

# Radiation-induced Hounsfield unit change correlates with dynamic CT perfusion better than 4DCT-based ventilation measures in a novel-swine model

**Antonia E. Wuschner** (✉ [wuschner@wisc.edu](mailto:wuschner@wisc.edu))

University of Wisconsin–Madison

**Eric M. Wallat**

University of Wisconsin–Madison

**Mattison J. Flakus**

University of Wisconsin–Madison

**Dhanansayan Shanmuganayagam**

University of Wisconsin–Madison

**Jennifer Meudt**

University of Wisconsin–Madison

**Gary E. Christensen**

University of Iowa

**Joseph M. Reinhardt**

University of Iowa

**Jessica R. Miller**

University of Wisconsin–Madison

**Michael J. Lawless**

University of Wisconsin–Madison

**Andrew M. Baschnagel**

University of Wisconsin–Madison

**John E. Bayouth**

University of Wisconsin–Madison

---

## Research Article

**Keywords:** Radiation, Hounsfield units, biomarker, swine model, CT perfusion

**Posted Date:** March 18th, 2021

**DOI:** <https://doi.org/10.21203/rs.3.rs-257237/v1>

**License:** © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

---

# Radiation-induced Hounsfield unit change correlates with dynamic CT perfusion better than 4DCT-based ventilation measures in a novel-swine model

Antonia E. Wuschner<sup>\*1</sup>, Eric M. Wallat<sup>1</sup>, Mattison J. Flakus<sup>1</sup>,  
Dhanansayan Shanmuganayagam<sup>1</sup>, Jennifer Meudt<sup>1</sup>, Gary E.  
Christensen<sup>2</sup>, Joseph M. Reinhardt<sup>2</sup>, Jessica R. Miller<sup>1</sup>, Michael J.  
Lawless<sup>1</sup>, Andrew M. Baschnagel<sup>1</sup>, John E. Bayouth<sup>1</sup>

(1) University of Wisconsin-Madison, Madison, WI, (2) University of Iowa, Iowa City, IA

Version typeset March 16, 2021

Author to whom correspondence should be addressed. email: wuschner@wisc.edu

## Abstract

**Purpose:** To analyze radiation induced changes in Hounsfield units and determine their correlation with changes in perfusion and ventilation. Additionally, to compare the post-RT changes in human subjects to those measured in a swine model used to quantify perfusion changes, and validate their use as a preclinical model.

**Methods:** A cohort of 5 Wisconsin Miniature Swine (WMS<sup>TM</sup>) were studied. Additionally, 19 human subjects were recruited as part of an IRB approved clinical trial studying functional avoidance radiation therapy for lung cancer and were treated with SBRT. Imaging (a contrast enhanced dynamic perfusion CT in the swine and 4DCT in the humans) was performed prior to and post-RT. Jacobian elasticity maps were calculated on all 4DCT images. Contours were created from the isodose lines to discretize analysis into 10 Gy dose bins. B-spline deformable image registration allowed for voxel-by-voxel comparative analysis in these contours between timepoints. The WMS<sup>TM</sup> underwent a research course of 60 Gy in 5 fractions delivered locally to a target in the lung using an MRI-LINAC system. In the WMS<sup>TM</sup> subjects, the dose-bin contours were copied onto the contralateral lung, which received < 5 Gy for comparison. Changes in HU and changes in Jacobian were analyzed in these contours.

**Results:** Statistically significant ( $p < 0.05$ ) changes in the mean HU value post-RT compared to pre-RT were observed in both the human and WMS<sup>TM</sup> groups at all timepoints analyzed. The HU increased linearly with dose for both groups. Strong linear correlation was observed between the changes seen in the swine and humans (Pearson coefficient  $> 0.97$ ,  $p < 0.05$ ) at all timepoints. Changes seen in the swine closely modeled the changes seen in the humans at 12 months post RT (slope=0.95). Jacobian analysis showed between 30-60% of voxels were damaged post-RT. Perfusion analysis in the swine showed a statistically significant ( $p < 0.05$ ) reduction in contrast inside the vasculature 3 months post-RT compared to pre-RT. The increases in contrast outside the vasculature was strongly correlated (Pearson Correlation 0.88) with the reduction in HU inside the vasculature but were not correlated with the changes in Jacobians.

**Conclusions:** Radiation induces changes in pulmonary anatomy at 3 months post-RT, with a strong linear correlation with dose. The change in HU seen in the non-vessel

41 lung parenchyma suggests this metric is a potential biomarker for change in perfusion.  
42 Finally, this work suggests that the WMS<sup>TM</sup> swine model is a promising pre-clinical  
43 model for analyzing radiation-induced changes in humans and poses several benefits  
44 over conventional swine models.

45

# 46 Contents

47	<b>I. Introduction</b>	<b>1</b>
48	<b>II. Methods</b>	<b>3</b>
49	II.A. Human Patient Dataset . . . . .	3
50	II.B. Animal Model . . . . .	3
51	II.B.1. Measurement Setup . . . . .	3
52	II.B.2. Description of Animal Model . . . . .	5
53	II.B.3. Irradiation Scheme for WMS <sup>TM</sup> . . . . .	5
54	II.B.4. CT Acquisition . . . . .	6
55	II.B.5. Subtracting Vasculature from the dynamic 4DCTs . . . . .	6
56	II.B.6. Measuring change in HU Value in the swine . . . . .	7
57	II.C. Measuring change in HU Value in the Human Subjects . . . . .	9
58	II.D. Assessing Correlation between the WMS <sup>TM</sup> and Humans . . . . .	9
59	II.E. Ventilation Change Measurement via Jacobian Analysis . . . . .	10
60	II.F. Statistical Analysis . . . . .	11
61	<b>III. Results</b>	<b>11</b>
62	III.A. Observations of HU Change . . . . .	11
63	III.A.1. Swine Subjects . . . . .	11
64	III.A.2. SBRT Human Subjects . . . . .	12
65	III.A.3. Correlations Between Swine and Humans . . . . .	12
66	III.B. Change in Jacobian Results . . . . .	12
67	III.C. Correlation between change in Jacobian and change in HU . . . . .	13
68	<b>IV. Discussion</b>	<b>17</b>
69	IV.A. Observed Radiation Induced Changes . . . . .	17
70	IV.A.1. Observed Increases in HU Post-RT outside vasculature . . . . .	17
71	IV.A.2. Correlation Between HU Changes in and out of Vasculature . . . . .	17
72	IV.A.3. Interpretation of Jacobian results . . . . .	19
73	IV.B. Correlations between animal model and humans . . . . .	20

74	IV.C. Clinical Impact . . . . .	20
75	<b>V. Conclusion</b>	<b>21</b>
76	<b>VI. Additional Information</b>	<b>22</b>
77	VI.A. Competing Interests . . . . .	22
78	VI.B. Acknowledgements . . . . .	22
79	<b>References</b>	<b>23</b>
80	<b>VII. Author Contribution Statement</b>	<b>28</b>
81	<b>VIII. Legends</b>	<b>30</b>

## 1. Introduction

Functional avoidance radiation therapy has been increasingly utilized for subjects being treated for lung cancer. To do this, accurate modeling of the normal tissue complications of thoracic radiation, local functional mapping, or dose-response modeling is needed to define treatment parameters for a given patient. Previous work has indicated that radiation pneumonitis and radiation fibrosis are the primary toxicities following thoracic radiation in lung cancer patients<sup>1234</sup>. In addition, these complications can be predicted using a linear quadratic model<sup>561234</sup>. In recent years, multiple studies have looked at using functional metrics to create risk assessments and have found that the inclusion of these metrics increased the predictive power of those models<sup>173458910</sup>.

Several groups have used four dimensional computed tomography (4DCT) based ventilation metrics to estimate lung function<sup>11121314</sup>. This methodology uses anatomical changes between breathing phases to assess how the lungs expand and contract regionally. These metrics show changes with radiation dose and are surrogates for pulmonary function. Multiple groups have used (or are currently using) these metrics in clinical trials to assess the efficacy of using these metrics for functional avoidance in treatment planning.<sup>45151617</sup>. However, the majority of these models focus on ventilation estimates and do not account for damage due to fibrosis, inflammation, or other physiological responses that may not be represented by 4DCT ventilation measures. In particular, changes in perfusion are often excluded from these functional avoidance studies which is a crucial component to the lung's ability to function. In a functional avoidance review, Ireland et al cited that perfusion may be more clinically relevant when performing functional avoidance than ventilation.<sup>15</sup>. Therefore, additional metrics are needed to assess this damage. One potential imaging metric is the change in Hounsfield Units (HU) prior to and post radiation therapy (RT). There have been several studies that have investigated using tissue density changes to assess lung damage<sup>7181920212214232425</sup>. Additionally, several studies have correlated these changes with radiation dose delivered<sup>718192021221423242526</sup>.

Previous work in the CT-Ventilation space has derived lung function directly using Hounsfield unit values of the time-averaged 4DCT<sup>27</sup>. An alternate approach uses the Jaco-

113 bian determinant of the transformation computed from image registration<sup>28</sup>. This method  
114 assumes that the expansion of a voxel is caused by the addition of air from ventilation and  
115 is the method used in this work. This method was also shown to yield the highest DICE  
116 similarity coefficients and voxel-wise and ROI-based Spearman correlations with ventilation  
117 maps derived from <sup>68</sup>Ga-aerosol PET and <sup>3</sup>He-MRI static ventilation maps, placing first in  
118 the 2019 AAPM Grand Challenge. In this work we investigate the changes in HU post-RT.  
119 We assess if correlations exist between changes in HU and dose delivered or Jacobian ven-  
120 tilation metrics. Additionally, we investigate changes in HU in different regions of anatomy  
121 (inside and outside vessels) and assess the relationship between changes in these different  
122 anatomical regions. This leads to further analysis of radiation induced changes in perfusion  
123 and lung injury.

124

125 A current challenge of validating imaging biomarkers is finding proper surrogates for  
126 human lung function. These surrogates allow one to investigate the physiological significance  
127 of the biomarkers in addition to testing potential remedies. The use of swine in radiotherapy  
128 research has become increasingly prevalent due to the swine's similar physiology to humans. In  
129 the past decade, extensive work has been done to validate the use of swine to model human  
130 physiology in a variety of applications<sup>29 30 31 32 33</sup>. However, previous work has used conven-  
131 tional swine which presents barriers in analysis due to their larger size and growth rates. In  
132 this work, we present a novel swine model, the Wisconsin Miniature-Swine (WMS)<sup>TM</sup> devel-  
133 oped at The University of Wisconsin-Madison. These swine were genetically engineered to  
134 present several benefits to conventional swine models. We believe these benefits allow our  
135 novel swine model to better predict these changes post-RT than conventional swine used  
136 historically.

137

138 This work provides a dose response analysis of lung density changes and Jacobian venti-  
139 lation changes in human subjects enrolled in a prospective IRB-approved clinical trial. The  
140 lung density analysis outside the vasculature is also compared to the humans to validate the  
141 swine's response to radiation therapy such that they can be used as a preclinical model for  
142 future analysis.

## 143 **II. Methods**

### 144 **II.A. Human Patient Dataset**

145 Nineteen human subjects from a prospective trial at UW-Madison (NCT02843568) investi-  
146 gating the use of 4DCT-based ventilation in functional planning were used. The patient co-  
147 hort included non-small cell lung cancer patients undergoing radiation therapy using Stereo-  
148 tactic body radiation therapy (SBRT). Per trial protocol, a set of two 4DCT datasets,  
149 acquired 5 minutes apart, were obtained before RT and at 3, 6, and 12 months post RT  
150 for each subject. During this analysis, audio coaching was used in an effort to normalize  
151 breathing patterns across subjects and reduce experimental variance. Informed consent was  
152 obtained from all participants and a summary of patient demographics is provided In Table  
153 1. All human clinical trial procedures were approved by the University of Wisconsin Health  
154 Sciences Institutional Review Board (IRB) to ensure compliance with federal and ethical  
155 guidelines.

156

### 157 **II.B. Animal Model**

#### 158 **II.B.1. Measurement Setup**

159 All Wisconsin Miniature Swine (WMS)<sup>TM</sup> subjects were ventilated to a consistent tidal vol-  
160 ume (1 L was chosen to match the average tidal volume of the human subject populations).  
161 This allowed for repeatable breathing patterns in addition to assuring a fixed breath hold  
162 which is difficult to achieve with human subjects. The subjects were sedated which elim-  
163 inated their susceptibility to motion artifacts in the scans and uncertainty due to patient  
164 motion in the radiation delivery. Since we were not treating a disease in the swine, we were  
165 able to design the dose distribution such that the contralateral lung was left un-irradiated for  
166 comparative analysis. Details regarding animal care and drugs administered can be found in  
167 the supplementary material. All procedures as well as animal care practices were approved  
168 by the University of Wisconsin Institutional Animal Care and Use Committee (IACUC). The  
169 drugs and methods of anesthesia and euthanasia were approved in compliance with Amer-  
170 ican Veterinary Medical Association (AVMA) guidelines for anaesthesia and euthanasia of  
171 swine.

Table 1: Summary of Human Patient Demographics

Number	19
Sex	
M	12
F	7
Age	
Mean	72
Min	59
Max	85
Prescription	
Rx Dose = 50 Gy	17
Rx Dose = 60 Gy	2
Fractionation	
5 fx	17
15 fx	2
Diagnosis	
adenocarcinoma	11
squamous cell carcinoma	7
endometrial metastases	1
Stage	
I	3
IA	11
IB	4
II	1

172

173 An additional benefit of using the animal model was that we had the ability to perform  
174 scans that would not typically be given to human subjects to avoid giving excess dose. This  
175 included a dynamic contrast enhanced scan.

### 176 II.B.2. Description of Animal Model

177 The Wisconsin Miniature Swine (WMS)<sup>TM</sup> possess several characteristics that make them  
178 an ideal model. In general, swine are well suited for biomedical studies pertaining to the  
179 development/validation of diagnostic and therapeutic technologies because of their genetic  
180 proximity to humans and similarities in anatomy<sup>29 31 32 33</sup>. The WMS<sup>TM</sup> were created by se-  
181 lective crossbreeding of several swine breeds such that their weight, size, and physiology are  
182 similar to humans and their body composition can be easily manipulated<sup>30</sup>. As they can be  
183 easily maintained at human size for any length of time, they will remain the same size from  
184 intervention to necropsy. In addition, we were able to select swine that had lung volumes  
185 that were within the range of those in our human subjects.

186

187 The WMS at the size used in our study had lungs that matched human adult lungs.  
188 Additionally, they were swine in their early adulthood (approximately equivalent to a human  
189 in their late 20s or early 30s). If we had used a conventional breed of swine as previous studies  
190 have done, the swine would have been approximately 3 months of age (in order to match the  
191 size of human lungs). Given that swine reach sexual maturity at 5 months of age, a 3-month  
192 old swine is equivalent to a pre-pubescent human child at 6-8 years of age. A conventional  
193 swine at this age has a rate of development where tissue remodeling and size changes are very  
194 rapid. The swine's ability to heal and response to radiation damage (i.e., pathophysiology)  
195 would not mimic that of a human adult. The WMS on the other hand allowed us to more  
196 closely model the pathophysiology observed in a human adult. A 14 month old WMS is  
197 close to a human in the mid-late twenties<sup>30</sup>.

### 198 II.B.3. Irradiation Scheme for WMS<sup>TM</sup>

199 Five WMS<sup>TM</sup> (14.4 ± 1.7 months old) were given a research course of 60 Gy in 5 fractions  
200 approved by the an Institutional Animal Care and Use Committee (IACUC). These fractions

201 were delivered following a standard clinical SBRT schedule receiving fractions with a day  
202 in between each delivery during weekdays and 2 days over the weekend. The PTV was  
203 designated as the bifurcation of a vessel in one of the lungs. Treatment delivery was done  
204 using an MRI-guided LINAC system.

#### 205 **II.B.4. CT Acquisition**

206 All human and swine CT images were acquired on a Siemens SOMATOM Definition Edge  
207 CT scanner. Each swine underwent six total imaging sessions (one session before each of  
208 the five fraction deliveries and one 3 months post-RT). In each session, a contrast enhanced  
209 dynamic 4DCT image was obtained. Details regarding the dynamic 4DCT procedure can  
210 be found in the supplementary material.

#### 211 **II.B.5. Subtracting Vasculature from the dynamic 4DCTs**

212 Vasculature was masked out prior to analysis in the WMS<sup>TM</sup> swine to enable the analysis of  
213 the non-vessel lung parenchyma and the vessels separately. We used the HU values in the  
214 dynamic 4DCTs to indicate regions where contrast was present. These regions were seg-  
215 mented as vessels based on the assumption that the presence of contrast indicates a location  
216 through which blood is flowing. The dynamic 4DCT images contain between 28 and 36  
217 frames. In the initial frames, contrast was not present as acquisition began before contrast  
218 was injected. After the acquisition of 4 frames, contrast injection began. As acquisitions  
219 continued after injection, the mean HU value in a vessel increased as contrast flowed in and  
220 decreased as contrast flowed out (See Figure 1).

221

222 Segmentation was performed on a maximum intensity projection (MIP) image of all  
223 frames to account for the timing offset of contrast flowing through different vessels. All  
224 frames were deformably registered to the first frame using the built in registration algorithm  
225 in MIM Software (Cleveland, OH) prior to MIP generation.

226

227 Next, a HU threshold was utilized to perform vessel segmentation. To determine this  
228 threshold, a histogram of all pixel values over the lung mask in the first frame of the dynamic  
229 4DCT (a frame where contrast was not present) was plotted. This histogram yielded a

230 bimodal plot as shown in Figure 1c. The threshold was set at 2 standard deviations above  
 231 the mean of the second mode. This was done such that vessels would only be segmented  
 232 if they experienced an increase in HU due to the presence of iodine contrast flow and not  
 233 due to other structures in the lung that appear with a moderately elevated HU value. The  
 234 standard deviation was computed using the full width at half maximum of the higher mode  
 235 peak using the relation below:

$$FWHM = 2.35\sigma \tag{1}$$

237 Finally, to segment the vasculature, a threshold was applied such that any voxel within the  
 238 lung mask having a HU above the threshold was segmented. An example scan showing the  
 239 segmented vasculature is shown in Figure 1d. Once the vasculature was segmented, another  
 240 contour with this vasculature subtracted was created to analyze regions outside the vascu-  
 241 lature as well.

242

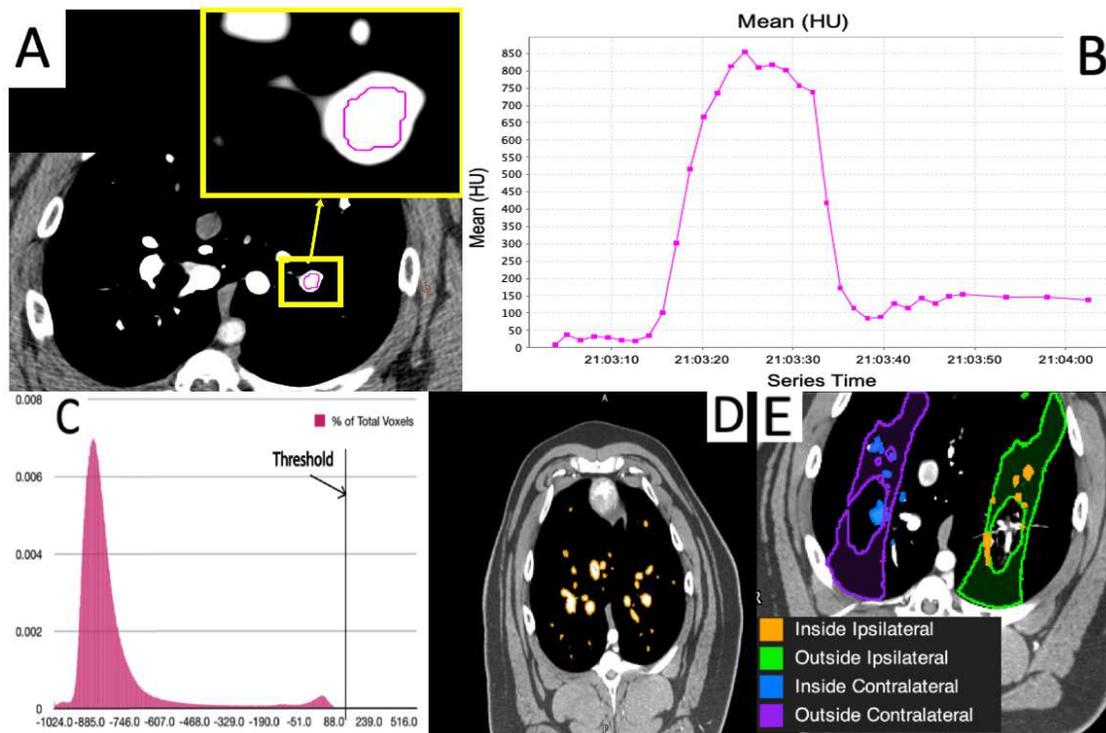


Figure 1:

243 II.B.6. Measuring change in HU Value in the swine

244 All post-RT scans were deformably registered to the pre-RT scan using a B-spline registra-  
 245 tion algorithm to allow for voxel-wise comparisons<sup>34 35</sup>. Analysis was performed in 10 Gy

246 dose bins. This was done by importing the planned dose distributions for each patient into  
247 MIM and creating contours from the isodose lines. The dose distribution was registered to  
248 CT scan used for treatment planning. All dose contours were copied onto the contralat-  
249 eral lung as shown in Figure 3c. Figure 1e shows an example of the 4 contours analyzed  
250 for each dose bin. The choice to use 10 Gy dose bins was determined through analysis of  
251 the variation in measurement as a function of the volume of the contour analyzed. As the  
252 volume of the contour analyzed decreases, the measurement becomes increasingly noisy. In  
253 SBRT treatments, high dose gradients are present thus the volume receiving a given dose is  
254 small. Our analysis found that for volumes greater than 30 cc we were able to minimize the  
255 standard deviation to below 25 % at all dose levels. This resulted in the bins being 10 Gy  
256 wide.

257

258 Figure 2 shows the two measurements that were obtained for this analysis. The mean  
259 HU value trace for the entire lung mask excluding the vasculature for one swine subject is  
260 plotted. Observe in 2b that the initial baseline values prior to contrast injection increase  
261 post-RT from the pre-RT value shown in 2a in the ipsilateral lung, but remain unchanged  
262 in the contralateral unirradiated lung.

263

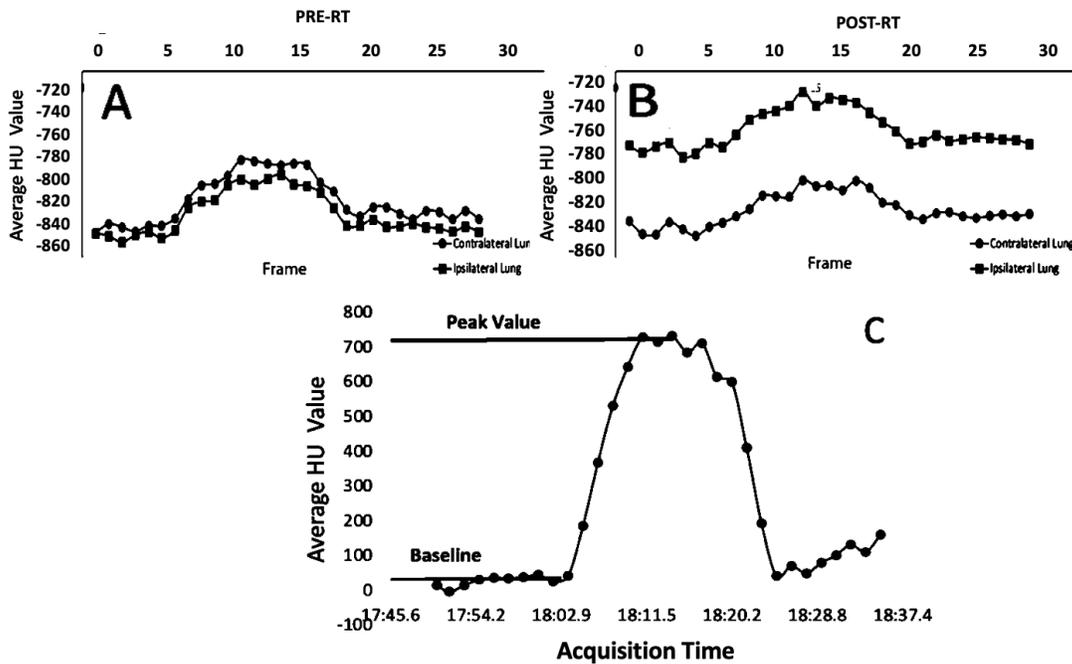


Figure 2:

264 The baseline measurement was the first measurement taken. For all swine, the first  
 265 frame of the Dynamic 4DCTs was used to obtain this value in both the contours where  
 266 vessels were subtracted as well as the contours of just the segmented vessels for each dose  
 267 bin (following the procedures above). The second measurement was the max HU value of  
 268 the trace which was taken only for the contours of the segmented vessels for each dose bin.  
 269 These measurements were recorded at pre and 3 months post-RT. The percent difference  
 270 was calculated using:

$$271 \quad \Delta HU(\%) = \frac{HU_{post} - HU_{pre}}{HU_{pre}} * 100\% \quad (2)$$

## 272 II.C. Measuring change in HU Value in the Human Subjects

273 The human subject analysis was similar to that of the swine with a few exceptions. Since the  
 274 treatment planning was not designed to reduce radiation dose below 5 Gy in the contralateral  
 275 lung, it was unable to be used as an "unirradiated" control. Additionally, since the human  
 276 subjects did not receive a contrast enhanced scan, vessel subtraction was not performed  
 277 and only measurements of the HU in regions of tissue that were delivered a given dose  
 278 were obtained. The HU analysis was performed on the exhalation phase of the 4DCT data  
 279 sets. This phase was chosen because it was believed to be the phase least susceptible to CT  
 280 artifacts. Since the human subjects received two 4DCTs at the same timepoint, we computed  
 281 the average of the results of the mean HU values found in each dose contour between the  
 282 two scans.

## 283 II.D. Assessing Correlation between the WMS<sup>TM</sup> and Humans

284 In an effort to account for differences in fractionation scheme, all plots and linear regressions  
 285 were created using equivalent dose in 2 Gy fractions (EQD2) as calculated below where D is  
 286 the total prescribed dose, and d is the dose per fraction. An alpha/beta ratio of 3 was used  
 287 for lung tissue<sup>36</sup>.

$$288 \quad EQD2 = D * \frac{d + \frac{\alpha}{\beta}}{2 + \frac{\alpha}{\beta}} \quad (3)$$

289 As shown in the dose distributions for an example patient in both the swine and humans in  
 290 Figures 3c and 3d, the volume of tissue irradiated was similar in both subject pools. However,  
 291 while both subject pools received SBRT-type plans and has similar volumes irradiated, the

292 human and swine populations received slightly different fractionation schemes (50 Gy in 5 fx  
 293 in the swine vs. 60 Gy in 5 fx in the humans). When calculating correlation coefficients to  
 294 compare the swine and human data, an adjusted data set was created such that there were  
 295 data points for the same EQD2 values in each human population. This was done using the  
 296 linear regression line of best fit equation on the swine data to calculate the percent changes in  
 297 lung density at the EQD2 values recorded in the human subjects. Since the linear regression  
 298 fit was so strongly correlated ( $R^2 = 0.987$ ) we expect the error resulting from doing this  
 299 to be negligible. These adjusted data sets were used to calculate the Pearson correlation  
 300 coefficients for the swine and the human subjects.

## 301 II.E. Ventilation Change Measurement via Jacobian Analysis

302 4DCT can be used to compute a surrogate for regional ventilation of lung tissue and provide  
 303 a spatial map of the local lung tissue expansion and contraction using Jacobian analysis.<sup>6</sup> In  
 304 this work, Jacobian analysis was used as a surrogate to assess change in ventilation in the hu-  
 305 man subjects. The Jacobian values were calculated on all images following the methodology  
 306 described in Patton et al<sup>6</sup>. A Jacobian value equal to one represents no local volume change,  
 307 greater than one represents expansion, and less than one represents contraction of the lung  
 308 tissue. Equivalent tidal volumes (ETV) were used to select the phases of the 4DCTs used  
 309 for the inspiration and expiration scans. This has been shown to increase the repeatability  
 310 of the ventilation measurement and isolate the effect of radiation in longitudinal changes<sup>37</sup>.

311

312 To assess the relative change in ventilation at different timpoints post-RT, the Jacobian  
 313 ratio was calculated using the relation:

$$314 \quad \text{Ratio} = \frac{J_{post}}{J_{pre}} \quad (4)$$

315 A Jacobian ratio of 1 indicates the voxel did not change from its pre-RT elasticity, a ratio of  
 316 less than one indicates the voxel was less elastic post-RT compared to pre-RT (loss of venti-  
 317 lation), and a ratio of greater than one indicates a more elastic voxel (improved ventilation)  
 318 post-RT. For this work, a threshold of a 0.95 Jacobian ratio was used to determine the voxels  
 319 that were considered damaged post-RT. This threshold was used in the works of Patton et.  
 320 al and Wallet et al and was chosen for consistency. Additionally, it is the threshold used  
 321 for evaluation of damage in the clinical trial the human patients were taken from.<sup>6,38</sup> Dose

322 binned analysis was performed using the same contours created for the HU analysis. In each  
323 dose bin the percent of voxels with a Jacobian ratio value of 0.95 or lower was calculated.

## 324 II.F. Statistical Analysis

325 Student paired two-tailed t-tests were used to compare the pre and post-RT HU values. A  
326 Kolmogorov-Smirnov Test was used to verify normality.<sup>39</sup> Correlations between the WMS™  
327 and human subjects as well as all percent increases with dose were assessed using Pearson  
328 correlation coefficients. Additionally, all linear fits to data were evaluated using the coeffi-  
329 cient of determination,  $R^2$ . To correct for multiple comparisons, Bonferroni adjustment was  
330 used. This method of adjustment is the most conservative of adjustments and yielded an  
331 adjusted p-value threshold of 0.01 for statistical significance at  $\alpha = 0.05$  level. All results  
332 were still statistically significant under these adjusted criteria.<sup>40</sup>

## 333 III. Results

### 334 III.A. Observations of HU Change

#### 335 III.A.1. Swine Subjects

336 Statistically significant increases in the mean HU value were seen in each dose bin of the swine  
337 in the non-vessel lung parenchyma. The percent increases showed a linear dependence with  
338 the EQD2 dose delivered with Pearson coefficient of 0.989 ( $p=0.001$ ). The linear regression  
339 fit had a slope of 0.167 %/Gy. In the contours copied onto the unirradiated lung, there was  
340 no statistically significant changes seen in the mean HU values. The percent increases of all  
341 contours are shown in Figure 3a.

342 The heat map shown in Figure 4a shows the percent change in HU for each voxel of the  
343 post-RT scan as compared to the pre-RT scan. Below the heat map is the dose distribution  
344 that was delivered to the subject. It can be seen that the large increases in HU are localized  
345 to irradiated regions.

346  
347 Additionally, statistically significant decreases ( $p < 0.05$ ) in the peak mean HU were  
348 seen in each dose bin of the swine in the segmented vasculature. The percent changes in

349 HU outside the vessels were strongly correlated to the percent change in peak HU inside the  
350 vessels at 3 months post RT as shown in Figure 5.

### 351 III.A.2. SBRT Human Subjects

352 Table 1 of the supplemental material and Figure 3b show statistically significant increases in  
353 all dose bins except 5-15 Gy (EQD2 = 13-39 Gy) for the human subjects. Table 1 lists the  
354 volume of each contour. The increases in HU were linearly correlated with dose delivered.  
355 Pearson correlation coefficients were 0.973, 0.899, and 0.844 for 3,6, and 12-months post-RT,  
356 respectively, with p-values all less than 0.00001.

357

358 The heat map shown in Figure 4b shows the percent change in HU for each voxel of  
359 the 3 months post-RT scan compared to the pre-RT scan for a single subject. Below the  
360 heat map is the dose distribution that was delivered to the subject. Notice that the large  
361 increases in HU are localized to irradiated regions.

362

### 363 III.A.3. Correlations Between Swine and Humans

364 Figures 6a and 6b shows the post-RT percent changes in mean HU value in the human  
365 subjects plotted vs. the percent change seen in the swine for the same dose bin at 3-months  
366 post-RT. Data was not obtained beyond 3 months post-RT in the swine and the swine data  
367 plotted is the adjusted data set derived from the linear best fit equation to match the EQD2  
368 values measured in the humans. The humans show strong linear correlation with the swine  
369 data. The Pearson coefficients for the humans were 0.973,0.899, and 0.844 ( $p<0.05$ ) and had  
370 linear fits had slopes of 0.785, 0.588, and 0.955 % change in human per % change in swine  
371 at 3, 6, and 12 months, respectively.

372

### 373 III.B. Change in Jacobian Results

374 Figures 7 shows the percent of damaged voxels in each dose bin for the humans at 3, 6, and  
375 12 months post-RT. The percent of damaged voxels increases as the dose delivered increases

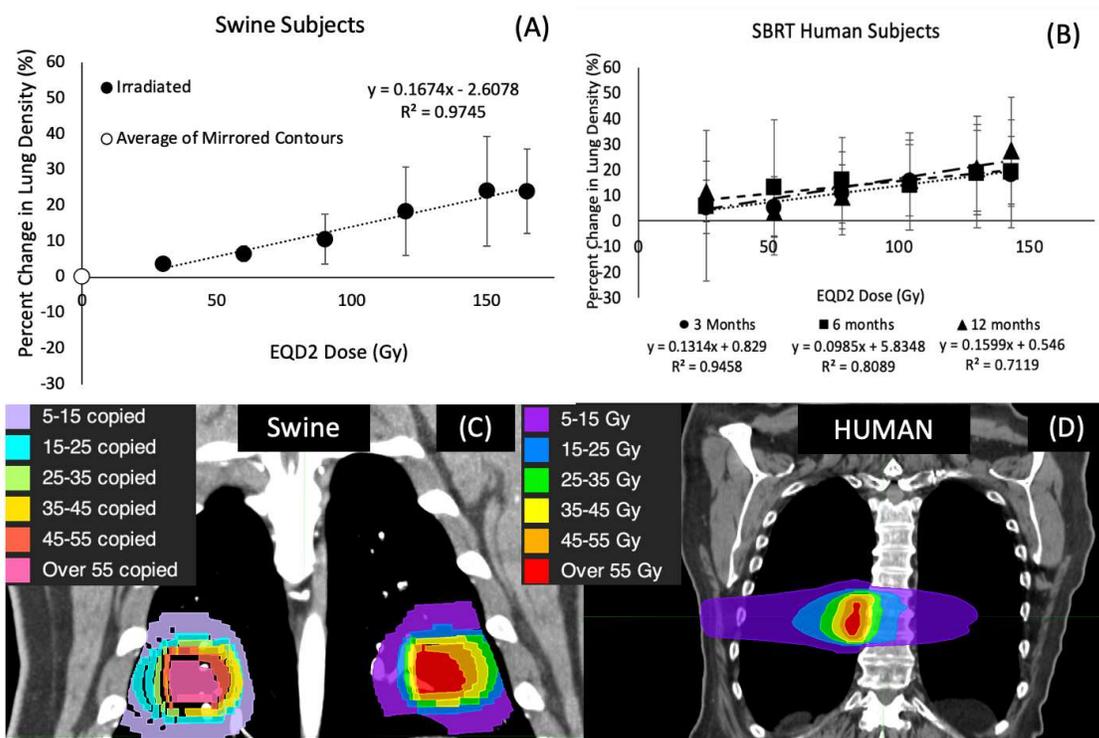


Figure 3:

376 and is correlated linearly at all timepoints. Additionally of note, there was little difference  
 377 between the datasets at the different timepoints in SBRT subjects.

### 378 III.C. Correlation between change in Jacobian and change in HU

379 Figure 8 shows the percent of damaged voxels plotted against the percent increase in HU  
 380 observed for the humans at the 3 and 12 month timepoints. Each point on the plot represents  
 381 a different dose bin analyzed. While strong correlation exists at both timepoints, the slope  
 382 of the linear fit is very low indicating that the changes in HU observed are not indicative of  
 383 the ventilation changes represented by the change in Jacobian.

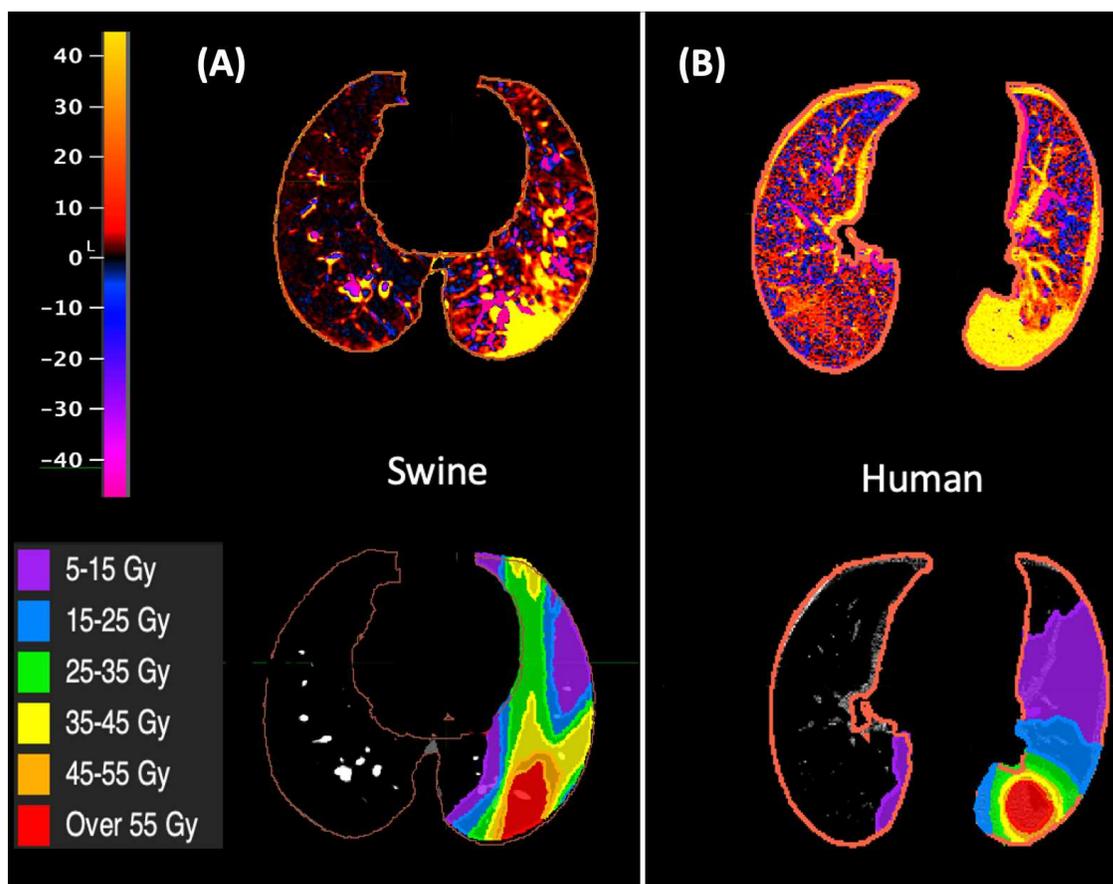


Figure 4:

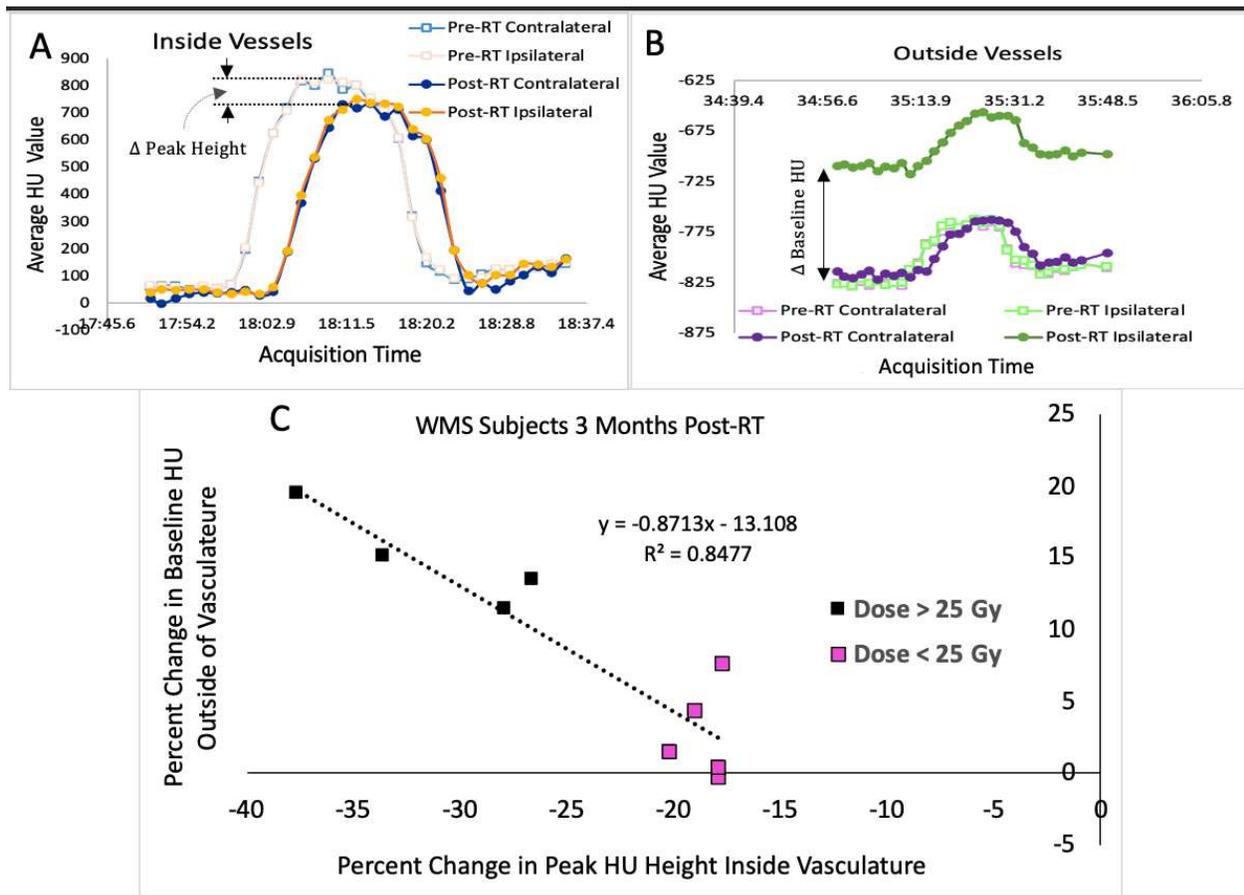


Figure 5:

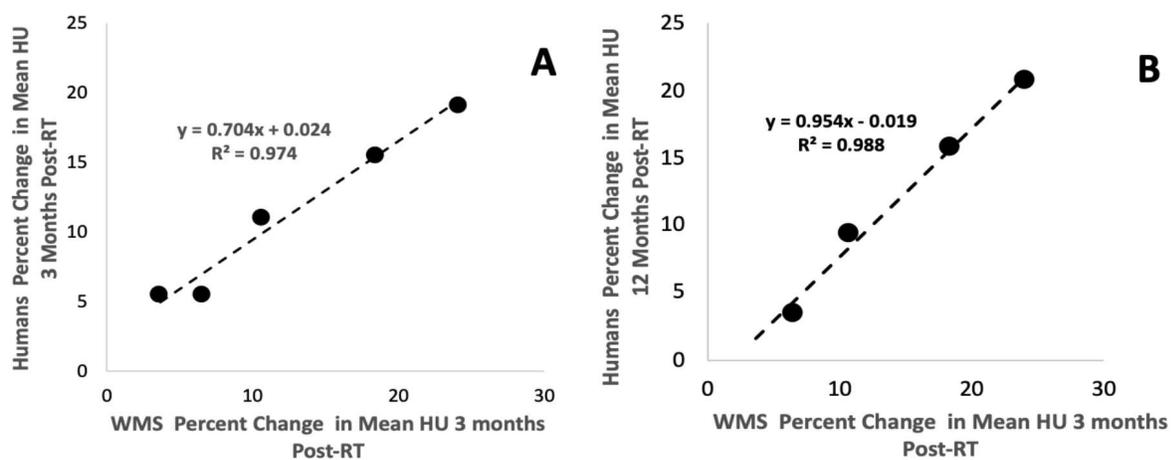


Figure 6:

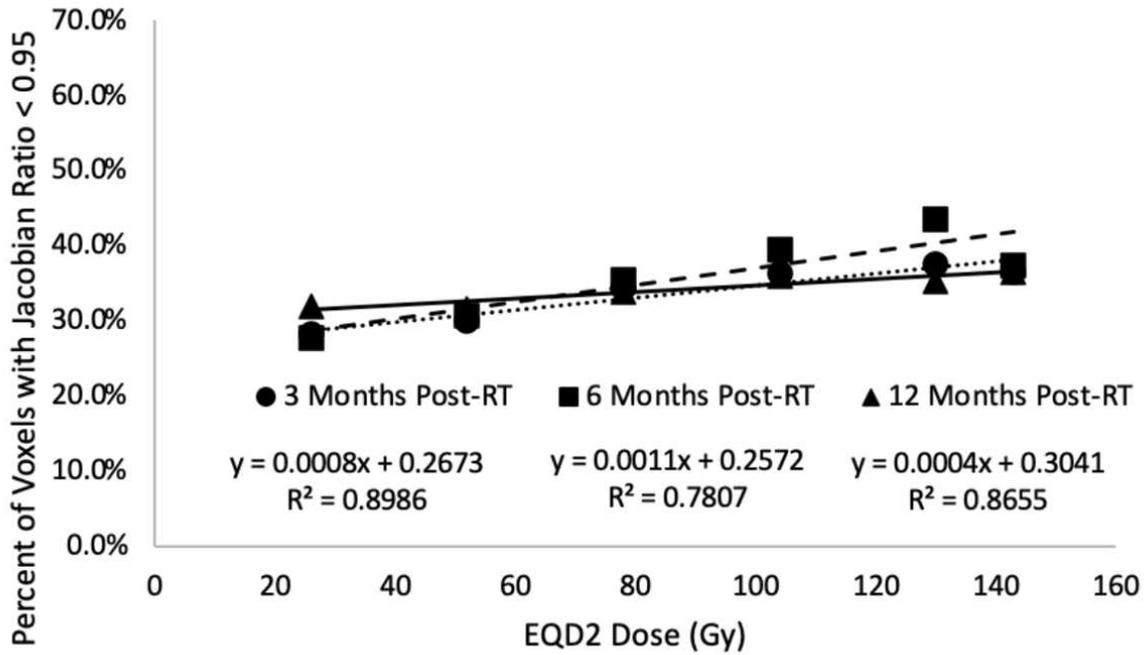


Figure 7:

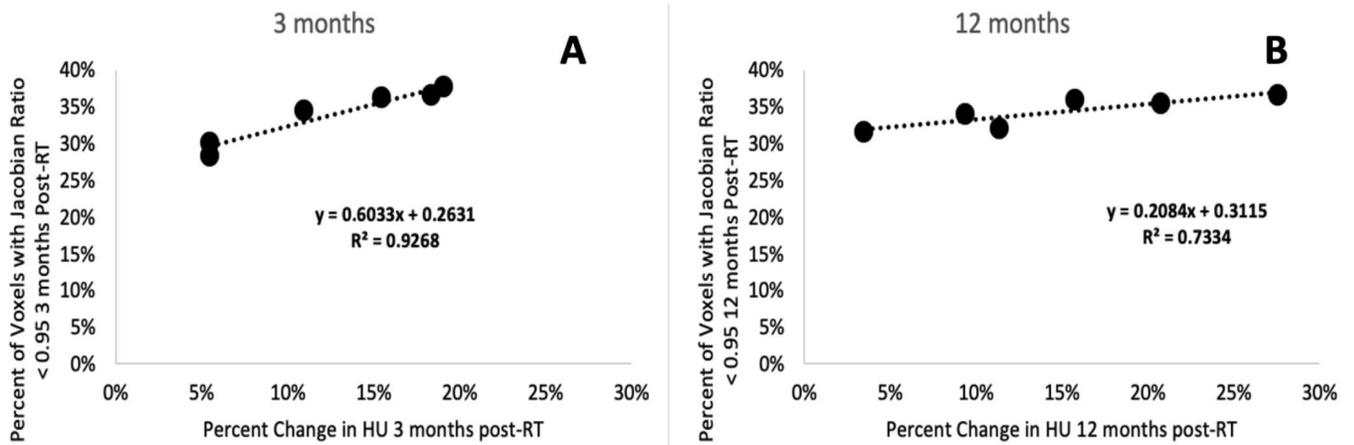


Figure 8:

## 384 **IV. Discussion**

### 385 **IV.A. Observed Radiation Induced Changes**

#### 386 **IV.A.1. Observed Increases in HU Post-RT outside vasculature**

387 Statistically significant increases in the mean HU value were observed in both the human  
388 subject and WMS groups 3 months post-RT. Increases ranged from 3.6% in the lowest dose  
389 bin to 30.0% at the largest doses and increased linearly. In regions receiving less than 5  
390 Gy in the swine, no statistically significant changes in HU were observed indicating changes  
391 were radiation-induced.

392 The percent increases in HU increased linearly with dose at all time points for the human  
393 subjects. We believe these changes model a combination of an inflammatory response that  
394 resolves with time combined with longer lasting damage that remains. The proportion of  
395 each effect's contribution cannot be distinguished from these measurements alone. Examples  
396 of longer lasting damage are pneumonitis which begins to develop 2-4 months post-RT, and  
397 fibrotic tissue which typically can develop 4-6 months post-RT<sup>1</sup>.

398 All heat maps showing the percent changes in HU indicated the changes in HU value are  
399 localized to regions receiving dose. In addition, the copied contours in unirradiated sections  
400 of lung in the swine showed no statistically significant differences post-RT. This indicates  
401 the changes we are measuring were radiation-induced. It is worth noting that by using the  
402 vessel subtraction method we chose, it is possible that the voxels at the edge of the vessel  
403 were excluded from the subtraction and thus analyzed in the regions classified as "non-vessel  
404 lung parenchyma." This is due to the increased HU inside the vessel from contrast being  
405 present setting the threshold for vessel classification higher. However, these voxels are not a  
406 significant contribution to our analysis when averaged with all the additional voxels in the  
407 contour. This is proven in the contralateral lung contours where vessels were subtracted in  
408 the same manner and yet we saw less than a 0.5 % increase in HU.

#### 409 **IV.A.2. Correlation Between HU Changes in and out of Vasculature**

410 A full understanding of the physiological processes the change in HU represents has yet to  
411 be developed. From our results looking at the change in HU inside and outside of vessels,

412 we speculate that part of the physiological response modeled by these changes in HU is a  
413 radiation-induced loss of perfusion. Radiation-induced changes in perfusion have been shown  
414 by other groups via PET and dual-energy CT (DECT) methods<sup>10 8 41 42</sup>. Additionally, func-  
415 tional avoidance trials have shown there is a benefit to avoiding highly perfusing regions.<sup>26</sup>  
416 The correlation of the increases in HU outside the vasculature combined with the decrease  
417 in mean peak HU in the regions of vasculature suggest that some of the HU increase is due  
418 to radiation induced vasculature leakage of contrast. While this does not directly confirm  
419 change in perfusion, a change that would have to be seen at the capillary level, our hypothesis  
420 is that this can be inferred from these results since the reduction in HU inside the vasculature  
421 would indicate a loss of contrast (a high HU material) and the corresponding raise in HU  
422 outside these vessels would explain where the contrast in addition to blood (another higher  
423 HU material) is going.

424 The reduction in contrast inside the vasculature was systemic and occurred in both  
425 irradiated and non-irradiated regions. However, the increases post-RT were only in the  
426 irradiated regions. This supports our hypothesis since leakage of contrast would result in  
427 less contrast circulating through all vasculature (showing a reduction in peak HU height),  
428 but the increase in HU due to contrast leakage would only be seen where the source of  
429 the leak is (in the irradiated lung tissue). Additionally, this result was observed across all  
430 swine subjects. When analyzing the plot in 5c, it appears as if there is a threshold dose  
431 above which this leakage occurs of 25 Gy. All dose bins plotted that are greater than 25  
432 Gy showed greater than a 10% increase in HU outside the vasculature while dose bins that  
433 were less than 25 Gy showed below 10% increases outside the vasculature. The slope of  
434 curve 5c is -0.87 which is is dominant but not a perfect negative relationship, indicating  
435 additional secondary factor(s) contributing to the increase in HU outside the vasculature.  
436 encompasses this change in perfusion as well as acute damage. As mentioned previously,  
437 there are other mechanisms of damage such as inflammation and presence of fibrotic tissue  
438 that would also contribute towards a raised HU value. Future work will involve acquiring  
439 these perfusion scans on human subjects receiving SBRT, in addition to analyzing damage  
440 to the pulmonary vasculature through post-RT lung pathology of the swine to confirm this  
441 speculation. If verified the change in HU may be a potential bio-marker for radiation-induced  
442 change in perfusion and understanding the dose threshold above which this damage occurs  
443 would provide additional guidance during planning.

#### 444 IV.A.3. Interpretation of Jacobian results

445 Figure 7 indicates that the human subjects showed no difference in the percent of damaged  
446 voxels (as indicated by a 0.95 or lower Jacobian ratio) across the 3 timepoints measured.  
447 Additionally, the percent of damaged voxels (those seeing a 5% or greater reduction in Ja-  
448 cobian) was independent of dose delivered above an EQD2 of 20 Gy.

449

450 One important point that should be considered is that these results encompass sub-  
451 jects pulled from a clinical trial studying the effectiveness of functional avoidance radiation  
452 therapy where highly ventilating regions were avoided. The results are not stratified by  
453 subjects in the control vs the experimental arm. It is possible the subjects in the experi-  
454 mental arm may have experienced different changes in Jacobian post-RT than those in the  
455 control arm since the tissue that was irradiated was selectively optimized to irradiate more  
456 low-functioning tissue.

457

458 As indicated in Figure 8, the percent of voxels showing a decrease in Jacobian were not  
459 correlated with the changes in HU observed with the exception of the 3 months timepoint.  
460 Additionally, while a significant percentage of voxels (greater than 30%) were damaged,  
461 the dose delivered did not heavily influence the number of voxels damaged as previously  
462 described and shown figure 7. However, the dose delivered did heavily influence the changes  
463 in HU. These results indicate that while radiation does induce changes in ventilation (which  
464 has been seen by several other groups as well)<sup>43 44 45</sup>, there are additional radiation induced  
465 changes taking place that are not indicated by the Jacobian metric and these changes are  
466 highly influenced by the magnitude of dose delivered.

467 This observation is important as there are several groups using the difference in HU  
468 between inhale and exhale images to estimate ventilation<sup>43 44 45</sup>. Therefore it would be rea-  
469 sonable to speculate that the changes in HU we see in this work are correlated with and  
470 partially caused by a change in ventilation. However, our results looking at Jacobian-based  
471 ventilation estimates show no correlation with change in HU. This indicates that the cause  
472 for the change in HU involves mechanisms other than ventilation. Combined with our con-  
473 trast data and results discussed previously, this work suggests that, in addition to some  
474 component of fibrotic damage, these increases in HU are representing leaked blood from

475 vasculature and ultimately perfusion change.

## 476 **IV.B. Correlations between animal model and humans**

477 Data was not obtained beyond 3 months post-RT in the WMS™ model. However, the human  
478 data sets showed a strong linear correlation at all timepoints with the WMS data. Addi-  
479 tionally, the linear fit for the 12 month post-RT SBRT human data vs Swine 3 month data  
480 showed a slope of 0.954 indicating that the swine results closely modeled the human results  
481 at 12 months. The linear fit for the 3 month data in the SBRT humans vs the 3 month  
482 swine data had a slope that was less than this (0.704). This indicates that the swine show  
483 an accelerated response to the humans. This accelerated response in the WMS™ model is  
484 consistent with reports in literature of swine models experiencing an expedited biological  
485 response when compared to humans.<sup>29 31 32 33</sup> This feature poses as a potential benefit to  
486 using the swine for pre-clinical modeling as it provides the ability to see response quicker in  
487 the animal model which could provide insight for human study design.

488

489 The six month data is not shown in Figure 6 and did not correlate with the swine data.  
490 We believe this is because at this timepoint there are multiple mechanisms present that are  
491 causing change as described in section IV.A.1. Some of these changes resolve themselves  
492 with time, producing a poor correlation at this timepoint.

## 493 **IV.C. Clinical Impact**

494 This work quantifies an anatomical response to radiation dose in a human population as well  
495 as in an animal model. Our results in the human population looking at changes in HU are  
496 in agreement with the behaviors reported in literature<sup>20 23</sup>. Additionally our results were  
497 part of a prospective clinical trial designed to standardize the acquisition of images for all  
498 subjects and also analyzed data longitudinally over multiple timepoints. We speculate that  
499 the observed increases in HU are indicative of mechanisms. Other groups have demonstrated  
500 radiation-induced changes in perfusion using SPECT<sup>42</sup>. Our findings in the swine indicate  
501 that there may be the ability to infer these changes from 4DCT which would immensely aid  
502 translation to a clinical setting since 4DCTs are already routinely collected for treatment  
503 planning and would not require the acquisition of additional scans. These results would

504 also present an opportunity for superior functional avoidance therapy as other groups have  
505 suggested that perfusion may be more clinically relevant when performing functional avoid-  
506 ance than ventilation<sup>15</sup>. However, further investigation is required to verify and validate  
507 this hypothesis and future work will involve attempting to confirm these speculations using  
508 pathology studies.

509

510 Correlations of the swine and human subjects shows the potential to use the swine as  
511 a pre-clinical model for human response. This work shows strong correlation with human  
512 outcomes and additionally shows accelerated response. This presents a benefit in being able  
513 to expedite the process of assessing response in the swine lungs in order to better design  
514 human clinical trials and investigate of potential treatments and intervention mechanisms.  
515 As mentioned previously, the novel swine used in this study present multiple benefits over  
516 conventional swine. These benefits combined with the results of this study presents these  
517 animals as an improved option for more longitudinal studies due to the swine's ability to  
518 stay the same size and mature similarly to an adult human.

519

520 After the 3-month post-RT scan, the swine lungs were extracted from the animal for  
521 future pathology studies. This work will provide further insight regarding the physiological  
522 response of these subjects and the damage done to the vasculature. Future work will also  
523 include a clinical trial analyzing the response of a larger sample size of these novel swine as  
524 well as an analysis of contrast enhanced scanning on humans. This will enable faster devel-  
525 opment of predictive models that would be able to be validated on existing human subject  
526 data from this trial. From there, clinical trials assessing the effectiveness of intervention  
527 mechanisms on human subjects may be initiated.

## 528 V. Conclusion

529 Radiation induces changes in pulmonary anatomy post-RT. This work quantifies those  
530 changes using a HU analysis and shows that the WMS<sup>TM</sup> model is a good surrogate for  
531 analyzing radiation-induced changes in humans treated with SBRT. The presentation of this  
532 model as a pre-clinical model yields an opportunity to expedite the development of pre-  
533 dictive models as well as human clinical trials to assess various response and intervention

534 mechanisms in human subjects. The observations connecting the changes in HU to changes  
535 in contrast present a potential bio-marker for analyzing functional changes in perfusion that  
536 can be derived off of 4DCT as opposed to requiring additional scans outside of clinical pro-  
537 tocol. This would allow these metrics to be considered in functional avoidance therapy and  
538 could provide a significant benefit to patient outcome.

## 539 **VI. Additional Information**

### 540 **VI.A. Competing Interests**

541 Joseph M. Reinhardt is a shareholder in VIDA Diagnostics, Inc  
542 John E. Bayouth is a co-owner of MR Guidance, LLC (MRG), which has performed business  
543 with vendors of technologies used in this publication.

544 Antonia E. Wuschner, Eric M. Wallet, Mattison J. Flakus, Gary E. Christensen, Jessica  
545 R. Miller, Michael J. Lawless, and Andrew M. Bachnagel have no conflicts of interest to  
546 disclose.

### 547 **VI.B. Acknowledgements**

548 This work was supported by the National Institutes of Health grant CA166703.

549 All figures were produced by Antonia Wuschner. All graphs were made in Microsoft  
550 Excel and Figures 1, 2c, 2d, and 3 were created in MIM Software version 6.6.

## References

- <sup>1</sup> S. McDonald, P. Rubin, T. L. Phillips, F. A. C. R. . Andlawrence, and B. Marks, Special Feature-Late Effects Consensus Conference: Injury to the Lung from Cancer Therapy: Clinical Syndromes, Measurable Endpoints, and Potential Scoring Systems., Technical Report 5.
- <sup>2</sup> L. B. Marks, S. M. Bentzen, J. O. Deasy, F. M. S. Kong, J. D. Bradley, I. S. Vogelius, I. El Naqa, J. L. Hubbs, J. V. Lebesque, R. D. Timmerman, M. K. Martel, and A. Jackson, Radiation Dose-Volume Effects in the Lung, *International Journal of Radiation Oncology Biology Physics* **76** (2010).
- <sup>3</sup> V. Mehta, Radiation pneumonitis and pulmonary fibrosis in non-small-cell lung cancer: Pulmonary function, prediction, and prevention, *International Journal of Radiation Oncology Biology Physics* **63**, 5–24 (2005).
- <sup>4</sup> P. R. Graves, F. Siddiqui, M. S. Anscher, and B. Movsas, Radiation pulmonary toxicity: From mechanisms to management, 2010.
- <sup>5</sup> T. J. Patton, J. E. Bayouth, B. P. Bednarz, S. B. Fain, J. M. Reinhardt, and J. Smilowitz, Quantifying and modeling radiation therapy-induced ventilation changes and investigating the robustness of 4DCT-based functional avoidance, Technical report, 2018.
- <sup>6</sup> T. J. Patton, S. E. Gerard, W. Shao, G. E. Christensen, J. M. Reinhardt, and J. E. Bayouth, Quantifying ventilation change due to radiation therapy using 4DCT Jacobian calculations, *Medical Physics* **45**, 4483–4492 (2018).
- <sup>7</sup> K. Mah and V. Dyk, Quantitative Measurements of Human Lung Density Following Irradiation, *Radiotherapy and Oncology* **11**, 169–179 (1988).
- <sup>8</sup> H. Koike, E. Sueyoshi, I. Sakamoto, and M. Uetani, Quantification of lung perfusion blood volume by dual-energy CT in patients with and without chronic obstructive pulmonary disease, *Journal of the Belgian Society of Radiology* **99**, 62–68 (2015).
- <sup>9</sup> S. R. Hopkins, M. O. Wielpütz, and H. U. Kauczor, Imaging lung perfusion, 2012.
- <sup>10</sup> J. Zhang, J. Ma, S. Zhou, J. L. Hubbs, T. Z. Wong, R. J. Folz, E. S. Evans, R. J. Jaszczak, R. Clough, and L. B. Marks, Radiation-Induced Reductions in Regional Lung

- 580 Perfusion: 0.1-12 Year Data From a Prospective Clinical Study, *International Journal*  
581 *of Radiation Oncology Biology Physics* **76**, 425–432 (2010).
- 582 <sup>11</sup> F. Hegi-Johnson, D. de Ruyscher, P. Keall, L. Hendriks, Y. Vinogradskiy, T. Yamamoto,  
583 B. Tahir, and J. Kipritidis, Imaging of regional ventilation: Is CT ventilation imaging  
584 the answer? A systematic review of the validation data, *Radiotherapy and Oncology*  
585 **137** (2019).
- 586 <sup>12</sup> J. Kipritidis et al., The VAMPIRE challenge: A multi-institutional validation study of  
587 CT ventilation imaging, *Medical Physics* **46** (2019).
- 588 <sup>13</sup> Y. Vinogradskiy, CT-based ventilation imaging in radiation oncology, *BJR—Open* **1**  
589 (2019).
- 590 <sup>14</sup> N. W. Bucknell, N. Hardcastle, M. Bressel, M. S. Hofman, T. Kron, D. Ball, and S. Siva,  
591 Functional lung imaging in radiation therapy for lung cancer: A systematic review and  
592 meta-analysis, 2018.
- 593 <sup>15</sup> R. Ireland, B. Tahir, J. Wild, C. Lee, and M. Hatton, Functional Image-guided Radio-  
594 therapy Planning for Normal Lung Avoidance, *Clinical Oncology* **28** (2016).
- 595 <sup>16</sup> T. Yamamoto, S. Kabus, M. Bal, P. Keall, S. Benedict, and M. Daly, The first patient  
596 treatment of computed tomography ventilation functional image-guided radiotherapy for  
597 lung cancer, *Radiotherapy and Oncology* **118** (2016).
- 598 <sup>17</sup> Y. Vinogradskiy, C. G. Rusthoven, L. Schubert, B. Jones, A. Faught, R. Castillo,  
599 E. Castillo, L. E. Gaspar, J. Kwak, T. Waxweiler, M. Dougherty, D. Gao, C. Stevens,  
600 M. Miften, B. Kavanagh, T. Guerrero, and I. Grills, Interim Analysis of a Two-  
601 Institution, Prospective Clinical Trial of 4DCT-Ventilation-based Functional Avoidance  
602 Radiation Therapy, *International Journal of Radiation Oncology\*Biological\*Physics* **102**  
603 (2018).
- 604 <sup>18</sup> Q. Diot, B. Kavanagh, T. Schefter, L. Gaspar, K. Stuhr, and M. Miften, Regional  
605 normal lung tissue density changes in patients treated with stereotactic body radiation  
606 therapy for lung tumors, *International Journal of Radiation Oncology Biology Physics*  
607 **84**, 1024–1030 (2012).
-

- 608 <sup>19</sup> S. Matsuoka, T. Yamashiro, S. Matsushita, A. Kotoku, A. Fujikawa, K. Yagihashi, and  
609 Y. Nakajima, Quantitative CT evaluation in patients with combined pulmonary fibrosis  
610 and emphysema: Correlation with pulmonary function, *Academic Radiology* **22**, 626–  
611 631 (2015).
- 612 <sup>20</sup> D. A. Palma, J. Van Sörnsen De Koste, W. F. Verbakel, A. Vincent, and S. Senan, Lung  
613 density changes after stereotactic radiotherapy: A quantitative analysis in 50 patients,  
614 *International Journal of Radiation Oncology Biology Physics* **81**, 974–978 (2011).
- 615 <sup>21</sup> S. O'Reilly, V. Jain, Q. Huang, C. Cheng, B. K. K. Teo, L. Yin, M. Zhang, E. Diffender-  
616 fer, T. Li, W. Levin, Y. Xiao, L. Dong, S. Feigenberg, A. T. Berman, and W. Zou, Dose  
617 to Highly Functional Ventilation Zones Improves Prediction of Radiation Pneumonitis  
618 for Proton and Photon Lung Cancer Radiation Therapy, in *International Journal of*  
619 *Radiation Oncology Biology Physics*, Elsevier Inc., 2020.
- 620 <sup>22</sup> Y. Li, M. Dykstra, T. D. Best, J. Pursley, N. Chopra, F. K. Keane, M. J. Khandekar,  
621 G. C. Sharp, H. Paganetti, H. Willers, F. J. Fintelmann, and C. Grassberger, Differential  
622 inflammatory response dynamics in normal lung following stereotactic body radiation  
623 therapy with protons versus photons, *Radiotherapy and Oncology* **136**, 169–175 (2019).
- 624 <sup>23</sup> Q. Diot, L. B. Marks, S. M. Bentzen, S. Senan, B. D. Kavanagh, M. V. Lawrence,  
625 M. Miften, and D. A. Palma, Comparison of radiation-induced normal lung tissue density  
626 changes for patients from multiple institutions receiving conventional or hypofractionated  
627 treatments, *International Journal of Radiation Oncology Biology Physics* **89**, 626–632  
628 (2014).
- 629 <sup>24</sup> K. A. Al Feghali, Q. C. Wu, S. Devpura, C. Liu, A. I. Ghanem, N. W. Wen, M. Ajlouni,  
630 M. J. Simoff, B. Movsas, and I. J. Chetty, Correlation of normal lung density changes  
631 with dose after stereotactic body radiotherapy (SBRT) for early stage lung cancer, *Clinical and Translational Radiation Oncology* **22**, 1–8 (2020).
- 633 <sup>25</sup> B. A. Hoff, E. Pompe, S. Galbán, D. S. Postma, J. W. J. Lammers, N. H. Ten Hacken,  
634 L. Koenderman, T. D. Johnson, S. E. Verleden, P. A. De Jong, F. A. Mohamed Hoesein,  
635 M. Van Den Berge, B. D. Ross, and C. J. Galbán, CT-Based Local Distribution Metric  
636 Improves Characterization of COPD, *Scientific Reports* **7** (2017).

- 637 <sup>26</sup> S. Siva, R. Thomas, J. Callahan, N. Hardcastle, D. Pham, T. Kron, R. J. Hicks, M. P.  
638 MacManus, D. L. Ball, and M. S. Hofman, High-resolution pulmonary ventilation and  
639 perfusion PET/CT allows for functionally adapted intensity modulated radiotherapy in  
640 lung cancer, *Radiotherapy and Oncology* **115** (2015).
- 641 <sup>27</sup> J. Kipritidis, M. S. Hofman, S. Siva, J. Callahan, P.-Y. Le Roux, H. C. Woodruff, W. B.  
642 Counter, and P. J. Keall, Estimating lung ventilation directly from 4D CT Hounsfield  
643 unit values, *Medical Physics* **43**, 33–43 (2015).
- 644 <sup>28</sup> J. M. Reinhardt, K. Ding, K. Cao, G. E. Christensen, E. A. Hoffman, and S. V. Bo-  
645 das, Registration-based estimates of local lung tissue expansion compared to xenon CT  
646 measures of specific ventilation, *Medical Image Analysis* **12**, 752–763 (2008).
- 647 <sup>29</sup> H. F. M. J. W. Hopewell M. Rezvani, The pig as a model for the study of radiation  
648 effects on the lung, *International Journal of Radiation Biology* **76**, 447–452 (2000).
- 649 <sup>30</sup> J. Reed, C. Krueger, D. Shanmuganayagam, T. Crenshaw, J. Reichert, and J. Parrish,  
650 Wisconsin Miniature Swine, Technical report.
- 651 <sup>31</sup> J. G. Lee, S. Park, C. H. Bae, W. S. Jang, S. J. Lee, D. N. Lee, J. K. Myung, C. H.  
652 Kim, Y. W. Jin, S. S. Lee, and S. Shim, Development of a minipig model for lung injury  
653 induced by a single high-dose radiation exposure and evaluation with thoracic computed  
654 tomography, *Journal of Radiation Research* **57**, 201–209 (2016).
- 655 <sup>32</sup> J. P. Williams, S. L. Brown, G. E. Georges, M. Hauer-Jensen, R. P. Hill, A. K. Huser,  
656 D. G. Kirsch, T. J. MacVittie, K. A. Mason, M. M. Medhora, J. E. Moulder, P. Okunieff,  
657 M. F. Otterson, M. E. Robbins, J. B. Smathers, and W. H. McBride, Animal Models  
658 for Medical Countermeasures to Radiation Exposure, *Radiation Research* **173**, 557–578  
659 (2010).
- 660 <sup>33</sup> D. T. Schomberg, A. Tellez, J. J. Meudt, D. A. Brady, K. N. Dillon, F. K. Arowolo,  
661 J. Wicks, S. D. Rousselle, and D. Shanmuganayagam, Miniature Swine for Preclinical  
662 Modeling of Complexities of Human Disease for Translational Scientific Discovery and  
663 Accelerated Development of Therapies and Medical Devices, 2016.
- 664 <sup>34</sup> K. Cao, K. Ding, J. M. Reinhardt, and G. E. Christensen, Improving Intensity-Based
-

- 665 Lung CT Registration Accuracy Utilizing Vascular Information, *International Journal*  
666 *of Biomedical Imaging* **2012**, 1–17 (2012).
- 667 <sup>35</sup> Y. Yin, E. A. Hoffman, and C.-L. Lin, Mass preserving nonrigid registration of CT lung  
668 images using cubic B-spline, *Medical Physics* **36**, 4213–4222 (2009).
- 669 <sup>36</sup> A. E. Scheenstra, M. M. Rossi, J. S. Belderbos, E. M. Damen, J. V. Lebesque, and  
670 J.-J. Sonke, Alpha/Beta Ratio for Normal Lung Tissue as Estimated From Lung Cancer  
671 Patients Treated With Stereotactic Body and Conventionally Fractionated Radiation  
672 Therapy, *International Journal of Radiation Oncology\*Biography\*Physics* **88**, 224–228  
673 (2014).
- 674 <sup>37</sup> K. Du, J. M. Reinhardt, G. E. Christensen, K. Ding, and J. E. Bayouth, Respiratory ef-  
675 fort correction strategies to improve the reproducibility of lung expansion measurements,  
676 *Medical Physics* **40** (2013).
- 677 <sup>38</sup> E. Wallat, M. Flakus, A. Wuschner, W. Shao, G. Christensen, J. Reinhardt,  
678 A. Baschnagel, and J. Bayouth, Modeling the impact of out-of-phase ventilation on  
679 normal lung tissue response to radiation dose, *Medical Physics* (2020).
- 680 <sup>39</sup> A. Ghasemi and S. Zahediasl, Normality Tests for Statistical Analysis: A Guide for  
681 Non-Statisticians, *International Journal of Endocrinology and Metabolism* **10** (2012).
- 682 <sup>40</sup> S.-Y. Chen, Z. Feng, and X. Yi, A general introduction to adjustment for multiple  
683 comparisons, *Journal of Thoracic Disease* **9** (2017).
- 684 <sup>41</sup> Y. Shioyama, S. Y. Jang, H. H. Liu, T. Guerrero, X. Wang, I. W. Gayed, W. D. Erwin,  
685 Z. Liao, J. Y. Chang, M. Jeter, B. P. Yaremko, Y. O. Borghero, J. D. Cox, R. Komaki,  
686 and R. Mohan, Preserving Functional Lung Using Perfusion Imaging and Intensity-  
687 Modulated Radiation Therapy for Advanced-Stage Non-Small Cell Lung Cancer, *Inter-*  
688 *national Journal of Radiation Oncology\*Biography\*Physics* **68** (2007).
- 689 <sup>42</sup> K. P. Farr, D. S. Møller, A. A. Khalil, S. Kramer, A. Morsing, and C. Grau, Loss of  
690 lung function after chemo-radiotherapy for NSCLC measured by perfusion SPECT/CT:  
691 Correlation with radiation dose and clinical morbidity, *Acta Oncologica* **54** (2015).

- 692 <sup>43</sup> T. Guerrero, K. Sanders, J. Noyola-Martinez, E. Castillo, Y. Zhang, R. Tapia, R. Guerra,  
693 Y. Borghero, and R. Komaki, Quantification of regional ventilation from treatment  
694 planning CT, *International Journal of Radiation Oncology\*Biography\*Physics* **62** (2005).
- 695 <sup>44</sup> T. Guerrero, K. Sanders, E. Castillo, Y. Zhang, L. Bidaut, T. Pan, and R. Komaki,  
696 Dynamic ventilation imaging from four-dimensional computed tomography, *Physics in  
697 Medicine and Biology* **51** (2006).
- 698 <sup>45</sup> B. A. Simon, Non-invasive imaging of regional lung function using X-ray computed  
699 tomography, *Journal of Clinical Monitoring and Computing* **16** (2000).

## 700 VII. Author Contribution Statement

701 **Antonia E. Wuschner, M.S.:** Antonia Wuschner performed all dynamic contrast CT  
702 data collection, performed all HU analysis in both the swine and humans, and performed  
703 analysis relating the Jacobians to the HU data. She also assisted in the human clinical data  
704 collection and completed the vast majority of manuscript preparation including writing,  
705 figure preparation, and formatting.

706 **Eric M. Wallat, M.S.:** Eric Wallat assisted in data collection for the human study  
707 as well as some of the jacobian data collection in the human subjects. He also assisted in  
708 editing of the manuscript.

709 **Mattison J. Flakus, M.S.:** Mattison Flakus assisted in data collection for the swine  
710 study as well as some of the jacobian data collection in the human subjects. She also assisted  
711 in editing of the manuscript.

712 **Dhanansayan Shanmuganayagam, PhD:** Dr. Shanmuganayagam was a lead in-  
713 vestigator in the development of the Wisconsin Miniature Swine used in this work. He  
714 provided extensive guidance and consultation regarding this swine breed, their characteris-  
715 tics, and information about their development for study development and inclusion in the  
716 supplementary information. He also assisted in the experimental design of the swine study.

717 **Jennifer Meudt, M.S.:** Jennifer Meudt was the primary animal care and husbandry  
718 coordinator. She assisted in the experimental swine study design specifically by assuring  
719 all procedures were approved by the IACUC committee. Additionally she coordinated all

720 animal care during the study and served as veterinary support during all imaging and treat-  
721 ments. This included monitoring and recording animals vitals during the study, anesthetiz-  
722 ing animals for imaging/treatment, delivering any necessary drugs, and performing animal  
723 husbandry in between sessions.

724 **Gary E. Christensen, PhD:** Dr. Christensen was a research advisor for this work.  
725 He helped guide image analysis techniques and interpretation specifically assisting with the  
726 image registration used in this work. He also developed the image registration algorithms  
727 used in this work and oversaw the development of the Jacobian calculation algorithms used  
728 in this work. He is also a co-investigator on the human clinical trial and provided substantial  
729 edits for the manuscript.

730 **Joseph M. Reinhardt, PhD:** Dr. Reinhardt was a research advisor for this work.  
731 He helped guide image analysis techniques and interpretation specifically in regards to the  
732 dynamic perfusion analysis. His expertise in lung imaging guided experimental design for  
733 the swine as well as post-processing techniques. He is also a co-investigator on the human  
734 clinical trial and provided substantial edits for the manuscript.

735 **Jessica R. Miller, PhD:** Dr. Miller assisted extensively in the swine study experimen-  
736 tal design. Specifically, she helped research and design the timing and imaging parameters for  
737 the dynamic perfusion CT. She also assisted with data collection in the swine and some anal-  
738 ysis and interpretation of the dynamic perfusion CTs. She also provided edits for manuscript  
739 preparation regarding the dynamic perfusion CT analysis.

740 **Micheal J. Lawless, PhD:** Dr. Lawless assisted extensively in the swine study ex-  
741 perimental design alongside Dr. Miller. He also was a crucial component in determining  
742 the timing and imaging parameters for the dynamic perfusion CT and assisted with swine  
743 data collection. He also provided edits for manuscript preparation regarding the dynamic  
744 perfusion CT analysis

745 **Andrew M. Baschnagel, MD:** Dr. Baschnagel was the attending physician for the  
746 human clinical trial subjects and assisted in accrual of subjects, experimental design of the  
747 swine study, and interpretation of the imaging biomarkers in relation to their physiological  
748 responses. He is also a co-investigator on the human clinical trial and provided substantial  
749 edits for manuscript preparation.

750 **John E. Bayoth, PhD:** Dr. Bayouth was the primary research advisor for this work.  
751 He helped guide all image analysis techniques and interpretation. He is also the principle  
752 investigator for the human clinical trial and the swine trial, coordinated each of these, and  
753 assisted with data collection and provided extensive edits to the manuscript during prepara-  
754 tion.

## 755 VIII. Legends

756 Figure 1:(a) Placement of an ROI in a region showing high contrast uptake (b) Plot of the  
757 mean HU value in that ROI over the timespan of the dynamic 4DCT scan for a subject. The  
758 increase in mean HU shows that contrast is flowing in and out of this vessel. (c) Histogram of  
759 HU values over the entire lung mask. A threshold was established based on the second mode  
760 and was placed 2 standard deviations from the mean of the second mode. (d) Example scan  
761 slice showing vasculature segmented using the threshold. (e) Example scan slice showing the  
762 four different contours created for each dose bin to perform analysis.

763 Figure 2: (a) Plot the mean HU value of the lung parenchyma pre-RT (b) Plot of the  
764 mean HU value of the lung parenchyma 3 months post-RT in a typical WMS swine and (c)  
765 Plot of the mean HU value of the vessels showing the two measurements obtained.

766 Figure 3: (a) Percent changes in mean HU post-RT in the swine (b) Percent changes  
767 in mean HU post-RT in the humans. (C) Irradiated dose contours and copied contralateral  
768 contours in the WMS swine (d) Dose distributions and contours for a typical human subject.  
769 Data is shown at 3, 6, and 12 months post RT in the humans and at 3 months for the swine.  
770 Copied contours in unirradiated regions were averaged and plotted as the 0 Gy data point  
771 in (a). Linear fits with  $R^2$  values are shown for all data sets. The percent increases in all  
772 subjects were linear with strong correlations. Additionally, the unirradiated regions of the  
773 swine showed no change post-RT.

774 Figure 4: (a) Heat map showing the percent change in HU in each voxel with the  
775 corresponding dose distribution for a typical WMS swine (b) for an SBRT treated human.  
776 In all cases, large increases in HU are localized to areas receiving radiation.

777 Figure 5: (a) Example contrast flow in the vessels pre- and post-RT for one WMS subject  
778 in the 20 Gy dose bin; by comparison the post-RT peak HU is reduced in magnitude. Here

779 the acquisition times shown are the time of day for the pre-RT data. Since the specific times  
780 are insignificant, the post-RT data is plotted on the same timepoints. The important feature  
781 is the difference in the pre and post-RT peak HU values (b) Example contrast flow outside  
782 of the vessels for the same WMS subject in the 20Gy dose bin; the post-RT ipsilateral region  
783 shows increased HU outside the vessels while the contralateral region does not increase. (c)  
784 Reduction in HU inside the vessels corresponds to an increase outside the vessels. Each  
785 point on the line corresponds to a different dose bin analyzed and is the average of all WMS  
786 subjects.

787 Figure 6: Correlation between the changes seen in the swine at 3 months post-RT and  
788 the humans at different timepoints post-RT. Each point represents a dose bin where analysis  
789 was performed. (a) Correlation between the humans at 3 months and swine at 3 months (b)  
790 Correlation between the humans at 12 months and swine at 3 months. A curve was fit to  
791 all data sets and Pearson stats showed strong correlation with the swine data.

792 Figure 7: (a) Percent of voxels experiencing a 5 % or greater reduction in Jacobian value  
793 post-RT at each dose level in the humans. Results show a linear increase in the percent of  
794 voxels experiencing the reduction in ventilation with increasing dose. Data is shown at 3, 6,  
795 and 12 months post-RT. The number of subjects analyzed at each timepoint are 19, 13, and  
796 11.

797 Figure 8: Correlation by dose bin of the percent of voxels with Jacobian ratio less than  
798 0.95 with the percent increase in HU at (a) 3 months post RT (b) 12 months post-RT. While  
799 there are strong correlation coefficients observed at 3 months, the slopes of the best fit lines  
800 are very low indicating no real relationship between the two metrics. Each point on the plots  
801 represents a dose bin analyzed

# Figures

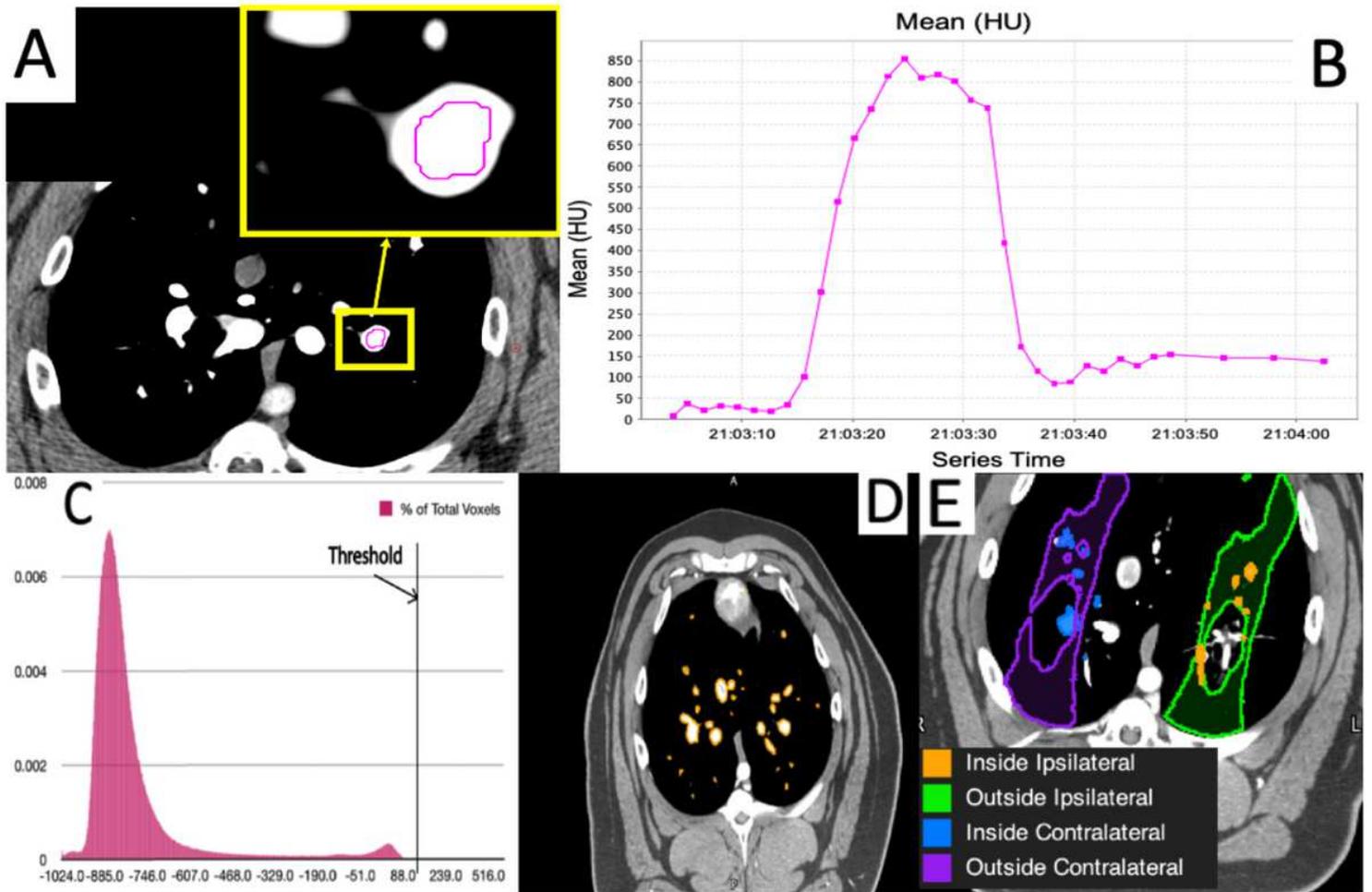


Figure 1

(a) Placement of an ROI in a region showing high contrast uptake (b) Plot of the mean HU value in that ROI over the timespan of the dynamic 4DCT scan for a subject. The increase in mean HU shows that contrast is owing in and out of this vessel. (c) Histogram of HU values over the entire lung mask. A threshold was established based on the second mode and was placed 2 standard deviations from the mean of the second mode. (d) Example scan slice showing vasculature segmented using the threshold. (e) Example scan slice showing the four different contours created for each dose bin to perform analysis.

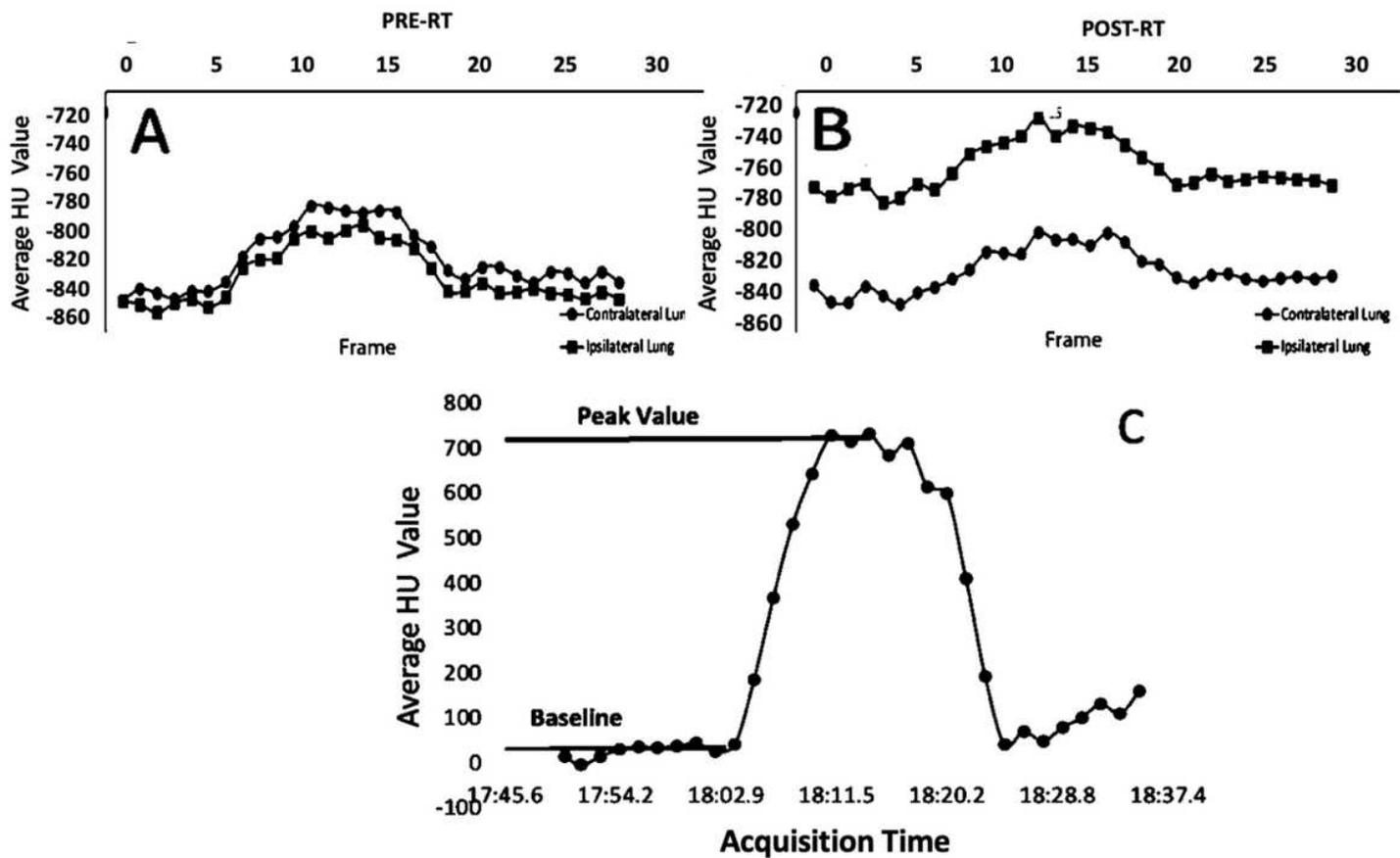
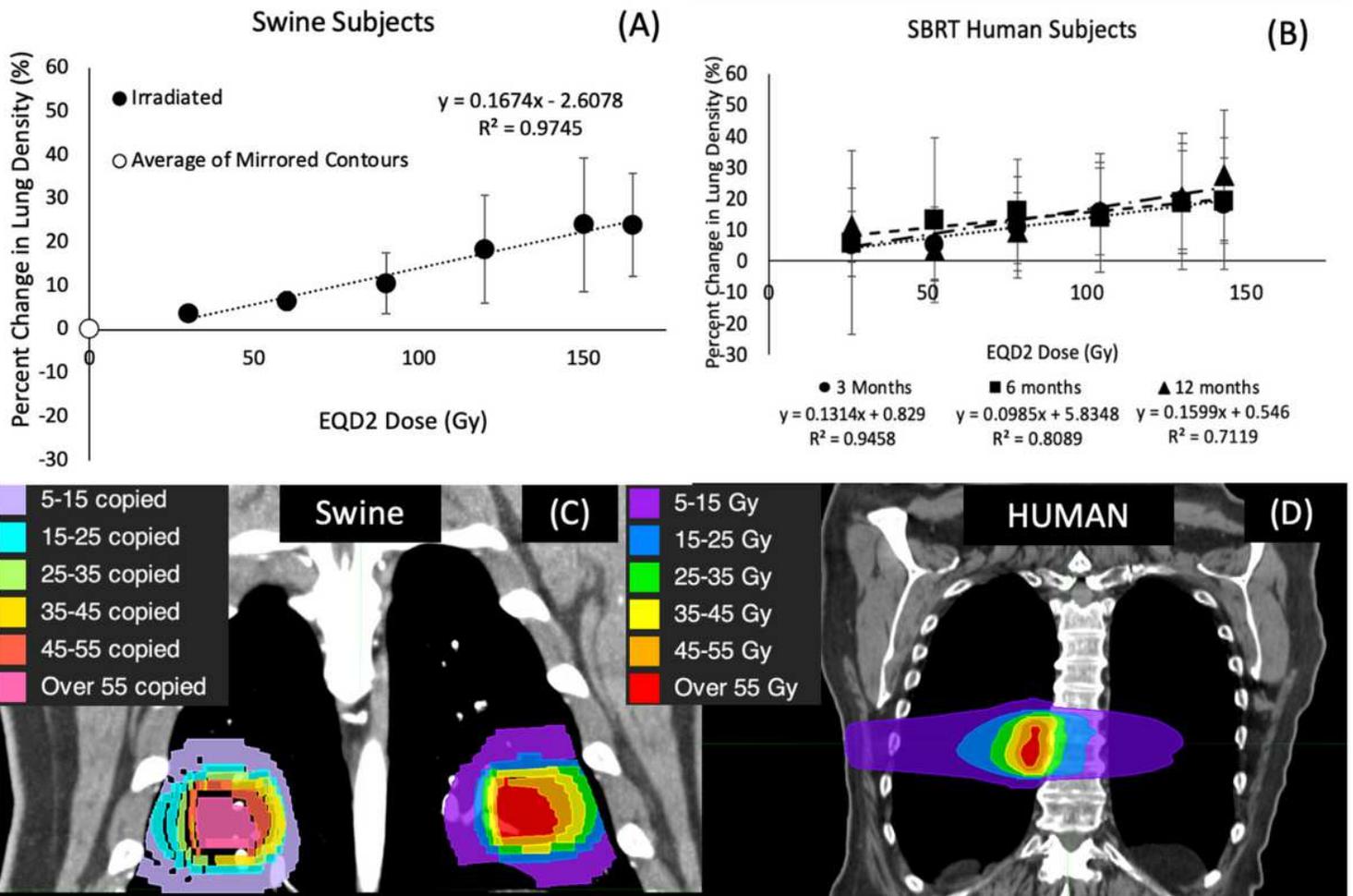


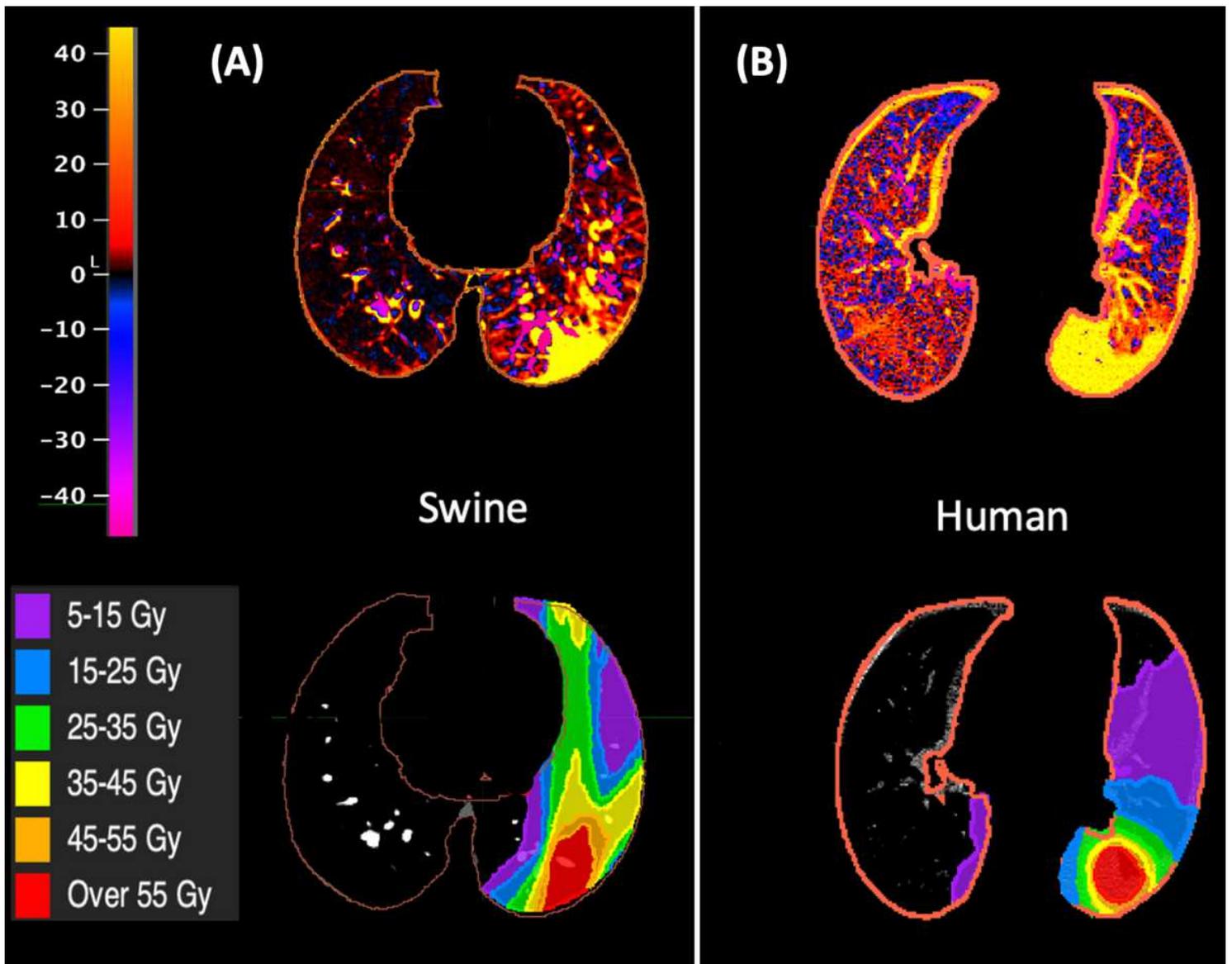
Figure 2

(a) Plot the mean HU value of the lung parenchyma pre-RT (b) Plot of the mean HU value of the lung parenchyma 3 months post-RT in a typical WMS swine and (c) Plot of the mean HU value of the vessels showing the two measurements obtained.



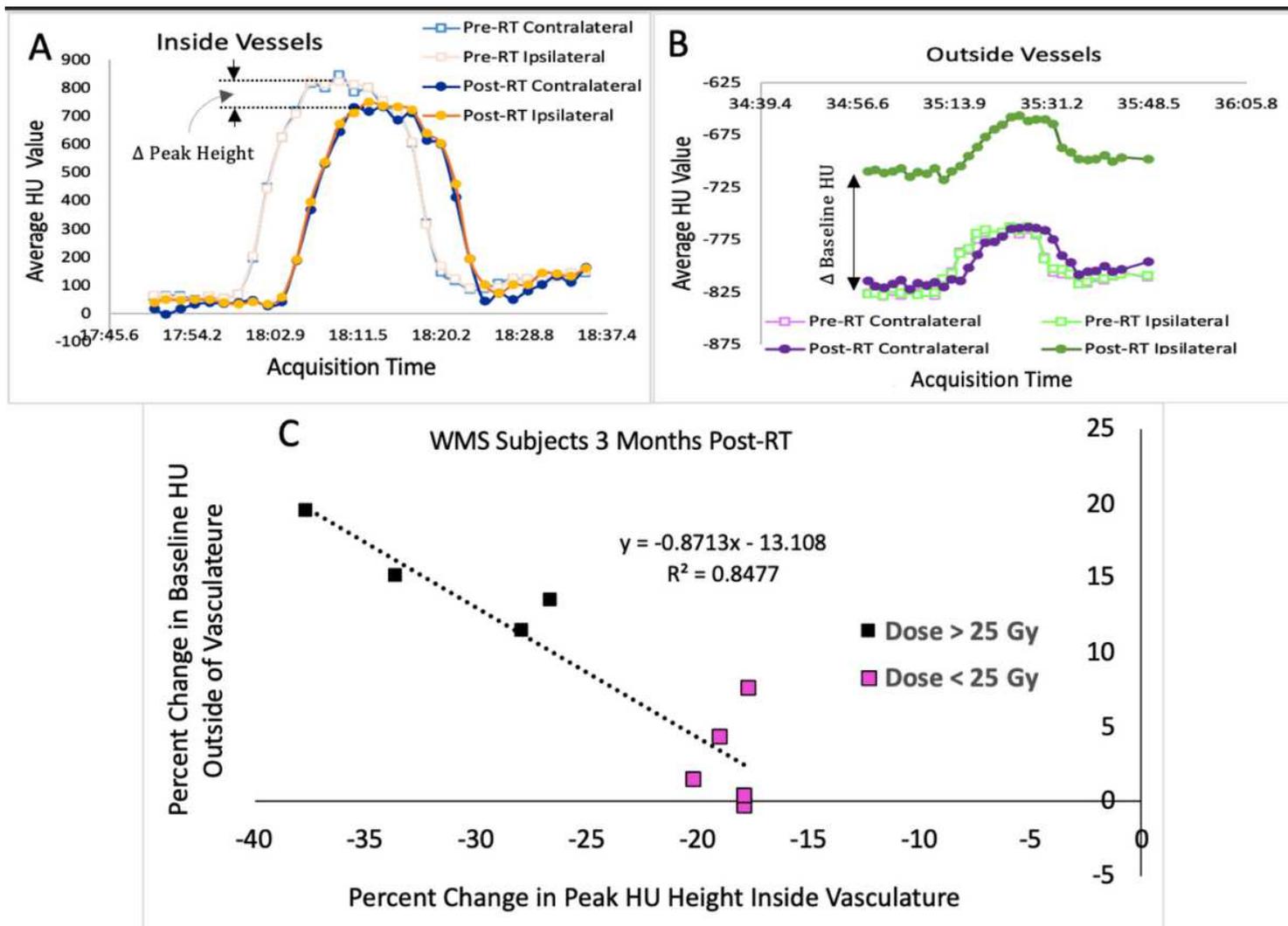
**Figure 3**

(a) Percent changes in mean HU post-RT in the swine (b) Percent changes in mean HU post-RT in the humans. (c) Irradiated dose contours and copied contralateral contours in the WMS swine (d) Dose distributions and contours for a typical human subject. Data is shown at 3, 6, and 12 months post RT in the humans and at 3 months for the swine. Copied contours in unirradiated regions were averaged and plotted as the 0 Gy data point in (a). Linear fits with R2 values are shown for all data sets. The percent increases in all subjects were linear with strong correlations. Additionally, the unirradiated regions of the swine showed no change post-RT



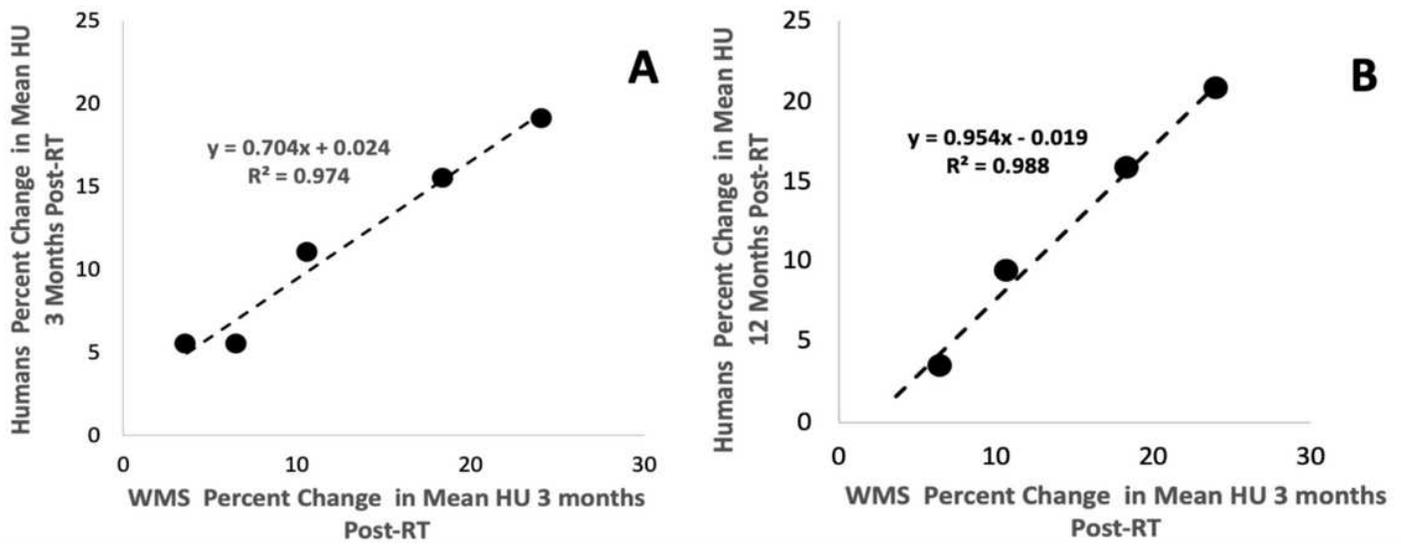
**Figure 4**

(a) Heat map showing the percent change in HU in each voxel with the corresponding dose distribution for a typical WMS swine (b) for an SBRT treated human. In all cases, large increases in HU are localized to areas receiving radiation.



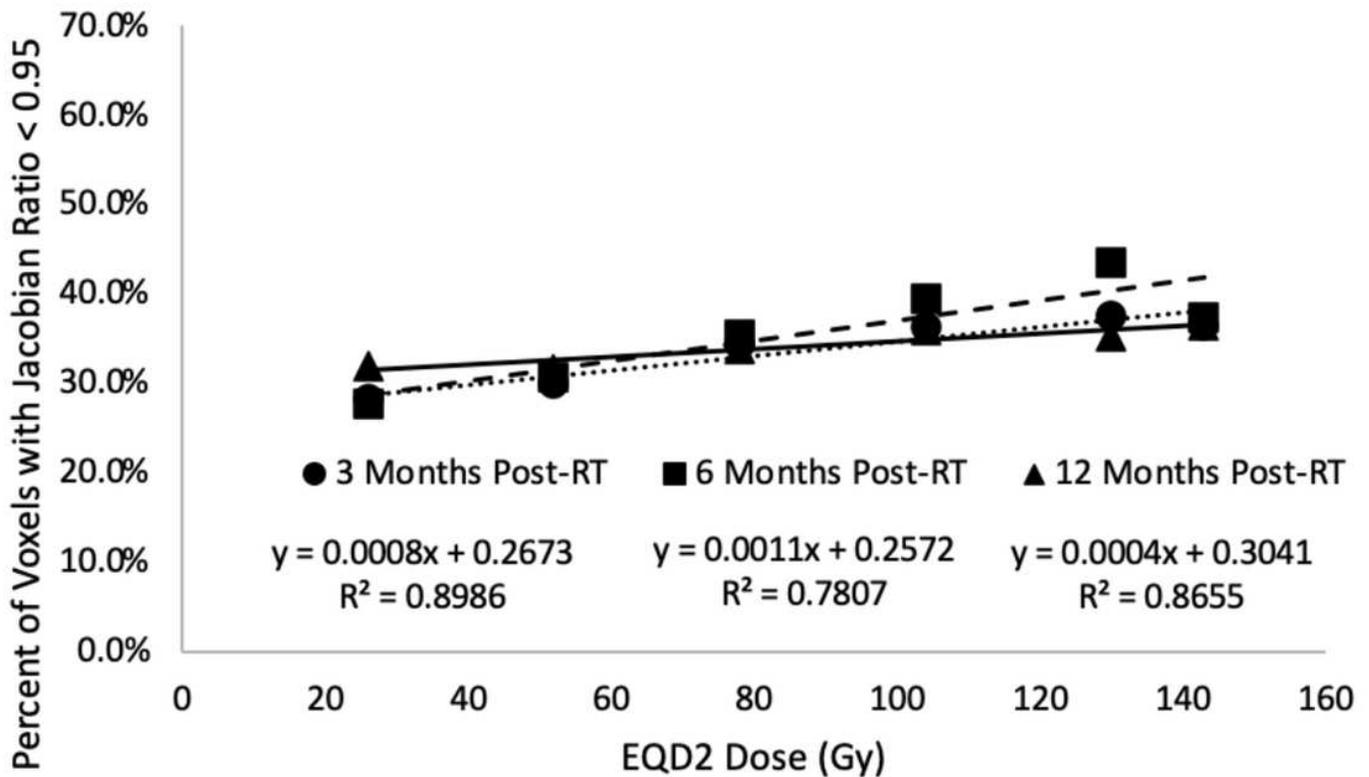
**Figure 5**

(a) Example contrast ow in the vessels pre- and post-RT for oneWMS subject in the 20 Gy dose bin; by comparison the post-RT peak HU is reduced in magnitude. Here the acquisition times shown are the time of day for the pre-RT data. Since the specific times are insignificant, the post-RT data is plotted on the same timepoints. The important feature is the difference in the pre and post-RT peak HU values (b) Example contrast ow outside of the vessels for the same WMS subject in the 20Gy dose bin; the post-RT ipsilateral region shows increased HU outside the vessels while the contralateral region does not increase. (c) Reduction in HU inside the vessels corresponds to an increase outside the vessels. Each point on the line corresponds to a different dose bin analyzed and is the average of all WMS subjects.



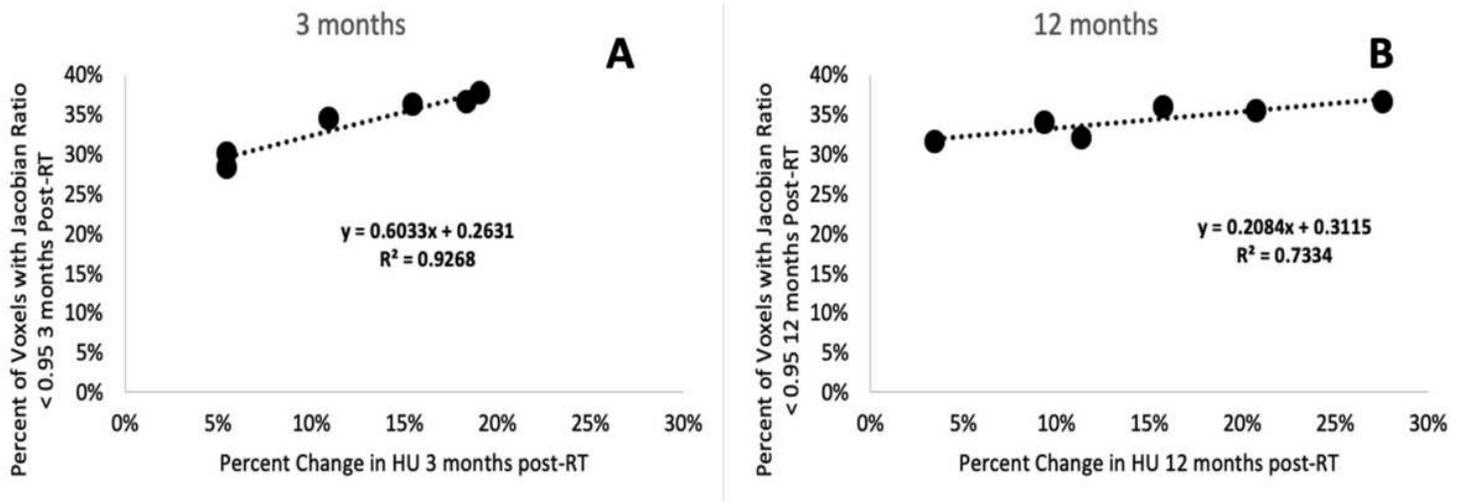
**Figure 6**

Correlation between the changes seen in the swine at 3 months post-RT and the humans at different timepoints post-RT. Each point represents a dose bin where analysis was performed. (a) Correlation between the humans at 3 months and swine at 3 months (b) Correlation between the humans at 12 months and swine at 3 months. A curve was fit to all data sets and Pearson stats showed strong correlation with the swine data.



**Figure 7**

(a) Percent of voxels experiencing a 5 % or greater reduction in Jacobian value post-RT at each dose level in the humans. Results show a linear increase in the percent of voxels experiencing the reduction in ventilation with increasing dose. Data is shown at 3, 6, and 12 months post-RT. The number of subjects analyzed at each timepoint are 19, 13, and 11.



**Figure 8**

Correlation by dose bin of the percent of voxels with Jacobian ratio less than 0.95 with the percent increase in HU at (a) 3 months post RT (b) 12 months post-RT. While there are strong correlation coefficients observed at 3 months, the slopes of the best fit lines are very low indicating no real relationship between the two metrics. Each point on the plots represents a dose bin analyzed.

## Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [DeltaHUSupplementalrevised.pdf](#)