

Negative assortative mating and maintenance of shell colour polymorphism in *Littorina* (*Neritrema*) species

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Research Article

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Abstract

Colour polymorphism is a widespread phenomenon in natural populations of several species. In particular, it is especially common on marine gastropod species from the genus *Littorina*. Recently, it has been argued that intrapopulation shell colour polymorphism in *Littorina fabalis* could be caused by negative frequency-dependent sexual selection via a mechanism of mate choice (indirectly estimated via negative assortative mating). Here we try to determine the existence of negative assortative mating in three species of the subgenus *Neritrema* (*L. fabalis*, *L. obtusata*, *L. saxatilis*) that share a similar shell colour polymorphism, in order to ascertain if this mechanism could represent an ancestral character in this subgenus that could be contributing to the maintenance of the colour polymorphism in each species. Here, we collected or reanalysed from previous studies a sample of mating pairs of the three species from seven locations from NW Spain and NE Russia and estimated assortative mating using the I_{PSI} index. Our results show that all species and populations show a systematic tendency towards negative assortative mating when shell colour is grouped in the broad categories: 'light' and 'dark'. Although, a more detailed analysis of each colour individually suggests that shell colour may not be the main target of assortative mating, but perhaps physically linked to another trait or through pleiotropic effects. This hypothesis opens interesting new lines of research in *Littorina* snails.

Introduction

Colour is a key trait in evolution, usually contributing to individual survivorship (Olsson et al. 2013; Rolán-Alvarez et al. 2015a) and reproduction (Cordero 1989; Hugall and Stuart-Fox 2012), and therefore potentially influencing fitness. In addition, colour is a relatively easy trait to be detected and measured, and so it has been thoroughly studied in several taxa (McKinnon and Pierotti 2010; Hugall and Stuart-Fox 2012; Svensson 2017; Cuthill et al. 2017). In particular, shell colour has been extensively studied in gastropod snails (reviewed in Williams 2017).

Of special interest in studies of colour is colour polymorphism, in which there exist various discrete and genetically determined colour phenotypes within a population (White and Kemp 2016). Colour polymorphisms are expected to be unfrequent, given that directional natural selection and genetic drift tend to erode genetic variability over time, therefore reducing observed polymorphisms (Lewontin 1974; Nielsen 2005). Nevertheless, under certain circumstances selection can maintain a polymorphism in a population, and even genetic drift can do so, although transitorily and lasting from a few to several generations depending on effective population size and gene flow (Wright 1977; Clarke, 1979; Kimura 1983; Hartl and Clark 2006). This makes rather difficult to identify the true causal factor responsible for the evolutionary maintenance of colour polymorphism within a population. One of the most effective types of selection able to maintain a polymorphism in wild populations is negative frequency-dependent selection (Ayala and Campbell 1974; Fitzpatrick et al. 2007; Svensson and Connallon 2019). Negative frequency-dependent selection occurs when the fitness of a particular genotype correlates with its frequency, in a manner that a decrease in frequency increases its fitness relative to alternative genotypes

(Ayala and Campbell 1974). Fitness will peak as frequency falls below a certain threshold, hence protecting the polymorphism from being lost in the population.

The genus *Littorina* has been shown to be a particularly useful model organism in the field of evolutionary ecology (Rolán-Alvarez et al. 2015a). Besides speciation being the main focus of the evolutionary research carried out in these species (e.g. Morales et al. 2019; Costa et al. 2020; Galindo et al. 2020), the maintenance of intrapopulation colour polymorphism has also been studied (Rolán-Alvarez and Ekendahl 1996; Rolán-Alvarez et al. 2012, 2015b; Johannesson and Butlin, 2017; reviewed in Rolán-Alvarez et al. 2015a). Recently, it has been shown that shell colour polymorphism in populations of the intertidal gastropod *Littorina fabalis* (Turton 1825) can be maintained by negative frequency-dependent sexual selection *via* mate choice (Estévez et al. 2020). Another two species of the subgenus *Neritrema* (genus *Littorina*), *L. saxatilis* (Olivi 1792) and *L. obtusata* (Linnaeus 1758), have also shown colour polymorphism within certain populations (Reid 1996; Rolán-Alvarez et al. 2015a). These three species are direct developers (Reid 1996), lacking a planktotrophic larval stage, and therefore showing a potential for fast adaptation to local conditions (reviewed in Johannesson 2003; Galindo and Grahame 2014; Rolán-Alvarez et al. 2015a). Another important characteristic in these (dioecious) *Littorina* species is the possibility to observe mating pairs, which allows to readily estimate assortative mating in the wild (reviewed in Rolán-Alvarez et al. 2015a). In this context, assortative mating is understood as the phenotypic correlation between mates (Jiang et al. 2013; Janicke et al. 2019), which can be measured by isolation indexes in the case of qualitative traits, like shell colour (reviewed Gilbert and Starmer 1985; Rolán-Alvarez and Caballero 2000). Assortative mating for colour and its evolutionary consequences have been widely studied in several species over the last few years (Calderon et al. 2010; Pryke 2010; De Lanuza et al. 2013; Gade et al. 2016; Hedrick et al. 2016; Yang et al. 2016; Fargevieille et al. 2017). Besides, frequent biases in the estimation of assortative mating as well as its potential solutions have been recently identified (Rolán-Alvarez et al. 2015b)

Although there are alternative explanations (see Taborsky et al. 2009), assortative mating in simple organisms such as marine gastropods is presumably caused by mate choice (Ng et al. 2019; Estévez et al. 2020). On the other hand, negative assortative mating is directly related to negative-frequency dependent selection, as it has been shown both theoretically (Pusey and Wolf 1996; Hedrick et al. 2016 and 2018) and empirically in several species (Takashi and Hori 2008; Field and Barrett 2012; Holman et al. 2013; Hedrick et al. 2016 and 2018; Estévez et al. 2020). This, in turn, makes it plausible to assume that negative assortative mating is a relatively good proxy for both mate choice and negative-frequency dependent selection. Here we study assortative mating for shell colour in new populations of *L. fabalis* and two other related species with a similar colour polymorphism, *L. obtusata* and *L. saxatilis*. The aim of this study is to determine the existence of a negative assortative mating pattern in these two new species, as well as to corroborate its presence in the new populations of *L. fabalis*. Given that previous results (e.g. Estévez et al. 2020) strongly suggest a causal link between negative assortative mating and both mate choice and negative frequency-dependent selection, we suggest that if a negative assortative mating pattern is found in these two new species, an ancestral plesiomorphic behavioural trait for mate choice may exist within the subgenus *Neritrema*, which encompasses *L. fabalis*, *L. obtusata* and *L.*

saxatilis. This could pave the way to design future studies to directly test the existence of mate choice and negative frequency-dependent selection in the the Littorina clade.

Material And Methods

Study species and trait

We selected three colour polymorphic species from the genus *Littorina* (*L. obtusata*, *L. fabalis* and *L. saxatilis*), subgenus *Neritrema*. This subgenus is the only one with non-planktotrophic development within the genus, and it is formed by 10 species with a Northern distribution (Reid 1996; Reid et al. 2012). Within this subgenus we find the group of flat periwinkles composed by *Littorina fabalis* and *L. obtusata*, which are two sibling species (split 0.85 Mya, Sotelo et al. 2020) that live on intertidal brown macroalgae (e. g. *Fucus* sp.). *L. fabalis* feeds on diatoms and other microalgae that grow on the seaweed surface, while *L. obtusata* feeds directly on the macroalgae (Reid 1996). They also differ in habitat, *L. fabalis* commonly dwelling lower on the shore while *L. obtusata* lives in the upper part of the intertidal range. In certain localities their distribution partially overlaps, whereas in others only one species is found depending on the environmental conditions of the shore. In NW Spain they can be observed in true sympatry living on the same *Fucus vesiculosus* plants (Rolán-Alvarez and Ekendahl 1996). On the other hand, they are both oviparous, laying egg masses on the algae from which juveniles hatch as miniature snails (Reid 1996).

The rough periwinkle, *L. saxatilis* (split with *L. fabalis* 2.5 Mya, Panova et al. 2011), is the only ovoviviparous species in this subgenus (Reid 1996). The embryos are developed in a brood pouch inside the female until eventually hatching as miniature snails. *L. saxatilis* lives on the rock surfaces across the whole intertidal range, feeding on diatoms, microalgae and detritus (Otero-Schmitt et al. 1997). The three species have very similar distribution across NE Atlantic coasts (Reid 1996) and in many shores they can be found overlapping, although their microhabitats might differ.

Shell colour in these species have been typically studied by visual (qualitative) codification (reviewed in Reid, 1996; Rolán-Alvarez et al. 2015a) but at least in one case (*L. fabalis* from Abelleira) the qualitative classes have been validated by reflectance spectrometry (Rolán-Alvarez et al 2015b). Shell colour variation in the three species has been systematically attributed to one or a few genes (reviewed in Rolán-Alvarez et al. 2015a).

Experimental methods

In order to study assortative mating for shell colour in *L. fabalis*, *L. obtusata* and *L. saxatilis*, the experimental design consisted in the capture of mating pairs and the surrounding unmated individuals in natural populations following previous studies (Rolán-Alvarez and Ekendahl 1996; Rolán-Alvarez et al. 2012, 2015b; Estévez et al. 2020). These three species show a similar mating strategy, where males usually perform an active search of females, following them by their mucus trails, and typically mate by size similarity (Saltin et al. 2013; Ng et al. 2019). In the field, a mating pair can be detected when the male

is located over the female in a certain position on the right-hand side of shell (with the axis of coiling of the two snails forming a 'V'-like pattern) and the penis is inserted in the mantle cavity of the female (Fig. 1A-B). Associated to every mating pair we collected the four closest unmated individuals of the same species (typically within 5 cm of the mating pair), and these six snails were defined as an individual microarea for further analyses (see Fig. 2 in Estévez et al. 2020). These samplings were done for several days during the summer for each year, since snail density and matings increase during this season (Rolán-Alvarez and Ekendahl 1996).

All the snails were taken alive to the laboratory and stored at -20° C. The mating pairs were dissected in order to discard male-male matings from the analyses, as they are known to be quite frequent in *L. saxatilis* (Rolán-Alvarez et al. 1999). Following these dissections, we discarded around 10% for *L. saxatilis* and 12% for flat periwinkles. Then, the background shell colour of each individual was scored using a colour printed model following Rolán-Alvarez et al. (2015b) and classified into the categories described in Fig. 1. The banding pattern present in some shells was ignored in further analyses, as its frequency differed between species and it also varied in fadedness. The three studied species shared yellow, orange and brown shells while olive is only present in flat periwinkles and white only in *L. saxatilis* (Fig. 1c-d), for the studied populations. As in previous studies of *L. fabalis* (Rolán-Alvarez et al. 2015b; Estévez et al. 2020) these colours were classified into two categories: light (white, yellow, orange) and dark (olive, brown). In addition, in this study we have also performed the analyses based on the individual colours, comparing each colour against the rest, on a binary manner (see Table S1).

The analyses were carried out with the colour of the mated and unmated individuals using a previously published dataset on *L. fabalis* (Estévez et al. 2020) and new collected data on the three species (data available at: <https://doi.org/10.6084/m9.figshare.13295873.v1>). Estévez et al. (2020) data is composed by samples from 7 consecutive years (2011–2017) from Abelleira (Ria of Muros-Noia; NW Spain; 42°47'45.9"N – 9°01'19.1"W) and two close sites (< 1 Km apart) from the White Sea (NE Russia; WS-1, 66°20'10.73"N – 33°37'53.00"E; WS-2, 66°20'9.80"N – 33°38'41.29"E) collected in 2015. Then a new population of *L. fabalis* was sampled in Cangas NW Spain (Ria of Vigo; NW Spain, 42°15'43.7"N – 8°45'28.4"W) in 2018. Another locality, Castelete, was sampled for *L. fabalis* and *L. obtusata* (Ria of Arousa; NW Spain, 42°32'36.9"N – 8°49'57.0"W) in 2016. In this case both species were differentiated based on penis morphology following Reid (1996) and assuming the lack of interspecific mating pairs. For *L. saxatilis* two close by localities were sampled in the Ria of Vigo (NW Spain), Portela (42°17'17.74"N – 8°37'21.87"W) and Soutoxuste (42°19'22.30"N – 8°36'57.44"W) in 2020. The number of collected mating pairs for each species and locality is shown in Table 1.

Table 1

Estimates of assortative mating (I_{PSI}) for shell colour in natural populations of *L. fabalis*, *L. obtusata* and *L. saxatilis*. The studied localities for each species and the number (N) of microareas (mating pair + unmated individuals) are indicated. The first estimate of I_{PSI} takes into account the colours grouped into dark (olive, brown) or light (white, yellow, orange). The second estimate is carried out for each separate colour vs the rest of the colours (e.g. white vs yellow/orange/olive/brown). For each I_{PSI} the standard deviation is shown.

Species	Locality	N	I_{PSI}					
			Dark/Light	White	Yellow	Orange	Olive	Brown
<i>L. fabalis</i>	Abelleira#	811	-0.29 ^{***} ± 0.068	-	-0.31 ^{**} ± 0.021	-0.03 ± 0.632	0.07 ± 0.403	-0.30 ^{**} ± 0.032
	Cangas	223	-0.24 ± 0.158	-	-0.23 ± 0.165	-0.16 ± 0.291	0.41 ± 0.304	-0.31 [?] ± 0.133
	Castelete	12	-0.55 ± 0.403	-	-	-	-	-
	WS-1 (Russia)#	93	-0.44 [?] ± 0.202	-	-0.35 [*] ± 0.132	-	-0.45 [*] ± 0.145	-
	WS-2 (Russia)#	77	-0.27 ± 0.403	-	-0.23 ± 0.165	-	-0.40 [*] ± 0.117	-0.35
			-0.36 [*] ± 0.123		-0.28 ^{**} ± 0.060	-0.09 ± 0.092	-0.09 ± 0.409	-0.32 ^{**} ± 0.026
<i>L. obtusata</i>	Castelete	44	-0.59 ^{***} ± 0.117	-	-	-	-	-
<i>L. saxatilis</i>	Sotoxuste	74	-0.36 ^{**} ± 0.035	-0.16 ± 0.291	-0.26 ± 0.269	-0.17 ± 0.247	-	-0.56 ± 0.262
	Portela	114	-0.31 [?] ± 0.113	-0.11 ± 0.156	-0.34 [*] ± 0.104	-0.24 ± 0.339	-	-0.33 ± 0.085
			-0.33 [*] ± 0.025	-0.14 ± 0.035	-0.30 [?] ± 0.057	-0.20 ± 0.049	-	-0.44 ± 0.163
# from Estévez et al. (2020); [?] P < 0.10; * P < 0.05; ** P < 0.01; *** P < 0.001								

Statistical analyses

Assortative mating for qualitative traits can be estimated by several indices (reviewed in Rolán-Alvarez and Caballero 2000), from which the I_{PSI} index is the most robust under different sampling scenarios (Pérez-Figueroa et al. 2005). This index can be estimated in a hypothetical population with two colours (brown, B; yellow, Y) as,

$$I_{PSI} = \frac{(PSI_{BB} - PSI_{BY} - PSI_{YB} + PSI_{YY})}{(PSI_{BB} + PSI_{BY} + PSI_{YB} + PSI_{YY})} ,$$

being PSI_{BB} the ratio between the observed number of brown female–brown male mating pairs and the expectation of this type of mating pair under random mating (using exclusively the mated individuals to estimate random mating; see further details in Carvajal-Rodríguez and Rolán-Alvarez 2006 and references therein). The remaining PSI values are calculated accordingly. The I_{PSI} index ranges from -1 (maximum negative assortative mating) to 1 (maximum positive assortative mating), being 0 complete random mating. This index has been used successfully in previous studies of assortative mating in *Littorina* (Rolán-Alvarez et al. 2015b; Estévez et al. 2020). The I_{PSI} index was calculated for the trait shell colour, categorized as a binary trait in two different analyses. The first one (pooling analysis), grouped colours into light (white, yellow, orange) and dark (olive, brown) as mentioned in the experimental design and following previous studies. The second analysis (individual analysis) considered every colour separately against the rest of the colours grouped. These two analyses were carried out for each species and population datasets.

The I_{PSI} index gives an estimate of assortative mating for qualitative traits similar to Pearson's correlation coefficient for quantitative traits (Rolán-Alvarez et al. 2015b). However, these authors also showed that the existence of trait heterogeneity at a smaller scale than the scale of mate choice would bias assortative mating towards positive values (i. e. the scale-of-choice effect *sensu* Rolán-Alvarez et al. 2015b). Therefore, in order to reduce the potential bias towards positive values of assortative mating, we also performed an analysis subdividing each dataset (species/population) into three different homogeneous groups of microareas (mating pair + four unmated individuals) based on the colour frequencies of the microareas (pooling analysis: light vs dark; individual analysis: one colour vs the rest) as follows: low frequency (0-33.3% light or other colour), intermediate frequency (33.4–66.6%) and high frequency (66.7–100%). Then the I_{PSI} was estimated within each of these three homogeneous groups of microareas and then averaged across groups (Rolán-Alvarez et al. 2015b; Ng et al. 2016; Estévez et al. 2018, 2020). The significance of the I_{PSI} indices was accomplished by t-tests (with null value of 0) using the three different homogeneous groups (low, intermediate, high frequency), and for both analyses (pooling and individual). On the other hand, due to the low sample size obtained in Castelete, the significance has been estimated by bootstrapping in JMATING (Carvajal-Rodríguez and Rolán-Alvarez 2008) following Rolán-Alvarez et al. (2015b).

Results And Discussion

Even though directional natural selection and genetic drift would preclude the persistence of genetic diversity (Lewontin 1974; Nielsen 2005), colour polymorphisms exist in many invertebrate and vertebrate species, and they have even been shown to persist along species radiations (Jamie and Meier 2020). Even the same colour polymorphism can be found in closely related species, suggesting the persistence of a shared evolutionary mechanism maintaining the polymorphism across species radiation. We have studied a group of marine gastropod species within the subgenus *Neritrema* (genus *Littorina*) that share a similar shell colour polymorphism (Fig. 2; Table S1). The results of the pooling analyses of assortative mating (light vs dark) using mating pairs from natural populations of the three studied species (*Littorina fabalis*, *L. obtusata* and *L. saxatilis*) show a consistent negative assortative mating across all populations and species (Table 1). *L. obtusata* showed the greatest negative assortative mating ($I_{PSI} = -0.59$) and *L. fabalis* from Cangas, the lowest (-0.24). A similar result is obtained from the I_{PSI} estimates for the homogeneous groups of colour frequency (Supplementary Table S2), which have been used to correct for the scale-of-choice effect (Rolán-Alvarez et al. 2015b). These estimates support previous results obtained for *L. fabalis* in Abelleira (Rolán-Alvarez and Ekendahl 1996; Rolán-Alvarez et al. 2012, 2015b; Estévez et al. 2020) and are also similar to those found in other species (Takashi and Hori 2008; Field and Barrett 2012; Holman et al. 2013; Hedrick et al. 2016 and 2018). Even though our results cannot unravel the evolutionary mechanisms responsible for the observed patterns, they strongly suggest that negative assortative mating for shell colour in *Neritrema* species is caused by mate choice and therefore it would be at least partially responsible for the maintenance of the colour polymorphism within population *via* negative frequency-dependent sexual selection. There are at least two different pieces of evidence that support this claim, as we have already suggested: first, theoretical analysis have shown that any mate-choice based negative assortative mating will render a negative frequency-dependent sexual selection that could be able to maintain the polymorphism (Pusey and Wolf 1996; Hedrick et al. 2016 and 2018). Second, recently Estévez et al. (2020) have shown that this relationship between negative assortative mating, mate choice and negative frequency-dependent selection holds for *L. fabalis* (see similar evidences for other model organisms: Takashi and Hori 2008; Field and Barrett 2012; Holman et al. 2013; Hedrick et al. 2016 and 2018). These two strains of evidence, in addition to the results presented in this study, strongly suggest that an ancestral plesiomorphic behavioural mechanism could exist for mate choice within the *Neritrema* subgenus. We find the ancestral behavioural mechanism hypothesis plausible, since, from an evolutionary point of view, it is reasonable to expect similar behavioural mechanisms operating under similar ecological conditions in closely related species. This is akin to what in evolutionary studies of cognition has been dubbed the “evolutionary parsimony” principle (de Waal 1999). Thus, if *L. fabalis*, *L. obtusata* and *L. saxatilis* show a similar behavioural pattern, live in the same ecosystem, and they all share a (relatively) recent common ancestor, it is reasonable to expect a common behavioural mechanism behind this behavioural pattern. Nevertheless, this hypothesis, although plausible, needs to be experimentally tested in the future before concluding its validity.

In any case, there are a few potential caveats in the present study that need to be addressed in order to dispel doubts about our results. First, the I_{PSI} and similar indexes allow us to estimate assortative mating, and are frequently used as a proxy of mate choice, at least in laboratory conditions (reviewed in Gilbert

and Starmer 1985; Rolán-Alvarez and Caballero 2000; Pérez-Figueroa et al. 2008). However, their capability to estimate mate choice has been discussed and several sources of bias have been identified (Rolán-Alvarez and Caballero 2000; Pérez-Figueroa et al. 2008). In any case, when they have been exclusively used to estimate assortative mating in the laboratory, these indices performed appropriately (Conde-Padín et al. 2008; Rolán-Alvarez et al. 2015b). On the other hand, it has been shown that correlation indexes could be potentially inflated towards positive values when estimated directly in the wild, especially in species with low mobility, as well as when the study species shows great variability (e. g. colour) at a scale smaller than the scale at which the samples were collected (Rolán-Alvarez et al. 2015b). This bias could be corrected by estimating the I_{PSI} in homogeneous groups (e. g. by similar colour), as it has been shown by simulations (Rolán-Alvarez et al. 2015b) and as it has been carried out here. In fact, this correction has been used previously in cases of negative and positive assortative mating (Rolán-Alvarez et al. 2015b; Ng et al. 2016; Estévez et al. 2018, 2020). Thus, the correction used here does not correct, *a priori*, towards either negative or positive values of assortative mating. For example, the same correction was used to estimate the real scale at which mate choice occurs in two species of marine gastropods (Estévez et al. 2018), one characterised by positive assortative mating (*Echinolittorina malaccana*) and the other by negative assortative mating (*L. fabalis*), highlighting that this method is suitable to estimate both types of assortative mating. Therefore, the correction that we applied in this study assures a more reliable estimate of assortative mating in natural populations.

Second, another potential bias of our results could be caused by the actual observation of the mating pairs during the collection of the samples. For example, mating pairs in flat periwinkles (*L. fabalis*, *L. obtusata*) are commonly found among a dense canopy of *Fucus vesiculosus*, typically dark brown, allowing for a possible sampling bias towards the most visible mating pairs (light-light, dark-light). Although, in order to avoid this potential bias, we performed the search of mating pairs very close to the algae (~ 30 cm) and for each sampled microarea we checked all the individual algae, so it was unlikely to miss dark-dark pairs more often than the rest. Moreover, this bias in the sampling procedure has been rejected in previous studies as light-light mating pairs, which are highly noticeable on the *Fucus* canopy, appear at significantly lower frequencies than expected under random mating (Rolán-Alvarez et al. 2015b; Estévez et al. 2020). This potential sampling bias is even more unlikely in *L. saxatilis*, given that in this case all mating pairs are collected on the rock surface, as this species grazes on microalgae that grows on the bare rock. Therefore, in the light of the current results it seems plausible a hypothesis where a mating strategy of negative assortative mating for shell colour could represent an ancestral character within the subgenus *Neritrema*.

Recently, Estévez et al. (2020) have investigated the behavioural mechanism responsible for the negative assortative mating observed in *L. fabalis* from Abelleira (NW Spain) and the White Sea (NE Russia) populations. They found indirect evidence that mate choice is involved in assortative mating, with males showing preference for females with different shell colours. This, in turn, points to negative frequency-dependent selection as the evolutionary mechanism maintaining colour polymorphism in this species. However, their results didn't provide any evidence regarding the potential contribution of colour to fitness,

so that it would be favoured by mate choice. Their results neither clarify if colour, and not any other genetically linked trait, is the real target of selection. Here, we present a complementary result that could help to understand the relationship between shell colours and mating in these species. In the analysis of each colour separately (Table 1 and Supplementary Table S2), we found an interesting pattern given that, although in general all colours showed the exact same trend as in those cases in which they are grouped in dark and light colours, there was an exception. The olive shell colour in *L. fabalis* changes from a pattern of random (positive but not significant estimates) mating in Spain to a pattern of negative assortative mating in the Russian populations (Table 1 and Supplementary Table S2; Fig. 3). This result is unexpected if the shell colour itself is the target of the putative behavioural mechanism causing the assortative mating, but it could be expected if the colour is linked to another adaptive trait, physically or through pleiotropy. A relevant question is, therefore, what circumstances make a particular mating behaviour to be present in one location but not in others. One hypothesis is that the colour genes are associated with several other genes in a chromosomal inversion. In such scenario, different alleles of the colour gene could be associated to different chromosome variants (inversed or not). In support of this view is the fact that colour vision has not been detected and olfactory cues seems more relevant than visual cues in determining movement in aquatic gastropods (Seyer, 1992; Wyeth 2019). Actually, Cephalopods, the animals with the most complex visual system within the phylum Mollusca, have only one visual pigment and therefore cannot see in colour (Nilsson 2013). If cephalopods do not have colour vision despite having a complex visual system, it seems implausible that gastropods could have it. Because of this, previous hypotheses about mate choice for shell colour in gastropods were based on the possibility that they can sniff different colours when males follow mucus trails of females in the context of mate searching (see Estévez et al. 2020). Actually, a relationship between assortative mating for colour forms and sex chromosomes have been claimed in one finch species (Pryke 2010). Chromosomal inversions have been recently detected and associated to presumably adaptive traits in several organisms (Wellenreuther and Bernatchez 2018), including littorinids (Westram et al. 2018; Faria et al. 2019), and in at least one case has been related to negative assortative mating too (Hedrick et al. 2018). This could mean that the trait responsible for the pattern of negative assortative mating would be linked to the colour gene within the inversion, but could not be the colour itself. Although speculative, this hypothesis can be tested in the future and it opens a potentially fruitful new line of research.

Declarations

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Consent to participate/publicationb: all included authors have given their consent to participate

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Figures

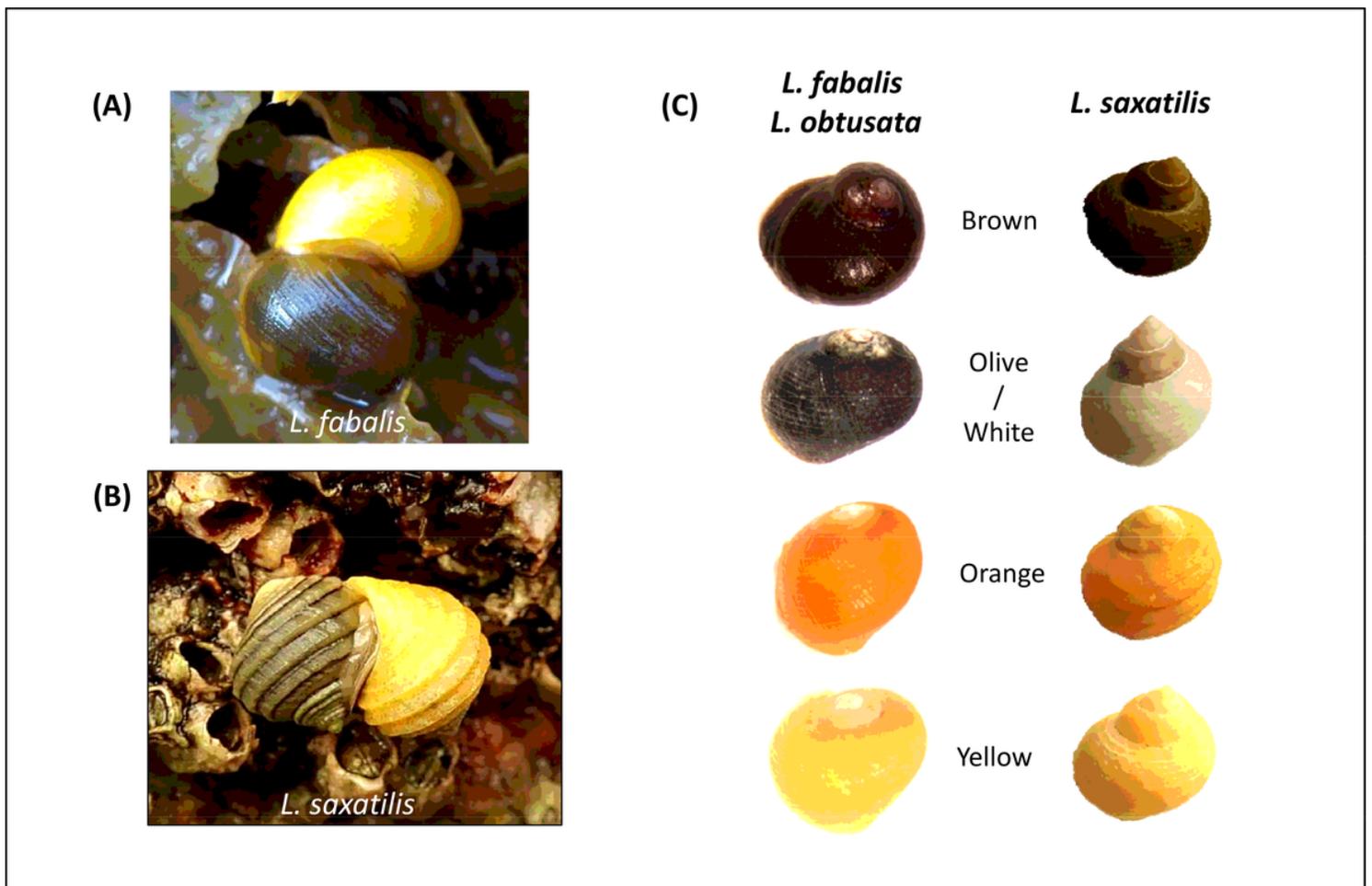


Figure 1

A) *Littorina fabalis* mating pair (yellow male and brown female). B) *L. saxatilis* mating pair (yellow male and brown female). C) Shell colour pattern in *L. fabalis*, *L. obtusata* and *L. saxatilis*.

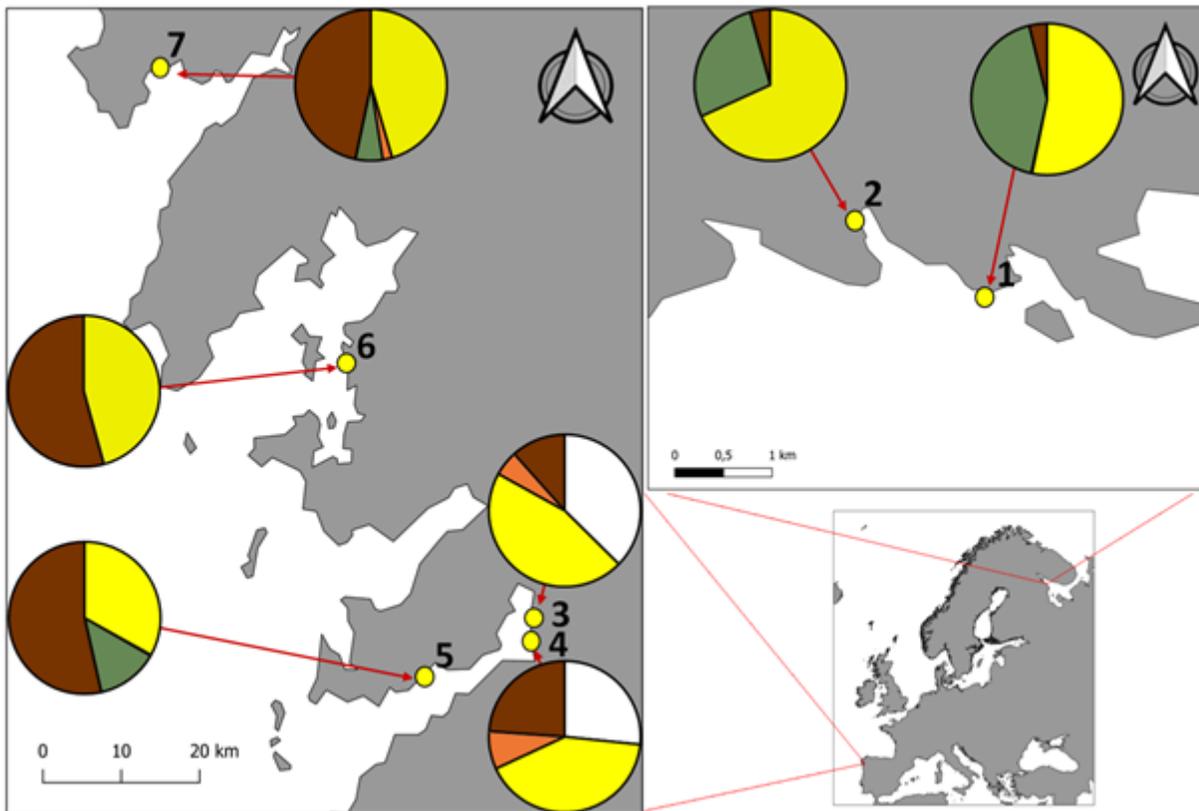


Figure 2

Geographical location of the different localities sampled for *L. fabalis* (1, 2, 5, 6, and 7), *L. obtusata* (6) and *L. saxatilis* (3 and 4). The pie charts represent the observed frequency of shell colours in each locality. Notice that in Castelete both *L. fabalis* and *L. obtusata* samples showed similar colour frequencies (Supplementary Table S1), and so only the *L. obtusata* pie chart is presented. Russia: 1 (WS-1) and 2 (WS-2). Spain: 3 (Portela), 4 (Soutoxuste), 5 (Cangas), 6 (Castelete) and 7 (Abelleira).

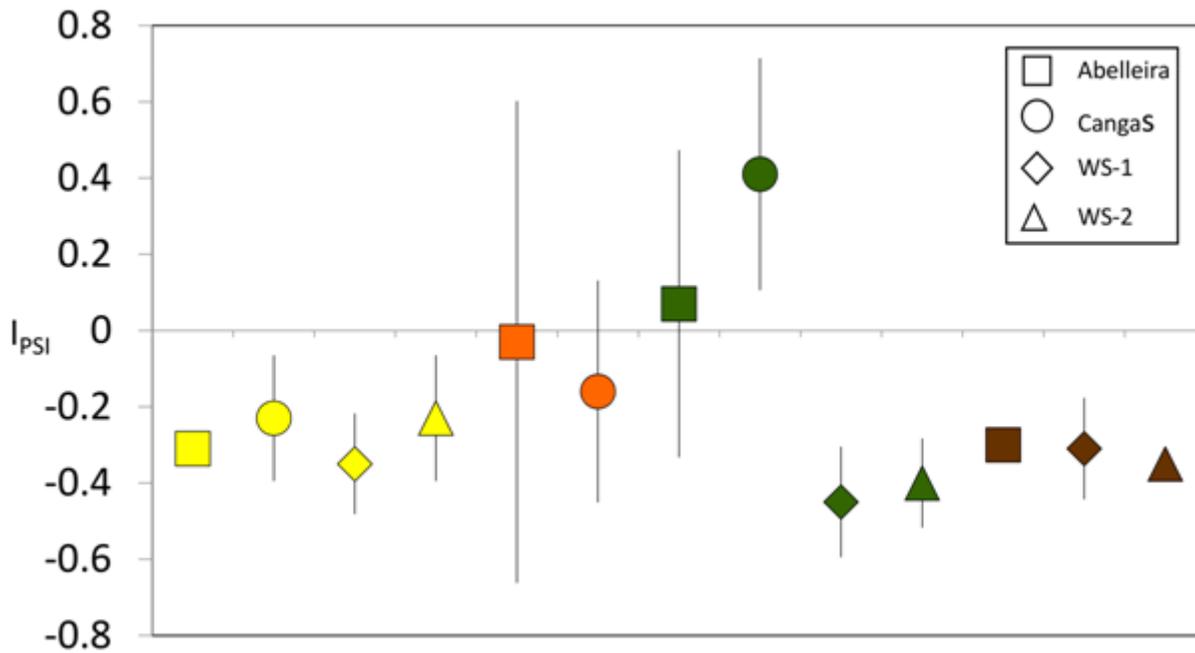


Figure 3

Estimates of corrected (for the scale-of-choice effect) IPSI values (with standard deviations) for the different populations of *L. fabalis* (from Table 1). Colours in the figure represent different shell colours. Notice that estimates for olive shell colour differ between Spain and Russia (Russia samples are significantly different from 0; see text). Notice that standard deviations for frequent colours in Abelleira are so small that fall inside symbols.

Supplementary Files

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