in epoxy resin, sections were cut and stained with uranyl acetate and lead citrate. Autophagy was visualized by transmission electron microscopy (TEM; Thermo Fisher Scientific, Waltham, MA).

Results

Proliferation, migration, and apoptosis of CD133⁺ stem cells. To investigate the role of GLUT-1 in LCSCs, we first sorted CD133⁺ Tu212 cells. Flow cytometry showed that the proportion of CD133⁺ Tu212 cells was ~8% before sorting and surpassed 90% after sorting. In addition, the Tu212 CD133⁺ cells showed good viability (Fig. 1a).

Next, we investigated the growth, proliferation, apoptosis, and migration of CD133⁺ Tu212 cells. The clonal number of Tu212 CD133⁺ cells was significantly greater than that of Tu212 CD133⁻ cells under hypoxic (1% O₂), low-glucose (2.5 mM glucose), and hypoxic+low-glucose conditions. However, the clonal number of CD133⁺ and CD133⁻ Tu212 cells was similar under normal culture conditions (Fig. 1b). Moreover, compared to CD133⁻ cells, the proliferation of Tu212 CD133⁺ cells was significantly increased under hypoxic, low-glucose, and hypoxic+low-glucose conditions (Fig. 1c). Flow cytometry showed that the apoptotic rate of Tu212 CD133⁺ cells was significantly lower than that of Tu212 CD133⁻ cells under hypoxic, low-glucose, and hypoxic+low-glucose conditions (Fig. 1d). In a Transwell assay, the migration of Tu212 CD133⁺ cells was greater than that of Tu212 CD133⁻ cells under all three stress conditions (Fig. 1e). By contrast, proliferation and apoptosis were comparable between Tu212 CD133⁺ cells and CD133⁻ cells under normal culture conditions, with CD133⁺ cells showing only a slight increase in migration. Hence, CD133⁺ laryngeal carcinoma stem cells showed greater proliferation and migration than CD133⁻ cells in the presence of stressors.

GLUT-1 expression and autophagy of CD133⁺ laryngeal carcinoma stem cells. GLUT-1 overexpression promotes the proliferation, invasion, and metastasis of cancer cells, particularly under hypoxia and starvation conditions [6,7]. Activation of autophagy promotes the functions of CD133⁺ cells under hypoxia [15,16], likely by enhancing glucose uptake [21–23]. Therefore, we explored whether the increased proliferation and migration of CD133⁺ Tu212 cells were associated with elevated GLUT-1 expression and autophagy. To this end, we investigated GLUT-1 expression and autophagy in Tu212 cells

under hypoxic and low-glucose conditions. The GLUT-1 mRNA level in Tu212 CD133⁺ cells was significantly higher than that in Tu212 CD133⁻ cells under normal, hypoxic, low-glucose, and hypoxic+low-glucose conditions. The GLUT-1 mRNA level of Tu212 CD133⁺ cells under hypoxic, low-glucose, and hypoxic+low-glucose conditions was higher than that under normal culture conditions (Fig. 2a). Western blotting showed that the GLUT-1 protein level in Tu212 CD133⁺ cells was significantly higher than that in Tu212 CD133⁻ cells irrespective of the culture conditions (Figs. 2b, S1).

Hypoxic or low-glucose conditions increased the expression of Beclin-1, Atg7, Atg5, and LC3 in Tu212 CD133⁺ cells, and to a lesser degree in Tu212 CD133⁻ cells. Compared to Tu212 CD133⁻ cells, the levels of these autophagy markers were significantly higher in Tu212 CD133⁺ cells under hypoxic, low-glucose, and hypoxic+low-glucose conditions (Figs. 2a, b, S1). TEM showed that the number of autophagosomes in Tu212 CD133⁺ cells was significantly greater than that in Tu212 CD133⁻ cells under the above three conditions, indicating enhanced autophagy (Fig. 2c). Thus, the increased survival and migration of laryngeal carcinoma stem cells under hypoxic and low-glucose conditions may be associated with high GLUT-1 expression and autophagy.

Association between GLUT-1 expression and autophagy markers in CD133⁺ laryngeal carcinoma stem cells. Next, we silenced GLUT-1 and beclin-1 expression using shRNAs, inhibited autophagy using 3-MA and chloroquine, and activated autophagy using rapamycin in Tu212 CD133⁺ cells. qRT-PCR and Western blotting showed that the levels of GLUT-1, beclin-1, Atg7, and Atg5, and the LC3II/LC3I ratio in Tu212 CD133⁺ cells were higher than those in Tu212 CD133⁻ cells under hypoxic+low-glucose conditions. After transfection with the GLUT-1 shRNA, the Beclin-1, Atg7, and Atg5 mRNA levels and the LC3II/LC3I ratio in Tu212 CD133⁺ cells were significantly decreased. After transfection with the beclin-1 shRNA, the LC3II/LC3I ratio in Tu212 CD133⁺ cells was significantly decreased, but the expression of GLUT-1, ATG7, and ATG5 was unaffected. The autophagy inhibitor 3-MA significantly decreased the levels of beclin-1, Atg7, and Atg5, and the LC3II/LC3I ratio in Tu212 CD133⁺ cells, whereas GLUT-1 expression was unaffected. By contrast, CQ did not affect the expression of GLUT-1, beclin-1, Atg7, and Atg5 or the LC3II/LC3I ratio in Tu212 CD133⁺ cells. Rapamycin significantly increased the expression of beclin-1 and the LC3II/LC3I ratio in Tu212 CD133⁺ cells (Figs. 3a, b, S2). Therefore, the expression of GLUT-1 and autophagy markers is closely associated in stressed laryngeal carcinoma stem cells. TEM showed that silencing of GLUT-1 and beclin-1, or inhibition of autophagy, significantly decreased the number of autophagosomes in Tu212 CD133⁺ cells under hypoxic+low-glucose conditions. By contrast, activation of autophagy by rapamycin significantly increased the number of autophagosomes in Tu212 CD133⁺ cells (Fig. 3c).

GLUT-1 knockdown and autophagy inhibition reduce the proliferation and migration of CD133⁺ laryngeal carcinoma stem cells. Silencing of GLUT-1 markedly decreased clone formation, proliferation, and migration of Tu212 CD133⁺ cells under hypoxic- and low-glucose conditions (Fig. 4). By contrast, the apoptotic rate of Tu212 CD133⁺ cells was significantly increased by the silencing of GLUT-1 (Fig. 4).

Similarly, beclin-1 silencing or autophagy inhibition (3-MA, CQ) significantly decreased the above malignant behaviors of CD133⁺ cells. Importantly, the survival and migration of CD133⁺ cells were reduced by GLUT-1 silencing or autophagy inhibition. Rapamycin also reduced the malignant behaviors of CD133⁺ cells, suggesting that excessive autophagy harms stem cells (Fig. 4). Therefore, enhanced glucose uptake and autophagy are responsible for maintaining the growth and migration of stressed laryngeal carcinoma stem cells.

Discussion

Glucose is the main energy source for tumor cells and GLUT-1 is a key transporter of extracellular glucose [6,7, 29]. GLUT-1 is overexpressed in many cancers, including laryngeal carcinoma, and is associated with their metastasis, drug resistance, and prognosis [6,7,30–36]. We investigated the role of GLUT-1 in Tu212 CD133⁺ laryngeal carcinoma cells under hypoxic and low-glucose conditions.

A stressful extracellular microenvironment, such as hypoxia or low glucose, may upregulate GLUT-1 expression [37]. Indeed, hypoxia promotes the proliferation, migration, and chemoresistance of CSCs [4–7,14,15]. In this study, the growth and migration of CD133⁺ Tu212 cells were greater than those of CD133[−] Tu212 cells under hypoxic and low-glucose conditions; this was associated with increased mRNA and protein levels of GLUT-1 in CD133⁺ Tu212 cells. These findings are consistent with previous reports that high GLUT-1 expression facilitates glucose uptake to meet the energy demand of cancer cells. This adaptive response enables cancer cells to overcome external stresses, such as hypoxia or nutrient deprivation [6,38], thereby suppressing apoptosis.

The levels of autophagy markers in Tu212 CD133⁺ cells were increased by hypoxia and low glucose, whereas GLUT-1 silencing reduced the levels of these proteins. These results suggest mutual regulation of glucose uptake and autophagy in stressed laryngeal carcinoma stem cells. Autophagy modulates various metabolic pathways, including that centered on glucose [23]. Hypoxia and glucose deprivation may induce autophagy, promoting glucose uptake by upregulating GLUT-1 expression to increase the glycolytic flux and maintain nutrient uptake under stress conditions [21–23,40,41]. In airway progenitor cells, a lack of GLUT-1 impacts its recycling but not its expression, facilitating glucose uptake [40]. In mouse embryonic fibroblasts, however, autophagy enhances glucose uptake by increasing GLUT-1 expression and promoting GLUT-1 trafficking [21]. In this study, silencing of GLUT-1 decreased the levels of the autophagy markers beclin-1, Atg7, and Atg5, as well as the LC3II/LC3I ratio. However, inhibition or activation of autophagy by the beclin-1 shRNA/3-MA/CQ or rapamycin did not affect GLUT-1 expression. By contrast, rapamycin-induced autophagy activation increased the frequency of apoptosis of laryngeal CSCs, consistent with reports that excessive autophagy induces cell death.

This study had some limitations. First, we did not assay glucose metabolism, instead using GLUT-1 as a surrogate for glucose uptake. Second, we did not explore the *in vivo* implications of our findings using animal models. Third, the signaling mechanisms responsible for the enhanced GLUT-1 expression and

autophagy in laryngeal carcinoma stem cells under hypoxic and low-glucose conditions warrant further investigation.

Conclusions

In summary, hypoxia and low glucose increased the growth and migration of CD133⁺ laryngeal carcinoma stem cells by enhancing the expression of GLUT-1 and activating autophagy.

Abbreviations

3-MA: 3-Methyladenine; GLUT-1: glucose transporter-1; CSCs: cancer stem cells; HNSCC: head and neck squamous cell carcinoma; ATG: autophagy-related proteins

Declarations

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Availability of data and materials

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Authors' contributions

XH C designed the study and wrote the manuscript. JL, JT Z and SH Z reviewed the literatures and analyzed results. SH Z designed the study and revised the manuscript. J F conducted the experiments. All authors reviewed the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

This study was conducted under the guidelines and with the approval of Second Hospital of Jiaxin City, Jiaxin City, Zhejiang Province, 314000, China (No. jxey-201003).

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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Figures

Figure 1

Proliferation, migration, and apoptosis of stressed CD133+ laryngeal carcinoma stem cells. (a) CD133+ Tu212 cells were isolated and the percentages of CD133+ cells before and after purification were determined by flow cytometry. (b-e) Tu212 cells were subjected to hypoxia or low glucose, singly or in combination. Proliferation as evaluated using a colony-formation assay (b) and a CCK-8 assay (c). (d) Apoptosis as assessed by Annexin V-PI staining and flow cytometry. (e) Cell migration as determined using a Transwell assay. Data are means $\pm$ SDs and are representative of at least three independent experiments. * $P < 0.05$; ** $P < 0.01$; two-tailed unpaired Student's t-test.

Figure 2

GLUT-1 and autophagy marker expression in stressed CD133⁺ laryngeal carcinoma stem cells. Tu212 cells were challenged with hypoxia or low glucose, singly or in combination. (a, b) mRNA and protein levels of GLUT-1, beclin-1, Atg7, Atg5, and LC3 as evaluated by qRT-PCR (a) and Western blotting (b). (c) Autophagy as examined by TEM. Data are means $\pm$ SDs and are representative of at least three independent experiments. * $P < 0.05$; ** $P < 0.01$; two-tailed unpaired Student's t-test.

Figure 3

Relationship between GLUT-1 expression and autophagy in stressed CD133⁺ laryngeal carcinoma stem cells. Tu212 cells were transfected with GLUT-1 and beclin-1 siRNAs or treated with an autophagy inhibitor (3-MA, CQ) or activator (rapamycin). Then the cells were challenged with hypoxia or low glucose singly (a-c) or in combination (d). mRNA and protein levels of GLUT-1, beclin-1, Atg7, Atg5, and LC3 as evaluated by qRT-PCR (a) and Western blotting (b). (c) Autophagy as examined by TEM. Data are means $\pm$ SDs and are representative of at least three independent experiments. * $P < 0.05$; ** $P < 0.01$; two-tailed unpaired Student's t-test.

Figure 4

Effects of GLUT-1 knockdown or autophagy modulation on the malignant behaviors of stressed CD133⁺ laryngeal carcinoma stem cells. Tu212 cells were transfected with GLUT-1 and beclin-1 siRNAs or treated with an autophagy inhibitor (3-MA, CQ) or activator (rapamycin). Then the cells were subjected to hypoxia plus low glucose. (a, b) Proliferation as evaluated using a colony-formation assay (a) and a CCK-8 assay (b). (c) Apoptosis as evaluated by Annexin V-PI staining with flow cytometry. (d) Cell migration as determined using a Transwell assay. Data are means $\pm$ SDs and are representative of at least three independent experiments. * $P < 0.05$; ** $P < 0.01$; two-tailed unpaired Student's t-test.

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