

Effect of Hormones (IBA & IAA) on the Propagation of Himalayan Yew in Pakistan: A Conservation Approach

Javaid Iqbal (✉ javaidenv@uop.edu.pk)

University of Peshawar <https://orcid.org/0000-0001-6349-8887>

Bushra Khan

University of Peshawar

Sardar Khan

University of Peshawar

Nasreen Ghaffar

Islamia College Peshawar

Ishaq Ahmad Mian

University of Agriculture Peshawar

Nowsher Yousaf

University of Peshawar

Iftikhar Ahmad

Pakistan Forest Institute

Sadaf Manzoor

Islamia College Peshawar

Research article

Keywords: Hormone, propagation, endangered, *Taxus wallichiana*, survival, conservation

Posted Date: June 22nd, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-27841/v1>

License:  This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Abstract

Background

Himalayan yew (*Taxus wallichiana*) is one of the endangered medicinal plants species having great importance due to the presence of anticancer drug Taxol. This metabolite is mainly used for the treatment of ovarian, breast, AIDS-related cancers, and other indications. The study being reported here was conducted for the propagation of Himalayan yew by using two different trials of Indole Butyric acid (IBA) and Indole acetic acid (IAA) hormones treatments (2000–7000 ppm) through stem cuttings (140 each). In the same way, 3840 cuttings were treated with IBA and IAA from November 2016 to November 2017 at Lalku valley, Swat, Khyber Pakhtunkhwa (KP), Pakistan.

Results

The influence of IBA treatment (7000 ppm) showed a survival of 85.22% (average number of roots = 10.4, average length of roots = 15.5 cm, average number of leaves = 92.4 and average number of sprouts = 3.3) while that of IAA treatment (7000 ppm) the survival of 81.11% (average number of roots = 9.1, average length of roots = 14.6 cm, average number of leaves = 84.0 and average number of sprout = 3.0) were more significant followed by 2000–6000 ppm (IBA and IAA). Lowest survival 40–45% (average number of roots = 4.2, average length of roots = 8.0 cm, average number of leaves = 32.2, average number of sprouts = 1.7) was noticed for controlled cuttings.

Conclusion

The present study enhanced the potential of conservation and propagation of *T. wallichiana*. Hence our study suggests and recommends the application of IBA (7000 ppm) as a better hormone for the conservation and propagation of Himalayan yew.

Background

T. wallichiana is one of the threatened medicinal plants of Himalayan Region (1) commonly known as Himalayan yew and belongs to family Taxaceae (2). Other names given to the species are Banrya in Pushto, Barmi in Hindi and Urdu, Common yew and Himalayan yew in English (3). It is a coniferous, evergreen and a slow-growing tree having needle-like leaves and bright red fruit (Fig. 1) (4) with lengthy seed dormancy period of about 1.5 to 2.0 years and shows growth rate (increase in its circumference) of 0.4 to 1.3 cm per year (5, 6). The species survive for an average life of approximately 600 years (7). It is found in Pakistan, Afghanistan, Bhutan, Indonesia, Nepal, China, Malaysia, Philippines, Vietnam, Myanmar, and India (8, 3). In Pakistan, it is present in moist temperate forests of Murree, Galliat, Kaghan, Kurram, Chitral, Kashmir, Swat and Hazara (9). Unlike other common coniferous species, the population of *Taxus* occurs in patches not continuous. Its habitat is mainly characterized by moist, mixed coniferous

tree forests or cool broad-leaved forests. Due to shade demanding nature, *Taxus* is usually found in association with large tree species such as *Abiespindrow*, *Betula utilizes Pinus wallichiana*, *Acer cesium*, *Rhododendron arboreuand*, *Quercus semecarpifolia* (10).

Taxus is very important to treat cancer and many other diseases like bronchitis, snake bites, epilepsy, asthma, aphrodisiac, internal injuries, scorpion, diabetes, and for the diseases of lungs (10, 11).

In Pakistan, lack of awareness, slow growth rate, agriculture, construction, habitat loss, forest fires, transformation, grazing, over-harvesting, decoration purposes, medicinal use, accidental mortality, lack of management policies, illicit cutting etc. are major threats to the species (10, 12, 13). Approximately 10 genera of *Taxus* are now declining at the Northern Hemisphere in temperate zones (14). During 2001–2005, Technology, Information, Forecasting and Assessment Council (TIFAC) has reported 45 threatened medicinal plants with specific recommendation for 7 plants including *Taxus* enlisted in Convention for International Trade in Endangered Species (CITES) (Appendix II) in 1995 (15, 16, 17, 18, 19, 20, 21). Recently the existence of Yew is receiving high conservation attention due to the high exploitation rate which has reduced its population by 87% (22, 23, 24, 25). Poor regeneration process, slow growth rate and lengthy seed dormancy period of the species (26) significantly contribute to hurdles in its conservation. Vegetative propagation, therefore, could be one of the practical options to enhance its natural regeneration. The *Taxus* species has high regeneration potential by adventitious rooting of fresh stem cuttings (27). Unlike other *Taxus* species, *T. wallichiana* is difficult to root and requires longer time (28). Rooting of *Taxus* stem cuttings is well documented (29, 30). The present experiment was aimed to enhance the potential of conservation and propagation of *T. wallichiana* using the stem cuttings of mature trees with the application of various doses of IBA and IAA.

Results

Test for normality

Results of Kolmogorov-Smirnov and Shapiro-Wilk tests are not significant therefore the data fulfill the normally assumption (Table 2)

Table 1
Experimental Design of Hormones applications.

Trials	No. of Rows	No. of Cuttings in each Row	Total Cuttings	Length of each cutting (inches)	Nos of Replications
1st _IBA	7	20	140	7–8	3
2nd _IAA	7	20	140	7–8	3

Table 2
Tests of Normality

Parameters	Kolmogorov-Smirnov ^a			Shapiro-Wilk		
	Statistic	df	Sig.	Statistic	df	Sig.
IAA Number of Roots	.089	140	.008	.971	140	.004
IAA Root Length(cm)	.109	140	.000	.896	140	.000
IAA Number of Leaves	.150	140	.000	.933	140	.000
IAA Number of Sprouts	.209	140	.000	.898	140	.000
a. Lilliefors Significance Correction						

The Q-Q plots was used to test the assumptions of normality for all the expected and observed values for the four parameters (number of roots, length of root, number of leaves, and number of sprouts). All the values were plotted on the Q-Q plots on a graph which shows that the data is normal (Fig. 4-7).

Multicollinearity

Table 3 shows that Correlation is significant at the 0.01 level (2-tailed). Correlations were done between the growth parameters (number of roots, root length (cm), number of leaves, and number of sprouts). There is strong to moderate correlation was found among number of roots-root length (0.736), number of roots-number of leaves (0.800), number of roots-number of sprouts (0.519). Significant correlation was also found among root length-number of leaves (0.718), root length-number of sprouts (0.421). Similarly, there is significant correlation among number of leaves-number of sprouts (0.541). Hence data contain no multicollinearity.

Table 3
Pearson Correlations for the Parameters i.e. number of roots, length of roots, number of leaves, and number of sprouts of *T. wallichiana* stems cuttings.

Parameters		Number of Roots	Root Length(cm)	Number of Leaves	Number of Sprouts
Number of Roots	Pearson Correlation	1	.736**	.800**	.519**
	Sig. (2-tailed)		.000	.000	.000
	N	140	140	140	140
Root Length (cm)	Pearson Correlation	.736**	1	.718**	.421**
	Sig. (2-tailed)	.000		.000	.000
	N	140	140	140	140
Number of Leaves	Pearson Correlation	.800**	.718**	1	.541**
	Sig. (2-tailed)	.000	.000		.000
	N	140	140	140	140
Number of Sprouts	Pearson Correlation	.519**	.421**	.541**	1
	Sig. (2-tailed)	.000	.000	.000	
	N	140	140	140	140

**. Correlation is significant at the 0.01 level (2-tailed).

Since the data fulfil all the assumption to apply MONOVA technique for further analysis.

Since the p value is < 0.05 (Table 4) which means different treatment at different level are significant to each other. Now to check the performance of different combination, which one performs better than the other, for that post hoc test was applied.

Table 4
Multivariate Analysis of Variance

Multivariate Tests ^a						
Effect		Value	F	Hypothesis df	Error df	Sig.
Intercept	Pillai's Trace	.988	2574.650 ^b	4.000	123.000	.000
	Wilks' Lambda	.012	2574.650 ^b	4.000	123.000	.000
	Hotelling's Trace	83.728	2574.650 ^b	4.000	123.000	.000
	Roy's Largest Root	83.728	2574.650 ^b	4.000	123.000	.000
Harmons	Pillai's Trace	.126	4.448 ^b	4.000	123.000	.002
	Wilks' Lambda	.874	4.448 ^b	4.000	123.000	.002
	Hotelling's Trace	.145	4.448 ^b	4.000	123.000	.002
	Roy's Largest Root	.145	4.448 ^b	4.000	123.000	.002
Concentration	Pillai's Trace	1.082	7.785	24.000	504.000	.000
	Wilks' Lambda	.109	15.905	24.000	430.306	.000
	Hotelling's Trace	6.468	32.742	24.000	486.000	.000
	Roy's Largest Root	6.204	130.275 ^c	6.000	126.000	.000
Harmons * Concentration	Pillai's Trace	.025	.130	24.000	504.000	1.000
	Wilks' Lambda	.976	.128	24.000	430.306	1.000
	Hotelling's Trace	.025	.126	24.000	486.000	1.000
	Roy's Largest Root	.017	.353 ^c	6.000	126.000	.907

a. Design: Intercept + Harmons + Concentration + Harmons * Concentration

b. Exact statistic

c. The statistic is an upper bound on F that yields a lower bound on the significance level.

For concentration levels (2000 PPM-7000 PPM), Post Hoc test was applied. As by having the same numbers of replications/concentrations of hormones/number of plant cuttings Tukey Test (HSD)

(Table 5) and correlation analysis (Table 4) have been done.

Table 5

Post Hoc Tests (Tukey HSD) applied showed the concentration 7000 ppm is more significant for similar number of cases in each treatment

Multiple Comparisons							
Tukey HSD							
Dependent Variable	(I) Quantity (PPM)	(J) Quantity (PPM)	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
						Lower Bound	Upper Bound
Number of Roots	Control	2000	-1.7000*	.44042	.003	-3.0197	-.3803
		3000	-2.3000*	.44042	.000	-3.6197	-.9803
		4000	-3.0500*	.44042	.000	-4.3697	-1.7303
		5000	-3.7000*	.44042	.000	-5.0197	-2.3803
		6000	-4.8000*	.44042	.000	-6.1197	-3.4803
		7000	-6.5000*	.44042	.000	-7.8197	-5.1803
2000	Control	2000	1.7000*	.44042	.003	.3803	3.0197
		3000	-.6000	.44042	.821	-1.9197	.7197
		4000	-1.3500*	.44042	.041	-2.6697	-.0303
		5000	-2.0000*	.44042	.000	-3.3197	-.6803
		6000	-3.1000*	.44042	.000	-4.4197	-1.7803
		7000	-4.8000*	.44042	.000	-6.1197	-3.4803
3000	Control	2000	2.3000*	.44042	.000	.9803	3.6197
		2000	.6000	.44042	.821	-.7197	1.9197
		4000	-.7500	.44042	.615	-2.0697	.5697
		5000	-1.4000*	.44042	.030	-2.7197	-.0803
		6000	-2.5000*	.44042	.000	-3.8197	-1.1803
		7000	-4.2000*	.44042	.000	-5.5197	-2.8803
4000	Control	2000	3.0500*	.44042	.000	1.7303	4.3697
		2000	1.3500*	.44042	.041	.0303	2.6697

Multiple Comparisons						
		3000	.7500	.44042	.615	-.5697
		5000	-.6500	.44042	.759	-1.9697
		6000	-1.7500*	.44042	.002	-3.0697
		7000	-3.4500*	.44042	.000	-4.7697
	5000	Control	3.7000*	.44042	.000	2.3803
		2000	2.0000*	.44042	.000	.6803
		3000	1.4000*	.44042	.030	.0803
		4000	.6500	.44042	.759	-.6697
		6000	-1.1000	.44042	.169	-2.4197
		7000	-2.8000*	.44042	.000	-4.1197
	6000	Control	4.8000*	.44042	.000	3.4803
		2000	3.1000*	.44042	.000	1.7803
		3000	2.5000*	.44042	.000	1.1803
		4000	1.7500*	.44042	.002	.4303
		5000	1.1000	.44042	.169	-.2197
		7000	-1.7000*	.44042	.003	-3.0197
	7000	Control	6.5000*	.44042	.000	5.1803
		2000	4.8000*	.44042	.000	3.4803
		3000	4.2000*	.44042	.000	2.8803
		4000	3.4500*	.44042	.000	2.1303
		5000	2.8000*	.44042	.000	1.4803
		6000	1.7000*	.44042	.003	.3803
Root Length (cm)	Control	2000	-2.0550*	.56146	.007	-3.7374
		3000	-2.4700*	.56146	.000	-4.1524
		4000	-2.7450*	.56146	.000	-4.4274

Multiple Comparisons

	5000	-2.9850*	.56146	.000	-4.6674	-1.3026
	6000	-4.1550*	.56146	.000	-5.8374	-2.4726
	7000	-4.5300*	.56146	.000	-6.2124	-2.8476
2000	Control	2.0550*	.56146	.007	.3726	3.7374
	3000	-.4150	.56146	.990	-2.0974	1.2674
	4000	-.6900	.56146	.882	-2.3724	.9924
	5000	-.9300	.56146	.646	-2.6124	.7524
	6000	-2.1000*	.56146	.005	-3.7824	-.4176
	7000	-2.4750*	.56146	.000	-4.1574	-.7926
3000	Control	2.4700*	.56146	.000	.7876	4.1524
	2000	.4150	.56146	.990	-1.2674	2.0974
	4000	-.2750	.56146	.999	-1.9574	1.4074
	5000	-.5150	.56146	.969	-2.1974	1.1674
	6000	-1.6850*	.56146	.049	-3.3674	-.0026
	7000	-2.0600*	.56146	.006	-3.7424	-.3776
4000	Control	2.7450*	.56146	.000	1.0626	4.4274
	2000	.6900	.56146	.882	-.9924	2.3724
	3000	.2750	.56146	.999	-1.4074	1.9574
	5000	-.2400	.56146	1.000	-1.9224	1.4424
	6000	-1.4100	.56146	.164	-3.0924	.2724
	7000	-1.7850*	.56146	.030	-3.4674	-.1026
5000	Control	2.9850*	.56146	.000	1.3026	4.6674
	2000	.9300	.56146	.646	-.7524	2.6124
	3000	.5150	.56146	.969	-1.1674	2.1974
	4000	.2400	.56146	1.000	-1.4424	1.9224
	6000	-1.1700	.56146	.369	-2.8524	.5124

Multiple Comparisons						
		7000	-1.5450	.56146	.094	-3.2274
6000		Control	4.1550*	.56146	.000	2.4726
		2000	2.1000*	.56146	.005	.4176
		3000	1.6850*	.56146	.049	.0026
		4000	1.4100	.56146	.164	-.2724
		5000	1.1700	.56146	.369	-.5124
		7000	-.3750	.56146	.994	-2.0574
7000		Control	4.5300*	.56146	.000	2.8476
		2000	2.4750*	.56146	.000	.7926
		3000	2.0600*	.56146	.006	.3776
		4000	1.7850*	.56146	.030	.1026
		5000	1.5450	.56146	.094	-.1374
		6000	.3750	.56146	.994	-1.3074
Number of Leaves	Control	2000	-30.5500*	2.49479	.000	-38.0257
		3000	-32.3000*	2.49479	.000	-39.7757
		4000	-33.8000*	2.49479	.000	-41.2757
		5000	-39.6000*	2.49479	.000	-47.0757
		6000	-47.8000*	2.49479	.000	-55.2757
		7000	-59.2500*	2.49479	.000	-66.7257
		2000	30.5500*	2.49479	.000	23.0743
2000		Control	30.5500*	2.49479	.000	38.0257
		3000	-1.7500	2.49479	.992	-9.2257
		4000	-3.2500	2.49479	.850	-10.7257
		5000	-9.0500*	2.49479	.007	-16.5257
		6000	-17.2500*	2.49479	.000	-24.7257
		7000	-28.7000*	2.49479	.000	-36.1757

Multiple Comparisons

	3000	Control	32.3000*	2.49479	.000	24.8243	39.7757
	2000		1.7500	2.49479	.992	-5.7257	9.2257
	4000		-1.5000	2.49479	.997	-8.9757	5.9757
	5000		-7.3000	2.49479	.060	-14.7757	.1757
	6000		-15.5000*	2.49479	.000	-22.9757	-8.0243
	7000		-26.9500*	2.49479	.000	-34.4257	-19.4743
	4000	Control	33.8000*	2.49479	.000	26.3243	41.2757
	2000		3.2500	2.49479	.850	-4.2257	10.7257
	3000		1.5000	2.49479	.997	-5.9757	8.9757
	5000		-5.8000	2.49479	.241	-13.2757	1.6757
	6000		-14.0000*	2.49479	.000	-21.4757	-6.5243
	7000		-25.4500*	2.49479	.000	-32.9257	-17.9743
	5000	Control	39.6000*	2.49479	.000	32.1243	47.0757
	2000		9.0500*	2.49479	.007	1.5743	16.5257
	3000		7.3000	2.49479	.060	-.1757	14.7757
	4000		5.8000	2.49479	.241	-1.6757	13.2757
	6000		-8.2000*	2.49479	.022	-15.6757	-.7243
	7000		-19.6500*	2.49479	.000	-27.1257	-12.1743
	6000	Control	47.8000*	2.49479	.000	40.3243	55.2757
	2000		17.2500*	2.49479	.000	9.7743	24.7257
	3000		15.5000*	2.49479	.000	8.0243	22.9757
	4000		14.0000*	2.49479	.000	6.5243	21.4757
	5000		8.2000*	2.49479	.022	.7243	15.6757
	7000		-11.4500*	2.49479	.000	-18.9257	-3.9743
	7000	Control	59.2500*	2.49479	.000	51.7743	66.7257

Multiple Comparisons

		2000	28.7000*	2.49479	.000	21.2243	36.1757
		3000	26.9500*	2.49479	.000	19.4743	34.4257
		4000	25.4500*	2.49479	.000	17.9743	32.9257
		5000	19.6500*	2.49479	.000	12.1743	27.1257
		6000	11.4500*	2.49479	.000	3.9743	18.9257
Number of Sprouts	Control	2000	-.3500	.25820	.824	-1.1237	.4237
		3000	-.5500	.25820	.342	-1.3237	.2237
		4000	-.8500*	.25820	.021	-1.6237	-.0763
		5000	-1.1000*	.25820	.001	-1.8737	-.3263
		6000	-1.4000*	.25820	.000	-2.1737	-.6263
		7000	-1.5500*	.25820	.000	-2.3237	-.7763
		2000	Control	.3500	.25820	.824	-.4237
2000		3000	-.2000	.25820	.987	-.9737	.5737
		4000	-.5000	.25820	.460	-1.2737	.2737
		5000	-.7500	.25820	.064	-1.5237	.0237
		6000	-1.0500*	.25820	.002	-1.8237	-.2763
		7000	-1.2000*	.25820	.000	-1.9737	-.4263
		3000	Control	.5500	.25820	.342	-.2237
		2000	.2000	.25820	.987	-.5737	.9737
3000		4000	-.3000	.25820	.907	-1.0737	.4737
		5000	-.5500	.25820	.342	-1.3237	.2237
		6000	-.8500*	.25820	.021	-1.6237	-.0763
		7000	-1.0000*	.25820	.003	-1.7737	-.2263
		4000	Control	.8500*	.25820	.021	.0763
		2000	.5000	.25820	.460	-.2737	1.2737
		3000	.3000	.25820	.907	-.4737	1.0737

Multiple Comparisons

	5000	-.2500	.25820	.960	-1.0237	.5237
	6000	-.5500	.25820	.342	-1.3237	.2237
	7000	-.7000	.25820	.104	-1.4737	.0737
5000	Control	1.1000*	.25820	.001	.3263	1.8737
	2000	.7500	.25820	.064	-.0237	1.5237
	3000	.5500	.25820	.342	-.2237	1.3237
	4000	.2500	.25820	.960	-.5237	1.0237
	6000	-.3000	.25820	.907	-1.0737	.4737
	7000	-.4500	.25820	.589	-1.2237	.3237
	Control	1.4000*	.25820	.000	.6263	2.1737
6000	2000	1.0500*	.25820	.002	.2763	1.8237
	3000	.8500*	.25820	.021	.0763	1.6237
	4000	.5500	.25820	.342	-.2237	1.3237
	5000	.3000	.25820	.907	-.4737	1.0737
	7000	-.1500	.25820	.997	-.9237	.6237
	Control	1.5500*	.25820	.000	.7763	2.3237
	2000	1.2000*	.25820	.000	.4263	1.9737
7000	3000	1.0000*	.25820	.003	.2263	1.7737
	4000	.7000	.25820	.104	-.0737	1.4737
	5000	.4500	.25820	.589	-.3237	1.2237
	6000	.1500	.25820	.997	-.6237	.9237

Based on observed means.

The error term is Mean Square (Error) = .667.

*. The mean difference is significant at the .05 level.

Since two different hormones IAA and IBA are being used, therefore we interpret the differences by simply looking at their mean values. The results revealed that in both trials for the regeneration of Himalayan yew (*T. wallichiana*) cuttings, by the treatment of hormones (IBA and IAA) with 2000–7000 ppm were

applied. Out of all the applied concentrations of hormone (2000–7000 ppm), 7000 ppm shows maximum marginal means for number of roots (10.4, 9.1), roots length (11.82 cm, 10.27 cm), and number of leaves (92.40, 88.30), sprout numbers (3.30, 3.0) and survival percentage (85.22%, 81.11%) throughout the experiment. The next best dozes were 6000 ppm followed by 5000 ppm, 4000 ppm, 3000 ppm, and 2000 ppm. The lowest growth was noticed in control cuttings (Table 6) (Fig. 8 to 12).

Table 6

General Linear Model (Descriptive Statistics), showing the estimated marginal means of the Parameter (number of roots, roots length (cm), number of leaves, and number of sprouts) of *T. wallichiana* cuttings by the application of various concentration of hormones (2000 PPM- 7000 PPM) of IBA, and IAA.

Parameters	Treatment	Quantity in PPM	Mean ± SD	N
Number of Roots	IAA	Control	2.8000 ± 1.54919	10
		2000PPM	4.8000 ± 1.03280	10
		3000PPM	5.2000 ± 1.39841	10
		4000PPM	5.9000 ± 1.19722	10
		5000PPM	6.6000 ± 1.17379	10
		6000PPM	7.6000 ± 1.07497	10
		7000PPM	9.1000 ± 1.19722	10
		Total	6.0000 ± 2.23931	70
	IBA	Control	3.7000 ± 1.63639	10
		2000PPM	5.1000 ± 1.19722	10
		3000PPM	5.9000 ± 1.19722	10
		4000PPM	6.7000 ± 1.56702	10
		5000PPM	7.3000 ± 1.33749	10
		6000PPM	8.5000 ± 1.26930	10
		7000PPM	10.4000 ± 2.22111	10
		Total	6.8000 ± 2.52868	70
	Total	Control	3.2500 ± 1.61815	20
		2000PPM	4.9500 ± 1.09904	20
		3000PPM	5.5500 ± 1.31689	20
		4000PPM	6.3000 ± 1.41793	20
		5000PPM	6.9500 ± 1.27630	20
		6000PPM	8.0500 ± 1.23438	20
		7000PPM	9.7500 ± 1.86025	20
		Total	6.4000 ± 2.41339	140
Root Length (cm)	IAA	Control	5.8600 ± 3.25958	10

Parameters	Treatment	Quantity in PPM	Mean ± SD	N
Growth Hormone	NAA	2000PPM	8.2000 ± 1.04243	10
		3000PPM	8.6400 ± 1.24383	10
		4000PPM	8.8800 ± 1.40063	10
		5000PPM	9.1300 ± 0.97985	10
		6000PPM	10.1100 ± 1.19019	10
		7000PPM	10.2700 ± 1.51441	10
		Total	8.7271 ± 2.11571	70
	IBA	Control	7.1700 ± 3.01517	10
Total	IBA	2000PPM	8.9400 ± 1.73666	10
		3000PPM	9.3300 ± 1.49596	10
		4000PPM	9.6400 ± 1.55863	10
		5000PPM	9.8700 ± 1.76638	10
		6000PPM	11.2300 ± 1.71856	10
		7000PPM	11.8200 ± 1.18115	10
		Total	9.7143 ± 2.27783	70
	Control	6.5150 ± 3.12903	20	
Number of Leaves	IAA	2000PPM	8.5700 ± 1.44481	20
		3000PPM	8.9850 ± 1.38499	20
		4000PPM	9.2600 ± 1.49399	20
		5000PPM	9.5000 ± 1.44113	20
		6000PPM	10.6700 ± 1.54923	20
		7000PPM	11.0450 ± 1.54255	20
		Total	9.2207 ± 2.24565	140
	Control	30.1000 ± 16.25799	10	
Root Length	IAA	2000PPM	60.8000 ± 5.11642	10
		3000PPM	62.1000 ± 7.48999	10
		4000PPM	64.0000 ± 6.97615	10
		5000PPM	69.7000 ± 4.71522	10

Parameters	Treatment	Quantity in PPM	Mean ± SD	N
Growth Hormone	IBA	6000PPM	77.7000 ± 6.91295	10
		7000PPM	88.3000 ± 5.01221	10
		Total	64.6714 ± 18.69645	70
	IBA	Control	32.1000 ± 11.87387	10
		2000PPM	62.5000 ± 6.32895	10
		3000PPM	64.7000 ± 7.45431	10
		4000PPM	65.8000 ± 7.19259	10
		5000PPM	71.7000 ± 3.83116	10
		6000PPM	80.1000 ± 6.08185	10
		7000PPM	92.4000 ± 6.39792	10
		Total	67.0429 ± 18.75350	70
	Total	Control	31.1000 ± 13.89396	20
		2000PPM	61.6500 ± 5.66870	20
		3000PPM	63.4000 ± 7.39417	20
		4000PPM	64.9000 ± 6.95777	20
		5000PPM	70.7000 ± 4.30544	20
		6000PPM	78.9000 ± 6.45552	20
		7000PPM	90.3500 ± 5.97605	20
		Total	65.8571 ± 18.69543	140
Number of Sprouts	IAA	Control	1.5000 ± 1.08012	10
		2000PPM	1.7000 ± 0.67495	10
		3000PPM	1.9000 ± 0.56765	10
		4000PPM	2.3000 ± 0.94868	10
		5000PPM	2.6000 ± 0.96609	10
		6000PPM	$2.9000 \pm .73786$	10
		7000PPM	$3.0000 \pm .66667$	10
		Total	$2.2714 \pm .96190$	70
	IBA	Control	$1.7000 \pm .94868$	10

Parameters	Treatment	Quantity in PPM	Mean ± SD	N
Hormone Concentration (PPM)	2000PPM	2.2000 ± .63246	10	
	3000PPM	2.4000 ± .51640	10	
	4000PPM	2.6000 ± .96609	10	
	5000PPM	2.8000 ± 1.03280	10	
	6000PPM	3.1000 ± .87560	10	
	7000PPM	3.3000 ± .48305	10	
	Total	2.5857 ± .92459	70	
	Total	1.6000 ± .99472	20	
Hormone Mode of Application	Control	1.6000 ± .99472	20	
	2000PPM	1.9500 ± .68633	20	
	3000PPM	2.1500 ± .58714	20	
	4000PPM	2.4500 ± .94451	20	
	5000PPM	2.7000 ± .97872	20	
	6000PPM	3.0000 ± .79472	20	
	7000PPM	3.1500 ± .58714	20	
	Total	2.4286 ± .95317	140	

The results depict that the cutting responded differently to various levels of concentration of hormones (2000 ppm-7000 ppm) and the modes of application. Table 4.5 shows the multiple comparisons of all the parameters (number of roots, length of roots cm, number of leaves, number of sprouts) marginal mean values by the applied various concentrations of hormones (2000 ppm-7000 ppm) to the cuttings of *T. wallichiana* for the significant. Therefore, Post Hoc test (Tukey HSD) has been applied to check the significance on different levels among the growth parameters (number of roots, length of roots cm, number of leaves, number of sprouts) by applied hormones various concentrations (2000 ppm-7000 ppm). The multiple comparison (Tukey HSD) shows that based on observed means the error term is Mean Square (Error) = 0.667 and (*) the mean difference is significant at the 0.05 level ($p < 0.05$) (Table 5).

Discussion

The current study indicates that hormone treatment (IBA & IAA) is the favourable and best option for the conservation and vegetative propagation of Himalayan yew. As our study depicts and recommends IBA the best rooting hormones in comparison to the Indole acitic acid (IAA). In fact, the auxins application is important to produce roots in fresh stem cuttings as they have been experienced for stimulating

secondary plant growth that results in the reserve food material mobilization to the root initiation site (31). Auxins application increases rooting and quality of roots in various tree species (32). It is a well-documented that the auxins application is important for the roots formation in the cuttings of plant stem as they have been found in stimulating the cambial activity which results in reserve food mobilization to the root initiation site (31; 33).

Also, a successful rooting in juvenile shoot cuttings of *T. wallichiana* with different auxins has been documented from the other temperate areas as well (34, 35, 36, 38, 39)

Our results are also similar with Nautiyal et al. (1991) (40), who treated hormones on stem cutting of teak for roots induction and concluded that IBA is the favourable and best auxins. Other scientists (41, 44, 37, 42, 45, 46, 43) also documented that the IBA application play a vital role in the propagation of stem cuttings of *Taxus*. Their results clearly mentioned that vegetative propagation (in-situ conservation) can improve the number of juvenile's plants of genus *Taxus*.

Several primary root formations under the influence of IBA application have been obtained in the present trials (7000 ppm). It must be accepted that a higher root to shoot is satisfactory for achieving a higher survival rate after transplantation of field. It has also been reported that more than optimums concentration of auxins is toxic to the root regeneration, while the optimum concentration of hormones is favorable (47, 48). In our study we had applied the maximum concentration of 7000 ppm of both the auxins (IBA and IAA) which shows favourable response to the growth of *T. wallichiana* cuttings.

The differential response to changing concentrations of hormones for survivals of *T. wallichiana* were noted in the present study could be like the action of increasing Auxins (2000 ppm – 7000 ppm). The IBA application may have an indirect effect by increasing the translocation speed and the movement of sugar to the cuttings base which in result stimulate rooting (31).

IBA application in the current study significantly increased the survival percentage (number of roots, length of roots, number of leaves, and number of sprouts). Same results were obtained in another study by applying different hormones, for the initiation of a maximum rooting percentage in the juvenile fresh stem cuttings of *T. wallichiana*; IBA is the effective and best hormone (50). It is now decided that IBA is the best rooting auxins for *Taxus* species and it was confirmed by many workers (36, 29). The time duration of our study was kept 20 weeks after the plantation of the cutting of *T. wallichiana*. Similarly, the auxins effect on growth and induction of adventitious roots of *T. wallichiana* cuttings after 4 and ½ months of application/treatment and plant growing. In general, it has been found that among various auxins IBA was the more active for root germination showed the maximum percentage of survival, rooting percentage, percentage of callusing roots per cutting, and length of root per cutting (37, 41, 42, 43, 45, 46). The researcher studied the application of 1000 ppm and 500 ppm of IBA on the *T. wallichiana* cuttings and found, IBA 500 ppm showed slightly less response than IBA 1000 ppm. IBA 1000 ppm showed the best rooting response on shoot cuttings of *T. wallichiana* in the spring season with a rooting percentage (95%) compared to other treatments in other seasons (51). In our study the best response was

given by 7000 ppm in case of both IBA and IAA. By looking into other studies, it clears that by increasing the concentration up to 7000 ppm the growth will be high.

By using IBA, with some other cultivar of yew (49) that showed that treatment of hormone was very important for successful conservation and propagation of *T. wallichiana*. By the treatment of hormones, stem cuttings of some other species were also published (55), their research showed that the best response for rooting due to IBA followed by IAA and NAA in rooting percentage, the cuttings heights, number of roots per cutting in Sage, Rosemary, and Elderberry. Similarly, the treatments of auxins (IBA, IAA, and NAA) on the root development were studied in species like *Melissa officinalis* (52), *Ficus Benjamina* (53), and *Oryza sativa* (54).

Auxins are involved in the formation of root, cambial cells activation, and lateral bud inhibition. They have been found as naturally existing compounds that promotes the formation of root and synthetic auxins also stimulating the emergence of root on cuttings. It is well documented and verified maximum time that auxins applied (naturally or artificially) is needed for adventitious root initiation on stem cuttings. Initial cells division of roots is dependent on either endogenous or applied auxins (56).

Our findings are also in similarity with (57) that showed that root development in stem cuttings of *T. wallichiana* using IBA followed by IAA and NAA and in control where no auxins treatment was given, less root formation was observed in those cuttings, and the lowest survival was found during the observation of cuttings. In this case further the survival becomes very difficult and ultimately the cuttings died. In this respect, our results are similar as obtained by (58).

Thus, our study suggests the utilization of auxins mainly IBA for regeneration of *T. wallichiana* under natural conditions. The results of our research will be beneficial for developing propagation protocol of *T. wallichiana* species especially for the moist temperate climate of the Himalayan Region.

Conclusion

This study was aimed to conserve and propagate the Himalayan yew (*T. wallichiana*) by finding out the best rooting hormone (IBA and IAA) with suitable concentration applied (2000 ppm-7000 ppm) for the regeneration of the fresh stem cuttings. Out of the two different trial, it has been found that 7000 ppm shows higher survival percentage (IBA 85.22% and IAA 81.11%) irrespective of the mode of application and results were highly significant followed by their corresponding. Hormones with concentration 6000 ppm was the next best suitable dose that shows maximum survival (IBA 66.67%, IAA 63.33%) followed by 5000ppm > 4000ppm > 3000ppm > 2000 ppm. Therefore, among all the treatments of auxins (IBA and IAA), IBA shows maximum growth (number of roots, root length (cm), number of leaves, and number of sprouts) and survival (IBA > IAA) (Fig. 8–12). The lowest survival percentages were noticed by control cuttings in both the trials.

Methods

Study Area

Our study was carried out with the aim to enhance the potential of conservation and propagation of the valuable and endangered medicinal plant *T. wallichiana* species in Lalku valley of District Swat, KP, Pakistan. Lalku valley has more Taxus density and offers a rich forest cover of pine and oak trees. It lies at an altitude of 1963 meters above sea level, latitude of 35°.1375 and longitude of 72°.38639. Forest area of Lalku forest range is 8580 ha with a Forest cover of 59.3%. In this region, temperature ranges from -2°C to 34°C. The average annual precipitation ranges from 1000–1200 mm. The study period was during November 2016 to November 2017.

Collection, Preparation, and Planting of Cuttings

The formal identification of the *T. wallichiana*'s plant materials was undertaken by the Directorate of Non-Timber Forest Produce (NTFP), Forest Department, Peshawar; Department of Environmental Sciences, University of Peshawar. Proper permission was granted by the University of Peshawar, Ethical review board. Voucher specimens no. Bot. 20156 (PUP) were deposited in the herbarium of Department of Botany, University of Peshawar.

Cuttings were collected from various mature patches of Himalayan yew. The cuttings were brought to the nursery, raised in Lalku Forest Research Station. The length of the final cuttings were kept 7 to 8 inches and 3 to 4 nodes were retained in each cutting. The needles at the basal portion (about 2 cm) of the stem cuttings were removed and sterilized using 2% benlate (fungicide) before planting. The cuttings were dipped in fungicidal solution for 5 minutes and dried for 20 to 25 minutes in an open environment. The dried cuttings were treated with 50% (Water: Ethanol) concentrated solution of the IBA and IAA (2000–7000 ppm) for 5 minutes and planted in the polythene bags containing soil. The soil was prepared by mixing forest soil, sand and agriculture soil of the area in 1:1:1 and sieved properly before filling into polythene bags (Fig. 2).

Experimental Design

The experiment was carried out in a randomized block design with a factorial treatment's arrangement. First trial treatment was carried out with 7 rows and each row containing 20 cuttings. Total of 140 cuttings were taken in the first trial of the experiment ($n = 140$; $x 20$ cuttings \times 1 type of cuttings \times 7 IBA treatments, 3 replications) (Fig. 3). Same was repeated for the second trial of IAA treatments 7 rows with 20 cuttings each, so a total of 140 cuttings were used ($n = 140$; $x 20$ cuttings \times 1 type of cuttings \times 7 IAA treatments, 3 replications) as shown in (Table 1). The trials were evaluated for the number of roots, length of roots, number of leaves, sprouts and survival percentages of cuttings after 20 weeks in 2016-17 of planting.

Statistical Analysis

A Statistical software SPSS version 25.0 was used for the analysis. Mean values of all the parameters (number of roots, length of roots (cm), number of leaves, number of sprouts) were calculated to determine the highest growth of each trial for every applied concentrations of hormones (2000 ppm-7000 ppm).

To compare the average effect of treatment at different level Multivariate Analysis of Variance (MANOVA) technique have been applied. The results of different assumptions which are acquired for applying MANOVA are discussed briefly.

Abbreviations

IBA: Indole Butyric acid; IAA: Indole acetic acid; KP: Khyber Pakhtunkhwa; TIFAC: Forecasting and Assessment Council; CITES: Convention for International Trade in Endangered Species; MANOVA: Multivariate Analysis of Variance; IAA: Indole acitic acid.

Declarations

Ethics approval and consent to participate:

There are no ethical guidelines for research on the conservation of *T. wallichiana*.

Consent for publication:

Not applicable.

Availability of data and materials:

The additional data regarding the *T. wallichiana*'s in-situ conservation (relationship between the species growth through hormones (IBA, IAA) applied) by looking into each mentioned parameters (Word docs 40 kb).

In current research study the datasets analysed are available from the corresponding author on reasonable request.

Competing interests:

The authors declare that they have no competing interests.

Funding:

This project was financially supported by the National Research Project for Universities (NRPU), Higher Education Commission (HEC) Pakistan. The project details are: No: 20-3690/NRPU/R&D/HEC/2014/816. The funding bodies have no role in the design and collection, analysis, interpretation of the data and in writing of the manuscript.

Author's contribution:

BK designed the research project plan ; JI did field and laboratory work, also wrote the manuscript; SK review and contributed in the writing; NG presented the concept idea; IAM hepled in field work of this study; NY review the manuscript also helped in the laboratory work; IA provide the field design for In-situ experiment at the forest nursery; SM helped to statistically analyse the results. Final manuscript is read and approved by all of the authors.

Acknowledgement:

The co-authors are very thankful to all of the organization (Department of Environmental Sciences, University of Peshawar; Pakistan Forest Institute (PFI); Directorate of Non-Timber Forest Produce, Forest Department, Peshawar; and Higher Education Commission Pakistan (HEC)) who contributed positively in the completion of this project.

Authors details:

¹Department of Environmental Sciences, University of Peshawar, 25120, Khyber Pakhtunkhwa, Pakistan

²Department of Soil and Environmental Sciences, The University of Agricultural Peshawar, 25000, Khyber Pakhtunkhwa, Pakistan

³Direcctorate of Non-Timber Forest Produce, Forest Department, Peshawar, Khyber Pakhtunkhwa, 25000, Pakistan

⁴Department of Statistic, Islamia College Peshawar, 25120, Khyber Pakhtunkhwa, Pakistan

References

1. Uniyal SK.2013. Bark removal and population structure of *Taxus wallichiana* Zucc in a temperate mixed conifer forest of western Himalaya. *Environmental Monitoring and Assessment.*, 185(4): 2921–2928.
2. Hussain A, Qarshi IA, Nazir H, Ullah I, Rashid M, Shinwari ZK. In vitro callogenesis and organogenesis in *Taxus wallichiana* ZUCC, The Himalayan Yew. *Pak J Bot.* 2013;45(5):1755–9.
3. Mulliken T. and P. Crofton.2008. Review of the status, harvest, trade and management of seven Asian CITES-listed medicinal and aromatic plant species. *BfN-Skripten, Federal Agency for Natural*

Conservation, Bonn, Germany.

4. Hussain A, Qarshi IA, Nazir H, Ullah I, Rashid M, Shinwari ZK. 2013. In vitro callogenesis and organogenesis in *Taxus wallichiana* ZUCC, The Himalayan Yew. *Pakistan Journal of Botany.*, 45(5): 1755–1759.
5. Anon. Wealth of India. New Delhi: Vol. X. Publications and Information Directorate, CSIR; 1976. 591 pp.
6. Chee PP. In vitro culture of zygotic embryos of *Taxus* species. *HortSci.* 1994;29:695–7.
7. Valis M, Koci J, Tucek D, Lutonsky T, Kopova J, Barton P, Vysata O. D.Kjíckova, J. Korabecny. 2014. Common yew intoxication: a case report. *Journal of Medical Case Report.*, 8:4.
8. Rahman S, Salehin F, Uddin MJ, Zahid A. *Taxus Wallichiana* Zucc. (Himalayan yew): insights on its anti-microbial and pharmacological activities. *OA Alternative Medicine.* 2013;1(1):3.
9. Shabir A. 2009. Woods of Pakistan. Higher Education Commission., 218–9.
10. Rikhari HC, Palni LMS, Sharma S. and S. K. Nandi. 1998. Himalayan yew: stand structure, canopy damage, regeneration and conservation strategy, *Environmental Conservation.*, 25 (4): 334–341.
11. Sharma P, Uniyal PR, Slowik J. Community involvement and conservation of *Taxus Baccata* in Pangi Valley, Himachal Pradesh. *Natural Areas Journal.* 2014;34(4):470–4.
12. *Iqbal, J., Meilan, R., & Khan, B. (2020). Assessment of risk, extinction and threats to Himalayan yew in Pakistan. Saudi Journal of Biological Sciences,* 27(2), 762–767.
13. Pant S. and S. S. Samant. 2008. Population ecology of the endangered Himalayan Yew in Khokhan Wildlife Sanctuary of North Western Himalaya for conservation management. *Journal of Mountain Science.*, 5(3): 257–264.
14. Nhut DT, Hien NTT, Don NT. and D.V.Khien. 2007. In vitro shoot development of *Taxus wallichiana* Zucc, a valuable medicinal plant. In *Protocols for Micropagation of Woody Trees and Fruits.*, 107–116.
15. Nimasow G, Dai Nimasow O, Singh Rawat J, Tsiring G. and T. Litin. 2016. Remote sensing and GIS-based suitability modeling of the medicinal plant (*Taxus baccata* Linn.) in Tawang district, Arunachal Pradesh, India. *Current Science.*, (00113891): 110 (2).
16. Sharma S, Thokchom R. A review on endangered medicinal plants of India and their conservation. *J Crop Weed.* 2014;10(2):205–18.
17. Yadav D, Kumar P, Tiwari SP. and D. K. Chauhan. 2013. Comparative palynological and wood anatomical studies of Indian *Taxus wallichiana* Zucc., *Cephalotaxusmannii* Hook. and *Cephalotaxusgriffithii* Hook. *Plant Systematics and Evolution.*, 299 (7): 1231–1242.
18. Nimachow G, Rawat JS, Dai O. Status of Himalayan yews in West Kameng district of Arunachal Pradesh. *Current Science.* 2010;98:1434–7.
19. Schippmann U. 2001. Medicinal plants significant trade study. *Bundesamt für Naturschutz.*
20. Saqib Z, Malik RN. and S. Z. Husain. 2006. Modeling potential distribution of *Taxus wallichiana* in Palas Valley, Pakistan. *Pakistan Journal of Botany.*, 38(3): 539.

21. Lange D. 2002. Medicinal and aromatic plants: trade, production, and management of botanical resources. In XXVI International Horticultural Congress. The Future for Medicinal and Aromatic Plants., 629:177–197.
22. Haq F. The critically endangered flora and fauna of District Battagram Pakistan. Advances in Life Sciences. 2012;2:118–23.
23. Joshi K. 2009. *Taxus wallichiana* (Himalayan Yew; Loth salla) in Nepal: Indigenous Uses, Conservation, and Agenda for Sustainable Management. Ethnobotanical Leaflets., (12):8.
24. Singh JS. 1992. Man, and forest interactions in Central Himalaya. Himalayan environment and development: Problems and Perspectives., 57–79.
25. Vishnu-Mitre. 1984. Floristic change in the Himalaya (southern slopes) and Shiwliks from the mid tertiary to recent times. In Whyte RO, editor, The Evolution of the East Asian environment., 483–503.
26. Steinfield D. 1992. Early lessons from propagating Pacific yew. Rocky Mountain Forest and Range Experiment Station Technical Research Report., 221.
27. Schneck V. Studies on the influence of clone on rooting ability and rooting quality in the propagation of cuttings from 40- to 350-year-old *Taxus baccata* L. ortets. *Silvae Genetica*. 1996;45:246–9.
28. Fordham AJ, Spraker LS. Propagation manual of selected gymnosperms. *Arnoldia*. 1977;37:1–88.
29. Nandi SK, Palni LMS, Rikhari HC. Chemical induction of adventitious root formation in *Taxus Baccata* cuttings. *Plant Growth Regulation*. 1996;19:11–22.
30. Khali RP. 2001. Ecological studies on *Taxus baccata* L. in relation to regeneration and conservation. Unpublished Ph. D. Thesis, FRI Deemed University, Dehra Dun.
31. Haissig BE. Influences of auxins and auxin synergists on adventitious root primordium initiation and development. *NZJ For Sci*. 1974;4(31):1–323.
32. Hartman HT, Kester DE. Plant Propagation: Principles and Practices. USAS: Prentice Hall; 1983. Englewood Cliffs., p. 662.
33. Philips IDJ. (1971). Introduction to the Biochemistry and Physiology of PG Hormones.
34. Khali RP, Sharma AK. Effect of phytohormones on propagation of Himalayan Yew (*Taxus Baccata* L.) through stem cuttings. *Indian Forester*. 2003;129:289–94.
35. Mishra A, Singh A, Kukreja AK, Patra NK. 2000. Adventitious root induction in stem cuttings of *Taxus baccata* Linn. For ex-situ cultivation. *J Non-Wood Forestry Prod*. 12:63 – 6.
36. Mitter H, Sharma A. Propagation of *Taxus baccata* Linn. by stem cuttings. *Indian forester*. 1999;125(2):159–62.
37. Chandra JP, Verma SD. (1989). Rootability of softwood cuttings of individual mother trees of Eucalyptus. In Proceeding Seminar on Vegetative Propagation, Held at Coimbatore, India (Vol. 27).
38. Dubey KP. Himalayan yew (*Taxus baccata*) conservation: A vegetative approach. *Indian forester*. 1997;123(12):1150–4.
39. Nandi SK, Rikhari HC, Nadeem M, &Palni LMS. Clonal propagation of *Taxus baccata* L.—a Himalayan asset under threat. *Physiol Mol Biol Plants*. 1997;3:15–24.

40. Nautiyal S, Singh U, Gurumurti K. Rooting response of branch cuttings of teak (*Tectonagrandis*) as influenced by growth hormones and position of the cutting on the crown. *Indian Forester*. 1992;118(2):112–21.
41. Aslam M, Arshid S, Rather MS, Salathia HS, Seth CM. Auxin induced rooting in *Taxus baccata* Linn. Stem cuttings. *Indian J Forstry*. 2007;30:221–6.
42. Gurumurti K, Bhandari HCS. Induction of rooting in cladode cuttings of *Casuarina equisetifolia*. *Curr Sci*. 1988;57(17):958–9.
43. Singh RR, Chander H. Effect of auxins on rooting behaviour of neem (*Azadirachtaindica*) branch cuttings. *Indian Forester*. 2001;127(9):1019–24.
44. Nautiyal S, Dhyani M, Kumar P, Bhandari HC. (2004, November). Rooting response of juvenile shoot cuttings of *Terminalia arjuna* under different hormonal treatment. In National Workshop on Potential and Strategies for Sustainable Development of Vanya Silks in the Himalayan States (pp. 70–4).
45. Blazich FA. (1988). Chemicals and formulations used to promote adventitious rooting. *Advances in plant sciences series (USA)*.
46. Pal M. Clonal propagation for yield improvement in forest plantations. *IPPTA*. 1992;4:61–1.
47. Chauhan KS, Reddy TS. Effect of growth regulators and mist on rooting in stem cuttings of plum (*Prunus domestica* L.). *Indian Journal of Horticulture*. 1974;31(3):229–31.
48. Avanzato D, Couvillon GA, Pokorny FA. 1998. The influence of P-ITB (phenyl indole-3-thiolobutyrate), an aryl ester of IBA, on the rooting of 'Redhaven' peach. In International Symposium on Vegetative Propagation of Woody Species., 227: 197–201.
49. Eccher T. 1987. Response of cuttings of 16 *Taxus* cultivars to rooting treatments. In International Symposium on Vegetative Propagation of Woody Species., 227: 251–253.
50. Aslam M, Rather MS. Macro-propagation of *Taxus baccata* Linn: a Novel Method for Conserving a Critically Endangered Medicinal Plant. *Indian Forester*. 2008;134(8):1058–66.
51. Nasir N, Kamili AN, Shah D, Zargar MY. (2018). Adventitious Rooting in shoot cuttings of *Taxus wallichiana* Zucc., an Endangered Medicinally Important Conifer of Kashmir Himalaya. *Forest Research*, 7 – 2..
52. Sevik H, Guney K. (2013). Effects of IAA, IBA, NAA, and GA3 on rooting and morphological features of *Melissa officinalis* L. stem cuttings. *The Scientific World Journal*, 2013.
53. Topacoglu O, Sevik H, Guney K, Unal C, Akkuzu E, Sivacioglu A. Effect of rooting hormones on the rooting capability of *Ficus benjamina* L. cuttings. *Šumarski list*. 2016;140(1–2):39–44.
54. Chhun T, Taketa S, Tsurumi S, Ichii M. The effects of auxin on lateral root initiation and root gravitropism in a lateral rootless mutant Lrt1 of rice (*Oryza sativa* L.). *Plant Growth Regul*. 2003;39(2):161–70.
55. Koleva Gudeva L, Trajkova F, Mihajlov L, Troiciki J. Influence of different auxins on rooting of rosemary, sage and elderberry. *Annual Research Review in Biology*. 2017;12(5):1–8.

56. Gill MK, Chauhan SK, Gossal SS. Macro-and micro-propagation of *Azadirachta indica*. Indian Forester. 2006;132(9):1159–66.
57. Aslam M, Raina PA, Rafiq RU, Siddiqi TO, Reshi ZA. (2017). Adventitious root formation in branch cuttings of *Taxus wallichiana* Zucc. (Himalayan yew): A clonal approach to conserve the scarce resource. Current Botany, 127–135.
58. Puri SC, Verma V, Amna T, Qazi GN, Spiteller M. An endophytic fungus from *Nothapodytes f oetida* that produces Camptothecin. Journal of natural products. 2005;68(12):1717–9.

Figures



Figure 1

Leaves and fruit (bright red berries) of Himalayan Yew (*Taxus wallichiana*) from Kalam, Swat (Iqbal et al., 2020).



Figure 2

(a) Mixing and sieving of forest, sand and agriculture soil (1:1:1); (b) Filling of Polythene bags; (c) Cuttings treatment with the concentrated hormonal solutions (2000-7000 ppm); (d) *T. wallichiana*'s cuttings preparation with sharp knife; (e) Plant cuttings washing with tap water.



Figure 3

Himalayan yew (*Taxus wallichiana*) cuttings grown in the Lalku Valley, Khyber Pakhtunkhwa, Pakistan.

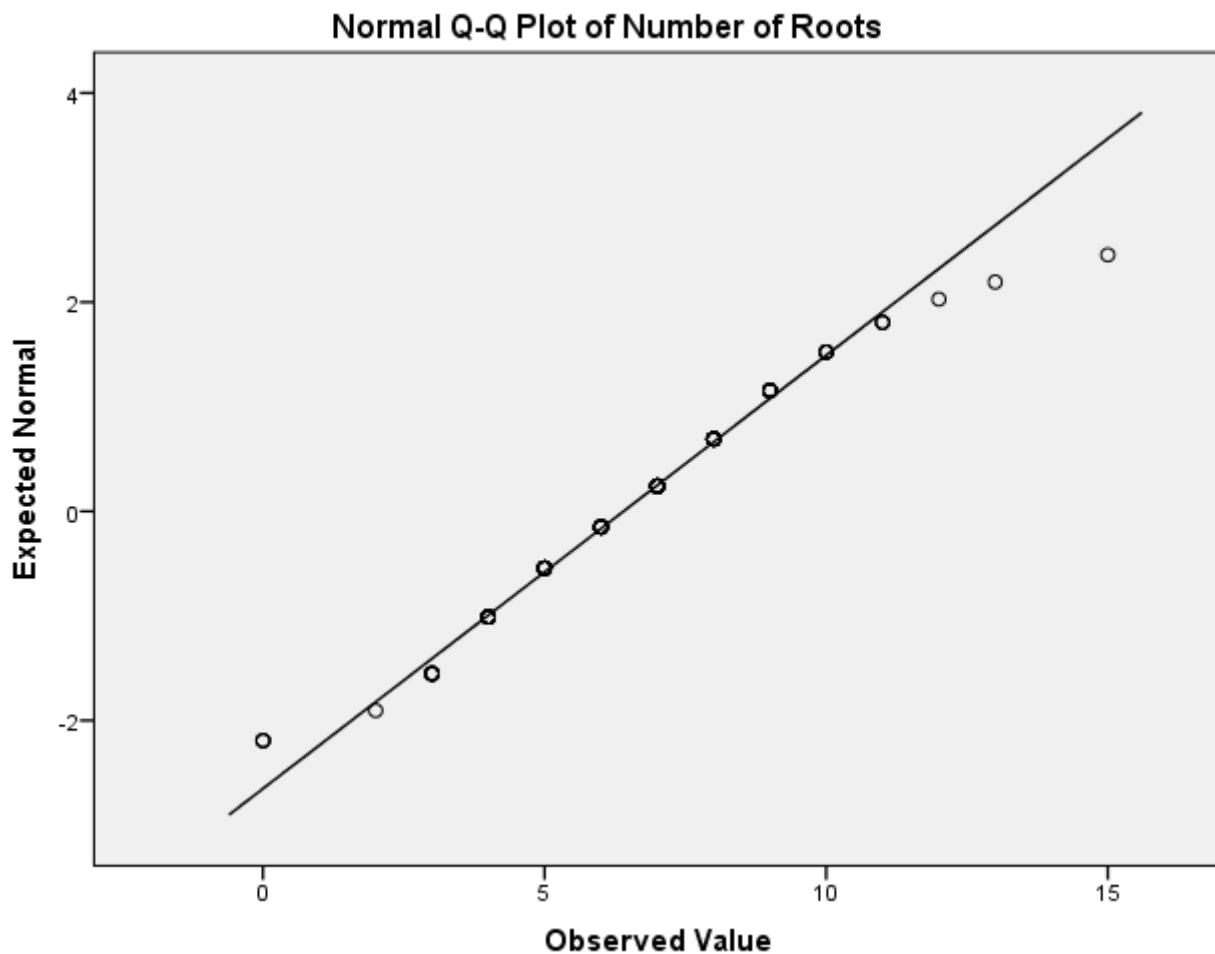


Figure 4

Normal Q-Q Plot of observed and expected values for number of roots of *T. wallichiana* Cuttings.

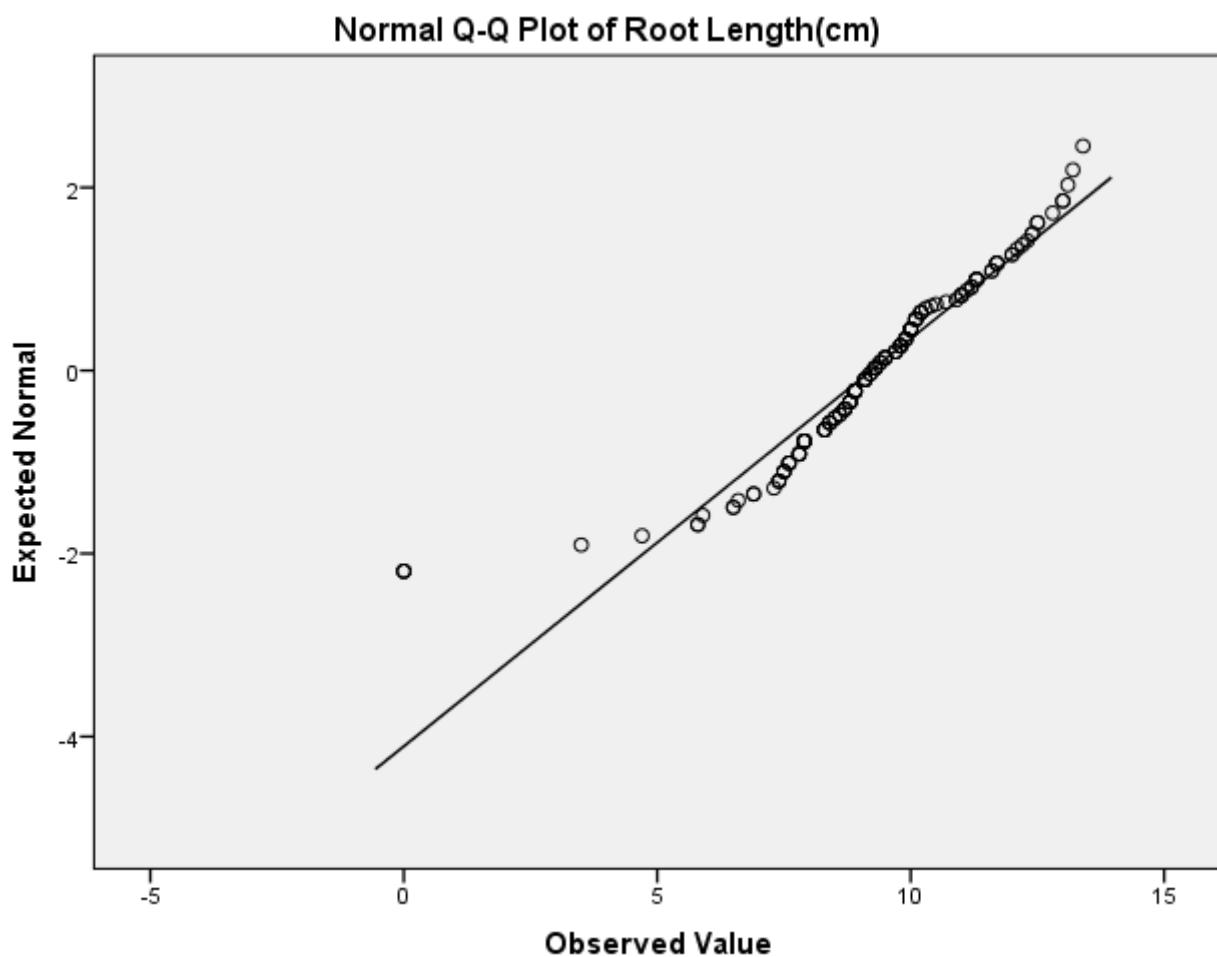


Figure 5

Normal Q-Q Plot of observed and expected values for length of roots of *T. wallichiana*'s cuttings.

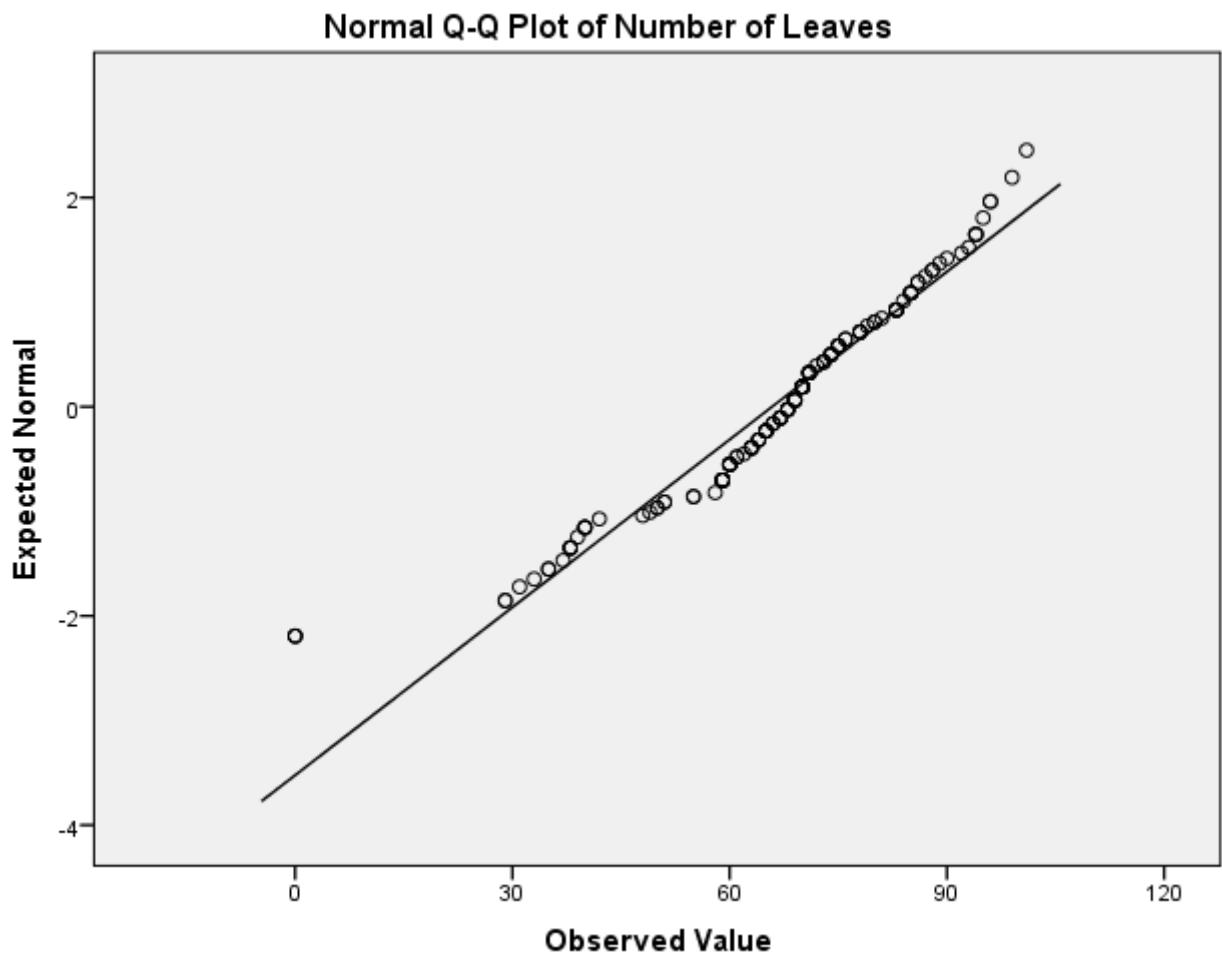


Figure 6

Normal Q-Q Plot of observed and expected values for number of leaves of *T. wallichiana*'s cuttings.

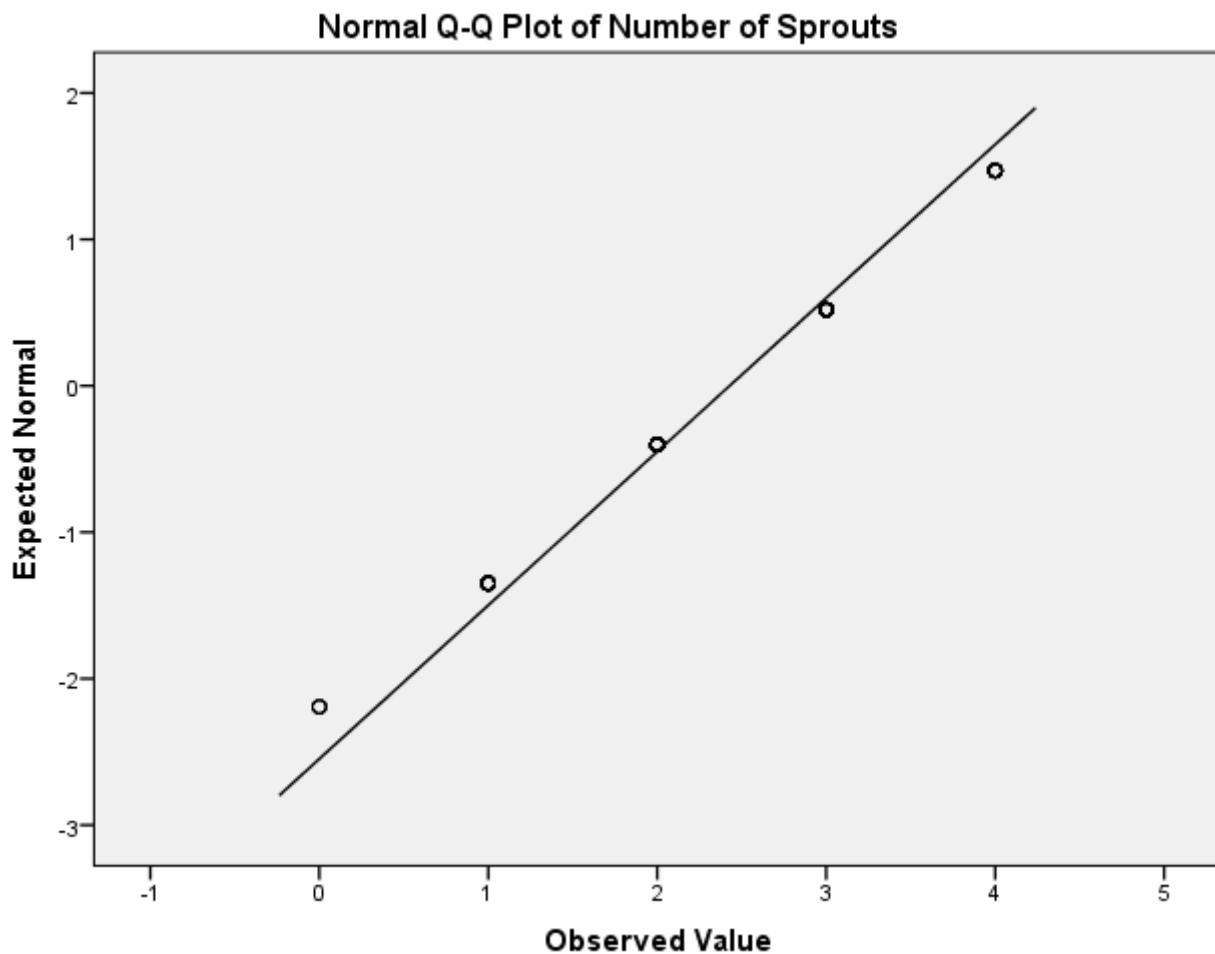


Figure 7

Normal Q-Q Plot of observed and expected values for number of sprouts of *T. wallichiana*'s cuttings.

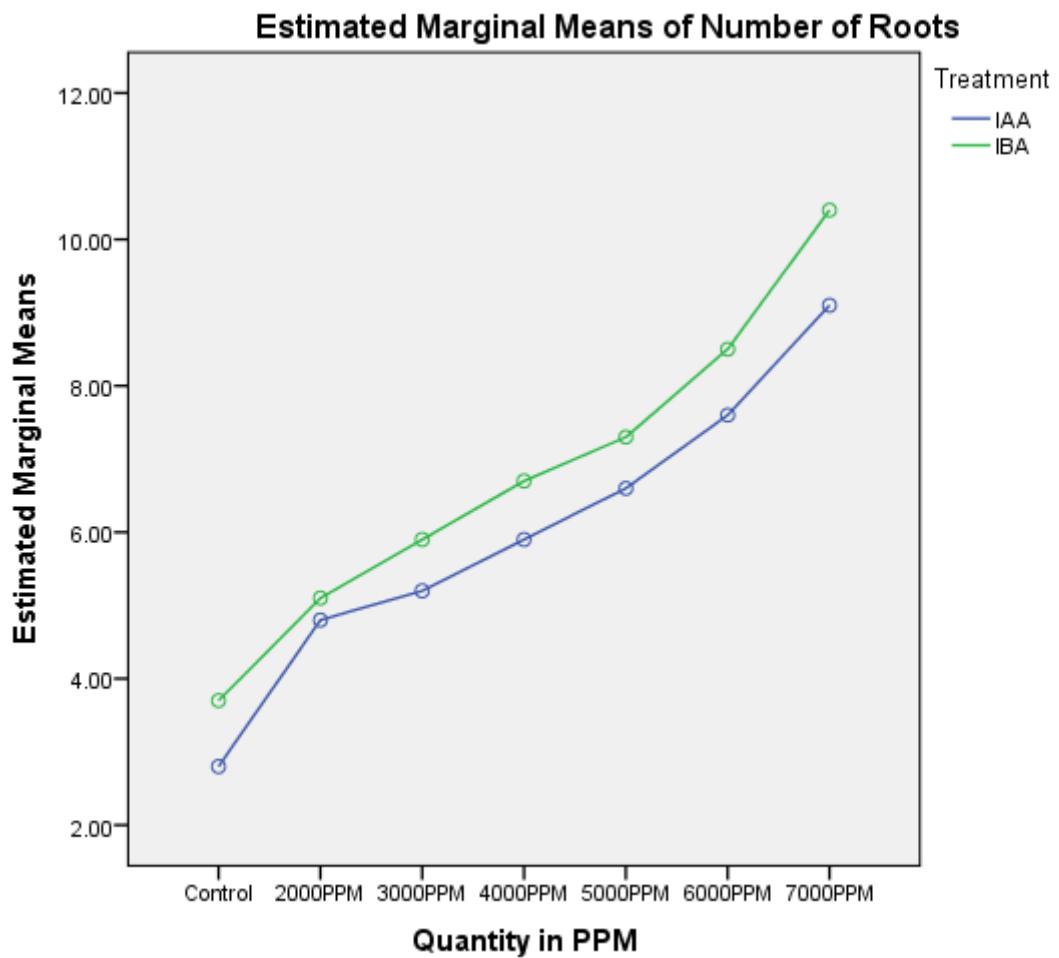


Figure 8

Comparison of number of roots of *T. wallichiana* fresh stem cuttings by the application of various concentrations of hormones (IBA, IAA).

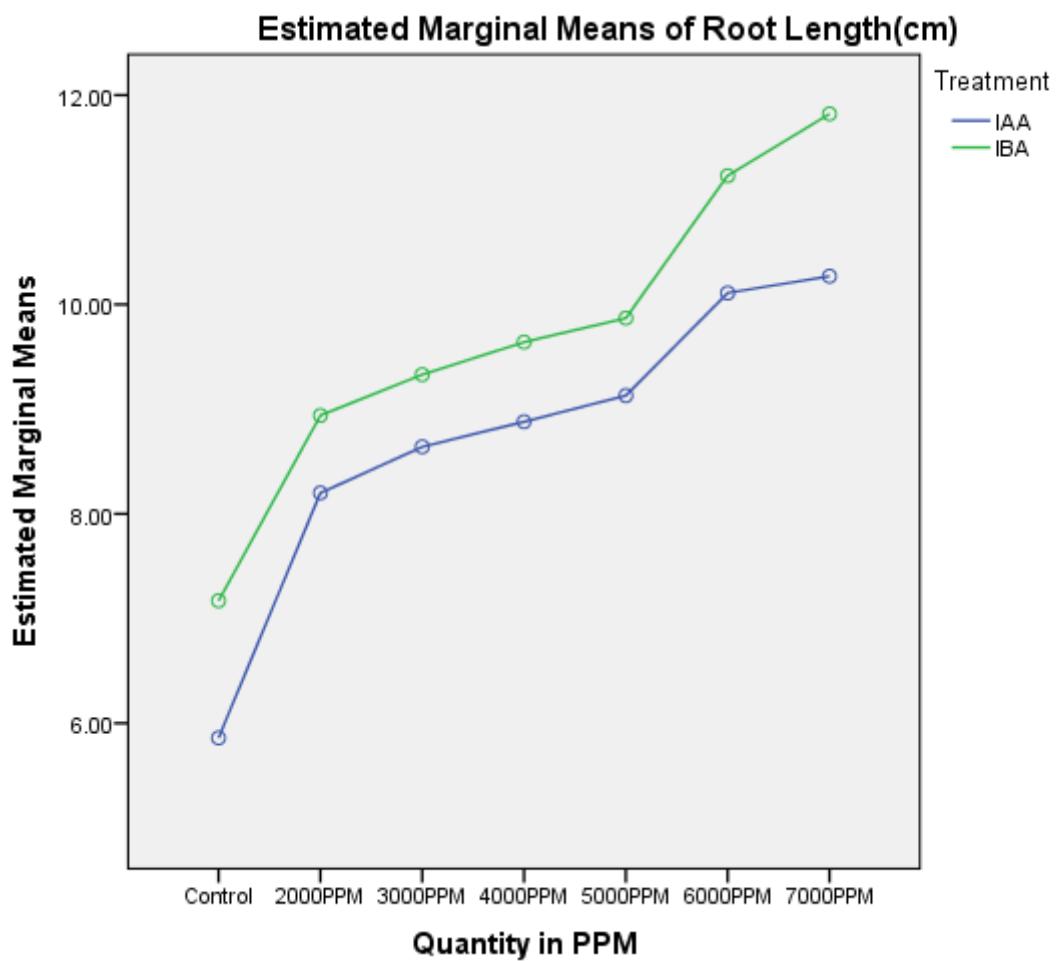


Figure 9

Comparison of length of roots (cm) of *T. wallichiana* fresh stem cuttings by the application of various concentrations of hormones (IBA, IAA).

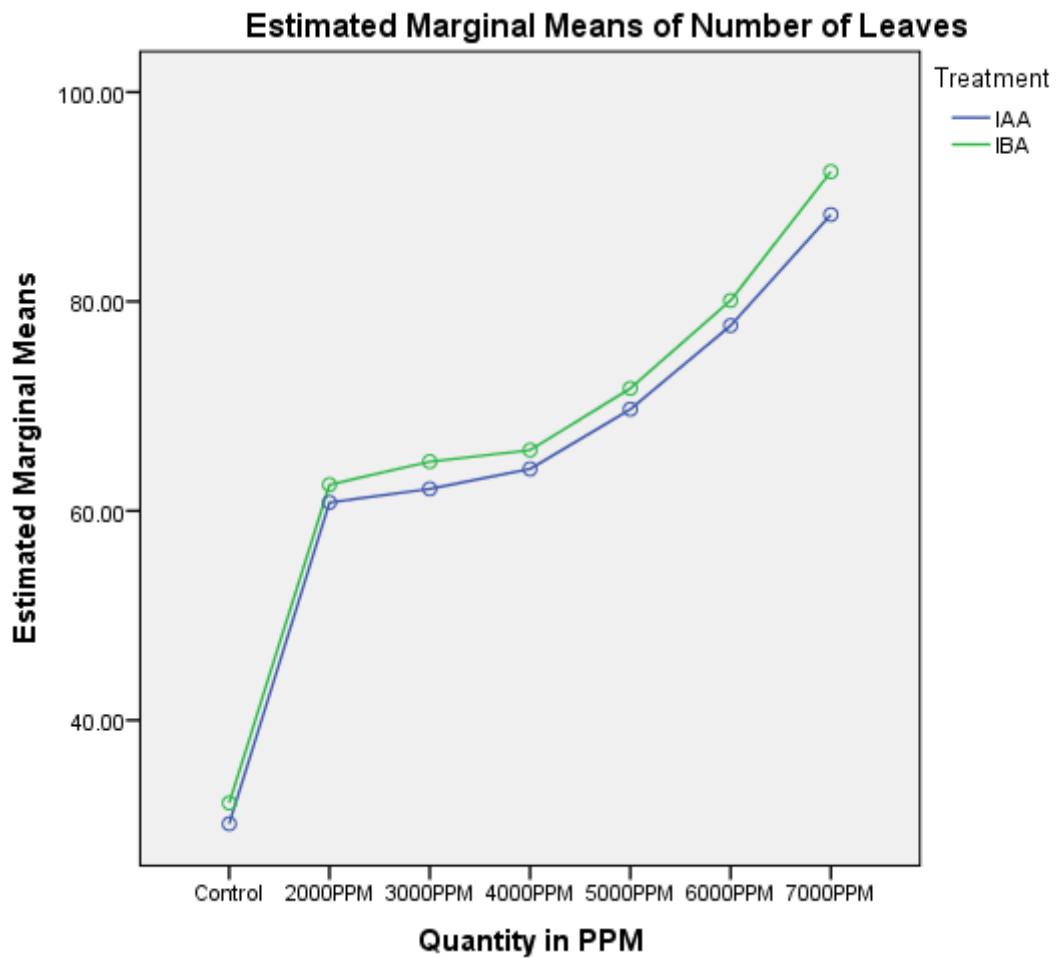


Figure 10

Comparison of number of leaves of *T. wallichiana* fresh stem cuttings by the application of various concentrations of hormones (IBA, IAA).

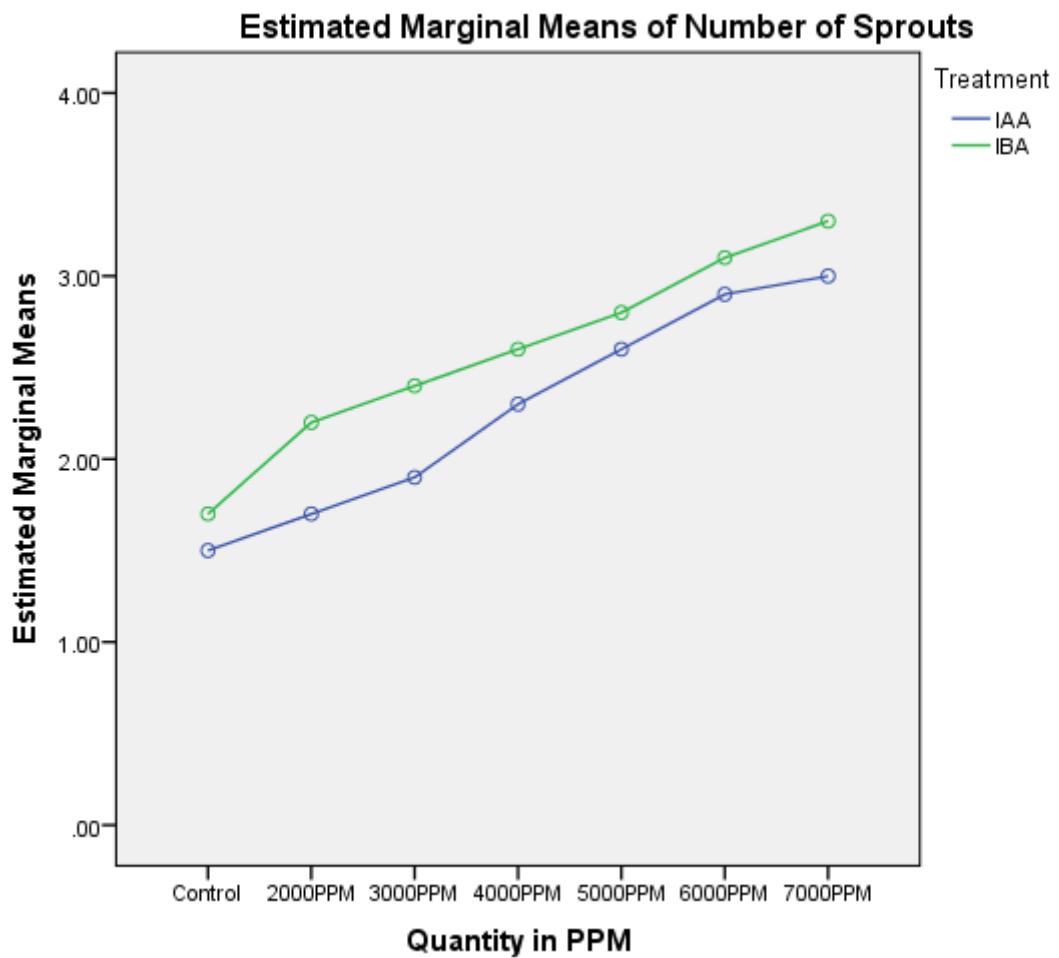


Figure 11

Comparison of sprouts of *T. wallichiana* fresh stem cuttings by the application of various concentrations of hormones (IBA, IAA).

Survival percentage by applied different concentrations of IBA and IAA

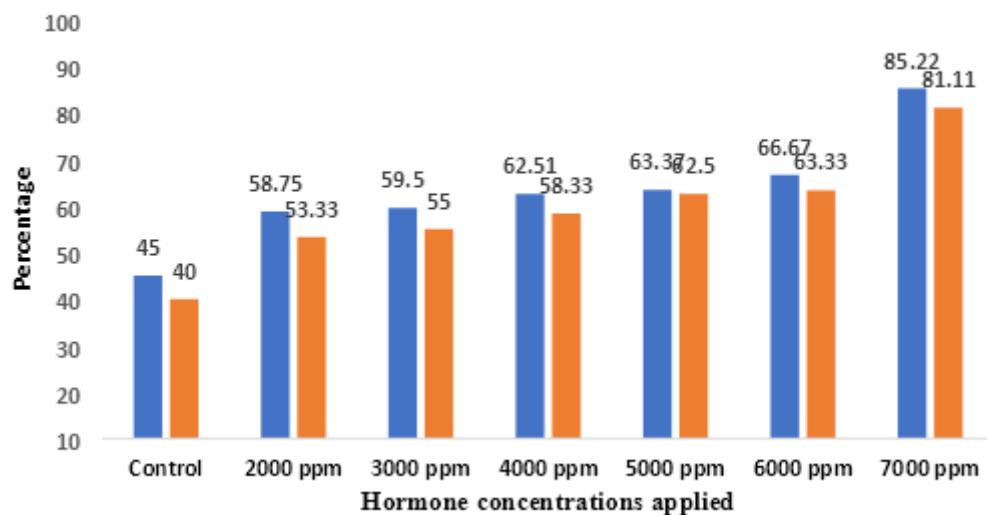


Figure 12

Survival percentage of *T. wallichiana* cuttings by the application of various concentrations of hormones (IBA and IAA).

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [Docs.docx](#)
- [ConflictofInterest.pdf](#)