

# NF- $\kappa$ B and pSTAT3 synergistically drive G6PD overexpression and facilitate sensitivity to G6PD inhibition in ccRCC

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## Primary research

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# Abstract

**Background** Glucose 6-phosphate dehydrogenase (G6PD) serves key roles in cancer cell metabolic reprogramming, and has been reported to be involved in certain carcinogenesis. Previous results from our laboratory demonstrated that overexpressed G6PD was a potential prognostic biomarker in clear cell renal cell carcinoma (ccRCC), the most common subtype of kidney cancer. G6PD could stimulate ccRCC growth and invasion through facilitating reactive oxygen species (ROS)-phosphorylated signal transducer and activator of transcription 3 (pSTAT3) activation and ROS-MAPK-MMP2 axis pathway, respectively. However, the reasons for ectopic G6PD overexpression and the proliferation repressive effect of G6PD inhibition in ccRCC are still unclear.

**Methods** The impact of ROS accumulation on NF- $\kappa$ B signaling pathway and G6PD expression was determined by real-time RT-PCR and Western blot in ccRCC cells following treatment with ROS stimulator or scavenger. The regulatory function of NF- $\kappa$ B signaling pathway in G6PD transcription was analyzed by real-time RT-PCR, Western blot, luciferase and ChIP assay in ccRCC cells following treatment with NF- $\kappa$ B signaling activator/inhibitor or lentivirus infection. ChIP and Co-IP assay was performed to demonstrate protein-DNA and protein-protein interaction of NF- $\kappa$ B and pSTAT3, respectively. MTS assay, human tissue detection and xenograft model were conducted to characterize the association between NF- $\kappa$ B, pSTAT3, G6PD expression level and proliferation functions.

**Results** ROS-stimulated NF- $\kappa$ B and pSTAT3 signaling over-activation could activate each other, and exhibit cross-talks in G6PD aberrant transcriptional regulation. The underlying mechanism was that NF- $\kappa$ B signaling pathway facilitated G6PD transcription via direct DNA-protein interaction with p65 instead of p50. p65 and pSTAT3 formed a p65/pSTAT3 complex, occupied the pSTAT3-binding site on G6PD promoter, and contributed to ccRCC proliferation following facilitated G6PD overexpression. G6PD, pSTAT3, and p65 were highly expressed and positively correlated with each other in ccRCC tissues, confirming that NF- $\kappa$ B and pSTAT3 synergistically promote G6PD overexpression. Moreover, G6PD inhibitor exhibited tumor-suppressor activities in ccRCC and attenuated the growth of ccRCC cells both in vitro and in vivo .

**Conclusion** ROS-stimulated aberrations of NF- $\kappa$ B and pSTAT3 signaling pathway synergistically drive G6PD transcription through forming a p65/pSTAT3 complex and occupy the pSTAT3-binding site on G6PD promoter. Moreover, G6PD activity inhibition may be a promising therapeutic strategy for ccRCC treatment.

## Background

Clear cell renal cell carcinoma (ccRCC) is the most prevalent subtype of kidney malignant tumor, which constitutes about 80–90% of renal cell carcinoma (RCC). Despite the diagnosis and treatment of ccRCC have improved obviously in recent years [1, 2], most ccRCC patients still confront difficulties in early diagnosis, drug resistance, high metastasis rate (~ 34%), and short median survival time (~ 13 months)

[1, 3]. Meanwhile, new cases and the number of deaths due to ccRCC are approximately 400,000 and 139,000, respectively, globally per year [4]. These situations emphasize the importance of studies on ccRCC tumorigenesis. However, the fundamental theory of ccRCC is still unclear, especially the critical molecular mechanisms underlying ccRCC initiation and development [5, 6]. Therefore, identifying novel molecular mechanisms involved in ccRCC tumorigenesis and providing evidence for effective treatment are the pivotal points for ccRCC investigation [6].

In recent years, increasing studies had focused on the metabolism disorder of human cancers [7–9], and ccRCC was generally regarded as a metabolic disease in this field [10–13]. It was reported that ccRCC had increased antioxidant capacity and carbon metabolic reprogramming [14]; tumor progression and metastasis are closely related to enhanced glutathione, cysteine/methionine and tryptophan metabolic pathways [14, 15]. These reports highlighted the importance of the ccRCC metabolic disorder in tumorigenesis. However, the important drivers of the aforementioned metabolic reprogramming, the molecular regulatory mechanisms that joint the malignant phenotype and metabolism disorder, and the maintenance of the overall metabolic homeostasis of tumor cells remain to be clarified [14, 15].

Previous studies conducted in our laboratory showed that glucose 6-phosphate dehydrogenase (G6PD), which is the first and rate-limiting enzyme of the pentose phosphate pathway (PPP) and plays critical roles in the generation of nucleotide precursors, lipid synthesis, redox balance, and maintenance of the whole cell metabolic homeostasis [16, 17], is highly expressed in ccRCC cells and predicts the poor overall survival of patients of ccRCC, indicating that G6PD exerted crucial functions in ccRCC metabolic reprogramming and tumorigenesis [18]. A further investigation described that the overexpressed G6PD positively regulated reactive oxygen species (ROS) generation by facilitating NADPH oxidase 4 activity, increased phosphorylated signal transducer and activator of transcription 3 (pSTAT3) activation and enhanced RCC development [18]. Moreover, G6PD could stimulate ccRCC invasion through facilitating ROS-MAPK-MMP2 axis pathway [19]. The aforementioned results suggested that how G6PD regulated the downstream target, influenced ROS production, and promoted RCC growth and invasion had been revealed, but why G6PD was aberrantly overexpressed during ccRCC tumorigenesis was still not completely known.

It has been reported that ROS dysregulation plays crucial impact on abnormal intracellular signal transduction [20, 21]. ROS could stimulate continuous activation of pSTAT3, NF- $\kappa$ B and MAPK signals [18, 19, 22–24], and promote the occurrence and development of various types of cancer, including RCC, skin cancer and lung cancer [18, 25, 26]. NF- $\kappa$ B signaling over-activation has been described to contribute to RCC cell migration and invasion [27], and the high expression level of pSTAT3 S727 is expected to be an independent prognostic molecule for ccRCC patients [28, 29]. Moreover, the interactions between NF- $\kappa$ B and STAT3 signaling pathways were discovered in several physiological and pathological processes, such as B-cell activation [30], cancer cell starvation [31, 32], osteoclastogenesis [33], and mitochondrial fusion [34, 35]. These two pathways were rapidly activated in response to various stimuli, including cytokines, ROS, and other stresses [36]. Additionally, it was reported that a series of tumorigenesis-related genes could be regulated by pSTAT3 and NF- $\kappa$ B, either synergistically or individually [32], and STAT3/NF-

$\kappa$ B signaling targeting could sensitize triple-negative breast cancer cells to cisplatin [37]. The interactions between ROS, pSTAT3, and G6PD in ccRCC have been reported previously by our laboratory [18]. We found that G6PD could stimulate ccRCC growth through facilitating ROS production and pSTAT3 signaling activities, and pSTAT3 showed a positive feedback regulation of G6PD transcription [18]. These evidences promote us to hypothesize that there may be a correlation between not only ROS and pSTAT3, but also NF- $\kappa$ B signaling pathway and G6PD overexpression in ccRCC. However, the cooperation between these factors, and the underlying mechanism of how they exert the role in ccRCC tumorigenesis have not yet been unraveled.

The present study explored the reasons for the high expression of G6PD in ccRCC. To do so, a reciprocal effect between ROS, NF- $\kappa$ B/STAT3 signaling pathway and G6PD aberrant overexpression in ccRCC was consolidated. Both *in vitro* and *in vivo* investigations were conducted to unravel the molecular mechanism underlying ROS-mediated NF- $\kappa$ B/STAT3 signaling pathway over-activation and ectopic G6PD transcription. Moreover, to evaluate the potential role of G6PD as a therapeutic target for ccRCC treatment, the effects of G6PD inhibition on ccRCC proliferation were assessed both *in vitro* and *in vivo*.

## Methods

### Cell culture, cell treatment, and cell proliferation assays

ACHN (ATCC, CRL-1611™), 786-O (ATCC, CRL-1932™) and Caki-1 (ATCC, HTB46™) cell lines were obtained from Chinese Academy of Sciences, Kunming Institute of Zoology (Kunming, China) and cultured in RPMI-1640 (11875-085, Gibco, Grand Island, NY, USA) containing 10% fetal bovine serum (FBS, 16140071, Thermo Fisher Scientific, Waltham, MA, USA) in a 37°C humidified incubator (5% CO<sub>2</sub>).

H<sub>2</sub>O<sub>2</sub> was purchased from Santa Cruz (sc-203336, CA, USA) and saved at 4 °C. N-acetyl-L-cysteine (NAC, A7250, Sigma, Louis, MO, USA), 6-aminonicotinamide (6-AN, A68203-1G, Sigma), STATTIC (sc-202818, Santa Cruz), and BAY11-7082 (S2913, Selleck, Shanghai, China) were dissolved in 100% DMSO to prepare a 600 mM, 250 mM, 40 mM, and 10 mM stock, respectively, and stored at -20 °C. Recombinant human IL-6 (206IL, R&D Systems, MN, USA) and TNF $\alpha$  (300-01A, Peprotech, Rocky Hill, USA) were dissolved in 0.1% BSA to prepare a 10  $\mu$ g/mL and 50  $\mu$ g/mL stock, respectively, and stored at -20 °C. p65-knocked down RNAi lentivirus (target sequence: CGGATTGAGGAGAAACGTAAA) and negative control were purchased from Genechem (Shanghai, China). An MTS cell proliferation assay kit purchased from Promega (CTB169, Beijing, China) was used for cell viability analysis according to the manufacturer's protocol.

### Real-time RT-PCR

Total RNA was isolated and real-time RT-PCR amplifications were performed according to previous reports [18, 38] using the following primers: G6PD: F: 5'-TGAGCCAGATAGGCTGGAA-3', R: 5'-TAACGCAGGCGATGTTGTC-3'; U6: F: 5'-CTCGCTTCGGCAGCACA-3', R: 5'-AACGCTTCACGAATTTGCGT-3'; CyclinD1: F: 5'-GCGTACCCTGACACCCCTCTC-3', R: 5'-CTCCTCTTCGCCTGATCC-3'; CDK4: F: 5'-

ACAGTTCGTGAGGTGGCTTTAC-3', R: 5'-GTCCTTAGGTCCTGGTCTACATG-3'; STAT3: F: 5'-AGAAGGAGGCGTCACTTTCA-3', R: 5'-TTTCCGAATGCCTCCTCCTT-3'.

## Western blot analysis

Cell lysates were prepared and Western blot analysis was performed as previously described [18, 38]. Following antibodies were used: anti-G6PD antibody (ab133525, Abcam, Cambridge, U.K.); anti-STAT3 antibody (4904, Cell Signaling Technology), anti-phospho-STAT3 antibody (Ser 727) (ab30647), anti-CyclinD1 antibody (ab16663, Abcam), Anti-CDK4 antibody (ab108357), Anti-p105/50 antibody (3035, Cell Signaling Technology), anti-p65 antibody (ab32536), anti- $\beta$ -actin antibody (4967, Cell Signaling Technology), and anti-GAPDH antibody (2118, Cell Signaling Technology).

## Luciferase report analysis

293T cells were seeded in a 24-well plate, and constructs of pGL3-BASIC-luc-G6PD vector (wild type or NF- $\kappa$ B-binding site deletion) were transfected using Lipofectamine 2000 Transfection Reagent (11668019, Invitrogen) when the cells reached 60% confluence. After TNF $\alpha$  or BAY11-7082 stimulation for 24 h, the luciferase activity was measured using a luciferase report assay kit (T002, Vigorous Biotechnology, Beijing, China). Data were evaluated following manufacturer's protocols.

## Chromatin Immunoprecipitation (ChIP) assays

ChIP experiments were performed as described earlier [18, 38] using the following primers covering a 101-bp potential region of the NF- $\kappa$ B-binding site on the G6PD promoter: F: 5'-acgagcaaacaggcatatga-3', R: 5'-atagagcgcagctcaagtttg-3', or a primer covering a 103-bp potential region of the pSTAT3-binding site on the G6PD promoter: F: 5'-AACGTCCGGGGGAAGTTTC-3', R: 5'-TTCTCACGTCTGACGGACTCT-3'. The following antibodies were used: anti-p105/50 antibody (3035, Cell Signaling Technology), anti-p65 antibody (ab32536), anti-STAT3 antibody (4904, Cell Signaling Technology), and anti-phospho-STAT3 antibody (Ser 727) (ab30647).

## Co-IP assays

The cell lysates were prepared using RIPA lysis buffer (P0013D, Beyotime, Shanghai, China) containing protease inhibitor cocktail (469313200, Roche Basle, Switzerland). Immunoprecipitation and Western blot analysis were carried out as described earlier using anti-phospho-STAT3 antibody (Ser 727) (ab30647, Abcam), anti-p65 antibody (ab32536, Abcam), or the control anti-immunoglobulin G antibody (X0936, DAKO A/S) [38].

## Immunohistochemistry (IHC)

All pathological specimens were collected from the department of pathology of the First Affiliated Hospital of Kunming Medical University, which was approved by the Research Ethics Committee of Kunming Medical University. Anti-G6PD antibody (ab133525, Abcam), anti-phospho-STAT3 antibody (Ser 727) (ab30647, Abcam), and anti-p65 antibody (ab32536, Abcam) were used for immunohistochemical analysis performed using the 2-step plus poly-HRP anti-mouse/rabbit IgG detection system (PV-9000,

ZSGB-BIO, Beijing, China). The experimental procedures were performed following the manufacturer's protocol and previous reports [18].

## **G6PD activity detection**

G6PD activity was analyzed using the G6PD assay kit (GMS70013.1, Genmed, Shanghai, China) following the manufacturer's protocol and previous report [18].

## **Animal experiments**

Six-week-old female BALB/c nude mice were injected subcutaneously with  $1 \times 10^7$  ccRCC cells suspended in 200  $\mu$ L of phosphate-buffered saline in the right outer flank. Tumor volumes were calculated as follows: length  $\times$  width<sup>2</sup>  $\times$  0.5. When the average of tumor volume reached 200 mm<sup>3</sup>, tumor-bearing nude mice were randomly divided into two groups ( $n = 5$  per group) and treated intratumor with vehicle or 6-AN at a dose of 23 mg/kg every day for 2 weeks. At the indicated time point, the mice were euthanized by cervical dislocation, and the tumors were harvested. All animal experiments were approved by the Institutional Animal Care and Use Committee of Kunming Medical University.

## **Statistical analysis**

The differences in G6PD, pSTAT3, and p65 expression levels between ccRCC and paired adjacent normal tissues in the immunohistological analysis was assessed using the  $\chi^2$  test. The correlations between G6PD, pSTAT3, and p65 expression levels were assessed using Pearson correlation analysis. The significance of the difference in tumor volumes between the groups in animal experiments was determined using Mixed ANOVA. For the other analyses, the differences between groups were determined using the Student *t* test ( $n = 2$ ) or one-way ANOVA ( $n \geq 3$ ). The data were expressed as means  $\pm$  standard deviation (SD) of three independent experiments. A *P* value of less than 0.05 was considered to be statistically significant.

## **Results**

### **ROS positively regulated G6PD and NF- $\kappa$ B signaling pathway in ccRCC cells**

A pilot study revealed that G6PD-facilitated ROS accumulation could positively induce G6PD transcription in RCC [18]. To further reveal the impact of ROS on G6PD expressional regulation and discover other relevant signaling factors involved in this process, ROS production was triggered or inhibited with H<sub>2</sub>O<sub>2</sub> or NAC in 786-O and Caki-1 cells. The results demonstrated that G6PD mRNA expression level was significantly decreased in NAC-treated 786-O (Fig. 1A) and Caki-1 (Fig. 1B) cells, whereas it was obviously increased in H<sub>2</sub>O<sub>2</sub>-stimulated ccRCC cells compared with the control. These results were consistent with previous study using different ccRCC cell lines [18], and further confirmed the reciprocal regulatory effect between ROS and G6PD in ccRCC. Therefore, the functional factors of signaling pathways simultaneously involved in ROS and G6PD regulation were analyzed using the MatInspector software

platform (will be discussed later in Fig. 2A). The results indicated that the NF- $\kappa$ B signaling pathway, which was demonstrated to be dysregulated and functional in the ROS-related chemotherapeutic resistance of ccRCC cells [23, 24], might intervene in the regulation of G6PD expression. To verify this hypothesis, the protein expression pattern of several important functional factors of NF- $\kappa$ B pathway, including p105, p50, and p65, which has been proved to participate in signaling transduction and closely link to ccRCC tumorigenesis [23, 24, 27], was examined in NAC- or H<sub>2</sub>O<sub>2</sub>-treated 786-O and Caki-1 cells. The results from Western blot showed that ROS positively regulated both the NF- $\kappa$ B signaling pathway and G6PD expression in ccRCC cells (Fig. 1C-D). Based on the previous conclusion that G6PD promoted RCC proliferation through positive feedback regulation of ROS-pSTAT3 axis [18], the present findings led to the hypothesis that ROS regulated G6PD overexpression might not only through pSTAT3 activation but also the NF- $\kappa$ B signaling pathway.

To further clarify the interactions between G6PD and NF- $\kappa$ B signaling pathway, TNF $\alpha$  or BAY11-7082, which could simulate or inhibit the activity of NF- $\kappa$ B signaling pathway, were used to treat 786-O and Caki-1 cells. Subsequently, the expression of G6PD and NF- $\kappa$ B pathway-related functional activators, including p50 and p65 at the protein level were evaluated. The results from western blot displayed that the protein expression levels of G6PD, p50 and p65 were increased in a dose-dependent manner in TNF $\alpha$ -treated cells, whereas they were decreased in BAY11-7082-treated cells (Fig. 1E-F). Additionally, the results from real-time RT-PCR demonstrated that the mRNA expression levels of G6PD was also increased in TNF $\alpha$ -treated cells, whereas it was decreased in cells following BAY11-7082 treatment (Fig. 1G-H), which was consistent with the expression changes of G6PD at the protein level. Taken together, these findings suggested that ROS-mediated NF- $\kappa$ B signaling pathway may be involved in increased G6PD transcription in ccRCC.

## **NF- $\kappa$ B signaling pathway facilitated G6PD expression via direct protein-DNA interaction with p65 instead of p50**

To explore the underlying mechanism of NF- $\kappa$ B signaling pathway regulated G6PD expression in ccRCC, MatInspector software platform was used to analyze the potential regulatory factors that could bind to G6PD promoter. Fortunately, the binding site of NF- $\kappa$ B signaling-related factor was found to occupy the -1913 to -1902 of the G6PD promoter (Fig. 2A). Subsequently, a sequence containing this potential binding site was cloned to pGL3-BASIC vector (named *G6PD-luc*), and luciferase report assay was performed to determine the impact of TNF $\alpha$  or BAY11-7082 on the luciferase activity in 293T cells. As presented in Fig. 2B-C, the luciferase activity of *G6PD-luc* was significantly increased by 4.2-fold following TNF $\alpha$  (100 ng/ml) treatment, whereas the luciferase activity was significantly decreased by about 37% following BAY11-7082 (5  $\mu$ M) treatment. Additionally, repeated luciferase reporter assay using a mutant vector with the NF- $\kappa$ B-binding sequence deleted (*G6PD-luc-D*) displayed that the luciferase activity was significantly decreased compared with the wild type vector. Moreover, in TNF $\alpha$ -treated cells, the luciferase activity was increased by only 2.3fold; however, in BAY11-7082-treated cells, the luciferase activity was decreased by no more than 21% (Fig. 2D), which demonstrated that the regulatory effect of NF- $\kappa$ B signaling pathway on G6PD expression was obviously attenuated without NF- $\kappa$ B-binding sequence.

These findings indicated that NF- $\kappa$ B signaling pathway may promote G6PD overexpression through a direct transcriptional regulatory effect.

In order to explore how NF- $\kappa$ B signaling pathway contributes to G6PD transcription, oligonucleotide primers covering the potential NF- $\kappa$ B binding site were designed for ChIP analysis in ACHN, Caki-1, and 786-O cells by using anti-p65 or anti-p105/50 antibodies. The results demonstrated that p65 instead of p105/50 displayed a space-occupying effect (Fig. 2E), indicating that p65 might exert a direct regulatory effect on the G6PD promoter. To further evaluate the role of p65 in G6PD expression, the endogenous expression of p65 was knocked down with RNAi lentivirus in 786-O and Caki-1 cells. The results from real-time RT-PCR and Western blot demonstrated that both G6PD mRNA and protein expression levels were significantly declined compared with the negative control (Fig. 2F), which suggested that p65 could directly regulate G6PD transcription by occupying the binding site on the G6PD promoter. Taken together, these findings confirmed the hypothesis that NF- $\kappa$ B signaling pathway facilitated G6PD expression via direct protein–DNA interaction with p65 instead of p50.

## **p65 and pSTAT3 presented a synergistic effect on G6PD transcriptional regulation**

In a previous study conducted in our laboratory, it was found that G6PD was regulated by pSTAT3 through directly binding to -1614 bp to -1595 bp of the G6PD promoter region [18]. Interestingly, the potential p65-binding site was just adjacent to the functionally verified pSTAT3-binding site (Fig. 3A), indicating that pSTAT3 and p65 might co-occupy the binding region and interact with each other to synergistically promote G6PD overexpression. To testify this hypothesis, the ChIP assay was performed in ACHN and Caki-1 cells. Firstly, cell lysates were immunoprecipitated with anti-p65 or anti-p105/50 antibody, and the eluates were amplified with primers covering the pSTAT3-binding site (Fig. 3B). Conversely, cell lysates were immunoprecipitated with anti-pSTAT3 (S727) or anti-STAT3 antibody, and the eluates were amplified with primers covering the NF- $\kappa$ B-binding site (Fig. 3C). The analysis revealed that p65 instead of p50 was capable of binding to the pSTAT3-binding site (Fig. 3B), but neither pSTAT3 (S727) nor STAT3 interacted with the NF- $\kappa$ B effective binding site (Fig. 3C). These results indicated that pSTAT3 and p65 might form a complex and bind to the pSTAT3- rather than the NF- $\kappa$ B-binding site. In this G6PD transcriptional regulatory model, although p65 had the potential to regulate G6PD transcription independently, it tended to form a p65/pSTAT3 transcriptional complex and occupy pSTAT3 instead of its own binding site while displaying a synergistic effect with pSTAT3.

To consolidate this hypothesis, the Co-IP assay was conducted in both ACHN and Caki-1 cells to investigate the protein–protein interaction between pSTAT3 and p65. The results demonstrated that the substantial interaction between pSTAT3 and p65 indeed existed in ccRCC cells (Fig. 3D), which suggested that there may be an interaction between STAT3 and NF- $\kappa$ B signaling pathways on G6PD transcriptional regulation. Furthermore, results from luciferase report assay demonstrated that the luciferase activity of *G6PD-luc* was significantly increased following the treatment of STAT3 or NF- $\kappa$ B signaling pathways activator (Fig. 3E), whereas it was decreased following the treatment of STAT3 or NF- $\kappa$ B signaling

pathways inhibitor (Fig. 3F). Moreover, synergistic effects could be seen in combinative stimulation with these two signaling pathways activator or inhibitors (Fig. 3E-F). Taken together, these findings demonstrated that STAT3 and NF- $\kappa$ B signaling pathways presented a cross-talk and synergistic effect on G6PD transcription. The underlying mechanism was that p65 and pSTAT3 formed a p65/pSTAT3 complex, occupied the pSTAT3-binding site of the G6PD promoter, and synergistically facilitated G6PD overexpression in ccRCC.

## **NF- $\kappa$ B and STAT3 activated each other and facilitated ccRCC proliferation synergistically**

The aforementioned results suggested that STAT3 and NF- $\kappa$ B signaling pathways over-activation may synergistically contribute to ccRCC proliferation following G6PD overexpression. To further evaluate the reciprocal effect between these two pathways and the role of NF- $\kappa$ B/STAT3-mediated G6PD overexpression in ccRCC growth, 786-O cells were stimulated with the STAT3 signaling activator (IL-6) or inhibitor (STATTIC) to promote or inhibit STAT3 phosphorylation, respectively. The results from Western blot showed that the expression of p50 and p65 were significantly increased following treatment with IL-6, whereas they were decreased following treatment with STATTIC (Fig. 4A), which suggested that the STAT3 phosphorylation status could influence the activation of NF- $\kappa$ B signaling pathway. Likewise, NF- $\kappa$ B over-activation or inhibition could also positively impacted the STAT3 signaling pathway activation in a dose-dependent manner (Fig. 4B). These findings indicated a reciprocal regulatory effect between NF- $\kappa$ B and STAT3 signaling pathways in ccRCC.

It has been reported that aberrant NF- $\kappa$ B activation depends on CDK4/6-mediation and contributes to tumor cell progression [39]. Previous results from our laboratory demonstrated that pSTAT3 excessive activation contributed to G6PD-stimulated RCC cell proliferation via upregulated CyclinD1 expression [18]. In the present study, the role of NF- $\kappa$ B and STAT3 cross-talk in ccRCC growth was further explored. It was found that NF- $\kappa$ B positively regulated not only CyclinD1 but also CDK4 expression at the protein level (Fig. 4B), which suggested that NF- $\kappa$ B might synergistically cooperate with STAT3 and in part contribute to ccRCC proliferation following G6PD overexpression. To confirm this hypothesis, endogenous p65 expression was knocked down by infected 786-O and Caki-1 cells with RNAi lentivirus. The results from real-time RT-PCR and Western blot demonstrated that the expression of STAT3, CyclinD1 and CDK4 were significantly decreased at the mRNA and protein level in p65 knocked down cells (Fig. 4C-E). These findings suggested that NF- $\kappa$ B and STAT3 signaling pathways could activate each other and exhibit a synergistical proliferation-promoting regulatory effect in ccRCC.

To further evaluate the role of NF- $\kappa$ B/STAT3-mediated G6PD overexpression in ccRCC growth, the MTS assay was conducted to determine the proliferation rate of ACHN and 786-O cells following treatment with STATTIC or BAY11-7082, respectively. As shown in Fig. 4F-G, both NF- $\kappa$ B and STAT3 inhibition presented a proliferation-inhibiting effect on ACHN and 786-O cells. However, whether G6PD overexpression is involved in this growth suppression effect needs to be clarified. Therefore, further MTS analyses were performed in G6PD-overexpressing ACHN and the relevant control cells to detect the

proliferation-inhibiting effect of STAT3 or BAY11-7082. As presented in Fig. 4H-I, G6PD overexpression could reverse the impact of STAT3 and NF- $\kappa$ B inhibitor on ACHN cell proliferation. Furthermore, ACHN or 786-O cells were treated with STAT3 and BAY11-7082 independently or in combination. The results from MTS assay demonstrated that these two inhibitors displayed a synergistic growth suppression effect in both ACHN and 786-O cells (Fig. 4J), and the overexpression of G6PD could rescue this tendency (Fig. 4K), indicating that G6PD overexpression is necessary for NF- $\kappa$ B/STAT3 promoted ccRCC proliferation. Taken together, these results suggested that NF- $\kappa$ B and STAT3 signaling pathways could activate each other, synergistically promote G6PD expression and contribute to ccRCC proliferation.

### **G6PD, pSTAT3, and p65 were highly expressed and positively correlated with each other in ccRCC tissues**

To verify the aforementioned mechanistic model *in vivo*, G6PD expression profile was evaluated in 27 ccRCC tumor species and matched adjacent normal tissues by real-time RT-PCR. The results demonstrated that 16 of 27 (59.3%) tumor species exhibited high G6PD expression levels (Fig. 5A), and the statistical analysis revealed that the mRNA expression level of G6PD was significantly increased in ccRCC species compared with normal tissues (Fig. 5B). Subsequently, all the specimens were subjected to Western blot analysis for the protein expression level detection of G6PD, pSTAT3, and p65. The results demonstrated that the expression levels of all these factors were significantly increased in ccRCC tumor tissues compared with adjacent normal tissues (Fig. 5C-F). In addition, the results from Pearson correlation analysis demonstrated that G6PD, pSTAT3, and p65 were positively correlated with each other at the protein level (Fig. 5G-I). Moreover, immunohistochemistry staining was performed to analyze the expression of G6PD, pSTAT3, and p65 in tumor tissues and adjacent normal tissues. As shown in Fig. 5J, G6PD and pSTAT3 was mainly localized in the cytoplasm and nucleus of the ccRCC tumor cells, respectively; whereas the predominant localizations of p65 was seen within the whole ccRCC cells. Moreover, G6PD along with both pSTAT3 and p65 were displayed high expression levels in tumor tissues compared with adjacent normal tissues (Fig. 5J-K). Taken together, these findings demonstrated that the persistent activation of both NF- $\kappa$ B and STAT3 signals existed in ccRCC tissues, which may contribute to ccRCC tumorigenesis partially through the synergistically mediated G6PD overexpression.

### **G6PD activity inhibition attenuated the growth of ccRCC cells both *in vitro* and *in vivo***

The aforementioned results showed that NF- $\kappa$ B and pSTAT3 signaling pathways were likely to drive G6PD overexpression through the constitutively activated effect of p65 and pSTAT3, indicating that G6PD, as an important common mediator of NF- $\kappa$ B and pSTAT3 signaling, might become a potential therapeutic target for ccRCC treatment. Therefore, it was necessary to investigate whether G6PD activity inhibitors had an anti-tumor effect both *in vitro* and *in vivo*. The G6PD competitive activity inhibitor 6-aminonicotinamide (6-AN), a nicotinamide analog [40], was first tested in 786-O cells. The results demonstrated that the G6PD activity was significantly decreased in a dose-dependent manner in 786-O cells following treatment with 6-AN (Fig. 6A). Moreover, the anti-proliferative effect was seen in 786-O cells after treated with 6-AN at the indicated time points and doses (Fig. 6B). In addition, the 786-O parental tumor-bearing nude mice were treated intratumor with vehicle or 6-AN to evaluate the tumor-

inhibitory activity of 6-AN *in vivo*. As presented in Fig. 6C-E, both the tumor volume (Fig. 6C-D) and tumor weight (Fig. 6E) were reduced following 6-AN treatment compared with the vehicle. Subsequently, the expression of G6PD, p-STAT3, and p65 in tumor xenografts was compared by Western blot analysis. The results showed that the expression levels of these factors were decreased in the 6-AN-treated tumor compared with the vehicle (Fig. 6F-G). Taken together, these *in vivo* data confirmed that there may be a positive correlation between G6PD overexpression and NF- $\kappa$ B/STAT3 signaling pathway over-activation. Moreover, G6PD inhibition exhibited tumor-suppressing activities in ccRCC, indicating that G6PD might be a potential therapeutic target for ccRCC treatment.

## Discussion

Increasing evidences including results from our laboratory demonstrated that aberrant expression and activation of G6PD promoted cell proliferation and adaptation in a serial of cancers and might serve as a potential anticancer target [16–18]. In these studies, the oncogenic functions and underlying molecular mechanisms of G6PD in cancer progression have partially been clarified. However, the question why G6PD is over-activated in different types of cancers is largely unknown.

G6PD activation could be regulated at both transcriptional and post-translational levels. It has been reported that Nrf2 could contribute to elevated G6PD expression in hepatocellular carcinoma through the transcriptional regulatory effect [41]. Ma *et al.* reported that Plk1 interacted with G6PD, promoted the formation of G6PD active homodimer, and therefore promoted cancer cell growth [42]. Meanwhile, Jiang *et al.* suggested that p53 formed a complex with G6PD, but inhibited its activities by restraining G6PD homodimer formation [43]. On the contrary, TAp73, a structural homolog of p53, could activate G6PD transcription, and facilitate oncogenic cell proliferation [44]. Although G6PD could be promoted by the positive feedback regulatory action of pSTAT3 through transcriptional regulation [18], the reason for ectopic G6PD gene overexpression in ccRCC still needs to be unraveled.

Previous studies from our laboratory demonstrated that G6PD overexpression lead to the increment of G6PD-NADPH-NOX4-dependent ROS accumulation and then pSTAT3 and MAKP signaling over-activation in ccRCC [18, 19]. However, several problems, such as alterations in ROS-triggered signaling pathways or other relevant regulators responsible for G6PD high expression, remained to be clarified. This study unraveled the interactive signaling pathways involved in G6PD overexpression, including ROS, NF- $\kappa$ B, and pSTAT3. Each of these pathways has been well studied in the tumorigenesis and development of a series of human cancers [21, 28, 32, 45]. ROS production and clearance are closely related to intracellular redox homeostasis and can be regulated by many factors. Compared with normal cells, tumor cells, especially derived from kidney cancers, are in a strong state of oxidative stress [46]. It is noteworthy that G6PD plays an important role in maintaining ROS homeostasis by balancing NADP/NADPH, GSH/GSSG, and other redox systems [16, 17]. Preliminary study showed that not only G6PD regulated ROS production and pSTAT3 activation, but ROS accumulation also impacted G6PD expression [18]. Therefore, it was hypothesized that ROS-involved signaling pathways might participate in G6PD dysregulation, perform cross-talk, and form a feedback loop to promote ccRCC tumorigenesis. The present study demonstrated

that significantly reduced or increased activities of the NF- $\kappa$ B signaling pathway was found in ccRCC cells following treatment with ROS scavenger or stimulator, respectively (Fig. 1), which was consistent with the changes in pSTAT3 signaling activity and G6PD expression. The aforementioned evidences suggested that pSTAT3 and NF- $\kappa$ B signaling were transcriptional regulators and might play synergistic effects on G6PD overexpression in ccRCC. After comprehensive investigation, it was concluded that ROS induced NF- $\kappa$ B and pSTAT3 signaling over-activation. These two signaling pathways not only activated each other, but also formed a p65/pSTAT3 transcriptional complex and exhibited a reciprocal regulatory effect on promoting G6PD transcription (Fig. 7).

As the first and rate-limiting enzyme of PPP, G6PD not only mediates glucose catabolism and maintains cell redox homeostasis, but also generates sufficient precursors and plays crucial roles in the biosynthesis of lipids and nucleic acids to meet the requirement of cancer cell for rapid proliferation and progression [17, 43]. The present study suggested novel proliferation strategies of ccRCC cancer cells via dual oncogenic transcriptional factors, NF- $\kappa$ B and STAT3, which are over-activated in ccRCC and cooperatively facilitate ccRCC proliferation through inducing G6PD overexpression and then cell cycle regulators such as CyclinD1 and CDK4. Additionally, previous results described that G6PD high expression was positively correlated to lymph node metastasis, Fuhrman grade, and TNM stage of ccRCC, indicating that G6PD might be involved in promoting ccRCC metastasis. Although the mechanism of G6PD in ccRCC was not fully revealed, it was faithful that in addition to the proliferation-promoting effect, the work model unraveled in present study—NF- $\kappa$ B and pSTAT3 synergistically drove G6PD overexpression (Fig. 7)—might display more functions, especially the potential to mediate tumor metabolic reprogramming and facilitate metastasis of ccRCC patients. Therefore, more investigations are necessary to be carried out for clarifying these hypotheses.

In addition, when the endogenous expression of p65 was knocked down by RNAi lentivirus, not only p65 and G6PD expression levels were declined, a significant reduction of p105 and p50 was also found in ccRCC cell lines (Fig. 2F-G). These results indicated an interaction between p65 and p50—two functional activators of NF- $\kappa$ B signaling pathway. Additionally, p50 also responded to ROS stimulation (Fig. 1C-D) and could form a complex with pSTAT3 (data not shown). However, p50 recruited on neither the potential NF- $\kappa$ B-binding site (Fig. 2E) nor the pSTAT3-binding site (Fig. 3B) on the G6PD promoter, suggesting that p50 may be potential to interact with pSTAT3 and regulate G6PD expression, but the complex might occupy other undiscovered binding sites. Similar experiments to verify the impact of p50 on G6PD expression and the interaction between p65, p50, and pSTAT3 in ccRCC still need to be performed in future studies.

Moreover, G6PD has been pinpointed as a new biomarker in acute myeloid leukemia (AML), and its overexpression positively correlated with poor prognosis of AML patients [47]. Meanwhile, targeting G6PD induces apoptosis and enhances chemotherapeutic antitumor effects via ROS-mediated damage in certain cancer, including AML, lung cancer, breast cancer, and colorectal cancer [47, 48]. Previous study also indicated that G6PD was a potential prognostic biomarker and a promising therapeutic target for ccRCC treatment [18]. However, whether G6PD inhibition exert any antitumor effects in ccRCC is far from

being clarified. The present study investigated whether the nicotinamide analog 6-AN could affect the proliferation of ccRCC both *in vitro* and *in vivo*. 6-AN, a known competitive inhibitor of G6PD, could modulate the cytotoxicity of antineoplastic treatments, and is undergoing preclinical investigation in certain cancers [40, 49]. As shown in Fig. 6A-B, a significantly reduced proliferation rate of 786-O cells was found *in vitro*. However, the inhibition of tumor growth was not as dramatic as that mediated by G6PD silencing in our previous study [18], which most probably due to low drug bioavailability. Nevertheless, the inhibition of tumor growth was associated with decreased levels of G6PD, pSTAT3, and p65 in isolated tumor samples (Fig. 6F-G), demonstrating that abnormally activated pSTAT3 and NF- $\kappa$ B signaling pathways were positively correlated to G6PD overexpression *in vivo* and G6PD inhibition exhibited tumor-suppressive activities in ccRCC. Although developing clinical application are still challenging, the present studies indicated that G6PD-based gene therapy might provide an adjunctive approach to ccRCC treatment.

## Conclusion

Collectively, this study aimed to unravel the reason for abnormal G6PD overexpression and further revealed the proliferation repressive effect of G6PD inhibition in ccRCC. The results demonstrated ROS-stimulated NF- $\kappa$ B and pSTAT3 signaling over-activation could activate each other, and perform a cross-talk in the process of G6PD transcriptional regulation. The underlying mechanism was that p65 and pSTAT3 formed a p65/pSTAT3 complex, occupied the pSTAT3-binding site on the G6PD promoter, synergistically facilitated G6PD overexpression, and contributed to ccRCC proliferation. Moreover, G6PD activity inhibition might be a potential therapeutic strategy for ccRCC treatment (Fig. 7). Despite the aforementioned work model, the question why G6PD exhibits aberrant activities in other types of human cancers is far from being answered. Meanwhile, G6PD is a crucial metabolic regulatory factor. It is tentatively to hypothesize that G6PD must participate in the entire metabolic reprogramming of ccRCC. Overall, the results shed new light on the mechanism underlying G6PD dysregulation. Other functions and relevant mechanisms of G6PD in ccRCC carcinogenesis and metabolic reprogramming should be explored in future study.

## Abbreviations

G6PD  
Glucose 6-phosphate dehydrogenase  
ccRCC  
clear cell renal cell carcinoma  
RCC  
renal cell carcinoma  
ROS  
reactive oxygen species  
STAT3

signal transducer and activator of transcription 3

PPP

pentose phosphate pathway

## Declarations

## Acknowledgments

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## Authors' contributions

YZ and YK were responsible for the experiment design and interpretation of results. YN performed most of the experiments related to signaling pathway inhibitors and activator with the help of QZ and QH. YK, XY, and ZY collected the human ccRCC specimens. QH, XY, and ZY were in charge of the immunohistochemistry and Western blot analysis of human specimens. HB performed ChIP, Co-IP, and luciferase experiments. The animal experiments were conducted by QZ. ZY and QZ checked all results, organized figures, and wrote the manuscript with the help of YZ and YA. All authors approved the final version of this manuscript.

## Data availability statement

The data used to support the findings of this study are available from the corresponding author upon request.

## Ethics approval and consent to participate

All experiments involving human participants were performed in accordance with the regulations of the Helsinki declaration and approved by the Ethics Committee, Kunming Medical University. All patients provided prior written informed consent. All animal experiments were performed according to the

Regulations for the Administration of Affairs Concerning Experimental Animals (China, 1988) and approved by the Institutional Animal Care and Use Committee, Kunming Medical University.

## Consent for publication

Not applicable.

## Competing interests

The authors declare that they have no competing interests.

## References

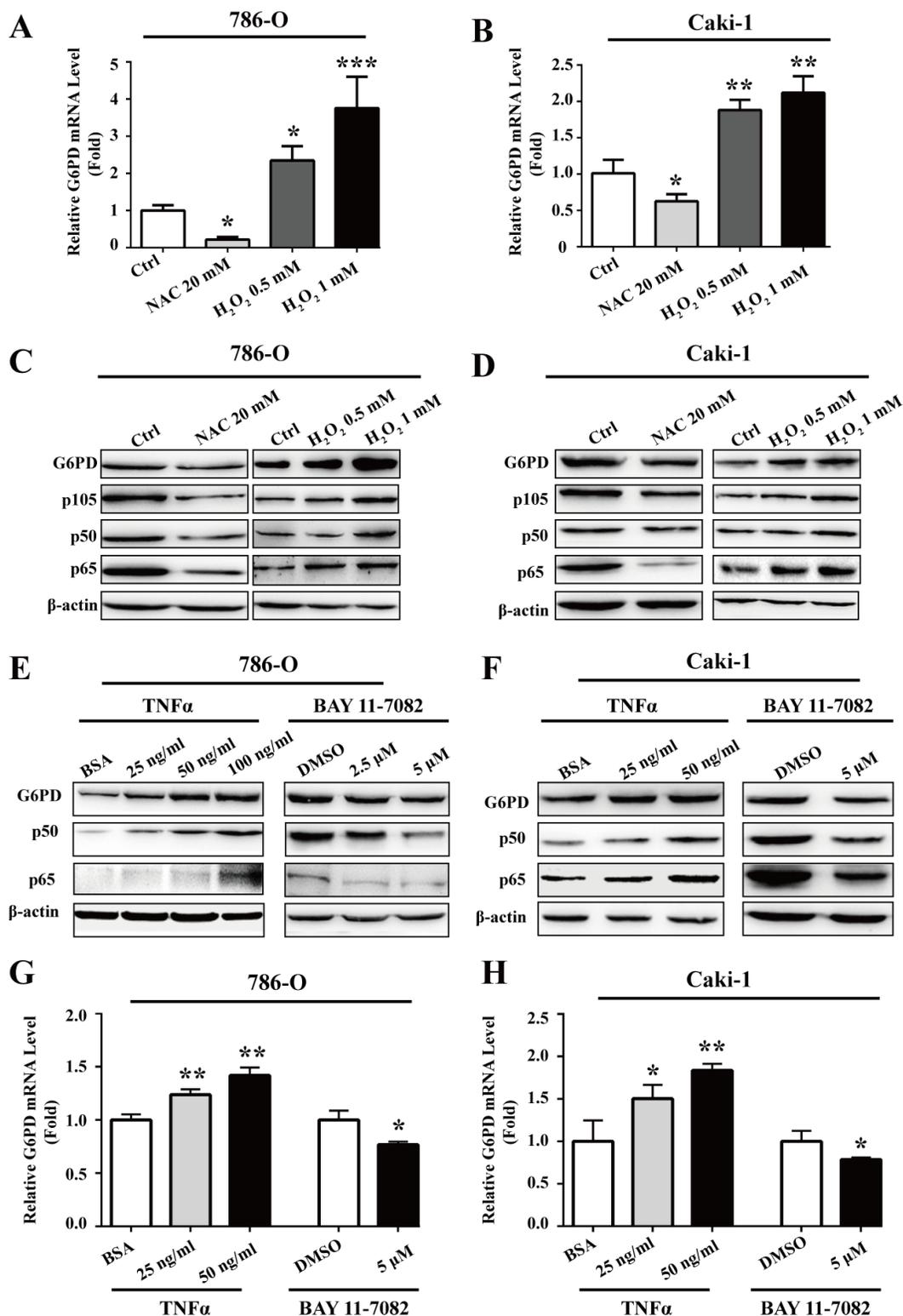
1. Hsieh JJ, et al. Renal cell carcinoma. *Nat Rev Dis Primers*. 2017 Mar;9:3:17009.
2. Escudier B, et al. Renal cell carcinoma: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up dagger. *Ann Oncol*. 2019 May 1;30(5):706 – 20.
3. Ghatalia P, et al. The Evolution of Clinical Trials in Renal Cell Carcinoma: A Status Report for 2013–2016 from the ClinicalTrials.gov Website. *Kidney Cancer*. 2017 Nov 27;1(2):151–9.
4. Global Burden of Disease. Cancer C, et al. Global, Regional, and National Cancer Incidence, Mortality, Years of Life Lost, Years Lived With Disability, and Disability-Adjusted Life-Years for 29 Cancer Groups, 1990 to 2017: A Systematic Analysis for the Global Burden of Disease Study. *JAMA Oncol*. 2019 Sep 27.
5. Hsieh JJ, et al. Genomic classifications of renal cell carcinoma: a critical step towards the future application of personalized kidney cancer care with pan-omics precision. *J Pathol*. 2018 Apr;244(5):525–37.
6. Clark DJ, et al. Integrated Proteogenomic Characterization of Clear Cell Renal Cell Carcinoma. *Cell*. 2019 Oct 31;179(4):964–83 e31.
7. Linehan WM, Ricketts CJ. The Cancer Genome Atlas of renal cell carcinoma: findings and clinical implications. *Nat Rev Urol*. 2019 Sep;16(9):539–52.
8. Chen W, et al. Targeting renal cell carcinoma with a HIF-2 antagonist. *Nature*. 2016 Nov 3;539(7627):112–7.
9. Sanchez DJ, Simon MC. Genetic and metabolic hallmarks of clear cell renal cell carcinoma. *Biochim Biophys Acta Rev Cancer*. 2018 Aug;1870(1):23–31.
10. Linehan WM, Srinivasan R, Schmidt LS. The genetic basis of kidney cancer: a metabolic disease. *Nat Rev Urol*. 2010 May;7(5):277–85.
11. Cancer Genome Atlas Research N. Comprehensive molecular characterization of clear cell renal cell carcinoma. *Nature*. 2013 Jul 4;499(7456):43–9.

12. Ricketts CJ, et al. SnapShot: Renal Cell Carcinoma. *Cancer Cell*. 2016 Apr 11;29(4):610- e1.
13. Wettersten HI, Aboud OA, Lara PN Jr, Weiss RH. Metabolic reprogramming in clear cell renal cell carcinoma. *Nat Rev Nephrol*. 2017 Jul;13(7):410–9.
14. Hakimi AA, et al. An Integrated Metabolic Atlas of Clear Cell Renal Cell Carcinoma. *Cancer Cell*. 2016 Jan 11;29(1):104–16.
15. Wettersten HI, et al. Grade-Dependent Metabolic Reprogramming in Kidney Cancer Revealed by Combined Proteomics and Metabolomics Analysis. *Cancer Res*. 2015 Jun 15;75(12):2541–52.
16. Yang HC, et al. The Redox Role of G6PD in Cell Growth, Cell Death, and Cancer. *Cells*. 2019 Sep 8;8(9).
17. Zhang C, Zhang Z, Zhu Y, Qin S. Glucose-6-phosphate dehydrogenase: a biomarker and potential therapeutic target for cancer. *Anticancer Agents Med Chem*. 2014 Feb;14(2):280–9.
18. Zhang Q, et al. Overexpression of G6PD Represents a Potential Prognostic Factor in Clear Cell Renal Cell Carcinoma. *J Cancer*. 2017;8(4):665–73.
19. Zhang Q, et al. G6PD facilitates clear cell renal cell carcinoma invasion by enhancing MMP2 expression through ROSMAPK axis pathway. *Int J Oncol*. 2020 Apr 8.
20. Rhee SG. Cell signaling. H<sub>2</sub>O<sub>2</sub>, a necessary evil for cell signaling. *Science*. 2006 Jun 30;312(5782):1882–3.
21. Pelicano H, Carney D, Huang P. ROS stress in cancer cells and therapeutic implications. *Drug Resist Updat*. 2004 Apr;7(2):97–110.
22. Fan TF, et al. Dihydromyricetin promotes autophagy and apoptosis through ROS-STAT3 signaling in head and neck squamous cell carcinoma. *Oncotarget*. 2016 Jul 25.
23. Morgan MJ, Liu ZG. Crosstalk of reactive oxygen species and NF-kappaB signaling. *Cell Res*. 2011 Jan;21(1):103–15.
24. Li G, et al. Activation of NF-kappaB pathways mediating the inflammation and pulmonary diseases associated with atmospheric methylamine exposure. *Environ Pollut*. 2019 Sep;252(Pt B):1216–24.
25. Xian D, et al. Emerging Perspective: Role of Increased ROS and Redox Imbalance in Skin Carcinogenesis. *Oxid Med Cell Longev*. 2019;2019:8127362.
26. Kim EK, et al. Redox-Mediated Mechanism of Chemoresistance in Cancer Cells. *Antioxidants (Basel)*. 2019 Oct 10;8(10).
27. Wang Y, et al. The interaction of YBX1 with G3BP1 promotes renal cell carcinoma cell metastasis via YBX1/G3BP1-SPP1- NF-kappaB signaling axis. *J Exp Clin Cancer Res*. 2019 Sep 3;38(1):386.
28. Lorente D, et al. The role of STAT3 protein as a prognostic factor in the clear cell renal carcinoma. Systematic review. *Actas Urol Esp*. 2019 Apr;43(3):118–23.
29. Cuadros T, et al. HAVCR/KIM-1 activates the IL-6/STAT-3 pathway in clear cell renal cell carcinoma and determines tumor progression and patient outcome. *Cancer Res*. 2014 Mar 1;74(5):1416-28.
30. Lafarge S, et al. Complexes between nuclear factor-kappaB p65 and signal transducer and activator of transcription 3 are key actors in inducing activation-induced cytidine deaminase expression and

- immunoglobulin A production in CD40L plus interleukin-10-treated human blood B cells. *Clin Exp Immunol*. 2011 Nov;166(2):171–83.
31. Kim SJ, Yoon S. Activated Rac1 regulates the degradation of I $\kappa$ B and the nuclear translocation of STAT3-NF $\kappa$ B complexes in starved cancer cells. *Exp Mol Med*. 2016 May;6:48:e231.
  32. Yoon S, et al. NF- $\kappa$ B and STAT3 cooperatively induce IL6 in starved cancer cells. *Oncogene*. 2012 Jul 19;31(29):3467–81.
  33. Joung YH, et al. Methylsulfonylmethane Inhibits RANKL-Induced Osteoclastogenesis in BMMs by Suppressing NF- $\kappa$ B and STAT3 Activities. *PLoS One*. 2016;11(7):e0159891.
  34. Nan J, et al. TNFR2 Stimulation Promotes Mitochondrial Fusion via Stat3- and NF- $\kappa$ B-Dependent Activation of OPA1 Expression. *Circ Res*. 2017 Aug 4;121(4):392–410.
  35. Albensi BC. What Is Nuclear Factor Kappa B (NF- $\kappa$ B) Doing in and to the Mitochondrion? *Front Cell Dev Biol*. 2019;7:154.
  36. Grivennikov SI, Karin M. Dangerous liaisons: STAT3 and NF- $\kappa$ B collaboration and crosstalk in cancer. *Cytokine Growth Factor Rev*. 2010 Feb;21(1):11–9.
  37. Kuo WY, et al. STAT3/NF- $\kappa$ B-Regulated Lentiviral TK/GCV Suicide Gene Therapy for Cisplatin-Resistant Triple-Negative Breast Cancer. *Theranostics*. 2017;7(3):647–63.
  38. Zhang Q, et al. ISL-1 is overexpressed in non-Hodgkin lymphoma and promotes lymphoma cell proliferation by forming a p-STAT3/p-c-Jun/ISL-1 complex. *Mol Cancer*. 2014 Jul;29:13:181.
  39. Jin X, et al. Phosphorylated RB Promotes Cancer Immunity by Inhibiting NF- $\kappa$ B Activation and PD-L1 Expression. *Mol Cell*. 2019 Jan 3;73(1):22–35 e6.
  40. Walker DL, et al. Murine pharmacokinetics of 6-aminonicotinamide (NSC 21206), a novel biochemical modulating agent. *Biochem Pharmacol*. 1999 Sep 15;58(6):1057–66.
  41. Liu B, et al. Hepatitis B virus stimulates G6PD expression through HBx-mediated Nrf2 activation. *Cell Death Dis*. 2015 Nov 19;6:e1980.
  42. Ma X, et al. Polo-like kinase 1 coordinates biosynthesis during cell cycle progression by directly activating pentose phosphate pathway. *Nat Commun*. 2017 Nov 15;8(1):1506.
  43. Jiang P, et al. p53 regulates biosynthesis through direct inactivation of glucose-6-phosphate dehydrogenase. *Nat Cell Biol*. 2011 Mar;13(3):310–6.
  44. Du W, et al. TAp73 enhances the pentose phosphate pathway and supports cell proliferation. *Nat Cell Biol*. 2013 Aug;15(8):991–1000.
  45. Pramanik KC, Makena MR, Bhowmick K, Pandey MK. Advancement of NF- $\kappa$ B Signaling Pathway: A Novel Target in Pancreatic Cancer. *Int J Mol Sci*. 2018 Dec 5;19(12).
  46. Xiao Y, Meierhofer D. Glutathione Metabolism in Renal Cell Carcinoma Progression and Implications for Therapies. *Int J Mol Sci*. 2019 Jul 26;20(15).
  47. Poulain L, et al. High mTORC1 activity drives glycolysis addiction and sensitivity to G6PD inhibition in acute myeloid leukemia cells. *Leukemia*. 2017 Nov;31(11):2326–35.

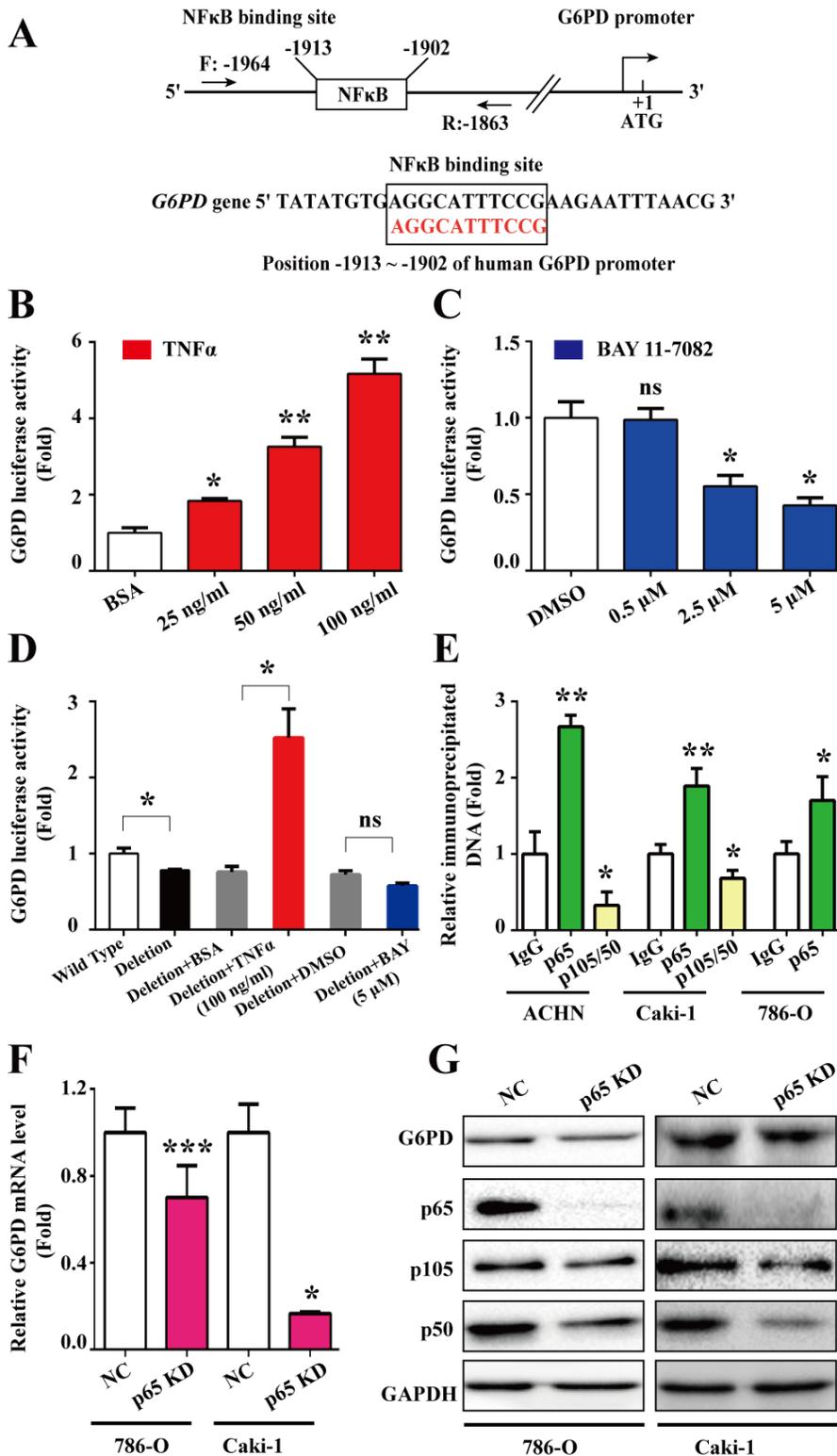
48. Ju HQ, et al. Disrupting G6PD-mediated Redox homeostasis enhances chemosensitivity in colorectal cancer. *Oncogene*. 2017 Nov 9;36(45):6282–92.
49. Budihardjo II, et al. Effect of 6-aminonicotinamide and other protein synthesis inhibitors on formation of platinum-DNA adducts and cisplatin sensitivity. *Mol Pharmacol*. 2000 Mar;57(3):529–38.

## Figures



## Figure 1

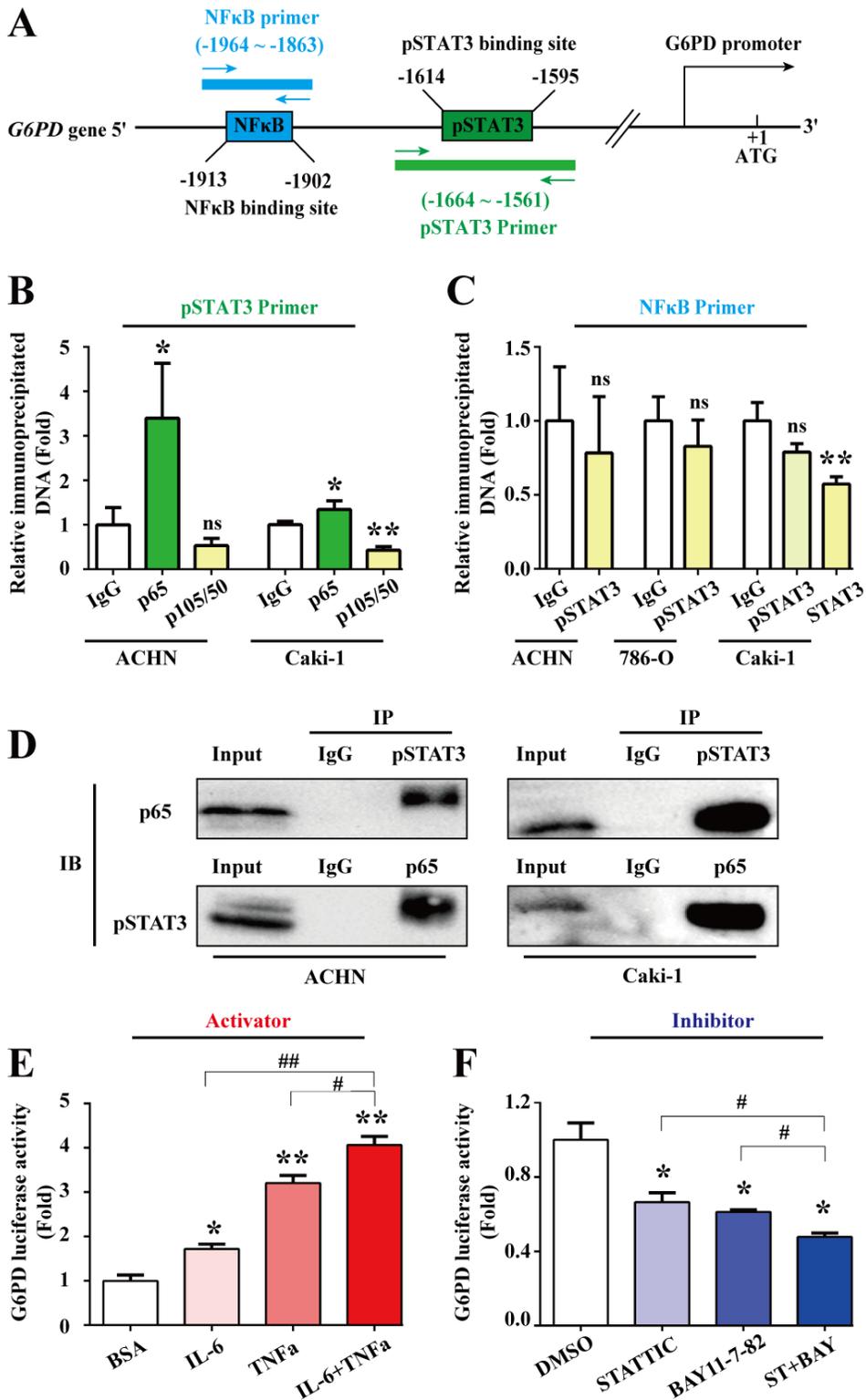
ROS positively regulated G6PD and NF- $\kappa$ B signaling pathway in ccRCC cells. (A-D) 786-O or Caki-1 cells were treated with NAC (24 h) or H<sub>2</sub>O<sub>2</sub> (2 h) to inhibit or induce ROS production, respectively. The changes in the expression of G6PD at the mRNA level and G6PD, p105, p50, and p65 at the protein level were detected using real-time RT-PCR (A-B) and Western blot (C-D) analysis, respectively. (E-H) 786-O or Caki-1 cells were treated with TNF $\alpha$  (24 h) or BAY11-7082 (24 h) at indicated doses to stimulate or inhibit NF- $\kappa$ B signaling pathway activities. The changes in the expression of G6PD, p50, and p65 at the protein level and G6PD at the mRNA level were detected using Western blot (E-F) and real-time RT-PCR (G-H) analysis, respectively.  $\beta$ -Actin was used as a loading control. Bars represent the mean  $\pm$  SD from three independent experiments, each performed in triplicate. \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001 vs each control.



**Figure 2**

NF-κB signaling pathway facilitated G6PD expression via direct protein–DNA interaction with p65 instead of p50. (A) MatInspector software platform showed that NF-κB binding site was localized at the G6PD promoter region and the primers were designed for ChIP analysis. (B-C) The G6PD promoter sequence containing the potential NF-κB binding site was cloned to a pGL3-BASIC-luc vector (named G6PD-luc) and conducted to luciferase report assay in 293T cells following treatment with TNFα (24 h) (B) or BAY11-

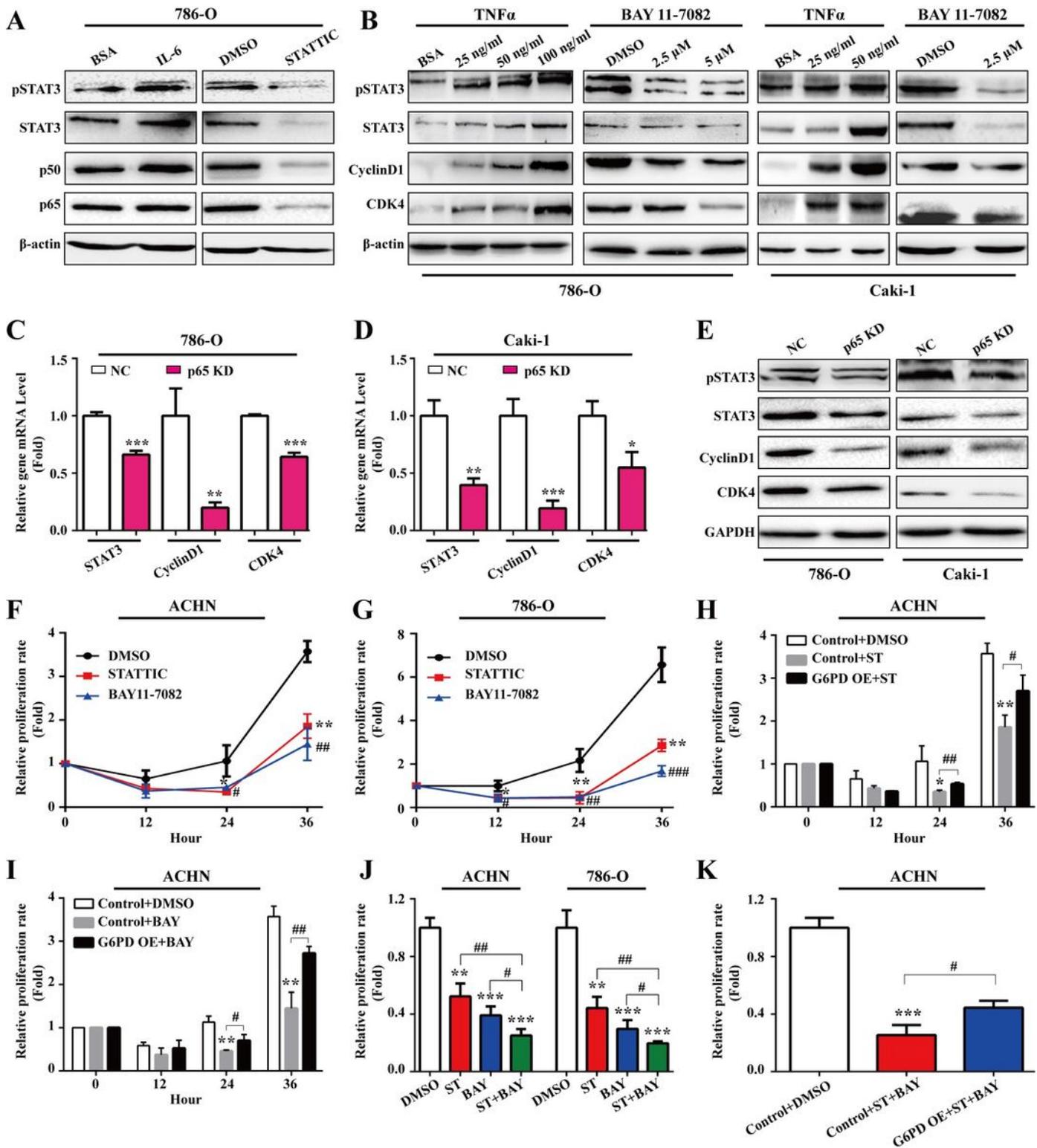
7082 (24 h) (C) at indicated doses. (D) The NF- $\kappa$ B-binding sequence deleted vector was used to repeat the luciferase reporter assay in 293T cells. (E) The critical factors involved in the NF- $\kappa$ B signaling pathway, including p65 and p50/105, were subjected to CHIP assay in ACHN, Caki-1, and 786-O cells. (F-G) p65 was knocked down by RNAi lentivirus infection in 786-O and Caki-1 cells. The changes in the expression of G6PD at the mRNA level and G6PD, p65, p105, and p50 at the protein level were detected using real-time RT-PCR (F) and Western blot (G) analysis, respectively. GAPDH was used as a loading control. Data are expressed as mean  $\pm$  SD from three independent experiments, each performed in triplicate. \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001; ns, nonsignificant vs each control. NC, Negative control lentivirus.



**Figure 3**

p65 and pSTAT3 presented a synergistic effect on G6PD transcriptional regulation. (A) MatInspector software platform showed that the potential NF- $\kappa$ B- and STAT3-binding sites localized in the G6PD promoter region were adjacent to each other. Primers covering each indicated transcriptional factor-binding region were designed. (B) ChIP assay was performed with anti-p65 or p50/105 antibodies in ACHN and Caki-1 cells, and the eluate was amplified by real-time RT-PCR with primers covering the

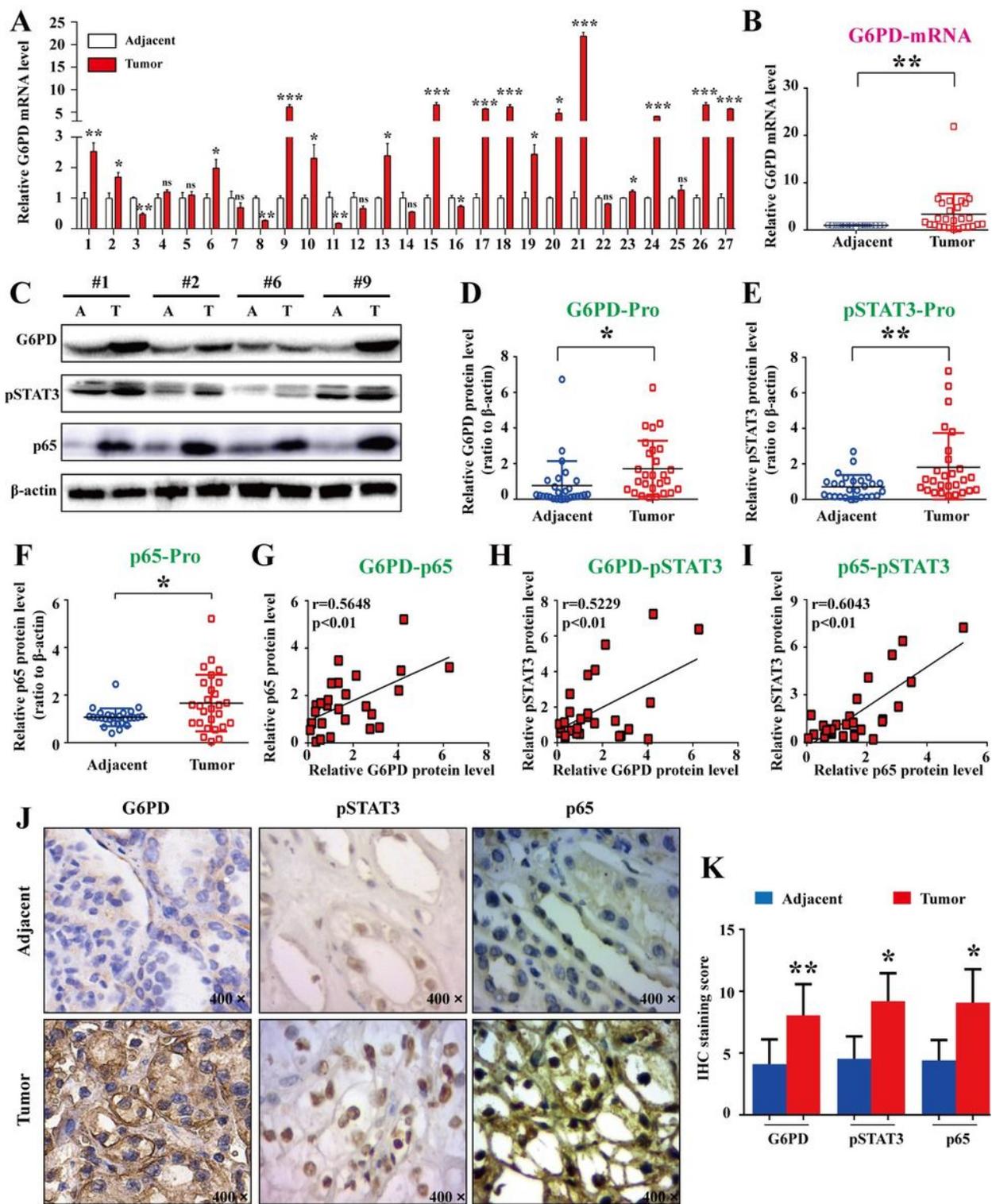
pSTAT3-binding site. (C) Similar experiments were repeated with anti-pSTAT3 or STAT3 antibody in ACHN, Caki-1, and 786-O cells, and primers covering the NF- $\kappa$ B-binding site was used. (D) The interaction between pSTAT3 and p65 was determined by Co-IP in ACHN (left panel) and Caki-1 (right panel) cells. (E-F) The luciferase activity of G6PD-luc were analyzed in 293T cells following treatment with the STAT3 or NF- $\kappa$ B signaling activator (IL-6, 2 ng/mL or TNF $\alpha$ , 50 ng/mL) (E), or inhibitor (STATTIC, 3  $\mu$ M or BAY11-7082, 2.5  $\mu$ M) (F) independently or jointly for 24 h. Data are expressed as mean  $\pm$  SD from three independent experiments, each performed in triplicate. \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001, #P < 0.05, ##P < 0.01; ns, nonsignificant vs each control. ST, STATTIC; BAY, BAY11-7082.



**Figure 4**

NF- $\kappa$ B and STAT3 activated each other and facilitated ccRCC proliferation synergistically. (A) 786-O cells were treated with pSTAT3 stimulator IL-6 (4 ng/mL) or inhibitor STATTIC (6  $\mu$ M) for 24 h. The changes in the expression of pSTAT3, STAT3, p50, and p65 at the protein level were detected using Western blot analysis. (B) 786-O or Caki-1 cells treated with TNF $\alpha$  (24 h) or BAY11-7082 (24 h) at indicated doses were subject to Western blot analysis to determine the protein expression changes of pSTAT3, STAT3,

CyclinD1, and CDK4. (C-E) 786-O or Caki-1 cells were infected with p65 RNAi lentivirus or negative control. The changes in the expression of STAT3, CyclinD1, and CDK4 at the mRNA level, and pSTAT3, STAT3, CyclinD1, and CDK4 expression at the protein level were determined by real-time RT-PCR (C-D) and Western blot (E) analysis, respectively. (F-G) The relative proliferation rates of ACHN (F) or 786-O (G) cells following treatment with DMSO (control), STATTIC (pSTAT3 inhibitor, 6  $\mu$ M), or BAY11-7082 (NF- $\kappa$ B inhibitor, 5  $\mu$ M) were measured by MTS assay at indicated time course. (H-I) The control or G6PD-overexpressing ACHN cells were treated with STATTIC (6  $\mu$ M) (H), or BAY11-7082 (5  $\mu$ M) (I) for 0, 12, 24, and 36 h, and the relative proliferation rate was determined by MTS assay. (J) ACHN and 786-O cells were treated with STATTIC (6  $\mu$ M) or BAY11-7082 (5  $\mu$ M) independently or jointly for 36 h, and the relative proliferation rate was measured by MTS assay. (K) The control or G6PD-overexpressing ACHN cells were treated with DMSO or combination of STATTIC (6  $\mu$ M) and BAY11-7802 (5  $\mu$ M) for 36 h, and the relative proliferation rate was determined by MTS assay.  $\beta$ -Actin or GAPDH was used as a loading control. Data are expressed as mean  $\pm$  SD from three independent experiments, each performed in triplicate. \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001, #P < 0.05, ##P < 0.01, and ###P < 0.001 vs each control. NC, negative control; ST, STATTIC; BAY, BAY11-7082; G6PD OE, G6PD overexpression.



**Figure 5**

G6PD, pSTAT3, and p65 were highly expressed and positively correlated with each other in ccRCC tissues. (A-B) G6PD mRNA expression level was detected by real-time RT-PCR in 27 ccRCC tumor specimens and adjacent normal tissues. (C-F) All specimens were subjected to Western blot analysis using anti-G6PD, pSTAT3, or p65 antibodies. The representative Western blot images (C) and the statistics of quantitative analysis were shown (D-F). (G-I) The correlations between G6PD, pSTAT3, and p65 at the protein level

were tested using Pearson correlation analysis. (J) Immunohistochemistry staining of G6PD, pSTAT3, and p65 in these specimens was conducted, and representative images are shown (200 $\times$ ). (K) Quantification of the staining score from (J). Data were expressed as the means  $\pm$  SD. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ ; ns, nonsignificant vs adjacent normal tissue.

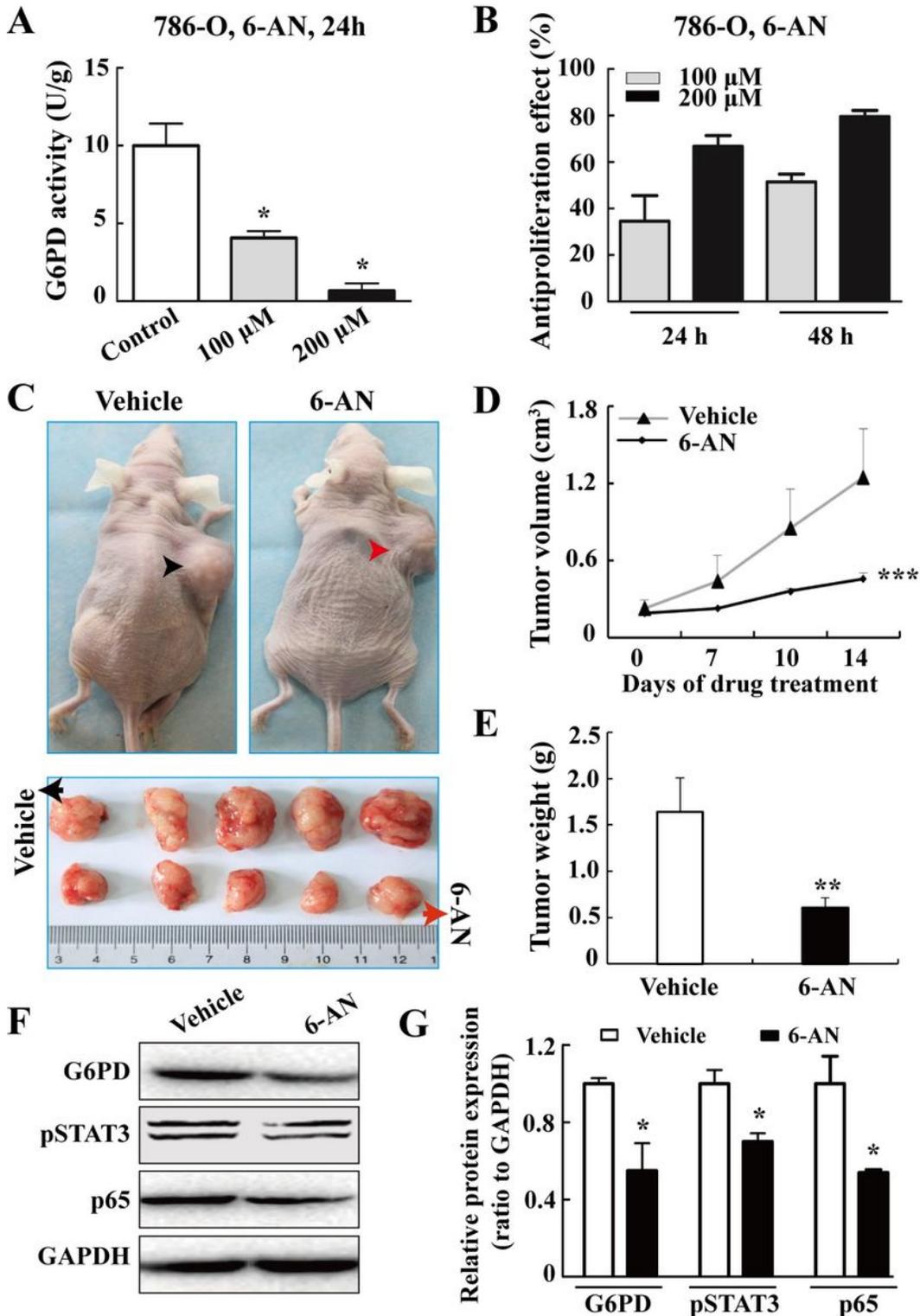


Figure 6

G6PD activity inhibition attenuated the growth of ccRCC cells both in vitro and in vivo. (A) 786-O cells were treated with 6-AN at indicated doses for 24 h, and reduced G6PD activities were verified using a G6PD Assay Kit. (B) After treating 786-O cells with 6-AN at indicated doses for 24 or 48 h, the anti-proliferative effect of 6-AN was analyzed by MTS assay. (C) 786-O parental tumor-bearing nude mice were treated with vehicle or 6-AN for 2 weeks. Representative images of tumor-bearing mice (top panel) and tumors isolated from each group (bottom panel) are shown. (D) Tumor volumes were measured at the indicated days. (E) The average tumor weights were calculated after euthanasia and tumor harvesting. (F-G) Western blot analysis (F) and grayscale scanning (G) for detecting G6PD, pSTAT3, and p65 levels in the whole-cell lysates of two random tumor samples. GAPDH levels served as loading controls. Each bar represents the mean  $\pm$  SD. \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001 vs control or vehicle.

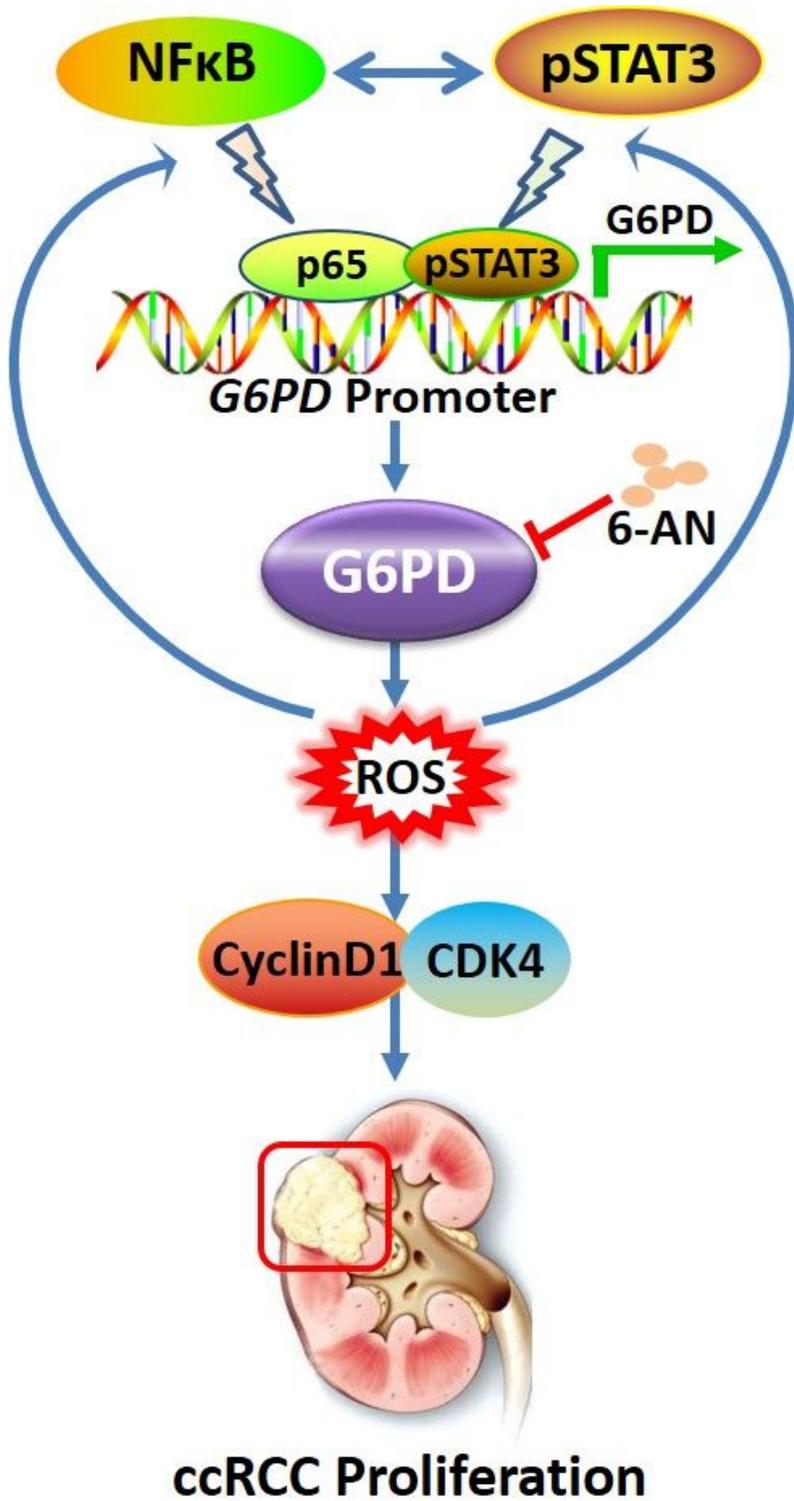


Figure 7

NF-κB and pSTAT3 synergistically drove G6PD overexpression and facilitated sensitivity to G6PD inhibition in ccRCC. ROS-stimulated NF-κB and pSTAT3 signaling over-activation could activate each other, and perform a cross-talk in the process of G6PD transcriptional regulation. The underlying mechanism was that p65 and pSTAT3 formed a p65/pSTAT3 complex, occupied the pSTAT3-binding site on the G6PD promoter, synergistically facilitated G6PD overexpression, and contributed to ccRCC

proliferation. Moreover, G6PD activity inhibition might be a potential therapeutic strategy for ccRCC treatment.