

Differentially expressed genes SNRPC and PRPF38A are potential biomarkers candidates for osteosarcoma

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Research

Keywords: PRPF38A, SNRPC, Osteosarcoma, GEO, DEGs

Posted Date: April 28th, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-24106/v1>

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2 **potential biomarkers candidates for osteosarcoma**

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11
12 **Abstract**

13 **Background:** Osteosarcoma (osteogenic sarcoma, OS) is a primary cause of morbidity
14 and mortality and is associated with poor prognosis in the field of orthopedic. Globally,
15 rates of OS are highest among 15 to 25-year-old adolescent. However, the mechanism
16 of gene regulation and signaling pathway is unknown.

17 **Material and Methods:** GSE9508, including 34 OS samples and 5 non-malignant bone
18 samples, was gained from Gene Expression Omnibus (GEO) database. The
19 differentially expressed genes (DEGs) were picked out by GEO2R online R soft tool.
20 Furthermore, the protein-protein interaction (PPI) network between the DEGs was
21 molded utilizing STRING online software. Afterward, PPI network of DEGs was
22 constructed. Gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes
23 (KEGG) pathway enrichment analysis of DEGs were carried out on DAVID online tool
24 and visualized via cytoscape software. Subsequently, module analysis of PPI was
25 performed by using MCODE app. What's more, prognosis-related genes were screened
26 by using online databases including GEPIA, UALCAN and cBioPortal databases.

27 **Results:** Totally, 671 DEGs were picked out, including 501 up-regulated genes and 170
28 down-regulated genes. Moreover, 22 hub genes were identified to be significantly

29 expressed in PPI network (16 up-regulated and 6 down-regulated). We found that
30 spliceosome signaling pathway may provide a potential target in OS. Furthermore, on
31 the basis of common crucial pathway, PRPF38A and SNRPC were closely associated
32 with spliceosome.

33 **Conclusion:** This study showed that SNRPC and PRPF38A are potential biomarkers
34 candidates for osteosarcoma.

35 Key words: PRPF38A, SNRPC, Osteosarcoma, GEO, DEGs

36 **Introduction**

37 The early detection of OS is limited. Most patients were diagnosed at a middle or
38 advance stage. Therefore, it is very valuable to find a biomarker for evaluation of the
39 of OS(1-4). Recently. High-throughput biochip technology has been prevalent in
40 screening of pre-symptomatic disease(5-7). Bioinformatics have led to using for data-
41 intensive statistical analysis in various fields of healthcare(8-12). Development of big-
42 data has attracted attention in recent years(13-16). It provided new insights toward
43 novel strategies for early testing and diagnosis of cancer(17-20). However,
44 bioinformatics study of OS has not been catalogued. We tested GSE9508(Platform
45 GPL676) from GEO. The dataset was divided into two part :34 OS sample and 5 non-
46 malignant bone sample. Microarray dataset was downloaded from GEO and analyzed
47 via GEO2R online tool. What' s more, DEGs between OS samples and non-malignant
48 bone samples were screened by GEO2R. Moreover, DAVID version 6.8 (21)was used
49 to analyze KEGG pathway and GO enrichment analysis with 501 up-regulated and 170
50 down-regulated DEGs. Multiple PPI network was calculated by using STRING online
51 tool version 11.0(22). Furthermore, we establish simulation of PPI network with DEGs
52 in Cytoscape software (version 3.7.2) (23)and then 23 core genes were picked out by
53 MCODE app(24). Next, these genes were imported into (25) cBioPortal
54 (26)databases for significant prognosis information($P < 0.05$). However, only 8 genes
55 were valid and then re-analyzed these central genes for KEGG pathway enrichment.
56 Last but not least, PRPF38A and SNRPC were obtained and enriched in spliceosome
57 pathway. The aim of present research was to provide a potential biomarker in OS.

58 **Materials and methods**

59 *Microarray and DEGs.* Microarray data containing OS related information were
60 download from the GEO database (<http://www.ncbi.nlm.nih.gov/gds/>), including 34 OS
61 samples and 5 non-malignant bone samples. A microarray dataset, GSE9508(GPL6076).
62 Analyzed via GEO2R online R soft tool(27). DEGs were selected according to adjust
63 P-value<0.05, $\log_{2}FC > 1$. All data were preserved in excel format. On this basis,
64 inclusion criteria for up-regulated was $\log_{2}FC > 1$. Likewise, $\log_{2}FC > -1$ was considered
65 as down-regulated.

66 *GO and KEGG pathway enrichment analysis.* Two common approaches for
67 identifying molecular function and biological process as mentioned above. Functional
68 enrichment analysis was performed DAVID Bioinformatics resources database version
69 6.8(28)(<http://david.ncifcrf.gov/>).

70 *Protein-protein interaction.* We cluster the DEGs PPI network to identify novel
71 gene utilizing STRING online tool version 11.0(22) (<https://string-db.org>). Finally, the
72 data were saved in tab-separated values (TSV format).

73 *Central genes module analysis.* Raw data files were imported into Cytoscape3.7.2,
74 we run Molecular Complex Detection(MCODE)(24), an app of cytoscape, to calculate
75 the significant modules in PPI network, with the threshold set as follows: Degree
76 cutoff=2, Node Score Cut off=0.2, Max. Depth=100, K-core=0.2.

77 *Overall survival of prognosis-related genes.* Survival prediction was performed
78 using cBioPortal(26)(<http://www.cbioportal.org/>). Statistical analyses for survival were
79 performed with log rank test. A P-values less than 5% was considered as statistically
80 significant. In addition, novel genes were pinpointed according to the threshold values
81 above.

82 **Result**

83 *Identification of DEGs in osteosarcoma.* There were 34 OS samples and 5 non-
84 malignant samples in our present research. Totally, 671 DEGs were picked up through
85 GEO2R online R software tool, involving 501 up-regulated (namely $\log_{2}FC > 1$) and 170
86 down-regulated (namely $\log_{2}FC < -1$) (Table I).

87 *GO and KEGG pathway enrichment analysis.* GO and KEGG pathway enrichment
88 analysis was performed using DAVID database. Within the biological process (BP)

89 categories, nuclear-transcribed mRNA poly(A) tail shortening, vesicle fusion, cellular
90 response to caffeine, female meiotic division, regulation of heart rate were dominant
91 term for up-regulated DEGs. Down-regulated were particularly featured in translation,
92 SRP-dependent cotranslational protein targeting to membrane, regulation of viral
93 transcription, nuclear-transcribed mRNA catabolic process, nonsense-mediated decay,
94 negative regulation of endopeptidase activity. For molecular function (MF)level, up-
95 regulated genes included in transcription factor activity, sequence-specific DNA
96 binding, nucleotide binding, extracellular matrix constituent, lubricant activity, poly(A)
97 RNA binding, RNA polymerase II regulatory region sequence-specific DNA binding.
98 However, down-regulated genes involved in structural constituent of ribosome, protein
99 binding, histone methyltransferase activity, U1 snRNA binding, endopeptidase
100 inhibitor activity. In the cellular component (CC)category, eukaryotic translation
101 initiation factor 4F complex, nucleoplasm, dendrite, cytoplasmic microtubule, primary
102 cilium were main enrichment term for up-regulated. Whereas, the down-regulated
103 genes were involved in photoreceptor inner segment, blood microparticle,
104 photoreceptor outer segment, ribosome, perinuclear region of cytoplasm (Table II).
105 KEGG pathway analysis was shown that up-regulated genes mainly play a role in
106 spliceosome(P-value<0.05). While the significance of identified pathway, down-
107 regulated related, was ribosome(P-value<0.05) (Table III).

108 *Protein-protein interaction.* The DEGs were mapped to PPI network by using
109 STRING (Fig. I). In addition, the network was identified via MCODE plugin in
110 cytoscape software. Additionally, 23 central genes were screened, which include 16 up-
111 regulated and 7 down-regulated (Fig. IIA and Fig. IIB).

112 *Overall survival of prognosis-related genes.* Analysis of central gene by using
113 cBioPortal database. A total of 8 prognosis-associated genes were identified using
114 overall survival analysis from above database (Table IV and Fig.III).

115 *Re-analysis of 8 central genes by KEGG pathway enrichment analysis.* A subset
116 of 8 genes, which related to prognosis, were selected for KEGG pathway enrichment
117 analysis. The result of the cluster shows an enrichment for terms related to spliceosome,
118 which SNRPC and PRF38A were directly involved in (Table V and Fig. IV).

119 **Discuss**

120 Early diagnosis and timely treatment contribute to improving the prognosis of
121 sarcoma(29-32). In this study, we analyzed GSE9508 dataset downloaded from GEO
122 database, and calculated 2 potential genes (SNRPC and PRPF38A) associated with OS
123 through a serious of bioinformatics method. It is worth nothing that 671 DEGs were
124 screened out after a comparative analysis between OS samples and non-malignant
125 samples via GEO2R online R software(33, 34)($|\log_{2}FC| > 1$ and adjust $P < 0.05$) including
126 501 up-regulated and 170 down-regulated DEGs. After bioinformatical analysis of GO
127 and KEGG pathway, we acquired that 1) for BP categories, up-regulated DEGs were
128 featured in regulation of heart rate, vesicle fusion, nuclear-transcribed mRNA poly(A)
129 tail shortening, cellular response to caffeine, female meiotic division were dominant
130 term for up-regulated DEGs. Down-regulated were particularly enriched in regulation
131 of viral transcription, translation, SRP-dependent cotranslational protein targeting to
132 membrane, nuclear-transcribed mRNA catabolic process, nonsense-mediated decay,
133 negative regulation of endopeptidase activity; 2) for MF, up-regulated genes included
134 in transcription factor activity, sequence-specific DNA binding, nucleotide binding,
135 extracellular matrix constituent, lubricant activity, poly(A) RNA binding, RNA
136 polymerase II regulatory region sequence-specific DNA binding. However, down-
137 regulated genes included in structural constituent of ribosome, protein binding, histone
138 methyltransferase activity, U1 snRNA binding, endopeptidase inhibitor activity; 3) for
139 CC, up-regulated DEGs were enriched in eukaryotic translation initiation factor 4F
140 complex, nucleoplasm, dendrite, cytoplasmic microtubule, primary cilium were main
141 enrichment term for up-regulated. Whereas, the down-regulated concentrated in
142 photoreceptor inner segment, blood microparticle, photoreceptor outer segment,
143 ribosome, perinuclear region of cytoplasm (Table II).

144 KEGG enrichment analysis provided details that up-regulated genes participated
145 in spliceosome signaling. Based on the KEGG pathway, down-regulated gene were in
146 ribosome (Table III, $P < 0.05$). Though PPI network construction, 23 core genes were
147 identified (involving 16 up-regulated and 6 down-regulated) via GEPIA and UALCAN
148 and cBioPortal databases. A total of 8 genes significantly associated with prognosis

149 were screened.

150 Additionally, re-analysis 8 novel genes by using DAVID shown that SNRPC and
151 PRPF38A as potential biomarkers and spliceosome had a significant, which provided a
152 new effective target to improve the prognosis of OS patients.

153 Small nuclear ribonucleoprotein poly peptide C(SNRPC), is also known as U1
154 snRNP, which is important for recognition of pre-mRNA and assembly of
155 spliceosome(35, 36). Another study proved that SNRPC is related to splice-site(37).
156 Some of these protein interact with SNRPC(38). Moreover, Wang et al indicated that
157 SNRPC, which is the main component of spliceosome(39). Lindén et al reported
158 alternation in pre-mRNA spliceosome is a promising biomarker, which plays an
159 important role in diagnosis of Stochastic Loewner Evolution (SLE)(40, 41).

160 Pre-mRNA splicing factor 38A (PRPF38A), is a center of spliceosome component,
161 which is related to protein-protein interaction(42). Previous research proved that knock
162 down of PRPF38A resulted in widespread intron retention (43). Whereas it was then
163 demonstrated that knock down of PRPF38A promotes apoptosis in triple negative
164 breast cancer(44).

165 To date, many studies confirm that SNRPC and PRPF38A were related to
166 numerous types of cancer progression(36, 45-48). Nevertheless, studies were very few
167 and far between in OS. Thus, these data may provide some functional information from
168 bench to bedside.

169 **Conclusions**

170 In summary, our study presents SNRPC and PRPF38A were identified as potential
171 biomarkers in OS by bioinformatical analysis. We provide a new strategy for
172 osteosarcoma treatment, however, which should be verified in the future.

173 **Abbreviations**

174 OS: Osteosarcoma; GEO: Gene Expression Omnibus; DEGs: differentially expressed
175 genes; PPI: protein-protein interaction; GO: Gene ontology; KEGG: Kyoto
176 Encyclopedia of Genes and Genomes; MCODE: Molecular Complex Detection;
177 BP :biological process; MF: molecular function; CC: cellular component; SNRPC:
178 Small nuclear ribonucleoprotein poly peptide C; SLE: Stochastic Loewner Evolution;

179 PRPF38A: Pre-mRNA splicing factor 38A;

180 **Ethical Approval and Consent to participate**

181 Not applicable.

182 **Consent for publication**

183 Not applicable.

184 **Availability of supporting data**

185 We declare that all data supporting our findings are provided in the manuscript.

186 **Conflict of Interest**

187 No conflict of interest declared.

188 **Acknowledgement**

189 We wish to thank Dr. Endo-Munoz LB for his contribution to the investigation of
190 osteosarcoma.

191 **Funding**

192 Award for “Liaoning Distinguished Professor”.

193 **Authors' contributions**

Dr. CY conceived and designed the study. CS contributed to data analysis and wrote the first draft of the manuscript.

References

1. Y. Suehara *et al.*, Clinical Genomic Sequencing of Pediatric and Adult Osteosarcoma Reveals Distinct Molecular Subsets with Potentially Targetable Alterations. *Clinical Cancer Research* **25**, 6346-6356 (2019).
2. T. Guo, H. Ma, Y. Zhou, Bioinformatics analysis of microarray data to identify the candidate biomarkers of lung adenocarcinoma. *PeerJ* **7**, (2019).
3. F. Li *et al.*, Identification of Key Biomarkers and Potential Molecular Mechanisms in Renal Cell Carcinoma by Bioinformatics Analysis. *J Comput Biol* **26**, 1278-1295 (2019).
4. L. Li, X. Chen, Z. Chen, Identification of Key Candidate Genes in Dairy Cow in Response to Escherichia coli Mastitis by Bioinformatical Analysis. *Front Genet* **10**, 1251 (2019).
5. R. Giorgi Silveira *et al.*, MicroRNAs expressed in neuronal differentiation and their associated pathways: Systematic review and bioinformatics analysis. *Brain Res Bull* **157**, 140-148 (2020).
6. L. Deng *et al.*, Identification and characterization of biomarkers and their functions for docetaxel-resistant prostate cancer cells. *Oncol Lett* **18**, 3236-3248 (2019).
7. G. Li, L. Cai, L. Zhou, Microarray gene expression profiling and bioinformatics analysis reveal key differentially expressed genes in clival and sacral chordoma cell lines. *Neurological Research* **41**, 554-561 (2019).
8. H. Lin *et al.*, Identification of key candidate genes and pathways in hepatitis B virus-associated

- acute liver failure by bioinformatical analysis. *Medicine (Baltimore)* **97**, e9687 (2018).
9. L. Liu *et al.*, Identification of Key Candidate Genes and Pathways in Endometrial Cancer by Integrated Bioinformatical Analysis. *Asian Pac J Cancer Prev* **19**, 969-975 (2018).
 10. X. X. Liu, L. Cai, F. J. Liu, An in silico analysis of human sperm genes associated with asthenozoospermia and its implication in male infertility. *Medicine* **97**, (2018).
 11. Z. Shang, Y. Zhao, K. Zhou, Y. Xu, W. Huang, PAX5 alteration-associated gene-expression signatures in B-cell acute lymphoblastic leukemia. *Int J Hematol* **97**, 599-603 (2013).
 12. S. Shen *et al.*, Identification of core genes and outcomes in hepatocellular carcinoma by bioinformatics analysis. *J Cell Biochem* **120**, 10069-10081 (2019).
 13. C. Su *et al.*, Microarraybased analysis of COL11A1 and TWIST1 as important differentially expressed pathogenic genes between left and rightsided colon cancer. *Mol Med Rep* **20**, 4202-4214 (2019).
 14. Y. Tu *et al.*, Identification of candidate aberrantly methylated and differentially expressed genes in thyroid cancer. *J Cell Biochem* **119**, 8797-8806 (2018).
 15. R. Wang, J. Wei, Z. Li, Y. Tian, C. Du, Bioinformatical analysis of gene expression signatures of different glioma subtypes. *Oncol Lett* **15**, 2807-2814 (2018).
 16. G. Xu *et al.*, Gene expression profile and bioinformatics analysis revealed key molecular characteristics of chordoma-before and after TNF- a treatment. *Medicine (Baltimore)* **99**, e18790 (2020).
 17. T. Xing, T. Yan, Q. Zhou, Identification of key candidate genes and pathways in hepatocellular carcinoma by integrated bioinformatical analysis. *Exp Ther Med* **15**, 4932-4942 (2018).
 18. X. Zhang *et al.*, Bioinformatics analysis and identification of potential genes related to pathogenesis of cervical intraepithelial neoplasia. *J Cancer* **11**, 2150-2157 (2020).
 19. Z. Hu *et al.*, Ferritin: A potential serum marker for lymph node metastasis in head and neck squamous cell carcinoma. *Oncol Lett* **17**, 314-322 (2019).
 20. K. Jiang, H. Liu, D. Xie, Q. Xiao, Differentially expressed genes ASPN, COL1A1, FN1, VCAN and MUC5AC are potential prognostic biomarkers for gastric cancer. *Oncol Lett* **17**, 3191-3202 (2019).
 21. W. Huang da, B. T. Sherman, R. A. Lempicki, Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat Protoc* **4**, 44-57 (2009).
 22. D. Szklarczyk *et al.*, STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. *Nucleic Acids Res* **47**, D607-D613 (2019).
 23. A. Treister, A. R. Pico, Identifier Mapping in Cytoscape. *F1000Res* **7**, 725 (2018).
 24. B. Demchak *et al.*, The Cytoscape Automation app article collection. *F1000Res* **7**, 800 (2018).
 25. D. S. Chandrashekar *et al.*, UALCAN: A Portal for Facilitating Tumor Subgroup Gene Expression and Survival Analyses. *Neoplasia* **19**, 649-658 (2017).
 26. E. Cerami *et al.*, The cBio Cancer Genomics Portal: An Open Platform for Exploring Multidimensional Cancer Genomics Data: Figure 1. *Cancer Discovery* **2**, 401-404 (2012).
 27. S. Yao, T. Liu, Analysis of differential gene expression caused by cervical intraepithelial neoplasia based on GEO database. *Oncology Letters*, (2018).
 28. W. Huang da, B. T. Sherman, R. A. Lempicki, Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists. *Nucleic Acids Res* **37**, 1-13 (2009).
 29. X. Wang, M. Gu, Y. Ju, J. Zhou, PIK3C3 Acts as a Tumor Suppressor in Esophageal Squamous Cell

- Carcinoma and Was Regulated by MiR-340-5p. *Med Sci Monit* **26**, e920642 (2020).
30. W. Hong *et al.*, Immune-related prognosis biomarkers associated with osteosarcoma microenvironment. *Cancer Cell Int* **20**, 83 (2020).
 31. L. Endo-Munoz *et al.*, Loss of osteoclasts contributes to development of osteosarcoma pulmonary metastases. *Cancer Res* **70**, 7063-7072 (2010).
 32. L. L. Cao *et al.*, Novel classifiers with clinical laboratory parameters for early detection of osteosarcoma. *J Clin Lab Anal*, e23189 (2020).
 33. C. Dong *et al.*, Cancer stem cell associated eight gene-based signature predicts clinical outcomes of colorectal cancer. *Oncol Lett* **17**, 442-449 (2019).
 34. L. Yang *et al.*, Bioinformatical analysis of Gene Expression Omnibus (GEO) database associates TAF7/CCNB1, TAF7/CCNA2, and GTF2E2/CDC20 pathways with glioblastoma development and prognosis. *World Neurosurg*, (2020).
 35. L. L. Knoop, S. J. Baker, The Splicing Factor U1C Represses EWS/FLI-mediated Transactivation. *Journal of Biological Chemistry* **275**, 24865-24871 (2000).
 36. N. Ohkura, H. Yaguchi, T. Tsukada, K. Yamaguchi, The EWS/NOR1 Fusion Gene Product Gains a Novel Activity Affecting Pre-mRNA Splicing. *Journal of Biological Chemistry* **277**, 535-543 (2002).
 37. Y. Muto *et al.*, The structure and biochemical properties of the human spliceosomal protein U1C. *J Mol Biol* **341**, 185-198 (2004).
 38. N. Ayoub, M. Alkhatatbeh, M. Jibreel, M. Ababneh, Analysis of circulating adipokines in patients newly diagnosed with solid cancer: Associations with measures of adiposity and tumor characteristics. *Oncol Lett* **13**, 1974-1982 (2017).
 39. H.-Y. Wang, L. Zhou, J.-F. Gui, Identification of a putative oocyte-specific small nuclear ribonucleoprotein polypeptide C in gibel carp. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology* **146**, 47-52 (2007).
 40. M. Lindén *et al.*, Sex influences eQTL effects of SLE and Sjögren's syndrome-associated genetic polymorphisms. *Biology of Sex Differences* **8**, (2017).
 41. D. L. Armstrong *et al.*, GWAS identifies novel SLE susceptibility genes and explains the association of the HLA region. *Genes Immun* **15**, 347-354 (2014).
 42. S. Blanton, A. Srinivasan, B. C. Rymond, PRP38 encodes a yeast protein required for pre-mRNA splicing and maintenance of stable U6 small nuclear RNA levels. *Mol Cell Biol* **12**, 3939-3947 (1992).
 43. A. E. Armstrong, D. O. Walterhouse, P. J. Leavey, J. Reichel, A. L. Walz, Prolonged response to sorafenib in a patient with refractory metastatic osteosarcoma and a somatic PDGFRA D846V mutation. *Pediatr Blood Cancer* **66**, e27493 (2019).
 44. S. Chan *et al.*, Basal-A Triple-Negative Breast Cancer Cells Selectively Rely on RNA Splicing for Survival. *Mol Cancer Ther* **16**, 2849-2861 (2017).
 45. H. Z. Ling *et al.*, Discovery of new serum biomarker panels for systemic lupus erythematosus diagnosis. *Rheumatology (Oxford)*, (2020).
 46. M. Koguchi *et al.*, BMP4 induces asymmetric cell division in human glioma stem-like cells. *Oncology Letters*, (2019).
 47. Q. Jiang, W. Feng, C. Xiong, Y. Lv, Integrated bioinformatics analysis of the association between apolipoprotein E expression and patient prognosis in papillary thyroid carcinoma. *Oncol Lett* **19**, 2295-2305 (2020).
 48. G. M. Calaf, J. Abarca-Quinones, Ras protein expression as a marker for breast cancer. *Oncol*

Figure I. Protein-protein interaction network showing experimentally verified and predicted interaction information among the proteins encoded by the differentially expressed genes.

Figure II . A sub-network of key proteins was constructed by using Cytoscape. Yellow and blue depict the up- or down-regulated differentially expressed genes, consecutively.

Figure III Prognostic survival analysis of CDC34, HNRNPA3, SRRT, HNRNPD, LRRC41, PRPF38A, SNRPC and KLHL22 genes via cBioPortal database. CDC34, cell division cycle 34; HNRNPA3, heterogeneous nuclear ribonucleoprotein A3; SRRT, serrate RNA effector molecule; HNRNPD, heterogeneous nuclear ribonucleoprotein D; LRRC41, leucine rich repeat containing 41; PRPF38A, pre-mRNA processing factor 38A; SNRPC, small nuclear ribonucleoprotein polypeptide C; KLHL22, kelch like family member 22.

Figure IV Significantly enriched pathway terms of DEGs in osteosarcoma. KEGG pathway annotations of the spliceosome pathway. Red star-marked means significant genes.

Table I All 671 DEGs were detected from GSE9508, involving 501 up-regulated and 170 down regulated genes in OS samples compared to non-malignant sample.

Table II GO enrichment analysis of DEGs in osteosarcoma.

Table III KEGG enrichment analysis of DEGs in osteosarcoma.

Table IV The prognosis-related information of 23 central genes.

Table V KEGG pathway enrichment analysis with 2 novel genes.

Figures

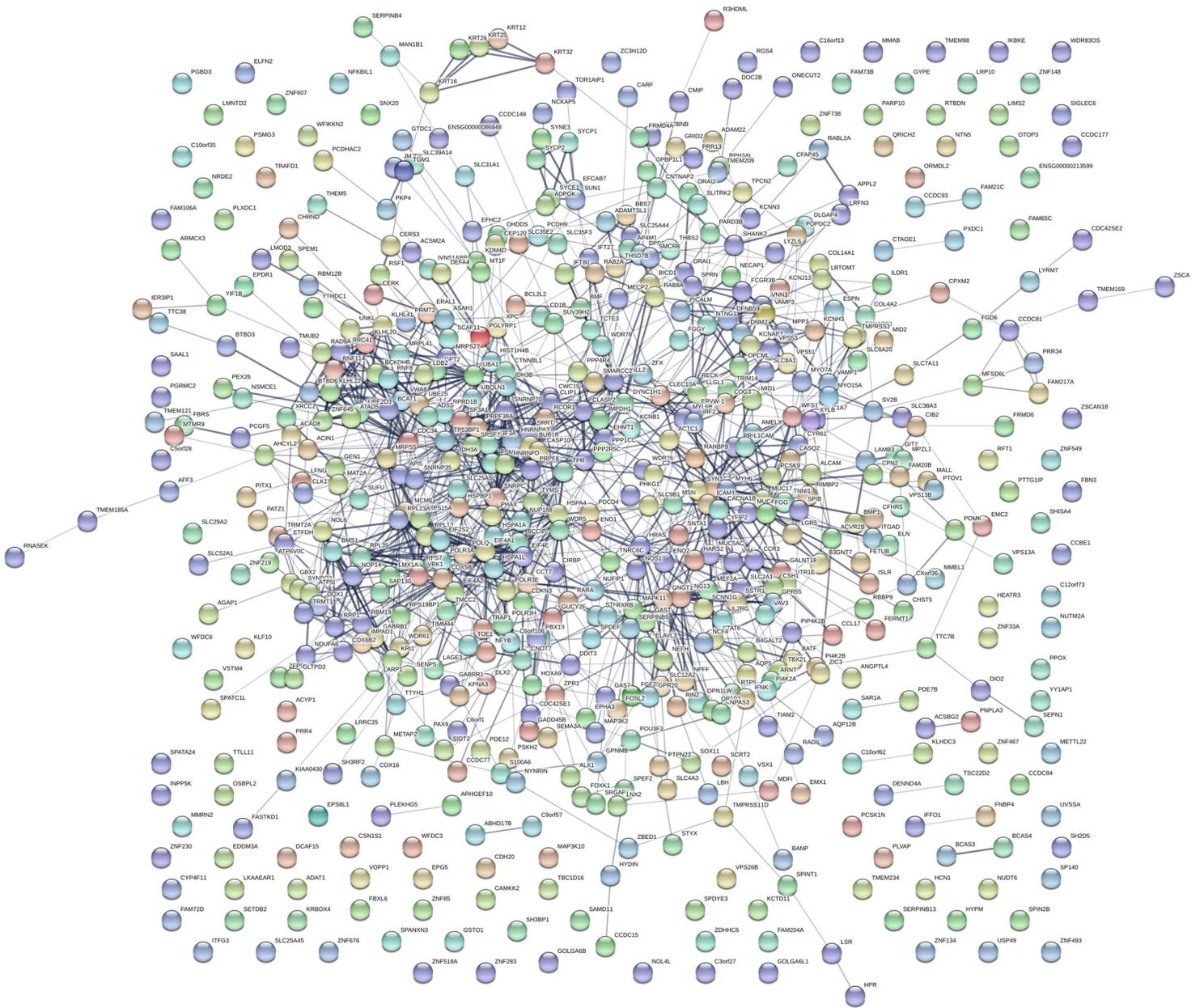


Figure 1

Protein-protein interaction network showing experimentally verified and predicted interaction information among the proteins encoded by the differentially expressed genes.

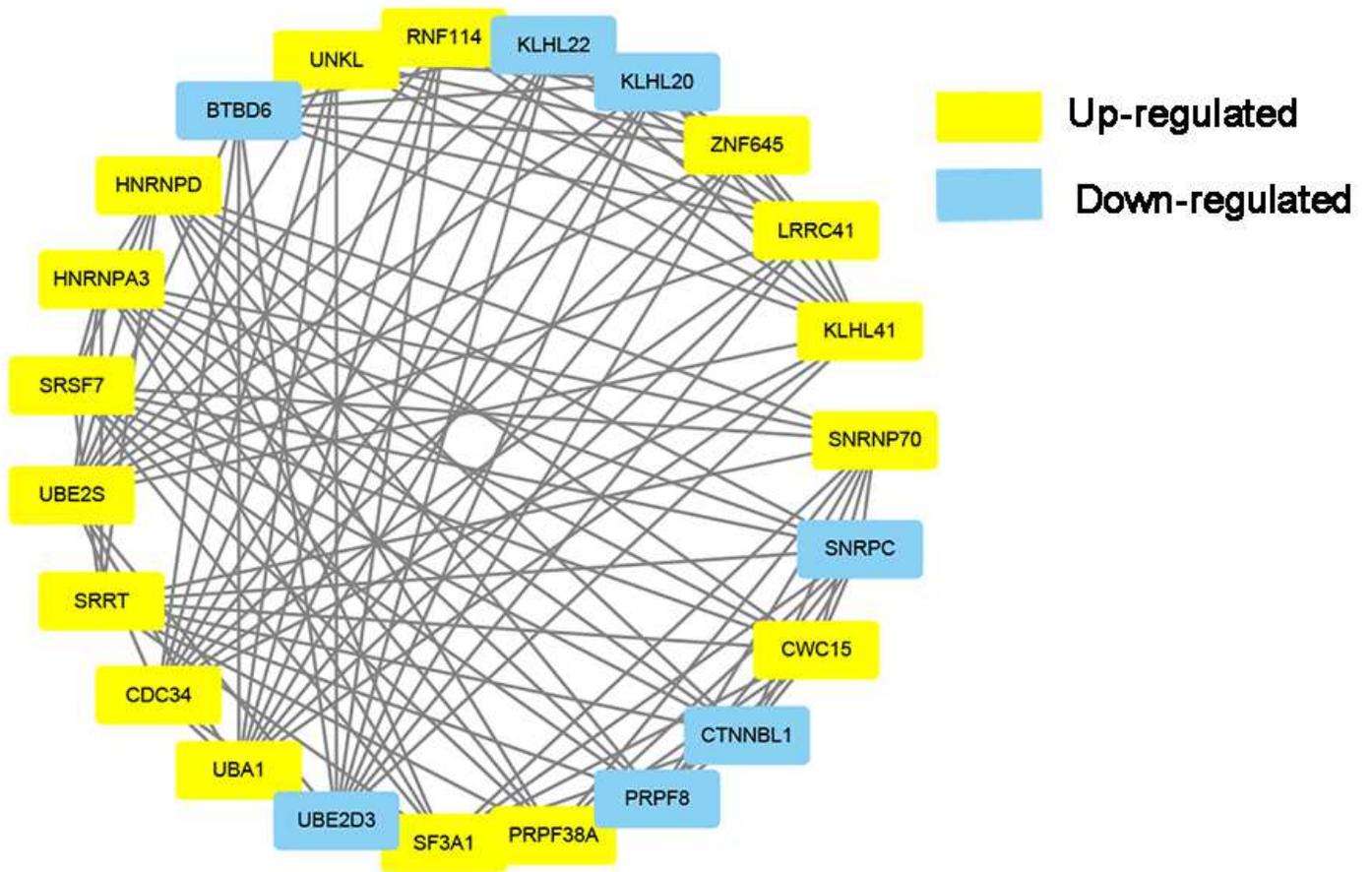


Figure 2

A sub-network of key proteins was constructed by using Cytocape. Yellow and blue depict the up- or down-regulated differentially expressed genes, consecutively.

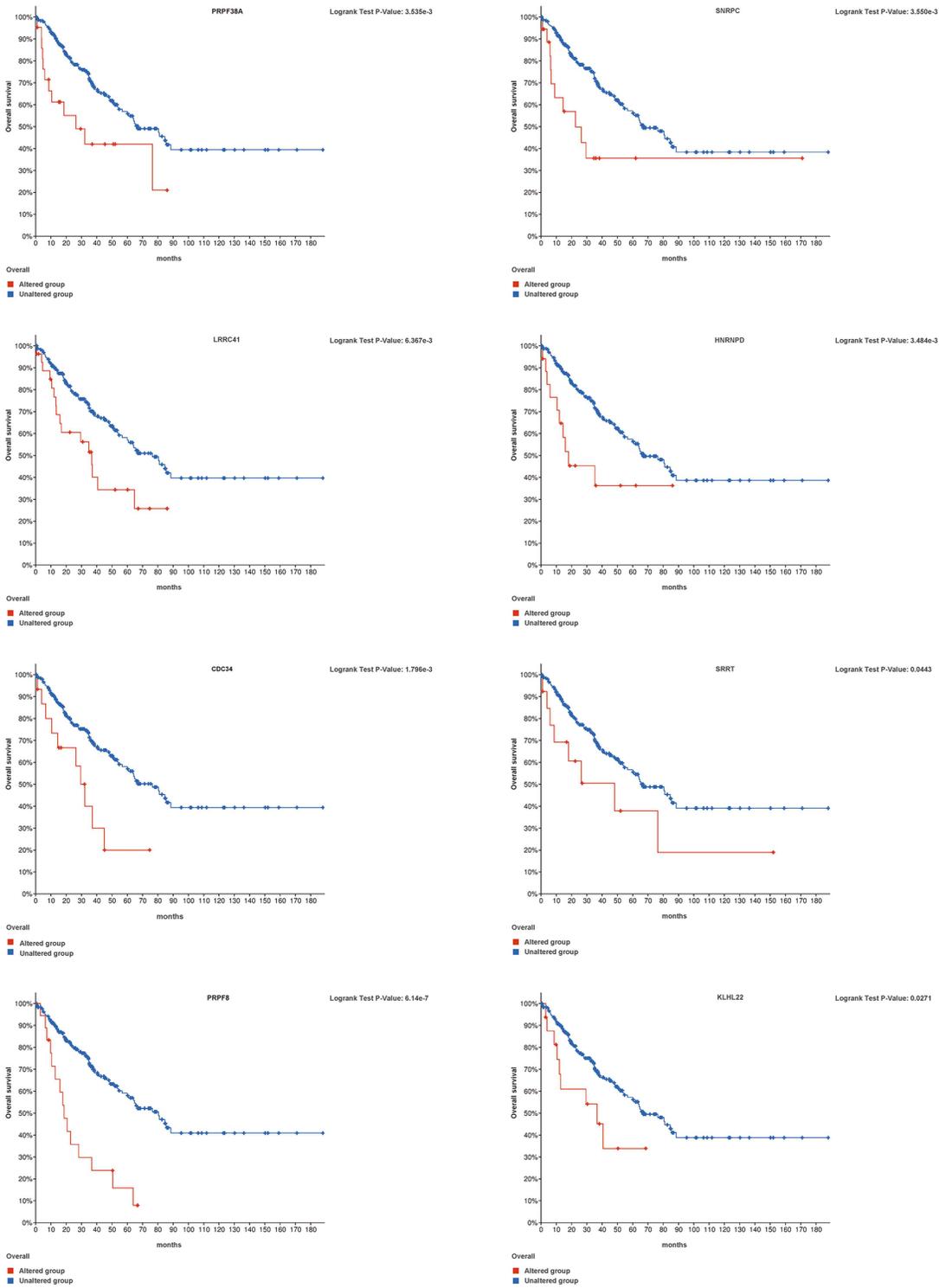
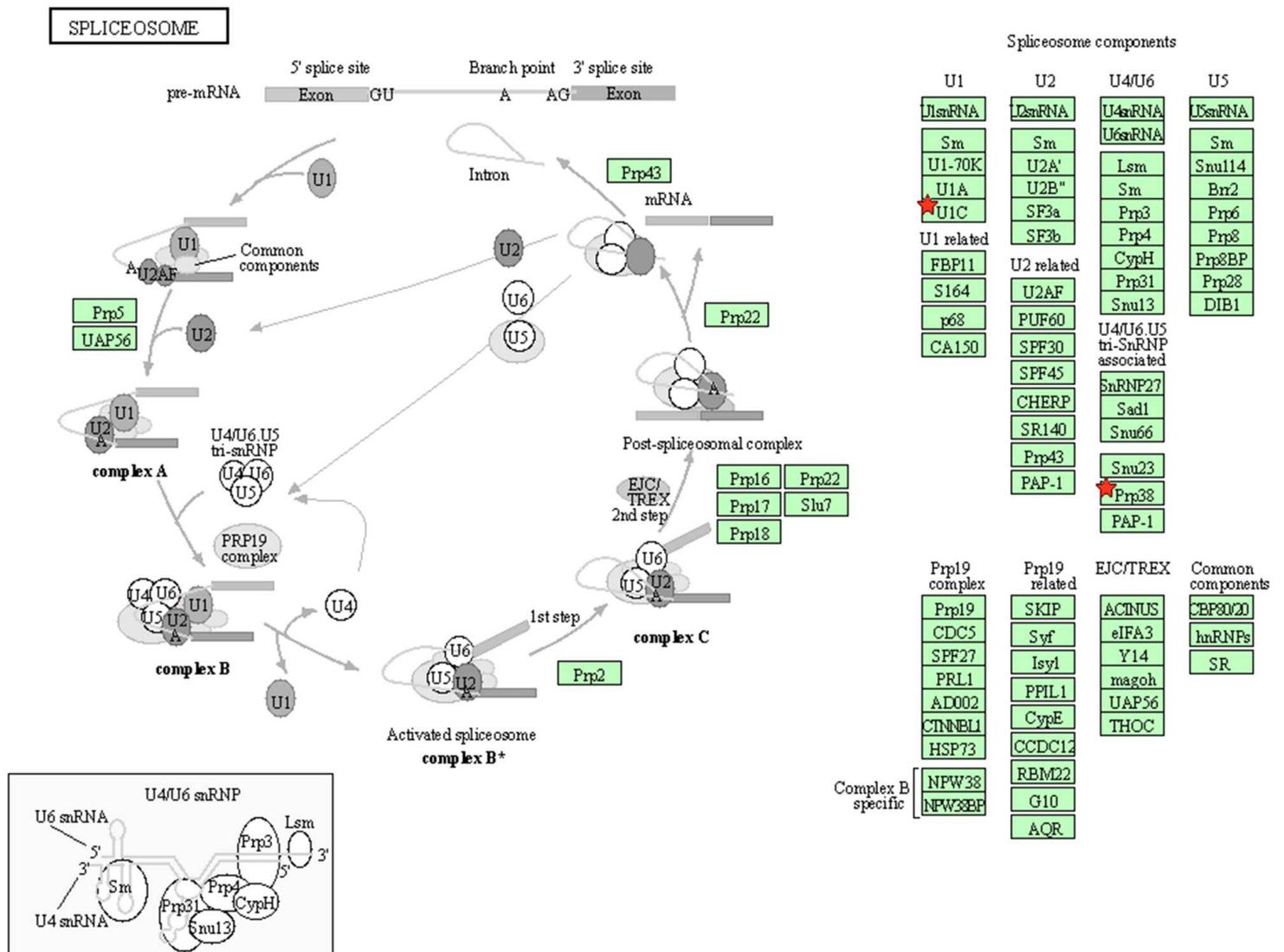


Figure 3

Prognostic survival analysis of CDC34, HNRNPA3, SRRT, HNRNPD, LRR41, PRPF38A, SNRPC and KLHL22 genes via cBioPortal database. CDC34, cell division cycle 34; HNRNPA3, heterogeneous nuclear ribonucleoprotein A3; SRRT, serrate RNA effector molecule; HNRNPD, heterogeneous nuclear ribonucleoprotein D; LRR41, leucine rich repeat containing 41; PRPF38A, pre-mRNA processing factor 38A; SNRPC, small nuclear ribonucleoprotein polypeptide C; KLHL22, kelch like family member 22.



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Figure 4

Significantly enriched pathway terms of DEGs in osteosarcoma. KEGG pathway annotations of the spliceosome pathway. Red star-marked splice means significant genes.

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