

ROCK2 promotes osteosarcoma growth and glycolysis by up-regulating HKII via phospho-PI3K/AKT signalling

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Abstract

Background

Osteosarcoma (OS) is a malignant bone tumour that exhibits a high mortality. While tumours thrive in a state of malnutrition, the mechanism by which OS cells adapt to metabolic stress through metabolic reprogramming remains unclear.

Methods

We analysed the expression of ROCK2 in osteosarcoma patients by RT-qPCR and Western blot. Cell proliferation, and colony formation were analysed using CCK8, EdU assays and colony formation assays. The level of Cell glycolysis was detected by glucose-6 phosphate, glucose consumption, lactate production and ATP levels.

Results

Herein, our study showed that ROCK2 expression in OS was higher than in adjacent tissues. Functional assays have demonstrated that ROCK2 contributes to the growth of OS cells by inducing aerobic glycolysis. The current study revealed that ROCK2 knockdown decreased the levels of mitochondrial hexokinase II (HKII). And also indicated that ROCK2 served as a key enzyme in glycolysis and that it served an important role in tumour growth and metastasis. A significant positive correlation was identified between the mRNA and protein expressions of ROCK2 and HKII, further demonstrating that ROCK2-induced glycolysis and proliferation was dependent on HKII in OS cells. Mechanistically, ROCK2 promotes HKII expression by activating the phospho-PI3K/AKT signalling pathway.

Conclusions

Taken together, the results of the current study linked the two drivers of OS growth and aerobic glycolysis, and identified a new mechanism of ROCK2 control in OS.

Background

Osteosarcoma (OS) is the most common primary malignant tumour of the bone, with an incidence rate of ~ 1–3% worldwide [1]. OS is particularly common in children and adolescents, followed by individuals over 50 years of age [2]. OS often occurs in the long bones of the limbs near the growth plate at the end of metaphyseal plate [3] and is composed of malignant osteoblasts that produce immature bone or osteoid tissue. Histologically, the cell types can be subdivided into conventional, low-grade central, periosteal, parotid gland cells, telangiectasia, chondrogenic cells and small cells [4, 5]. OS is highly malignant and prone to distant metastasis at early stages. Despite the rapid development of clinical

laboratory technology and surgery, the long-term survival rate of patients with OS remains low, particularly in patients with chemotherapy-resistant tumours, as evidenced by a 5-year survival rate of ~20% [6, 7]. OS treatment is limited by a lack of understanding regarding the mechanism behind OS recombination and metabolism in a unique tumour microenvironment. Therefore, studies assessing the regulatory metabolic targets of OS are urgently required, and insight into this molecular mechanism may reveal novel therapies and approaches that may improve the prognosis of patients.

Rho-associated coiled-coil-containing protein kinase 2 (ROCK2) is a downstream effector of the Rho subfamily of small GTPases [8]. They are activated following interaction with Rho GTPases and regulate the actin cytoskeleton through a series of pathways that regulate cell migration, cell adhesion and tumour cell invasion [9]. In the present study, ROCK2 regulated the contraction, adhesion, migration, proliferation and apoptosis of tumour cells [10]. Previous studies have also determined that ROCK2 is highly expressed in various types of tumours, including liver [11], breast [12], prostate [13] and lung cancer [14], and that high expression of ROCK2 is closely associated with poor prognosis. Overexpression of ROCK2 enhances tumour growth, while ROCK2 silencing markedly inhibits tumour growth in many types of cancer [15]. In addition, previous studies have reported that ROCK2 acts as a protein kinase, increasing the phosphorylation of substrate proteins [16, 17]. However, it is unclear whether ROCK2 affects the progression and regulation of OS.

Aerobic glycolysis, also known as the Warburg effect, is an important feature of cancer and is characterized by an increase in glucose conversion to lactic acid, independent of oxygen [18]. The Warburg effect allows tumour cells to metabolise glucose to a greater extent than normal cells, providing an energy source for tumour cell growth. Glycolysis provides sufficient energy for tumour cells, which enhances the tolerance of cells to ischemia and hypoxia, allows cells to avoid apoptosis caused by the inhibition of oxidative phosphorylation and allows tumour cells to synthesize cellular constituents during rapid growth [19–21]. Metabolism provides a rich source of material and growing evidence has indicated that aerobic glycolysis serves a critical role in the proliferation of OS cells [22, 23]. The rate-limiting enzyme in glycolysis is hexokinase II, which encodes a rate-limiting gluconeogenesis enzyme that has been reported to serve a tumour-promoting effect in many types of cancer [24–26]. HKII overexpression is associated with poor prognosis in patients with cancer [27]. However, the underlying mechanism of HKII regulation in OS remains unclear. In addition, previous studies have confirmed that ROCK2 affects the glycolysis of tumour cells [28], which is congruent to the results of the present study, which demonstrated that ROCK2 and HKII expression was increased in OS. The current study hypothesized that ROCK2 may affect the aerobic glycolysis of OS by regulating the expression of HKII. Previous studies have demonstrated that the HKII protein can be activated by the PI3K/AKT signalling pathway [29] and other studies have reported that ROCK2 affects the phosphorylation of substrate proteins [17]. Therefore, the current study speculated that ROCK2 promoted HKII expression by activating PI3K/AKT signalling via phosphorylation.

The present study revealed that ROCK2 was expressed in patients with OS, indicating that ROCK2 affected the level of glycolysis and growth in OS cells by promoting the expression of HKII. In addition,

ROCK2 was demonstrated to further regulate HKII expression by activating the PI3K/AKT signalling pathway via phosphorylation. In summary, the results of the present study may provide new targets for the treatment of patients with OS.

Materials And Methods

Tissue samples

A total of 43 OS samples and adjacent specimens were obtained from the Second Affiliated Hospital of Nanchang University and the Jiangxi Cancer Hospital. A pathologist diagnosed all specimens as OS and samples were frozen and stored at -80°C for until further analysis. The study protocol was approved by the Medical Ethics Committee of Second Affiliated Hospital of Nanchang University. This study was performed in accordance with the ethical standards of the Declaration of Helsinki.

Cell Lines And Cell Culture

Human osteosarcoma cells (U2-OS, Saos-2, 143B and MG-63) and the normal cell line, hfoBI-19, were purchased from The Cell Bank of Type Culture Collection of the Chinese Academy of Sciences. Cells were cultured in 10% FBS and maintained at 37°C with 5% CO₂. Medium was changed every 1–2 days, and cells were plated at 80–90% confluency for a 1:3 passage.

Reverse Transcription-quantitative Pcr (RT-qPCR), Western Blotting And Co-immunoprecipitation

RT-qPCR, western blotting and co-immunoprecipitation (Co-IP) were performed as previously described [9]. The primer sequences for PCR were as follows: ROCK2 forward, 5'-GGTATCTGTACATGGTAATGG-3' and reverse, 5'-GGAGTGTATTGCATCCAGAG-3'; HKII forward, 5'-GGCAATGAAACCAAAGCCAGGAG-3' and reverse, 5'-GGAAGGAGGAGCCAGAAGC-3'; GAPDH forward, 5'-CCTGCCGGTGACTAACCCCTG-3' and reverse, 5'-AGTTAAAAGCAGCCCTGGTG-3'. The antibodies of Western blot were as follow: ROCK2(Abcam 1:1000 dilution), HKII(Abcam 1: 1000 dilution),BCL-2(Abcam 1×2000 dilution), BAX(Abcam 1×2500 dilution), AKT/P-AKT(Abcam 1×500 dilution)

PI3K/P-PI3K(Abcam 1: 1000 dilution),GAPDH(Abcam 1×5000 dilution).

Immunohistochemistry (IHC)

OS and adjacent tissues were fixed in paraffin, sectioned, deparaffinized and blocked using 5% bovine serum albumin for 30 min. Sections were then incubated with anti-ROCK2 (1:200; Abcam) and anti-HKII

(1:200; Abcam) antibodies. Secondary antibodies were subsequently applied for visualization. Subjective relative nuclear staining intensity was assessed to grade ROCK2 and HKII expression.

Short Interfering (sh)RNA Plasmids And Construct

Plasmids encoding shRNA against ROCK2 and HKII were synthesized by Shanghai GenePharma Co., Ltd. Vectors and pcDNA3.1(+)-ROCK2 and HKII were also obtained from Shanghai GenePharma Co., Ltd. OS cells were transfected with shRNA plasmids, pcDNA3.1(+)-ROCK2 and HKII using Lipofectamine® 3000 (Invitrogen; Thermo Fisher Scientific, Inc.) in accordance with the manufacturer's protocol.

Cell Growth Assays

Following trypsinization, OS cell lines were seeded (1.0×10^4 cells/well) into 96-well plates with different treatment factors and counted. After a specified length of time, cell growth was assessed under different processing factors using a cell counting kit (CCK8) assay in accordance with the manufacturer's protocol. An EdU assay was performed as previously described[30].

Oxygen Consumption Rate (OCR) And Extracellular Acidification Rate (ECAR)

The Extracellular Flow Analyzer XF96 (Seahorse Bioscience) was used to measure cell glycolysis. Cell mitochondrial respiration was determined using the XF Cell Mitosis Stress Test kit and the Glycolysis Stress Test kit (each, Seahorse Bioscience), respectively. Each assay was performed according to the manufacturer's protocol.

Tumourigenicity Assay

All animal experiments were approved by the Animal Experiment Ethics Committee of Nanchang University. A total of 1×10^7 OS cells in 100 μ l PBS were counted following trypsinization and injected into the dorsal side of nude mice (male BALB/c-nu/nu; age, 5 weeks). After tumour formation, mice were observed every 5 days. Tumour volumes were measured for 35 days. After anaesthesia, tumour tissues from each group were collected and weighed, after which they were fixed in 4% neutral paraformaldehyde for subsequent experimentation. All animal experiments were approved by the Animal Experimental Ethics Committee of the Second Affiliated Hospital of Nanchang University and was performed in accordance with the "Guide for the Care and Use of Laboratory Animals" (revised 1985).

Statistical analysis.

All data obtained from the current study were expressed as the mean \pm standard deviation and analysed using GraphPad Prism 7 and SPSS 19.0 software following at least 3 independent repeats. Differences between two groups were analyzed using t-tests or Student's t-tests, Multiple groups comparisons were performed with the one-way analysis of variance (ANOVA), and the post hoc test was Student-Newman-Keuls test. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

ROCK2 is upregulated in OS tissue and cells.

To clarify the role of ROCK2 in the development of OS, ROCK2 expression was detected in OS and corresponding adjacent tissues. Following RT-qPCR and western blotting, the mRNA and protein levels of ROCK2 were revealed in tumour tissue (Fig. 1A and B). IHC was subsequently performed, the results of which revealed an increased expression of ROCK2 in cancer tissue. As presented in Fig. 1C, ROCK2 was overexpressed in 72.09% (31/43) of all OS tissues examined, while weak positive staining was observed in corresponding adjacent non-tumour tissues. The expression of ROCK2 in OS cell lines (MG-63, 143B U2-OS, and Saos-2) and in the normal cell line, hfoBI-19, was determined. The results revealed an increased expression of ROCK2 in OS cells (Fig. 1D and E). These data indicated that ROCK2 is overexpressed in OS tissue and cells.

ROCK2 promotes the growth of OS in vitro and in vivo

To further explore the functional role of ROCK2 in OS, ROCK2 shRNA was used to knockdown the expression of ROCK2 in U2OS and MG-63 OS cells (Fig. 2A and B; Fig. S1). As presented in Fig. 2C-D and Fig. S4 A-B, the results of the CCK8 and EdU assays revealed that ROCK2 downregulation significantly inhibited the growth of OS cells *in vitro*. Additionally, a colony formation assay was performed to detect OS cell clones in shROCK2- and shNC-treated cells (Fig. 2E). The results of flow cytometry demonstrated a significant increase in apoptosis following ROCK2 downregulation (Fig. 2F and Fig. S4 C). Additionally, the expression of B-cell lymphoma-2 (Bcl-2) and Bcl-2-associated protein X was analysed via western blotting (Fig. S2). The expression of ROCK2 following ROCK2 overexpression was subsequently evaluated (Fig. 3A and B). As Presented in Fig. 3C-D and Fig. S5 A-B, OS cell proliferation was significantly increased in P-ROCK2 cells. Consistent with the aforementioned results, the colony formation assay revealed that ROCK2 overexpression increased OS cell growth (Fig. 3E). Furthermore, the results of flow cytometry indicated a significant decrease in apoptosis following ROCK2 upregulation (Fig. 3F and Fig. S5 C).

To confirm the role of ROCK2 in tumour formation *in vivo*, OS cells (U2-OS and 143B) were over- and under-expressed with ROCK2. The effect of ROCK2 on tumour formation in nude mice was subsequently established. After 5 weeks of growth, ROCK2-knockdown U2-OS cells (U2-OS/shROCK2) exhibited reduced tumour growth in nude mice (Fig. 2H; Fig. S3). Consistent with this result, the average tumour weights of mice bearing U2-OS/shROCK2 cells were significantly decreased (Fig. 2I and J), while ROCK2-overexpressing 143B cells (143B/p-ROCK2) resulted in significantly increased tumour growth compared

with respective control cells (Fig. 3H). Tumours in mice bearing 143B/p-ROCK2 cells were markedly increased in size compared with those of their respective controls (Fig. 3I and J). In summary, these data indicated that ROCK2 regulated the growth of OS *in vivo* and *in vitro*.

ROCK2 Affects The Level Of Glycolysis In OS Cells

Given that aerobic glycolysis is indicative of changes to cellular metabolism, this shift is ubiquitous in tumour cells. Therefore, the current study investigated whether ROCK2 affected the metabolism OS cells. As presented in Fig. 4A-D, glucose-6 phosphate levels, glucose consumption, lactate production and ATP levels were all significantly reduced in OS cells where ROCK2 expression was downregulated. In contrast, ROCK2 overexpression produced the opposite affects in Saos-2 cells (Fig. 4E-H).

To further confirm the role of ROCK2 in the glycolytic hydrolysis of OS, OS cell glycolysis was determined via ECAR. As presented in Fig. 4I, ROCK2 knockdown significantly decreased the glycolytic rate and capacity of U2-OS cells, whereas ROCK2 overexpression increased ECAR in Saos-2 cells (Fig. 4K). The current study assessed OCR, an indicator of mitochondrial respiration. The results revealed that U2-OS/shROCK2 cells exhibited an increased OCR (Fig. 4J), whereas ROCK2 overexpression produced the opposite effect in Saos-2 cells (Fig. 4L). In summary, the results indicated that ROCK2 may promote aerobic glycolysis and inhibit mitochondrial respiration in OS cells.

ROCK2 Positively Regulates HKII Expression

Hexokinase, a key enzyme in glycolysis, has been revealed to serve a crucial role in the regulation of glycolysis. Among the enzymes that participate in glycolysis, HKII is of particularly importance. Previous studies have confirmed that HKII is upregulated in liver [31], colorectal [32] and lung cancer [33]. Since HKII is closely associated with the proliferation of tumour cells and in order to confirm the expression of HKII in OS, the current study detected the expression of HKII in OS tissues via RT-qPCR and western blotting (Fig. 5A-C). The results of IHC confirmed the increased expression of HKII in OS (Fig. 5D). Scatter plots revealed that ROCK2 and HKII mRNA and protein levels were positively correlated with OS tissues (Fig. 5E and F). The expression of HKII in OS cells was further confirmed via RT-qPCR and western blotting (Fig. 5G-H).

To verify whether ROCK2 regulates HKII expression, ROCK2 was up- and down-regulated in OS cells, the results of which demonstrated that ROCK2 knockdown significantly reduced the mRNA and protein expression of HKII in U2-OS and MG-63 cells. In contrast, ROCK2 overexpression significantly increased HKII protein and mRNA levels in Saos-2 and 143B cells (Fig. 5I and J). Taken together, these results indicated that ROCK2 positively regulates the expression of HKII in OS cells.

ROCK2 regulates glycolysis and cell proliferation depending on the expression of HKII

As aforementioned, HKII was determined to be a downstream gene of glycolysis. The current study therefore upregulated the expression of HKII in U2-OS cells that were previously treated to knockdown ROCK2. Successful HKII overexpression was confirmed via immunoblotting (Fig. 6A). The results of CCK8 and EdU assays revealed that HKII overexpression restored the proliferation of shROCK2-treated cells (Fig. 6B-C and Fig. S6 A). The results also demonstrated that HKII overexpression restored glucose-6 phosphate levels, glucose consumption, lactate production and ATP levels in shROCK2-treated cells (Fig. 6D-G).

The expression of HKII was silenced in p-ROCK2 OS cells. Western blot analysis confirmed that HKII expression was increased in P-ROCK2 cells and that its expression was inhibited following HKII silencing (Fig. 6H). Similarly, OS cell proliferation was also inhibited (Fig. 6I-J and S6 B). Further studies confirmed that silencing HKII in P-ROCK2 inhibited glucose-6 phosphate levels, glucose consumption, lactate production and ATP levels (Fig. 6K-N). In conclusion, ROCK2 regulated glucose metabolism and proliferation in OS cells depending on the expression of HKII.

ROCK2 regulates HKII expression by activating the phosphorylated PI3K/AKT signalling pathway

To further investigate the mechanism by which ROCK2 regulates HKII expression, it was determined whether ROCK2 and HKII could be intrinsically bound. However, the results of Co-IP revealed no intrinsic link between them (Fig. 7A). Additionally, it has been reported that HKII is affected by PI3K/AKT signalling pathways [29] and that AKT phosphorylates HKII at Thr-473 [34]. Previous studies have also confirmed that ROCK2 acts as a protein kinase that affects the phosphorylation of substrate proteins [16, 17]. Therefore, the current study hypothesized that ROCK2 may promote HKII expression by activating the PI3K/AKT signalling pathway. To verify this hypothesis, ROCK2 expression was reduced and immunoblotting revealed that total AKT and PI3K levels did not change. However, levels of phosphorylated AKT and PI3K decreased significantly (Fig. 7B). Furthermore, AKT and PI3K phosphorylation was significantly increased in cells in which ROCK2 was upregulated (Fig. 7C). To further verify whether ROCK2 regulates the expression of HKII by activating phosphorylated PI3K/AKT and whether it affects OS growth, ROCK2 was overexpressed and OS cells were treated with the PI3K inhibitor LY294002 (10 μ M). Western blot analysis confirmed that HKII expression was inhibited (Fig. 7D). The results of the CCK8 and EdU assay also indicated that OS growth was inhibited (Fig. 7E and F; Fig. S4). The results indicated that ROCK2 activated AKT and PI3K via phosphorylation, which induced HKII expression and further promoted the growth of OS cells.

Discussion

Although the prognosis and treatment of OS has improved, patient survival rates remain poor [35]. Therefore, a better understanding of the biological properties of OS is essential. In recent years, the importance of metabolic reprogramming in tumorigenesis and progression has been confirmed. Metabolic reprogramming is considered one of the important features of cancer. OS seriously affects adolescent health and due to OS exhibiting varying degrees of metabolic dependence, survival rates are

extremely low. Therefore, understanding the basis of OS from a metabolic perspective may provide novel insights into this disease. The current study demonstrated that ROCK2 expression was upregulated in OS and that it serves a key role in the reprogramming of glucose metabolism in this type of cancer. Therefore, studying the molecular and regulatory mechanisms of glucose metabolism reprogramming may provide more evidence for OS targeted therapy.

Members of the Rho GTPase family are important regulators of cell migration, proliferation and apoptosis [10]. The stimulation of contraction and adhesion is achieved by inducing the formation of actin stress fibres and focal adhesions of fibroblasts, as well as by modulating surface integrins to aggregate platelets and lymphocytes. Recent research has confirmed that ROCK2 is highly expressed in liver, colon and kidney cancer, and that its expression is closely associated with poor patient prognosis [36, 37]. ROCK2 silencing significantly inhibits tumour cell growth and metastasis, and promotes cell apoptosis. The current study revealed a novel mechanism by which OS promotes glycolysis and cell proliferation. The present study confirmed that ROCK2 and HKII were upregulated in OS tissues and cells, and that these two proteins were positively correlated. Furthermore, experimental data also demonstrated that HKII expression was significantly reduced after ROCK2 expression was silenced and that aerobic glycolysis and OS cell proliferation were also inhibited. HKII upregulation via ROCK2 knockdown reversed the decrease in OS cell proliferation and aerobic glycolysis, whereas HKII downregulation significantly attenuated the increase in cell proliferation and aerobic glycolysis induced by ROCK2 overexpression. Overall, the results revealed a novel mechanism whereby ROCK2 regulates HKII and thus affects the growth and glycolysis of OS cells.

The present study evaluated the molecular mechanism by which ROCK2 regulates HKII. Since ROCK2 stabilizes the expression of substrate proteins, the results of Co-IP confirmed that ROCK2 and HKII cannot bind. However, previous studies have reported that HKII is regulated by the PI3K/AKT signalling pathway [29]. ROCK2 activates substrate proteins via phosphorylation. The present study silenced the expression of ROCK2 and demonstrated that total levels of PI3K and AKT did not change following treatment. In addition, the level of PI3K/AKT phosphorylation was significantly inhibited. In contrast, ROCK2 overexpression activated PI3K and AKT phosphorylation. Following ROCK2 overexpression and subsequent treatment with the PI3K inhibitor, LY294002, it was demonstrated that HKII expression was inhibited. The results revealed a novel mechanism for ROCK2 in the regulation of HKII expression via the activation of PI3K/AKT by means of phosphorylation.

Conclusions

The current study demonstrated that ROCK2 is upregulated in OS. The results also revealed that ROCK2 may promote OS cell growth and aerobic glycolysis. Furthermore, the aerobic glycolysis and proliferation of OS cells was demonstrated to rely on HKII. The results also indicated that ROCK2 activated phosphorylated PI3K/AKT and regulated HKII expression. Based on the current results, ROCK2 may be used as a potential biomarker for the future diagnosis and treatment of OS.

Abbreviations

ROCK2: Rho-associated coiled-coil-containing protein kinase 2. HKII: Hexokinase II. OS: Osteosarcoma. RT-qPCR: Reverse transcription-quantitative PCR. shRNA: Short interfering RNA. OCR: Oxygen consumption rate. ECAR: Extracellular acidification rate.

Declarations

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Authors' contributions

Liang Hao and Xueqiang Deng conceived and designed the study. Xuan Yi and Jianguo Deng performed the experiments and collected data; Yeqing Zou performed the data analysis; All authors discussed and interpreted the data. Xueqiang Deng wrote the manuscript. All authors reviewed and edited the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials

All the data in the manuscript are available upon reasonable request.

Ethics approval and consent to participate

In the study, the use of human data and human tissue was clearly approved by the Ethics Committee of the Second Affiliated Hospital of Nanchang University. The use of All animal experiments were approved by the Animal Experimental Ethics Committee of the Second Affiliated Hospital of Nanchang University and was performed in accordance with the "Guide for the Care and Use of Laboratory Animals" (revised 1985)

Consent for publication

Not applicable

Competing interests

The author declare that they have no competing interests

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Figures

Figure 1

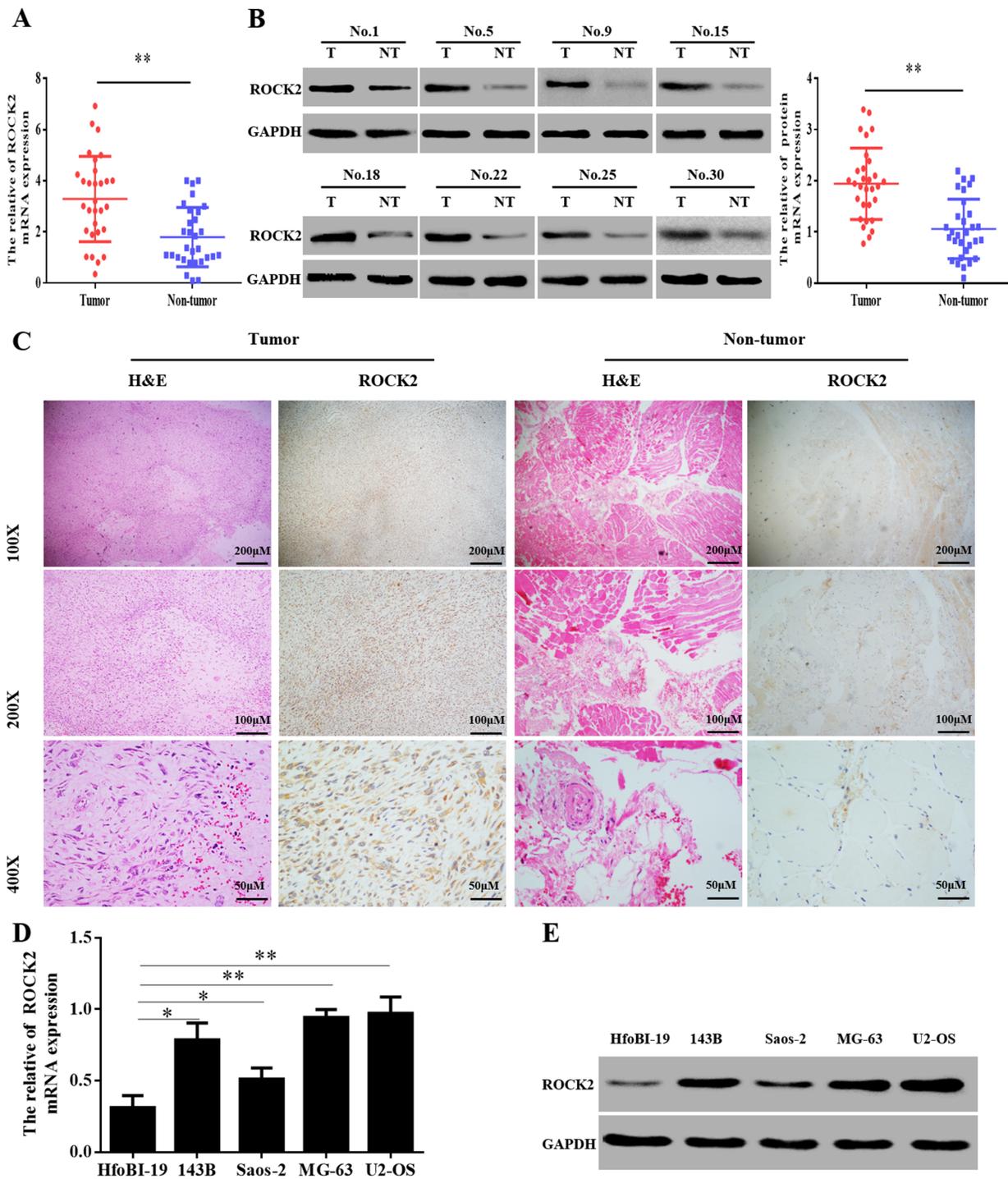


Figure 1

ROCK2 is upregulated in OS tissue and cells. (A) RT-qPCR analysis of ROCK2 mRNA expression in 30 cases of OS and corresponding adjacent tissues. (B) Analysis of ROCK2 protein expression in OS and corresponding adjacent tissues, as determined via immunoblotting. GAPDH was used as a loading control. (C) Representative images of ROCK2 staining in 43 paired OS tissues (scale bar, 50 μ m). (D and E) ROCK2 expression in OS cells (U2-OS, Saos-2, 143B and MG-63) and normal hfoBI-19 cells was

detected via RT-qPCR and western blotting. GAPDH was used as a loading control. * $P < 0.05$, ** $P < 0.01$. ROCK2, Rho-associated coiled-coil-containing protein kinase 2; OS, osteosarcoma; RT-qPCR, reverse transcription-quantitative PCR.

Figure 2

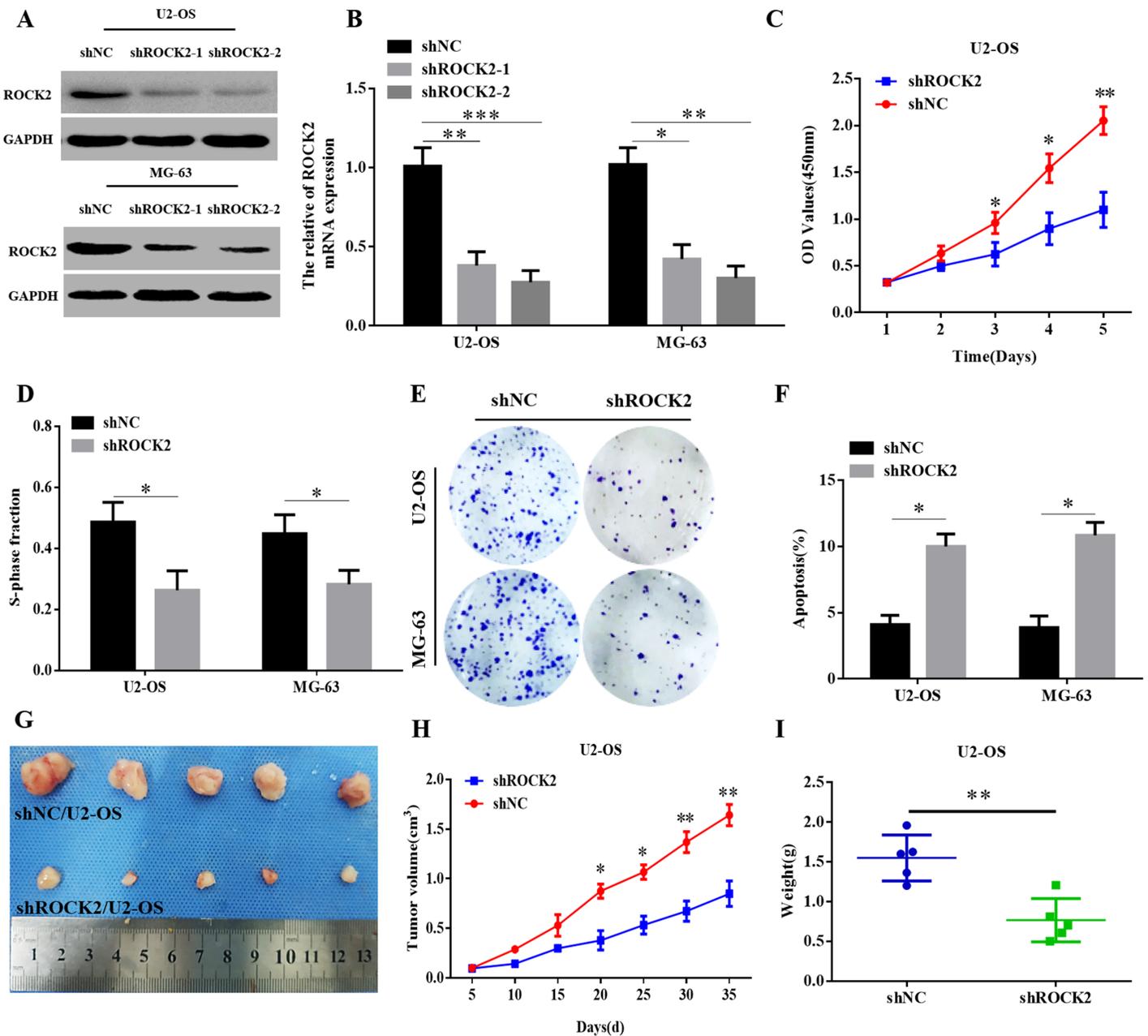


Figure 2

Knockdown of ROCK2 expression inhibits OS growth in vitro and in vivo. (A) mRNA and (B) protein levels of ROCK2 in U2-OS and MG-63 cells following transfection with shROCK2-1, shROCK2-2 and shNC (control) were determined via reverse transcription-quantitative PCR and western blotting. (C) Cell counting kit 8 and (D) EdU assays demonstrated a decrease in shROCK2-transfected OS cell (U2-OS and MG-63) growth compared with shNC-transfected cells (scale bars:100 μ m). (E) Representative images of

the colony formation assay performed using OS cells transfected with shROCK2 or shNC. (F and G) Flow cytometry analysis of OS cell apoptosis following ROCK2 silencing. (H and J) shROCK2/U2-OS and shNC/U2-OS cells were subcutaneously injected into nude mice, after which tumour volumes were measured on the indicated days. At the endpoint, tumours were dissected, imaged and weighed (n=5). *P<0.05, **P<0.01 and ***P<0.001. ROCK2, Rho-associated coiled-coil-containing protein kinase 2; OS, osteosarcoma; sh, short interfering; NC, negative control.

Figure 3

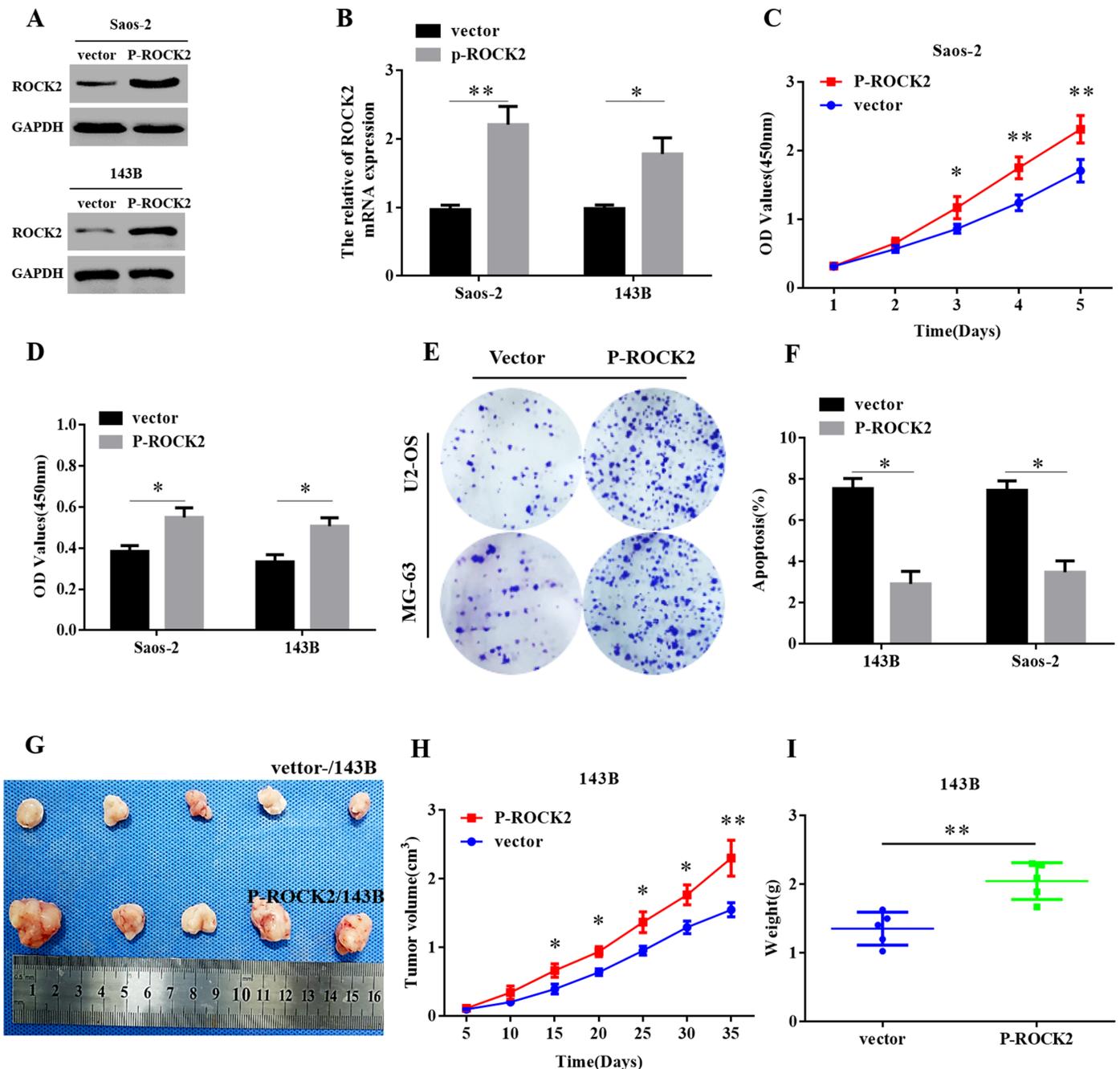


Figure 3

Overexpression of ROCK2 promotes the growth of OS in vitro and in vivo. (A and B) Saos-2 and 143B OS cell lines were transfected with pcDNA3.1(+)-ROCK2 and vectors. ROCK2 mRNA and protein expressions were subsequently determined via reverse transcription-quantitative PCR and western blotting, respectively. Following ROCK2 overexpression, (C) CCK8 and (D) EdU assays confirmed that the proliferation of OS cells was significantly increased (scale bars:100 μ M). (E) Representative images obtained from the colony formation assay of OS cells transfected with p-ROCK2 and vectors. (F and G) following ROCK2 overexpression, flow cytometry was performed to determine the apoptosis rate of OS cells. (H and J), p-ROCK2/143B and vector/143B cells were subcutaneously injected into nude mice, and tumour volumes were measured on the indicated days. At the endpoint, tumours were dissected, imaged and weighed (n=5). *P<0.05 and **P<0.01. ROCK2, Rho-associated coiled-coil-containing protein kinase 2; OS, osteosarcoma; p, phosphorylated.

Figure 4

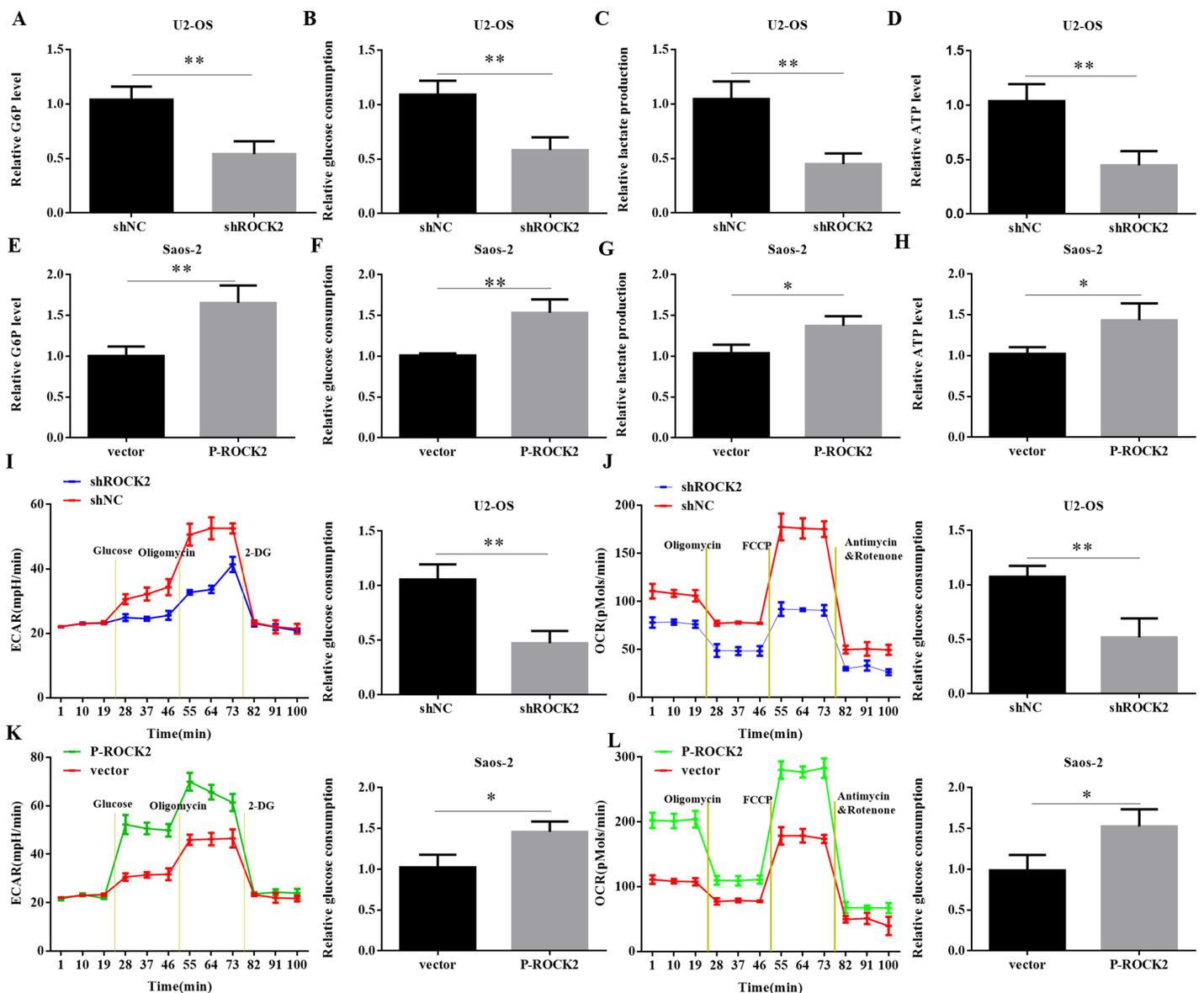


Figure 4

ROCK2 affects the level of glycolysis in OS cells. (A) Cellular G6P levels, (B) glucose consumption, (C) lactate production and (D) ATP levels were analysed in shROCK2 and shNC-treated cells. (E and H) Cellular G6P levels, glucose consumption, lactate production and ATP levels were analysed in OS cells after transfection with p-ROCK2 or vectors. Extracellular acidification rate data revealed the glycolytic rate and capacity in (I) shROCK2 or (J) p-ROCK2 cells. Glucose (10 mM), the oxidative phosphorylation inhibitor oligomycin (1 μ M) and the glycolytic inhibitor 2-deoxyglucose (50 mM) were sequentially added to each well at the indicated time point. All measurements were normalized to the cell number calculated using crystal violet at the end of the experiment. Oxygen consumption rate results revealed the basal respiration and maximum respiration of (K) shROCK2 cells and (L) p-ROCK2 cells. Oligomycin (1 μ M), the mitochondrial uncoupler carbonyl cyanide p-trifluoromethoxy phenylhydrazone (1.0 μ M) and the mitochondrial complex I inhibitor rotenone in addition to the mitochondrial complex III inhibitor antimycin A (0.5 μ M) were sequentially added. All measurements were normalized to the cell number calculated using crystal violet at the end of the experiment. * $P < 0.05$ and ** $P < 0.01$. ROCK2, Rho-associated coiled-coil-containing protein kinase 2; OS, osteosarcoma; sh, short interfering; NC, negative control; p, phosphorylated.

Figure 5

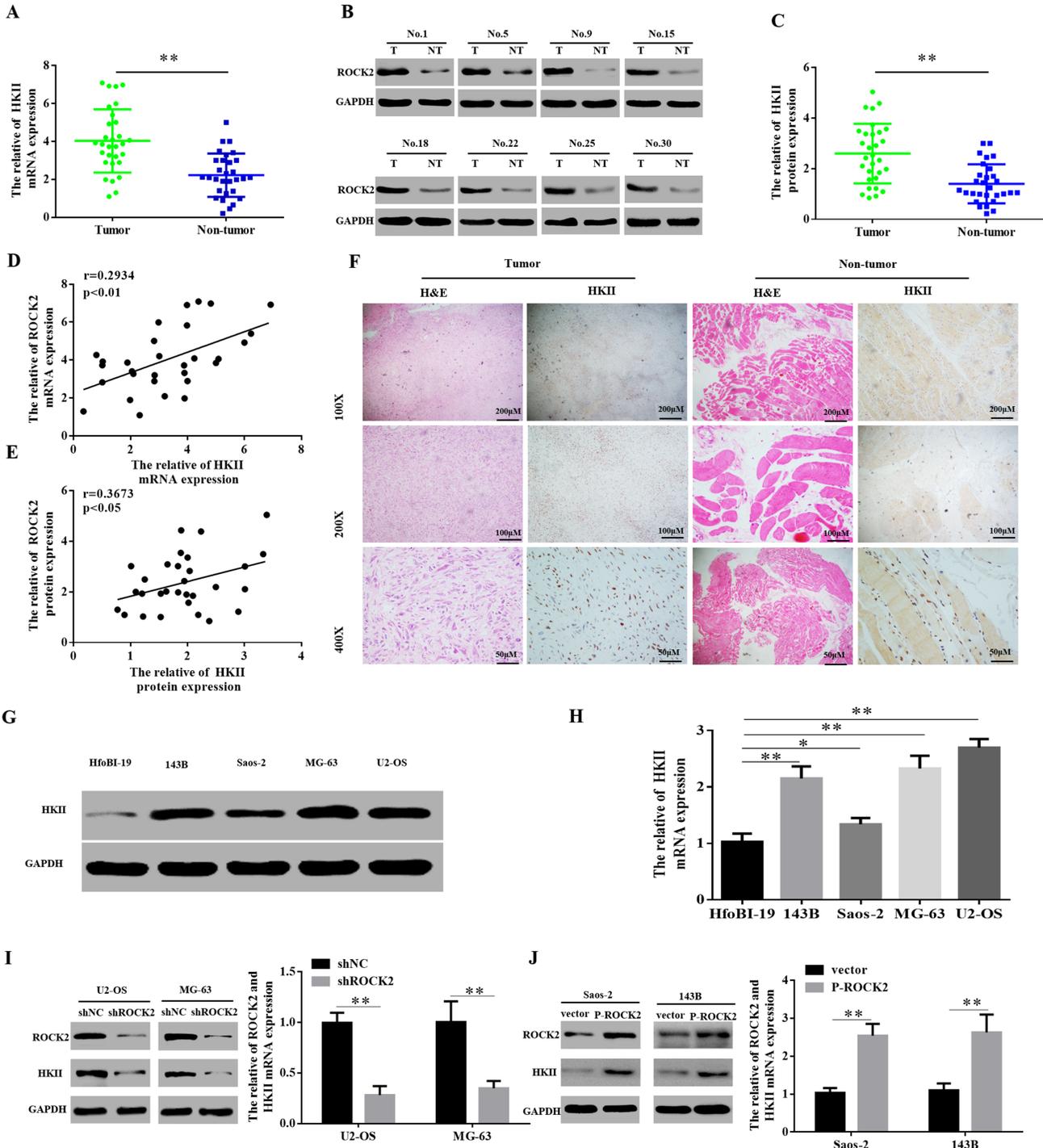


Figure 5

ROCK2 positively regulates HKII protein levels. (A-C) RT-qPCR and western blot analysis were performed to detect the mRNA and protein expression of HKII in 30 patients with OS and their corresponding adjacent tissues, respectively. (D and E) Scatter plots of ROCK2 and HKII mRNA and protein expression in OS tissue. (F) Representative images of HKII staining in 43 paired OS tissues (scale bar, 50 μ m). (G) RT-qPCR and (H) western blotting were performed to detect the mRNA and protein expression of HKII,

respectively, in OS cells (U2-OS, Saos-2, 143B and MG-63) and normal hfoBI-19 cells. (I) Western blotting and (J) RT-qPCR analyses of HKII levels in U2-OS cells stably transfected with shNC or shROCK2 plasmids and in Saos-2 cells stably transfected with the vector or p-ROCK2 plasmid. **P<0.01. ROCK2, Rho-associated coiled-coil-containing protein kinase 2; HKII, mitochondrial hexokinase II; RT-qPCR, reverse transcription-quantitative PCR; OS, osteosarcoma; sh, short interfering; NC, negative control; p, phosphorylated.

Figure 6

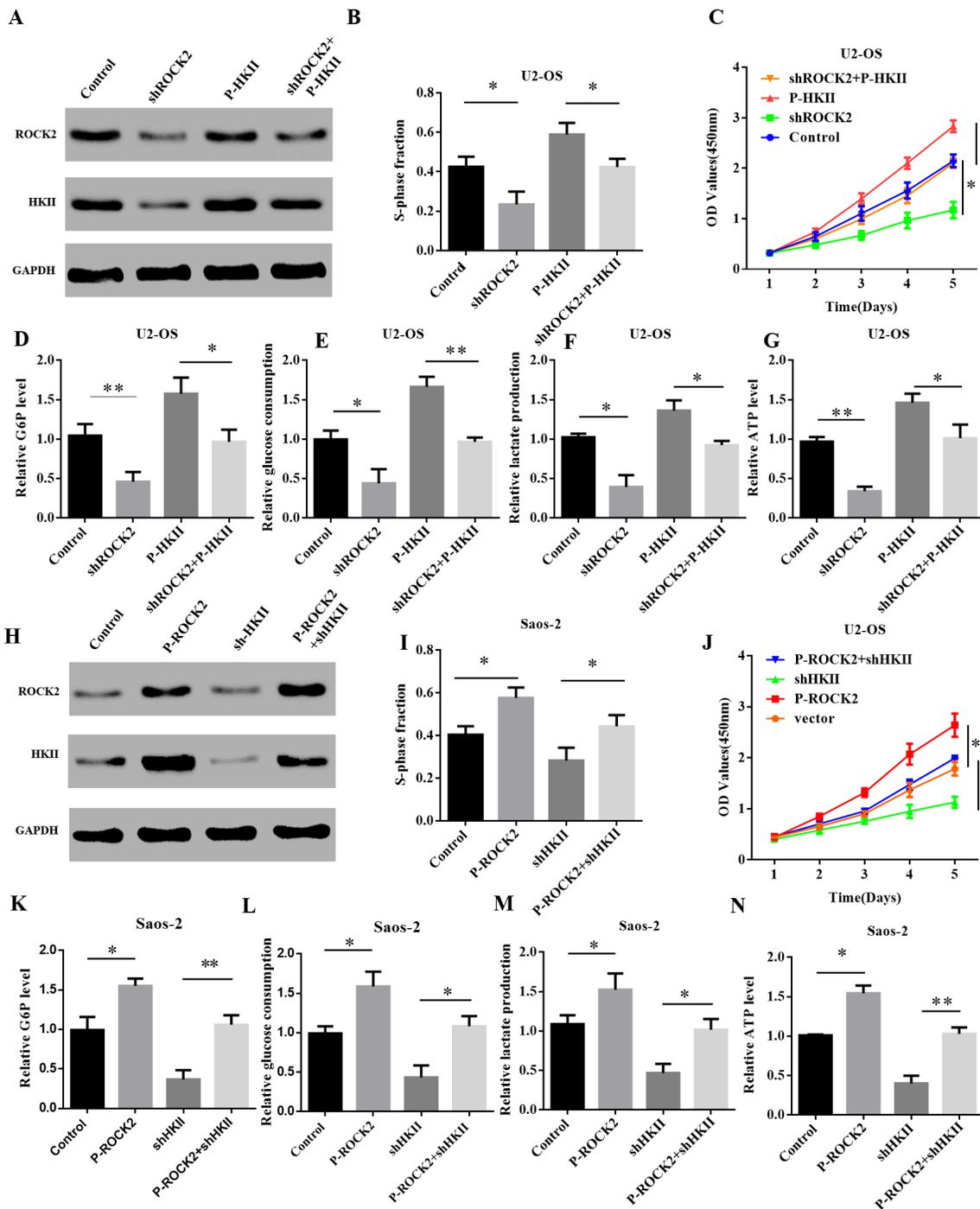


Figure 6

ROCK2 regulates glycolysis and cell proliferation depending on the expression of HKII. (A) Western blotting was performed to detect the expression of ROCK2 and HKII in P-HKII-transfected shROCK2/U2-OS cells. (B) Cell counting kit 8 and (C) EdU results demonstrated the proliferation of U2-OS shROCK2 cells stably transfected with P-HKII. (D) Cellular G6P levels, (E) glucose consumption, (F) lactate production and (G) ATP levels in P-HKII-transfected U2-OS shROCK2 cells. (H) Western blotting was performed to detect ROCK2 and HKII expression in p-ROCK2-transfected Saos-2 cells in the presence or absence of shHKII treatment. (I) CCK8 and (J) EdU results demonstrated the proliferation of Saos-2 cells transfected with p-ROCK2 in the presence or absence of shHKII. (K) Cellular G6P levels, (L) glucose consumption, (M) lactate production and (N) ATP levels were detected in p-ROCK2-transfected Saos-2 cells in the presence or absence of shHKII. * $P < 0.05$ and ** $P < 0.01$. ROCK2, Rho-associated coiled-coil-containing protein kinase 2; HKII, mitochondrial hexokinase II; sh, short interfering; OS, osteosarcoma; p, phosphorylated.

Figure 7

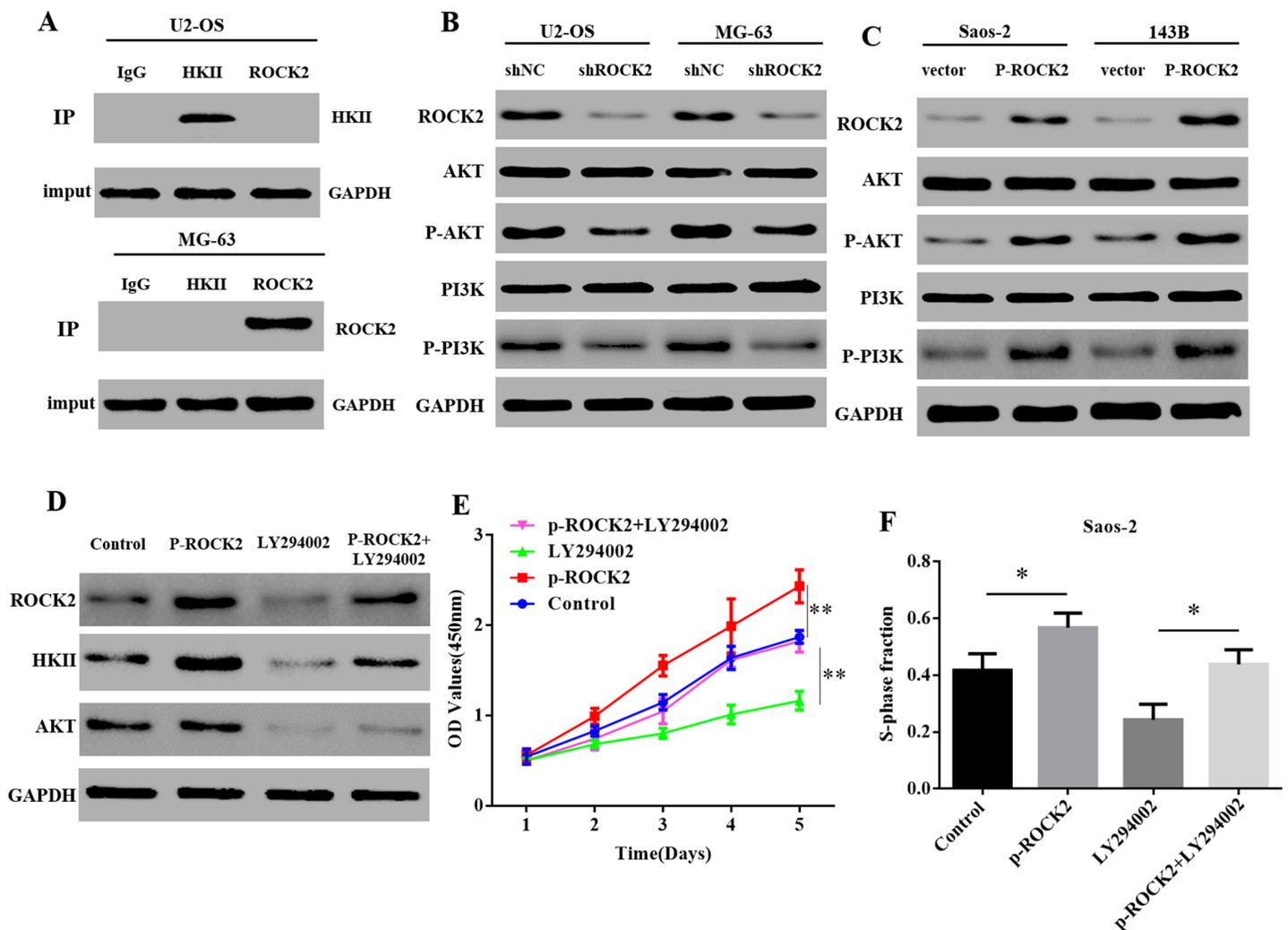


Figure 7

ROCK2 regulates HKII expression by activating the PI3K/AKT signalling pathway via phosphorylation. (A) The results of co-immunoprecipitation indicated that endogenous ROCK2 and HKII were not directly

bound. (B) Western blotting was performed to determine the expression of PI3K/AKT and p-PI3K/p-AKT protein in shROCK2 or shNC cells. (C) Western blotting was additionally performed to determine PI3K/AKT and p-PI3K/p-AKT protein expression in p-ROCK2 or vector cells. (D) Western blotting was applied to analyse HKII expression in p-ROCK2 cells after the inhibition of PI3K/AKT signalling. (E) CCK8 and (F) EdU results demonstrated the proliferation of p-ROCK2 cells after PI3K/AKT signalling inhibition (scale bars:100µM). *P<0.05 and **P<0.01. ROCK2, Rho-associated coiled-coil-containing protein kinase 2; HKII, mitochondrial hexokinase II; p, phosphorylated; sh, short interfering; NC, negative control.

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