

Endocrine disruptors concentrations in drinking water samples from México and their health implications

Karla Ximena Vargas-Berrones

Higher Technological Institute of Rioverde: Instituto Tecnologico Superior de Rioverde

Juan Manuel Izar-Landeta

Higher Technological Institute of Rioverde: Instituto Tecnologico Superior de Rioverde

Luis Armando Bernal-Jácome

Universidad Autónoma de San Luis Potosí: Universidad Autonoma de San Luis Potosi

Jennifer Iridian Sánchez-García

Universidad Autónoma de San Luis Potosí: Universidad Autonoma de San Luis Potosi

Rogelio Flores-Ramírez (rfloresra@conacyt.mx)

Universidad Autonoma de San Luis Potosi https://orcid.org/0000-0003-2263-6280

Research Article

Keywords: drinking water, nonylphenol, surfactant, emerging pollutant, Mexico, gas chromatographymass spectrometry

Posted Date: January 18th, 2023

DOI: https://doi.org/10.21203/rs.3.rs-2394953/v1

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Abstract

Nonylphenol ethoxylate, used mainly in detergent production, is transformed under environmental conditions into the endocrine disruptor, Nonylphenol (NP). 4-Nonylphenol (4-NP) was identified in drinking water samples from a developing country without regulations (Mexico) to establish exposure and environmental concentrations. The extraction and quantification of 4-NP were performed using solid phase microextraction (SPME) combined with gas chromatography-mass spectrometry (GC-MS). A derivatization process was carried out to increase sensitivity in the method. Eighty percent of the samples showed concentrations above the detection limit, and 57% of the samples presented concentrations above the Directive on the Quality of Water intended for human consumption (0.3 µg L-1). Our data gives an overview of the exposure levels and the environmental and health risks that these may represent. According to the results, continuous monitoring and regulations of this pollutant are highly recommended to prevent exposure and ecological and health effects.

1. Introduction

Water guality research has focused on nutrients, microbial contaminants, heavy metals, and priority pollutants. Recently, a new type of pollutants (emerging pollutants) has been recognized (EPA 2005). Currently, more than 1036 emerging pollutants are listed in the European Aquatic Environment NORMAN Network (www.norman-network.net) significantly affecting water quality and causing potential public health and security problems (Bilal et al. 2019). However, due to its recent detection and low concentrations (µg L⁻¹, ng L⁻¹), there is a gap in the knowledge about its occurrence, fate, behavior, risk assessment, and ecological and human effects (Vargas-Berrones et al. 2020a). In addition, population growth has resulted in major environmental impacts, being water bodies the most affected in terms of availability and quality (Peña-Guzmán et al. 2019). One of the most reported organic pollutants in wastewater, effluents, rivers, drinking water, sediments, and soil are the Alkylphenol Ethoxylates (APEs) (Belmont et al. 2006, Chen et al. 2013, Dong et al. 2015, Jie et al. 2017, Van Zijl et al. 2017). APEs are non-ionic surfactants widely used in detergent manufacturing, plastic additives, emulsifiers, and pesticides (Ferrara et al. 2011). Above 95% of APEs are removed from wastewaters with conventional treatment plants processes; however, the main problem is the formation of resistant metabolites classified as endocrine disruptors (Petrović et al. 2003). The most used APE is the nonylphenol ethoxylated (NPE) due to its capacity to form micelles in solution (Araujo et al. 2018). After their disposal, NPE is degraded by microorganisms or ultraviolet light under environmental conditions transforming them into nonylphenol (NP), including mainly 4-nonylphenol (4-NP) (Cheng et al. 2017). NP is classified as an endocrine disruptor which are external agents that interfere with the formation, elimination, transport, attachment, activity, or displacement of natural hormones that maintain homeostasis development, reproduction, and behavior (EPA 1997). Endocrine disruptors are hardly reviewed and regulated, so there is scarce information regarding their occurrence, fate, and health impacts (Gavrilescu et al. 2015). Therefore, NP has been included as a priority pollutant by some authorities. The Water Framework Directive of the European Union allows maximum concentrations in surface water of 2 µg L⁻¹

(European Union 2013), and the Environmental Protection Agency (EPA) of the United States establishes a maximum concentration of 6.6 µg L⁻¹ in surface water (EPA 2005). Also, regulations to restrict the use of NPE in industries have been introduced. For example, the European Union (EU) with the Directive 2003/53/EC establishes that NPE "...may not be placed on the market or used as a substance or constituent of preparations in concentrations equal or higher than 0,1 % by mass..." (Union 2003), and Directive 775/2004(02/2076) prohibits the use of NPE in pesticides formulations (Union 2006). On the other hand, the EPA added NPE to the Toxics Release Inventory (TRI) list and meets the toxicity listing criteria of the EPA's Emergency Planning and Community Right-to-know Act (EPCRA) section 313(d)(2) (C) indicating that NPE is highly toxic to aquatic organisms (EPA 2018). However, regulations in developing countries that restrict levels of NP in water are null, and their health and environmental effects are not well understood (Shannon et al. 2008).

The main exposure pathway of NP is through food and water intake, which leads to bioaccumulation and biomagnification. Effects of NP exposure in animals have been previously reported in concentrations from 1 to >195 µg L⁻¹ (Lussier et al. 2000, Scaia et al. 2019, Tabassum et al. 2017); however, effects in humans are still debated and require more investigation. Some studies have suggested potential health effects like decreased sperm count, reproductive malformations, immune deficiency, an increase in prostate, breast, ovarian and testicular cancer, neurological effects, poor intellect development in children, and psychological effects being the more vulnerable population fetuses and newborns (Bolong et al. 2009, Lussier et al. 2000, Tijani et al. 2016). The predicted no-effect concentration (PNEC) of NP has not been established because there is not enough toxicity data, and the specific mechanism in organisms is still unclear (Bakke 2003). Therefore, the World Health Organization (WHO) suggests a maximum concentration of 0.3 μ g L⁻¹ in drinking water based on the precautionary values that comply with existing environmental quality standards (WHO 2017). There is a current concern about the future and transport of NP through the environment and humans because of the continuous detection and identification of NP in water sources. Thus, the objective of this study was to monitor the concentrations of 4-NP in drinking water from countries without regulation (Mexico) to demonstrate potential human health risks from water intake (Figure 1). It is paramount to implement monitoring strategies in water to contribute to the generation of regulatory framework in developing countries due to the toxicity associated with NP and the extensive use of NPEs.

2. Materials And Methods

2.1 Reagents and chemicals

Stock standard solution of 4-NP (1000 µg L-1) was prepared in acetone and stored in the dark at -40°C until further analysis. SPME fiber DVB/CAR/PDMS (50/30 µm stableflex divinylbenzene/carboxen/polydimethylsiloxane) was supplied by Supelco (Edo. Mexico) and it was conditioned before its use following the manufacturer's instructions. A 0.1 M hydrochloric acid (HCl) solution (JT Baker, Edo. Mexico), sodium chloride (NaCl) (JT Baker, Edo Mexico), N-Methyl-

bis(trifluoroacetamide) (MBTFA) \geq 97.0% GC (Sigma Aldrich, Edo. Mexico), and Milli-Q deionized water (18.3 M Ω , Millipore) were used for the derivatization process. Samples were analyzed in a gas chromatograph (GC) (Agilent 6890) coupled to a mass spectrometry detector (MS) (Agilent 5975) in electron impact ionization mode (EI). The injection port was operated in splitless mode with a 0.75 mm liner without glass wool. GC separation was performed on a HP 5MS (60 m x 0.25 mm x 0.25 µm) column (Agilent). Helium, used as carrier gas, was controlled at a flow rate of 1 mL min-1 and 36 psi. The injection port temperature was set at 230°C, and the oven temperature program used was as follows: 90°C (2 min), 180°C (30°C min-1), 200°C (1°C min-1), 230°C (30°C min-1) and held for 5 min with a run time of 31 min. The tune parameters were: emission: 34.6; energy: 69.9; repeller: 26.6 and EMVolts: 1341. The SCAN mode (50–500 m/z) was used to identify the compound. The identification and quantification ions were selected for SIM mode (203/316 m/z). Results were obtained and processed with Chemstation Software (Agilent).

2.2 Sampling preparation

Directed monitoring of drinking water samples was performed. Since tap water is not potable in Mexico, water samples were collected from jugs of different water purifiers in Mexico. One-liter plastic bottles previously rinsed with Milli-Q water were used. No detergent was applied to prevent contamination. Immediately after sampling, they were stored in the dark at -20°C until further analysis.

2.3 4-NP quantification

4-NP quantification was performed based on the methodology described by Vargas-Berrones et al (Vargas-Berrones et al. 2020b). The lineal range ($r^2 = 0.99$) of the method was from 0.5 to 50 µg L⁻¹, the detection (LOD) and quantification (LOQ) limits were determined by blank signal and the obtained values were 0.01 µg L⁻¹ and 0.15 µg L⁻¹, respectively. One milliliter of water sample, 20 µL of HCl (0.1 M), and NaCl (3%) were added to a 10 mL sealed amber vial with gentle agitation. HCl and NaCl were used to adjust pH and enhance ionic strength, respectively. Solid phase microextraction (SPME) was carried out with a DVB/CAR/PDMS (50/30 µm stable flex divinylbenzene/carboxen/polydimethylsiloxane) sorbent. The fiber was exposed to the headspace at 80°C for 20 min magnetically stirred at 600 rpm. After the extraction of 4-NP, a derivatization process with N-Methyl-bis(trifluoroacetamide) (MBTFA) was carried out to improve the volatility and sensitivity of the method. This consisted of exposing the fiber to the headspace of a solution of acetone (1 mL) with MBTFA (100 µL) at 60°C for 10 min magnetically stirred at 600 rpm. After derivatization, samples were analyzed by gas chromatography (GC) (Agilent 6890) coupled to a mass spectrometry detector (MS) (Agilent 5975) in electron impact ionization mode (EI).

3. Results And Discussion

Concentrations in drinking water from different water purifiers are shown in Table 1.

Sample	Concentration		Concentration	Sample	Concentration
1	1.33	21	0.47	41	3.05
2	<lod< td=""><td>22</td><td>0.89</td><td>42</td><td>3.42</td></lod<>	22	0.89	42	3.42
3	2.32	23	0.19	43	0.79
4	2.64	24	0.10	44	1.50
5	6.08	25	0.12	45	2.48
6	40.29	26	0.40	46	1.42
7	3.99	27	0.13	47	0.85
8	<lod< td=""><td>28</td><td>0.37</td><td>48</td><td>4.62</td></lod<>	28	0.37	48	4.62
9	<lod< td=""><td>29</td><td>3.66</td><td>49</td><td>0.40</td></lod<>	29	3.66	49	0.40
10	<lod< td=""><td>30</td><td>0.14</td><td>50</td><td>0.09</td></lod<>	30	0.14	50	0.09
11	<lod< td=""><td>31</td><td>0.11</td><td>51</td><td>0.22</td></lod<>	31	0.11	51	0.22
12	<lod< td=""><td>32</td><td>2.99</td><td>52</td><td>2.17</td></lod<>	32	2.99	52	2.17
13	<lod< td=""><td>33</td><td>0.91</td><td>53</td><td>0.84</td></lod<>	33	0.91	53	0.84
14	<lod< td=""><td>34</td><td>0.20</td><td>54</td><td>0.95</td></lod<>	34	0.20	54	0.95
15	0.85	35	0.23	55	0.31
16	<lod< td=""><td>36</td><td>0.94</td><td>56</td><td>4.19</td></lod<>	36	0.94	56	4.19
17	<lod< td=""><td>37</td><td>3.34</td><td>57</td><td>0.25</td></lod<>	37	3.34	57	0.25
18	<lod< td=""><td>38</td><td>1.83</td><td>58</td><td>0.17</td></lod<>	38	1.83	58	0.17
19	<lod< td=""><td>39</td><td>6.43</td><td>59</td><td>0.16</td></lod<>	39	6.43	59	0.16
20	15.29	40	1.82	60	0.09
*Units: μg L ⁻¹ ; LOD: 0.01 μg L ⁻¹					

Table 1 Concentrations of 4-NP in different samples of drinking water from water purifiers of countries without regulation (Mexico).

Values above the LOD were detected in 80% of the collected samples, and 57% of the samples presented concentrations above the Directive on the quality of water intended for human consumption (0.3 μ g L⁻¹) (WHO 2017). Reported values are similar to previous studies **(**Table 2**)**.

Table 2	
Concentrations of 4-NP in drinking water from different countries	

	Country	n	Min	Median	Max	Reference
Drinking water	Mexico	5	<lod< td=""><td>2.48</td><td>6.08</td><td>(Vargas-Berrones et al., 2020b)</td></lod<>	2.48	6.08	(Vargas-Berrones et al., 2020b)
	China	15	0.01	0.05	2.7	(Shao et al., 2005)
	Czech Republic	6	0.029	0.0335	0.045	(Pernica et al., 2015)
	China	62	ND	0.027	0.558	(Fan et al., 2013a)
	Italy	35	< 0.0077	0.0149	0.084	(Maggioni et al., 2013)
	China	21	0.108	0.170	0.298	(Li et al., 2010)
	China	6	0.196	0.502	1.073	(Li et al., 2010)
	France	8	<loq< td=""><td>0.0159</td><td>0.0594</td><td>(Dupuis et al., 2012)</td></loq<>	0.0159	0.0594	(Dupuis et al., 2012)
	China	8	0.0082	0.577 (media)	0.918	(Jie et al., 2017)
	Taiwan	18	0.017	0.032 (media)	0.195	(Cheng et al., 2016)
	Japan	9	0.016	0.076	0.078	(Toyo et al., 2000)
	Greece	6	NR	NR	0.15	(Amiridou and Voutsa, 2011)
	United States	18	NR	100	93	(Benotti et al., 2009)
	United States	12	NR	NR	1.1	(Stackelberg et al., 2007
	Germany	10	0.0025	0.0066	0.016	(Kuch and Ballschmiter, 2001)

Occurrence of NP has increased significantly because of its great industrial demand (Silva et al. 2018). It is difficult to determine the source of contamination in water samples; however, the most common use of NPE is in detergents (Kim et al. 2019, Priac et al. 2017). For example, 41% of 90 domestic detergents in Taiwan contained from 0.2 to 21% of NP (Huang et al. 2014). This suggests that the presence of NP in water is mainly due to the use of detergents, tourism, people washing clothes, illegal water discharges in rivers, and inadequacy and lack of maintenance in drainage networks (Fenet et al. 2003, Gambolati et al. 2006). In countries where NP and its ethoxylates are not regulated, like Mexico, it is common to find these compounds in detergents due to their excellent surfactant properties (Merrettig-Bruns &Jelen 2009) and low cost (Perron &Juneau 2011). In this country, water jugs are washed each time before being refilled.

So, a poor rinse may explain the high concentrations of NP found in drinking water. Also, conventional water purification processes do not remove endocrine disruptors like NP (Van Zijl et al. 2017), and NPE may be degraded into shorter ethoxylated chains in the treatment processes (Soares et al. 2008). Furthermore, the chlorination process in water supply systems forms byproducts like monochloro-NP (CNP) and dichloro-NP (DCNP). These compounds have previously demonstrated estrogenic activity (Fan et al. 2013, Takemura et al. 2005).

Previous studies have demonstrated adverse effects in biota (Table 3) and humans (Table 4). However, more research in this regard is paramount to understanding the risks and effects due to the exposure of this xenobiotic. A risk assessment regarding NP exposure through water intake could be performed as future work with these results. The estimation of the non-carcinogenic risk obtained by the hazard quotation (HQ) for water intake would indicate the potential risks of adverse health effects according to the values established by the EPA (0.1 mg kg⁻¹ day⁻¹) (Bakke 2003). Risk assessment regarding NP exposure has been previously reported in sludge (González et al. 2010, Kollmann et al. 2003, Roberts et al. 2006), surface water, wastewaters (Chen et al. 2014, Gao et al. 2014, Jin et al. 2014), and aquatic organisms (Lee et al. 2015, Pachura-Bouchet et al. 2006, Servos et al. 2003). Nevertheless, limited information about risk assessment in humans is available because of high analysis costs and the lack of scientific data in this area (Tijani et al. 2016). The United Nations Environment Programme has established that the highest estimated value for human exposure through the environment is 5.31×10^{-3} mg kg⁻¹day⁻¹ and the maximum intake combined from the air, water, and food exposure is 6.4 mg kg⁻¹ ¹day⁻¹. However, there is uncertainty in the daily intake estimated making it difficult to determine accurate predictions in this regard (Bontje D. 2002). These values have only been considered in adults though children are more vulnerable to hormone impacts caused by environmental xenobiotics. This vulnerability is associated with physiological differences like constant increase in weight, higher respiration and ventilation range, higher relative consumption of water and food, and faster brain development (Longnecker et al. 2003, McElroy 2008, Mishra & Vankar 2002, Norgil Damgaard et al. 2002, Selevan et al. 2000). Previous studies have shown that NP is ubiquitous in baby food representing a daily intake from 0.23 to 0.65 μ g kg⁻¹ bw d⁻¹ (Raecker et al. 2011). However, low concentrations in humans may have virtually no chance to compete with natural hormones in the unions of free receptors, implying that the health risks of endocrine disruptors may be insignificant (Autrup et al. 2020). Therefore, exposure effects of endocrine disruptors at low doses during the development of humans have been underestimated (Welshons et al. 2006). A greater potential risk for infants and babies is expected due to the higher vulnerability to hormonal effects and their higher relative NP consumption through food and water compared to adults; also, a reference dose for infants has not been established yet.

Organism	Specie	Compound	Concentration	Effects	Reference
Plants	Vigna radiata	NP	1000 mg/kg	Leaf vein necrosis	(Kim 2019)
Fish	Orechromis niloticus	NP	16 µg/L	Alterations in the female gonads	(Rivero 2008)
	Bream and black rockfish	4-NP	50 μg /L	Reduced gonadosomatic index	(Saravanan 2019)
	Oncorhynchus mykiss	NP	1 µg /L	Alterations in the immune system	(Hébert 2009)
	Xiphophorus maculatus	NP	>0.96 mg/L	Negative effects on testicular morphology and male fertility	(Kinnberg 2000)
Oyster	Crassostrea gigas	4-NP NP	0.1–10 µg /L	Development of abnormalities	(Nice HE 2000)
Oyster	Crassostrea gigas	4-NP NP	1-100 µg /L	Increased incidence of hermaphroditism	(Nice HE 2003)
Crustacean	Elminus modestus	4-NP	0.1–10 µg /L	DNA adduct formation, mutations and genomic rearrangements	(Atiezar FA 2002)
Amphibians	Xenopus laevis	NP	0.1–10 µg /L	Increased mortality, morphological deformations, and increased apoptosis	(Bevan CL 2003)
Mice	Ratones	NP	50-500 μg /L	Negative effects on spermatogenesis and sperm quality	(Kyselova V 2003)

Table 3 NP and 4-NP concentrations in different species.

Table 4
Health effects in human by NP and/or 4-NP exposure

Sample	Analyte	Concentration	Exposure	Effects	Reference
Semen	NP	<7 pg/mL	Normal food and water intake	No significant exposure	(Katayama et al. 2003)
Urine	4-NP	< 110 ng/mL	Normal food and water intake	Not available	(Inoue et al. 2003)
Human milk	NP	< 56.3 ng/mL	Normal food and water intake	No significant exposure	(Ademollo et al. 2008)
Human breast carcinoma cell line (MCF-7)	NPEOs	NA	100-200 μΜ	γ-H2AX generation caused by direct chemically induced DNA damage	(Zhao et al. 2015)
Human breast carcinoma cell line (MCF-7)	NPEOs	NA	100-500 μΜ	Generation of γ-H2AX means the formation of DSBs (DNA damage)	(Toyooka et al. 2012)
CHO-K1 cells	NP	NA	0.025- 0.1 μM	DNA damage (sister- chromatid exchange)	(Tayama et al. 2008)
Jurkat cells	4-NP	NA	Not specified	Induced loss of mitochondrial membrane potential, caspase-8 activation, and internucleosomal DNA fragmentation	(Yao et al. 2007)
Jurkat cells	NP	NA	9.72− 38.9 µM	Induced DNA damage	(Park &Choi 2007)
<i>Saccharomyces cerevisiae</i> cells	4-NP	NA	50 mg/L	Induced significant cytotoxic effect	(Frassinetti et al. 2011)
Spermatozoa	NP	NA	5 µL	Oxidative stress and DNA damage	(Bennetts et al. 2008)

Some limitations were considered when interpreting the results of this study. 1) Only 4-NP was considered because of its commercial availability (Calafat et al. 2005). It is also important to acknowledge that NP is a mixture of approximately 20 para-substituted isomers with different branched alkyl chains and with intermediate structures compounds that make the mixture more toxic (leda et al.

2005, Ruß et al. 2005, Thiele et al. 2004, Wheeler et al. 1997). Previous studies have reported that 4-NP only represents 2.2% of the total mass in water, and represents only 26% of all the risk (Fenner et al. 2002). Moreover, NP isomers have relevant differences among them regarding their disruptive endocrine activity. Therefore, it is not adequate to take any isomer as a general reference to establish models, activities, structure relations, and/or risk assessments (Preuss et al. 2006). Thus, to achieve a complete analysis, it would be necessary to examine all isomers in the sample. Even though our study only considers 4-NP, it provides a reference point and allows to establish minimum exposure concentrations. 2) Other pollutants classified as endocrine disruptors may be found in drinking water. Consequently, it may be practically impossible to predict health problems caused by each compound since they can act independently or in synergy with others. For example, Bisphenol A (BPA) is a monomer used for polycarbonates and epoxy resin production. This product is used as a coating for food cans, water containers, water pipes, reusable milk containers, food storage vessels, and baby bottles. Its incomplete polymerization during manufacture and the temperature increment through bottling processes may cause the compound to leach into food and beverages (Markey et al. 2001). Accordingly, phthalates (PEs) are endocrine disruptors mainly used as plasticizers and in paints, adhesives, dyes, and cosmetics manufacture. Usually, PEs are incorporated into food and water through packaging and manufacturing processes (Serodio & Nogueira 2006). Good practices and environmental policies are needed to mitigate potential risks to human and ecologic health established on the precautionary principle approach, based on evidence of potential harm without compelling absolute scientific certainty.

4. Conclusions

According to the literature, this is the first monitoring of 4-nonylphenol in Mexico in drinking water samples. This study aimed to provide a reference to establish minimum NP exposure concentrations through water intake. An exposure scenario of 4-NP in drinking water is shown in countries without regulations (Mexico). Our results demonstrate that 4-NP was detected in 80% of the water samples. Fifty seven percent of the samples presented concentrations above the Directive on the quality of water intended for human consumption. Although these results may not represent a significant risk for the consumption of NP through water intake, there is no consensus among the scientific community about this issue. Moreover, it is critical to acknowledge that current parameters are considered only for adults, and water intake in children may represent a greater risk. Efforts in investigation and strategies to promote consciousness of the impact of NP as a pollutant are required. These developments would help to establish a responsible approach regarding the use and handling of NP and its ethoxylates. The search for environmental quality to protect human and ecological health is a compulsory long-term challenge shared by most modern societies and civilizations worldwide.

Declarations

Acknowledgements

This work was supported by the National Council for Science and Technology – Initial Academic Postdoctoral Research CVU 815662 and CB-CONACYT (#2017-2018- A1-S-28176).

Ethical Approval

The paper reflects the authors' own research and analysis in a truthful and complete manner.

Consent to Participate and Publish

All persons who meet authorship criteria are listed as authors, and all authors consent to participate and publish in the work to take public responsibility for the content

Author Contributions

All persons who meet authorship criteria are listed as authors, and all authors certify that they have participated sufficiently in the work to take public responsibility for the content, including participation in the concept, design, analysis, writing, or revision of the manuscript. Conceptualization, Sampling, Analytical methods, Writing and manuscript editing by Karla Ximena Vargas-Berrones; Analytical methods by Juan Manuel Izar-Landeta, Luis Armando Bernal-Jácome, and Jennifer Iridian Sánchez-García; Analytical methods and Conceptualization, Writing- Reviewing, Editing and Funding by Rogelio Flores-Ramírez.

Funding

The authors acknowledge grants and fellowships from the National Council on Science and Technology-Sectoral Research Fund for Education Basic-Science # A1-S-28176 and Initial Academic Postdoctoral Research CVU 815662.

Disclosure of potential conflicts of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper

Availability of data and materials

Data available on request from the authors

References

- 1. Ademollo N, Ferrara F, Delise M, Fabietti F, Funari E (2008) Nonylphenol and octylphenol in human breast milk. Environ Int 34:984–987
- 2. Araujo FG, Bauerfeldt GF, Cid YP (2018) Nonylphenol: Properties, legislation, toxicity and determination. Anais da Academia Brasileira de Ciencias 90:1903–1918

- 3. Atiezar FABZ, Depledge MH (2002) 4-n-nonylphenol and 17- β estradiol may induce common DNA effects in developing barnacle larvae. Environ Pollut 120:735–738
- 4. Autrup H, Barile FA, Berry SC, Blaauboer BJ, Boobis A, Bolt H, Borgert CJ, Dekant W, Dietrich D, Domingo JL, Gori GB, Greim H, Hengstler J, Kacew S, Marquardt H, Pelkonen O, Savolainen K, Heslop-Harrison P, Vermeulen NP (2020) Human exposure to synthetic endocrine disrupting chemicals (S-EDCs) is generally negligible as compared to natural compounds with higher or comparable endocrine activity: how to evaluate the risk of the S-EDCs? Arch Toxicol 94:2549–2557
- 5. Bakke D (2003) : Human and Ecological Risk Assessment of Nonylphenol Polyethoxylate-based (NPE) Surfactants in Forest Service Herbicide Applications
- Belmont MA, Ikonomou M, Metcalfe CD (2006) Presence of nonylphenol ethoxylate surfactants in a watershed in central Mexico and removal from domestic sewage in a treatment wetland. Environ Toxicol Chem 25:29–35
- 7. Bennetts LE, De Iuliis GN, Nixon B, Kime M, Zelski K, McVicar CM, Lewis SE, Aitken RJ (2008) Impact of estrogenic compounds on DNA integrity in human spermatozoa: Evidence for cross-linking and redox cycling activities. Mutat Research/Fundamental Mol Mech Mutagen 641:1–11
- 8. Bevan CLPD, Prasad A, Howard M, Henderson LP (2003) Environmental estrogens alter early development in Xenopus laevis. Environ Heath Persp 111:488–496
- 9. Bilal M, Adeel M, Rasheed T, Zhao Y, Iqbal HMN (2019) Emerging contaminants of high concern and their enzyme-assisted biodegradation A review. Environ Int 124:336–353
- 10. Bolong N, Ismail AF, Salim MR, Matsuura T (2009) A review of the effects of emerging contaminants in wastewater and options for their removal. Desalination 239:229–246
- 11. Bontje DHJ, Vermeire T, Damstra T (2002) : INTEGRATED RISK ASSESSMENT: NONYLPHENOL CASE STUDY WHO/IPCS/IRA/12/04 In: SAFETY RPFTWUIIPOC (Hrsg.)
- Calafat AM, Kuklenyik Z, Reidy JA, Caudill SP, Ekong J, Needham LL (2005) Urinary concentrations of bisphenol A and 4-nonylphenol in a human reference population. Environ Health Perspect 113:391– 395
- 13. Chen HW, Liang CH, Wu ZM, Chang EE, Lin TF, Chiang PC, Wang GS (2013) Occurrence and assessment of treatment efficiency of nonylphenol, octylphenol and bisphenol-A in drinking water in Taiwan. Sci Total Environ 449:20–28
- 14. Chen R, Yin P, Zhao L, Yu Q, Hong A, Duan S (2014) Spatial-temporal distribution and potential ecological risk assessment of nonylphenol and octylphenol in riverine outlets of Pearl River Delta, China. J Environ Sci (China) 26:2340–2347
- 15. Cheng Y, Shan Z, Zhou J, Bu Y, Li P, Lu S (2017) Effects of 4-nonylphenol in drinking water on the reproductive capacity of Japanese quails (Coturnix japonica). Chemosphere 175:219–227
- 16. Dong C-D, Chen C-W, Chen C-F (2015) Seasonal and spatial distribution of 4-nonylphenol and 4-tertoctylphenol in the sediment of Kaohsiung Harbor. Taiwan Chemosphere 134:588–597
- 17. EPA (1997) : Special report on environmental endocrine disruption: an effects assessment and analysis. Office of Research and Development, Washington, DC. EPA/630/R-96/012

- 18. EPA (2005) : Aquatic life ambient water quality criteria-nonylphenol
- 19. EPA (2018) : Addition of NPEs Category to TRI List Final Rule
- 20. European Union (2013) : Directive 2013/39/EU of the European Parliament and the Council of 12. August 2013 Amending Directives 2000/60/EC and 2008/105/EC as Regards
- 21. Priority Substances in the Field of water policy
- 22. Fan Z, Hu J, An W, Yang M (2013) Detection and Occurrence of Chlorinated Byproducts of Bisphenol A, Nonylphenol, and Estrogens in Drinking Water of China: Comparison to the Parent Compounds. Environ Sci Technol 47:10841–10850
- 23. Fenet H, Gomez E, Pillon A, Rosain D, Nicolas JC, Casellas C, Balaguer P (2003) Estrogenic Activity in Water and Sediments of a French River: Contribution of Alkylphenols. Arch Environ Contam Toxicol 44:0001–0006
- 24. Fenner K, Kooijman C, Scheringer M, Hungerbühler K (2002) Including Transformation Products into the Risk Assessment for Chemicals: The Case of Nonylphenol Ethoxylate Usage in Switzerland. Environ Sci Technol 36:1147–1154
- 25. Ferrara F, Ademollo N, Orrù MA, Silvestroni L, Funari E (2011) Alkylphenols in adipose tissues of Italian population. Chemosphere 82:1044–1049
- 26. Frassinetti S, Barberio C, Caltavuturo L, Fava F, Di Gioia D (2011) Genotoxicity of 4-nonylphenol and nonylphenol ethoxylate mixtures by the use of Saccharomyces cerevisiae D7 mutation assay and use of this text to evaluate the efficiency of biodegradation treatments. Ecotoxicol Environ Saf 74:253–258
- 27. Gambolati G, Putti M, Teatini P, Gasparetto Stori G (2006) Subsidence due to peat oxidation and impact on drainage infrastructures in a farmland catchment south of the Venice Lagoon. Environ Geol 49:814–820
- 28. Gao P, Li Z, Gibson M, Gao H (2014) Ecological risk assessment of nonylphenol in coastal waters of China based on species sensitivity distribution model. Chemosphere 104:113–119
- 29. Gavrilescu M, Demnerova K, Aamand J, Agathos S, Fava F(2015) : Emerging pollutants in the environment: present and future challenges in biomonitoring, ecological risks and bioremediation. N Biotechnol 32, 147 56
- 30. González MM, Martín J, Santos JL, Aparicio I, Alonso E (2010) Occurrence and risk assessment of nonylphenol and nonylphenol ethoxylates in sewage sludge from different conventional treatment processes. Sci Total Environ 408:563–570
- 31. Hébert N, Gagné F, Cyr D, Pellerin J, Blaise C, Fournier M (2009) Effects of 4-nonylphenol on the immune system of rainbow trout (oncorhynchus mykiss). Fresenius Environ Bull 18:757–761
- 32. Huang YF, Wang PW, Huang LW, Yang W, Yu CJ, Yang SH, Chiu HH, Chen ML (2014) Nonylphenol in pregnant women and their matching fetuses: placental transfer and potential risks of infants. Environ Res 134:143–148

- 33. leda T, Horii Y, Petrick G, Yamashita N, Ochiai N, Kannan K (2005) Analysis of Nonylphenol Isomers in a Technical Mixture and in Water by Comprehensive Two-Dimensional Gas Chromatography – Mass Spectrometry. Environ Sci Technol 39:7202–7207
- 34. Inoue K, Kawaguchi M, Okada F, Takai N, Yoshimura Y, Horie M, Izumi S-i, Makino T, Nakazawa H (2003) Measurement of 4-nonylphenol and 4-tert-octylphenol in human urine by column-switching liquid chromatography–mass spectrometry. Anal Chim Acta 486:41–50
- 35. Jie Y, Jie Z, Ya L, Xuesong Y, Jing Y, Yu Y, Jiaqi Y, Jie X (2017) Pollution by Nonylphenol in river, tap water, and aquatic in an acid rain-plagued city in southwest China. Int J Environ Health Res 27:179–190
- 36. Jin X, Wang Y, Jin W, Rao K, Giesy JP, Hollert H, Richardson KL, Wang Z (2014) Ecological Risk of Nonylphenol in China Surface Waters Based on Reproductive Fitness. Environ Sci Technol 48:1256– 1262
- 37. Katayama M, Matsuda Y, Shimokawa K-I, Ishikawa H, Kaneko S (2003) Preliminary Monitoring of Bisphenol A and Nonylphenol in Human Semen by Sensitive High Performance Liquid Chromatography and Capillary Electrophoresis After Proteinase K Digestion. Anal Lett 36:2659–2667
- 38. Kim AR, Ha NR, Jung IP, Kim SH, Yoon MY(2019) : Development of a ssDNA aptamer system with reduced graphene oxide (rGO) to detect nonylphenol ethoxylate in domestic detergent. J Mol Recognit32, e2764
- 39. Kim D, Kwak JI, An Y-J (2019) Physiological response of crop plants to the endocrinedisrupting chemical nonylphenol in the soil environment. Environ Pollut 251:573–580
- 40. Kinnberg K, Korsgaard B, Bjerregaard P, Jespersen AS (2000) Effects of nonylphenol and 17betaestradiol on vitellogenin synthesis and testis morphology in male platyfish Xiphophorus maculatus. J Exp Biol 203:171–181
- 41. Kollmann A, Brault A, Touton I, Dubroca J, Chaplain V, Mougin C (2003) Effect of Nonylphenol Surfactants on Fungi following the Application of Sewage Sludge on Agricultural Soils. J Environ Qual 32:1269–1276
- 42. Kyselova VPJ, Buckiova D, Boubelik M (2003) Effects of p-n-nonylphenol and resveratrol on body and organ weight and in vivo fertility of outbred CD-I mice. Reproduct. Biol. Endocrinol
- 43. Lee CC, Jiang LY, Kuo YL, Chen CY, Hsieh CY, Hung CF, Tien CJ (2015) Characteristics of nonylphenol and bisphenol A accumulation by fish and implications for ecological and human health. Sci Total Environ 502:417–425
- 44. Longnecker MP, Bellinger DC, Crews D, Eskenazi B, Silbergeld EK, Woodruff TJ, Susser ES (2003) An approach to assessment of endocrine disruption in the National Children's Study. Environ Health Perspect 111:1691–1697
- 45. Lussier SM, Champlin D, LiVolsi J, Poucher S, Pruell RJ (2000) Acute toxicity of para-nonylphenol to saltwater animals. Environ Toxicol Chem 19:617–621
- 46. Markey CM, Michaelson CL, Sonnenschein C, Soto AM (2001) Alkylphenols and Bisphenol A as Environmental Estrogens. In: Metzler M (ed) Endocrine Disruptors – Part I. Springer Berlin Heidelberg,

Berlin, Heidelberg, pp 129-153

- 47. McElroy JA(2008) : Environmental Exposures and Child Health: What we Might Learn in the 21st Century from the National Children's Study?Environmental Health Insights2, EHI.S1061
- 48. Merrettig-Bruns U, Jelen E (2009) Anaerobic Biodegradation of Detergent Surfactants. Mater (Basel) 2:181–206
- 49. Mishra R, Vankar PS (2002) Pesticide residue analysis of infant formula in India. Bull Environ Contam Toxicol 69:667–673
- 50. Nice HEMD, Crane M, Thorndyle M (2003) Long-term and transgenerational effects of nonylphenol exposure at a key stage in the development of Crassostrea gigas Possible endocrine disruption? Mar. Ecol Prog Series 256:293–300
- 51. Nice HETM, Morrit D, Steele S, Crane M (2000) Development of Crassostrea gigas larvae is affected by 4-nonylphenol. Mar Pollut Bull 40:491–496
- 52. Norgil Damgaard I, Main KM, Toppari J, Skakkebaek NE (2002) Impact of exposure to endocrine disrupters in utero and in childhood on adult reproduction. Best practice & research. Clin Endocrinol metabolism 16:289–309
- 53. Pachura-Bouchet S, Blaise C, Vasseur P(2006) : Toxicity of nonylphenol on the cnidarian Hydra attenuata and environmental risk assessment. Environmental toxicology 21, 388 94
- 54. Park SY, Choi J (2007) Cytotoxicity, genotoxicity and ecotoxicity assay using human cell and environmental species for the screening of the risk from pollutant exposure. Environ Int 33:817–822
- 55. Peña-Guzmán C, Ulloa-Sánchez S, Mora K, Helena-Bustos R, Lopez-Barrera E, Alvarez J, Rodriguez-Pinzón M (2019) Emerging pollutants in the urban water cycle in Latin America: A review of the current literature. J Environ Manage 237:408–423
- 56. Perron MC, Juneau P (2011) Effect of endocrine disrupters on photosystem II energy fluxes of green algae and cyanobacteria. Environ Res 111:520–529
- 57. Petrović M, Gonzalez S, Barceló D (2003) Analysis and removal of emerging contaminants in wastewater and drinking water. TRAC Trends Anal Chem 22:685–696
- 58. Preuss TG, Gehrhardt J, Schirmer K, Coors A, Rubach M, Russ A, Jones PD, Giesy JP, Ratte HT (2006) Nonylphenol isomers differ in estrogenic activity. Environ Sci Technol 40:5147–5153
- 59. Priac A, Morin-Crini N, Druart C, Gavoille S, Bradu C, Lagarrigue C, Torri G, Winterton P, Crini G (2017) Alkylphenol and alkylphenol polyethoxylates in water and wastewater: A review of options for their elimination. Arab J Chem 10:S3749–S3773
- 60. Raecker T, Thiele B, Boehme RM, Guenther K (2011) Endocrine disrupting nonyl- and octylphenol in infant food in Germany: Considerable daily intake of nonylphenol for babies. Chemosphere 82:1533–1540
- 61. Rivero CLG, Barbosa AC, Ferreira MFN, Dorea JG, Grisolia CK (2008) Evaluation of genotoxicity and effects on reproduction of nonylphenol in Oreochromis niloticus (Pisces: cichlidae). Ecotoxicology, pp 732–737

- 62. Roberts P, Roberts JP, Jones DL (2006) Behaviour of the endocrine disrupting chemical nonylphenol in soil: Assessing the risk associated with spreading contaminated waste to land. Soil Biol Biochem 38:1812–1822
- 63. Ruß AS, Vinken R, Schuphan I, Schmidt B (2005) Synthesis of branched para-nonylphenol isomers: Occurrence and quantification in two commercial mixtures. Chemosphere 60:1624–1635
- 64. Saravanan M, Nam S-E, Eom H-J, Lee D-H, Rhee J-S (2019) Long-term exposure to waterborne nonylphenol alters reproductive physiological parameters economically important marine fish. Comp Biochem Physiol C: Toxicol Pharmacol 216:10–18
- 65. Scaia MF, de Gregorio LS, Franco-Belussi L, Succi-Domingues M, de Oliveira C (2019) Gonadal, body color, and genotoxic alterations in Lithobates catesbeianus tadpoles exposed to nonylphenol. Environ Sci Pollut Res 26:22209–22219
- 66. Selevan SG, Kimmel CA, Mendola P (2000) Identifying critical windows of exposure for children's health. Environ Health Perspect 108(Suppl 3):451–455
- 67. Serodio P, Nogueira JM (2006) Considerations on ultra-trace analysis of phthalates in drinking water. Water Res 40:2572–2582
- 68. Servos MR, Maguire RJ, Bennie DT, Lee H-B, Cureton PM, Davidson N, Sutcliffe R, Rawn DFK(2003) : An Ecological Risk Assessment of Nonylphenol and Its Ethoxylates in the Aquatic Environment. Human and Ecological Risk Assessment: An International Journal 9, 569–587
- 69. Shannon MA, Bohn PW, Elimelech M, Georgiadis JG, Mariñas BJ, Mayes AM (2008) Science and technology for water purification in the coming decades. Nature 452:301–310
- 70. Silva A, de Oliveira CD, Quirino A, Silva F, Saraiva R, Silva-Cavalcanti J (2018) Endocrine Disruptors in Aquatic Environment: Effects and Consequences on the Biodiversity of Fish and Amphibian Species. Aquat Sci Technol 6:35
- 71. Soares A, Guieysse B, Jefferson B, Cartmell E, Lester JN (2008) Nonylphenol in the environment: a critical review on occurrence, fate, toxicity and treatment in wastewaters. Environ Int 34:1033–1049
- 72. Tabassum H, Ashafaq M, Parvez S, Raisuddin S (2017) Role of melatonin in mitigating nonylphenolinduced toxicity in frontal cortex and hippocampus of rat brain. Neurochem Int 104:11–26
- 73. Takemura H, Ma J, Sayama K, Terao Y, Zhu BT, Shimoi K (2005) In vitro and in vivo estrogenic activity of chlorinated derivatives of bisphenol A. Toxicology 207:215–221
- 74. Tayama S, Nakagawa Y, Tayama K (2008) Genotoxic effects of environmental estrogen-like compounds in CHO-K1 cells. Mutat Research/Genetic Toxicol Environ Mutagen 649:114–125
- 75. Thiele B, Heinke V, Kleist E, Guenther K (2004) Contribution to the Structural Elucidation of 10 Isomers of Technical p-Nonylphenol. Environ Sci Technol 38:3405–3411
- 76. Tijani JO, Fatoba OO, Babajide OO, Petrik LF (2016) Pharmaceuticals, endocrine disruptors, personal care products, nanomaterials and perfluorinated pollutants: a review. Environ Chem Lett 14:27–49
- 77. Toyooka T, Kubota T, Ibuki Y (2012) Nonylphenol polyethoxylates induce phosphorylation of histone H2AX. Mutat Research/Genetic Toxicol Environ Mutagen 741:57–64

- 78. Union E(2003) : DIRECTIVE 2003/53/EC June 2003 amending for the 26th time Council Directive 76/769/EEC relating to restrictions on the marketing and use of certain dangerous substances and preparations (nonylphenol, nonylphenol ethoxylate and cement)
- 79. Union E(2006) : Directive 775/2004 (02/2076)
- 80. Van Zijl MC, Aneck-Hahn NH, Swart P, Hayward S, Genthe B, De Jager C (2017) Estrogenic activity, chemical levels and health risk assessment of municipal distribution point water from Pretoria and Cape Town, South Africa. Chemosphere 186:305–313
- 81. Vargas-Berrones K, Bernal-Jácome L, Díaz de León-Martínez L, Flores-Ramírez R (2020a) Emerging pollutants (EPs) in Latin América: A critical review of under-studied EPs, case of study -Nonylphenol. Sci Total Environ 726:138493
- 82. Vargas-Berrones K, Díaz de León-Martínez L, Bernal-Jácome L, Rodriguez-Aguilar M, Ávila-Galarza A, Flores-Ramírez R (2020b) Rapid analysis of 4-nonylphenol by solid phase microextraction in water samples. Talanta 209:120546
- 83. Welshons WV, Nagel SC, vom Saal FS (2006) Large Effects from Small Exposures. III. Endocrine Mechanisms Mediating Effects of Bisphenol A at Levels of Human Exposure. Endocrinology 147:s56-s69
- 84. Wheeler TF, Heim JR, LaTorre MR, Janes AB (1997) Mass Spectral Characterization of p-Nonylphenol Isomers Using High-Resolution Capillary GC–MS. J Chromatogr Sci 35:19–30
- 85. WHO (2017) :Drinking Water Parameter Cooperation Project
- 86. Yao G, Ling L, Luan J, Ye D, Zhu P (2007) Nonylphenol induces apoptosis of Jurkat cells by a caspase-8 dependent mechanism. Int Immunopharmacol 7:444–453
- 87. Zhao X, Yang G, Toyooka T, Ibuki Y (2015) New mechanism of γ-H2AX generation: Surfactantinduced actin disruption causes deoxyribonuclease I translocation to the nucleus and forms DNA double-strand breaks. Mutat Res Genetic Toxicol Environ Mutagen 794:1–7

Figures

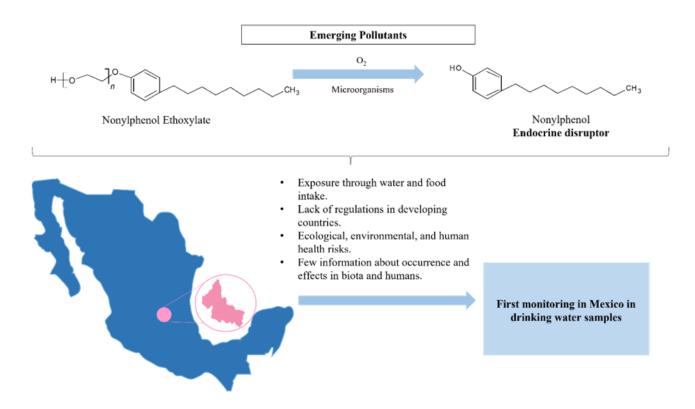


Figure 1

Context of nonylphenol analysis in drinking water samples to determine potential exposure risks through water intake in developing countries.