

**Analysis of the heterogeneity of the BCR H-CDR3 repertoire in the bone marrow and spleen of
3-,12-,and20-month old mice**

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SUMMARY

The number of central and peripheral B cells and their responsiveness are decreased in aged mice. The diversity of mouse central and peripheral B cell repertoires with increasing age has not been elucidated. In this study, we demonstrated that there were significant differences in the usage of some *V*, *D*, and *J* genes in the BCR H-CDR3 repertoire of bone marrow B cells, spleen B cells and spleen memory B cells in 3-, 12-, and 20-month-old mice. In the productive, pseudogene, and out-of-frame sequences, bone marrow B cells had significant differences in 5'J trimming with age; peripheral spleen B cells and memory B cells had significant differences in N1 insertion, N2 insertion, P5'D insertion, and 5'D trimming with age. The BCR H-CDR3 repertoire diversity of mouse bone marrow B cells, spleen B cells and spleen memory B cells decreased with increasing age. The proportion of overlap in bone marrow and spleen B cells, but not spleen memory B cells, of mice at different ages was lower at 3 months than at 12 and 20 months. This study is the first to report the homogeneity and heterogeneity of the CDR3 repertoire of central and peripheral B cells change as mice age, to further investigation of the decline and response of B cell immunity in young/middle/old-aged mice.

INTRODUCTION

The percentage of B cells and antibody category conversion and reorganization decrease with age[1, 2].

The response B cells produced by aging animals under the same intensity of antigen stimulation are 1/10-1/50 that produced by adult animals[3]. B cell clone proliferation changes with age, and the genetic lineage of B cells also undergoes corresponding dynamic changes. In particular, the increase in memory B cell clones is closely related to the immune system status of elderly individuals[4-6].

The diversity of the naive BCR H-CDR3 repertoire is derived from the rearrangement of the germline *V(D)J* gene in the bone marrow[7] (high-frequency mutations in the periphery that increase the diversity of the B cell repertoire[8]). A shift of the reading frame during the rearrangement process can result in an out-of-frame sequence, and rearrangement of the *V(D)J* pseudogene will not produce a functional sequence[9], although an out-of-frame and pseudogene rearrangement failure on one chromosome may be the start *V(D)J* on another chromosome to continue rearrangement. With the application of HTS to the CDR3 repertoire of T/B cells, the dynamic changes in the body's immune system can be explored by comparing the effective rearrangement of functional genes (in frame) and the rearrangement of the out-of-frame and pseudogene sequences.

Human studies have shown that the lifespan of B cells in elderly individuals is increased, and the production in the bone marrow is reduced[10], which has a certain influence on the number of B cells and changes in the repertoire[11]. B cell expansion clones increase with age[12, 13]. The repertoire is closely related to changes in age before and after immunization in mice[14]. At present, how the homogeneity and heterogeneity of the CDR3 repertoire of central and peripheral B cells change as mice age is not fully elucidated.

In elderly individuals, immunoglobulins IgM and IgD are reduced, and naive B cells are transformed into memory B cells[12]. Plasma cells produce reduced IgG antibodies, which limit memory B cell diversity[15, 16]. As the mouse age increases, how the corresponding changes in the peripheral memory B cell repertoire has not been elucidated.

In this study, we selected 3-, 12-, and 20-month-old mice and used HTS to detect the bone marrow B cell, spleen B cell, and spleen memory B cell BCR H-CDR3 repertoire. The homogeneity and heterogeneity of the productive, pseudogene, and out-of-frame sequences in the BCR H-CDR3 repertoire were compared and analyzed. The main components and characteristics of the central and peripheral BCR H-CDR3 repertoire of mice at different ages were further studied.

RESULTS

Preparation of mouse spleen samples and tissue HE staining

The spleen tissue of mice aged 3, 12, and 20 months was taken (Fig. 1a): the spleen length was approximately 1.5-2 cm, and the color was bright red. The spleen white pulp structure became irregular with increasing age (Fig. 1b). The purity of spleen tissue memory B cells (CD45R⁺CD27⁺) by Miltenyi bead sorting was more than 85% (Fig. 1c).

Mouse bone marrow B cell, spleen B cell and memory B cell CDR3 repertoire

The concentration and purity of multiplex PCR products were in accordance with HTS requirements (Fig. S1). The CDR3 repertoire from the HTS of each tissue sample of 3-, 12-, and 20-month-old mice was compared and statistically analyzed according to the productive, pseudogene, and out-of-frame unique and total sequences (Tab. 1).

Gene frequency in the BCR H-CDR3 repertoire

V gene usage (Fig. 2-1, Fig. S2-1 and Fig. S2-2): (1) Productive sequences: 3-, 12-, and 20-month-old mice were given high-frequency *IGHV1-4*, *IGHV14-1*, *IGHV1-50*, and *IGHV1-64*. With the change in age, Bone marrow B cell *IGHV1-54* and *IGHV1-84*; spleen B cell *IGHV4-1*, *IGHV1-87*, *IGHV5-12*, and *IGHV9-1*; and spleen memory B cell *IGHV1-4*, *IGHV5-12-1*, and *IGHV5-9* showed significant differences at different months ($P < 0.05$). (2) Pseudogene sequences: 3-, 12-, and 20-month-old mice were given high-frequency *IGHV1-67* and *IGHV1-83*. Bone marrow B cell *IGHV1-25* and *IGHV1-83* and spleen B cell *IGHV1-32* and *IGHV1-67* showed significant differences with changes in age ($P < 0.05$). (3) Out-of-frame sequences: 3-, 12-, and 20-month-old mice were given high-frequency *IGHV1-4*, *IGHV14-1*, *IGHV1-50*, and *IGHV1-64*. Bone marrow B cell *IGHV11-1* and *IGHV14-1*; spleen B cell *IGHV11-1*, *IGHV1-4*, *IGHV1-87*, *IGHV1S-45*, and *IGHV9-1*; and spleen memory B cell *IGHV1-14*, *IGHV1-4*, *IGHV1-42*, *IGHV1-50*, *IGHV1-87*, and *IGHV1S-45* showed significant differences with changes in age ($P < 0.05$).

D gene usage (Fig. 2-2): (1) Productive, pseudogene, and out-of-frame sequence high-frequency usage genes were concentrated in *IGHD1-1*, *IGHD2-1*, *IGHD2-3*, and *IGHD2-4*. (2) Productive sequences: Spleen B cell *IGHD2-5*, *IGHD3-3*, and *IGHD6-3* and spleen memory B cell *IGHD2-3*, *IGHD3-1*, *IGHD3-2*, and *IGHD3-3* showed significant differences with age ($P < 0.05$). (3) Pseudogene sequences: Bone marrow B cell *IGHD2-3*, and *IGHD2-4* and spleen B cell *IGHD3-1*, *IGHD3-3*, and *IGHD5-1* showed significant differences with changes in age ($P < 0.05$). (4) Out-of-frame sequences: The frequency of spleen B cell and memory B cell *IGHD2-4* in 20-month-old mice was greater than that in 12- and 3-month old mice; bone marrow B cell *IGHD2-2*, *IGHD2-4*, *IGHD3-2*, and *IGHD5-1*, spleen B cell *IGHD2-5*, and spleen memory B cell *IGHD1-3*, *IGHD2-1*, *IGHD2-12*, *IGHD2-5*, *IGHD3-1*, *IGHD3-2*, *IGHD4-1*, and *IGHD6-2* showed significant differences with changes in age ($P < 0.05$).

J gene usage (Fig. 2-3): Productive sequences: The spleen B cell *IGHJ2* gene showed significant differences with changes in age ($P < 0.05$). Pseudogene sequences: The spleen B cell *IGHJ1* gene showed significant differences with changes in age ($P < 0.05$).

V-J pairing in the BCR H-CDR3 repertoire (Fig. 3, Fig. S3-1 and Fig. S3-2): In the productive, pseudogene, and out-of-frame sequences, the V-J advantage pairing usage was performed differently. Cluster analysis showed that the bone marrow B cells, spleen B cells, and spleen memory B cells in the productive sequences had closer clustering distances at 12 months and 20 months. The pseudogene sequences had closer clustering of the bone marrow and spleen B cells at 3 months and 12 months, while spleen memory B cells were closer clustered at 12 months and 20 months. Out-of-frame sequences had closer clustering of bone marrow B cells and spleen memory B cells at 3 months and 12 months of age, while spleen B cells were closer clustered at 12 months and 20 months.

Insertions and deletions of nucleotides in the BCR H-CDR3 repertoire

The CDR3 diversity results from the “N” nucleotides at the V→D(N1) and D→J(N2) junctions, exonuclease trimming (3'V trimmed, 5'D trimmed and 5'J trimmed) and the addition of palindromic “P” nucleotides (P3'V, P5'D and P5'J)[17]. According to the classification method of nucleotide insertion and deletion reported by Murugan et al.[18], it was found that in the productive, pseudogene, and out-of-frame sequences, bone marrow B cells had significant differences in 5'J trimming with age ($P < 0.05$); spleen B cell and memory B cell N1 insertion, N2 insertion, P5'D insertion, and 5'D trimming were significantly different ($P < 0.05$) (Fig. 4).

Productive sequence clone proliferation in the BCR H-CDR3 repertoire

An analysis of productive sequence clone proliferation was conducted using the inverse of Simpson's diversity index ($1/DS$), calculated as $1/DS = 1 / \sum \{n_i * (n_i - 1)\} / \{n * (n - 1)\}$, where n_i refers to the total

number of the i th sequence [19, 20]. The higher the $1/DS$ value, the richer the diversity and the lower the clonal proliferation [21]. The frequencies of the unique sequences are ordered from high to low, and a \log_{10} scale is used on the Y-axis to reveal the frequency (Fig. S4).

The BCR H-CDR3 repertoire of mouse bone marrow B cells, spleen B cells and spleen memory B cells decreased with the increase in the $1/DS$ index (Fig. 5a). In the BCR H-CDR3 repertoire of mouse spleen B cells, the $1/DS$ index at 12 months and 20 months was significantly lower than that at 3 months ($P < 0.05$) (Fig. 5b). The spleen memory B cell BCR H-CDR3 repertoire $1/DS$ index of 3-month-old mice was greater than that of 12- and 20-month-old mice (Fig. 5c).

CDR3 length and amino acid usage in productive sequences

The distribution of CDR3 lengths at different ages and in different tissues was similar and showed a Gaussian distribution (Fig. 6a,b,c). There was a statistically significant difference in the use of asparagine (N) and isoleucine (I) in the spleen B cell repertoire with increasing age ($P < 0.05$). There was a statistically significant difference in the use of asparagine (N) in the spleen memory B cell repertoire with increasing age ($P < 0.05$) (Fig. 6d,e,f).

Overlap in the productive sequences of the CDR3 repertoire

The ratio of overlap to the number of unique amino acids in the mice at different ages was calculated. The proportion of overlap in bone marrow and spleen B cells in 3-month old mice was lower than that in 12- and 20-month-old mice. This observation was not seen in spleen memory B cells (Tab. 2-1, 2-2, 2-3).

DISCUSSION

As the age of mice increases, the number and response capacity of central and peripheral B cells will change accordingly. The homogeneity and heterogeneity of the composition and characteristics of the

central and peripheral B cells in younger/middle/old-aged mice is an important basis for the study of the responsive ability and mechanism of the aging immune system.

In this experiment, 3-, 12-, and 20-month-old mice were used as subjects, and HTS was used to compare and analyze the homogeneity and heterogeneity of the productive, pseudogene, and out-of-frame sequences of the BCR H-CDR3 repertoire in bone marrow (central) and spleen (peripheral).

The study found that the white pulp of the spleen was more regular at 3 months of age, the density of lymphocytes was larger, and the shape of the shifting area is obvious. With the increase in the age of the mice, the structure of the white pulp became irregular at the ages of 12 months and 20 months. Some of them are relatively loose, with only a small number of lymphocytes, and the shape of the area became irregular. The white pulp structure is typically associated with antigenic stimulation; as the mouse age increased, the mice received more antigenic stimulation, resulting in an irregular white pulp structure in aged mice, which is consistent with previous literature reports[22]. The diversity of the BCR H-CDR3 repertoire is derived from the *V(D)J* gene rearrangement, insertion, deletion, and high-frequency mutations in somatic cells. As mice age, changes in BCR H-CDR3 repertoire diversity can be demonstrated by random combinations of light and heavy chains[23]. The frequency of the *V*, *D*, and *J* genes was related to the advantages of naive rearrangement, B cell self-tolerance selection, B cell clonal proliferation, immune response, etc. In theory, the frequency of the usage of the *V*, *D*, and *J* genes reflects the key features of the CDR3 recognition of specific antigens. This study found a high frequency of the usage of *IGHV1-4*, *IGHV14-1*, *IGHV1-50*, and *IGHV1-64* in the productive and out-of-frame sequences of bone marrow B cells, spleen B cells and spleen memory B cells at different ages. In the pseudogene sequences, the frequency of *IGHV1-67* and *IGHV1-83* was significantly higher

than that of other genes; *IGHV1* had high-frequency usage at different ages and in different tissues, showing homogeneity, which is consistent with other researchers' reports[8]. This finding suggests that in the naive rearrangement, there is a significant advantage of some *V* gene in the naive rearrangement, resulting in its high frequency in the self-tolerance selection and peripheral response BCR repertoire.

In the productive sequences, mouse bone marrow B cell *IGHV1-54* and *IGHV1-84*; spleen B cell *IGHV4-1*, *IGHV1-87*, *IGHV5-12*, and *IGHV9-1*; and spleen memory B cell *IGHV1-4*, *IGHV5-12-1*, and *IGHV5-9* usage significantly changed with age. Significant changes in gene usage may be related to B cell tolerance, clonal proliferation, and the frequency of the immune response and further indicate that the BCR H-CDR3 repertoire is partially heterogeneous at different ages and in different tissues.

A pseudogene sequence is produced by the deletion of the initiation codon and the premature introduction of a termination codon[9]. A pseudogene sequence also participates in rearrangement. Because of the inefficiency of rearrangement, the pseudogene sequence better reflects the frequency characteristics of the *V*, *D* and *J* genes in the naive rearrangement, and, together with the out-of-frame rearrangement of functional genes, it can be used to compare and analyze the composition and specificity of the productive rearrangement of functional genes[24]. The pseudogene sequence showed significant differences in bone marrow B cell *IGHV1-25*, *IGHV1-83*, *IGHD2-3* and *IGHD2-4* with age and significant differences in spleen B cell *IGHV1-32*, *IGHV1-67*, *IGHD3-1*, *IGHD3-3*, and *IGHD5-1* genes. Compared with the productive sequences, there were differences in *V* and *D* gene usage, suggesting that the pseudogene sequence better reflects naive rearrangement.

An out-of-frame sequence is the result of non-productive rearrangement. When we analyzed the *V* genes usage, we found that the out-of-frame sequence and the Productive sequence have many similarities, mainly because the out-of-frame sequence is a translational reading frame change

caused by insertion and deletion of nucleotides during gene rearrangement and cannot be translated into functionality AA. Therefore, we believe that the difference in the out-of-frame sequence in the bone marrow tissue with age changes is mainly affected by gene rearrangement, or it may be the cause of individual differences. Therefore, we believe that the difference in the age of the out-of-frame sequence in the central and peripheral tissues is mainly affected by gene rearrangement, or it may be affected by the antigen response in the periphery.

The frequency of the acquisition of the *V*, *D*, and *J* genes in mice was almost the same, and there were some differences with changes in mouse age, which is consistent with the study of the *V*, *D*, and *J* genes in the peripheral blood of newborns and adults[25].

The closer the cluster analysis is, the more similar the gene access is. The cluster analysis of *V*-*J* pairing in spleen B cells and memory B cells shows that the clustering distances of 12 months and 20 months are the closest, while the cluster distance of 3 months is more. Indicating that the 12-month-old and 20-month-old genes are more similar, suggesting that it may be similar to the longer-term in vitro antigen stimulation, and B cells produce corresponding responses and clonal proliferation. The clustering distance of pseudogene sequences was closer at 3 months and 12 months than at 20 months in the bone marrow and spleen B cells. In the out-of-frame sequences in bone marrow B cells and spleen memory B cells at the ages of 3 months and 12 months, the clustering distance was closer than that at 20 months. It is suggested that more *V* and *J* gene naive rearrangements have higher advantages and matching. The specific sources and mechanisms of various *V*-*J* pairing B cell populations need to be further explored.

For the insertions and deletions of the BCR H-CDR repertoire, in the productive, pseudogene, and out-of-frame sequences, the bone marrow B cells had significant differences in 5'J trimming with age

and may be mainly pro-B cells and pre-B cells. In the pro-B cell repertoire, the rearrangement is generally random, but there is a certain difference in the involvement of the *J* gene in functional and nonfunctional rearrangements. There were significant differences in spleen B cell and memory B cell N1 insertion, N2 insertion, P5'D insertion, and 5'D trimming, suggesting that the BCR H-CDR3 repertoire and B cell self-tolerance selection, immune response, and clonal proliferation are related. Due to the response of multiple autoantibodies and external antigens, the CDR3 repertoire participates in this process in addition to the naive rearrangement, high-frequency mutations of somatic cells and the secondary rearrangement of the BCR, resulting in N1 insertion, N2 insertion, P5'D insertion, and 5'D trimming differences.

The diversity of the BCR H-CDR3 repertoire in different ages of mice showed that the diversity in bone marrow B cells and spleen memory B cells of 3-month-old mice was higher than that of 12- and 20-month old mice. In spleen B cells, the diversity of the 3-month-old mouse repertoire was significantly higher than that of the 12- and 20-month old mouse repertoire. The diversity of the BCR H-CDR3 repertoire is closely related to gene rearrangement and the immune response generated by external antigen stimulation. The diversity of the repertoire in the bone marrow is mainly due to the rearrangement/insertion/deletion of the *V*, *D*, and *J* genes[26]. As the mouse age increases, the B cell output from the bone marrow to the periphery decreases, and the diversity decreases[5]. The findings may have occurred in the bone marrow, and this reduction may also reflect changes in the subset[20, 23].

The response of spleen B cells to external antigen stimulation, as the body ages to increase the body's depletion of B cells, also leads to a decrease in B cell diversity, and B cell clonal proliferation changes with increasing age. Not only are the unique CDR3 sequence species reduced, but the clonal

proliferation that occurs is also greater. This phenomenon also indicates that the clonal distribution caused by the steady-state proliferation and peripheral selection in the aging process is more unbalanced. Related studies indicate that the imbalance of clonal distribution can reflect the response ability and the peripheral selection of self-identification[27]. The changes in the diversity of the central and peripheral BCR H-CDR3 repertoire in mice at 3, 12, and 20 months of age suggest that they are due to both central and peripheral causes, providing further insight into immune system aging and results from diversity studies, basic data and new research techniques.

The AA composition of the BCR H-CDR3 region was found to be high-frequency tyrosine in the B cells of different ages and tissue-derived B cells, consistent with the literature[28] . The use of isoleucine (I) in the spleen B cell repertoire was significantly higher at 12 months than at 3 months and 20 months, and the use of asparagine (N) was significantly lower at 20 months than at 12 months. In the spleen memory B cell repertoire, the frequency of asparagine (N) was significantly lower at 3 months and 20 months than at 12 months. It is suggested that the difference in AA in the peripheral BCR repertoire of mice at different ages may be related to the immune response generated by specific antigen stimulation.

The length distribution analysis of CDR3 revealed that the length of the bone marrow B cells, spleen B cells and spleen memory B cells of mice at different ages showed a Gaussian distribution with 11-12 AA residues. This is consistent with the result previously reported for mouse CDR3, which was an average length of 11.5 ± 1.9 AA residues[17] . Compared with 3 months of age, the CDR3 length distribution in the BCR H-CDR3 repertoire of bone marrow B cells, spleen B cells, and spleen memory B cells shifted to the left at 12 months and shifted to the right at 20 months, suggesting that with the

increase in age, recognition antigen-producing responses, mutations, and clonal proliferation have a tendency to become longer and may be related to the antigens that are exposed[20].

The proportion of overlap in the bone marrow B cells and spleen B cells of mice at different ages was lower at 3 months and higher at 12 months and 20 months. The overlap of unique amino acids at different ages reflects not only clonal proliferation but also the presence of new, effective, unique sequences. Our results showed that the overlap rate of bone marrow and spleen B cells at 3 months was lower, and the overlap rate at 12 and 20 months was higher, which suggests that clonal proliferation increases with age. However, the spleen memory B cells have no obvious regularity, which is mainly affected by the stimulation of the external antigen response.

CONCLUSION

The degradation of the immune system related to aging is a dynamic process that affects the diversity and response capacity of the body's immune repertoire. In this study, HTS was used to investigate the effects of aging on the characteristics of the BCR H-CDR3 repertoire in mouse bone marrow B cells, spleen B cells and spleen memory B cells. We found that the productive, pseudogene, and out-of-frame sequences of bone marrow B cells, spleen B cells and spleen memory B cells in 3-, 12-, and 20-month-old mice have different compositions, and some features show significant heterogeneity, which further provides a basis for investigating the decline and response of B cell immunity in younger/middle-aged/older mice.

METHODS

Study subjects

Three-month-old, 12-month-old, and 20-month-old SPF female BALB/c mice were purchased from Chongqing Tengxin Biotechnology Co., Ltd., and introduced into the Central Laboratory Animal

Center of Zunyi Medical University (SPF feeding conditions). All animals and experiments were performed in accordance with the guidelines of the Animal Care and Use of Laboratory Animals (Ministry of Health, China, 1998) and approved by the Laboratory Animal Ethics Committee of Zunyi Medical University.

Sample preparation

(1) Mouse bone marrow B cell samples (5 mice per month): 3 months old (M3BM): M3BM1, M3BM2, M3BM3, M3BM4, M3BM5; 12 months old (M12BM): M12BM1, M12BM2, M12BM3, M12BM4, M12BM5; and 20 months old (M20BM): M20BM1, M20BM2, M20BM3, M20BM4, M20BM5. (2) Mouse spleen B cell samples (5 mice per month): 3 months old (M3S): M3S1, M3S2, M3S3, M3S4, M3S5; 12 months old (M12S): M12S1, M12S2, M12S3, M12S4, M12S5; and 20 months old (M20S): M20S1, M20S2, M20S3, M20S4, M20S5. (3) Mouse spleen memory B cell samples (3 mice per month): 3 months old (M3SP⁺): M3S7, M3S8, M3S9; 12 months old (M12SP⁺): M12S7, M12S8, M12S9; and 20 months old (M20SP⁺): M20S7, M20S8, M20S9.

Preparation of single-cell suspension

Mice were dissected to remove the spleen and bone marrow for tissue sectioning. A single-cell suspension of memory B cells (CD45R⁺CD27⁺) was prepared with Miltenyi bead sorting, and the purity of the sorted cells was identified. The medullary cavity was washed of the bone marrow cells with PBS to prepare a single-cell suspension, and then, the genomic DNA in the bone marrow and in the spleen single-cell suspension of the memory B cell sample was extracted.

BCR H-CDR3 sequencing

The design and synthesis of 16 upstream primers and 4 downstream primers of the mouse, as well as 2 GAPDH primers, were conducted by Shanghai Invitrogen Biotechnology Co., Ltd. (Tab. S1-1; S1-2;

S1-3). The DNA was used as a template to PCR-amplify the BCR H-CDR3 region, with 100-200 bp PCR product agarose gel recovery (Fig. S1), and the HTS of the CDR3 repertoire was completed by BGI.

Data analysis

The CDR3 sequence of the HTS was converted into FASTA format and uploaded to the IMGT database. The sequences with no results, unknown or <90% V/J-region identity, AA junctions that did not have a C at the beginning or a W at the end, fewer than 6 nucleotides, blanks and ORFs were filtered out from the data downloaded by IMGT. The selected productive, pseudogene, and out-of-frame sequences were statistically analyzed using Excel, GraphPad Prism, HeIm, SPSS (one-way ANOVA), and Draw Venn Diagram online software. For statistical significance, * indicates $P < 0.05$, ** indicates $P < 0.01$, and *** indicates $P < 0.001$.

Supplementary information

Supplemental Information can be found online at immunity.

Abbreviations

1/DS: The inverse Simpson's diversity index; AA: Amino Acid; BCR: B Cell Receptor; SPF: Specific Pathogen Free; H-CDR3: Heavy chain Complementarity Determining Region 3; HTS: High-Throughput Sequencing; IGHV: Immunoglobulin Heavy chain Variable gene; IGHD: Immunoglobulin Heavy chain Diversity gene; IGHJ: Immunoglobulin Heavy chain Joining gene; ORF: Open Reading Frame; C: Cysteine; W: Tryptophan.

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Authors contributions

Xinsheng Yao designed the research. Lina Ma did the experiment, analyzed the data and wrote the paper. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

All animals and experiments were performed in accordance with the guidelines of the Animal Care and Use of Laboratory Animals (Ministry of Health, China, 1998) and approved by the Laboratory Animal Ethics Committee of Zunyi Medical University.

Consent for publication

Written consent for publication was obtained from Zunyi Medical University.

Competing interests

The authors declare that they have no competing interests.

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