

Isolation, identification and 16S rRNA gene analysis of *Yersinia enterocolitica* strains isolated from the *Gymnocypris Przewalskii*

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Abstract

Background: *Yersinia enterocolitica* is a human-animal-fish-associated infectious diarrhea pathogen that has caused widespread international attention in recent years. Many strains of *Yersinia enterocolitica* were identified from different animal species, but there is no information reported *Yersinia enterocolitica* in *Gymnocypris przewalskii*. The *Gymnocypris przewalskii* is a very important species in the Qinghai Lake. They were listed in the China's second-class protected animal species. Preliminary research on the distribution of *Yersinia enterocolitica* and the drug sensitive fauna test on *Gymnocypris przewalskii* was a urgent solving problem for which maintain the original ecological symbiotic system and restore *Gymnocypris przewalskii* resource. In order to solve these issues, we performed this research.

Results: The results showed that 5 strains *Yersinia enterocolitica* were obtained, positive ratio was 6.67% (5/75). The average drug-resistant of 5 strain *Yersinia enterocolitica* was 54.29% (38/70) to 14 kinds of antibiotic. The result of 16S rRNA gene of *Yersinia enterocolitica* identified showed that one piece of 1419 bp specific braid was obtained. The homologies of nucleotide of 16S rRNA were 91%-95% between 15 strains of *Yersinia enterocolitica* from GenBank by measuring sequence.

Conclusion: *Yersinia enterocolitica* can infect *Gymnocypris przewalskii*. Some strains have drug-resistant effect. The 16S rRNA gene matches 91%-95% with other strains nucleotides download from GenBank and forms a unique branch separated from them.

Background

Yersinia enterocolitica is a human-animal-fish-associated infectious diarrhea pathogen that has caused widespread international attention in recent years. Yersiniosis caused by *Yersinia enterocolitica* is the third most common zoonotic disease followed by *Salmonella* and *Campylobacter* in some European countries[1]. It can often cause the respiratory, cardiovascular systems, bones and connective tissues diseases[2, 3], making the disease with poor prognosis or death due to sepsis[4]. At present, the bacteria have been isolated from various mammals, oviparous animals, agricultural products and animal products in China and abroad[5–10]. However, no reports of *Yersinia enterocolitica* from *Gymnocypris przewalskii* in Qinghai Lake have been seen.

The *Gymnocypris przewalskii* belongs to the *Gymnocypris*, *Schizothoracinae*, *Cyprinidae*, *Cypriniformes* and common called 'Huangyu'. It is the only aquatic economic animal in Qinghai lake and its water system. At the end of the last century, due to the shrinking water area of the Qinghai Lake, the deterioration of the water environment, the frequent occurrence of various diseases and the overfishing of spawning and spawning, the phenomenon of spawning and spawning of broods tock was repeatedly banned. In addition, the hatching rate of fertilized eggs of broods tock in the natural state was less than 5% and their growth was very slow (Gains about 0.15 kg every 10 years), leading to a sharp decline in the population base to about 2000t. With the increasing intensity of lake closure and artificial seedling release in Qinghai Lake in recent years, although the population base has reached 88,000 tons, it is still

less than one third of the 1960s. At present, researches on *Gymnocypris przewalskii* in Qinghai Lake are mostly focused on biology and parasitic diseases, while research on pathogenic microorganisms is almost blank.

Results

Routine isolation and identification of *Yersinia enterocolitica* from naked carp of Qinghai Lake

Colony morphology and culture characteristics

Red bull-eye colonies appeared on CIN-1. Colorless and transparent, non-sticky colonies appeared on modified Y. Gram staining microscopic examination showed short rod-shaped Gram-negative bacilli or cocci, mostly scattered individually, sometimes arranged in short chains or piles, without spores and capsules.

Screening of bacteria

After preliminary screening tests, 20 strains of KIA with yellow and non-gas-producing strains were obtained on the slope and bottom of KIA. Fifteen urea-positive strains were obtained by rescreening. Two rescreening yielded 5 strains of 26 ° C cultured with motility, and 37 ° C cultured non-motility strains, which were classified as Y1301, Y1302, Y1303, Y1304, Y1305.

Biochemical identification

See Table 1 for details.

Table 1 The result of biochemical characteristics of *Yersinia enterocolitica*

Items		Y1301	Y1302	Y1303	Y1304	Y1305
GLU		0	0	0	0	0
LAC		0	0	0	0	0
MAL		0	0	0	0	0
SUC		0	0	0	0	0
Trehalose		0	0	0	0	0
RHA		0	0	0	0	0
Raffinose		0	0	0	0	0
ARA		0	0	0	0	0
XYL		0	0	0	0	0
MAN		0	0	0	0	0
SOR		0	0	0	0	0
DUL		0	0	0	0	0
SAL		0	0	0	0	0
INO		0	0	0	0	0
ODC		0	0	0	0	0
TRP		0	0	0	0	0
CIT		0	0	0	0	0
NIT		0	0	0	0	0
URE		0	0	0	0	0
Semi-solid	26°C	0	0	0	0	0
	37°C	0	0	0	0	0
GEL		0	0	0	0	0
MR		0	0	0	0	0
V-P	26°C	0	0	0	0	0
	37°C	0	0	0	0	0
H ₂ S		0	0	0	0	0
IND		0	0	0	0	0

Note: "+" means positive, "-" means negative

Drug sensitive test

The results are as follows (Table 2)

Table 2 Sensitive Test of 14 kinds of Medicine for 5 strains *Yersinia enterocolitis*

Antibiotics	Y1301	Y1302	Y1303	Y1304	Y1305	Resistance	intermediary	Sensitivity
AZI	S	R	R	S	R	60%	0.00%	40%
GMIO	S	S	S	S	I	0.00%	20%	80%
CH	R	R	R	R	R	100%	0.00%	0.00%
E	R	R	S	I	R	60%	20%	20%
AM	S	R	S	R	I	40%	20%	40%
CAZ	R	R	I	R	S	60%	20%	20%
PIP	R	S	R	S	I	40%	20%	40%
TE	I	I	S	R	I	20%	60%	20%
TM	S	S	S	R	S	20%	0.0%	80%
CMZ	R	R	R	R	S	80%	0.00%	20%
K	R	R	R	S	I	60%	20%	20%
S ₃₀₀	R	R	R	I	R	80%	20%	0.00%
SXT	R	R	R	R	I	80%	20%	0.00%
C	R	S	R	R	S	60%	0.00%	40%

16S rRNA gene clone

PCR amplification of 16S rRNA gene of *Yersinia enterocolitica* from naked carp of Qinghai Lake (Fig. 1).

PCR specificity test of wild isolates and control bacteria

The PCR specificity test of 16 s rRNA gene was performed on Y1301 and Y1302 strains with P1 and P2 primers, respectively. The target fragment of 1419 bp in size consistent with the expected results was obtained (Fig. 2).

Sensitivity test of isolated strains

A single clone of Y1301 was aseptically inoculated into LB Amp + liquid medium, cultured at 26 ° C for 12 hours, and genomic DNA was extracted. The extracted DNA was diluted 10-fold to 10⁻¹⁰ in turn, and P1 and P2 primer pairs were applied respectively. The PCR amplification of 16S rRNA gene was performed, and the obtained PCR product was subjected to electrophoresis, and the results showed that a dilution of 10⁻⁶ was detectable (Fig. 3).

Homologous analysis of sequence of *Y.e* 16S rRNA gene from naked carp of Qinghai Lake

The nucleotide sequence comparison showed that the nucleotide homology of Y1301, Y1303 and Y1304 was 100%; the homology of Y1302 and Y1305 was 100%, and there were no mutation sites. AJ639645, HE803738, HE803739, HE803740, HE803741, HE803742, HE803743, HE803744, HE803745, HE803748,

HE803750, HE803756, HE803758, HE803762, HE803792 are in sites of 28th, 72th, 75th, 81th, 84th, 101th, 112nd, 154th, 299th, 325th, 346th, 347th, 351st, 352nd, 359th, 361st, 362nd, 366th, 367th, 481st, 485th, 491st, 507th, 511st, 515th, 529th, 540th, 702nd, 729th, 739th, 806th, 816th, 867th, 892nd, 893th, 897th, 912nd, 913th, 928th, 929th, 992-997th, 998th -1006th, 118th, 1010th -1012nd, 1016th -1017th 1020th -1025th, 1041st -1042nd, 1046th, 1050th - 1055th, 1057th -1067th, 1070th -1073th, 1076th -1077th, 1078th -1079th, and 1081st -1082nd have changed. Y1301 had base deletions at 915 and 976, and Y1302 had base deletions at 816 and 867, respectively (Fig. 4, Fig. 5).

Conclusion

In this test, 5 isolated strains of *Yersinia enterocolitica* were identified via traditional biochemical, Pathogenicity identification and drug resistance testing. The result is that they were identified as *Yersinia enterocolitica* with severe drug resistance. At the same time, a phylogenetic tree was constructed based on the 16S rRNA gene of *Yersinia enterocolitica*, and the results showed that *Yersinia enterocolitica* isolated from Qinghai Lake naked carp located in the Qinghai-Tibet Plateau was an independently evolved cluster. Compared to other strains isolated from non-Tibet Plateau, they show clear regional differences. By routine identification and identification of *Yersinia enterocolitica* from naked carp of Qinghai Lake and detection of drug-susceptible flora, the interspecies distribution and drug distribution of *Yersinia* from naked carp of Qinghai Lake can be found out. Sensitive fauna lays a theoretical foundation for future research on key control points for preventing the disease. The theoretical and practical significance of protecting local special animal resources, exploring ways and capabilities to adapt to the environment, and laying a research foundation for comprehensively protecting the biodiversity of the Qinghai-Tibet Plateau.

Discussion

The results of the drug susceptibility test showed that the resistance rate of five strains to 14 types of drug susceptibility tests exceeded 50.00%. Y1301 is resistant to 9 drugs: CH, E, CAZ, K, AMP, PIP, CMZ, K, P, S300, SXT, and C. The drug resistance rate is 56.25% (9/16). No. 2 is resistant to 11 drugs: AZI, CH, E, AN, CAI, AMP, CMZ, P, K, P, E, S300, and SXT, with a drug resistance rate of 68.75% (11/16). No. 3 has resistance to 10 drugs, AZI, CH, PIP, AMP, K, P, S300, SXT, and C, with a drug resistance rate of 62.50% (10/16). No. 4 has resistance to 9 drugs: CH, AN, CAZ, TE, TM, CMZ, P, SXT, and C. The drug resistance rate is 56.25% (9/16). No. 5 is resistant to five drugs, AZI, CH, E, AMP, P, and S300, with a resistance rate of 31.25% (5/16). The results of the drug sensitivity test showed that the isolate was highly sensitive to two drugs, gentamicin and tobramycin, but resistant to penicillin. Therefore, these high-sensitivity drugs can be selected for prevention and control in production. This result is different from the drug sensitivity characteristics of *Yersinia* isolated from spot fork tails infected by *Y.e.* The possible reason for the difference in drug sensitivity characteristics is that the strains in different regions and different water environments are exposed to the effects of different drug environmental effects, resulting in differences in drug resistance variations.

Because biochemical characteristics of *Y.e* are more complex, accurate biochemical test results are essential to identify *Y.e*. The isolates tested in this test were sucrose-positive, raffinose-negative, VP (26 °C) positive, and VP (37 °C) negative, all of which were in line with typical *Ye* characteristics. In addition, the biochemical characteristics of other bacteria belonging to the genus *Enterobacteriaceae* were used. The biochemical characteristics of indole-positive and fermented sucrose can be distinguished from *Yersinia pseudotuberculosis*. At the same time, according to the biochemical characteristics of the isolates and the typical culture characteristics on CIN-1 and modified Y medium, it can be clearly distinguished from the genera *Proteus*, *Salmonella* and *Shigella*. Finally, according to the Berger's Bacteria Identification Manual and Cai Miaoying's "Enterobacteriaceae Retrieval Line", the isolate was determined to be *Y.E*. Because this bacterium is a psychrophilic pathogen, it brings great difficulty to the isolation and identification. According to the National Standard for "*Yersinia enterocolitica* Test" (GB4789.8-2008) issued by the Ministry of Health of the People's Republic of China, by using the alkali treatment method in combination with CIN-1 to select media and improve Y, a better Separate selection effects was succeeded. The detection medium for naked carp's solid organ *Y.e* shortened the inspection time by 3d, saved about 40% of raw materials, and improved the detection rate. On the one hand, it demonstrates the superiority of this method, and on the other hand, it also suggests the carrier rate in local naked carps. Five *Y.e* were isolated from the organs and gills of 75 naked carps collected, and the positive rate was 6.67% (5/75).

Methods

Samples collection

75 pancreas of Qinghai Lake naked carp that had died one after another in a fishery of a naked carp rescue center in Qinghai Province were aseptically collected and numbered, and placed in a refrigerator at 4 °C for use.

Control strains

Escherichia coli (ATCC25922) and *Salmonella typhimurium* (ATCC14028) were donated by the Department of Preventive Veterinary Medicine, China Agricultural University.

Growth medium and molecular reagents

The Improved Phosphate Buffer Solution (PBS), modified Y and CIN-1 plates, and modified Kirschner's disaccharide iron bevel (KIA) were made in our laboratory. Glucose and other 24 biochemical media were purchased from Beijing Luqiao Technology Co., Ltd. (batch number: 20180611). Bacterial DNA extraction kit, gel recovery kit, Taq DNA polymerase and pMD18-T vector were purchased from TaKaRa.

Primers

The primers sequences are P1 5'CGCGGATCCATTGAACGCTGGCGGCAG3' and P2 5'GGGGTACCCCTACGGTTACCTTGTTACGACTTC3'. They were synthesized by Shanghai Biotech

Drug sensitive paper

14 kinds of drug sensitive paper such as chloramphenicol were purchased from Hangzhou Microbial Reagent Co., Ltd. (Lot No.: 20180290).

Isolation and identification of *Yersinia enterocolitica* from naked carp of Qinghai Lake.

Sample enrichment

Cut a few samples aseptically and inoculate them in PSB, and incubate them at 26 °C for 48 ± 2 h.

Sample alkali treatment

Take 1 mL of the bacterial growth solution and add it to 9 mL of 0.4% KOH solution (containing 0.5% NaCl, prepared before use), which is warmed to 26 °C, and mix for 30 s.

Isolation and purification of bacteria

Alkali-treated bacteria-enriching solution was streaked on CIN-1 and modified Y plates, and cultured at 26 °C for 48 ± 2 h. After that, the suspect colony smears were picked, and Gram staining microscopy was performed, and several purification cultures were performed.

Screening of bacteria

Five suspicious colonies on CIN-1 and modified Y plate were picked in turn and tested on KIA oblique urea and semi-solid medium, and the results were observed.

Smear, Gram stain and microscopy

Bacteria that were positive at 26 °C and negative at 37 °C in the secondary rescreening test were picked for smear and Gram staining microscopy.

Biochemical identification

The suspected strains screened by the primary and secondary screening tests were aseptically inoculated into 27 biochemical reaction tubes such as glucose and cultured at 26 °C for 48 ± 2 h, and the results were observed.

Drug sensitive test

The purified strains were individually inoculated into a modified Y slant, cultured at 26 °C for 24 hours, and then washed under sterile conditions with sterile physiological saline, and were turbid with a Macquarie tube. A bacterial suspension at a concentration of 9×10^8 live bacteria / mL was selected, and the bacterial solution was dipped in a sterile cotton swab to uniformly coat the MH (A) plate 3 times. Then, 16 kinds of drug-sensitive papers were respectively put on in order, 3 pieces were put on each plate, and the results were determined by incubating at 37 °C for 18–24 h.

Cloning of 16S rRNA gene

Template DNA Extraction

The operation steps are performed according to the instruction manual of MiniBEST Bacterial Genomic DNA Extraction Kit.

PCR reaction system and conditions

PCR reaction system (50 μ L): Premix Taq 25 μ L, P1 and P2 primers 1 μ L each, DNA template 1 μ L, ddH₂O 22 μ L; PCR reaction conditions: 94 °C pre-denaturation for 5 min, 94 °C denaturation for 30 s, 55 °C 30 s, 72 °C for 30 s, a total of 35 cycles, 72 °C extended 10 minutes.

16S rRNA Gene Cloning and Sequence Analysis

The PCR amplified product recovered from the gel was cloned, PCR identified and sequenced 3 times. The 16S rRNA sequence of *Yersinia enterocolitica* from naked carp of Qinghai Lake was performed on GenBank and 15 representative strains of China and abroad Homology analysis.

Abbreviations

PBS: Phosphate Buffer Solution; GLU: glucose; LAC: lactose; MAL: maltose; SUC: sucrose; RHA: rhamnose; ARA: arabinose; XYL: xylose; MAN: mannitol; SOR: sorbitol; DUL: dulcitol; SAL: saligenin; INO: inositol; ODC: Ornithine decarboxylase culture; TRP: Tryptophan; CIT: citrate; NIT: nitrate; URE: urease; MR: methyl red test ; IND: indol; AZI: azithromycin; GMIO: gentamicin; CH: cephalosporin; E: erythrocin; AM: amikacin; CAZ: ceftazidime; PIP: piperacillin; TE: tetracycline; TM: tobramycin; CMZ: cefuroxime; K: kanamycin; S300: streptomycin; SXT: selectrin; C: chloromycetin

Declarations

Ethics approval and consent to participate

No specific permits were required for this study.

Consent for publication

Not applicable.

Availability of data and materials

All data generated or analyzed during this study and supporting the conclusions of this article are included within the article.

Competing interests

The authors declare that they have no competing interests.

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Authors' contributions

ZH, CQ and ZJ conceived and designed the experimental concept. FY and KF collected the samples and extracted the DNA. TL and GX conducted the lab experiments. ZH and CQ wrote the paper. All authors reviewed the manuscript approved the final manuscript.

Acknowledgments

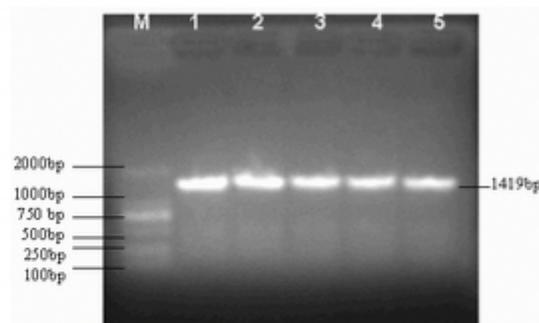
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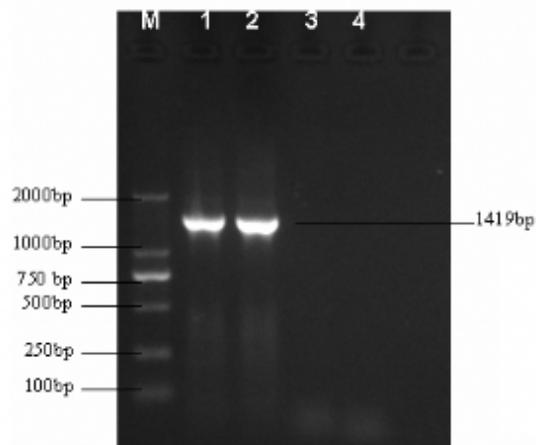
Figures



M: DL 2000 Marker; 1: Y1301; 2: Y1302; 3: Y1303; 4: Y1304; 5: Y1305

Figure 1

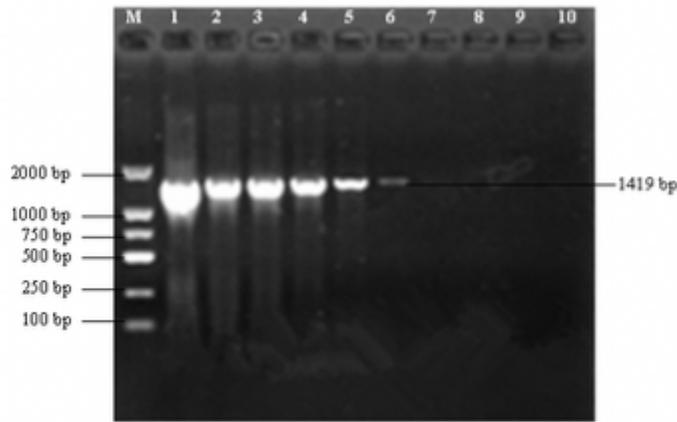
PCR Amplification of gene of 16S rRNA of Y1301, Y1302 by use of P1, P2 Primer



M: DL 2000 Marker; 1: Y1301; 2: Y1302; 3: Escherichia coli (ATCC25922); 4: Salmonella typhimurium (ATCC14028)

Figure 2

Specificity of single PCR of Y1301, Y1302



M: DL 2000 Marker; 1: 10^{-1} ; 2: 10^{-2} ; 3: 10^{-3} ; 4: 10^{-4} ; 5: 10^{-5} ; 6: 10^{-6} ; 7: 10^{-7} ; 8: 10^{-8} ; 9: 10^{-9} ; 10: 10^{-10}

Figure 3

Sensitivity of single PCR of Y1301

		Percent Identity																					
		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20		
Divergence	1	■	98.8	99.6	99.3	99.5	99.3	99.6	99.1	99.1	99.3	99.6	97.9	97.9	98.2	97.9	94.1	93.9	94.1	93.9	93.9	1	HF558392.seq
	2	1.2	■	98.3	98.1	98.3	98.0	98.3	98.0	98.1	98.1	98.3	97.1	97.1	97.5	97.1	94.7	94.8	94.7	94.8	94.8	2	AJ639645.seq
	3	0.4	1.7	■	99.6	99.9	99.6	100.0	99.5	99.5	99.6	100.0	98.3	98.3	98.0	98.3	93.5	93.4	93.5	93.4	93.4	3	HE803738.seq
	4	0.7	2.0	0.4	■	99.6	99.6	99.6	99.5	99.9	100.0	99.6	98.7	98.7	98.4	98.7	93.7	93.5	93.7	93.5	93.5	4	HE803739.seq
	5	0.5	1.7	0.1	0.4	■	99.6	99.9	99.4	99.4	99.6	99.9	98.2	98.2	98.0	98.2	93.5	93.4	93.5	93.4	93.4	5	HE803740.seq
	6	0.4	1.6	0.0	0.0	0.1	■	99.6	99.9	99.5	99.6	99.6	98.3	98.3	98.0	98.3	93.5	93.4	93.5	93.4	93.4	6	HE803741.seq
	7	0.4	1.7	0.0	0.4	0.1	0.0	■	99.5	99.5	99.6	100.0	98.3	98.3	98.0	98.3	93.5	93.4	93.5	93.4	93.4	7	HE803742.seq
	8	0.4	1.6	0.0	0.0	0.1	0.0	0.0	■	99.6	99.5	99.5	98.2	98.2	97.9	98.2	93.5	93.4	93.5	93.4	93.4	8	HE803743.seq
	9	0.8	2.0	0.4	0.1	0.5	0.1	0.4	0.0	■	99.9	99.5	98.5	98.5	98.2	98.5	93.8	93.6	93.8	93.6	93.6	9	HE803744.seq
	10	0.7	2.0	0.4	0.0	0.4	0.0	0.4	0.0	0.1	■	99.6	98.7	98.7	98.4	98.7	93.7	93.5	93.7	93.5	93.5	10	HE803745.seq
	11	0.4	1.7	0.0	0.4	0.1	0.0	0.0	0.0	0.4	0.4	■	98.3	98.3	98.0	98.3	93.5	93.4	93.5	93.4	93.4	11	HE803748.seq
	12	2.1	2.9	1.7	1.4	1.8	1.4	1.7	1.4	1.4	1.4	1.7	■	99.9	99.6	99.9	93.4	93.1	93.4	93.1	93.1	12	HE803750.seq
	13	2.0	2.8	1.6	1.3	1.7	1.3	1.6	1.3	1.4	1.3	1.6	0.0	■	99.7	100.0	93.4	93.1	93.4	93.1	93.1	13	HE803756.seq
	14	1.7	2.4	1.9	1.6	2.0	1.6	1.9	1.6	1.6	1.6	1.9	0.3	0.3	■	99.7	93.2	93.0	93.2	93.0	93.0	14	HE803758.seq
	15	2.0	2.8	1.6	1.3	1.7	1.3	1.6	1.3	1.4	1.3	1.6	0.0	0.0	0.3	■	93.4	93.1	93.4	93.1	93.1	15	HE803762.seq
	16	6.2	5.5	6.8	6.6	6.8	6.4	6.8	6.2	6.4	6.6	6.8	7.0	6.9	7.1	6.9	■	98.3	100.0	98.3	98.3	16	Y1305.seq
	17	6.3	5.4	6.9	6.8	6.9	6.6	6.9	6.4	6.6	6.8	6.9	7.2	7.1	7.3	7.1	1.8	■	98.3	100.0	100.0	17	Y1301.seq
	18	6.2	5.5	6.8	6.6	6.8	6.4	6.8	6.2	6.4	6.6	6.8	7.0	6.9	7.1	6.9	0.0	1.8	■	98.3	98.3	18	Y1302.seq
	19	6.3	5.4	6.9	6.8	6.9	6.6	6.9	6.4	6.6	6.8	6.9	7.2	7.1	7.3	7.1	1.8	0.0	1.8	■	100.0	19	Y1303.seq
	20	6.3	5.4	6.9	6.8	6.9	6.6	6.9	6.4	6.6	6.8	6.9	7.2	7.1	7.3	7.1	1.8	0.0	1.8	0.0	■	20	Y1304.seq
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20			

Figure 4

Analysis of genetic evolution of Y.e 16S rRNA gene from naked carp in Qinghai Lake

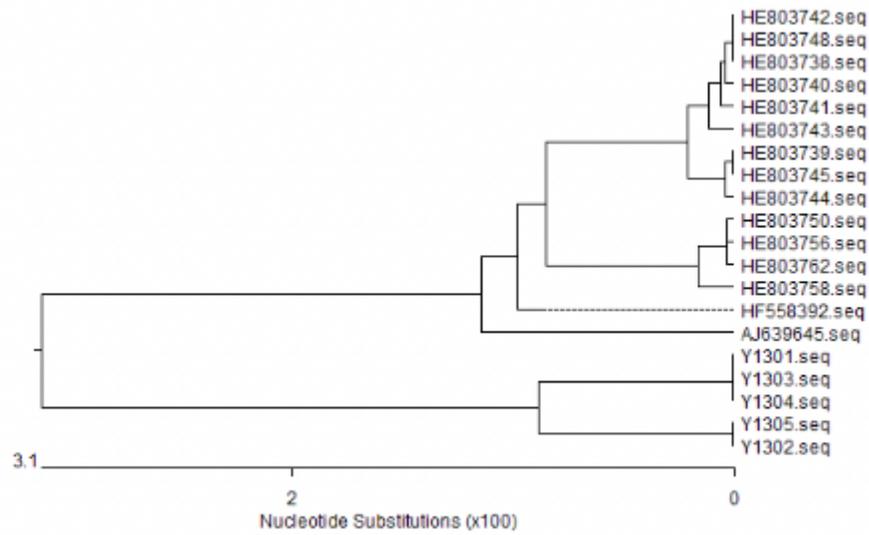


Figure 5

Phylogenetic Tree based on nucleotide sequence of 16S rRNA gene