

Evolutionary loss of thermal acclimation accompanied by periodic monocarpic mass flowering in *Strobilanthes* species

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Abstract

While life history, physiology and molecular phylogeny in plants have been widely studied, understanding how physiology changes with the evolutionally life historical change remains largely unknown. In two closely related understory *Strobilanthes* plants, the molecular phylogeny has previously shown that the monocarpic 6-year masting *S. flexicaulis* have evolved from a polycarpic perennial, represented by the basal clade *S. tashiroi*. The polycarpic *S. tashiroi* exhibited seasonal thermal acclimation with increased leaf respiratory and photosynthetic metabolism in winter, whereas the monocarpic *S. flexicaulis* showed no thermal acclimation. The monocarpic *S. flexicaulis* required rapid height growth after germination under high intraspecific competition, and the respiration and N allocation were biased toward nonphotosynthetic tissues. By contrast, in the long-lived polycarpic *S. tashiroi*, these allocations were biased toward photosynthetic tissues. The life-history differences between the monocarpic *S. flexicaulis* and the polycarpic *S. tashiroi* are represented by the “height growth” and “assimilation” paradigms, respectively, which are controlled by different patterns of respiration and nitrogen regulation in leaves. The obtained data suggest that the monocarpic *S. flexicaulis* with the evolutionary loss of thermal acclimation will more often exhibit increased vulnerability to global warming.

Introduction

Atmospheric carbon dioxide (CO₂) concentrations are increasing, because of extensive fossil fuel burning. Currently, the average air temperature is increasing by 0.85°C per year in globally¹, but the increase in Japan reaches 1.15°C per year (according to the Japan Meteorological Agency). Leaf gas exchange of vascular plants plays an important role in regulating global carbon cycling^{2,3}. To avoid thermal damage and to ensure carbon assimilation under seasonal variations in air temperature, terrestrial vascular plants usually adjust their photosynthesis and respiration to altered air temperatures²⁻⁷. Even tropical plants and C₄ plants from warm habitats exhibit photosynthetic acclimation to altered air temperatures⁸⁻⁹. To improve the prediction of the effects of plant thermal acclimation on terrestrial carbon cycling under global warming, numerous experiments have been conducted under regulated environments over short to long periods, and these results have been fed into meta-analyses across various functional types and biomes^{3,7,10-13}.

The range of seasonal variations in air temperature exceeds 20°C in the warm-temperate forests of southern Japan. The values of Q₁₀ values (proportional change in respiration rates per 10°C change in temperature) for the respiration rates of many vascular plants are around 2, meaning that the respiration rates increase two-fold with a 10°C increase in temperature. The reported seasonal variation in Q₁₀ has not been large or has been nonsignificant for various plant species^{3,14,15}. Species-specific thermal acclimation of cell metabolism, which is measured at a given temperature over seasons, could be thus considered a physiologically important determinant of the vulnerability of plant growth and survival under global warming. The temperature response of respiration rates reflects metabolic states of plant tissues^{16,17}. Nevertheless, the information on thermal acclimation in rare and/or understory plants is

virtually missing. Therefore, for these plants, we cannot evaluate the predicted rapid loss of biodiversity under global warming.

We examined two closely-related sympatric understory plants of the genus *Strobilanthes* species (Acanthaceae), *S. tashiroi* Hayata and *S. flexicaulis* Hayata, in warm-temperate, overwintering evergreen forests in Japan (Fig. 1). They have contrasting life histories: *S. tashiroi* is a perennial herb with continuous flowering and non-masting phenology, whereas *S. flexicaulis* is a shrub with periodic monocarpic mass flowering and subsequent death every six years¹⁸. Molecular phylogeny shows that the periodical phenology of *S. flexicaulis* has locally evolved from the polycarpic phenology of *S. tashiroi*¹⁹.

Thermal acclimation of cell metabolism is expected to be different between the two plants with distinctive phenology. Costly thermal acclimation may be disadvantageous for the monocarpic *S. flexicaulis*, which must undergo rapid height growth during its short lifespan, whereas *S. tashiroi*, thermal acclimation should be advantageous to ensure the effective photosynthesis over its long lifespan. Here, we examined the thermal acclimation and its related physiological states of the two *Strobilanthes* species in the field. The photosynthetic and dark respiration rates in leaves, stems, and roots were compared between summer and winter on Okinawa Island. We found that the monocarpic *S. flexicaulis* leaves did not show any seasonal acclimation in respiration and photosynthesis, whereas the polycarpic *S. tashiroi* leaves showed increased respiratory and photosynthetic faculties in winter. We discuss the loss of thermal acclimation in *S. flexicaulis* and its implications in the context of global warming causing the extinction of rare species and loss of biodiversity. This study is the first report that integrates physiological ecology (thermal acclimation) and plant phenology (life history) with molecular phylogeny. Until now, there has been no study linking thermal acclimation with life history strategies in any plant.

Results

Seasonal variations in air temperature in the study site. From 2017 to 2020, the yearly maximum, minimum, and average air temperatures were $29.2 \pm 0.49^\circ\text{C}$, $5.8 \pm 0.99^\circ\text{C}$, and $19.3 \pm 0.23^\circ\text{C}$ (mean \pm 1 SD), respectively. No conspicuous variations in the seasonal trends were found among the past four years (Supplementary Fig. 1). The difference between the annual maximum and minimum air temperatures was $23.4 \pm 0.69^\circ\text{C}$ (mean \pm 1 SD) from 2017 to 2020. This large difference indicates a considerable seasonal variation in air temperature even in the forest understory. The observed results are typical climatological characteristics in the warm-temperate, evergreen forests in East Asia. There were no conspicuous drought and disturbance events, such as tropical typhoons, since 2016, when their germination of *S. flexicaulis* began.

Leaf area-based photosynthesis increases in winter in the polycarpic *S. tashiroi*. The light-response curves of the photosynthesis rates were measured in winter and summer under nonregulated air temperature conditions in a field setting (Supplementary Fig. 2). The measured leaf temperature was 20–24°C in winter, whereas it was 28–29°C in summer. The two species exhibited different seasonal variations in leaf gas exchange (Fig. 2). In the monocarpic *S. flexicaulis*, the maximum net assimilation

rates (A_{\max}) did not show seasonal variations, but dark respiration rates (R_d) decreased significantly in winter. In contrast, in the polycarpic *S. tashiroi*, A_{\max} increased significantly in winter, but R_d did not vary even though the winter temperatures were low. The A_{\max}/R_d ratio was significantly higher in summer than in winter in both species. However, the summer-to-winter ratios in A_{\max}/R_d were 1.9 and 1.6 in *S. flexicaulis* and *S. tashiroi*, respectively. This obvious difference in the ratios indicates that the polycarpic *S. tashiroi* maintains a more homeostatic carbon balance (see ref. No. 3) than the monocarpic *S. flexicaulis*. *S. tashiroi* did not show the seasonal variations in light compensation points (LCPs), whereas the LCPs in *S. flexicaulis* decreased significantly in winter, because of low dark respiration rates in winter. The initial slopes of the light-response curves were kept at relatively high values (> 0.061) over the seasons in both species, indicating that the leaves did not suffer from conspicuous photoinhibition²⁰⁻²².

Cell metabolisms increases in winter in the polycarpic *S. tashiroi*, but not in the monocarpic *S. flexicaulis*. Mass-based dark respiration rates in leaves, stems, and roots were measured in winter and summer in the field. To standardize the variations in measurement temperature and to evaluate the seasonal variations in cell metabolism between summer and winter, the obtained values were standardized at 19°C and 28°C by using $Q_{10} = 2$ (Fig. 3a-f); these temperatures corresponded to the mean values of plant temperature measured in winter and summer in the field setting, respectively. The two species exhibited seasonally different responses in the respiration rates, especially in the aboveground tissues. Comparing between winter and summer, the respiration rates at a given temperature (i.e., metabolic activity) increased significantly in winter for the leaves and stems of polycarpic *S. tashiroi*. The respiratory metabolic activity of *S. tashiroi* leaves increased 1.7 times from summer to winter; this phenomenon corresponds to the increase in A_{\max} observed in winter. In contrast, in the monocarpic *S. flexicaulis*, no metabolic change over seasons was found for any tissues. For the roots, no seasonal metabolic change in cell respiration was found in either species. Comparing between the two species, no significant differences in the mass-based root respiration rates were found, whereas the mass-based leaf and stem respiration rates were significantly lower in the monocarpic *S. flexicaulis* than in the polycarpic *S. tashiroi* ($P = 0.001$ for leaves; $P < 0.001$ for stems) in both seasons (Supplementary Table S2).

Similar to cell metabolisms, no seasonal variations in nitrogen (N) concentrations were not found in any tissues in the monocarpic *S. flexicaulis* (Fig. 3g-i). In the polycarpic *S. tashiroi*, no seasonal variations in N concentrations in leaves were found, but those in stems and roots increased significantly in summer. Therefore, the seasonal variations in N concentrations were not consistent with those in the respiratory metabolism of *S. tashiroi* cells.

Photosynthetic N use efficiency (PNUE; N-based photosynthetic rate) was estimated from the mean values of A_{\max} and foliar N. The values of PNUE in the polycarpic *S. tashiroi* were 0.191 and 0.147 mmol mmol N⁻¹ in winter and summer, respectively; i.e., PNUE increased 1.3 times from summer to winter because A_{\max} increased in winter, but foliar N remained unchanged. In the monocarpic *S. flexicaulis*, the PNUE values were 0.126 and 0.139 mmol mmol N⁻¹ in winter and summer, respectively; i.e., PNUE did not vary largely between seasons. These data indicate that the N within the lamina is allocated mostly to

photosynthetic enzymes (such as Rubisco) in winter in the polycarpic *S. tashiroi* leaves, whereas the allocation of N within the lamina did not vary largely in the monocarpic *S. flexicaulis* leaves. Thus, Rubisco abundance rather than N concentrations within the lamina underpins the thermal acclimation response in polycarpic *S. tashiroi* leaves.

The respiration, N, and dry matter allocations vary between the two species. The two species have contrasting plant forms in relation to the differences in their life histories (Fig. 4a-d). The monocarpic *S. flexicaulis* had a significantly higher stem mass ratio than did the polycarpic *S. tashiroi*, whereas *S. tashiroi* had a significantly higher leaf mass ratio (resulting in a high leaf area ratio) than did *S. flexicaulis*. The high dry matter allocation into stems in the monocarpic *S. flexicaulis* is due to its unique life history, which requires rapid height growth during a short period (6 years) after germination to escape from high intraspecific competition (i.e., shading by the other individual plants). No significant difference between species was found in root dry mass. The values of leaf mass per area (LMA) were 48.3 ± 2.4 and 37.2 ± 4.5 g m⁻² (mean \pm 1 SD) in *S. flexicaulis* and *S. tashiroi*, respectively; LMA was significantly higher in *S. flexicaulis* than in *S. tashiroi* ($P < 0.001$).

The whole-plant respiration rates in monocarpic *S. flexicaulis* were 1.57 and 2.92 nmol g⁻¹ s⁻¹ in winter (19°C) and summer (28°C), respectively, and those in polycarpic *S. tashiroi* were 2.59 and 3.70 nmol g⁻¹ s⁻¹ in winter and summer, respectively. In terms of interspecific variation, the whole-plant respiration rates in *S. tashiroi* were 1.7–1.3 times of those in *S. flexicaulis*, because the polycarpic *S. tashiroi* was leafier. In terms of seasonal variation, the summer-to-winter ratios of the whole-plant respiration rates were 1.86 and 1.43 in *S. flexicaulis* and *S. tashiroi*, respectively. Therefore, with regard to whole-plant respiration, the polycarpic *S. tashiroi* maintained higher homeostasis over seasons through the thermal acclimation of leaf respiration (see ref. No. 17) than did the monocarpic *S. flexicaulis*.

Interspecific differences were also found in respiration and N allocation among tissues (Fig. 4e-f). In the polycarpic *S. tashiroi*, leaf respiration amounted to approximately 42–55% of the whole-plant respiration rates, and leaf N amounted to approximately 42–50% of the whole N contents. On the other hand, in the monocarpic *S. flexicaulis*, leaf respiration amounted to approximately 29% of the whole-plant respiration rates, and leaf N amounted to approximately 34–36% of the whole N contents.

Discussion

This is the first paper combining physiological traits and phenology in plants. Until now, life history and phenology are not directly compared with physiological acclimation capacity, because the phenological differences are represented in distant phylogeny and direct comparisons are impossible. Furthermore, we cannot deeply know what physiological traits are correlated to the evolutionary life history change. However, in this study, we examined two closely related plants with distinct life histories and phenologies, and elucidated the ecophysiological differences that have supported their own life history.

For understory plants in stable overwintering evergreen forests, thermal acclimation is the primary factor for determining carbon assimilation, because the seasonal variations in light are relatively small, and prolonged drought is rare. Polycarpic *S. tashiroi*, having a long-lived lifespan, exhibits winter thermal acclimation, accompanied by a resource allocation bias; its respiration and N allocations are biased toward photosynthetic tissues. In contrast, the monocarpic *S. flexicaulis*, which requires rapid height growth for six years after germination, has lost its winter thermal acclimation ability. In this species, respiration and N allocation are biased toward nonphotosynthetic tissues. These differences between the two species are represented by the “assimilation” paradigm in *S. tashiroi* and the “height growth” paradigm in *S. flexicaulis* in life-history strategies (Fig. 5). The lack of thermal acclimation in *S. flexicaulis* leaves will be well adjusted to their unique evolution towards the monocarpic, masting phenology within the local *Strobilanthes* group. Taking into account the huge energy costs for thermal acclimation and the required rapid height growth under intensive intraspecific competition, the costly thermal acclimation of foliar cells will decrease the instantaneous height-growth rates and resultantly reduce the plant size and seed production in *S. flexicaulis* because of the shading by the other individual plants. Therefore, the monocarpic *S. flexicaulis* would be unable to adopt the costly thermal acclimation that involves cell metabolism and N reallocation within the lamina, especially under relatively shaded understory environments. The energy costs of protein turnover amount to approximately 16–61% of ATP production rates in *Alocasia* and *Phaseolus* plants^{23,24}. Active growth of plant tissues requires large amounts of ATP and NADPH for the construction respiration, which possibly modifies the temperature sensitivity of respiration^{17,25}. These facts reveal a previously unknown trade-off related to the regulation of respiration between rapid height growth for a short lifespan (i.e., height growth paradigm) and seasonal physiological acclimation over a long lifespan (i.e., assimilation paradigm).

This finding is parallel to the previously well-known trade-offs related to leaf lifespan at the individual leaf level. Among plant species, long-lived, thick leaves must pay large N, C, and energy costs to ensure that long leaf lifespan^{26–28}, but leaf mechanical resistance is not directly correlated with photosynthetic capacity²⁹. Furthermore, a shade-to-sun and sun-to-shade transfer experiment in potted seedlings showed that the acclimation in photosynthetic capacity under altered light intensity was smaller in a pioneer tree with a short leaf lifespan than in late-successional trees with long leaf lifespans³⁰. Thus, the newly proposed trade-off at the individual plant level is analogous to the well-known trade-offs at the individual leaf level. In this study, we try to provide a link between the “height growth” (for the monocarpic *S. flexicaulis*) and “assimilation” (for the polycarpic *S. tashiroi*) paradigms within life-history strategies with physiological acclimation capacity related to plant lifespan, involving interspecific variations in resource and respiration allocations within a plant body. The monocarpic *S. flexicaulis* has long stems and a high stem mass ratio, while the polycarpic *S. tashiroi* effectively allocates C and N resources and respiration costs into leaves (Fig. 5); the mean stem heights were 113.5 cm and 54.3 cm in the *S. flexicaulis* and *S. tashiroi* plants examined in this study, respectively (data not shown). The low stem respiration rates in *S. flexicaulis* (Fig. 3; Supplementary Table 2) were due to the lignification of long stems to maintain their large overall plant size.

In the polycarpic *S. tashiroi*, the enhanced or maintained carbon gains during cold winters are important for maintaining the long lifespan of the forest understory. This is because carbon starvation within the plant body directly induces plant/tissue death^{31–35}. The seasonal variations in photosynthetic and respiration rates were unable to be explained by those in N concentrations in the polycarpic *S. tashiroi* leaves (Fig. 3g-i). However, the enhanced cell metabolism and PNUE in winter would contribute to substantial carbon gains under cold air temperature and ensure their long lifespan of this species, even securing the huge energy costs for thermal acclimation. Seasonal acclimation, which increases the contents of metabolic protein, Rubisco, photoprotective pigments, and osmoles within single leaves in winter, has been reported in many long-lived, overwintering evergreen trees^{36–38}. Recent studies using *Arabidopsis* have clarified that plant acclimation to lowered temperatures requires the expression of many genes and is a complex process with multiple steps^{39–41}. The polycarpic *S. tashiroi* leaves, but not those of *S. flexicaulis*, exhibited a significant increase in photosynthetic capacity and metabolic respiration in winter. Nevertheless, the molecular mechanisms underlying the lack of acclimation in *S. flexicaulis* are not yet known.

Thermal acclimation of the dark respiration rates of foliar cells has been reported in many vascular plant species in long-term experiments^{3, 7, 42}. Thermal acclimation in the respiratory metabolism of cells depends not only on plant species but also on tissue/organ types^{43–45}. The results for *S. tashiroi* also demonstrated that seasonally thermal acclimation is dependent on tissue type; acclimation is greater in photosynthetic tissues than in nonphotosynthetic tissues. However, the opposite results were also reported in a short-term experiment. A one-week experiment showed that thermal acclimation is less notable in photosynthetic tissues than in nonphotosynthetic tissues in some plant species⁴⁵. This contradiction may be because we have used field-grown understory plants, which have evolutionarily adapted to seasonal temperature variations over the long term. When incorporating the effects of altered temperatures on plant acclimation into climate-warming models, this contradiction may become crucial for predicting global carbon cycling. A meta-analysis clarified that the thermal acclimation of leaf respiration increases with increasing the duration of experimental treatments¹³. Thus, there may still be a gap between the results of laboratory experiments and those of field observations.

We hypothesize that the thermal acclimation capacity has been lost during the evolutionary process from the long-lived polycarpic phenology in *S. tashiroi* to the periodic monocarpic mass flowering phenology in *S. flexicaulis*, accompanied with the life-history change from the “assimilation” to the “height growth” paradigms. This study is the first report to analyze physiological acclimation between closely related plants with distinct life-history strategies, together with the known evolutionary history based on molecular evidence. The lack of thermal acclimation in monocarpic *S. flexicaulis* would be associated with a disadvantage under climate warming, because of the lack of homeostatic regulation. The current results suggest that rare understory plants evolving a fast-growing life history are highly vulnerable to global warming. Approximately 400 *Strobilanthes* species are distributed in the understory of tropical and subtropical forests in the world^{46, 47}, including many species assessed as VU (vulnerable), CR (critically endangered) and EN (endangered) in the IUCN red list category. Many of these species are also reported

to exhibit periodic mass-flowering phenology; for example, *S. flexicaulis*, with masting every 6 years in Japan, *S. fragrans* J.R.I. Wood, with masting every 6 years in Thailand⁴⁸, *S. cerna* Bl. with masting every 9 years in Indonesia⁴⁹, and *S. kunthiana* (Wall. Ex Nees) T. Anders ex Benth. with masting every 12 years in India⁵⁰. Moreover, many *Strobilanthes* species are polycarpic perennials with non-masting phenology, such as *S. tashiroi* and *S. cusia* (Nees) Kuntze in Japan. However, molecular phylogenetic analysis has not yet been completed, and information on flowering phenology is also largely lacking for these species¹⁹. Further detailed study is greatly needed to elucidate the causal mechanisms of the unique periodic monocarpic phenology and to clarify the linkage among life strategy, physiological acclimation, and the molecular mechanisms underlying the thermal acclimation response in plants. The genus *Strobilanthes* could be a sound model taxon for linking several disciplines in plant biology and for contributing to biodiversity conservation under global warming, especially for rare and endangered species.

Methods

All the plant experiments were carried out in accordance with relevant guidelines. The plant materials used in this study were identified by Dr. Satoshi Kakishima in National Museum of Nature and Sciences.

Study sites. The study sites were located in the subtropical forests on Okinawa Island on the Ryukyu Islands of Japan. The population of *Strobilanthes flexicaulis* Hayata (26° 38' N, 127° 55' E, 280 m a.s.l.) and the population of *Strobilanthes tashiroi* Hayata (26° 48' N, 128° 16' E, 365 m a.s.l.) were located on Mt. Yae-dake and Mt. Nishimei-dake, respectively. The *S. tashiroi* population was located at approximately 39 km northeast of the *S. flexicaulis* population. *S. flexicaulis* is a monocarpic, perennial shrub and has the periodic mass flowering of every six years, whereas *S. tashiroi* is a polycarpic perennial herb and has a non-masting phenology [18]. Almost all *S. flexicaulis* plants flowered in autumn and winter of 2015, and the seed germination and seedling growth started in spring 2016. The seedling density under the parent plants was $780 \pm 324 \text{ m}^{-2}$ (mean \pm 1SD) in June 2016, showing high intraspecific competition following germination in the monocarpic *S. flexicaulis*. Their seeds do not have intrinsic dormancy after dropping to the ground. In *S. flexicaulis*, all plants examined in summer in 2019 and in winter in 2020 were thus 3 years old.

Plant materials. Phylogenetic analysis based on the chloroplast and nuclear DNA sequences of the small *Strobilanthes* group within the Ryukyu and Taiwan Islands, has been published. According to the results, *S. flexicaulis* and *S. tashiroi* are closely related species. Furthermore, their evolutionary processes have been estimated as follows: polycarpic perennial phenology via monocarpic perennial phenology, to monocarpic mass/synchronous flowering phenology within the local area¹⁹. Therefore, it is predicted that the periodical phenology of *S. flexicaulis* has locally evolved from the polycarpic phenology of *S. tashiroi*¹⁹.

Climatological measurements. The air temperature at approximately 1 m high was measured at every 1 hr. with a thermistor sensor (TER-51i, T&D Co Ltd, Nagano, Japan) at just near the *S. flexicaulis*

population from 2017 to 2020. From 2017 to 2020, the mean annual precipitation was 2415 mm at the closest climatological station in Nago (observation by the Japan Meteorological Agency). No snow accumulation was not found in the understory.

Photosynthetic measurements. The photosynthetic light-response curves and water-vapor stomatal conductance were measured in the fully expanded young leaves on Sep. 23 and Sep. 24 (summer) in 2019 and on Jan. 24 and 25 (winter) in 2020 with a portable open gas exchange system (LI-6400; LI-COR Inc., Lincoln, NE, USA) in the field. The measurements were conducted with $400 \mu\text{mol mol}^{-1} \text{CO}_2$ in the inlet gas stream. The exposed photon flux density (PFD) was stepwise decreased from 1500, 1000, 700, 400, 300, 200, 100, 70, 50, 30, 20, 10, 7, 4 and $0 \mu\text{mol m}^{-2} \text{s}^{-1}$ with red-blue light emitting diodes. The relative humidity (RH) in the outlet gas stream was adjusted to the ambient air RH. All leaf gas exchange measurements were conducted before noon to avoid the midday stomatal closure. In each season, the fully expanded young leaves of 7–8 individual plants from each species were used for the measurements.

The initial slope (Φ), the light compensation point (LCP) and dark respiration rates (R_d) were calculated for each leaf from the linear regression between low PFDs (10, 7, 4 and $0 \mu\text{mol m}^{-2} \text{s}^{-1}$) and the corresponding net assimilation rates (A_n) ($r^2 = 0.999 - 0.858$). Light-response curves in net assimilation rates (A_n) were calculated by using the following equation:

$$A_n = \frac{\Phi I + A_{max} - \sqrt{(\Phi I + A_{max})^2 - 4\theta\Phi I A_{max}}}{2\theta} - R_d$$

where A_{max} are the maximum gross assimilation rates, I is the PFD, and θ is the curvature. When $\theta = 0$, the fitting curve is converted into rectangular hyperbola equations; when $\theta = 1$, it is converted into Blackman equations. The values of θ in *S. flexicaulis* were 0.805 and 0.678 in winter and summer, respectively; those in *S. tashiroi* were 0.669 and 0.701 in winter and summer, respectively.

Dark respiration measurements. After the photosynthetic measurements, we collected seven individual plants for which the top canopies were not directly covered by the other plants, in each season or each species. Their stem heights were 113.5 ± 9.0 and 54.3 ± 16.9 cm (mean \pm 1 SD) for *S. flexicaulis* and *S. tashiroi*, respectively. Their stem diameters near the ground were 9.0 ± 2.0 and 4.8 ± 1.1 mm for the monocarpic *S. flexicaulis* and the polycarpic *S. tashiroi*, respectively.

We separated the individual plants into three parts (leaves, stems, and roots) with scissors, and immediately measured their dark respiration rates under the field conditions. The plant parts were put into a closed plastic box with a small fan (1.3 L, 5 L or 14.5 L in volume; the size of the plastic box was selected according to the volume or length of the samples), and then the box was covered with black cloth. To measure the air temperature, copper-constantan thermocouples were placed in the box together with the plant parts. Here, we assumed that the plant temperature was in equilibrium with the air

temperature in the box. We measured the increasing rates in air CO₂ concentration in the box for approximately 3–5 min, using a nondispersive infrared CO₂ gas analyzer (GMP343, Vaisala Inc., Helsinki, Finland) connected with a data logger (GL240, Graphtec Co. Ltd., Yokohama, Japan). From the CO₂ increasing rates and the box volume, we calculated the dark respiration rates. After the field measurements, the plant samples were dried at 60°C for at least three days and then the dry mass was weighed. Dry mass-based respiration rates were calculated. Furthermore, to evaluate metabolic activity in cells at a given temperature, the respiration rates were standardized at 19°C and 28°C in both seasons by using $Q_{10} = 2^{3,15}$. The chosen values of 19°C and 28°C were the mean box temperatures in winter and summer, respectively.

Allocations of dry matter, whole-plant respiration and N in plants. To evaluate the interspecific variations in plant form, the plant samples used for respiration measurements were used to calculate dry matter allocation. Leaf areas of individual plants were calculated from the leaf dry mass in individual plants and the mean values of leaf mass per area (LMA) in each species. The standardized whole-plant respiration rates were calculated from the mean dry matter allocation in each species and the mean dark respiration rates of each tissue in each season.

The N concentrations in the plant tissues were also examined. Each tissue in individual plants was milled into powder with a cutter mill (ABSOLUTE 3, Osaka Chemical Co. Ltd., Osaka, Japan). The N concentrations were measured with an NC analyzer (Vario Max CN, Elementar Analysensysteme GmbH Co. Ltd., Hanau, Germany). The standardized N and respiration allocations among tissues in the plants were calculated from the mean dry matter allocation in each species and the mean values of N and respiration in each tissue and each season.

Statistics. Statistical analyses were conducted with R Ver. 3.0.2 (R Development Core Team, R Foundation for Statistical Computing, Vienna, Austria). To evaluate seasonal acclimation, the significant differences between winter and summer were examined with ANOVA at $P < 0.05$ (electronic supplementary material, table S1). The significant differences between species were also examined with ANOVA at $P < 0.05$ (electronic supplementary material, table S2). The mean (1 SD) values and all statistical results are shown in the supplementary materials.

Declarations

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Author contributions

A.I., T.N. and S.K. conducted the field observation. T.N. calculated respiration rates and S.T.S. measured nitrogen contents. A.I., J.Y. and S.K. wrote this MS. All authors discussed and commented on the manuscript. All authors reviewed the manuscript. All images of photograph and illustration are created by A.I.

Additional Information

Supplementary information accompanies this paper at <https://doi.org/>

Competing interests: The authors declare no competing interests.

References

1. IPCC AR5. Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report to the Intergovernmental Panel on Climate Change. (Cambridge University Press, 2013).
2. Atkin, O. K. *et al.* 2008 Using temperature-dependent changes in leaf scaling relationships to quantitatively account for thermal acclimation of respiration in a coupled global climate–vegetation model. *Global Change Biol.* **14**, 2709–2726. (2008).
3. Reich, P. B. *et al.* Boreal and temperate trees show strong acclimation of respiration to warming. *Nature.* **531**, 633–636 (2016).
4. Hikosaka, H., Ishikawa, K., Borjigidai, A., Muller, O. & Onoda, Y. Temperature acclimation of photosynthesis: mechanisms involved in the changes in temperature dependence of photosynthetic rate. *J. Exp. Bot.* **57**, 291–302 (2006).
5. Ow, L. F., Griffin, K. L., Whitehead, D., Walcroft, A. S. & Turnbull, M. H. Thermal acclimation of leaf respiration but not photosynthesis in *Populus deltoides* × *nigra*. *New Phytol.* **178**, 123–134 (2008).
6. Way, D. A. & Yamori, W. Thermal acclimation of photosynthesis: on the importance of adjusting our definitions and accounting for thermal acclimation of respiration. *Photosynth. Res.* **119**, 89–100 (2014).
7. Heskell, M. A. *et al.* Convergence in the temperature response of leaf respiration across biomes and plant functional types. *Proc. Natl. Acad. Sci. USA* **113**, 3832–3837(2016).
8. Sage, R. F. & Kubien, D. S. The temperature response of C3 and C4 photosynthesis. *Plant Cell Environ.* **30**, 1086–1106 (2007).
9. Scafaro, A. P. *et al.* Strong thermal acclimation of photosynthesis in tropical and temperate wet-forest tree species: the importance of altered Rubisco content. *Global Change Biol.* **23**, 2783–2800 (2017).
10. Atkin, O. K. & Tjoelker, M. G. Thermal acclimation and the dynamic response of plant respiration to temperature. *Trend. Plant Sci.* **8**, 343–351 (2003).

11. Yamori, W., Hikosaka, K. & Way, D. A. Temperature response of photosynthesis in C3, C4, and CAM plants: temperature acclimation and temperature adaptation. *Photosynth. Res.* **119**, 101–117 (2014).
12. Atkin, O. K. *et al.* Global variability in leaf respiration in relation to climate, plant functional types and leaf traits. *New Phytol.* **206**, 614–636 (2015).
13. Slot, M. & Kitajima, K. General patterns of acclimation of leaf respiration to elevated temperatures across biomes and plant types. *Oecologia.* **177**, 885–900 (2015).
14. Atkinson, L. J., Hellicar, M. A., Fitter, A. H. & Atkin, O. K. Impact of temperature on the relationship between respiration and nitrogen concentration in roots: an analysis of scaling relationships, Q10 values and thermal acclimation ratios. *New Phytol.* **173**, 110–120 (2007).
15. Wei, X. *et al.* Consistent leaf respiratory response to experimental warming of three North American deciduous trees: a comparison across seasons, years, habitats and sites. *Tree Physiol.* **37**, 285–300 (2017).
16. Kruse, J., Hopmans, P. & Adams, M. A. Temperature responses are a window to the physiology of dark respiration: differences between CO₂ release and O₂ reduction shed light on energy conservation. *Plant Cell Environ.* **31**, 901–914 (2008).
17. Kruse, J., Rennenberg, H. & Adams, M. A. Steps towards a mechanistic understanding of respiratory temperature responses. *New Phytol.* **189**, 659–677 (2011).
18. Kakishima, S., Yoshimura, J., Murata, H. & Murata, J. 6-year periodicity and variable synchronicity in a mass-flowering plant. *PLoS ONE.* **6**, e28140 <https://doi.org/10.1371/journal.pone.002814> (2011).
19. Kakishima, S. *et al.* Evolutionary origin of a periodical mass-flowering plant. *Ecol. Evol.* **9**, 4373–4381 (2019).
20. Björkman, O. & Denning, B. Photon field of O₂ evolution and chlorophyll fluorescence characteristics at 77K among vascular plants of diverse origins. *Planta.* **170**, 489–504 (1987).
21. Stirling, C. M., Aguilera, C., Baker, N. R. & Long, S. P. Changes in the photosynthetic light response curve during leaf development of field-grown maize with implications for modeling canopy photosynthesis. *Photosynth. Res.* **42**, 217–225 (1994).
22. Ishida, A., Uemura, A., Koike, N., Matsumoto, Y. & Ang, L. H. Interactive effects of leaf age and self-shading on leaf structure, photosynthetic capacity and chlorophyll fluorescence in the rain forest tree. *Dryobalanops aromatica.* *Tree Physiol.* **19**, 741–747 (1999).
23. Bouma, T. J. *et al.* Respiratory energy requirements and rate of protein turnover in vivo determined by the use of an inhibitor of protein synthesis and a probe to assess its effect. *Physiol. Plant.* **92**, 585–594 (1994).
24. Noguchi, K. *et al.* Costs of protein turnover and carbohydrate export in leaves of sun and shade species. *Aust. J. Plant Physiol.* **28**, 37–47 (2001).
25. Stephanie, S. Y. *et al.* Respiratory alternative oxidase responds to both low- and high-temperature stress in *Quercus rubra* leaves along an urban-rural gradient in New York. *Funct Ecol.* **25**, 1007–1017 (2011).

26. Reich, P. B., Walters, M. B. & Ellsworth, D. S. From tropics to tundra: Global convergence in plant functioning. *Proc. Natl. Acad. Sci. USA* **94**, 13730–13734(1997).
27. Wright, I. J. *et al.* The worldwide leaf economics spectrum. *Nature*. **428**, 821–827 (2004).
28. Ishida, A. *et al.* Coordination between leaf and stem traits related to leaf carbon gain and hydraulics across 32 drought-tolerant angiosperms. *Oecologia*. **156**, 193–202 (2008).
29. He, P. *et al.* Leaf mechanical strength and photosynthetic capacity vary independently across 57 subtropical forest species with contrasting light requirements. *New Phytol.* **223**, 607–618(2019).
30. Yamashita, N., Koike, N. & Ishida, A. Leaf ontogenetic dependence of light acclimation in invasive and native subtropical trees of different successional status. *Plant, Cell Environ.* **25**, 1341–1356 (2002).
31. McDowell, N. *et al.* Mechanisms of plant survival and mortality during drought: why do some plants survive while others succumb to drought? *New Phytol.* **178**, 719–739(2008).
32. Sala, A., Piper, F. & Hoch, G. Physiological mechanisms of drought-induced tree mortality are far from being resolved. *New Phytol.* **186**, 274–281 (2010).
33. O'Brien, M. J., Leuzinger, S., Philipson, C. D., Tay, J. & Hector, A. Drought survival of tropical tree seedlings enhanced by non-structural carbohydrate levels. *Nat. Clim. Change*. **4**, 710–714 (2014).
34. Saiki, S. T., Ishida, A., Yoshimura, K. & Yazaki, K. Physiological mechanisms of drought-induced tree die-off in relation to carbon, hydraulic and respiratory stress in a drought-tolerant woody plant. *Sci. Rep.* **7**, 2995 <https://doi.org/10.1038/s41598-017-03162-5> (2017).
35. Kono, Y. *et al.* Initial hydraulic failure followed by late-stage carbon starvation leads to drought-induced death in the tree *Trema orientalis*. *Commun. Biol.* **2**, 8 <https://doi.org/10.1038/s42003-018-0256-7> (2019).
36. Ebbert, V., Adams, I. I. I. W. W., Mattoo, A. K., Sokolenko, A. & Demming-Adams, B. Up-regulation of a photosystem II core protein phosphatase inhibitor and sustained D1 phosphorylation in zeaxanthin-retaining, photoinhibited needles of overwintering Douglas fir. *Plant Cell Environ.* **28**, 232–240 (2005).
37. Harayama, H., Ikeda, T., Ishida, A. & Yamamoto, S. I. Seasonal variations in water relations in current-year leaves of evergreen trees with delayed greening. *Tree Physiol.* **26**, 1025–1033(2006).
38. Yasumura, Y. & Ishida, A. Temporal variation in leaf nitrogen partitioning of a broad-leaved evergreen tree, *Quercus myrsinaefolia*. *J. Plant Res.* **124**, 115–123 (2011).
39. Fowler, S. & Thomashow, M. F. Arabidopsis transcriptome profiling indicates that multiple regulatory pathways are activated during cold acclimation in addition in to the CBF cold response pathway. *Plant Cell*. **14**, 1675–1690 (2002).
40. Oono, Y. *et al.* Monitoring expression profiles of Arabidopsis genes during cold acclimation and deacclimation using DNA microarrays. *Funct. Integr. Genomics*. **6**, 212–234 (2006).
41. Nakaminami, K. *et al.* Analysis of differential expression patterns of mRNA and protein during acclimation and de-acclimation in Arabidopsis. *Mol. Cell. Proteomics*. **13**, 3602–3611 (2014).

42. Wang, H. *et al.* Acclimation of leaf respiration consistent with optimal photosynthetic capacity. *Global Chang Biol.* **26**, 2573–2583 (2020).
43. Maseyk, K., Grünzweig, J. M., Rotenberg, E. & Yakir, D. Respiration acclimation contributes to high carbon-use efficiency in a seasonally dry pine forest. *Global Change Biol.* **14**, 1553–1567 (2008).
44. Jarvi, M. P. & Burton, A. J. Acclimation and soil moisture constrain sugar maple root respiration in experimentally warmed soil. *Tree Physiol.* **33**, 949–959 (2013).
45. Smith, N. G., Li, G. & Dukes, J. S. Short-term thermal acclimation of dark respiration is greater non-photosynthetic than in photosynthetic tissues. *AoBP Plants.* **11**, 1–9 (2019).
46. Bennett, J. R. & Scotland, R. E. A revision of *Strobilanthes* (Acanthaceae) in Java. *Kew. Bull.* **58**, 1–82 (2003).
47. Wood, J. R. I. & Scotland, R. W. New and little-known species of *Strobilanthes* (Acanthaceae) from India and South East Asia. *Kew. Bull.* **64**, 3–47 (2009).
48. Vongkamjan, S. & Sampson, F. B. Phenology, seed germination and some vegetative features of *Strobilanthes fragrans* (Acanthaceae), a recently described unusual species, found only in a single Forest Park in Thailand. *Thai Forest Bull. Bot.* **44**, 6–10 (2016).
49. Tsukaya, H., Kakishima, S., Hidayat, A., Murata, J. & Okada, H. Flowering phenology of the nine-year plant, *Strobilanthes cernua* (Acanthaceae). *Tropics.* **20**, 79–85 (2011).
50. Sharma, M. V., Kuriakose, G. & Shivanna, K. R. Reproductive strategies of *Strobilanthes kunthianus*, an endemic, semelparous species in southern Western Ghats, India. *Bot. J. Linn. Soc.* **157**, 155–163 (2008).

Figures

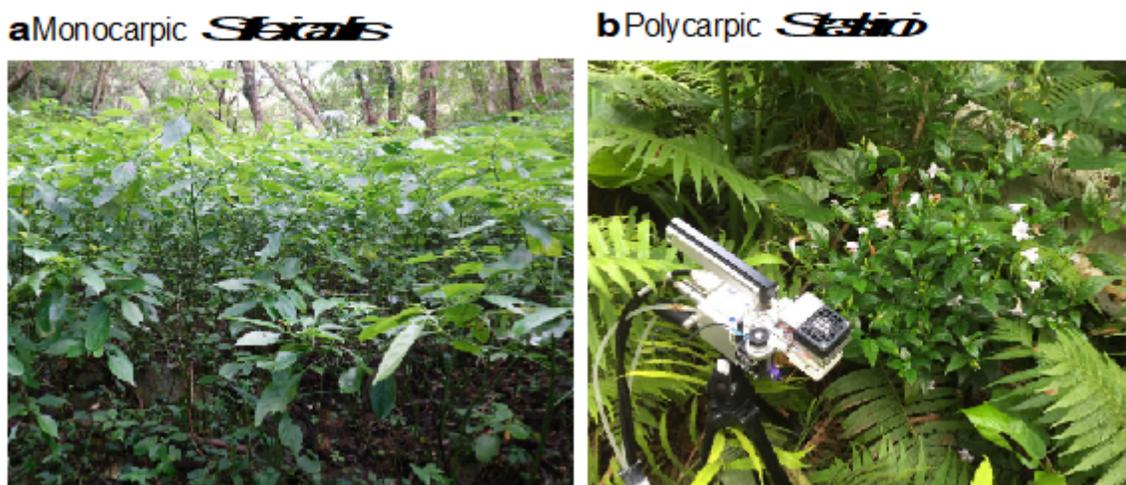


Figure 1

Two *Strobilanthes* species growing in the understory of evergreen forests in Japan. (a) Photographs of the monocarpic *S. flexicaulis*. (b) The polycarpic *S. tashiroi* with the chamber head of a portable open gas exchange system (LI-6400). The maximum top-canopy heights are approximately 2 m and 0.5 m high above the ground in *S. flexicaulis* and *S. tashiroi*, respectively. When the germination started in *S. flexicaulis* in spring 2016, the number of seedlings often exceeded 780 m⁻² under the parent plants.

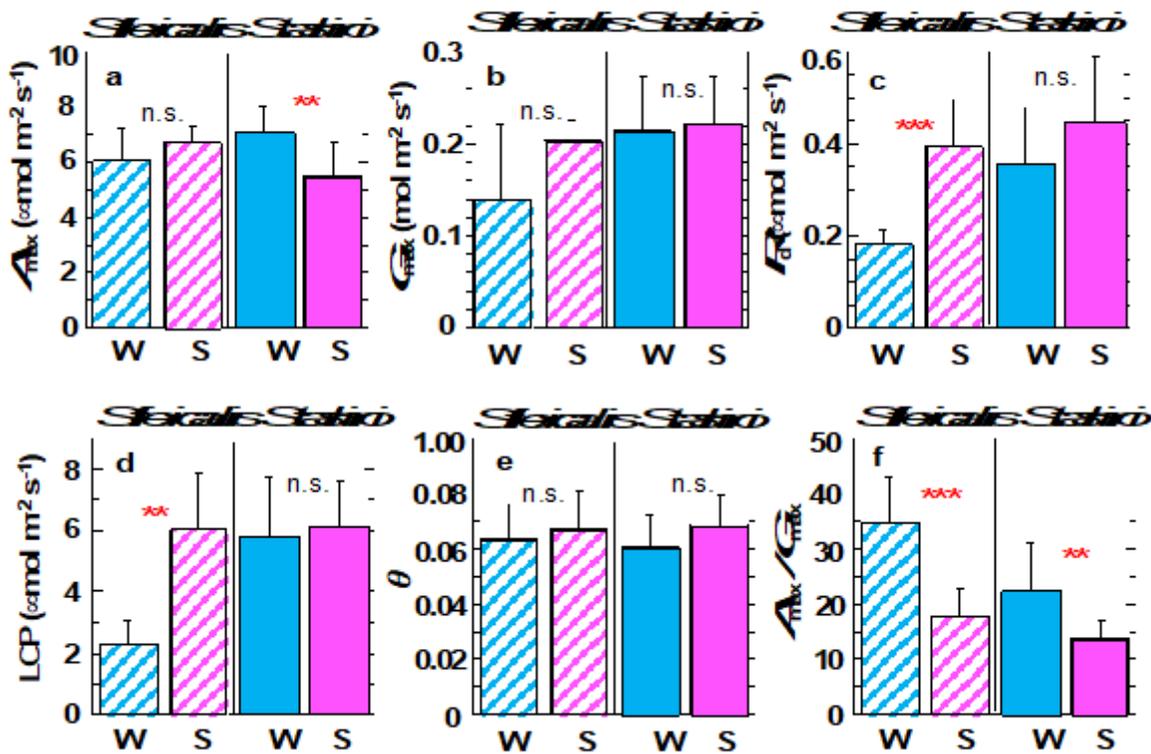


Figure 2

Leaf area-based gas exchange traits measured in field settings in the monocarpic *Strobilanthes flexicaulis* and the polycarpic *S. tashiroi* in winter (blue) and summer (red). (a) A_{max} : maximum net photosynthetic rate. (b) G_{max} : max. water-vapor stomatal conductance. (c) R_d : dark respiration rate. (d) LCP: light compensation point. (e) Φ : initial slope of the light-response curve. (f) A_{max}/G_{max} ratio. W and S show the winter and summer measurements, respectively. Bars show + 1 S.D. Significant differences were compared between seasons in each species (***: P < 0.001, **: P < 0.01, n.s.: P > 0.05).

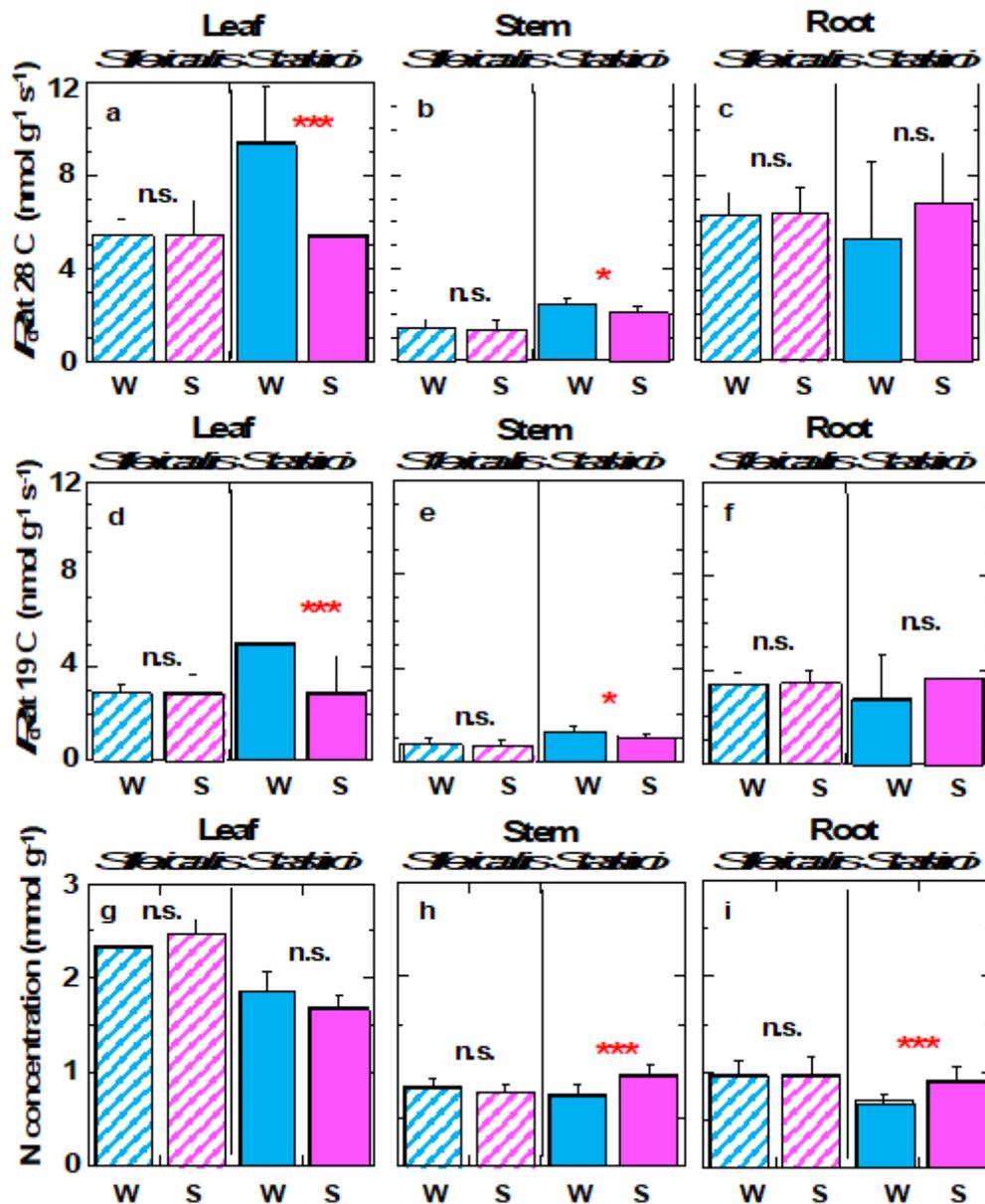


Figure 3

Dry mass-based dark respiration rates and N concentrations in each tissue in the monocarpic *Strobilanthes flexicaulis* and the polycarpic *S. tashiroi*. The respiration rates were measured in the field in each season, and the obtained values were standardized to a constant temperature (19°C and 28°C) by using Q10=2. (a), (d), and (g) show data from leaves, (b), (e), and (h) show data from stems, and (c), (f), and (i) show data from roots. (a), (b), (c) Data at 28°C, and (d), (e), (f) data at 19°C. W and S show the winter and summer measurements, respectively. Bars show + 1 S.D. Significant differences were compared between seasons in each species (***: P<0.001, *: P<0.05, n.s.: P>0.05).

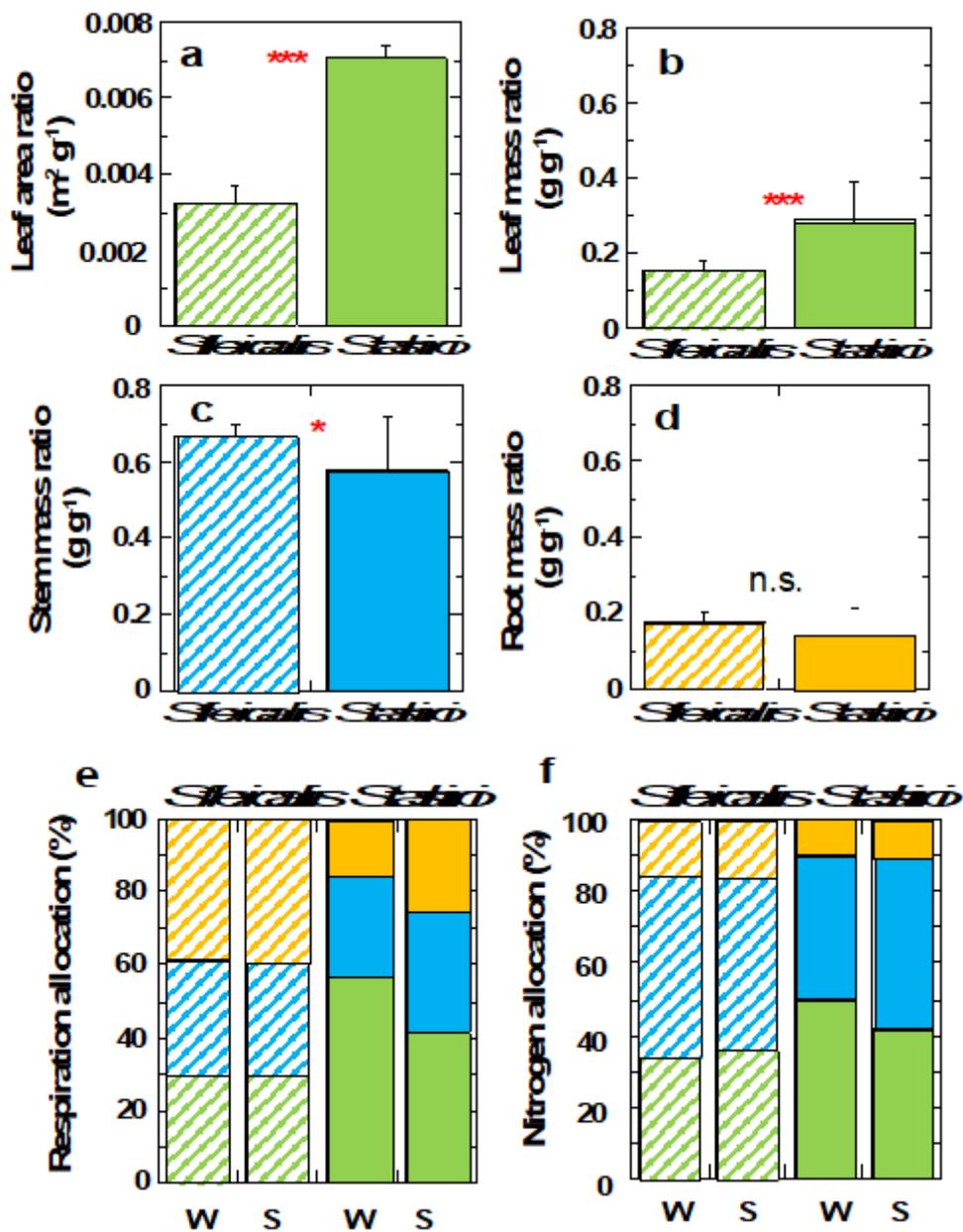


Figure 4

The allocation of dry matter, whole-plant respiration rates and nitrogen contents among tissues in the monocarpic *Strobilanthes flexicaulis* and the polycarpic *S. tashiroi*. (a) Leaf area ratio, (b) leaf mass ratio, (c) stem mass ratio, and (d) root mass ratio in plant dry matter. Bars show + 1 S.D. Significant differences in dry matter allocation were compared between species (***: $P < 0.001$, *: $P < 0.05$, n.s.: $P > 0.05$). e Dark respiration and f nitrogen allocations. W and S show the winter and summer measurements, respectively. Green: leaves; blue: stems; yellow: roots.

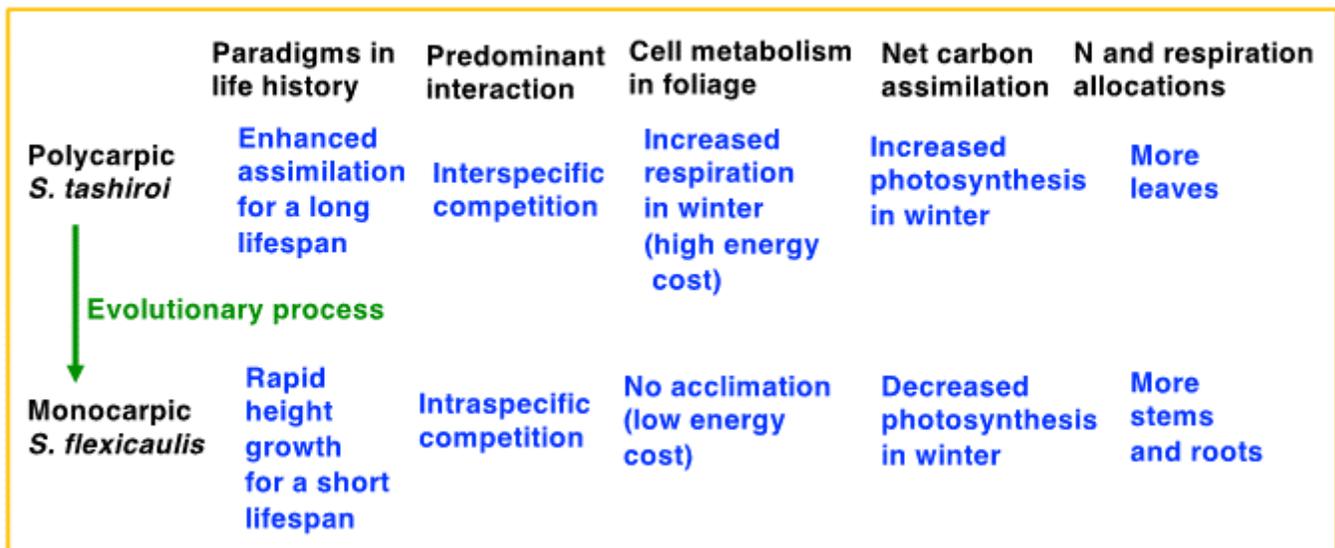


Figure 5

Link between the “height growth” and “assimilation” paradigms in life-history strategies with leaf respiration and nitrogen (N) regulation. Physiological thermal acclimation in *Strobilanthes* leaves accompanied by the evolution from the polycarpic, perennial flowering phenology to the monocarpic, mass-flowering phenology. See the text for more details.

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